# **RESEARCH ARTICLE**



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# Effect of *epimedium pubescen* flavonoid on bone mineral status and bone turnover in male rats chronically exposed to cigarette smoke

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# Abstract

**Background:** *Epimedii herba* is one of the most frequently used herbs in formulas that are prescribed for the treatment of osteoporosis in China and its main constituent is *Epimedium pubescen* flavonoid (EPF). However, it is unclear whether EPF during chronic exposure to cigarette smoke may have a protective influence on the skeleton. The present study investigated the effect of EPF on bone mineral status and bone turnover in a rat model of human relatively high exposure to cigarette smoke.

**Methods:** Fifty male Wistar rats were randomized into five groups: controls, passive smoking groups and passive smoking rats administered EPF at three dosage levels (75, 150 or 300 mg/kg/day) in drinking water for 4 months. A rat model of passive smoking was prepared by breeding male rats in a cigarette-smoking box. Bone mineral content (BMC), bone mineral density (BMD), bone turnover markers, bone histomorphometric parameters and biomechanical properties were examined.

**Results:** Smoke exposure decreased BMC and BMD, increased bone turnover (inhibited bone formation and stimulated its resorption), affected bone histomorphometry (increased trabecular separation and osteoclast surface per bone surface; decreased trabecular bone volume, trabecular thickness, trabecular number, cortical thickness, bone formation rate and osteoblast surface per bone surface), and reduced mechanical properties. EPF supplementation during cigarette smoke exposure prevented smoke-induced changes in bone mineral status and bone turnover.

**Conclusion:** The results suggest that EPF can prevent the adverse effects of smoke exposure on bone by stimulating bone formation and inhibiting bone turnover and bone resorption.

**Keywords:** Smoking, *Epimedium pubescen* flavonoid, Bone mineral density, Bone mineral content, Bone turnover, Bone histomorphometry

# Background

Recent epidemiological studies have yielded increasingly strong evidence that even low level of tobacco smoke exposure poses a risk for health, particularly affecting the lung and skeleton [1-12]. Smoking was also associated with lower areal BMD and reduced cortical thickness in young men [13]. Systemic nicotine may have a significant adverse impact on bone wound healing and inhibit the bone matrix-related gene expressions required for wound healing [14,15].

Tobacco has been identified as the cause for the death of half of its regular users, but insufficient consideration has been given to the hazard of smoking to passive smokers. Thus, a growing interest has been focused on factors that can protect from the harmful action of cigarette smoke.

*Epimedii herba* (Yinyanghuo) is one of the most frequently used herbs in formulas that are prescribed for the treatment of osteoporosis in China and its main



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constituent is *Epimedium pubescen* flavonoid (EPF). Recently, EPF which consist of three phytoestrogenic compounds (Icariin, Genistein and Daidzein), has been fractioned by a series of standardized extractionisolation procedures. Studies have shown the extract of *Epimedii herba* reduces the occurrence of osteoporosis not only in experimental models [16,17] but also in clinical studies [18].

It is unclear whether EPF during chronic exposure to cigarette smoke may have a protective influence on the skeleton. Therefore, this study was designed to evaluate the effects of EPF on BMD, BMC, bone turnover, bone histomorphometry and biomechanical properties in a rat model of human relatively high exposure to cigarette smoke.

# Methods

## Animals and experimental protocol

Fifty adult 8-month-old (body weight  $352 \pm 24$  g) male Wistar rats were purchased from SLRC Laboratory Animal (Shanghai, China). The experiment was performed with the approval of the Animal Research Committee of the Xiangya Hospital, and the animals were cared for in the Experimental Animal Center of Xiangya Hospital. During the whole experiment the animals were housed in stainless steel cages (one rat per cage) at conventional controlled conditions (temperature  $25 \pm 2^{\circ}$ C, relative humidity of  $50 \pm 10\%$ , 12 h light–dark cycle) and had free access to the standard laboratory food and tap water.

The rats were acclimated to conditions for one week before the experiment and randomly assigned to five groups of 10 rats each. One group drank tap water and it served as a control. The smoke-exposed groups were kept in an environment with smoking for 4 months and it served as passive smoking groups, whereas the remaining three groups were supplemented with EPF during exposure to cigarette smoke. EPF (Jiuhui pharmaceutical Corp., Hunan, China) was administered at three dosage levels (75, 150 or 300 mg/kg per day) in drinking water for 4 months and expressed as L-EPF, M-EPF, and H-EPF.

# **Cigarette smoking**

The rats were exposed to sidestream cigarette smoke (CS) from an ashtray which was placed 10 cm below the rat cage in an exposure box (30 cm  $\times$  5 cm  $\times$  15 cm) made of polypropylene, which was placed in a laboratory draft chamber [19]. Using this equipment, the time, amount, and interval of smoke to be sent into the chamber was defined, and 10 cigarettes could be set at a time. All blood samples were collected from the abdominal vena cava into heparinized tubes at the same time of day (9:00–10:00 AM) and centrifuged at 1500 r.p.m. and

4°C for 5 min. Separated plasma was stored at -70°C for subsequent determination of blood nicotine concentrations. The blood concentration of nicotine was measured to reflect the intensity of exposure to smoke. Cigarette smoke was sent for 10 min and then the box was ventilated with room air for 10 min. This procedure was repeated 10 times at 1-h intervals such that the amount of cigarette smoking per day in the rats was equal to in heavy smokers who smoke equal to 30 cigarettes per day [20]. The impact of passive smoking was evaluated by measuring blood nicotine concentrations [21].

## The length of femur and height of the lumbar

The length of the femur and the posterior height and anterior height of the L4 were measured with a precision calliper ( $\pm 0.02$  mm; China Machine Building International Corporation Hunan Co., Hunan, China). These measurements were made in duplicate by the same examiner. The results of the length of the femur were expressed as the mean value of both femurs.

# Bone densitometry

The bone mineral content (BMC) and density (BMD) at femur neck and lumbar vertebrae (L4-L6) were measured ex-vivo by dual-energy X-ray absorptiometry (DXA; Hologic QDR-4500A) equipped with appropriate software for bone assessment in small animals [22]. The scan resolution was  $1.0 \times 1.0$  mm and scan speed was 10 mm/s. The measurements with repositioning of the bones were repeated for three times to calculate the Coefficients of variation (CV). The CV for BMC and BMD on femur neck in our laboratory were 2.1% and 2.2%, and on L4-L6 were 0.9% and 0.8%, respectively. The results of the BMC or BMD in femur neck were expressed as the mean value of both femurs. Similarly to the femur neck, the results of the BMC or BMD in lumbar spine were expressed as the mean value of L4-L6.

# Biomarkers of bone turnover

Serum osteocalcin concentration was measured by an enzyme-linked immunosorbent assay (Rat Mid<sup>TM</sup> Osteocalcin ELISA kit, IDS Inc., Fountain Hills, AZ, USA) according to the manufacturer's instructions. The intraand inter-assay variations of this measurement in our laboratory were 6.8% and 8.9%, respectively. Serum b-ALP was measured by immunoassay using the Access Ostase Assay (Beckman Access, Beckman Coulter Inc., Fullerton, CA, USA). The intra-assay and interassay coefficients of variation were 4.1% and 5.2%, respectively. Serum TRACP 5b level was measured by a solid phase immunofixed enzyme activity assay (Rat TRACP<sup>TM</sup> Assay, IDS Inc., Fountain Hills, AZ, USA) according to the manufacturer's instructions to assess bone resorption.

Group	Control	Smoke	Smoke + L-EPF	Smoke + M-EPF	Smoke + H-EPF
Initial weights(g)	$353\pm23$	351 ± 28	351±16	355 ± 25	352±17
Final weights(g)	$362\pm22$	$360 \pm 26$	$358 \pm 27$	368±29	$364 \pm 28$
Length of femur (mm)	$39.85\pm0.54$	$39.73 \pm 0.46$	39.18±0.71	39.88±0.59	$39.57 \pm 0.66$
Anterior height of L4 (mm)	$7.32 \pm 0.28$	$7.30 \pm 0.24$	$7.35 \pm 0.29$	7.33 ±0.30	7.31 ±0.26
Posterior height of L4 (mm)	$7.63 \pm 0.38$	$7.62 \pm 0.35$	$7.68 \pm 0.34$	7.65 ± 0.29	7.70 ±0.41

Table 1 Body weight, femur length and L4 height

Values are means (for 10 rats) with their standard deviation.

The intra- and inter-assay variations of this measurement in our laboratory were 4.9% and 7.3%, respectively.

#### Bone histomorphometry

The one-third proximal segment of left tibia was embedded in methyl methacrylate (MMA) after Villanueva's staining. A 10- $\mu$ m-thick mid-sagittal section was obtained using a microtome (MR2065; Reichert-Jung, Heidelberg, Germany). Histomorphometry variables were analyzed in both trabecular bone and cortical bone using an imageanalyzing computer (Cosmozone 1 S; Nikon, Tokyo, Japan) linked to a microscope. The parameters measured included trabecular bone volume, trabecular number, trabecular thickness, cortical thickness, BFR/BS (bone formation rate), trabecular separation, osteoclast surface per bone surface and osteoblast surface per bone surface. Standard bone histomorphometrical nomenclatures, symbols, and units were used as described in the report of the American Society of Bone and Mineral Research Histomorphometry Nomenclature Committee [23]. The sections were also carefully polished, etched with a solution of HCl 0.1 N for 60 s, and gold palladium-coated for backscattered scanning electron microscopic (SEM) analysis.





#### Bone biomechanical testing

Biomechanical properties were estimated in a compression test of the L4 performed by AG-IS Universal Testing Instrument (Shanghai, Shimadzu, Japan) [21]. The L4 was compressed until failure along its longitudinal axis at a displacement rate of 2 mm/min using the flat-tipped pin 4.5 mm in diameter. Load– displacement curves were recorded on-line and analyzed for the following bone biomechanical properties: yield load (the force causing the first bone damage), ultimate load (the force at which fracture occurs), displacement at yield (maximal bone deformation under elastic conditions) and at ultimate (bone deformation at the fracture point), stiffness (the slope of the linear, elastic part of the load–displacement curve), and work to the L4 failure.

#### Statistical analysis

Results of all measurements were presented as mean  $\pm$  S.D. for ten rats in each group. A one-way analysis of variance (ANOVA) was first performed to determine whether there were statistically significant (p < 0.05) differences among the experimental groups. Further, the Duncan's multiple range post-hoc test was used for comparisons between individual groups and to determine which means differed statistically significantly (p < 0.05).

#### Results

# Body weight, blood nicotine concentrations, femur length and L4 height

The rats were weighed weekly throughout the whole experiment. The initial weights and final weights are shown in Table 1. At the beginning of the study and during the 4-month experimental period there were no statistically significant differences in the body weight of rats among the five groups (P > 0.05). At termination, the body weight of rats receiving passive smoking or/ and EPF was similar to that noted in the control group reaching  $362.3 \pm 22.2$  g. The mean water consumption was  $11.5 \pm 1.8 \text{ ml}/100 \text{ g body weight/day}$ . Total food and water supply in smoke-exposed groups were similar to that in the control groups.

The concentrations in the controls and the 4-month smoke-exposed rats were  $2.4 \pm 1.3$  ng/ml and  $40.6 \pm 5.9$  ng/ml, respectively. The blood nicotine concentrations in the 75, 150 and 300 mg/kg/d EPF groups receiving passive smoking were  $39.2 \pm 3.7$  ng/ml,  $40.7 \pm 3.5$  ng/ml and  $41.2 \pm 3.9$  ng/ml, respectively. The blood nicotine concentration in the rats receiving passive smoking or/ and EPF was significantly higher than those in controls (P < 0.05).

The femur length and the anterior height and posterior height of L4 in the control rats were 39.85  $\pm$ 



0.54 mm,  $7.32 \pm 0.28 \text{ mm}$  and  $7.63 \pm 0.38 \text{ mm}$ , respectively, after 4 months of the experiment. The femur length and L4 height were unchanged by any treatment (Table 1).

## Bone densitometry

BMD and BMC of the femur neck and lumbar vertebrae are demonstrated in Figure 1. BMD and BMC of the femur neck and lumbar vertebrae were lower (P < 0.05) in the smoke-exposed rats compared to control groups after 4 months of the experiment. There was no significant difference in the BMD and BMC at the femur neck and lumbar vertebrae between the controls and 75 mg/kg/day EPF groups receiving passive smoking. The 150 or 300 mg/kg/day EPF groups receiving passive smoking had greater BMD and BMC at the femur neck and lumbar vertebrae than control groups.

## Bone turnover

Figure 2 shows the results of the bone turnover measurements. Significantly lower osteocalcin concentration and b-ALP activity and higher TRACP 5b levels were found in the smoke-exposed male rats compared to controls. There was no significant difference in serum osteocalcin concentration and b-ALP activity and TRACP 5b levels among the controls, 75, 150, and 300 mg/kg/d EPF groups receiving passive smoking (P > 0.05) (Figure 2).

# SEM and histomorphometric findings

Figure 3 displays five images obtained by SEM examination of the tibia. The rats of the control group revealed normal compactness of the diaphysis and competent trabeculae (Figure 3A). The passive smoking animals showed sparse, uniform thinning of the trabeculae resulting in widened intertrabecular spaces (Figure 3B). The passive smoking and L-EPF group exhibited significant remodeling progress with mineralization (Figure 3C). The group treated with M-EPF (Figure 3D) and H-EPF (Figure 3E) showed nearly essential features of normal bone.

Results concerning bone histomorphometry of the tibia are summarized in Figure 4. In the smoke-exposed rats, trabecular bone volume, trabecular thickness, cortical thickness, trabecular number, BFR/BS and osteoblast surface per bone surface were lower (by 43.5%, 24.2%, 12.5%, 52.7%, 39.4% and 25.4%, respectively), whereas trabecular separation and osteoclast surface per bone surface were greater (by 128.8 and 30.6%, respectively) compared to the control group. Compared to control group, lower trabecular bone volume and BFR/BS (bone formation rate) were found in the 75 mg/kg/d EPF group receiving passive smoking. No differences were noted in the other bone histomorphometry measurements between this group and the control group. In the 150 and 300 mg/kg/d EPF group receiving passive smoking, greater osteoblast surface per bone surface was noted, but there was no significant difference in the other bone histomorphometry measurements, when compared to the control group.

#### **Biomechanical properties**

The biomechanical properties of L4 are shown in Table 2. The bone yield load, ultimate load, displacement at yield, displacement at ultimate, stiffness and work to failure were, respectively 23.8%, 26.5%, 9.2%, 21.9%, 21.8% and







#### (See figure on previous page.)

**Figure 4 Histomorphometric parameters of tibia.** Significantly greater trabecular separation (**F**) and osteoclast surface per bone surface (**H**), lower trabecular bone volume (**A**), trabecular thickness (**B**), trabecular number (**C**), cortical thickness (**D**), BFR/BS (**E**), osteoblast surface per bone surface (**G**) were noted in the smoke-exposed male rats compared to controls. EPF returned the histomorphometric parameters to control levels. Mean  $\pm$  SD (n = 10 in each group). \**P* < 0.05 compared to smoke-exposed rats; \**P* < 0.05 compared to controls.

29.1% lower in smoke-exposed compared to control rats. No significant difference of the biomechanical properties of L4 was noted among the controls, 75, 150, and 300 mg/kg/d EPF groups receiving passive smoking.

## Discussion

The damaging action of chronic smoke exposure on the skeleton and its possible mechanisms have been extensively studied by us and reported in details [24-31]. We demonstrated that the BMD and BMC of the femur neck and lumbar vertebrae were lower in 4-month smoke exposed rats than in controls. Moreover, lower serum osteocalcin concentration and activity of b-ALP and greater TRACP 5b level were noted in the passive smoking group, consistent with the results of previous report [21,28,29]. These results indicated that smoke exposure suppressed bone formation and increased bone resorption. In addition, significant lower trabecular bone volume, trabecular thickness, trabecular number, cortical thickness, bone formation rate and osteoblast surface per bone surface and significant greater trabecular separation and osteoclast surface per bone surface were found in the passive smoking groups. Taking into account the observations made in the present study and the previous reports [24,27,31] the damaging action of smoke exposure on the skeleton might result from its direct action. This mechanism involved stimulation of bone resorption and inhibition of its formation through a direct influence on both differentiation and activity of osteoblastic and osteoclastic bone cells.

Chinese herbal medicine has been widely used for thousands of years in the treatment of fracture and joint diseases. EPF was found to have a preventive effect on osteoporosis by ovariectomy rat models [16,32] and exert a beneficial effect on preventing bone loss for postmenopausal women [33]. Icariin is a major constituent of flavonoids isolated from the *herb Epimedium*. The

previous studies reported that icariin can stimulate proliferation and differentiation of human osteoblasts by increasing production of BMP-2, BMP-4, NO synthesis, promoting the ALP activity and type I collagen expression, subsequently regulating Cbfa1/Runx2, OPG, and RANKL gene expressions [17,34-36]. Meanwhile, icariin also was found to can inhibit osteoclast formation by RANKL and macrophage-colony stimulating factor, and inhibit osteoclast differentiation and bone resorption by suppression of MAPKs/NF- $\kappa$ B regulated HIF-1 $\alpha$  and PGE(2) synthesis[37,38].

However, the beneficial influence of EPF on the skeleton at the exposure to cigarette smoke has not been shown until now. The enhanced serum osteocalcin concentration and b-ALP activity due to EPF indicated an enhanced osteoblastic activity in the whole skeleton. Our findings about TRACP 5b, would indicate that EPF can prevent the smokeinduced increased osteoclast activities. The pattern of change in bone turnover was consistent with the findings from previously published study on EPF in an ovariectomized rat model [39]. Icariin, one of the major flavonoids in EPF (77%), has been reported to stimulate proliferation of rat bone marrow stromal cells and increase the number of colony unit forming-fibroblasts staining positive for alkaline phosphatase in a dose-dependent manner, whereas the alkaline phosphatase activity, OC secretion, and calcium deposition level of rat bone marrow stromal cells were also increased by icariin in a dose-dependent manner, suggesting a potential anabolic effect of icariin on bone [40]. Furthermore, Genistein and Daidzein, the other two flavonoids within EPFs (23%), have also been reported to have a stimulatory effect on protein synthesis and on alkaline phosphatase release by various types of osteoblast cells in vitro [41-43]. Thus, the beneficial effect of EPF consisted in the stimulation of osteoblast activity as well as the reduction of smoke-induced bone resorption.

Table 2 Biomechanical properties of L4 in five groups

Group	Yield load (N)	Ultimate Ioad (N)	Displacement at yield (mm)	Displacement at ultimate (mm)	Stiffness (N/mm)	Work to failure (J)
Control	126.24±8.01*	235.1 ± 38.6*	0.791 ± 0.051*	1.888±0.245*	254.0±64.4*	0.491 ± 0.045*
Smoke	96.24 ± 14.67	172.9±15.9	$0.721 \pm 0.043$	$1.475 \pm 0.136$	198.6±42.3	$0.398 \pm 0.054$
Smoke + L-EPF	117.43±17.09*	225.1 ± 38.6*	$0.782 \pm 0.052^{*}$	1.723±0.167*	238.2±54.7*	0.466±0.039*
Smoke + M-EPF	115.52±15.12*	238.1 ± 36.2*	0.786±0.058*	1.779±0.138*	248.2 ± 46.2*	$0.465 \pm 0.077^*$
Smoke + H-EPF	128.52 ± 16.39*	245.8±38.2*	$0.795 \pm 0.060^{*}$	1.783±0.196*	258.2 ± 35.5*	0.488±0.063*

Mean  $\pm$  SD (n = 10 in each group).

\*P < 0.05 compared to smoke-exposed rats.

Kim BS et al. [44] showed that nicotine suppressed osteoblast proliferation and inhibited the expression of some key osteogenic (TGF- $\beta$ 1, BMP-2) and angiogenic mediators (PDGF-AA, VEGF) in the in vitro experimental model. In addition, Ma L et al. [45] reported that Calcium accumulation, ALP activity, and mRNA levels of ALP, bone sialoprotein (BSP), collagen type I  $\alpha$  1 (Col1 $\alpha$ I), and runt-related transcription factor 2 (Runx2) were significantly decreased by treatment with nicotine, while osteocalcin transcripts decreased by treatment with nicotine. Thus, nicotine may interact with EPF since nicotine also regulates proliferation and differentiation of human osteoblasts and modulates bone metabolism-associated gene expression in osteoblast cells [44,46].

In this study, the 8-month rats were in the final phase of bone mass formation (bone mass consolidation) and its metabolic processes in the bone slow down. The cellular mechanism behind the anabolic effect of EPF has not been established clearly yet. EPF-treatment may induce a positive bone balance in the remodeling cycle, but this has not been shown directly by histomorphometry. Alternatively, part of the bone anabolic effect of icariin or other EPF components may result from the induction of bone formation by modelling. Such an effect has been demonstrated already to be part of the bone anabolic action of parathyroid hormone by Lindsay and Dempster in iliac crest biopsies [47], and the same may be true for icariin. Our study showed that EPF can succeed to bring the histomorphometric parameters and biomechanical properties back to control levels and prevent from all of the adverse effects of smoke exposure on bone. This study also showed that EPF can exhibit a beneficial effect on preventing bone loss in passive smoking rats, which was shown by maintained BMD and BMC at the femur neck and lumbar vertebrae in the EPF treatment group compared with significantly decreased BMD and BMC in the passive smoking group. In addition, it is important to note that the higher mean values of the densitometric parameters were found in the EPF group receiving passive smoking than in the passive smoking group or the control group. The alternative explanations for the increase in BMD may include induction of periosteal and endocortical bone modeling (increased cortical thickness), induction of trabecular micro modeling and prevention of marrow cavity expansion, etc. These results may suggest that EPF via its positive effect on bone might also somewhat induce bone modeling.

The influence of EPF on the bone mineral status and bone turnover during the exposure to cigarette smoke somewhat differed depending on EPF dose. Additional daily consumption of 75 mg EPF/kg body wt was noted to be sufficient in this regard; however, the daily dose of 150 or 300 mg EPF/kg body wt may offer stronger protection. The osteoblast surface and many static parameters of bone geometry indicated that the bone formation was dose dependent. The results suggested that there was a bone formation inducing component in EPF which induced a bone anabolic response independent of its effect on bone resorption. As mentioned above this could be in the form of induction of bone modeling, which was independent of bone resorption.

The effects of EPF on bone have been reported [16]. Thus, this study has not a series of rats treated only with EPF (without smoking). Considering how this medication could be used in the future, it would be necessary to include an additional control group that is kept under normal environment without smoke but receives EPF. That is exactly what we are planning to do in the near future.

Though the present study demonstrate that EPF has a protective effect on the skeleton in smoke-exposed rats, it is not clear whether EPF has a protective effect on the cardio-respiratory effects of smoking and the increased risk of cancer up to now. Even if the smoker can inhibit their bone loss by taking this medicine, smoking also is not ok. According the 2004 Surgeon's General Report, every smoker lives on average 13– 14 years less than non-smokers. Smoking also causes cancer, damages organs, and weakens the immune system. In addition, smoking can also harm those nearby, in terms of second-hand smoke. Therefore, it would still be recommended to quit smoking or never initiate it in the first place.

# Conclusion

In summary, the present study, in a male rat model of relatively high human exposure to cigarette smoke, suggests that EPF supplementation during chronic exposure to cigarette smoke can prevent from smoke-induced bone loss by stimulating bone formation and inhibiting bone turnover and bone resorption, without resulting in a detectable effect on the bone.

#### Competing interests

The author(s) declare that they have no competing interests.

#### Authors' contributions

SGG, LC and GHL were involved in the conceptual discussion and design of the study. WHL and SGG carried out the biochemical analysis. MX, WJ and LCW performed the bone densitometry analysis. FJZ, WFX and YLX performed the bone biomechanical testing. TJ, CZ JPS and QX carried out the bone histomorphometric analysis. SGG and GHL drafted the manuscript. KHL and GHL obtained the funding for National Natural Science Foundation of China. All authors read and approved the final manuscript.

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#### References

- Pleasance ED, Stephens PJ, O'Meara S, McBride DJ, Meynert A, Jones D, Lin ML, Beare D, Lau KW, Greenman C, Varela I, Nik-Zainal S, Davies HR, Ordoñez GR, Mudie LJ, Latimer C, Edkins S, Stebbings L, Chen L, Jia M, Leroy C, Marshall J, Menzies A, Butler A, Teague JW, Mangion J, Sun YA, McLaughlin SF, Peckham HE, Tsung EF, Costa GL, Lee CC, Minna JD, Gazdar A, Birney E, Rhodes MD, McKernan KJ, Stratton MR, Futreal PA, Campbell PJ: A small-cell lung cancer genome with complex signatures of tobacco exposure. Nature 2010, 463:184–190.
- Clauss M, Voswinckel R, Rajashekhar G, Sigua NL, Fehrenbach H, Rush NI, Schweitzer KS, Yildirim AÖ, Kamocki K, Fisher AJ, Gu Y, Safadi B, Nikam S, Hubbard WC, Tuder RM, Twigg HL 3rd, Presson RG, Sethi S, Petrache I: Lung endothelial monocyte-activating protein 2 is a mediator of cigarette smoke-induced emphysema in mice. J Clin Invest 2011, 121:2470–2479.
- Götting M, Ph Addicks J, Spallek M, Jensen AM, Groneberg DA: Exposure to smoke and its effects on pulmonary diseases. *Versicherungsmedizin* 2011, 63:11–16. German.
- Coleman AL, Seitzman RL, Cummings SR, Yu F, Cauley JA, Ensrud KE, Stone KL, Hochberg MC, Pedula KL, Thomas EL, Mangione CM: Study Of Osteoporotic Fractures Research Group: The association of smoking and alcohol use with age-related macular degeneration in the oldest old: the Study of Osteoporotic Fractures. Am J Ophthalmol 2010, 149:160–169.
- Giampietro PF, McCarty C, Mukesh B, McKiernan F, Wilson D, Shuldiner A, Liu J, LeVasseur J, Ivacic L, Kitchner T, Ghebranious N: The role of cigarette smoking and statins in the development of postmenopausal osteoporosis: a pilot study utilizing the Marshfield Clinic Personalized Medicine Cohort. Osteoporos Int 2010, 21:467–477.
- Bjarnason NH, Nielsen TF, Jørgensen HL, Christiansen C: The influence of smoking on bone loss and response to nasal estradiol. *Climacteric* 2009, 12:59–65.
- Jenkins MR, Denison AV: Smoking status as a predictor of hip fracture risk in postmenopausal women of northwest Texas. *Prev Chronic Dis* 2008, 5:A09.
- Wong PK, Christie JJ, Wark JD: The effects of smoking on bone health. Clin Sci (Lond) 2007, 113:233–241.
- Donzé J, Ruffieux C, Cornuz J: Determinants of smoking and cessation in older women. Age Ageing 2007, 36:53–57.
- Oncken C, Prestwood K, Kleppinger A, Wang Y, Cooney J, Raisz L: Impact of smoking cessation on bone mineral density in postmenopausal women. J Womens Health (Larchmt) 2006, 15:1141–1150.
- Bernaards CM, Twisk JW, Snel J, van Mechelen W, Lips P, Kemper HC: Smoking and quantitative ultrasound parameters in the calcaneus in 36-year-old men and women. Osteoporos Int 2004, 15:735–741.
- Kanis JA, Johnell O, Oden A, Johansson H, De Laet C, Eisman JA, Fujiwara S, Kroger H, McCloskey EV, Mellstrom D, Melton LJ, Pols H, Reeve J, Silman A: Smoking and fracture risk: a meta-analysis. Osteoporosis Int 2005, 16(Tenenhouse A):155–156.
- Lorentzon M, Mellstrom D, Haug E, Ohlsson C: Smoking is associated with lower bone mineral density and reduced cortical thickness in young men. J Clin Endocrinol Metab 2007, 92:497–503.
- Berley J, Yamano S, Sukotjo C: The effect of systemic nicotine on osseointegration of titanium implants in the rat femur. J Oral Implantol 2010, 36:185–193.
- Yamano S, Berley JA, Kuo WP, Gallucci GO, Weber HP, Sukotjo C: Effects of nicotine on gene expression and osseointegration in rats. *Clin Oral Implants Res* 2010, 21:1353–1359.

- Qin L, Zhang G, Hung WY, Shi Y, Leung K, Yeung HY, Leung P: Phytoestrogen-rich herb formula "XLGB" prevents OVX-induced deterioration of musculoskeletal tissues at the hip in old rats. *J Bone Miner Metab* 2005, 23:55–61.
- 17. Qian G, Zhang X, Lu L, Wu X, Li S, Meng J: Regulation of Cbfa1 expression by total flavonoids of Herba epimedii. *Endocr J* 2006, **53**:87–94.
- An SJ, Li T, Li E: Effect of kidney-tonifying herbs on ovary function and bone mass in postmenopausal women. *Chin J Osteoporosis* 2000, 6:55–59. Chinese.
- Maehira F, Miyagi I, Kawano N, Kawano M, Shinjoh S: Tobacco smoke exposure alters cholestryl esterase activities and causes accumulation of cholesteryl esters in the rat aorta. J Clin Biochem Nutr 1995, 18:145–155.
- Epping-Jordan MP, Watkins SS, Koob GF, Markou A: Dramatic decreases in brain reward function during nicotine withdrawal. *Nature* 1998, 393:76–79.
- Gasser JA, Ingold P, Venturiere A, Shen V, Green JR: Long-term protective effects of zoledronic acid on cancellous and cortical bone in the ovariectomized rat. J Bone Miner Res 2008, 23:544–551.
- Deyhim F, Stoecker BJ, Brusewitz GH, Devareddy L, Arjmandi BH: Dried plum reverses bone loss in an osteopenic rat model of osteoporosis. *Menopause* 2005, 12:755–762.
- Parfitt AM, Drezner MK, Glorieux FH, Kanis JA, Malluche H, Meunier PJ, Ott SM, Recker RR: Bone histomorphometry: standardization of nomenclature, symbols, and units. Report of the ASBMR Histomorphometry Nomenclature Committee. J Bone Miner Res 1987, 2:595–610.
- Gao SG, Li KH, Xu M, Jiang W, Shen H, Luo W, Xu WS, Tian J, Lei GH: Bone turnover in passive smoking female rat: relationships to change in bone mineral density. *BMC Musculoskelet Disord* 2011, 12:131.
- César-Neto JB, Benatti BB, Manzi FR, Sallum EA, Sallum AW, Nociti FH: The influence of cigarette smoke inhalation on bone density. A radiographic study in rats. *Braz Oral Res* 2005, 19:47–51.
- 26. Ajiro Y, Tokuhashi Y, Matsuzaki H, Nakajima S, Ogawa T: Impact of passive smoking on the bones of rats. *Orthopedics* 2010, **33**:90–95.
- Hapidin H, Othman F, Soelaiman IN, Shuid AN, Luke DA, Mohamed N: Negative effects of nicotine on bone-resorbing cytokines and bone histomorphometric parameters in male rats. J Bone Miner Metab 2007, 25:93–98.
- Cornuz J, Feskanich D, Willett WC, Colditz GA: Smoking, smoking cessation, and risk of hip fracture in women. Am J Med 1999, 106:311–314.
- Hermann AP, Brot C, Gram J, Kolthoff N, Mosekilde L: Premenopausal smoking and bone density in 2015 perimenopausal women. J Bone Miner Res 2000, 15:780–787.
- Broulik PD, Rosenkrancová J, Růzicka P, Sedlácek R, Kurcová I: The effect of chronic nicotine administration on bone mineral content and bone strength in normal and castrated male rats. *Horm Metab Res* 2007, 39:20–24.
- Hapidin H, Othman F, Soelaiman IN, Shuid AN, Mohamed N: Effects of nicotine administration and nicotine cessation on bone histomorphometry and bone biomarkers in Sprague–Dawley male rats. *Calcif Tissue Int* 2011, 88:41–47.
- Zhang G, Qin L, Hung WY, Shi YY, Leung PC, Yeung HY, Leung KS: Flavonoids derived from herbal Epimedium Brevicornum Maxim prevent OVX-induced osteoporosis in rats independent of its enhancement in intestinal calcium absorption. *Bone* 2006, 38:818–825.
- Zhang G, Qin L, Shi Y: Epimedium-derived phytoestrogen flavonoids exert beneficial effect on preventing bone loss in late postmenopausal women: a 24-month randomized, double-blind and placebo-controlled trial. J Bone Miner Res 2007, 22:1072–1079.
- Zhang DW, Cheng Y, Wang NL, Zhang JC, Yang MS, Yao XS: Effects of total flavonoids and flavonol glycosides from Epimedium koreanum Nakai on the proliferation and differentiation of primary osteoblasts. *Phytomedicine* 2008, 15:55–61.
- Zhao J, Ohba S, Shinkai M, Chung UI, Nagamune T: Icariin induces osteogenic differentiation in vitro in a BMP- and Runx2-dependent manner. Biochem Biophys Res Commun 2008, 369:444–448.
- Hsieh TP, Sheu SY, Sun JS, Chen MH, Liu MH: Icariin isolated from Epimedium pubescens regulates osteoblasts anabolism through BMP-2, SMAD4, and Cbfa1 expression. *Phytomedicine* 2010, 17:414–423.
- Hsieh TP, Sheu SY, Sun JS, Chen MH: Icariin inhibits osteoclast differentiation and bone resorption by suppression of MAPKs/NF-κB regulated HIF-1α and PGE(2) synthesis. *Phytomedicine* 2011, 18:176–185.

- Chen KM, Ge BF, Liu XY, Ma PH, Lu MB, Bai MH, Wang Y: Icariin inhibits the osteoclast formation induced by RANKL and macrophage-colony stimulating factor in mouse bone marrow culture. *Pharmazie* 2007, 62:388–391.
- Nian H, Ma MH, Nian SS, Xu LL: Antiosteoporotic activity of icariin in ovariectomized rats. *Phytomedicine* 2009, 16:320–326.
- Chen KM, Ge BF, Ma HP, Liu XY, Bai MH, Wang Y: Icariin, a flavonoid from the herb Epimedium enhances the osteogenic differentiation of rat primary bone marrow stromal cells. *Pharmazie* 2005, 60:939–942.
- 41. Sugimoto E, Yamaguchi M: Anabolic effect of genistein in osteoblastic MC3T3-E1 cells. Int J Mol Med 2000, 5:515–520.
- 42. Sugimoto E, Yamaguchi M: Stimulatory effect of daidzein in osteoblastic MC3T3-E1 cells. *Biochem Pharmacol* 2000, 59:471–475.
- Yamaguchi M, Sugimoto E: Stimulatory effect of genistein and daidzein on protein synthesis in osteoblastic MC3T3-E1 cells: Activation of aminoacyl-t-RNA synthetase. Mol Cell Biochem 2000, 214:97–102.
- Kim BS, Kim SJ, Kim HJ, Lee SJ, Park YJ, Lee J, You HK: Effects of nicotine on proliferation and osteoblast differentiation in human alveolar bone marrow-derived mesenchymal stem cells. *Life Sci* 2012, 90:109–115.
- Ma L, Zwahlen RA, Zheng LW, Sham MH: Influence of nicotine on the biological activity of rabbit osteoblasts. *Clin Oral Implants Res* 2011, 22:338–342.
- Rothem DE, Rothem L, Dahan A, Eliakim R, Soudry M: Nicotinic modulation of gene expression in osteoblast cells, MG-63. Bone 2011, 48:903–909.
- Dempster DW, Cosman F, Kurland ES, Zhou H, Nieves J, Woelfert L, Shane E, Plavetić K, Müller R, Bilezikian J, Lindsay R: Effects of daily treatment with parathyroid hormone on bone microarchitecture and turnover in patients with osteoporosis: a paired biopsy study. J Bone Miner Res 2001, 16:1846–1853.

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