

Omicron Variant (B.1.1.529): Infectivity, Vaccine Breakthrough, and Antibody Resistance

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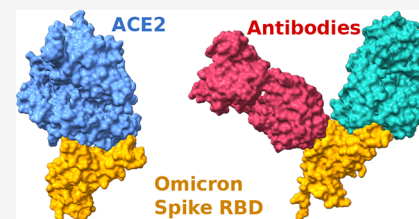


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ABSTRACT: The latest severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) variant Omicron (B.1.1.529) has ushered panic responses around the world due to its contagious and vaccine escape mutations. The essential infectivity and antibody resistance of the SARS-CoV-2 variant are determined by its mutations on the spike (S) protein receptor-binding domain (RBD). However, a complete experimental evaluation of Omicron might take weeks or even months. Here, we present a comprehensive quantitative analysis of Omicron's infectivity, vaccine breakthrough, and antibody resistance. An artificial intelligence (AI) model, which has been trained with tens of thousands of experimental data and extensively validated by experimental results on SARS-CoV-2, reveals that Omicron may be over 10 times more contagious than the original virus or about 2.8 times as infectious as the Delta variant. On the basis of 185 three-dimensional (3D) structures of antibody–RBD complexes, we unveil that Omicron may have an 88% likelihood to escape current vaccines. The U.S. Food and Drug Administration (FDA)-approved monoclonal antibodies (mAbs) from Eli Lilly may be seriously compromised. Omicron may also diminish the efficacy of mAbs from AstraZeneca, Regeneron mAb cocktail, Celltrion, and Rockefeller University. However, its impacts on GlaxoSmithKline's sotrovimab appear to be mild. Our work calls for new strategies to develop the next generation mutation-proof SARS-CoV-2 vaccines and antibodies.



1. INTRODUCTION

On November 26, 2021, the World Health Organization (WHO) announced a new severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) variant Omicron (B.1.1.529), as a variant of concern (VOC). This variant carries an unusually high number of mutations, 32, on the spike (S) protein, the main antigenic target of antibodies generated by either infections or vaccination. In comparison, the devastating Delta variant has only five S protein mutations, which posed a high potential global risk and has spread internationally. Therefore, the “panic button” has been pushed in several cases worldwide, and many countries have enacted travel restrictions to prevent the rapid spread of the Omicron variant.

The mutations on the Omicron variant are widely distributed on multiple proteins of SARS-CoV-2 such as NSP3, NSP4, NSP5, NSP6, NSP12, NSP14, S protein, envelope protein, membrane protein, and nucleocapsid protein. The focus is the mutations on the S protein receptor-binding domain (RBD) for the potential impact on infectivity and antibody resistance caused by this new variant. This is due to the fact that the RBD located on the S protein facilitates the binding between the S protein and the host angiotensin-converting enzyme 2 (ACE2). Such S-ACE2 binding helps SARS-CoV-2 enter the host cell and initiates the viral infection process. Several studies have shown that the binding free energy (BFE) between the S RBD and the ACE2 is proportional to the viral infectivity.^{1–5} As such, an antibody that binds strongly to the RBD would directly neutralize the virus.^{6–8} Indeed, many RBD binding antibodies are generated

by the human immune response to infection or vaccination. Monoclonal antibodies (mAbs) targeting the S protein, particularly the RBD, are designed to treat viral infection. As a result, any mutation on the S protein RBD would cause immediate concerns about the efficacy of existing vaccines, mAbs, and the potential of reinfection.

The global panic brought by the emergence of the Omicron variant drives the scientific community to immediately investigate how much this new variant could undermine the existing vaccines and mAbs. However, relatively reliable experimental results from experimental laboratories will take a few weeks to come out. Therefore, an efficient and reliable in silico analysis is imperative and valuable for such an urgent situation. Here, we present a comprehensive topology-based artificial intelligence (AI) model called TopNetmAb^{9,10} to predict the BFE changes of S and ACE2/antibody complexes induced by mutations on the S RBD of the Omicron variant. The positive BFE change induced by a specific RBD mutation indicates its potential ability to strengthen the binding of an S protein–ACE2/antibody complex, while a negative BFE

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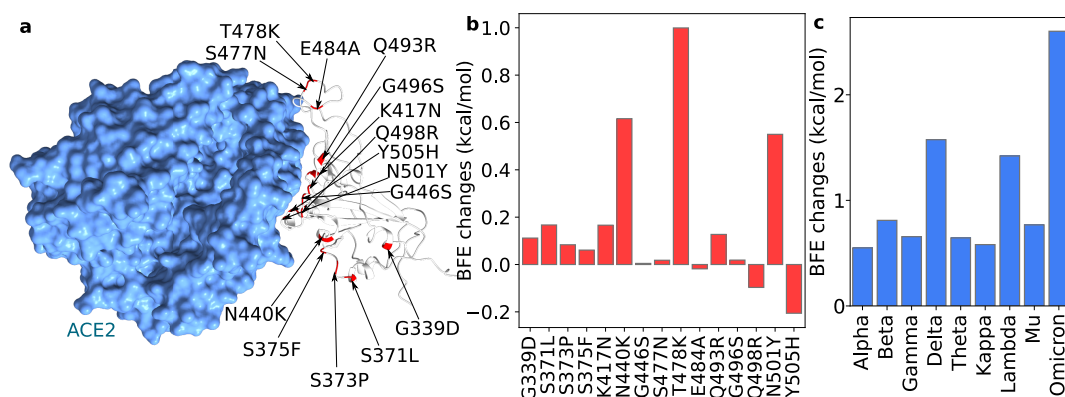


Figure 1. Illustration of the Omicron RBD and ACE2 interaction and RBD mutation-induced BFE changes. (a) 3D structure of the ACE2 and RBD complex (PDB: 6M0J²⁰). Omicron mutation sites are labeled. (b) Omicron mutation-induced BFE changes. Positive changes strengthen the binding between ACE2 and S protein, while negative changes weakened the binding. (c) Comparison of predicted mutation-induced BFE changes for few variants.

change suggests a likely capacity to reduce the binding strength of an S protein–ACE2/antibody complex.

The TopNetmAb model that we proposed has been extensively validated over the past 1.9 years.^{10,11} Initially, in early 2020, we applied our TopNetmAb model to successfully predict that residues 452 and 501 “have high chances to mutate into significantly more infectious COVID-19 strains”.⁹ Such findings have been confirmed due to the emergency of multiple variants such as Alpha, Beta, Gamma, Delta, Theta, Lambda, Mu, and Omicron that carry L452R/Q and N501Y mutations. In April 2021, we provided a list of 31 RBD mutations that may weaken most of the binding to antibodies, such as W353R, I401N, Y449D, Y449S, P491R, P491L, and Q493P.¹⁰ Notably, experimental results have also shown that mutations at residues Y449, E484, Q493, S494, and Y505 might enable the virus to escape antibodies.¹² Meanwhile, in the same work, we also revealed that variants found in the United Kingdom and South Africa in late 2020 may strengthen the binding of the RBD–ACE2 complex, which is consistent with the experimental results.¹³ Later on, we provided a list of the most likely vaccine escape RBD mutations predicted by TopNetmAb, such as S494P, Q493L, K417N, F490S, F486L, R403K, E484K, L452R, K417T, F490L, E484Q, and A475S,¹⁴ and mutations such as S494P, K417N, E484K/Q, and L452R are all detected in the variants of concern and variants of interest denounced by the WHO. Last but not least, the correlation between the experimental deep mutational data¹⁵ and our AI-predicted RBD-mutation-induced BFE changes for all possible 3686 RBD mutations on the RBD–ACE2 complex is 0.7, which indicates the reliability of our TopNetmAb model predictions.¹¹ As a baseline, one may keep in mind that experimental deep mutational results for SARS-CoV-2 PPIs from two different laboratories only have a correlation of 0.67.^{15,16}

This work aims to analyze how the RBD mutations on the Omicron variant will affect the viral infectivity and efficacy of existing vaccines and antibody drugs. Fifteen Omicron RBD mutations, including S371L, S373P, S375F, K417N, N440K, G446S, S477N, T478K, E484A, Q493R, G496S, N501Y, and Y505H, are studied in this work. Additionally, three-dimensional (3D) structures of the RBD–ACE2 complex and 185 antibody–RBD complexes, including many mAbs, are examined to understand the impacts of Omicron RBD mutations. We reveal that Omicron may be over 10 times

more contagious than the original SARS-CoV-2, more infectious than any other named variants, and over twice as infectious as the Delta variant, mainly due to its RBD mutations N440K, T478K, and N501Y. Additionally, Omicron has a high potential to disrupt the binding of most 185 antibodies with the S protein, mainly due to its RBD mutations K417N, E484A, and Y505H, indicating its stronger vaccine-breakthrough capability than the Delta or any other named variants. We have also unveiled that Omicron may seriously reduce the efficacy of the Eli Lilly mAb cocktail because of Omicron RBD mutations K417N, E484A, and Q493R. The Regeneron mAb cocktail may be impaired by Omicron RBD mutations K417N and E484A, and G446S. The efficacy of the AstraZeneca mAb cocktail tixagevimab and cilgavimab may be moderately reduced by Omicron RBD mutation Q498R. Celltrion antibody Regdanvimab may be disrupted by Omicron RBD mutations E484A, Q493R, and Q498R. Omicron RBD mutation E484A may also disrupt Rockefeller University mAbs. However, Omicron’s impacts on GlaxoSmithKline’s mAb are predicted to be mild.

We stated in an earlier work that “we anticipate that as a complementary transmission pathway, vaccine breakthrough or antibody-resistant mutations, like those in Omicron, will become a dominating mechanism of SARS-CoV-2 evolution when most of the world’s population is either vaccinated or infected”.¹⁷ Our present finding shows it is high time to develop a new generation of vaccines and mAbs that will not be prone to viral mutations.

2. RESULTS

2.1. Infectivity. The infectivity of SARS-CoV-2 is mainly determined by the binding affinity of the ACE2 and RBD complex, although the furin cleavage site plays a crucial role as well.¹⁸ Omicron has three mutations at the furin cleavage site and 15 mutations on the RBD, suggesting a significant change in its infectivity. Due to natural selection, the virus enhances its evolutionary advantages at the RBD either by mutations to strengthen the ACE2–RBD binding affinity or by mutations to escape antibody protection.¹⁹ Since the virus has optimized its infectivity in human cells, one should not expect a dramatic increase in the viral infectivity by any single mutation. An effective infection pathway is for the virus to have multiple RBD mutations to cumulatively enhance its infectivity, which appears to be the case for Omicron.

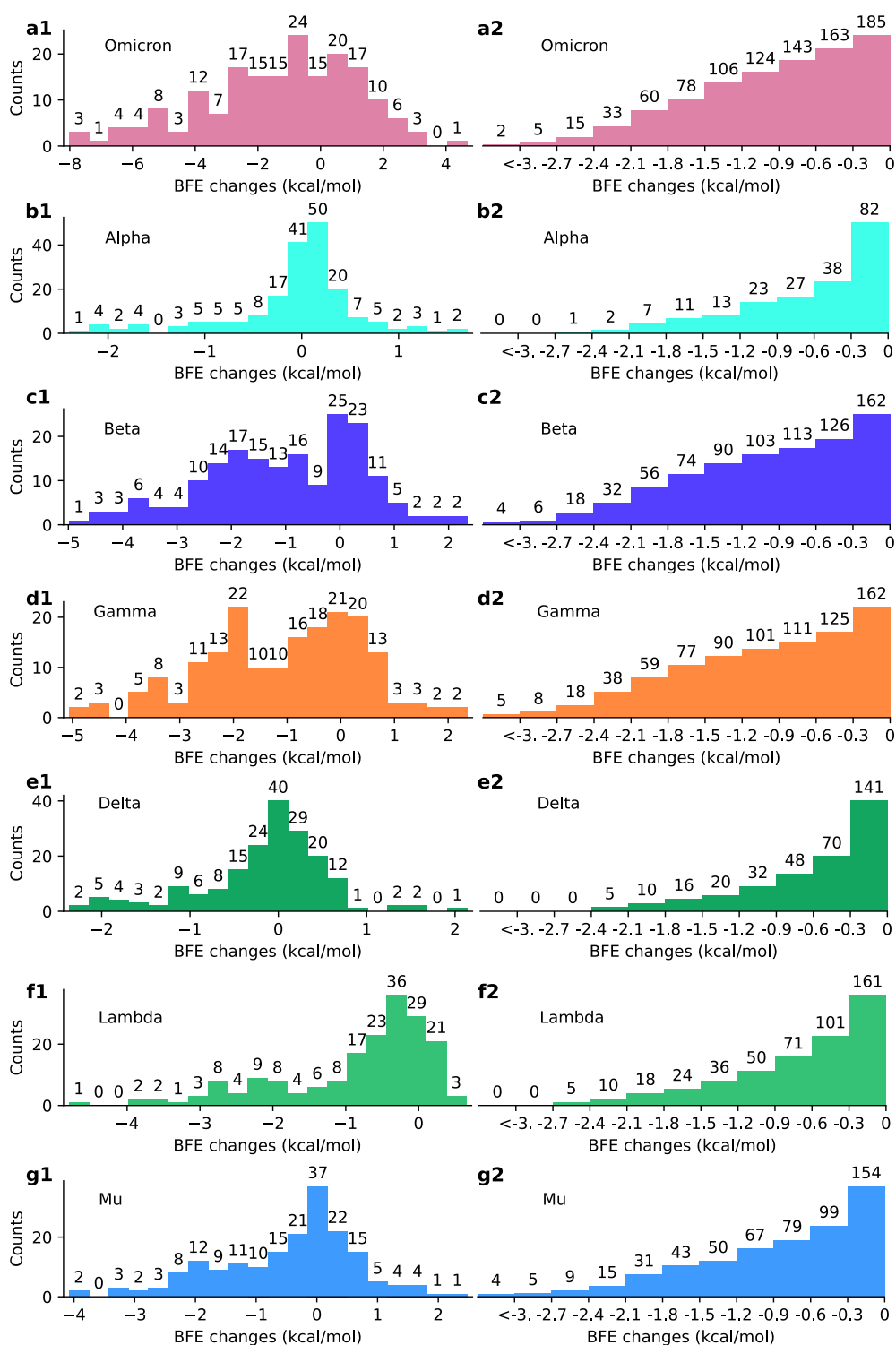


Figure 3. Analysis of variant mutation-induced BFE changes of 185 antibody and RBD complexes. (a1, b1, c1, d1, e1, f1, g1) Distributions (counts) of accumulated BFE changes induced by Omicron, Alpha, Beta, Delta, Gamma, Lambda, and Mu mutations, respectively, for 185 antibody and RBD complexes. Overall, there are more complexes that are weakened upon RBD mutations than complexes that are strengthened. (a2, b2, c2, d2, e2, f2, g2) Numbers (counts) of antibody–RBD complexes regarded as disrupted by Omicron, Alpha, Beta, Delta, Gamma, Lambda, and Mu mutations, respectively, under different thresholds ranging from 0 kcal/mol to -0.3 kcal/mol to less than -3 kcal/mol.

viral-vector vaccines, DNA/RNA vaccines, and protein-based vaccines.²² Essentially, the current COVID-19 vaccines in use mainly target the S protein.²³ The 32 amino acid changes, including three small deletions and one small insertion in the spike protein, suggest that Omicron may be induced by antibody resistance.¹⁷ As a result, these mutations may

dramatically enhance the variant's ability to evade current vaccines.

In general, it is essentially impossible to accurately characterize the full impact of Omicron's S protein mutations on the current vaccines in the world's populations. First, different types of vaccines may lead to different immune

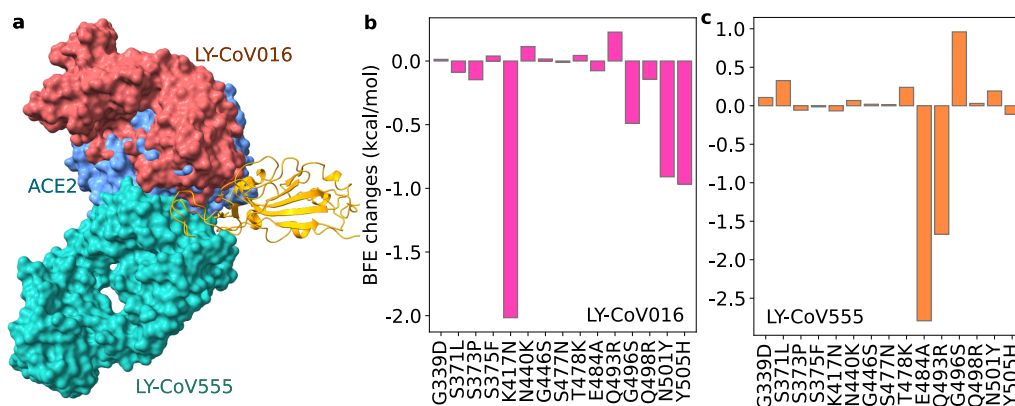


Figure 4. Illustration of the Omicron RBD and Eli Lilly antibody interaction and RBD mutation-induced BFE changes. (a) 3D structure of the ACE2 and Eli Lilly antibody complex. LY-CoV555 (PDB ID: 7KMG²⁴) and LY-CoV016 (PDB ID: 7C01²⁵) overlap on the S protein RBD. ACE2 is included as a reference. (b) Omicron mutation-induced BFE changes for the complex of RBD and LY-CoV016. (c) Omicron mutation-induced BFE changes for the complex of RBD and LY-CoV555.

responses from the same individual. Additionally, different individuals characterized by race, gender, age, and underlying medical conditions may produce different sets of antibodies from the same vaccine. Moreover, the reliability of statistical analysis over populations may be limited because of the inability to fully control various experimental conditions.

This work offers a molecule-based data-driven analysis of Omicron's impact on vaccines through a library of 185 known antibody and S protein complexes. We evaluate the binding free energy changes induced by 15 RBD mutations on these complexes to understand the potential impact of Omicron's RBD mutations to vaccines. To ensure reliability, our study does not include a few known antibody–S protein complexes that are far away from the RBD, such as those in the N-terminal domain (NTD), due to limited experimental data in our antibody library.^{10,11}

Figure 2a, b1, and b2 depict the Omicron RBD mutation-induced BFE changes of 185 known antibody and RBD complexes. Overall, Omicron RBD mutations can significantly change the binding pattern of known antibodies. Positive changes strengthen the binding between antibody and RBD complexes, while negative changes weaken the binding. In the color bar, the largest negative change is more significant than the largest positive change, indicating more severe disruptive impacts. In general, there are more negative BFE changes than positive ones, as shown in Figure 2, indicating that the Omicron mutations favor the escape of current vaccines.

Among 15 RBD mutations, K417N, also part of the Beta variant that originated in South Africa, causes the most significant disruption of known antibodies. Notably, E484A is another mutation that leads to overwhelmingly disruptive effects to many known antibodies. It is worth mentioning that most of E484A's disruptive effects are complementary to those of K417N, which makes Omicron more effective in vaccine breakthroughs. The third disruptive mutation is Y505H. It is also able to weaken many known antibody and RBD complexes.

Mutation G339D creates a mild impact on various antibody–RBD complexes. One of the reasons is that it locates pretty far away from the binding interfaces of most known antibodies. Its change from a noncharged amino acid to a negatively charged amino acid induces mostly favorable bindings among many antibody–RBD complexes. S371L,

S373P, and S375F are other mutations that have mild impacts due to their locations.

For a comparison, ACE2 is also included in Figure 2b1. The impact of Omicron on ACE2 is significantly weak, indicating the SARS-CoV-2 has already optimized its binding with ACE2, and there is a relatively limited potential for the virus to improve its infectivity. However, due to the increase in the vaccination rate, variants can become more destructive to vaccines in years to come.¹⁷

Figure 2a gives a separate plot of the impacts of Omicron on a few mAbs. Similarly, there are dramatic reductions in their efficacy. A more specific discussion is given in the next section.

Figure 3 provides the analysis of variant mutation-induced BFE changes of 185 antibody–RBD complexes induced by Omicron, Alpha, Beta, Delta, Gamma, Lambda, and Mu mutations. From Figure 3a1, it is clear that most complexes have negative accumulated BFE changes, indicating Omicron may disrupt most antibody–RBD binding complexes. In contrast, Delta's distribution focuses on a smaller domain as shown in Figure 3e1. The BFE changes are essentially distributed around zero, suggesting Delta RBD mutations may not disrupt most known antibody–RBD binding complexes. The distributions of Beta and Gamma, respectively, in Figure 3c1 and d1 also indicate potential antibody–RBD binding complex disruption.

It becomes very subtle to judge whether a mutation would disrupt an antibody and RBD complex as Omicron involves multiple vaccine-escape RBD mutations, which may generate multiple cancellations for each antibody–RBD complex over different mutations. It is useful to focus on disruptive mutations, i.e., mutations leading to negative BFE changes. Therefore, we previously have used -0.3 kcal/mol as a threshold to judge whether a mutation disrupts an antibody–RBD complex, which would give us a total of 163 disrupted antibody–RBD complexes as shown in Figure 3a2, suggesting a rate of 0.88 (i.e., 163/185) for potential vaccine breakthrough. As a comparison, Delta has 70 counts and a rate of 0.37 (70/185) in a similar estimation as shown in Figure 3e2. One would have 143 and 48 disrupted antibody and RBD complexes, respectively, for Omicron and Delta if the threshold is increased to -0.6 kcal/mol. In both cases, Omicron is over twice more likely to disrupt antibody–RBD complexes. Note that Beta and Gamma in Figure 3c2 and d2 show a similar pattern.

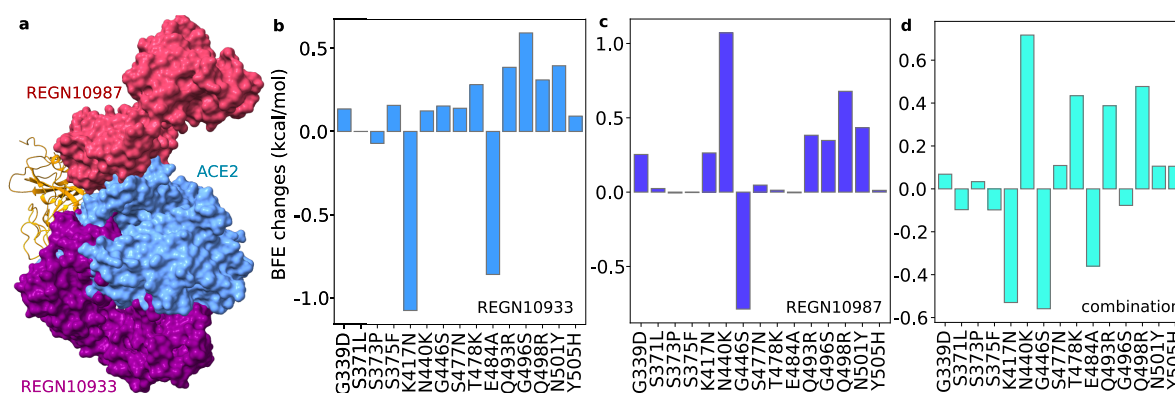


Figure 5. Illustration of the Omicron RBD and Regeneron antibody interaction and RBD mutation-induced BFE changes. (a) 3D structure of the ACE2 and Regeneron antibody complex. REGN10987 and REGN10933 do not overlap on the S protein RBD (PDB ID: 6XDG²⁶). ACE2 is included as a reference. (b) Omicron mutation-induced BFE changes for the complex of RBD and REGN10933. (c) Omicron mutation-induced BFE changes for the complex of RBD and REGN10987. (d) Omicron mutation-induced BFE changes for the complex of RBD, REGN10933, and REGN10987.

2.3. Antibody Resistance. The assessment of Omicron's mutational threats to FDA-approved mAbs and a few other mAbs in clinical development is of crucial importance. Our AI-based predictions of similar threats from other variants, namely, Alpha, Beta, Gamma, Delta, Epsilon, and Kappa, have shown excellent agreements with experimental data.¹¹ In this section, we select a few mAbs, specifically, mAbs from Eli Lilly (LY-CoV016 and LY-CoV555), Regeneron (REGN10933, REGN10987, and REGN10933/10987), AstraZeneca (AZD1061 and AZD8895), GlaxoSmithKline (S309), Celltrion (CT-P59), and the Rockefeller University (C135 and C144). Among them, mAbs from Eli Lilly, Regeneron, AstraZeneca, and GlaxoSmithKline have had FDA approval. In addition, Celltrion's COVID-19 antibody treatment had the EU drug agency's recommendation in November 2021. Rockefeller University's mAbs are still in clinical trials. Our analysis focuses on disruptive RBD mutations.

2.3.1. Eli Lilly mAbs. Eli Lilly mAb LY-CoV555 (PDB ID: 7KMG²⁴) is also known as Bamlanivimab and is used in combination with LY-CoV016 (aka Etesevimab, PDB ID: 7C01²⁵). Antibody LY-CoV016 is isolated from patient peripheral blood mononuclear cells convalescing from COVID-19. It was optimized based on the SARS-CoV-2 virus. The interaction of Eli Lilly mAbs with the S protein RBD is depicted in Figure 4a. ACE2 is included as a reference, indicating both LY-CoV016 and LY-CoV555 can directly neutralize the virus. Clearly, LY-CoV555 has a competing relationship with LY-CoV016, which might complicate our predictions slightly. In this work, we carry out the analysis of Eli Lilly mAbs separately.

Omicron mutation-induced BFE changes for the antibody LY-CoV016 and RBD complex is given in Figure 4b. It appears that LY-CoV555 was optimized with respect to the original S protein but is sensitive to mutations. This complex may be weakened by K417N and N501Y as predicted in our earlier work.¹¹ New mutation Y505H may also reduce LY-CoV016's efficacy. Overall, the complex may be significantly weakened by Omicron, leading to the efficacy reduction of Etesevimab.

The predicted BFE changes of LY-CoV555 are shown in Figure 4c. Mutation E484A induces a negative BFE change of -2.79 kcal/mol for the LY-CoV555 and RBD complex. The BFE change may translate into a dramatic efficacy reduction of

16 times for LY-CoV555, making it less competitive with ACE2 as most Omicron mutations strengthen the S protein and ACE2 binding. Similarly, Q493R may also reduce the efficacy by about 5 times. However, G496S may enhance the binding of the complex. The impacts of other mutations are mild. Therefore, Omicron is expected to reduce LY-CoV555 efficacy significantly. A previous study indicated that LY-CoV555 is prone to the E484K mutation presented in Beta and Gamma variants, for which the Eli Lilly mAb cocktail was taken off the market for many months in 2021.

Although LY-CoV555 and LY-CoV016 might slightly complement, they are both prone to Omicron mutation-induced efficacy reduction. We predict that the Eli Lilly mAb cocktail will be retaken off the market if Omicron becomes a prevailing variant in the world.

2.3.2. Regeneron mAbs. Regeneron mAbs REGN10933 and REGN10987 (aka Casirivimab and Imdevimab, respectively) are FDA-approved antibody cocktails (PDB ID: 6XDG²⁶) against COVID-19. Their 3D structures in complex with the S protein RBD are depicted in Figure 5a. ACE2 is included as a reference. Unlike the Eli Lilly mAb cocktail, the Regeneron mAbs do not overlap each other and bind to different parts of the RBD. Our 3D alignment shows that the antibody REGN10987 does not directly compete with ACE2 on their binding interfaces with the RBD but still spatially conflicts with ACE2. As a result, REGN10987 can directly neutralize the virus but is less sensitive to infectivity-induced RBD mutations. In contrast, REGN10933 overlaps with ACE2 both spatially and on the RBD binding interface. Consequently, REGN10933 is prone to infectivity-induced RBD mutations.

Figure 5b plots our AI predicted BFE changes of the REGN10987-RBD complex. There are mixed responses to various Omicron mutations. Although G446K and K417N induce a negative BFE change, many other mutations may enhance the binding of the complex.

Omicron-induced BFE changes of the REGN10933-RBD complex are given in Figure 5c. Apparently, K417N and E484A induce BFE changes of -1.08 and -0.86 kcal/mol, respectively. However, most other Omicron mutations may strengthen the binding of the complex.

It is interesting to study how the two Regeneron mAbs are affected by Omicron when they are combined. Figure 5d shows

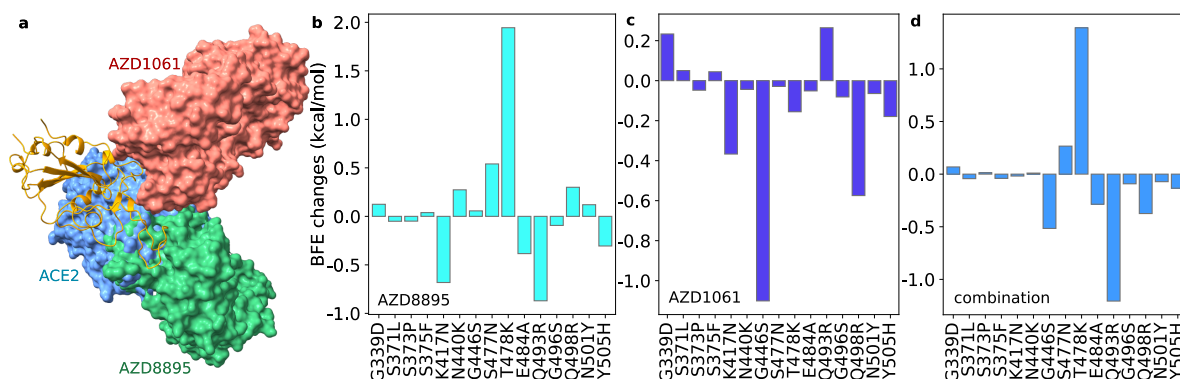


Figure 6. Illustration of the Omicron RBD and AstraZeneca antibody interaction and RBD mutation-induced BFE changes. (a) 3D structure of the ACE2 and AstraZeneca antibody complex. AZD1061 and AZD8895 do not overlap on the S protein RBD (PDB ID: 7L7E²⁷). ACE2 is included as a reference. (b) Omicron mutation-induced BFE changes for the complex of RBD and AZD8895. (c) Omicron mutation-induced BFE changes for the complex of RBD and AZD1061. (d) Omicron mutation-induced BFE changes for the complex of RBD, AZD8895, and AZD1061.

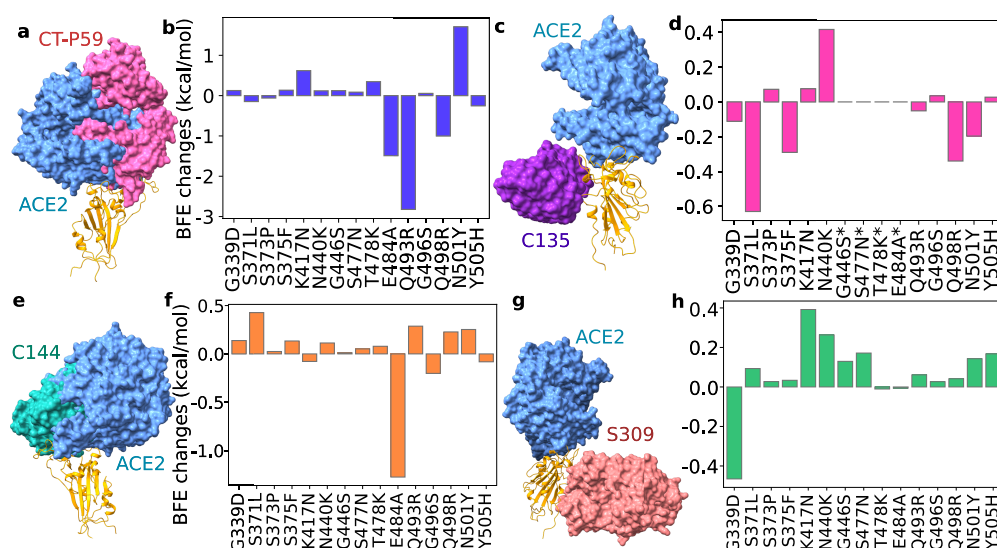


Figure 7. Illustration of the Omicron RBD and other antibodies and RBD mutation-induced BFE changes. (a) Antibody CT-P59 in reference with ACE2. (b) BFE changes of Omicron mutation induced on the binding of CT-P59 and RBD. (c) Antibody C135 in reference with ACE2. (d) BFE changes of Omicron mutation induced on the binding of C135 and RBD. *: no results due to incomplete structure of C135. (e) Antibody C144 in reference with ACE2. (f) BFE changes of Omicron mutation induced on the binding of C144 and RBD. (g) Antibody S309 in reference with ACE2. (h) BFE changes of Omicron mutation induced on the binding of S309 and RBD.

the BFE changes of the complex induced by various Omicron mutations. We note that amplitudes of both positive and negative BFE changes have significantly reduced. However, Omicron RBD mutations K417N, G446S, and E484A may still weaken the cocktail binding to the RBD. We predict that Omicron will have a negative impact on the Regeneron cocktail efficacy.

2.3.3. AstraZeneca mAbs. AstraZeneca mAbs are designed as a cocktail of tixagevimab (AZD8895, PDB ID: 7L7D) and cilgavimab (AZD1061, PDB ID: 7L7E) as in Figure 6a. AZD8895 competes with ACE2 for the same binding interface and thus is able to directly neutralize the virus. However, it is also prone to the infection-induced RBD mutations. As shown in Figure 6b, AZD8895 can be slightly weakened by Q493R and K417N. In contrast, AZD1061 can be significantly disrupted by G477N as shown in 6c. Omicron mutation Q493R can also lead to the binding affinity reduction of the RBD and AZD1061 complex. Omicron impacts on the AstraZeneca cocktail are slightly alleviated as shown in 6d. Since mutation Q493R affects both AZD8895 and AZD1061,

it may give rise to a significant BFE reduction and disrupt the efficacy of the cocktail.

2.3.4. Other mAbs. Celltrion's antibody CT-P59 (aka Regdanvimab, PDB ID: 7CM4) is used as cocktail with CT-P63, for which we do not have its 3D structure. Figure 7a shows that antibody CT-P59 binds the RBD in a competing region with ACE2 and thus might play a more important role than CT-P63 in combating the virus. Figure 7b shows that mutations E484A, Q493R, and Q498R, respectively, lead to BFE changes of -1.49 , -2.82 , and -1.0 kcal/mol for the CT-P59-RBD complex. These disruptive effects may be slightly offset by a positive BFE change of 1.71 kcal/mol due to mutation N501Y, which was reported in our earlier work.¹¹ The impacts of other mutations are relatively mild. Overall, CT-P59 may still be impaired by Omicron. Previously, we have shown that CT-P59 is prone to L452R in Delta and Q439R and S494P.¹¹ Due to the lack of the CT-P63 structure, we cannot provide an inclusive estimation for Celltrion's cocktail but would recommend caution toward the use of Celltrion's Regdanvimab in the wake of Omicron infections.

We also analyze the Rockefeller University antibodies C135 (PDB ID: 7K8Z) and C144 (PDB ID: 7K90), whose binding complexes with the RBD are given in Figure 7c and e, respectively. Antibody C135 has a relatively small region of interface with RBD and does not overlap with ACE2. Our earlier study indicates that C135 is prone to R346K and R346S mutations.¹¹ Mutation S317L induces a BFE change of -0.63 kcal/mol, indicating a relatively weak negative impact on C135's efficacy. In contrast, antibody C144 shares part of its binding domain with ACE2 and has more dramatic responses to Omicron mutations (see Figure 7f). Our earlier study indicates that the efficacy of C144 can be significantly reduced by E484K in the Delta variant.¹¹ Mutation E484A may cause a BFE change of -1.27 kcal/mol. Therefore, we predict that the efficacy of C144 may be also undermined by Omicron RBD mutations.

Finally, we study antibody S309 (PDB ID: 6WPS) which is the parent antibody for Sotrovimab developed by GlaxoSmithKline and Vir Biotechnology, Inc. The alignment of S309 with ACE2 was given an earlier study²⁸ and is also presented in Figure 7g. Since S309 does not overlap with ACE2 both spatially and on the RBD binding interface, infectivity-induced mutations will not affect S309 very much. Figure 7h shows that Omicron-induced BFE changes are from -0.47 to 0.39 kcal/mol. Therefore, Omicron may have minor impacts on S309.

3. DATA, METHODS, AND VALIDITY

Data. To deliver an accurate and reliable machine learning model, a data set collection is of paramount importance among other steps. Both the BFE changes and next-generation sequencing enrichment ratios indicate the mutation-induced effects on protein–protein interactions (PPIs) binding affinities. Our methods integrate these two types of data sets to improve the prediction accuracy.^{10,11} Considering the urgency of COVID-19, the scattered SARS-CoV-2 data concerning BFE changes are reported inconsistently, while the sequencing enrichment ratio data is relatively easy to obtain but consistently has particular protein–protein interaction problems. The method is set up based on the BFE change data set, SKEMPI 2.0,²⁹ together with SARS-CoV-2 related data sets. These data sets are obtained from the mutational scanning on ACE2 binding to the S protein RBD,³⁰ the mutational scanning on RBD binding to ACE2,^{15,16} and the mutational scanning on RBD binding to CTC-445.2 and on CTC-445.2 binding to RBD.¹⁵ We have also collected a library of 185 3D structures of antibody–RBD complexes.¹¹

Methods. Our deep learning model for predicting BFE changes induced by mutations is constructed in two main steps. First, once 3D structures of PPI complexes are obtained, mathematical features and biochemical/biophysical features are extracted. Biochemical/biophysical features provide the chemical and physical information, such as surface areas, partial charges, Coulomb interactions, van der Waals interaction, electrostatics, and more. Mathematical features, including the element-specific and site-specific persistent homology (algebraic topology), are implemented to simplify the structural complexity of PPI complexes.^{9,31} Second, a deep learning algorithm, artificial neural networks (ANNs), is constructed to tackle the massive features and mutational scanning data for predictions,¹¹ which is available at TopNetmAb. Notice that our early model was constructed by integrating convolutional neural networks (CNNs) with gradient boosting trees (GBTs) and was trained with a large data set of 8338 PPI entries from

the SKEMPI 2.0 data set,²⁹ which had already achieved a high accuracy.³¹ In the following, the idea of persistent homology is introduced briefly, which plays a key role in the feature processing. Moreover, the Supporting Information, Sections S3 and S4, present more detailed descriptions of data preprocessing, feature generation, and machine learning methods.

Recent years have seen a booming development of topological data analysis in a wide variety of scientific and engineering problems, whose main workhorse is persistent homology.^{32,33} By using persistent homology, molecular atoms can be modeled as a set of point clouds. Vertices, edges, faces, and more can be treated as simplices σ with their collections to be simplicial complexes X . For a simplicial complex X , a chain is a finite sum of simplices as $\sum_i \alpha_i \sigma_i^k$ with coefficients α_i , and the set of all chains is a group $C_k(X)$, where $k = 0, 1, 2, 3$. Thus, the boundary operator ∂_k maps $C_k(X) \rightarrow C_{k-1}(X)$ defined as $\partial_k \sigma^k = \sum_{i=0}^k (-1)^i [v_0, \dots, \hat{v}_i, \dots, v_k]$, where $\sigma^k = \{v_0, \dots, v_k\}$ and $[v_0, \dots, \hat{v}_i, \dots, v_k]$ is a $(k-1)$ -simplex excluding v_i . This is followed by an important property of boundary operators which is that $\partial_{k-1} \partial_k = 0$ and

$$\dots \xrightarrow{\partial_{k+1}} C_k(X) \xrightarrow{\partial_k} C_{k-1}(X) \xrightarrow{\partial_{k-1}} \dots \xrightarrow{\partial_2} C_1(X) \xrightarrow{\partial_1} C_0(X) \xrightarrow{\partial_0} 0 \quad (1)$$

and the k th homology group H_k is defined by $H_k = Z_k/B_k$, where $Z_k = \ker \partial_k = \{c \in C_k \mid \partial_k c = 0\}$ and $B_k = \text{im } \partial_{k+1} = \{\partial_{k+1} c \mid c \in C_{k+1}\}$. The Betti numbers are defined by the ranks of k th homology group H_k . This, in practice, is counting holes in the k dimension, such as β_0 reflects the number of connected components, β_1 gives the number of loops, and β_2 is the number of cavities. Then, persistent homology can be devised to track Betti numbers along a filtration in order to describe the topological, spatial, and geometry information, which generates features for machine learning.

Validity. In more recent work,^{10,11,19} with the help of the aforementioned deep mutational data sets associated with SARS-CoV-2, our predictions are highly consistent with experimental data. The predictions for the binding of CTC-445.2 and S protein RBD were compared with experimental data with a Pearson correlation of 0.7.^{11,15} In the same work,¹¹ the predictions of emerging mutations on clinical trial antibodies had a Pearson correlation of 0.8 with the natural log of experimental escape fractions.³⁴ In addition, the predicted mutation-induced BFE changes on L452R and N501Y for the ACE2-RBD complex have a near perfect correlation with experimental luciferase data.^{11,35}

Our TopNetmAb model assumes that the RBD mutations are independent, which is very reasonable for Delta and other variants as they involve only one, two, or three isolated RBD mutations. As shown in Figure 1a, adjacent Omicron mutations S477N and T478K are dependent on each other. Similarly, S373P is just one residue away from S371L and S375F and is deemed to be depending on its neighbors. However, these three mutations are pretty far away from the ACE2 binding interface and play less important roles in our predictions. Mutations G496S and Q498R are also one amino acid apart, albeit their predicted BFE change amplitudes are very small. Overall, we expect a larger error in our prediction of Omicron infectivity compared to our earlier successful predictions.^{10,11,14} However, we are still confident that the predicted trend of the Omicron infectivity change is correct. Figure 3 shows that the most severe antibody disruptions are not obtained from the interdependent mutations (i.e., S371L,

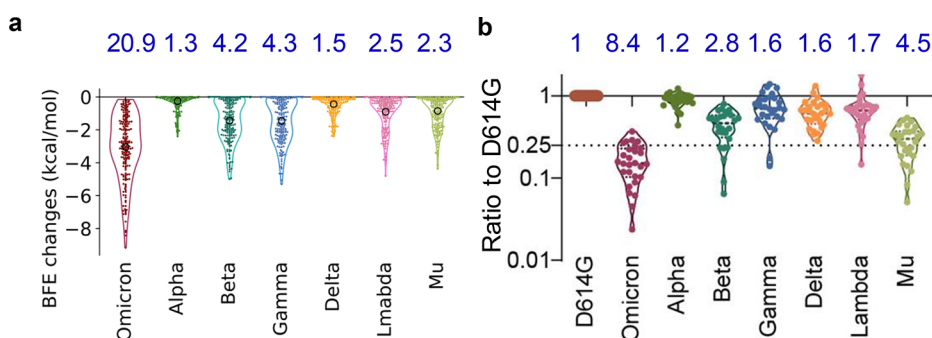


Figure 8. Comparison of predicted variant vaccine-breakthrough potential with experimental data. (a) Accumulated negative BFE changes induced by Omicron, Alpha, Beta, Delta, Gamma, Lambda, and Mu mutations, respectively, for 185 antibody–RBD complexes. For each variant, the number on the top is the fold of affinity reduction computed by $e^{-\text{BFEchange}_{\text{average}}}$, where $\text{BFEchange}_{\text{average}}$, denoted by a circle, is the mean value of 185 antibody–RBD negative BFE changes. (b) The comparison of neutralization activity against Omicron, Alpha, Beta, Delta, Gamma, Lambda, and Mu variants based on 28 convalescence sera.³⁷ For each variant, the number on the top is the ratio of neutralization ED_{50} compared to the reference strain D614G.

	REGN10933	REGN10987	LY-CoV016	LY-CoV555	S2X259	S309	DH1047	Fab 2-7
BFE changes								
Fold changes								

Figure 9. Comparison of predicted antibody-breakthrough potential with experimental data.³⁸ Colors indicate three different ranges. For BFE changes: dark red, BFE changes ≤ -2 kcal/mol; median red, -2 kcal/mol $<$ BFE changes ≤ -1 kcal/mol; light red, BFE changes > -1 kcal/mol. For fold changes: dark red, fold changes < -1000 ; median red, $-1000 \leq$ fold changes < -100 ; light red, fold changes ≥ -100 .

S373P, S375F, S477N, T478K, G496S, and Q498R), suggesting the predicted trends of antibody disruptions are still valid. The reliability and accuracy of our assumption for Omicron are to be validated by experimental data, which may become available in a few weeks.

4. NOTE ADDED IN PROOF

After the publication of our manuscript in arXiv on December 1, 2021,³⁶ two sets of experimental results have been released recently.^{37,38} The first set of experimental results is the sensitivity of serum samples from COVID-19 convalescent patients,³⁷ and the second set of experimental results is about antibody evasion impacted by the Omicron variant.³⁸

Figure 8a provides a comparison of accumulated BFE changes for variants Omicron, Alpha, Beta, Gamma, Lambda, and Mu. For each antibody–RBD complex, the accumulated negative BFE change is obtained by the summation over RBD mutations (e.g., 15 mutations for Omicron and two for Delta) with positive BFE changes being set to zero, so that only disruptive effects are compared. Therefore, there are 185 accumulated BFE changes for each variant. The mean value of these 185 values is used to compute the fold of affinity reduction, which can be compared for different variants against the original variant ($\text{BFEchange}_{\text{average}} = 0$). It appears that Omicron is near 14 folds as capable as Delta and near 5 folds as capable as Gamma to escape vaccines.

In Figure 8b, the sensitivity of 28 serum samples from COVID-19 convalescent patients infected with the SARS-CoV-2 original strain was tested against pseudotyped Omicron, Alpha, Beta, Gamma, Delta, Lambda, and Mu.³⁷ The mean neutralization ED_{50} of these sera against Omicron decreased about 8.4 folds compared to the D614G reference strain. In contrast, Delta neutralization capability decreased 1.6 fold. Both our prediction and serum experiment indicate that Omicron has the highest capability to evade vaccines. The

overall correlation between our prediction and experiment is 0.9.

Figure 9 illustrates the comparison of BFE change predictions and the experimental data of fold changes in IC_{50} compared with the wild type.³⁸ The BFE changes for each antibody are calculated by the peak negative BFE changes. From the figure, it is shown that only predictions for antibodies from Regeneron do not highly match to the experimental data, but the rest of predictions are perfectly consistent with the experimental results.

5. CONCLUSION

The identification of Omicron as a variant of concern (VOC) by the World Health Organization (WHO) has triggered countries around the world to put in place travel restrictions and precautionary measures. At this moment, the scientific community knows little about Omicron's infectivity, vaccine breakthrough, and antibody resistance. Since the spike (S) protein, particularly, its receptor-binding domain (RBD), plays a vital role in viral infection, it has been a key target of vaccines and antibody drugs. Therefore, the study of Omicron's 15 RBD mutations can lead to valuable understanding of Omicron's infectivity, vaccine breakthrough, and antibody resistance.

On the basis of a well-tested and experimentally confirmed deep learning model trained with tens of thousands of experimental data, we investigate the impacts of Omicron's RBD mutations to its infectivity. We show that Omicron is about 10 times more infectious than the original virus or about 2.8 times as infectious as the Delta variant. Using the structures of 185 known antibody–RBD complexes, we reveal that Omicron's vaccine-escape capability is near 14 times as high as that of the Delta variant. We unveil that Omicron may completely abolish the Eli Lilly antibody cocktail. Omicron RBD mutations may also compromise monoclonal antibodies (mAbs) from Regeneron, AstraZeneca, Celltrion, and Rock-

efeller University. However, mAbs from GlaxoSmithKline might not be affected much. Our results call for the development of a new generation of vaccines and mAbs that will not be easily affected by viral mutations.

6. DATA AND SOFTWARE AVAILABILITY

The structural information on 185 antibody–RBD complexes with their corresponding PDB IDs and the results of BFE changes of PPI complexes induced by Omicron mutations can be found in the [Supporting Information, Section S2](#). The analysis of observed SARS-CoV-2 RBD mutations is available at Mutaton Analyzer. The TopNetTree model is available at TopNetmAb. The detailed methods can be found in the [Supporting Information, Sections S3 and S4](#). The validation of our predictions with experimental data can be located in the [Supporting Information, Section S5](#).

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jcim.1c01451>.

BFE changes of antibodies disrupted by Omicron mutations and list of antibodies with corresponding PDB IDs ([ZIP](#))

BFE changes of 185 antibodies induced by mutations R346K, K417T, L452R/Q, E484K/Q, F490S occurred in prevailing variants. Preprocessing and feature generation methods. Machine learning methods. Validations of machine learning predictions with experimental data ([PDF](#))

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Notes

The authors declare no competing financial interest.

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