### LETTER

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# Co-existence of OXA-48 and NDM-1 in colistin resistant *Pseudomonas aeruginosa* ST235

Cansel Vatansever<sup>a</sup>, Sirin Menekse<sup>b</sup>, Ozlem Dogan<sup>a</sup>, Lal Sude Gucer<sup>a</sup>, Berna Ozer<sup>a</sup>, Onder Ergonul<sup>a</sup> and Fusun Can<sup>a</sup>

<sup>a</sup>Department of Infectious Diseases and Clinical Microbiology, Koc University School of Medicine, Istanbul, Turkey; <sup>b</sup>Department of Infectious Diseases and Clinical Microbiology, Kosuyolu State Hospital, Istanbul, Turkey

#### ABSTRACT

Here, we presented 11 cases with colistin-resistant *Pseudomonas aeruginosa* infection and co-existence of OXA-48 and NDM-1 in the ST235 high-risk clone. The molecular analyses were performed by Sanger sequencing and RT–PCR. The eight patients (72.7%) had an invasive infection and three (27.3%) had colonization. The 30-day mortality rate was 87.5% (7/8). Three patients (37.5%, 3/8) received colistin therapy before isolation of *P. aeruginosa*. In the Multilocus sequence typing (MLST) analysis of 11 isolates, eight (72.7%) isolates belonged to *P. aeruginosa* ST235 clone. All isolates were NDM-1 positive, and nine isolates (81.8%) were found to be positive for both OXA-48 and NDM-1. Sequences of *pmrAB* and *phoPQ* revealed numerous insertions and deletions in all isolates. In 10 isolates *pmrAB* and *phoPQ* were found to be upregulated. In conclusion, the co-existence of OXA-48 and NDM-1 genes in colistin-resistant *P. aeruginosa* ST235 high-risk clone indicates the spread of carbapenemases in clinical isolates and highlights need of continuous surveillance for high-risk clones of *P. aeruginosa*.

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KEYWORDS P. aeruginosa; colistin resistance; high-risk clone; ST235; OXA-48; NDM-1

Pseudomonas aeruginosa (P. aeruginosa) is one of the most common causes of healthcare-related infections [1]. The ST235 high-risk clone of P. aeruginosa has high capacity to acquire antibiotic resistance and is disseminating worldwide. The ST235 clone harbours nearly 39 types of beta-lactamases especially IMP, NDM and VIM type Metallo- $\beta$ -lactamases (MBLs) [2]. However, up to date, there is no report on co-existence of NDM-1 and OXA-48 in P. aeruginosa. Colistin resistance among *P. aeruginosa* is rare (<1%) in Europe [3]. However, it was found to be 7.4% in Korea, and 8.8% in Turkey [4,5]. Dissemination of colistin resistance in high-risk clones is concerning because of increased fatality and lack of antimicrobial therapy options. The overexpression of phoPQ and pmrAB two-component regulatory systems contribute colistin resistance by reducing the negative charge of the outer membrane in P. aeruginosa [6].

In this correspondence, we presented 11 cases with colistin-resistant *P. aeruginosa* infection and reported the presence of OXA-48 along with NDM-1 in the isolates belonging to the ST235 high-risk clone. We also analysed mutations and expressions of *phoPQ* and *pmrAB* systems in 11 colistin-resistant *P. aeruginosa* isolates.

Patients who were diagnosed with colistin-resistant *P. aeruginosa* infection or colonization in the ICU

unit of a Cardio-Pulmonary Surgery Hospital in Istanbul between July 2017 and December 2018 were included in the study. The demographic and clinical data were recorded on a standardized case form. The patients were followed up for clinical outcomes.

The colistin resistance was determined by the broth microdilution method according to EUCAST criteria [7]. In strain typing, Multilocus sequence typing (MLST) was performed by amplifying seven housekeeping genes, according to the protocol on Pseudomonas aeruginosa MLST website (https://pubmlst. org/P.aeruginosa/). Allelic profiles and sequence types (STs) were determined using Applied Math Bionumerics V7.6 software. Clonal relatedness was determined by the repetitive PCR (rep-PCR) (Diversilab, Biomerieux). Isolates that had a similarity index >95% were considered as clonally related. Carbapenem and colistin resistance genes (bla<sub>IMP</sub>, bla<sub>VIM</sub>, bla<sub>OXA</sub>, bla<sub>NDM</sub>, bla<sub>KPC</sub>, mcr-1) were screened by PCR using primers as described previously, and the amplicons were confirmed by sequencing (6). For colistin resistance mechanisms, mutations in *pmrAB* and *phoPQ* were detected by Sanger Sequencing [6]. Expressions of pmrA and phoP were studied by qRT-PCR [6]. The *rpsl* was selected for normalization and P. aeruginosa ATCC 27853 was for calibration.

\*🔀 fucan@ku.edu.tr

CONTACT Fusun Can 🖾 cvatansever17@ku.edu.tr

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Among 11 patients, eight (72.7%) had an invasive infection and three (27.3%) had colonization with colistin-resistant P. aeruginosa. Eight (72.7%) patients stayed in ICU and six (54.5%) had lung transplantation. Sepsis was diagnosed in three (37.5%) of eight patients. The 30-day mortality rate was 88% (7/8). The overall mortality of P. aeruginosa infections in the hospital was 46% and it was 68% for carbapenem-resistant colistin susceptible P. aeruginosa infections. In a recent report from Spain, the overall 30day mortality of bacteremia cases caused by ST235 was found to be 82%, however, it was 42.2% in other clones [10]. In our study, the eight of 11 (72.7%) isolates belonged to P. aeruginosa ST235 high-risk clone. The three isolates were identified as a novel allele (ST3078) referred by Pasteur Institutes MLST database (https://pubmlst.org/paeruginosa/)(Table 1). In Germany, the mortality rate of P. aeruginosa bacteremia was reported as 26% in 937 ICU units [8]. The increased mortality rates are associated with multidrug resistance [9]. The ST235 is associated with MDR or PDR profile and fatal infections [2]. In genotyping K741-K748 and K752-K753 were found to be clonally related (>95%). Other isolates belong to different clone.

All isolates were found to be carbapenem-resistant. All of them were NDM-1 positive, and nine (81.8%) harbour both OXA-48 and NDM-1 beta-lactamases. Carbapenem-resistant OXA-48 positive *P. aeruginosa* was isolated in Sudan and India [11] and NDM-1 positive *P. aeruginosa* was detected in Serbia [12]. However, this is the first report of co-existence of OXA-48 and NDM-1 producing *P. aeruginosa* isolation in Turkey and Europe.

Colistin use is one of the major factors responsible for the development of colistin resistance. In our study, only four patients (36,3%) received colistin therapy before the isolation of P. aeruginosa (Table 1). The MICs for colistin were between 4 and >64 mg/L. In all isolates, sequences of pmrAB and phoPQ revealed numerous insertions and deletions. In the ten of them, *pmrAB* and *phoPQ* were found to be upregulated. Relative expressions of pmrA and phoQ genes were between 0.3-59.9-fold (mean 12.9fold) and 0.9-6.9-fold (mean 4.15-fold), respectively. In nine isolates, colistin MICs and pmrAB-phoPQ expressions were found to be related (Figure 1). These results suggested that there could be additional mechanisms contributing to colistin resistance in P. aeruginosa.

In conclusion, colistin resistance is emerging in *P. aeruginosa* ST235 global high-risk clone. The coexistence of OXA-48 and NDM-1 genes in colistinresistant *P. aeruginosa* ST235 high-risk clone indicates the spread of carbapenemases in clinical isolates and highlights need of continuous surveillance for highrisk clones of *P. aeruginosa*.

Table	1. Clini	ical and lai	boratory ch	haracteristics o	Table 1. Clinical and laboratory characteristics of study population.	on.							
			Colistin	Meropenem			Pre-exposure time to		Duration of Active Colistin		30-day	Clonal	
Code	ST	Source	MIC	MIC	Carbapenemase	Empirical Therapy	colistin	Active Therapy	Therapy	Survival	Mortality	Relatedness	
K704	235	235 Catheter	>64	16	OXA-48, NDM-1 Meropenem	Meropenem	0	0	0	EX	1		
K741	235	BAL	16	16	vancomycı OXA-48, NDM-1 Moxifloxacin	vancomycın Moxifloxacin	0	Meropenem Colistin Ertanenem	1	Ĕ	2	*	
K740	3078	BAL	>64	8	OXA-48, NDM-1	Ы	0	Meropenem Colistin	27	Ē	30		
K748	235	DTA	64	16	0XA-48, NDM-1	l azobactam 0	0	Ertapenem 0	0	Discharge	Discharge	*	
K752	3078	DTA	4	16	OXA-48, NDM-1	0	0	0	0	Discharge	Discharge	**	
K753	3078	BAL	4	8	OXA-48, NDM-1	Piperacillin	0	Meropenem Colistin	14	Discharge	Discharge	**	
						Tazobactam							
K783	235	BAL	16	16	OXA-48, NDM-1	Meropenem Colistin	10	Meropenem Colistin	10	Ĕ	7		
K970	235	Catheter	32	16	NDM-1	Levofloxacin Colistin	21 days	0	0	Ĕ	10		
K982	235	BAL	16	16	OXA-48, NDM-1	Colistin (inhalation)	25 days	0	0	Ĕ	1		
K989	235	Catheter	>64	16	NDM-1	Meropenem	0	Meropenem Colistin	38	Ĕ	42		
K1009	235	DTA	16	16	OXA-48, NDM-1	Colistin (inhalation)	8 days	0	0	EX	19		
Notes: BAL: Broncho *clone 1, **clone 2.	AL: Bror , **cloné	achoalveolar a 2.	lavage; DTA.	Notes: BAL: Bronchoalveolar lavage; DTA: Deep tracheal aspirate. *clone 1, **clone 2.	aspirate.								$\cup$

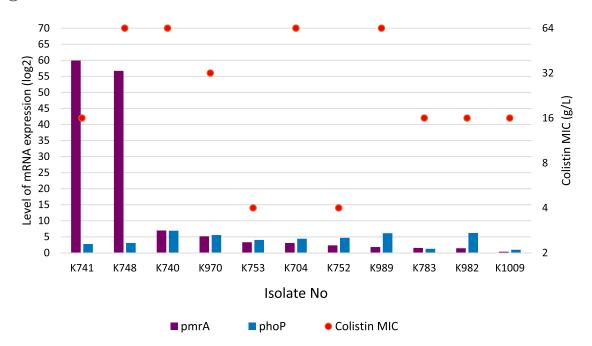


Figure 1. PmrA and PhoP expressions of the colistin-resistant P. aeruginosa in correlation with colistin MIC values.

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## **Disclosure statement**

No potential conflict of interest was reported by the authors.

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