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Genetic diversity of the Central European wild boar (*Sus scrofa scrofa*) population and domestic pig (*Sus scrofa domesticus*) breeds based on a microsatellite DNA locus

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Abstract. The results of studies of the genetic structure of the Central European wild boar (*Sus scrofa scrofa*) population and four breeds of domestic pigs (Duroc, Yorkshire, Large White and Landrace) bred in the Central Black Earth region of Russia are presented in this work. Based on 12 microsatellite loci, a significant ($p < 0.05$) decrease in the level of genetic variability in bred breeds was shown. The expected heterozygosity and Shannon index were as follows: in the wild boar, $H_o = 0.763 \pm 0.026$, $I = 1.717 \pm 0.091$; in the Duroc breed, $H_o = 0.569 \pm 0.068$, $I = 1.191 \pm 0.157$; in the Landrace, $H_o = 0.618 \pm 0.062$, $I = 1.201 \pm 0.147$; in the Large White, $H_o = 0.680 \pm 0.029$, $I = 1.362 \pm 0.074$; and in the Yorkshire, $H_o = 0.642 \pm 0.065$, $I = 1.287 \pm 0.156$. The results of checking genotypic Hardy–Weinberg equilibrium based on the G-test of maximum likelihood demonstrated that the overwhelming majority of loci in the wild boar population were in the state of said equilibrium. By contrast, in pig breed populations, some loci demonstrated a significant deviation from the indicated equilibrium. In addition, the Yorkshire, Large White, and Landrace populations had loci, for which the hypothesis of neutrality was reliably rejected based on the results of the Ewens–Watterson test. The revealed private alleles, characteristic of the wild boar and breeds, can later be used to identify them. The ordination of the centroids of different herds in the space of the first two principal coordinates based on the matrix of pairwise estimates of Nei's genetic distances showed that the most distant populations are the Duroc and Boar breeds, and the most genetically close are the Yorkshire and Landrace breeds. The closest to the wild boar population was the Large White breed. The assessment of the effective size, carried out using the method based on the linkage disequilibrium and the molecular coancestry method, showed that in all studied groups, including the wild boar population, the effective size was less than 100 individuals. The low effective size of the wild boar population ($N_e = 21.8$, $N_{eb} = 4.0$) is probably caused by the death and shooting of animals due to *Pestis africana suum*.

Key words: wild boar; pig breeds; microsatellite loci; genetic structure; effective population size.

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Генетическое разнообразие популяции центральноевропейского кабана (*Sus scrofa scrofa*) и пород домашних свиней (*Sus scrofa domesticus*) на основе микросателлитных локусов ДНК

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Аннотация. В работе приведены результаты исследований генетической структуры популяции центральноевропейского кабана (*Sus scrofa scrofa*) и четырех пород домашних свиней (дюрок, йоркшир, крупная белая и ландрас), разводимых в Центрально-Черноземном регионе России. На основе 12 микросателлитных локусов установлено достоверное ($p < 0.05$) снижение генетической изменчивости в разводимых породах. Ожидаемая гетерозиготность и индекс Шеннона были равными: у кабана – $H_o = 0.763 \pm 0.026$, $I = 1.717 \pm 0.091$;

у пород дюрок – $H_o = 0.569 \pm 0.068$, $I = 1.191 \pm 0.157$; ландрас – $H_o = 0.618 \pm 0.062$, $I = 1.201 \pm 0.147$; крупная белая – $H_o = 0.680 \pm 0.029$, $I = 1.362 \pm 0.074$; йоркшир – $H_o = 0.642 \pm 0.065$, $I = 1.287 \pm 0.156$. Результаты проверки генотипического равновесия Харди–Вайнберга на основе G-теста максимального правдоподобия показали, что в популяции кабана большинство локусов находилось в состоянии генотипического равновесия Харди–Вайнберга. Напротив, в популяциях различных пород свиней часть локусов демонстрирует достоверное отклонение от отмеченного равновесия. Кроме того, в популяциях йоркшир, крупная белая и ландрас присутствовали локусы, для которых достоверно отвергалась гипотеза о нейтральности на основании результатов теста Эвенса–Ваттерсона (Ewens–Watterson test). Обнаруженные приватные аллели, характерные для кабана и различных пород, в дальнейшем могут быть использованы для их идентификации. Ординация центроидов разных стад в пространстве первых двух главных координат на основании матрицы попарных оценок генетических дистанций M. Nei показала, что наиболее удаленные популяции – породы дюрок и кабан, а самые генетически близкие – йоркшир и ландрас. Ближе всех к популяции кабана была порода крупная белая. Оценка эффективной численности, проведенная с использованием метода, основанного на неравновесии по сцеплению (linkage disequilibrium) и МС-метода (the molecular coancestry method), продемонстрировала, что во всех изученных группах, включая и популяцию кабана, эффективный размер оказался меньше 100 особей. Низкое значение эффективного размера популяции кабана ($N_e = 21.8$, $N_{eb} = 4.0$), вероятно, является следствием падежа и отстрела животных из-за африканской чумы свиней (*Pestis africana suum*). Ключевые слова: кабан; породы свиней; микросателлитные локусы; генетическая структура; эффективная численность популяции.

Introduction

According to various estimates, the domestication of the wild boar began 7–9 thousand years ago. During this time, more than 730 breeds of these animals have been created by man. It is obvious that for such a long “cultural” evolution, various breeds, being so-called clean lines, have largely lost the natural genetic potential that provides homeostatic mechanisms. As a result, maintaining the stability of existing breeds, like any artificially created systems, requires significant financial investments. In this regard, the study of the genetic potential of natural populations of wild boars for a possible increase in the resistance of pig breeds (for example, by methods of genome editing, etc.) is highly relevant.

In this regard, the study of wild boar genetics is now receiving much attention, both in Russia and other countries (Gladyr et al., 2009; Zinovieva et al., 2013; Rębała et al., 2016; Mihalik et al., 2020). In addition, due to constant outbreaks of the African swine fever (*Pestis africana suum*) in Russia, the wild boars are regularly shot as potential carriers of this disease. At the same time, the population structure of this animal is not taken into account, which can cause depletion of the gene pool and, against the background of increasing anthropogenic pressure, lead to the extinction of some groups. Examples of a significant reduction in the population of commercial species occurring due to the data on the state of the gene pool being neglected are well known (Altukhov, 2003).

Also, in the practice of molecular genetic laboratories, for forensic purposes, it is often necessary to diagnose tissue samples from illegally caught wild boars and prove their belonging to a wild species, and not to domesticated forms of pigs (Kipen et al., 2016; Lorenzini et al., 2020), or identify wild boar tissue in food (Szemethy et al., 2021). In this regard, the identification of private alleles for natural populations is also an urgent problem.

Microsatellite DNA loci (STR markers), which are tandem repeats of the noncoding part of nuclear DNA, are very convenient markers for studying genetic processes in populations. There are many works on the assessment of population gene pools of both domestic pigs and wild boars in different regions (Vernesi et al., 2003; Ferreira et al., 2009; Nikolov et al., 2009;

da Silva et al., 2011; Choi et al., 2014; Sahoo et al., 2016; Ryabtseva et al., 2018; Han et al., 2021; Snegin et al., 2021).

The purpose of this work is to assess the genetic diversity of microsatellite loci in the population of the Central European wild boar (*Sus scrofa scrofa*) and the most common pig breeds (*Sus scrofa domesticus*) bred in the Central Black Earth region of Russia. It should be noted that such studies have not previously been conducted in this region.

Materials and methods

A total of 320 animals were involved in the study. A sample of 30 wild boars was taken from the populations of the Oryol region (districts: Korsakovsky, Zalogoshchensky, Novosilsky, Pokrovsky, Shablykinsky), Russia. The wild boars were caught during the hunting season of 2018. For comparison, samples from four populations of different breeds of domestic pigs bred on the farms of the Central Black Earth Region of Russia were used: Durok – 67 individuals (Belgorod region), Yorkshire – 108 individuals (Kursk region), Landrace – 50 individuals (Belgorod region), large white – 65 individuals (Voronezh region). All analyzed animals are Canadian breeds.

All 12 microsatellite loci recommended by ISAG-FAO (International Society for Animal Genetics, Food and Agriculture Organization) (FAO SoW-AnGR... 2006) and arranged in one multiplex panel (*S0101*, *S0155*, *S0228*, *S0355*, *S0386*, *SW24*, *SW240*, *SW72*, *SW857*, *SW911*, *SW936*, *SW951*) were used as DNA markers (Table 1). Primers for PCR were selected taking into account the amplification of all 12 loci in one tube. The size of all amplified PCR products, taking into account all known alleles, was < 300 base pairs.

In the domestic pigs, DNA extraction was carried out from ear pits, and in the wild boars – from muscle tissue samples. For this purpose, we used kits with proteinase K DNA-Extran-2 (Syntol, Russia). The PCR reaction was carried out on a Verity amplifier (Applied Biosystems, USA) in 20 µL of a mixture containing 20 ng of genomic DNA, PCR buffer (10 mmol Tris-HCl (pH 8.3), 50 mmol KCl, 2 mmol MgCl₂), 0.25 mmol dNTP, 0.5 µmol primer, 1 unit *Taq* DNA polymerase (inhibited for hot start).

Table 1. The characteristics of the microsatellite loci recommended by ISAG for determining the reliability of the origin pigs

Locus	Allele length, bp	Fluorescent Dye	Primers (5'-----3')
<i>S0101</i>	193–221	R6G	F: GAATGCAAAGAGTTCAGTGTAGG R: GTCTCCCTCACACTTACCGCAG
<i>S0155</i>	142–166	TAMRA	F: TGTTCTCTGTTTCTCTCTGTTTG R: AAAGTGAAAGAGTCAATGGCTAT
<i>S0228</i>	218–270	TAMRA	F: GGCATAGGCTGGCAGCAACA R: AGCCACCTCATCTTATCTACT
<i>S0355</i>	223–277	FAM	F: TCTGGCTCTACTCTTCTTGATG R: TTGGGTGGGTGCTGAAAAATAGGA
<i>S0386</i>	164–182	FAM	F: GAACTCCTGGGTCTTATTTTCTA R: GTCAAAAATCTTTTATCTCCAACAGTAT
<i>SW24</i>	95–124	ROX	F: CTTGGGTGGAGTGTGTGC R: ATCCAAATGCTGCAAGCG
<i>SW240</i>	92–124	R6G	F: AGAAATTAGTGCCTCAAATTGG R: AAACCATTAAGTCCCTAGCAA
<i>SW72</i>	97–125	TAMRA	F: ATCAGAACAGTGCGCCGT R: TTTGAAAATGGGGTGTTC
<i>SW857</i>	137–161	R6G	F: TGAGAGGTCAAGTACAGAAGACC R: GATCCTCCTCCAATCCAT
<i>SW911</i>	149–177	ROX	F: CTCAGTTCTTTGGGACTGAACC R: CATCTGTGAAAAAAGCC
<i>SW936</i>	81–117	FAM	F: TCTGGAGCTAGCATAAGTGCC R: GTGCAAGTACACATGCAGGG
<i>SW951</i>	124–134	FAM	F: TTTCACTCTGGCACCAG R: GATCGTGCCCAATGGAC

PCR parameters: 94 °C – 3 min; (98 °C – 30 sec, 59 °C – 120 sec, 72 °C – 90 sec) – 4 cycles; (94 °C – 30 sec, 59 °C – 120 sec, 72 °C – 90 sec) – 6 cycles; (90 °C – 30 sec, 59 °C – 120 sec, 72 °C – 75 sec) – 20 cycles; 68 °C – 30 min. The heating rate from 59 to 72 °C was no more than 0.3 °C/1 sec.

Fragment analysis of the PCR products was carried out on an ABI PRISM 3500 automatic capillary DNA sequencer (Applied Biosystems, USA) using 50 cm capillaries and a POP-7™ polymer matrix. Fragment size analysis was performed using GeneMapper R Software v. 4.1 (Applied Biosystems).

For statistical processing of the data obtained we used the GenAIEx v. 6.5 (Peakall, Smouse, 2012) and PorGene 1.32 (Yeh et al., 1999).

Results

In Table 2, the allele frequencies of the microsatellite loci used for analysis are presented. The data provide insight into the distribution of various alleles among populations of domestic pigs and wild boars.

The presence of private alleles in different populations is shown in Table 3. The results show a high content of unique alleles in the wild boar population (16 alleles). The highest frequency of private alleles was found at the loci *SW24* and *SW72* (0.25 in each). At the same time, the allele 97 at the *SW24* locus, and the allele 99 at the *SW72* locus are found

more often than others among private alleles. The pig of the Duroc breed is slightly inferior to the wild boar in the number of private alleles, while the 105 and 111 alleles at the *SW936* locus (0.246 and 0.276, respectively) had the highest frequency of occurrence. The Large White breed is almost three times inferior in the number of private alleles to the population of wild boar and Duroc pigs. However, some of the original alleles are found in this group of animals with noticeable frequency. For example, the allele 141, unique for this breed, at the *SW857* locus was observed in half of the animals analyzed (frequency 0.5). No private alleles were found in the Landrace population, and only one private allele was noted in the Yorkshire population.

The wild boar population has significantly high values of the indicators of genetic variability in comparison with the pig breeds. The comparison was carried out using the Pearson χ^2 test ($p < 0.05$) (Table 4).

The level of inbreeding (F) in the studied groups turned out to be low, and the populations of Yorkshire, Landrace and wild boar received a negative value due to the prevalence of observed heterozygosity over the theoretically expected heterozygosity (see Table 4).

The results of checking the genotypic balance of Hardy–Weinberg based on the G-test of maximum likelihood demonstrated that in the wild boar population the overwhelm-

Table 2. Frequencies of the microsatellite loci alleles in the populations of various pig breeds and wild boars

Locus	Population					Locus	Population					Locus	Population				
<i>S0101</i>	1	2	3	4	5	<i>S0386</i>	1	2	3	4	5	<i>SW72</i>	1	2	3	4	5
193	0	0	0.169	0	0	164	0.022	0	0.008	0	0.017	119	0.500	0.361	0	0.250	0.017
195	0	0.055	0	0.090	0.017	166	0.045	0	0.054	0.140	0.133	121	0.097	0	0	0	0.050
197	0	0	0	0.020	0.150	168	0	0.231	0.015	0.060	0.150	123	0.052	0	0	2	0
199	0	0.097	0	0.030	0.017	170	0.007	0	0.131	0.090	10	<i>SW857</i>	1	2	3	4	5
201	0	0.028	0	0	0.083	172	0.060	0.014	0.023	0	0.117	137	0.007	0	0	0	0
203	0	0	0	0	0.050	174	0.112	0.14	0	0.010	0.017	141	0	0	0.500	0	0
205	0.007	0	0.015	0	0	176	0.015	0	0.215	0	0.033	143	0	0	0.008	0	0
207	0	0.319	0.008	0.310	0.317	178	0.022	0.630	0.346	0.630	10	147	0.007	0	0	0	0
209	0	0.241	0.061	0.280	0.200	180	0.649	0.083	0.208	0.040	0.017	149	0	0	0.231	0	0.317
211	0.851	0.014	0.746	0.210	0.100	182	0.067	0.028	0	0.030	11	151	0	0.352	0.100	0.370	0.150
213	0.142	0.176	0	0.060	0	<i>SW24</i>	1	2	3	4	5	153	0.052	0.194	0.077	0	0.333
215	0	0.069	0	0	2	95	0	0	0	0	0.050	155	0.627	0.014	0	0	0.050
217	0	0	0	0	2	97	0	0	0	0	0.183	157	0	0.241	0.054	0.470	0.067
<i>S0155</i>	1	2	3	4	5	99	0	0	0	0	0.017	159	0.298	0.143	0.031	0.160	0.083
144	0.015	0	0	0	0	101	0.037	0	0	0	0.017	161	0.007	0.056	0	0	0
146	0.022	6	0.061	0	0.433	105	0	0	0.046	0.410	0.050	<i>SW911</i>	1	2	3	4	5
148	0.007	0.352	0.008	0.250	0.117	107	0	0.245	0	0	0.050	149	0	0	0.046	0	0.017
150	0	0	0	0	0.017	109	0.455	0.157	0	0	0.050	151	0.007	0	0	0	0
152	0.007	0	0	0	0	111	0	0	0.261	0.010	0.017	153	0.022	3	0	0	0
154	0	0	0	0	0.133	113	0	0.018	0.046	0	0	155	0	0	0.208	0	0
156	0.589	0	0.208	0	0.017	115	0.134	0.111	0.069	0.150	0.067	157	0.187	0.217	0.100	0.350	0
158	0	0.264	0.346	0.420	0.100	117	0	0.060	0.354	0.310	0.267	159	0	0	0.485	0	0.200
160	0.022	0.356	0.223	0.330	0.083	119	0.052	0.380	0.215	0.070	0.067	161	0	0.444	0.054	0.430	0.117
162	0.328	0	0.154	0	0.083	121	0.216	0.028	0.008	0.030	0.133	163	0.537	0	0.008	0	0.117
164	0.007	0	0	0	0	123	0.097	0	0	0.020	0.033	165	0	0	0.015	0	0.333
166	0	0	0	0	0.017	125	0.007	0	0	0	0	167	0	0.264	0.085	0.130	0.017
<i>S0228</i>	1	2	3	4	5	<i>SW240</i>	1	2	3	4	5	<i>SW911</i>	1	2	3	4	5
220	0.015	0.236	0.008	0.240	0	93	0	0.305	0	0	0	169	0.119	0.060	0	0.090	11
222	0.134	0.278	0	0.060	0.017	97	0.007	0	0.231	0	0.017	171	0.112	0	0	0	0.017
224	0.157	0.018	0	0.040	0	99	0	0	0.008	0	0.367	177	0.015	0	0	0	0
226	0	0	0.061	0.030	0	101	0.060	0	0.185	0	0.033	<i>SW936</i>	1	2	3	4	5
228	0	0.056	0.023	0.050	0.433	103	0.082	0	0	0	0.017	85	0	0	0	0.500	0.033
230	0.037	0	0	0.020	0.017	105	0.254	0	0.200	0	0	87	0.015	0.500	0	0	0
232	0.022	0.014	0	0.090	0.017	107	0.164	0	0	0.030	0	91	0	0	0	0.500	0.383
234	0.022	0.167	0	0.330	0	109	0.246	0.046	0	0.120	0	93	0	0.500	0.346	0	0.150
236	0.007	0.060	0	0.040	0.267	111	0	0.028	0	0.030	0	95	0	0	0.254	0	0
238	0	0.028	0.061	0.010	0.050	113	0	0.018	0.315	0	0.233	97	0	0	0	0	0.150
240	0.007	0.055	0.015	0.010	0.033	115	0	0.055	0	0	0.133	99	0	0	0	0	0.050
242	0	0.028	0.015	0.020	0.083	117	0.030	0.088	0.054	0.430	0	101	0	0	0	0	0.017
244	0	0	0.515	0.020	0.050	119	0.127	0.296	0	0	0	103	0.007	0	0	0	0.033
246	0.366	0.032	0.031	0	0	121	0.015	0.162	0.008	0.390	0.033	105	0.246	0	0	0	0
248	0.231	0.028	0.269	0.040	0.033	123	0.015	0	0	0	0.167	107	0.022	0	0.231	0	0.183
<i>S0355</i>	1	2	3	4	5	<i>SW72</i>	1	2	3	4	5	109	0.045	0	5	0	0
243	0	0	0.415	0	0.667	99	0	0	0	0	0.233	111	0.276	0	0	0	0
245	0	0.125	0	0.110	0.083	101	0.015	0	0	0	0.033	113	0.246	0	0.061	0	0
247	1.0	0	0.300	0	0	103	0	0	0	0	0.017	115	0.097	0	0.069	0	0
249	0	0.088	0	0.190	0	105	0	0	0.162	0.420	0.133	117	0.045	0	0	0	0
255	0	0	0	0	0.033	107	0	0.245	0.115	0	0.017	<i>SW951</i>	1	2	3	4	5
259	0	0.301	0	0.210	0.100	109	0.298	0.171	0	0	0.050	126	0.134	0	0.154	0	0.383
261	0	0	0	0	0.050	111	0.007	0	0	0	0	128	0.045	1.0	0.246	1.0	0.200
269	0	0	0	0	0.067	113	0.015	0	0.531	0	0.133	130	0.619	0	0.054	0	0.067
271	0	0	0.285	0.160	0	115	0	0.181	0.192	0.220	0.150	132	0.134	0	0.546	0	0.350
273	0	0.486	0	0.330	0	117	0.015	0.042	0	0.090	0.167	134	0.067	0	0	0	0

Note. Population: 1 – Duroc; 2 – Yorkshire; 3 – Large White; 4 – Landrace; 5 – Wild Boar.

Table 3. Private alleles in the studied populations of various breeds of pigs and wild boars

Population	Locus									
	S0101	S0155	S0355	SW24	SW240	SW72	SW857	SW911	SW936	SW951
Duroc		144, 152, 164		125		111	137, 147	151, 177	105, 111, 117	134
Yorkshire					93					
Large White	193						141, 143	155	95	
Wild Boar	203, 217	150, 154, 166	255, 261, 269	95, 97, 99		99, 103			97, 99, 101	

Note. No private alleles were found in the Landrace pig population.

Table 4. The indicators of the genetic diversity in the studied groups of pigs and wild boars (Mean ± SE)

Population	N	P, %	Aa	Ae	I	Ho	He	F	N _{pa}
Duroc	67	91.7	6.917±0.802	2.913±0.396	1.191±0.157	0.525±0.079	0.569±0.068	0.076±0.076	1.083±0.336
Yorkshire	108	91.7	5.667±0.847	3.452±0.384	1.287±0.156	0.716±0.086	0.642±0.065	-0.128±0.104	0.083±0.083
Large White	65	100.0	6.167±0.534	3.350±0.241	1.362±0.074	0.660±0.060	0.680±0.029	0.022±0.075	0.417±0.193
Landrace	50	91.7	5.250±0.978	3.124±0.336	1.201±0.147	0.713±0.081	0.618±0.062	-0.175±0.101	0.000
Wild Boar	30	100.0	8.583±0.712	4.702±0.444	1.717±0.091	0.844±0.038	0.763±0.026	-0.106±0.033	1.333±0.414

Note. N – the number of individuals; P – the percentage of polymorphic loci; Aa – the average number of alleles; Ae – the effective number of alleles; I – Shannon's index; Ho – observed heterozygosity; He – expected heterozygosity; F – the coefficient of inbreeding; N_{pa} – the average number of private alleles per locus; Mean ± SE – mean ± standard error.

Table 5. The results of checking the genotypic balance of Hardy–Weinberg for 12 loci of MS-DNA in herds of pigs of different breeds and wild boar based on the G-test of maximum likelihood (likelihood ratio G-test)

Locus	Population				
	Duroc	Yorkshire	Large White	Landrace	Wild Boar
S0101	ns	< 0.001	ns	ns	ns
S0155	ns	ns	ns	0.019	ns
S0228	0.014	< 0.001	ns	ns	ns
S0355	mono	ns	ns	< 0.001	ns
S0386	< 0.001	< 0.001	< 0.001	< 0.001	ns
SW24	ns	< 0.001	< 0.001	0.004	ns
SW240	ns	< 0.001	< 0.001	< 0.001	ns
SW72	ns	< 0.001	ns	0.001	ns
SW857	ns	< 0.001	ns	< 0.001	ns
SW911	ns	< 0.001	ns	0.032	ns
SW936	ns	< 0.001	0.014	< 0.001	0.002
SW951	< 0.001	mono	0.017	mono	< 0.001

Note. ns – correspondence to Hardy–Weinberg equilibrium; p > 0.05; mono – monomorphic locus.

ing majority of the loci were in the state of the genotypic equilibrium of Hardy–Weinberg (Table 5). On the contrary, in the populations of the different breeds of pigs, some of the loci demonstrated a significant deviation from the indicated equilibrium. In addition, the monomorphic loci were noted in three groups (Duroc, Yorkshire, and Landrace). Also, in the populations of Yorkshire, Large White, and Landrace, the

MC-loci were present, for which the hypothesis of neutrality was reliably rejected based on the results of the Ewens–Watterson test (Ewens, 1972; Watterson, 1978) (Table 6). Most of these loci were found in the Yorkshire and Landrace breeds.

The degree of similarity of the genetic structure of the pigs was assessed using the principal component analysis, which resulted in the ordination of the centroids of different herds

Table 6. The results of the Ewens-Watterson test for 12 MS-DNA loci for various breeds of pigs and boars (only the loci for which the hypothesis of neutrality is reliably rejected are shown)

Population	Locus	Obs. F	L95–U95
Yorkshire	S0155	0.321	0.332–0.906
	SW72	0.254	0.282–0.893
	SW936	0.500	0.502–0.991
Large White	S0355	0.344	0.367–0.970
Landrace	S0155	0.348	0.371–0.961
	S0355	0.227	0.267–0.831
	SW936	0.500	0.503–0.980

Note. Obs. F – the actual sum of squares of allele frequencies; L95, U95 – lower and upper values of the 95 % confidence interval of the Obs F estimate, calculated from 1000 simulations.

Table 7. The values of genetic distances (according to M. Nei) between the studied groups of pigs and wild boars

Population	Duroc	Yorkshire	Large White	Landrace	Wild Boar
Duroc	0.000				
Yorkshire	1.831	0.000			
Large White	0.988	1.209	0.000		
Landrace	1.976	0.318	1.087	0.000	
Wild Boar	1.803	1.091	0.707	0.968	0.000

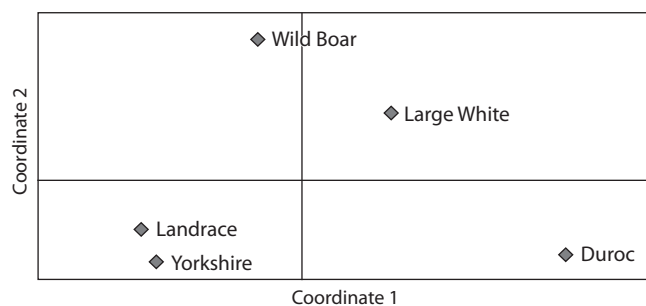
Table 8. The estimates of the effective population size (N_e , N_{eb}), calculated using the LD- and MC-method between 12 loci of MS-DNA

Population	Linkage disequilibrium		Molecular coancestry	
	N_e	95 % CI	N_{eb}	95 % CI
Duroc	86.1	54.9–164.2	19.4	2.3–54.0
Yorkshire	7.4	5.9–8.9	7.6	4.3–11.7
Large White	44.6	34.0–60.9	15.5	6.7–28.0
Landrace	9.0	6.6–11.8	2.1	1.3–3.5
Wild Boar	21.8	17.1–28.9	4.0	2.7–5.6

Note. 95 % CI – limits of 95 % confidence interval.

in the space of the first two Main Coordinates based on the matrix of pairwise estimates of genetic distances by M. Nei (Table 7, see the Figure). According to the data obtained, the most distant populations are the Duroc and Boar breeds, and the most genetically close are the Yorkshire and Landrace breeds. The closest to the wild boar population was the Large White breed.

The effective population size was estimated using the linkage disequilibrium (LD) method (Hill 1981; Waples, 2006; Waples, Do, 2010), as well as the molecular coancestry method (Nomura, 2008). The calculations were performed using the NeEstimator V2 software (Do et al., 2014). The results are shown in Table 8.



The ordination of centroids based on a matrix of pairwise estimates of M. Nei's genetic distances based on the distribution of allele frequencies of 12 MS-DNA loci.

Table 9. Level of actual heterozygosity for microsatellite markers in different wild boar populations

Country	<i>H_o</i>	References	Country	<i>H_o</i>	References
Portugal	0.627	Ferreira et al., 2009	Japan	0.473	Choi et al., 2014
South Korea	0.682	Han et al., 2021	Indonesia	0.658	
	0.422–0.673	Choi et al., 2014	Russia		
Bulgaria	0.620	Nikolov et al., 2009	Primorsky region	0.710	Choi et al., 2014
Germany	0.460		Kirov region	0.463	Gladyr et al., 2009
Italy	0.520-0.720	Vernesi et al., 2003	Yaroslavl region	0.535	
Hungary	0.750		Orenburg region	0.722	
China	0.845	Choi et al., 2014	Krasnodar region	0.614	
Vietnam	0.859		Khabarovsk region	0.670	

Discussion

The reliably high values of genetic variability ($p < 0.05$), noted in the wild boar population, are quite an expected phenomenon. This is despite the fact that the sample from the analyzed group was smaller than the samples from herds of domestic pigs. This clearly demonstrates the consequences of panmixis and genetic-automatic processes, which, in combination with the natural selection, form the gene pools of natural populations. Moreover, in the wild boar population we analyzed, the level of actual heterozygosity turned out to be significantly higher than in the populations of this animal both in Europe and Asia. At the same time, the Oryol group, in terms of the level of genetic diversity, turned out to be similar to the Chinese and Vietnamese wild boar populations (Table 9).

On the contrary, the transition of a number of loci to the monomorphic state and the lack of equilibrium according to Hardy–Weinberg, noted by us for most loci in domestic pig breeds, is a consequence of long-term artificial selection work, as a result of which many alleles of the “wild” type were lost, which led to the loss of genetic variety. This is also evidenced by the significant deviation from neutrality of some microsatellite loci in the Landrace, Yorkshire, and Large White breeds, which was revealed by us using the Evens–Watterson test (see Table 6).

Allelic diversity results in a large number of private alleles observed in the wild boar population. At the same time, the Duroc population, despite the large number of private alleles, turned out to be the most monomorphic among the studied groups. This probably indicates a long-term selection of this red color breed in the conditions of the North American continent, isolated from crossing with other pig breeds, including white breeds of European origin (Large White, Yorkshire and Landrace). This can explain the significant genetic distance of the Duroc breed, both from the wild boar and the European pig breeds. The presence of original alleles in the studied populations may be used in the future for identifying both the breed of pigs and belonging to a wild boar population.

It should be noted that the results obtained are in part consistent with the data obtained in the work of E.A. Glydyr et al. (2009). In this study, the level of actual heterozygosity in three

out of five wild boar populations was found to be higher than in domestic pigs. However, the average number of the effective alleles was the same ($A_e = 2.6$). In terms of the number of private alleles, the wild boar population also surpassed the domestic pigs (21 versus 10, respectively).

The calculation of the effective population size based on the LD method showed that in almost all the studied groups, including the wild boar population, the effective size was less than 50 individuals. The only exception was the Duroc breed ($N_e = 86.1$). The data indicate the observed linkage disequilibrium, probably caused by closely related mating in the analyzed groups of domestic pigs. This genetic drift has generated a non-random association between alleles at different loci.

Calculations carried out using the MC method showed even lower values. This phenomenon is most likely associated with a small number of parental individuals (primarily males) who founded the studied populations. It is worth noting that a similar picture was obtained in the work on assessing the effective population size of the endangered *Gochu Asturcelta* pig breed (Menendez et al., 2016).

If the results for breeds of domestic pigs were comparable with data obtained in other studies, where N_e varied in general from 20 to 92 individuals (Šveistienė, Razmaitė, 2013; Krupa et al., 2015; Zanella et al., 2016; Lugovoy et al., 2018), then in relation to the wild boar population, the result was somewhat unexpected, since in previous studies N_e ranged from 180 to 1477 animals in natural populations (Cowled et al., 2008; Herrero-Medrano et al., 2013).

Such a low N_e value noted in the wild boar population of the Oryol region can be partly explained by a small sample, however, reasons behind it may be more serious. In particular, it is known that outbreaks of African swine fever (*Pestis africana suum*) (<https://www.kommersant.ru/doc/4236233>), resulting in the death of wild boars are often recorded on the territory of the Central Black Earth Region, which includes the specified area. In addition, in order to prevent the spread of infection, hunting farms are forced to shoot a significant part of the animals. It is likely that these phenomena affect the effective size of the wild boar populations.

Conclusion

Thus, on the basis of the studies carried out, a clear reduction in the genetic diversity of the domestic pig breeds in comparison with the natural wild boar population was demonstrated. The presence of private alleles can further aid in the identification of wild boar and different breeds of pigs. Low values of the effective size of the studied groups require attention from breeders in relation to the breeds of pigs. In particular, the pig breeding companies in the region under study need to use a larger number of producers (primarily males) to obtain replacement livestock. With regard to wild boar populations, prophylactic shooting and harvesting should be carried out under the control of environmental authorities with mandatory monitoring of the state of the population gene pools.

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