

Transgelin is a TGF β -inducible gene that regulates osteoblastic and adipogenic differentiation of human skeletal stem cells through actin cytoskeleton organization

M Elsafadi^{*,1,2}, M Manikandan¹, RA Dawud^{3,4}, NM Alajez¹, R Hamam¹, M Alfayez¹, M Kassem^{1,2}, A Aldahmash^{1,2,5} and A Mahmood^{*,1}

Regenerative medicine is a novel approach for treating conditions in which enhanced bone regeneration is required. We identified transgelin (*TAGLN*), a transforming growth factor beta (TGF β)-inducible gene, as an upregulated gene during *in vitro* osteoblastic and adipocytic differentiation of human bone marrow-derived stromal (skeletal) stem cells (hMSC). siRNA-mediated gene silencing of *TAGLN* impaired lineage differentiation into osteoblasts and adipocytes but enhanced cell proliferation. Additional functional studies revealed that *TAGLN* deficiency impaired hMSC cell motility and *in vitro* transwell cell migration. On the other hand, *TAGLN* overexpression reduced hMSC cell proliferation, but enhanced cell migration, osteoblastic and adipocytic differentiation, and *in vivo* bone formation. In addition, deficiency or overexpression of *TAGLN* in hMSC was associated with significant changes in cellular and nuclear morphology and cytoplasmic organelle composition as demonstrated by high content imaging and transmission electron microscopy that revealed pronounced alterations in the distribution of the actin filament and changes in cytoskeletal organization. Molecular signature of *TAGLN*-deficient hMSC showed that several genes and genetic pathways associated with cell differentiation, including regulation of actin cytoskeleton and focal adhesion pathways, were downregulated. Our data demonstrate that *TAGLN* has a role in generating committed progenitor cells from undifferentiated hMSC by regulating cytoskeleton organization. Targeting *TAGLN* is a plausible approach to enrich for committed hMSC cells needed for regenerative medicine application.

Cell Death and Disease (2016) 7, e2321; doi:10.1038/cddis.2016.196; published online 4 August 2016

Regenerative medicine through employing stem cell transplantation is a novel approach for treating conditions in which enhanced bone regeneration is required. A number of stem cell types have been envisaged as candidates for use in therapy. Human bone marrow-derived stromal (also known as skeletal or mesenchymal) stem cells (hMSCs) is one of the most promising candidates. Optimal use of hMSC in therapy requires detailed understanding of molecular mechanisms of lineage commitment and differentiation as well as identifying regulatory factors that can be targeted for controlling hMSC differentiation and functions.

Global hypothesis generating methods, for example, DNA microarrays, proteomic analysis, and miRNA microarrays have been employed by our group in order to identify factors

relevant to hMSC biology and functions and that exhibit significant changes during lineage-specific differentiation.^{1–5} This approach has led to the identification of several factors that control osteoblast or adipocyte differentiation of hMSC.³ Using transcriptomic profiling of differentiating hMSC, we identified transgelin (*TAGLN*) as a highly upregulated gene in hMSC during osteoblast and adipocyte differentiation.

TAGLN is a transforming growth factor beta (TGF β)-inducible gene⁶ that functions as an actin-crosslinking/gelling protein of the calponin family. It is localized to the cytoskeleton and is expressed by endothelial, smooth muscle cells, fibroblasts as well as several immune cells.⁷ It has several names: mouse p27, *WS3-10*, and *SM22*.⁸ It is transiently expressed in heart and skeletal muscle cells during early

¹Stem Cells Unit, Department of Anatomy, College of Medicine, King Saud University, Riyadh 11461, Kingdom of Saudi Arabia; ²KMEB, Department of Endocrinology, University Hospital of Odense and University of Southern Denmark, Odense, Denmark; ³Berlin-Brandenburg Center for Regenerative Therapies, Charité-Universitätsmedizin Berlin, Berlin, Germany; ⁴Department of Comparative Medicine, King Faisal Specialist Hospital and Research Centre, Riyadh, Kingdom of Saudi Arabia and ⁵Prince Naif Health Research Center, King Saud University, Riyadh 11461, Kingdom of Saudi Arabia

*Corresponding author: M Elsafadi, Stem Cells Unit, Department of Anatomy, College of Medicine, King Saud University, PO Box 295577, Riyadh 11351, Kingdom of Saudi Arabia. Tel: +966 1 503117199; Fax: +966 1 14653362

or A Mahmood, PO Box 2925 (28), Riyadh 11461, Kingdom of Saudi Arabia. Tel: +966 1 536161760; Fax: +966 1 14671300;

E-mail: monasafadi@gmail.com or ammahmood@ksu.edu.sa

Abbreviations: *TAGLN*, Transgelin; TGF β , transforming growth factor beta; hMSC, human bone marrow-derived stromal (skeletal) stem cells; CL1, hMSC clone 1 high osteogenic cells; CL2, hMSC clone 2 low osteogenic cells; mRNA, messenger RNA; siRNA, small interfering RNA; qRT-PCR, quantitative real-time polymerase chain reaction; RUNX2, runt related runt related transcription factor 2; ALPL, alkaline phosphatase, liver/bone/kidney; MAPK, mitogen-activated protein kinase; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; ABL, Applied Biosystem; OC, osteocalcin; ADIPOQ, adiponectin; PPARG2, peroxisome proliferator activated receptor gamma; LPL, lipoprotein lipase; COL1A, collagen type I alpha; POSTN, periostin; DKK2, DICKKOPF-related protein 2; HA/TCP, hydroxyapatite–tricalcium phosphate; NOD/SCID, Non-obese diabetic/ severe combined immunodeficiency; hFF, human foreskin fibroblast; PD, population doubling; RTCA, Real-time cell analysis; FITC, Fluorescein isothiocyanate

Received 13.1.16; revised 08.4.16; accepted 30.5.16; Edited by Y Shi

mouse embryogenesis.⁹ It is known as one of the earliest commitment markers of differentiated smooth muscle cells,^{10–12} and has been suggested to regulate their contractile functions. TAGLN-deficient mice were fertile and developed normally;¹³ however, vascular smooth muscle cells exhibited a pronounced alterations in the distribution of the actin filament and changes in cytoskeletal organization.¹⁴

The aim of this study was thus to examine the novel biological role for TAGLN in regulating hMSC lineage-specific differentiation and functions. We employed a number of cellular and molecular approaches including loss-of-function and gain-of-function studies. Our data suggest that TAGLN is important for differentiation progression of hMSC through regulation of distribution of actin filaments and cytoskeletal organization.

Results

Activation of TGF β 1 signaling induces TAGLN expression and enhances hMSC differentiation. We have identified TAGLN as one out of 11 genes that were upregulated during osteogenic differentiation and adipogenic differentiation of hMSC as well as enriched in the hMSC clone 1 high osteogenic cell (CL1) cell line, which is an hMSC cell line that exhibits enhanced osteogenic and adipogenic differentiation (Figure 1a). We chose TAGLN as its role in regulating hMSC differentiation has not been investigated. Given the known role of TGF β signaling in regulating TAGLN expression, we subsequently assessed the effect of TGF β treatment on TAGLN expression and hMSC differentiation. Adding TGF β 1 (10 ng/ml) to osteoblast induction medium enhanced osteoblast differentiation as evidenced by increased extracellular mineralized matrix formation (Figures 1b and c) and expression of osteoblast lineage gene markers: *RUNX2* (runt related transcription factor 2), *ALPL* (alkaline phosphatase, liver/bone/kidney) and osteocalcin (*OC*; Figure 1d). Similarly, TGF β 1 enhanced adipocytic differentiation as evidenced by increased formation of mature lipid-filled adipocytes (Figures 1e and f) and expression of adipocyte lineage gene markers: *PPARG2* (peroxisome proliferator activated receptor gamma), *LPL* (lipoprotein lipase), and adiponectin (*ADIPOQ*; Figure 1g). Adding TGF β 1 signaling inhibitor SB 431542 (SB; 10 μ m) to the induction media abolished TGF β 1-stimulatory effect (Figures 1b–g). As seen in Figure 1h, the enhanced osteoblast and adipocyte differentiation by TGF β 1 treatment was associated with increased expression of TAGLN. Dose–response experiments revealed induction of TAGLN by TGF β 1 in a concentration range 0.1–20 ng/ml following 24 or 48 h treatment (Figure 1i). The specificity of TGF β 1 induction of TAGLN was confirmed by adding SB 431542 (10 μ m) to the culture media for variable time (2 h–72 h) that resulted in inhibition of TAGLN expression already after 2 h treatment and with a maximal effect observed after 24 h and it continued for 72 h (Figure 1j). We confirmed the induction of TAGLN by TGF β 1 and inhibition by SB using western blot analysis (Supplementary Figure S1A). To investigate whether the TGF β 1-induced TAGLN expression is mediated by the SMAD2–/3 pathway, we tested phosphorylation levels of SMAD2 and found that TGF β 1 treatment increased phospho-SMAD2 levels (Supplementary Figure S1B).

TAGLN-depleted cells exhibited impaired osteoblast and adipocyte differentiation. As the enhanced osteoblastic and adipocytic differentiation by TGF β 1 was associated with increased TAGLN expression, we employed a loss-of-function approach to assess the role of TAGLN in the differentiation processes. Small interfering RNA (siRNA)-targeting TAGLN downregulated TAGLN gene expression (Figure 2a) even in the presence of TGF β 1 (10 ng/ml; Figure 2a). Western blot analysis confirmed suppression of TAGLN expression at the protein level (Supplementary Figure S1C).

As shown in Figure 2b, TAGLN-siRNA cells exhibited impaired osteoblast differentiation demonstrated by significant reduction in mineralized matrix formation in absence or presence of TGF β 1 treatment (10 ng/ml; Figure 2b) that was confirmed by alizarin red quantification (Figure 2c), and decreased expression of osteoblastic gene markers: *RUNX2*, *ALPL*, *COL1A1* (collagen type I alpha), *COL1A2*, periostin (*POSTN*), and *DICKKOPF-related protein 2* (*DKK2*; Figure 2d and Supplementary Figure S1D). The following markers *RUNX2*, *POSTN*, and *DKK2* were upregulated in the presence of TGF β 1 and were significantly decreased in TAGLN-siRNA cells (Supplementary Figure S1D). Similarly, TAGLN-siRNA cells exhibited impaired adipocytic differentiation shown by decreased number of lipid-filled mature adipocytes quantified by Nile Red staining (Figure 2e) and decreased expression of adipocytic gene markers: *PPARY2*, *LPL*, *AP2*, and *ADIPOQ* (Figure 2f). The effects of TAGLN deficiency on hMSC differentiation were further confirmed by establishing hMSC with a stable knockdown of TAGLN using shRNA (TAGLN-shRNA), where similar results were obtained (Supplementary Figure S2).

TAGLN overexpression exhibited enhanced osteoblast and adipocyte differentiation of hMSC. We established a TAGLN stably overexpressing hMSC-TERT (TAGLN-hMSC) by lentiviral transduction. The overexpression of TAGLN was confirmed by quantitative real-time polymerase chain reaction (qRT-PCR; Figure 3a), western blot analysis (Figure 3b), and immunocytochemical staining (Figure 3c). To examine the *in vivo* differentiation capacity, TAGLN-hMSC cells were mixed with hydroxyapatite–tricalcium phosphate (HA/TCP) and implanted subcutaneously into non-obese diabetic/severe combined immunodeficiency (NOD/SCID) mice. Histological analysis of the implants revealed significant increased formation of ectopic bone in TAGLN-hMSC, as assessed by twofold increase in quantification of newly formed bone of TAGLN-hMSC comparing with the control (Figure 3d). Following differentiation induction, TAGLN-hMSC exhibited enhanced differentiation to osteoblastic cells evidenced by increased Alizarin Red S staining for formed mineralized matrix and expression of osteogenic gene markers (Figures 3e–g). In addition, adipocyte differentiation was enhanced as shown by increased number of Nile Red-positive mature adipocytes and significant upregulation of adipocytic gene expression (Figures 3h–j).

TAGLN-overexpressing hFFs exhibited enhanced osteoblastic and adipocytic differentiation. To confirm that TAGLN's osteogenic and adipogenic induction potential is not restricted to BM-derived hMSC, we sought to assess the role of TAGLN-hFFs.⁴ The TAGLN-hFF cells showed

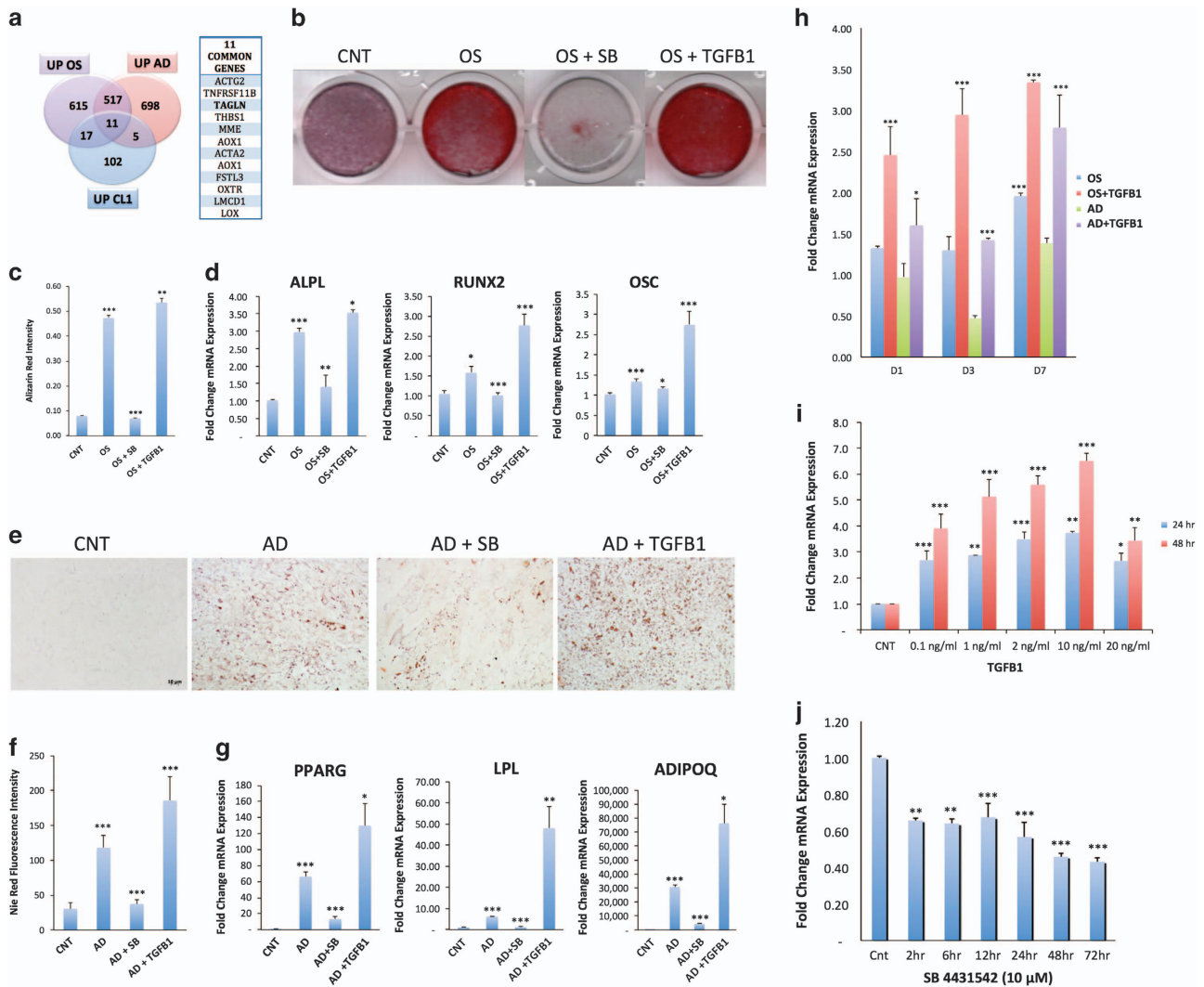


Figure 1 Activation of TGF β 1 signaling induces TAGLN expression and enhances hMSC differentiation. (a) Venn diagram depicting the overlap between the differentially upregulated genes during osteogenic differentiation of hMSC, the upregulated genes during adipogenic differentiation of hMSC and the upregulated genes of CL1 versus CL2 cells. CL1 cells were differentiated into osteoblasts by osteogenic mixture for 7 days. (b) Mineralized matrix stained by Alizarin Red S ($\times 20$, magnification). (c) Quantification of Alizarin Red S staining: control non-induced culture (NI), osteoblast-induced cultures (OS), with TGF β 1 (10 ng/ml) or SB 431542 (10 μ m). (d) qRT-PCR performed for osteogenic markers, including *ALPL*, *RUNX2*, and *OSC*. Cells were differentiated into adipocytes by adipogenic induction mixture for 7 days. (e) Oil red-O staining of mature lipid-filled adipocytes ($\times 20$, magnification). (f) Quantification of Nile Red staining of mature lipid-filled adipocytes: non-induced controls (NI), adipocyte-induced (AD), with TGF β 1 (10 ng/ml), or SB 431542 (10 μ m). (g) qRT-PCR for adipogenic markers: *LPL*, aP2 (adipocyte protein 2), *PPARG*, and *ADIPOQ*. (h) qRT-PCR of gene expression of *TAGLN* gene following osteogenic and adipogenic induction: D0 (non-induced), D1, D3, and D7 with and without TGF β 1 induction. D = day. (i) Time- (24 and 48 h) and dose- (TGF β 1: Control, 0.1, 1, 2, 10, and 20 ng/ml) response induction of *TAGLN* gene expression following treatment with TGF β . (j) Time-response suppression of *TAGLN* gene expression in response to SB 431542 (SB). Expression of target gene was normalized to GAPDH. Data are shown as mean \pm S.D. of three independent experiments, * $P < 0.05$; ** $P < 0.01$, *** $P < 0.005$

upregulation of *TAGLN* messenger RNA (mRNA) compared with control hFFs (Figure 4a), and increased protein expression as shown by immunocytochemical staining (Figure 4b) and western blot analysis (Figure 4c). *TAGLN*-hFFs exhibited decreased in proliferation rate, enhanced cell motility, and cell differentiation (Figures 4d–i).

TAGLN inhibited hMSC cell proliferation. To study the effects of *TAGLN* on hMSC on the overall changes in cell number, we employed one standard assay that measured cell viability, which is Alamar blue assay and a state-of-the-art real-time monitoring cell number using real-time cell analysis

(RTCA, DP system). In addition, we applied growth curves based on direct cell counting and expressed as cumulative population doublings (PDs) over 18 days of culture, in which *TAGLN-shRNA* exhibited higher PD rate compared with control cells (Figure 5a). RTCA DP system and Alamar blue assays showed increased proliferation in *TAGLN-shRNA* (Figures 5b and c). On the other hand, *TAGLN*-hMSC exhibited decreased cell viability (Figure 5d).

TAGLN enhanced hMSC migration. *TAGLN* has previously been implicated in cell migration,^{6,15–19} however, the role of *TAGLN* expression in hMSC migration has not been studied.

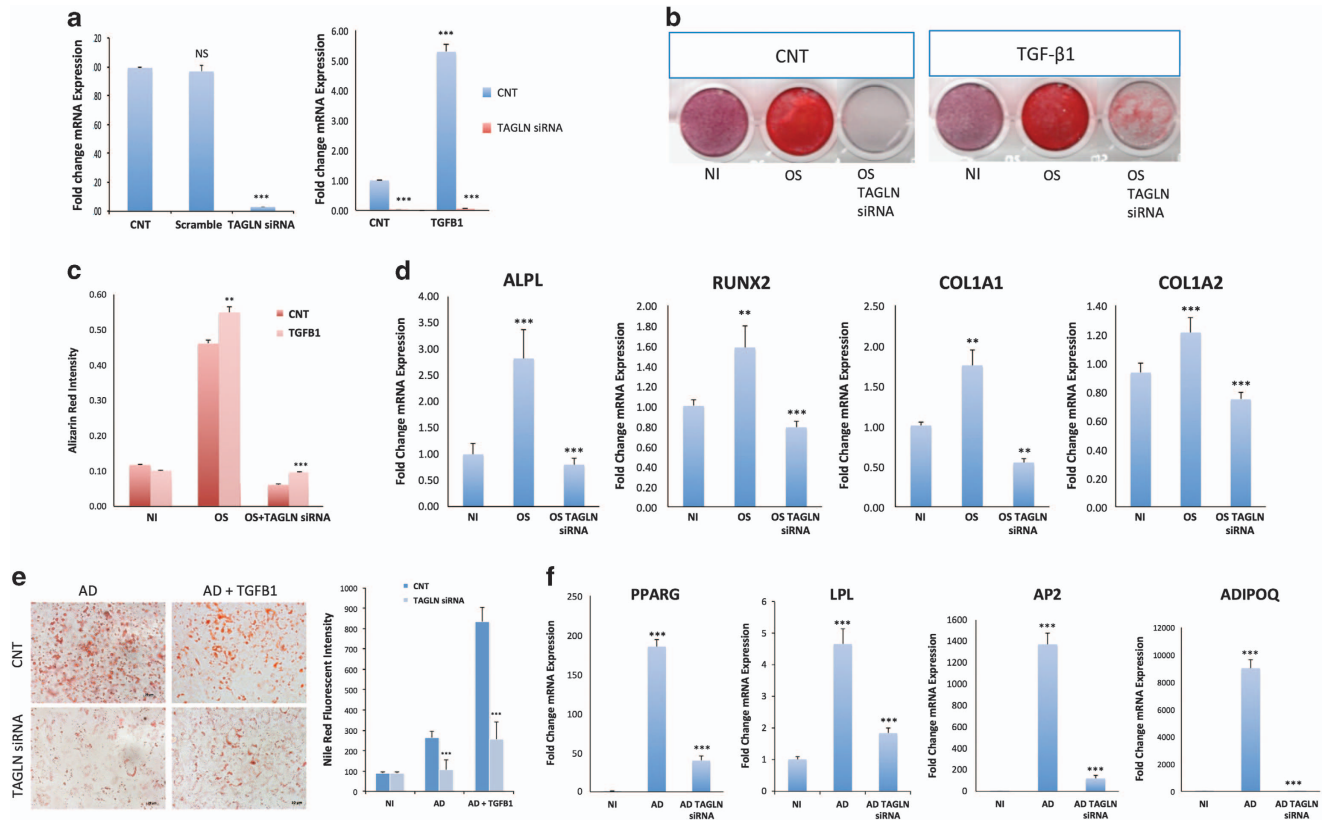


Figure 2 siRNA-mediated TAGLN suppression inhibits osteogenesis and adipogenesis. (a) Quantitative qRT-PCR for *TAGLN* gene expression 3 days post-TAGLN-siRNA, or scramble-siRNA transfection. Data are presented as fold induction. All further controls represent scramble-transfected cells. (b) Alizarin Red S staining for mineralized matrix formation. (c) Quantification of Alizarin Red staining: NI, OS, and TAGLN-siRNA cells cultured in osteoblast induction media in presence or absence of TGFβ1. (d) qRT-PCR of gene expression of osteogenic markers (*ALPL*, *RUNX2*, *COL1A1*, and *COL1A2*): NI, OS, and TAGLN-siRNA cells cultured in osteoblast-induction media. (e) Cells were induced using adipocyte induction mixture for 5 days and stained by Nile red for quantification of mature lipid-filled adipocytes: NI, AD, and adipocyte-induced cultures in presence of TGFβ1 (AD+TGFβ1) in control cells or cells transfected with TAGLN-siRNA. (f) qRT-PCR was performed for adipogenic markers' gene expression: *PPARG*, *LPL*, *ADIPOQ* (adipocyte protein 2), and *ADIPOQ*: NI, AD, and adipocyte-induced cultures in presence of TGFβ1 (AD+TGFβ1). Expression of each target gene was normalized to GAPDH. Data are presented as mean ± S.D. from three independent biological samples from at least two independent experiments. ** $P < 0.01$, *** $P < 0.005$

Thus, we studied the effects of TAGLN deficiency and overexpression on hMSC cell migration in two independent assays. In scratch assays, TAGLN-hMSC migrated faster and tended to close the scratch defect compared with control cells or TAGLN-shRNA (Figures 6a and b). We have also conducted transwell migration assay using the RTCA DP system that revealed impaired migration of TAGLN-shRNA compared with control cells (Figure 6c).

TAGLN-shRNA cells exhibited cellular and nuclear morphological changes. The observed effects of TAGLN on hMSC cell migration prompted us to study its effects on cell morphology and cytoskeleton. For this, we employed high content imaging using Opretta of Phalloidin-fluorescein isothiocyanate (FITC) staining for F-actin. As shown in Figure 6, TAGLN-shRNA cells exhibited increased cell roundness, percentage of rounded cells, index of nuclei mean roundness, and nuclei width to length ratio, but reduced cell area and nuclear area (Figure 6d). F-Actin staining revealed decreased staining intensity in TAGLN-shRNA cells (Figure 6e).

In addition, we employed transmission electron microscopic examination of TAGLN-shRNA cells and TAGLN-hMSC (Figure 7, Supplementary Figures S3–S6). In control cells, we observed prominent actin filament organized as bundles/aggregates and distributed in the whole cytoplasm and in perinuclear locations. In addition, the cells contained surface-branched microvilli. In contrast, TAGLN-shRNA contained relatively fewer actin filaments, whereas TAGLN-hMSC showed high content of actin filaments. Furthermore, rough endoplasmic reticulum (rER) was dilated in TAGLN-hMSC, suggesting increased protein synthesis activity. Interestingly, cellular processes were observed extending from TAGLN-hMSC that may enhance cellular motility. The ultrastructural characteristics of the examined cells are summarized in Supplementary Table C.

Gene expression profiling of TAGLN-depleted cells. In order to determine the molecular mechanisms underlying TAGLN functions in hMSC biology, we performed global gene expression profiling in TAGLN-siRNA cells and control cells. Hierarchical clustering revealed that control and TAGLN-siRNA samples clustered separately (Figure 8a). We found

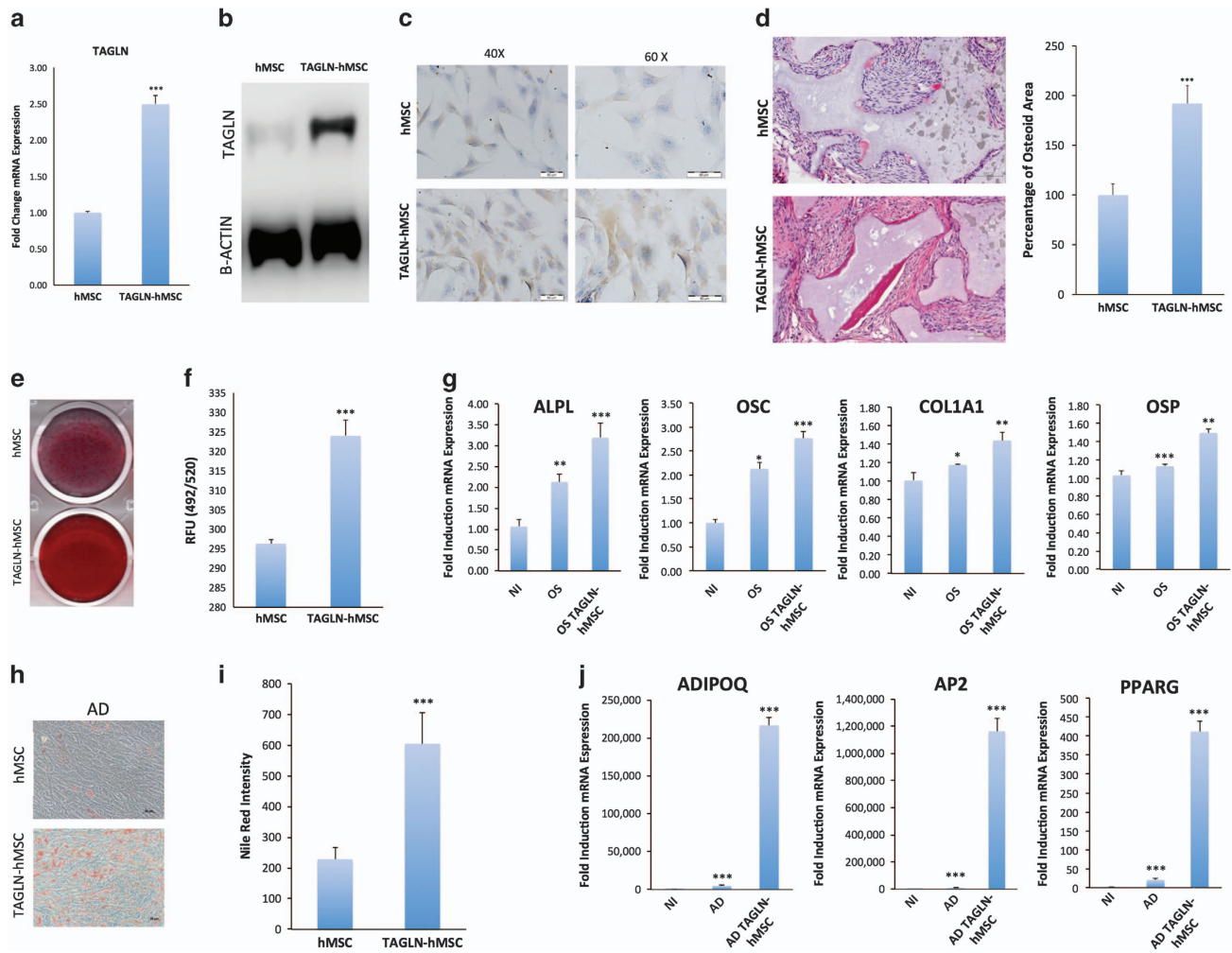


Figure 3 TAGLN overexpression induces osteogenesis and adipogenesis in hMSC. (a) All controls represent empty vector-transfected cells. qRT-PCR of *TAGLN* gene expression in control (empty vector) hMSC and TAGLN-overexpressing line (TAGLN-hMSC). (b) Western blotting for TAGLN in TAGLN-hMSC (upper panel) and B-Actin (lower panel). (c) TAGLN immunocytochemical staining in control hMSC and TAGLN-hMSC cells at magnification $\times 40$ and $\times 60$. Cells were differentiated into osteoblasts by osteogenic or adipogenic induction mixture for 14 days. (d) hMSC and TAGLN-hMSC cells were implanted with HA/TCP subcutaneously into NOD/SCID mice. The histology of *in vivo* formed bone was examined with H&E staining (original magnification $\times 200$). High-resolution whole-slide digital scans of all histological sections were created with a ScanScope scanner and bone formed was quantified. (e) Mineralized bone matrix formation in control hMSC (empty vector) cells and TAGLN-hMSC stained for Alizarin Red. (f) Quantification of Alizarin Red staining in control hMSC and TAGLN-hMSC. (g) qRT-PCR for osteogenic markers: ALPL, OSC, COL1A1 (collagen type I), and OSP: NI, OS, and osteoblast-induced TAGLN-hMSC (OS TAGLN-hMSC). (h) Both control (empty vector) hMSC, and TAGLN-hMSC were induced to adipocyte differentiation using adipocyte induction medium for 7 days. Mature lipid-filled adipocytes were stained by oil red. (i) Quantification of Nile Red staining for mature adipocytes: control (empty vector) hMSC, TAGLN-hMSC. (j) qRT-PCR for adipogenic markers: ADIPOQ, LPL, AP2 (adipocyte protein 2), and PPARG in NI, AD, and adipo-induced TAGLN-hMSC line (AD TAGLN-hMSC). Data are shown as mean \pm S.D. from at least two independent experiments, * $P < 0.05$; ** $P < 0.01$, *** $P < 0.005$

that 6351 genes were upregulated and 3159 genes were downregulated in TAGLN-siRNA cells compared with control cells (fold change (FC) ≥ 2.0 , $P < 0.02$, Supplementary Table F). Among the significantly downregulated genes were integrin family (*ITGA4*, *ITGA10*, and *ITGB1BP1*), *TPM1*, *Fos*, *SRF*, and *CSRP3* as well as mesodermal genes such as twist family (*TWIST1* and *TWIST2*), and MFH-1, *FoxC2* (Figure 8b). In addition, osteoblast-associated genes such as *ALPL* and *BMP4* were downregulated (Figure 8b). In addition, Venn diagram showed 60 common downregulated genes in TAGLN-deficient cells during osteoblast and adipocyte differentiation including *NR1H3*, *CDF*, *IGF2*, and *IGFBP2*, which could potentially have a role in commitment of hMSC to

osteoblastic and adipocytic lineage (Figure 8c). As a confirmation of the microarray data, qRT-PCR showed good concordance between microarray and qRT-PCR for a selected gene panel and are known to be involved in cell differentiation and the TGF β pathway (Figure 8d).

Pathway analysis of significantly regulated genes in TAGLN-siRNA cells. TAGLN-siRNA cells exhibited downregulation in several intracellular signaling pathways including pathways regulating actin cytoskeleton, focal adhesions (FAs), endochondral ossification, adipogenesis, TGF β signaling, and MAPK cascade (Figure 8e, Supplementary Figures S10–S13). Figure 8e shows a pie chart of

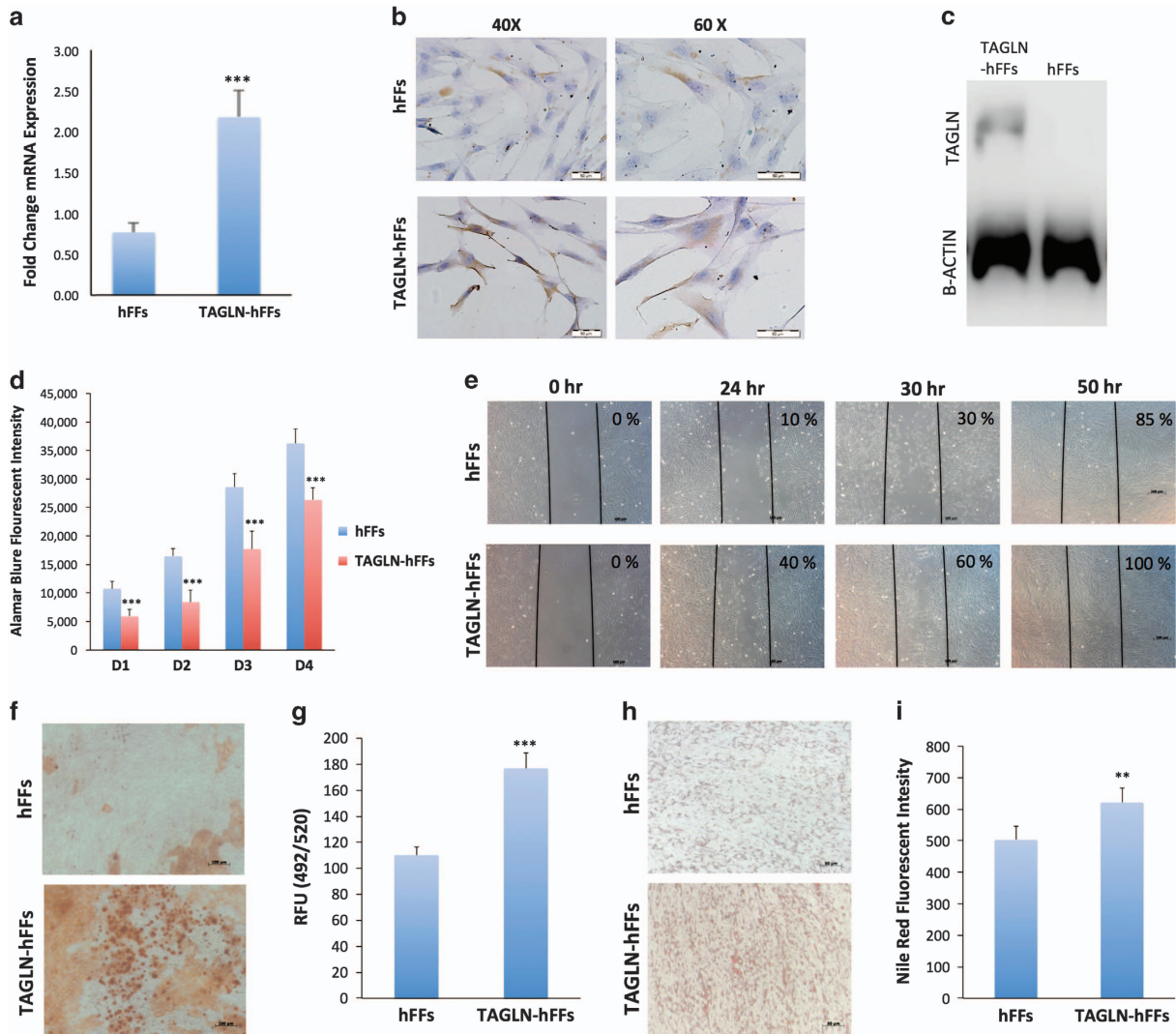


Figure 4 Effects of TAGLN overexpression on hFFs functions. (a) qRT-PCR of *TAGLN* gene expression in control (empty vector) TAGLN-hFFs. (b) Immunocytochemical staining for TAGLN protein in control hFFs, and TAGLN-hFFs at magnifications $\times 40$ and $\times 60$. (c) Western blot of TAGLN (upper panel) and B-Actin (lower panel) in control hFFs and TAGLN-hFFs. Cells were induced to differentiate into osteoblasts or adipocytes by either osteogenic or adipogenic mixture for 24 days. (d) Alamar blue staining for cell viability in control hFFs and TAGLN-hFFs on day (D): 1D, 2D, 3D, and 4D. (e) Scratch assay where the motility of hFFs and TAGLN-hFFs cells was observed at different time intervals: 0, 24, 30, and 50 h. (f) Alizarin Red S staining for mineralized matrix formation. (g) Quantification of mineralized matrix formation in control hFFs cells and TAGLN-hFFs using Osteo-Image Mineralization Assay. (h) Control hFFs and hFF-TAGLN were induced to adipocyte differentiation by incubation in adipocyte induction medium for 24 days and stained by oil red for lipid-filled mature adipocytes. (i) Quantification of Nile red staining of mature adipocytes formed in control hFF cells and TAGLN-hFFs. Data are shown as mean \pm S.D. of three independent experiments. ** $P < 0.01$, *** $P < 0.005$

the top significantly regulated signaling pathways in TAGLN-siRNA cells. Interestingly, 21 FA genes and 18 actin cytoskeleton-regulating genes were downregulated in TAGLN-siRNA cells including *AKT1*, *AKT2*, *PFN1*, *ARPC5*, *VIL2*, and integrins (Supplementary Table D).

Gene expression profiling in TAGLN-siRNA during osteoblast differentiation. Transcriptomic profiling was conducted on CL1 control and TAGLN-depleted cells on day 3 of osteogenic differentiation. As shown in Supplementary Figure S7A, hierarchical clustering revealed clear separation of the control and TAGLN-siRNA samples. A list of the 17 osteogenic genes that were downregulated in TAGLN-siRNA cells is shown in Supplementary Figure S7B and

includes *BMP4*, *COL12A1*, *CL8A1*, *FZD1*, *IGF2*, and *SMAD6*. Comparing the list of the downregulated genes in TAGLN-siRNA cells and the upregulated genes in the CL1 control cells, in the presence of osteogenic induction medium, identified 256 overlapped genes that are predicted to be involved in osteogenic differentiation (Supplementary Figure S7C). Furthermore, using significance analysis, 6004 up- and 2752 downregulated genes were identified in TAGLN-depleted cells during osteogenesis (2.0 FC, $P < 0.02$, Supplementary Table F). Pathway analysis on the downregulated genes using GeneSpring GX revealed significant changes in several pathways related to osteogenesis including osteoblast signaling, osteopontin signaling, TGF β 1 signaling, adipogenesis, endochondral ossification,

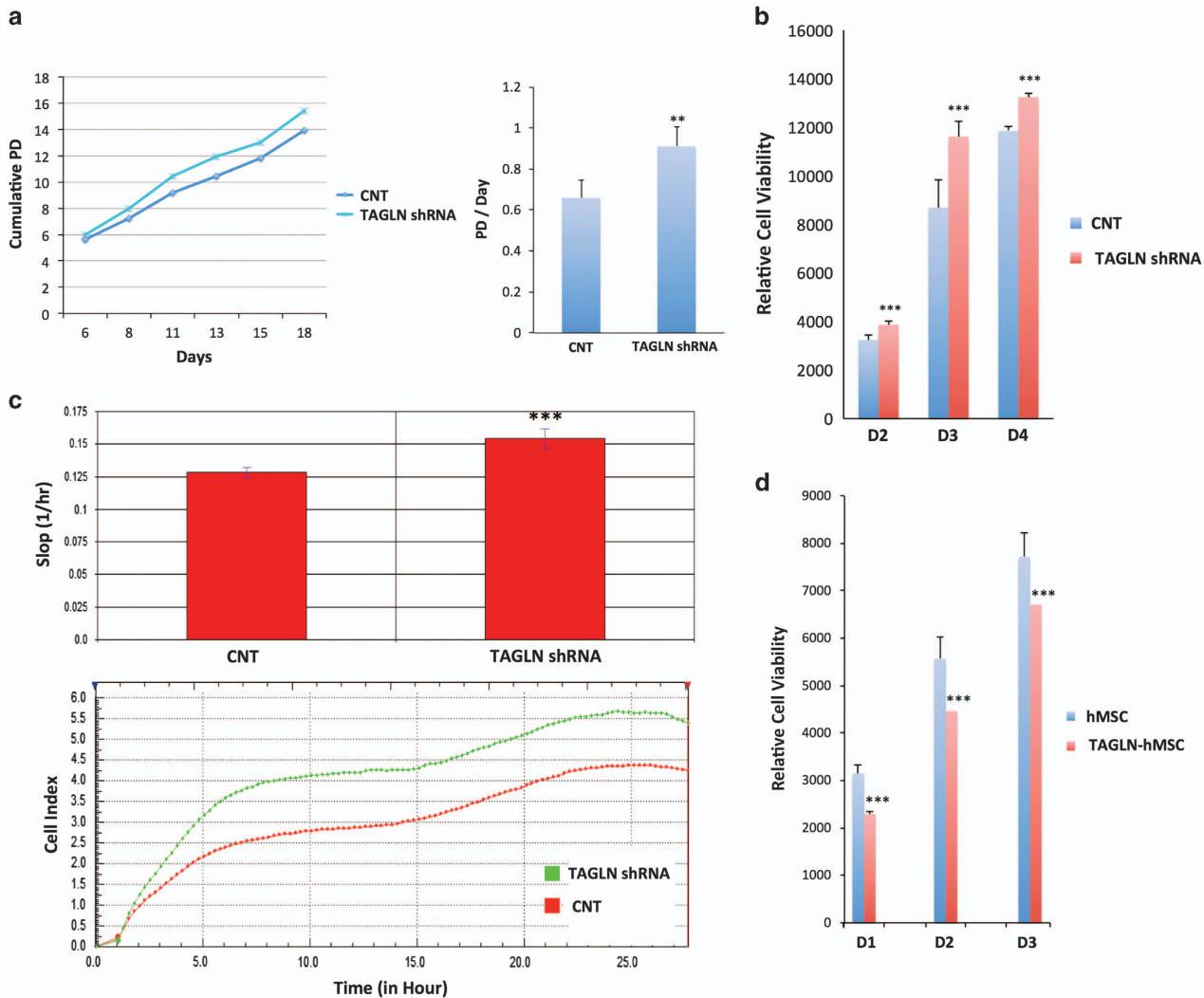


Figure 5 TAGLN inhibits cells proliferation. (a) Short-term growth curve showing cumulative PD over 18-day culture of control CL1 (CNT) and TAGLN-deficient cells (TAGLN-shRNA). (b) Alamar blue staining showing cell viability in control cells (CNT) and TAGLN-shRNA on days (D) 2, D3, and D4. (c) Proliferation assay using DIC device with the RTCA DP system. CNT and TAGLN-shRNA. Cell proliferation was measured at 15-min intervals for 27 h (lower panel). Upper panel shows cell proliferation slope during the 27 h observation period. (d) Alamar blue staining for control hMSC and TAGLN-overexpressing hMSC (TAGLN-hMSC) during 4 days of culture. Data are shown as mean \pm S.D. of at least two independent experiments. ** $P < 0.01$, *** $P < 0.005$

and regulation of microtubule cytoskeleton (Supplementary Figure S7D).

Gene expression profiling in TAGLN-depleted cells during adipocytic differentiation. Global gene expression profiling was conducted on parental and TAGLN-siRNA cells on day 3 of adipogenic differentiation. As shown in Supplementary Figure S8A, hierarchical clustering revealed clear separation of the CL1 control cells and TAGLN-siRNA cells. Using significance analysis, 2990 up- and 4717 downregulated genes were identified in TAGLN-depleted cells during adipogenesis (2.0 FC, $P < 0.02$, Supplementary Table F). In addition, the adipogenic inhibition in TAGLN-siRNA cells was associated with downregulation of adipogenic markers including *FABP4*, *LPL*, *ADIRF*, *LIPE*, *APOL4*, *APOL6*, *AOC3*, and *CEBPA* (Supplementary Figure S8B). Comparing the list of the downregulated genes in TAGLN

siRNA cells and the upregulated genes in control cells, in the presence of adipogenic induction medium, revealed 729 common genes, which are predicted to be involved in adipogenic differentiation (Supplementary Figure S8C). Pathway analysis revealed significant enrichment in several pathways related to the TGF β 1 signaling pathway, adipogenesis, fatty-acid biosynthesis, integrin-mediated cell adhesion, and cell differentiation (Supplementary Figure S8D).

Discussion

Identifying regulatory factors that have a role in hMSC proliferation and differentiation is prerequisite for their optimal use in regenerative medicine protocols. In the present study, we unraveled novel roles for TAGLN in hMSC proliferation, migration, and differentiation. TAGLN was observed to be upregulated during hMSC lineage differentiation and in

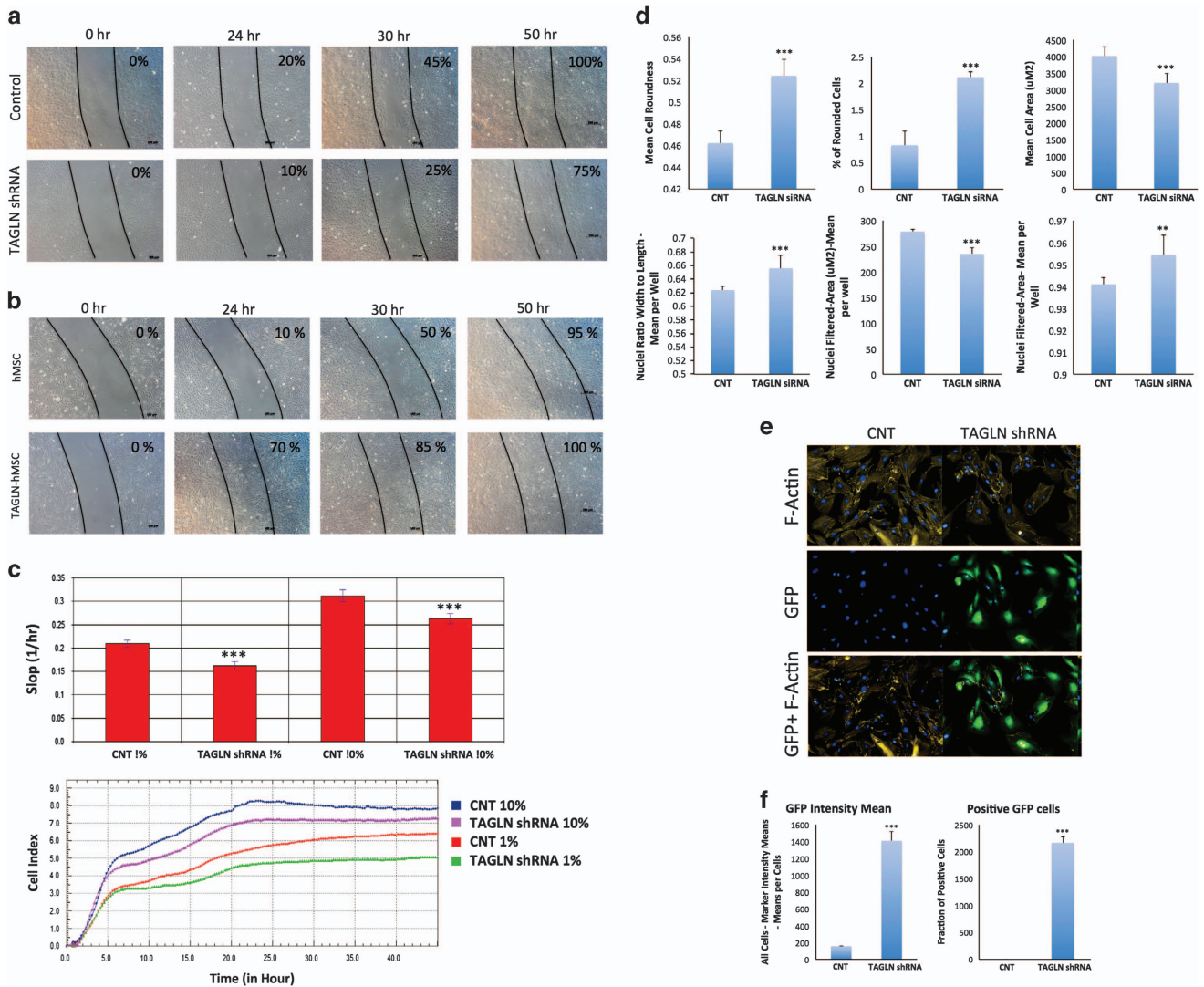


Figure 6 TAGLN induces changes in cell morphology and enhances transwell cell migration. Scratch assay where a scratch is made in 50–70% confluent cells and cell motility is observed at 0, 24, 30, and 50 h in (a) control CL1 (control) and TAGLN-deficient cells (TAGLN-shRNA), and (b) control hMSC and TAGLN-overexpressing hMSC (TAGLN-hMSC). (c) Transwell migration assay using DIC device with the RTCA DP system. FBS was used as chemoattractant. The curves show the number of cell migrated up to 40 h (lower panel). Control empty vector cells (CNT) and TAGLN-shRNA cultured in presence of 1 or 10% FBS. The cell migration was measured at 15 min intervals for 45 h. Upper panel shows the average cell migration slope during 24 h. (d) High content image analysis using Opreta of cell morphology in TAGLN-shRNA as compared with CNT. (e) Immunocytochemical staining for F-Actin protein in CNT and TAGLN-shRNA. (f) High content image analysis of GFP intensity and the number of GFP-positive cells in CNT and TAGLN-shRNA

response to TGF β activation. TAGLN-mediated TGF β -dependent enhancement of lineage differentiation into osteoblasts and adipocytes inhibited hMSC cell proliferation and enhanced their migratory capacity. These effects are possibly mediated through changes in actin distribution and cytoskeleton dynamics.

We observed that TGF β 1 upregulated TAGLN expression in hMSC, and these effects were specific, as they were abolished by TGF β -signaling inhibitor SB 431542. TAGLN expression was associated with increased SMAD phosphorylation. Previous studies have reported that TGF β 1 upregulated TAGLN expression in a number of cell models, for example, smooth muscle cells, epithelial cells, neural crest stem cells Monc-1, and embryonic fibroblast cells (BALB 3T3).^{6,20,21}

In addition, TGF β 1-induced TAGLN expression is mediated by the SMAD2/3 signaling pathway.^{6,22} Thus, TGF β -mediated TAGLN expression represents a general phenomenon.

Interestingly, adding TGF β 1 to hMSC induction media induced both osteogenic and adipogenic differentiation, and these effects were dependent on TAGLN as they were abolished in TAGLN-deficient hMSC. TGF β 1 exerts complex effects on hMSCs. It has been demonstrated that it regulates proliferation and differentiation of osteoprogenitor cells.^{23–26} We observed that TAGLN exerted opposing effects on hMSC proliferation (inhibition) and differentiation (enhancement). Similar phenomenon has been observed in rat calvarial cultures where osteoblast differentiation coincided with decreased cell proliferation.²⁷ It is plausible that

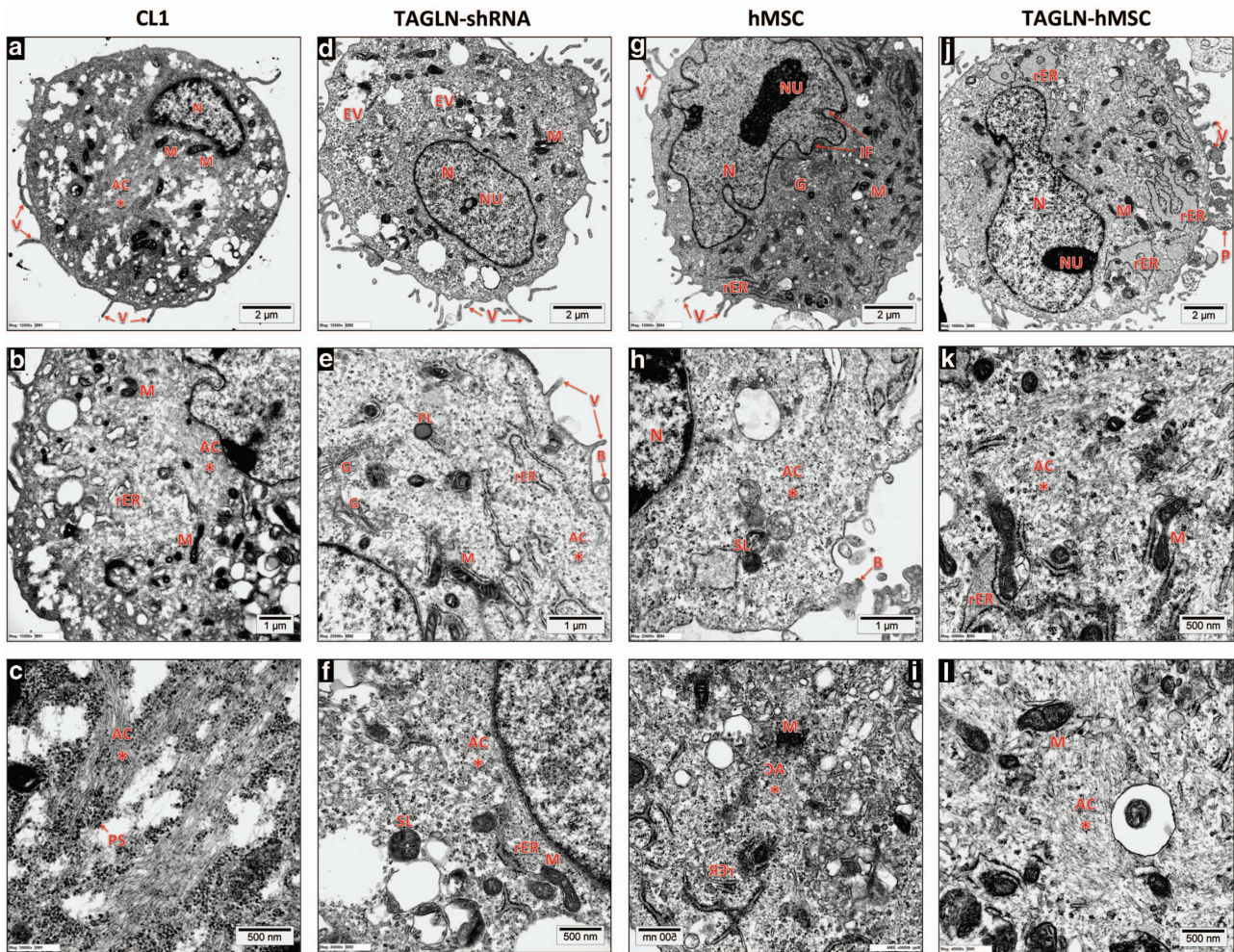


Figure 7 Transmission electron microscopy (TEM) analysis. Ultra-thin sections from resin embedded samples were ultrastructural analyzed using TEM. (a-c) Control empty vector (CL1) cells with magnification x12 000, x15 000 and x50 000 respectively. (d-f) TAGLN deficient cells (TAGLN-shRNA) with magnification x12 000, x25 000 and x40 000 respectively. (g-i) Control empty vector hMSC (hMSC) cells with magnification x12 000, x25 000 and x40 000 respectively. (j-l) hMSC overexpressing TAGLN (TAGLN-hMSC) cells with magnification x10 000, x25 000 and x40 000 respectively. AC, actin filaments; B, cell blebs; EV, endocytotic vacuole; G, Golgi bodies; IF, infolding of nuclear membranes; M, mitochondria; N, nucleus; Nu, nucleolus; P, cell processes; PL, primary lysosome; rER, rough endoplasmic reticulum; SL, secondary lysosome; V, microvilli

TAGLN-mediated effects are based on increasing the committed precursor pool, thus allowing more cells to differentiate into osteoblasts and adipocytes. Taken together, we proposed that the molecular changes mediated via TGF β -induced TAGLN defines the biological process of hMSCs, and their osteogenic and adipogenic differentiation and proliferation potential.

In our studies, TAGLN enhanced hMSC migration. Similar to our findings, TAGLN silencing inhibited TGF β 1-induced cell migration in lung fibrosis model.⁶ Interestingly, it has been hypothesized that TGF β 1 released from the bone matrix during bone resorption phase of bone remodeling mediates coupling of bone resorption to bone formation by inducing migration of hMSC to the bone-formation sites.^{28,29} It is plausible that the enhanced hMSC motility and migration is achieved through upregulation of TAGLN and may thus have a role in recruiting progenitor cells to the bone-formation sites.

TAGLN overexpression or deficiency resulted in significant changes in cell morphology, cytoskeleton, and functional capacity for migration. It has been reported that TAGLN is localized to cellular cytoskeleton and filamentous actin, which are the principal components of the cytoskeleton,^{13,30–32} and thus can actively contribute to regulation of cell shape and cell polarity. The observed changes in cell morphology and cytoskeleton may have a role in hMSC differentiation. It has been reported that significant changes took place in the cytoskeleton during lineage commitment and differentiation of hMSC.^{33,34} During hMSCs' lineage commitment, the cells undergo significant modifications in morphology and actin cytoskeletal organization, and this participates in cell-fate determination.³³ In addition, during osteoblast differentiation, hMSCs morphologically change from fibroblast-like cells to cuboidal morphology, which is associated with changes in the cytoskeleton.³⁵ Similarly, during differentiation of hMSCs to

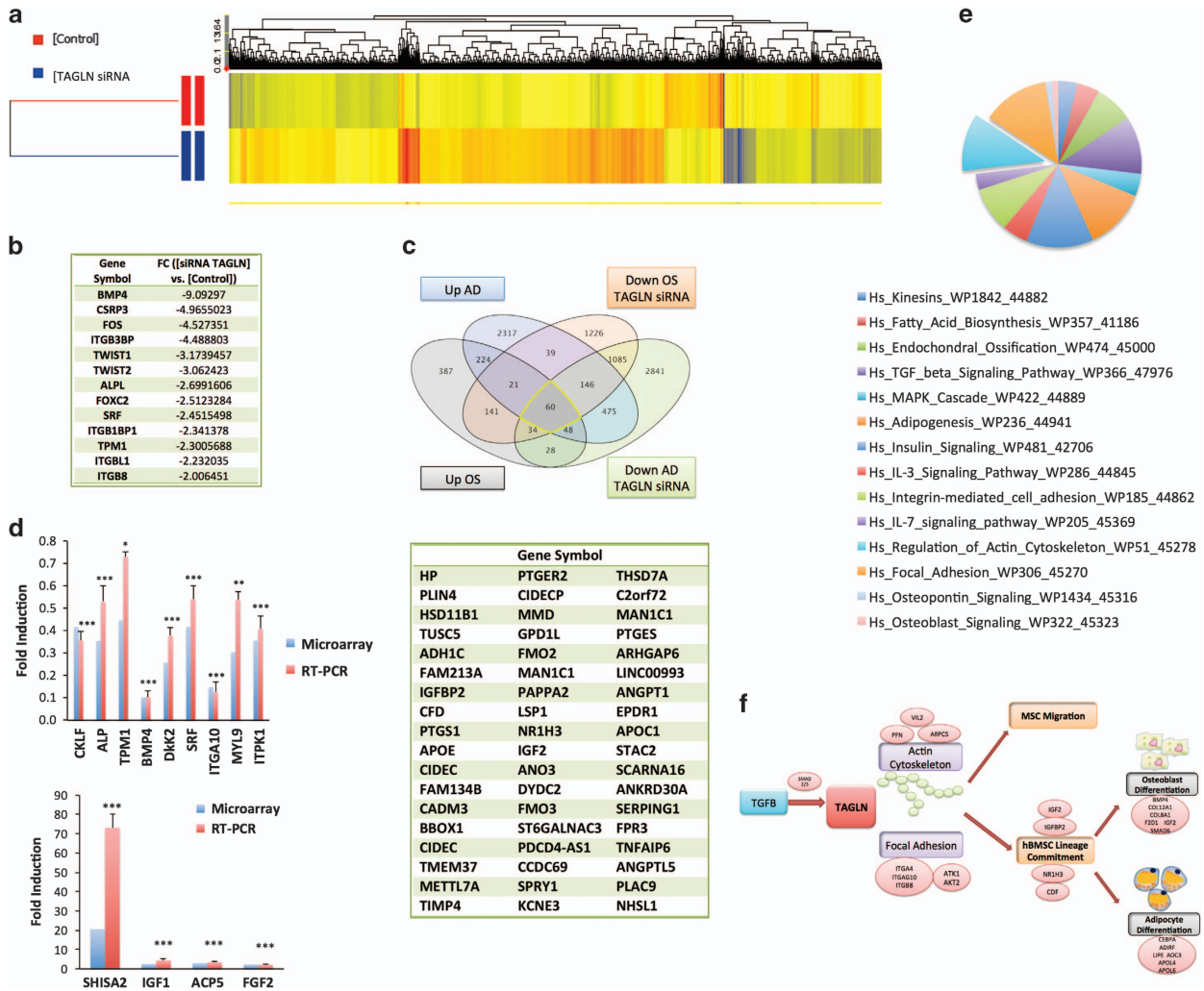


Figure 8 Gene expression profiling of TAGLN-depleted cells (TAGLN-siRNA). (a) Hierarchical clustering of control cells (control) and TAGLN-siRNA. Each row represents a biological sample and each column represents a transcript. Expression level of each gene in a single sample is depicted according to the color scale. (b) List of differentiation-associated genes that were downregulated in TAGLN-siRNA. (c) Venn diagram revealing overlap between the upregulated genes and the downregulated genes in TAGLN-siRNA following osteogenic and adipogenic differentiation. The list of the 60 common genes shown in the Venn diagram. (d) qRT-PCR validation of selected genes $*P < 0.05$; $***P < 0.001$. (e) Pie chart illustrating the distribution of the top pathways enriched in the downregulated genes in TAGLN-siRNA identified during osteoblast differentiation. (f) A working model illustrating the effect of TGFβ-induced TAGLN expression on hMSC migration, and osteoblastic and adipocytic differentiation by modulating actin cytoskeletal organization and focal adhesion molecules and suggesting NRH3, CDF, IGFBP2, and IGF2 genes as downstream targets

adipocytes, significant changes in cell morphology takes place.^{36,37} Our data are in concordance with previous work showing that TGFβ1 treatment suppresses MSC proliferation and participates in cytoskeleton changes and thus hMSC differentiation.³⁸ Finally, osteoblast differentiation in response to mechanical stimulation is associated with alteration in actin cytoskeletal organization and is regulated by signals transmitted through membrane-associated adhesion molecules.³⁹ The molecular mechanisms linking changes of the cytoskeleton and hMSC differentiation are under investigation. One of the extensively studied mechanisms involves Rho GTPase/Rho-associated Kinase, adhesion kinases, and extracellular matrix components that regulate Actin Depolymerization Factor (ADF) expression, for example, Destrin, Cofilin1, and Cofilin2.⁴⁰⁻⁴² A recent study from our group has demonstrated that, during osteoblast differentiation of hMSCs, alteration in

ADF expression leads to changes in the ratio of G-actin (monomer) and F-actin (polymerized), and inhibit actin depolymerization.³³

Treating bone marrow-derived hMSCs with TGFβ1 has previously reported to regulate differentiation potential of MSCs.⁴³ On the basis of our study, we propose that the mechanism by which the TGFβ1/TAGLN axis regulates hMSC differentiation into osteoblasts and adipocytes is through changes in the cell cytoskeleton. Given that TAGLN is an actin-crosslinking or actin-gelling protein,^{9,44} our studies demonstrate that the structure of actin cytoskeleton determines the transition from undifferentiated to committed differentiating hMSC. It is also plausible that osteoblast/adipocyte-differentiated phenotype requires a special actin cytoskeleton structure, which is distinct from the cytoskeletal structure of proliferating undifferentiated cells.

Interestingly, our TEM studies have revealed that rER was cystically dilated in TAGLN-hMSC, suggesting an increased protein synthesis activity. It has been shown by TEM that the ultrastructure of human bone marrow hMSCs is rich in rER with dilated cisternae⁴⁵ and that there is a reduction in the abundance of rER following osteoblast differentiation.⁴⁶ Palomäki *et al.*⁴⁷ have shown the presence of dilated rER in undifferentiated bone marrow hMSCs, whereas, after osteoblast differentiation, narrow rERs with more extracellular matrix were formed. Our data suggest that the presence of cystically dilated rER, which is an indication of hyperactivity of rER, is a prerequisite at the early stage of hMSC commitment to cell differentiation when enhanced protein synthesis is required, and this is accompanied with cytoskeletal changes, which regulate ER dynamics and activity. It is known that during mitosis the cisternal ER network changes its intracellular localization, because of cytoskeleton changes, to the peripheral membrane away from the mitotic spindle and chromosomes in order to not interfere with the segregation of chromosomes.^{48,49} This might explain the increased cell proliferation in TAGLN-depleted cells, which constitute abundant of long-stalk cisternal rER.

It has been shown that cytoskeletal and FA cooperate to influence the shape, mechanical properties and differentiation of MSCs.^{50–52} FAs (cell–matrix adhesions) are large macromolecular assemblies through which mechanical force and regulatory signals are transmitted between the extracellular matrix and an interacting cell.⁵³ FA molecules such as ATK1, ATK2, and integrins (for example, ITGA10, ITGA4, and ITGB8) have an essential role in directing MSCs to osteogenic differentiation by changing the cytoskeleton that initiates extracellular matrix-induced osteogenic differentiation.^{54–56} Concordantly, *AKT1*, *AKT2*, *PFN1*, *ARPC5*, *VIL2*, and integrins were significantly downregulated upon TAGLN depletion, suggesting their implication in actin polymerization-mediated MSC differentiation.

In summary, we proposed a model (Figure 8f) where TGFB regulates TAGLN expression and this leads to enhanced MSC cell migration and osteoblast and adipocyte differentiation through modulating actin cytoskeletal organization and FA molecules. A number of possible molecules, for example, *NRIH3*, *CDF*, *IGFBP2*, and *IGF2*, are possibly implicated in this process. Our study thus provides a novel molecular insight into the role of the TGF β intracellular signaling pathway in bone and bone marrow adipose tissue formation. In order to control the lineage fate of hMSCs, TAGLN is postulated as a potential target for pharmacological intervention.

Materials and Methods

Cell culture. We have employed the hMSC-TERT cell line, which was created from primary normal human MSC by overexpressing human telomerase reverse transcriptase gene.^{1,4} The hMSC-TERT cells have been extensively characterized, and they exhibit similar cellular and molecular phenotypes to primary MSC.⁴ For ease, we will refer to this cell line as hMSC for the remaining part of the manuscript. For the current experiments, we employed two subclones derived from hMSC called CL1 and hMSC clone 2 low osteogenic cell (CL2) that exhibits enhanced and reduced differentiation potential to osteoblasts and adipocytes, respectively. In addition, hFFs have been used to extend our functional studies of TAGLN. The cells were cultured in Dulbecco's Modified Eagle Medium (DMEM) supplemented with D-glucose 4500 mg/l, 4 mM L-Glutamine and 110 mg/l Sodium Pyruvate, 10% fetal

bovine serum (FBS), 1 \times penicillin–streptomycin (Pen–strep), and non-essential amino acids (all purchased from Gibco-Invitrogen, Waltham, MA, USA).

siRNA-mediated transfection of hMSC. For transfection, CL1 cells in the logarithmic growth phase were reversed-transfected with Silencer Select Pre-designed TAGLN-siRNA (25 nM; Ambion, Carlsbad, CA, USA; ID: s13739 and Cat. No. 4392420; Ambion, The RNA Company, USA) using Lipofectamine RNAiMAX Reagent (Invitrogen, CA, USA) plus serum-free Opti-MEM1 medium (Invitrogen) under the conditions described by the manufacturer. On day 3 of transfection, the cells were induced to osteogenic or adipogenic differentiation for an additional 1 week. Detailed information about siRNA reagents is described in Supplementary Table E.

Establishing a stable TAGLN-deficient (TAGLN-shRNA) line. CL1 cells were seeded in 24-well plates for 1 day, followed by forward transfection by 50 μ l serum-free Opti-MEM1 medium (Invitrogen) mixed with 0.5 μ l of Lipofectamine 2000 Reagent (Invitrogen). The cells were incubated for 5 min and 1 μ g of TAGLN-shRNA (GeneCopoeia, Inc., Rockville, MD, USA; Cat. No. HSH017877-2-CH1) was mixed with 50 μ l serum-free Opti-MEM1 medium (Invitrogen) and incubated for 5 min. One hundred microliters of shRNA and lipofectamine mixture were added to 0.5 ml pen–strep-free DMEM per well. Transfection medium was changed next day with Pen–Strep-free DMEM and cultured for 3 days. Puromycin (1.5 mg/ml) was added when the cells reached confluence. A OmicsLink shRNA Expression vector was used (Cat. No.: HSH017877-1-CH1, OriGene Technologies (Rockville, MD, USA)) for knocking down TAGLN with Puromycin as selection marker and GFP as reporter gene. Detailed information about shRNA reagents is described in Supplementary Table E and Supplementary Figure S18.

Establishing TAGLN-hMSC line. Lentiviral transduction was employed to create stable hMSC cell lines overexpressing TAGLN. Lentiviral transduction was conducted as previously described.⁵ Lentifect lentiviral particles encoding human TAGLN (LP-G0046-Lv213-200) or control lentiviral particles (LP-FLUC-LV105-0205) were purchased from GeneCopoeia Inc. Briefly, hundred thousand cells were seeded in DMEM in 24-well plate. Forty-eight hours later (~80 confluency), media were removed and then 20 μ l of lentiviral particles in 500 μ l of DMEM+5% heat-inactivated serum (Invitrogen) and 1% Pen–Strep supplemented with polybrene (8 μ g/ml; Sigma, St. Louis, MO, USA) were added to the cells. Seventy-two hours later, media were removed and transduced cells were expanded in T25 and selected using puromycin (1.5 mg/ml; Sigma).

In vitro osteoblast differentiation. Cells were grown in standard DMEM growth medium in six-well plates at 0.3×10^6 cells/ml. When 70–80% confluence, the cells were cultured in DMEM supplemented with osteoblast induction mixture containing 10% FBS, 1% Pen–strep, 50 μ g/ml L-ascorbic acid (Wako Chemicals, Neuss, Germany), 10 mM β -glycerophosphate (Sigma), and 10 nM calcitriol (1 α ,25-dihydroxy vitamin D3; Sigma), and 10 nM dexamethasone (Sigma). The media were replaced three times per week.

In vitro adipocyte differentiation. Cells were grown in standard DMEM growth medium in six-well plates at 0.3×10^6 cells/ml. At 90–100% confluence, cells were cultured in DMEM supplemented with adipogenic induction mixture containing 10% FBS, 10% horse serum (Sigma), 1% Pen–strep, 100 nM dexamethasone, 0.45 mM isobutyl methyl xanthine⁵⁷ (Sigma), 3 μ g/ml insulin (Sigma), and 1 μ M Rosiglitazone⁵⁸ (Novo Nordisk, Bagsvaerd, Denmark). The media were replaced three times per week.

Cytochemical staining

Alizarin Red S staining for mineralized matrix: The cell layer was washed with PBS, and then fixed with 4% paraformaldehyde for 15 min at room temperature. After removing the fixative, the cell layer was rinsed in distilled water and stained with the 2% Alizarin Red S Staining Kit (ScienceCell, Research Laboratories, Cat. No. 0223) for 20–30 min at room temperature. Excess dye was washed off with water. For quantifying the Alizarin Red S staining, the Alizarin Red S dye was eluted in 800 μ l of acetic acid incubated in each well for 30 min at room temperature as described⁵⁹ and measured in spectrophotometer (BioTek, Epoch) at 405 nm.

Osteo-image mineralization assay: The *in vitro* formed mineralized matrix was quantified using the Osteo-Image Mineralization Assay Kit (Lonza, Walkersville,

MD, USA, Cat. No. PA-1503). Culture media were removed and cells washed once with PBS, and fixed with 70% cold ethanol for 20 min. Appropriate amount as recommended by the manufacturer of diluted staining reagent was added, and plates were incubated in dark for 30 min at room temperature. Cells were washed, and staining quantitation was performed using fluorescent plate reader at 492/520 excitation emission wavelengths.

Oil red-O staining for lipid droplets: Mature adipocytes filled with cytoplasmic lipid droplets were visualized by staining with Oil Red-O. After washing with PBS, the cells were fixed in 4% formaldehyde for 10 min at room temperature, and then rinsed once with 3% isopropanol and stained for 1 h at room temperature with filtered Oil Red-O staining solution (prepared by dissolving 0.5 g Oil red-O powder in 60% isopropanol). To quantify the mature adipocytes formed, Oil Red-O stain was eluted by adding 100% isopropanol to each well and color intensity was measured with spectrophotometer at 510 nm (Biotek Spectrophotometer, Epoch).

Nile red fluorescence determination and quantification of mature adipocytes: Stock solution of Nile red (1 mg/ml) in DMSO was prepared and stored at -20°C protected from light. Staining was performed on fixed cells with 4% paraformaldehyde (Sigma) for 15 min. Cultured undifferentiated and differentiated cells were washed once with PBS. The dye was added directly to the cells (5 $\mu\text{g}/\text{ml}$ in PBS), and the cells were incubated for 10 min at RT. Fluorescent signal was measured using SpectraMax/M5 fluorescence spectrophotometer plate reader (Molecular Devices Co., Sunnyvale, CA, USA) using the bottom well-scan mode where nine readings were taken per well using excitation 485 nm and emission 572 nm spectra.

Immunocytochemical staining. The cells were fixed in 4% paraformaldehyde (Sigma) for 15 min, and were permeabilized with 0.1% Triton X-100 (Sigma) for 10 min. To block nonspecific binding, cells were treated with 3% bovine serum albumin (Sigma) for 30 min, and then incubated with TAGLN antibody (Thermo-Fisher Scientific, Rockford, IL, USA, CAT# PA5-29767) diluted in PBS (1:100) at 4°C overnight. After washing the cells with PBS, they were treated with secondary antibody (Goat polyclonal to anti-mouse and anti-rabbit IgG, Abcam) and incubated for 1 h at room temperature.

Cell proliferation assays

Cell number: Cell number was determined after 2 or 3 days of culture. The cells were cultured in T25 tissue culture flask dish at cell density 0.5×10^6 cells (28 000 cells/cm²). At confluence, the cells were trypsinized and counted manually with hemocytometer. The proliferation curve is shown as cell number at each passage against number of days in culture.

Alamar blue cell viability assay: Cell viability was measured using Alamar blue assay according to the manufacturer's recommendations (AbD Serotec, Raleigh, NC, USA). In brief, to cultured cells in 96-well plates 10 μl of Alamar blue substrate was added and the plates were incubated in the dark at 37°C for 1 h. Reading was subsequently taken using fluorescent mode (Ex 530 nm/Em 590 nm) using BioTek Synergy II microplate reader (BioTek Inc., Winooski, VT, USA).

RTCA cell proliferation assay: The RTCA DP system was used to measure the cell proliferation rate. Cells were cultured for 24 h in 1% FBS-DMEM, followed by adding 100 μl 10% FBS-DMEM to each well. E-plate 16 (ACEA Biosciences Inc., San Diego, CA, USA, ID: 691315, China) was placed onto PTCA DP analyzer inside the 37°C incubator. After 1 h, the background measurements were performed in RTCA SW. This is followed by adding 5000 cells in 100 μl 10% FBS-media per well to each well, and CIM plates containing the cells were placed in the DP system inside the incubator followed by measurements with 15 min intervals.

Scratch assay. This assay was conducted as previously described.⁴⁴ Overall, 0.2×10^6 cells/dish were plated on 30-mm cell culture dish. At confluence, the cell monolayer was scraped in a straight line to create a 'scratch' with a p10 pipet head. The debris was removed and markings to be used as reference points close to the scratch were created by tip marker, and the plates were placed under the phase-contrast microscope. Images of the scratch were acquired, and then the plates were placed in the tissue culture incubator. After incubation, the plates were examined with a phase-contrast microscope, and the reference points were matched. The previously acquired photographed regions were aligned and additional images were

acquired. To determine the rate of cell migration, lines were drawn along the edges of the scratch and the percentage of migrated cells at each time point was estimated.

RTCA cell migration assay. RTCA migration assay measures the effect of perturbations in electrical impedance in a label-free real-time setting. Migration assay was applied using DIC device with the RTCA DP system using FBS as a positive chemoattractant control. After 24 h incubation in 1% FBS-DMEM, the bottom chamber wells of the RTCA Cim-16 plate (ACEA Biosciences Inc., ID: 368179) were filled with 160 μl media (for each sample two wells were used, one filled with 1% FBS-media as a control and the other filled with 10% FBS-media as chemoattractant). The upper chamber was placed over the bottom one and filled with 25–50 μl 1% FBS-media to cover the membrane surface. CIM plate was placed on PTCA DP analyzer inside the 37°C incubator for 1 h incubation to allow the CIM membrane surface to reach the equilibrium with the media. After 1 h the background measurement was performed in RTCA SW. Cells (40 000) in 100 μl 1% FBS-media were added to each well of the top chamber. The CIM plates containing the cells were placed in the DP system inside the incubator, followed by the measurement with 15 min intervals.

Western blotting. Whole-cell lysates were prepared as previously described. Soluble proteins were analyzed by immunoblotting with TAGLN antibody (Thermo-Fisher Scientific, CAT# PA5-29767, diluted 1 : 50 00), total SMAD, P-SMAD2 (Cell Signaling, Boston, MA, USA, Cat. No. 9523, diluted 1:500), and anti- β -ACTIN (Sigma, A3854, diluted 1:10 000). Reactivity was detected with horseradish peroxidase-conjugated secondary antibodies (Santa Cruz Biotechnology, Dallas, TX, USA) and Clarity western ECL substrate (Bio-Rad, Hercules, CA, USA) for chemiluminescence using C-Digit Blot Scanner (LI-COR).

High content imaging using Operetta. Cells were plated in 96-cell carrier microtiter plates (Perkin Elmer) at 1500 cells/well in standard culture medium supplemented with 10% FCS at 37°C . After 24 h, the cells were fixed in 4% paraformaldehyde for 10 min, washed with PBS and stained for F-actin with Phalloidin-FITC (Sigma) and for nuclear staining with DAPI. Fluorescent images were analyzed using the Operetta high content imaging system (Perkin Elmer, Waltham, MA, USA) at a $\times 20$ magnification. Different parameters of cytoskeletal changes were measured by using Harmony High Content Imaging and Analysis Software (Perkin Elmer).

Transmission electron microscopy. After trypsinization, the cells were washed with PBS and the cell pellets were resuspended in 2.5% glutaraldehyde fixative (Electron Microscopy Sciences, Hatfield, PA, USA, Cat. No. 16500) in 0.1 M phosphate buffer (pH 7.2) and kept at 4°C for 4 h. The cells were washed in 0.1 M phosphate buffer (pH 7.2) and transferred to 1% osmium tetroxide (OsO_4) in 0.1 M phosphate buffer (pH 7.2). The cells were dehydrated in ascending grades of ethanol (10, 30, 50, 70, 90, and 100%) for 15 min each. The cells were then resuspended in acetone for 15 min and were aliquoted into BEEM embedding capsules and infiltrated with an acetone: resin mixture. Polymerization of the resin was accomplished in an oven at 70°C for 12 h. Semi-thin sections (0.5 μm thickness) were prepared and stained with 1% Toluidine Blue. Ultrathin sections (70 nm thickness) were prepared and mounted on copper grids. Ultrathin sections were first contrasted with uranyl acetate (saturated ethanol solution) for 30 min, rinsed, contrasted with Reynold's lead citrate for 5 min, and finally rinsed with distilled water. The contrasted ultrathin sections were examined and photographed under a TEM (Jeol 1010, Jeol, Tokyo, Japan).⁶⁰

In vivo subcutaneous implantation studies. All animal experimental procedures were approved by the animal care and use committees of the University of South Denmark. Cells were harvested via trypsinization, washed in PBS, and resuspended in PBS. Approximately 5×10^5 cells were mixed with 40 mg of hydroxyapatite-tricalcium phosphate ceramic powder per each implant (HA/TCP, Zimmer Scandinavia, Albertslund, Denmark) and implanted subcutaneously into the dorsal surface of 8-week-old female NOD/SCID mice (NOD/LtSz-Prkdcscid), as described in more detail previously.⁶¹ Three mice were used with two implants in each mouse. After 14 weeks, the implants were recovered, fixed in 4% paraformaldehyde, decalcified using formic acid solution (0.4 M formic acid and 0.5 M sodium formate), and embedded in paraffin.

Histologic analyses. Tissue blocks were sectioned at 4 mm. Immunohistochemical staining was performed on implants using DAKO En Vision and PowerVision according to the manufacturer's instructions (DAKO, Glostrup, Denmark). Human-specific staining was performed for human Vimentin. Briefly, paraffin sections were incubated for 1 h at room temperature with primary antibodies diluted in ChemMate (DAKO). Sections were washed subsequently in Tris-buffered saline (TBS, 0.05 M, pH 7.4), incubated for 30 min with secondary anti-mouse Ig/HR-conjugated polymers (K4001, En Vision., DAKO), and visualized with 3.30 diaminobenzidine tetrahydrochloride (DAB, S3000, DAKO) according to the manufacturer's instruction. Controls were performed without addition of primary antibodies and processed under identical conditions.

Image analysis methodology. High-resolution whole-slide digital scans of all histological sections were created with a ScanScope scanner (Aperio Technologies Inc., Buffalo Grove, IL, USA). Five images were captured for each section. The digital slide images were examined using the Aperio's ImageScope viewing software (Aperio Technologies Inc.). Random snapshots were taken from each section with $\times 10$ objective magnification. Each snapshot measured $1\,809\,171.65\ \mu\text{m}^2$ ($\sim 1.8\ \text{mm}^2$). The images were then subjected to image analysis using the ImageJ software (National Institute of Health). Each image was color-thresholded so as to select only the pink color of osteoid tissue (as stained with hematoxylin and eosin) and measured its area percentage relative to the total area of the image. The analysis output results were then exported to Excel sheets and subjected to statistical analysis.

DNA microarray global gene expression profiling. Total RNA was extracted using PureLink RNA Mini Isolation Kit (Ambion by Life Technologies, USA, Cat. No.: 12183018A) as recommended by the manufacturer. One hundred and fifty nanograms of total RNA were labeled and then hybridized to the Agilent Human SurePrint G3 Human GE $8 \times 60\ \text{k} \times 16$ microarray chip (Agilent Technologies, Santa Clara, CA, USA). All microarray experiments were conducted at the Microarray Core Facility (Stem Cell Unit, King Saud University College of Medicine). Normalization and data analyses were conducted using the GeneSpring GX software (Agilent Technologies). Pathway analysis was conducted using the Single Experiment Pathway analysis feature in GeneSpring 12.0 (Agilent Technologies) as described in (ref. 5). Twofold cutoff with $P < 0.02$ was used.

Quantitative real-time PCR (qRT-PCR). Total RNA was extracted using the PureLink Kit (Ambion by Life Technologies, Cat. No.: 12183018A) as recommended by the manufacturer. Total RNA was quantified using Nanodrop spectrophotometer (Nanodrop 2000, ThermoScientific). Complementary DNA (cDNA) was synthesized from $1\ \mu\text{g}$ of the RNA using High Capacity cDNA Reverse Transcription Kit (Applied Biosystem (ABI), USA) using Labnet, Multigene thermocycler according to the manufacturer's instructions. Relative levels of mRNA were determined from cDNA using real-time PCR (ABI-Real-Time PCR Detection System) with Power SYBR Green PCR Kit (ABI, UK), or with TaqMan Universal master Mix II, no UNG (ABI) according to the manufacturer's instructions. Following normalization to the reference gene glyceraldehyde 3-phosphate dehydrogenase (GAPDH), quantification of gene expression was carried out using a comparative Ct method where ΔCT is the difference between the CT values of the target and reference gene. Primers employed are listed in Supplementary Tables A and B.

Statistical analysis. All of the results were presented as the mean and S.D. of at least three independent experiments. Student's *t*-test was used for testing differences between groups. *P*-values < 0.05 was considered statistically significant.

Conflict of Interest

The authors declare no conflict of interest.

Acknowledgements. These studies are supported by a grant from the College of Medicine Research Centre (CMRC), Deanship of Scientific Research, King Saud University, Riyadh, Saudi Arabia.

- immortalized by human telomerase reverse transcriptase gene despite [corrected] extensive proliferation. *Biochem Biophys Res Commun* 2005; **326**: 527–538.
- Abdallah BM, Al-Shammary A, Skagen P, Abu Dawud R, Adjaye J, Aldahmash A et al. CD34 defines an osteoprogenitor cell population in mouse bone marrow stromal cells. *Stem Cell Res* 2015; **15**: 449–458.
 - Abdallah BM, Jafari A, Zaher W, Qiu W, Kassem M. Skeletal (stromal) stem cells: an update on intracellular signaling pathways controlling osteoblast differentiation. *Bone* 2015; **70**: 28–36.
 - Al-Nbaheen M, Vishnubalaji R, Ali D, Bouslimi A, Al-Jassir F, Megges M et al. Human stromal (mesenchymal) stem cells from bone marrow, adipose tissue and skin exhibit differences in molecular phenotype and differentiation potential. *Stem Cell Rev* 2013; **9**: 32–43.
 - Hamam D, Ali D, Vishnubalaji R, Hamam R, Al-Nbaheen M, Chen L et al. microRNA-320/RUNX2 axis regulates adipocytic differentiation of human mesenchymal (skeletal) stem cells. *Cell Death Dis* 2014; **5**: e1499.
 - Yu H, Königshoff M, Jayachandran A, Handley D, Seeger W, Kaminski N et al. Transgelin is a direct target of TGF-beta/Smad3-dependent epithelial cell migration in lung fibrosis. *FASEB J Off Publ Fed Am Soc Exp Biol* 2008; **22**: 1778–1789.
 - Sheppard D. Transforming growth factor beta: a central modulator of pulmonary and airway inflammation and fibrosis. *Proc Am Thorac Soc* 2006; **3**: 413–417.
 - Lawson D, Harrison M, Shapland C. Fibroblast transgelin and smooth muscle SM22alpha are the same protein, the expression of which is down-regulated in many cell lines. *Cell Motil Cytoskeleton* 1997; **38**: 250–257.
 - Li L, Miano JM, Cserjesi P, Olson EN. SM22 alpha, a marker of adult smooth muscle, is expressed in multiple myogenic lineages during embryogenesis. *Circ Res* 1996; **78**: 188–195.
 - Untergasser G, Gander R, Lilg C, Lepperding G, Plas E, Berger P. Profiling molecular targets of TGF-beta1 in prostate fibroblast-to-myofibroblast transdifferentiation. *Mech Ageing Dev* 2005; **126**: 59–69.
 - Sinha S, Hoofnagle MH, Kingston PA, McCanna ME, Owens GK. Transforming growth factor-beta1 signaling contributes to development of smooth muscle cells from embryonic stem cells. *Am J Physiol Cell Physiol* 2004; **287**: C1560–C1568.
 - Cevallos M, Riha GM, Wang X, Yang H, Yan S, Li M et al. Cyclic strain induces expression of specific smooth muscle cell markers in human endothelial cells. *Differ Res Biol Divers* 2006; **74**: 552–561.
 - Zhang JC, Kim S, Helmke BP, Yu WW, Du KL, Lu MM et al. Analysis of SM22alpha-deficient mice reveals unanticipated insights into smooth muscle cell differentiation and function. *Mol Cell Biol* 2001; **21**: 1336–1344.
 - Zeidan A, Swärd K, Nordström I, Ekblad E, Zhang JCL, Parmacek MS et al. Ablation of SM22alpha decreases contractility and actin contents of mouse vascular smooth muscle. *FEBS Lett* 2004; **562**: 141–146.
 - Peng J, Zhang Q, Ma Y, Wang Y, Huang L, Zhang P et al. A rat-to-human search for proteomic alterations reveals transgelin as a biomarker relevant to colorectal carcinogenesis and liver metastasis. *Electrophoresis* 2009; **30**: 2976–2987.
 - Maeda S, Hayashi M, Komiya S, Imamura T, Miyazono K. Endogenous TGF-beta signaling suppresses maturation of osteoblastic mesenchymal cells. *EMBO J* 2004; **23**: 552–563.
 - Lin Y, Buckhaults PJ, Lee JR, Xiong H, Farrell C, Podolsky RH et al. Association of the actin-binding protein transgelin with lymph node metastasis in human colorectal cancer. *Neoplasia N Y N* 2009; **11**: 864–873.
 - Zhang J, Song M-Q, Zhu J-S, Zhou Z, Xu Z-P, Chen W-X et al. Identification of differentially expressed proteins between early submucosal non-invasive and invasive colorectal cancer using 2D-DIGE and mass spectrometry. *Int J Immunopathol Pharmacol* 2011; **24**: 849–859.
 - Li S-Y, An P, Cai H-Y, Bai X, Zhang Y-N, Yu B et al. Proteomic analysis of differentially expressed proteins involving in liver metastasis of human colorectal carcinoma. *Hepatobiliary Pancreat Dis Int HBPD INT* 2010; **9**: 149–153.
 - Chen S, Kulik M, Lechleider RJ. Smad proteins regulate transcriptional induction of the SM22alpha gene by TGF-beta. *Nucleic Acids Res* 2003; **31**: 1302–1310.
 - Qiu P, Feng XH, Li L. Interaction of Smad3 and SRF-associated complex mediates TGF-beta1 signals to regulate SM22 transcription during myofibroblast differentiation. *J Mol Cell Cardiol* 2003; **35**: 1407–1420.
 - Massagué J, Wotton D. Transcriptional control by the TGF-beta/Smad signaling system. *EMBO J* 2000; **19**: 1745–1754.
 - Jian H, Shen X, Liu I, Semenov M, He X, Wang X-F. Smad3-dependent nuclear translocation of beta-catenin is required for TGF-beta1-induced proliferation of bone marrow-derived adult human mesenchymal stem cells. *Genes Dev* 2006; **20**: 666–674.
 - Erlebacher A, Filvaroff EH, Ye JQ, Derynck R. Osteoblastic responses to TGF-beta during bone remodeling. *Mol Biol Cell* 1998; **9**: 1903–1918.
 - Filvaroff E, Erlebacher A, Ye J, Gitelman SE, Lotz J, Heilman M et al. Inhibition of TGF-beta receptor signaling in osteoblasts leads to decreased bone remodeling and increased trabecular bone mass. *Dev Camb Engl* 1999; **126**: 4267–4279.
 - Wang D, Park JS, Chu JSF, Krakowski A, Luo K, Chen DJ et al. Proteomic profiling of bone marrow mesenchymal stem cells upon transforming growth factor beta1 stimulation. *J Biol Chem* 2004; **279**: 43725–43734.
 - Lian JB, Stein GS. Development of the osteoblast phenotype: molecular mechanisms mediating osteoblast growth and differentiation. *Iowa Orthop J* 1995; **15**: 118–140.
 - Bonewald LF, Mundy GR. Role of transforming growth factor-beta in bone remodeling. *Clin Orthop* 1990; **250**: 261–276.

1. Abdallah BM, Haack-Sørensen M, Burns JS, Elsnab B, Jakob F, Hokland P et al. Maintenance of differentiation potential of human bone marrow mesenchymal stem cells

29. Tang Y, Wu X, Lei W, Pang L, Wan C, Shi Z *et al*. TGF-beta1-induced migration of bone mesenchymal stem cells couples bone resorption with formation. *Nat Med* 2009; **15**: 757–765.
30. Shapland C, Hsuan JJ, Totty NF, Lawson D. Purification and properties of transgelin: a transformation and shape change sensitive actin-gelling protein. *J Cell Biol* 1993; **121**: 1065–1073.
31. Shapland C, Lowings P, Lawson D. Identification of new actin-associated polypeptides that are modified by viral transformation and changes in cell shape. *J Cell Biol* 1988; **107**: 153–161.
32. Small JV, Gimona M. The cytoskeleton of the vertebrate smooth muscle cell. *Acta Physiol Scand* 1998; **164**: 341–348.
33. Chen L, Shi K, Frary CE, Ditzel N, Hu H, Qiu W *et al*. Inhibiting actin depolymerization enhances osteoblast differentiation and bone formation in human stromal stem cells. *Stem Cell Res* 2015; **15**: 281–289.
34. Yourek G, Hussain MA, Mao JJ. Cytoskeletal changes of mesenchymal stem cells during differentiation. *ASAIO J Am Soc Artif Intern Organs* 1992 2007; **53**: 219–228.
35. Lomri A, Marie PJ, Escurat M, Portier MM. Cytoskeletal protein synthesis and organization in cultured mouse osteoblastic cells. Effects of cell density. *FEBS Lett* 1987; **222**: 311–316.
36. Fan JY, Carpentier JL, van Obberghen E, Grunfeld C, Gorden P, Orci L. Morphological changes of the 3T3-L1 fibroblast plasma membrane upon differentiation to the adipocyte form. *J Cell Sci* 1983; **61**: 219–230.
37. Antras J, Hilliou F, Redziniak G, Pairault J. Decreased biosynthesis of actin and cellular fibronectin during adipose conversion of 3T3-F442A cells. Reorganization of the cytoarchitecture and extracellular matrix fibronectin. *Biol Cell Auspices Eur Cell Biol Organ* 1989; **66**: 247–254.
38. Talele NP, Fradette J, Davies JE, Kapus A, Hinz B. Expression of α -smooth muscle actin determines the fate of mesenchymal stromal cells. *Stem Cell Rep* 2015; **4**: 1016–1030.
39. Sakai D, Kii I, Nakagawa K, Matsumoto HN, Takahashi M, Yoshida S *et al*. Remodeling of actin cytoskeleton in mouse periosteal cells under mechanical loading induces periosteal cell proliferation during bone formation. *PLoS One* 2011; **6**: e24847.
40. Harmey D, Stenbeck G, Nobes CD, Lax AJ, Grigoriadis AE. Regulation of osteoblast differentiation by *Pasteurella multocida* toxin (PMT): a role for Rho GTPase in bone formation. *J Bone Miner Res Off J Am Soc Bone Miner Res* 2004; **19**: 661–670.
41. Hamidouche Z, Fromigué O, Ringe J, Häupl T, Vaudin P, Pagès J-C *et al*. Priming integrin $\alpha 5$ promotes human mesenchymal stromal cell osteoblast differentiation and osteogenesis. *Proc Natl Acad Sci USA* 2009; **106**: 18587–18591.
42. Mathews S, Bhonde R, Gupta PK, Totey S. Extracellular matrix protein mediated regulation of the osteoblast differentiation of bone marrow derived human mesenchymal stem cells. *Differ Res Biol Divers* 2012; **84**: 185–192.
43. Park JS, Chu JS, Tsou AD, Diop R, Tang Z, Wang A *et al*. The effect of matrix stiffness on the differentiation of mesenchymal stem cells in response to TGF- β . *Biomaterials* 2011; **32**: 3921–3930.
44. Liang C-C, Park AY, Guan J-L. *In vitro* scratch assay: a convenient and inexpensive method for analysis of cell migration *in vitro*. *Nat Protoc* 2007; **2**: 329–333.
45. Karaöz E, Okçu A, Gacar G, Sağlam O, Yürüker S, Kenar H. A comprehensive characterization study of human bone marrow mscs with an emphasis on molecular and ultrastructural properties. *J Cell Physiol* 2011; **226**: 1367–1382.
46. Pietilä M, Palomäki S, Lehtonen S, Ritamo I, Valmu L, Nystedt J *et al*. Mitochondrial function and energy metabolism in umbilical cord blood- and bone marrow-derived mesenchymal stem cells. *Stem Cells Dev* 2012; **21**: 575–588.
47. Palomäki S, Pietilä M, Laitinen S, Pesälä J, Sormunen R, Lehenkari P *et al*. HIF-1 α is upregulated in human mesenchymal stem cells. *Stem Cells Dayt Ohio* 2013; **31**: 1902–1909.
48. Lu L, Ladinsky MS, Kirchhausen T. Cisternal organization of the endoplasmic reticulum during mitosis. *Mol Biol Cell* 2009; **20**: 3471–3480.
49. Friedman JR, Voeltz GK. The ER in 3D: a multifunctional dynamic membrane network. *Trends Cell Biol* 2011; **21**: 709–717.
50. Shih Y-RV, Tseng K-F, Lai H-Y, Lin C-H, Lee OK. Matrix stiffness regulation of integrin-mediated mechanotransduction during osteogenic differentiation of human mesenchymal stem cells. *J Bone Miner Res Off J Am Soc Bone Miner Res* 2011; **26**: 730–738.
51. Treiser MD, Yang EH, Gordonov S, Cohen DM, Androulakis IP, Kohn J *et al*. Cytoskeleton-based forecasting of stem cell lineage fates. *Proc Natl Acad Sci USA* 2010; **107**: 610–615.
52. Salasznyk RM, Klees RF, Williams WA, Boskey A, Plopper GE. Focal adhesion kinase signaling pathways regulate the osteogenic differentiation of human mesenchymal stem cells. *Exp Cell Res* 2007; **313**: 22–37.
53. Chen CS, Alonso JL, Ostuni E, Whitesides GM, Ingber DE. Cell shape provides global control of focal adhesion assembly. *Biochem Biophys Res Commun* 2003; **307**: 355–361.
54. Mathieu PS, Lobo EG. Cytoskeletal and focal adhesion influences on mesenchymal stem cell shape, mechanical properties, and differentiation down osteogenic, adipogenic, and chondrogenic pathways. *Tissue Eng Part B Rev* 2012; **18**: 436–444.
55. Kawamura N, Kugimiya F, Oshima Y, Ohba S, Ikeda T, Saito T *et al*. Akt1 in osteoblasts and osteoclasts controls bone remodeling. *PLoS One* 2007; **2**: e1058.
56. Xue X, Wang X, Liu Y, Teng G, Wang Y, Zang X *et al*. SchA-p85-FAK complex dictates isoform-specific activation of Akt2 and subsequent PCBP1-mediated post-transcriptional regulation of TGF β -mediated epithelial to mesenchymal transition in human lung cancer cell line A549. *Tumour Biol J Int Soc Oncodevelopmental Biol Med* 2014; **35**: 7853–7859.
57. Hildebrand A, Romaris M, Rasmussen LM, Heinegård D, Twardzik DR, Border WA *et al*. Interaction of the small interstitial proteoglycans biglycan, decorin and fibromodulin with transforming growth factor beta. *Biochem J* 1994; **302** (Pt 2): 527–534.
58. Serra R, Chang C. TGF-beta signaling in human skeletal and patterning disorders. *Birth Defects Res* 2003; **69**: 333–351.
59. Gregory CA, Gunn WG, Peister A, Prockop DJ. An Alizarin red-based assay of mineralization by adherent cells in culture: comparison with cetylpyridinium chloride extraction. *Anal Biochem* 2004; **329**: 77–84.
60. Ahmad R, Raina D, Trivedi V, Ren J, Rajabi H, Kharbanda S *et al*. MUC1 oncoprotein activates the I κ B kinase beta complex and constitutive NF-kappaB signalling. *Nat Cell Biol* 2007; **9**: 1419–1427.
61. Abdallah BM, Ditzel N, Kassem M. Assessment of bone formation capacity using *in vivo* transplantation assays: procedure and tissue analysis. *Methods Mol Biol Clifton NJ* 2008; **455**: 89–100.



Cell Death and Disease is an open-access journal published by **Nature Publishing Group**. This work is licensed under a **Creative Commons Attribution 4.0 International License**. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2016

Supplementary Information accompanies this paper on Cell Death and Disease website (<http://www.nature.com/cddis>)