Enhanced Extracellular Matrix Breakdown Characterizes the Early Distraction Phase of Canine Knee Joint Distraction

CARTILAGE 2021, Vol. 13(Suppl 2) 1654S–1664S © The Author(s) 2021

Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/19476035211014595 journals.sagepub.com/home/CAR

SAGE

Michelle Teunissen^{1*}, Alberto Miranda Bedate^{1*}, Katja Coeleveld², Frank M. Riemers¹, Björn P. Meij¹, Floris P. J. G. Lafeber², Marianna A. Tryfonidou^{1*}, and Simon C. Mastbergen^{2*}

Abstract

Objective. Joint distraction triggers intrinsic cartilage repair in animal models of osteoarthritis (OA), corroborating observations in human OA patients treated with joint distraction. The present study explores the still largely elusive mechanism initiating this repair process. Design. Unilateral OA was induced in the knee joint of 8 dogs using the groove model; the contralateral joint served as a control. After 10 weeks, 4 animals received joint distraction, the other 4 serving as OA controls. Halfway the distraction period (after 4 weeks of a standard 8-week distraction treatment), all animals were euthanized, and joint tissues were collected. A targeted quantitative reverse transcription polymerase chain reaction (qRT-PCR) analysis was performed of commonly involved processes including matrix catabolism/anabolism, inflammation, and known signaling pathways in OA. In addition, cartilage changes were determined on tissue sections using the canine OARSI (Osteoarthritis Research Society International) histopathology score and collagen type II (COL2A1) immunostaining. *Results.* Midway distraction, the distracted OA joint showed an upregulation of proteolytic genes, for example, *ADAMTS5*, *MMP9*, *MMP13*, compared to OA alone and the healthy joints, which correlated with an increased OARSI score. Additionally, genes of the transforming growth factor (TGF)- β and Notch pathway, and markers associated with progenitor cells were increased. *Conclusions.* Joint distraction initiates both catabolic and anabolic transcriptional responses. The enhanced turnover, and thereby renewal of the matrix, could be the key to the cartilage repair observed in the months after joint distraction.

Keywords

cartilage regeneration, groove model, osteoarthritis

Introduction

Osteoarthritis (OA) is a progressive joint disease characterized by inflammation and structural changes of the joint, causing pain and functional disability. The prevalence of knee OA approaches 5% of the global population, and is expected to rise due to increased age and prevalence of obesity of the population.¹⁻³ This will significantly affect societal health and economic costs. For patients with significant joint damage and severe OA symptoms despite conservative therapy, knee arthroplasty is considered an effective therapy. However, when patients are relatively young (<60 years of age), the prostheses' limited life span brings a greater risk of a future revision surgery.^{4,5} Therefore, there is a need for alternative treatment strategies that can delay, or even prevent, knee arthroplasty. Within this context, joint distraction has been proposed as a joint preserving treatment strategy. During joint distraction, the 2 bony ends of a joint are temporarily (6-9 weeks) distracted using an external fixation frame. The clinical application and efficacy of knee joint

¹Department of Clinical Sciences, Faculty of Veterinary Medicine, Utrecht University, Utrecht, The Netherlands ²Rheumatology & Clinical Immunology, UMC Utrecht, Utrecht University, Utrecht, The Netherlands

*Shared first author and shared last author.

Supplementary material for this article is available on the *Cartilage* website at https://journals.sagepub.com/home/car.

Corresponding Author:

Simon C. Mastbergen, Rheumatology & Clinical Immunology, UMC Utrecht, Utrecht University, G02.228, PO Box 85500, GA, Utrecht 3508, The Netherlands. Email: S.Mastbergen@umcutrecht.nl distraction (KJD) have been reviewed,⁶⁻¹⁰ and although the number of studies is limited and the sample size relatively small, there is evidence for a prolonged clinical benefit.¹⁰⁻¹⁴

At the biological level, there are indications that joint distraction facilitates cartilage regenerative effects. This reparative activity is evaluated in clinical studies by surrogate markers such as imaging and (serum/urine) biochemical markers. The strongest tissue repair is observed at 1 and 2 years after distraction by radiographic and magnetic resonance imaging evaluation (both quantitative and qualitative, such as dGEMRIC and T2 relaxation), the latter demonstrating the presence of hyaline cartilage.^{11,12,15} In addition, increased serum levels of the collagen type II synthesis marker, PIIANP (N-propeptide of collagen IIA), and decreased levels of the collagen type II breakdown marker, CTXII (C-Telopeptide of type II collagen), at 1 and 2 years post distraction treatment, show an increased ratio of synthesis over breakdown in this period.^{11,16} After this period, the degenerative, progressive nature of OA takes over again, and this results in a gradual waning of the effect. However, especially considering the natural progression in case of only conservative treatment, there is still improvement after 5 to 10 years compared to pretreatment situation.^{13,14} Complementary, a canine OA model, in which OA was induced in a period of 10 weeks using the groove model,¹⁷ followed by 8 weeks of KJD, showed structural improvement of the cartilage at 25 weeks follow-up after distraction.¹⁸ This experimental study showed that OA-knee joints treated with KJD had improved macroscopic and histopathology OARSI scores (the canine Osteoarthritis Research Society International [OARSI] assessment system),¹⁹ higher proteoglycan (PG) content, better retention of newly formed PG and less collagen damage compared to the OA control knee joints after the same prolonged follow-up.¹⁸

Altogether, the aforementioned findings support the notion that KJD elicits a reparative response, but the underlying mechanisms of action during distraction remain elusive. Involvement of multiple mechanisms have been postulated, including the temporary absence of mechanical loading while preserving joint fluid pressure oscillation, enhanced periarticular bone turnover, and/or stem cell modulation as a result of joint distraction.^{11,20-23} In order to further strengthen these existing hypotheses, the present study explored the initial transcriptional response of the cartilage and adjacent joint tissues during KJD treatment in the groove model, a canine OA model. More specifically, genes related to cartilage matrix turnover and bone remodeling, (cartilage) progenitor cell markers, cytokines, and signaling pathways involved in OA were investigated. As the first (bio)molecular changes are thought to start already during the treatment with joint distraction, a time point halfway (4 weeks) the common 8-week distraction period was selected for the analysis.

Methods

Animal Procedures

Skeletally mature Mongrel dogs (n = 8 females, mean \pm standard deviation [SD] age 29 ± 7.6 months, mean weight 23 ± 2.8 kg) were used. Upon ethical approval by the local ethics committee on animal experimentation (2013. III.08.054), OA was induced in all dogs unilaterally (the right knee joint) according to the groove model.¹⁷ Grooves were applied using a Kirschner-wire (1.5-mm diameter) bent 0.4 mm from the top at 90° to ensure that the depth of the grooves was restricted to the cartilage depth and not to the subchondral bone. The contralateral, left knee joint served as healthy control (control) without further treatment. Ten weeks post OA induction, dogs were randomly divided into 2 groups of 4 animals. One group received no additional treatment (OA), while the other group received KJD (distraction). Briefly, to initiate KJD, bone pins were drilled into the femur and tibia and connected to external fixation frames in a 3-point fixation with the use of commercially available connectors. Subsequently, the external fixation frames on the femur and the tibia were connected by hinges medially and laterally of the knee joint. Distraction of the joint was carried out by extending the connecting rods and was visualized by fluoroscopy using a C-arm, while smooth motion of the joint during flexion and extension was maintained. Halfway the commonly used distraction period of 8 weeks, thus after 4 weeks of joint distraction, all 8 animals were euthanized, and material was collected for further analysis. A full description of experimental procedures can be found in the Supplementary Material.

Collection of Material Postmortem and Tissue Processing

Within 1 hour after euthanasia, high-resolution photographs of the joint surfaces were obtained for macroscopic grading of cartilage damage after which tissues where processed. Cartilage tissue, excluding any subchondral bone, was collected from the weight-bearing area of the femoral condyles and tibial plateaus and processed for 2 purposes: fixed in 4% phosphate-buffered formalin containing 2% sucrose (pH 7.0) for (immuno)histochemistry, and snap frozen for RNA isolation. Additionally, tissue samples of fat pad, suprapatellar synovium, meniscus, and tibial and femoral subchondral bone samples were collected and snap frozen for RNA isolation.

Cartilage Quality Assessment and Immunohistochemistry

Cartilage damage was macroscopically graded (2 observers; FPL, SCM) and microscopically graded on Safranin-O/Fast

green stained sections (3 observers; SCM, MAT, and AMB) according to the OARSI canine scoring system.¹⁹ All samples were randomized and observers were blinded for the source of material studied. Data are provided as mean OARSI score \pm SD. Furthermore, immunopositivity for collagen type-1 (COL1A1), -2 (COL2A1), and -10 (COLX) of the cartilage matrix was evaluated (Supplementary Material).

Transcriptional Profiling

Tissue samples were reduced to powder (cartilage, meniscus, subchondral bone) and/or submitted to a short Tissuelyser (Qiagen) cycle (25 shakes/second for 4 minutes; fat pad, suprapatellar synovium). Thereafter, total RNA was extracted using the miRCURY RNA Isolation Kit (Exiqon, Vedbaek, Denmark) for cartilage and the RNeasy Mini Kit (Qiagen, Venlo, the Netherlands) for the other tissues, according to the manufacturer's instructions with an additional on column DNase treatment (Qiagen). RNA quality and quantity were measured with a Bioanalyzer (Nano-chip, Agilent Technologies, Amstelveen, the Netherlands). cDNA was produced using the iScriptTM cDNA Synthesis Kit (Bio-Rad, Veenendaal, the Netherlands) with a similar RNA input for all samples following manufacturer's instructions.

Total RNA was profiled with a panel of 63 genes by quantitative reverse transcriptase polymerase chain reaction (qRT-PCR; Supplementary Material). Investigated targets and/or pathways included genes related to cartilage matrix metabolism, bone remodeling, the TGF/BMP pathway,²⁴ the IGF pathway,²⁵ the Notch pathway,²⁶ the Wnt pathway,²⁷ the Indian Hedgehog pathway,27 and several cytokines, and progenitor cell associated markers.^{28,29} Based on initial analysis, additional target genes (n = 13) within these pathways were studied specifically for the cartilage tissue. Quantitative PCR was performed using a CFX384 Touch Real-Time PCR Detection System and IQ SYBR Green SuperMix (both from Biorad) according to the manufacturer's protocols. Standard curves consisted of 4-fold serial dilutions of the cDNA template. For each standard curve, the amplification efficiency was between 90% and 110%. The normalization of gene expression was performed with 7 reference genes: RPS19, SDHA, YWHAZ, TBP, RPS5, RPL13, and HPRT.

Statistical Analysis

 Δ Ct values, and OARSI histopathology scores, were statistically analyzed (R version 3.6.3,³⁰ RStudio version 1.2.5033³¹) for the 3 comparison groups (OA vs. control, distraction vs. OA, and distraction vs. control). Linear models were employed for the analysis of variance (ANOVA). For each parameter, the selection of random effects for the different linear models was performed, considering variables "donor" and "location" (tibial plateaus/femoral condyles [if applicable depending on the tissue]). If the variable "location" was considered a significant variable for the model, an additional analysis was run on the separated data of the tibial plateaus and femoral condyles of the cartilage and subchondral bone. Normality of the residuals, homoscedasticity, independence of errors, and the presence of outliers were assessed for each linear model. If any of the assumptions were not held, a power transformation of the dCt values with the lambda coefficient as exponent was performed, reassessing all the assumptions. If the assumptions were not passed, an exact Wilcoxon-Mann-Whitney test, which is a permutation based nonparametric test, was used. P values were subjected to corrections for multiple testing (Benjamini-Hochberg false discovery rate). Effect sizes (ES) and ES's 95% confident intervals (CI) were provided as Hedge's g for normally distributed data and Cliff's delta for nonnormally distributed data. In the event a specific gene could not be detected in 1 of the 2 groups, this was considered a biologically relevant difference in expression between groups. These comparisons were included as significant with a fold change >10. Specific details of the statistical analysis can be found in the Supplementary Material.

Results

Cartilage Integrity Is Still Deteriorated Midterm Distraction Treatment

Fourteen weeks after OA induction, the OA group (without distraction) clearly showed macroscopic cartilage damage in comparison to the control (OARSI score: femur 2.9 \pm 0.5 vs. $0.0 \pm 0.0, P < 0.005$; tibia 1.0 ± 0.4 vs. $0.0 \pm 0.0, P <$ 0.015; Fig. 1). A similar degree of cartilage damage was found in the distraction group (femur 2.5 \pm 0.4, P < 0.015; tibia 1.3 ± 0.3 , P < 0.005 [vs. control]); OA and distraction groups did not differ (Fig. 1). These macroscopic observations were confirmed by histological analysis. The average histopathology OARSI score was significantly higher in the OA group compared to control joints (femur 6.8 ± 2.3 vs. 3.0 \pm 2.4, P < 0.02; tibia 10.6 \pm 2.4 vs. 4.8 \pm 3.0, P < 0.02; Fig. 2B). The distraction group showed on average a higher histopathology OARSI score compared to the OA condition (femur 9.8 \pm 1.9, P = 0.08, with a very large ES; tibia 13.5 \pm 4.4, P = 0.2, with a large ES; Fig. 2B, Supplementary Material). COL1A1 and COLX proteins were undetectable in the cartilage tissues (Supplementary Material). A loss of COL2A1 staining into the intermediate cartilage layer was observed in the distraction and OA group (Fig. 2A).

Gene Expression Profiling of the OA Cartilage and OA Subchondral Bone Shows Minimal Changes at the Transcriptional Level

Taken together, the most differentially expressed (DE) genes were detected in cartilage and subchondral bone



Figure 1. Macroscopic cartilage damage assessment at midterm (4 weeks) of the distraction treatment. (**A**) Macroscopic photographs of canine cartilage after 4 weeks of the distraction for tibial plateaus (Tibia) and femoral condyles (Femur) in the experimental groups: control cartilage (Healthy), OA cartilage (OA), and distracted OA cartilage (Distraction). The grooves that were surgically applied were still visible (black arrowheads) with additional surrounding degeneration, while the condylar cartilage of the contralateral control knees was intact. (**B**) OARSI scoring of macroscopic cartilage after 4 weeks of distraction for femoral condyles (Femur) and tibial plateaus (Tibia) in control cartilage (Healthy), OA and OA distracted cartilage (Distraction). "Y" axes represent OARSI grade and "X" axes the experimental conditions. Asterisks indicate statistically significant differences compared to the healthy control (*P < 0.05; **P < 0.01).



Figure 2. Representative immunostainings and OARSI grading of the cartilage at midterm (4 weeks) distraction treatment. (**A**) Representative images of Safranin-O/Fast Green staining and collagen type II immunohistochemistry from control (healthy), osteoarthritic (OA), and OA + KJD (Distraction) joints after 4 weeks of distraction of the tibial plateaus (tibia) and femoral condyles (femur). Scale bar = 100 μ m. (S) = superficial layer; (I) = intermediate layer. (**B**) OARSI scoring of the histology after 4 weeks of distraction is given for all conditions, in cartilage from the femoral condyles and tibial plateaus. Asterisks indicate statistically significant differences between the indicated groups within the locations with at least medium effect sizes (*P < 0.05; **P < 0.01).



Figure 3. The transcriptional profile of cartilage, subchondral bone, synovium, meniscus, and fat pad. Cartilage and subchondral bone (Bone) registered the highest numbers of differentially expressed (DE-) genes from all analyzed tissues based on the corresponding DE-genes heat-map. Scale bars represent inverted, significant $\Delta\Delta$ Ct values in a color gradient (dark blue [highly downregulated gene], white [no differences], and dark red [highly upregulated gene]). Not significant DE-genes are depicted in gray. For each tissue, the heat map is divided in comparisons and anatomical locations. For the cartilage and subchondral bone the following anatomical locations were used: Whole joint (no differentiation between anatomical locations), Tibia (tibial plateaus), Femur (femoral condyles). The comparisons depicted on top of the heat map include: (1) osteoarthritic (OA) compared to healthy joints (OA vs. healthy [green comparison]), (2) distracted joints (KJD) compared to healthy joints (KJD vs. OA [lavender comparison]). Furthermore, DE-genes are divided in functional groups, characterized at the left side of heat map by colored boxes.

(Fig. 3). Genes with at least 2-fold difference were considered biologically relevant to report and discuss.

Compared to the control joints, 25% (19/76) DE-genes were detected in the OA cartilage, of which 10 showed more than a 2-fold change (**Fig. 3**). Interestingly, the cartilage matrix gene *COL2A1* was upregulated in the OA cartilage (*COL2A1*, P < 0.001), while catabolic genes, generally associated with OA, Matrix Metalloproteinase 13 (*MMP13*), andADisintegrin and Metalloproteinase with Thrombospondin Motifs 5 (*ADAMTS5*) did not differ significantly. Additionally, genes related to the hypertrophic differentiation *COLX*, Bone Gamma-Carboxyglutamate Protein (*BGLAP*), and Vascular Endothelial Growth Factor A (*VEGFA*) were significantly upregulated (P < 0.05). In the subchondral bone, only 9% (9/63) of the genes were significantly different, of which 7 were more than 2-fold regulated (**Fig. 3**). Noteworthy is the upregulation of *MMP13* (P < 0.01), while *ADAMTS5* was downregulated in the bone (P < 0.001). Furthermore, *COLX* (P < 0.05) and *COL1A1* (P < 0.05), as well as the inflammatory marker Prostaglandin E Synthase (*PTGES*)-1 (P < 0.05) were significantly upregulated.

In the synovium, 10% (6/63) of the genes were detected as DE-genes, all more than 2-fold different. Most of these genes were involved in the IGF pathway, including Growth Hormone Receptor (*GHR*), IGF 1 Receptor (*IGF1R*), and IGF Binding Proteins 2 and 5 (*IGFBP-2*, -5), and were significantly downregulated (P < 0.05) in OA. In addition, comparable with the bone, *ADAMTS5* was downregulated (P < 0.01). No significant DE-genes were found in the osteoarthritic fat pad and meniscus compared to the healthy joints.

Anabolic and Catabolic Transcriptional Responses Coincide in the Distracted Cartilage

In the distracted cartilage 58% (44/76) DE-genes were detected compared to the OA cartilage, of which 42 showed more than a 2-fold change. A clear catabolic transcriptional response was observed; *MMP9*, *MMP13*, and *ADAMTS5* were highly upregulated in the distracted cartilage (P < 0.001) compared to OA. Furthermore, the cartilage matrix genes Aggrecan (*ACAN*), *COL2A1*, Cartilage Oligomeric Matrix Protein (*COMP*), and the master chondrogenic regulator SRY-Box Transcription Factor 9 (*SOX9*) were severely downregulated (P < 0.001), while *COL1A1*, a marker of fibrocartilage, was upregulated (P < 0.001) compared to OA cartilage. Interestingly, although RUNX Family Transcription Factor 2 (*RUNX2*, P < 0.001), a marker of chondrocyte hypertrophy, was upregulated, *COLX* was downregulated (P < 0.001).

At the same time, putative anabolic transcriptional responses were found, including genes in the Notch pathway (upregulation of NOTCH1 [P < 0.001] and Hairy/ enhancer-of-split related with YRPW motif protein 1 [*HEY1*], P < 0.05), and the TGF- β pathway (upregulation of activin receptor-like kinase-1 [ALK1]; P < 0.0001), plasminogen activator inhibitor 1 (PAI1; P <0.0001) and the TGF- β receptor II (*TGF* β *RII*) specifically in the tibial plateaus (P < 0.05). Additionally, several cytokines were only expressed in distracted cartilage including interleukin 6 (IL-6), chemokine C-C motif ligand 2 (CCL2), and the anti-inflammatory cytokine IL-10 and its receptor, *IL-10R*, while they were undetectable in OA cartilage. Furthermore, several markers associated with progenitor cells were upregulated such as the surface markers CD105, CD90, CD166, and CD146 (all P <0.001).

Subchondral Bone of the Distracted Joined Showed Certain Coincidental Transcriptional Features with Cartilage

In the subchondral bone of the distracted joint, 26 (37%) DE-genes were found compared to the OA joint, of which 15 showed more than a 2-fold regulation. Noteworthy was the downregulation of *COL1A1* and *BGLAP* in the subchondral bone of the distracted joint compared to the OA joint (P < 0.001). Furthermore, in correspondence with the distracted cartilage, a downregulation of *ACAN* (P < 0.001) and *COL2A1* (P < 0.05) and an upregulation of *MMP9* (P < 0.01) was found.

Changes in the Synovium, Fat Pad, and Meniscus of the Distracted Joint Are Limited and Coincide with the Distracted Cartilage and Subchondral Bone

In the synovium of the distracted joint, only 4 DE-genes were detected that showed more than a 2-fold change compared to the OA joint. A significant upregulation of *MMP9* (P < 0.05) and *PAI1* (P < 0.05) was found as well as a significant upregulation of *IGFBP6* (P < 0.05). In the distracted joint, *COL2A1* was significantly downregulated in the meniscus compared to the OA condition (P < 0.05), and *MMP9* and *IL6* were only detected in distracted cartilage, coinciding with the changes in the cartilage and subchondral bone. In the fat pad, *MMP13* was upregulated in the distracted joint compared to the OA control.

Discussion

Although the clinical efficacy of joint distraction has been assessed, the underlying regenerative mechanisms behind distraction remain poorly understood. This study explores transcriptional regulation in all relevant joint tissues midway the distraction period. These unpresented results demonstrate that the regenerative response of KJD, 25 weeks after the 8-week distraction in a canine OA model,¹⁸ is fronted by an increased breakdown of the extracellular matrix (ECM) of the OA cartilage during the distraction phase. This is corroborated by an increased histological OARSI grade compared to the OA joint, with concomitant loss of collagen type II into the intermediate cartilage layer, and an increased expression of catabolic proteolytic genes midway distraction. At the same time, several transcriptional signals were detected compatible with cartilage regenerative responses, possibly constituting the initiation of cartilage repair activity that is seen at 25 weeks follow-up.18

The groove model has been shown to encompass hallmarks of progressive OA.^{32,33} In line with these previous reports, the present study revealed clear degenerative cartilage changes in the OA condition, at 14 weeks post OA induction,^{17,33} indicated by a loss of PG-rich matrix. At the transcription level, the present study demonstrated mild upregulation of matrix catabolic genes ADAMTS5 and MMP13, and hypertrophy like changes in chondrocytes with increased VEGFA and osteocalcin (BGLAP), known to play a role during OA.³⁴ ACAN and COL2A1 showed a higher expression compared with the control joints. This is in line with the increased proteoglycan synthesis reported in cartilage samples from the groove model at 10 weeks post OA induction,³² and other reports of the expression of ACAN and COL2A1 at early OA stages.35-37 This enhanced chondrocyte activity in OA cartilage is an attempt for repair considered to be ineffective, as the newly formed molecules are also lost at a higher rate, resulting in a net loss of tissue.38

This study demonstrates for the first time that joint distraction initiates mainly a catabolic response midway distraction, mostly concentrated in cartilage and subchondral bone as shown on histology and the coinciding transcriptional regulations. Joint distraction elicited a higher OARSI cartilage score compared to OA. This increased ECM degradation corresponded with the distinct upregulation of MMP9, MMP13, and ADAMTS5 and further reduction of their respective proteolytic targets, ACAN and COL2A1. The latter was further corroborated by a decrease of collagen type II staining into the intermediate cartilage layer. The observed imbalance between matrix anabolism and catabolism during distraction seems to be contra-intuitive in respect of the final outcome of distraction; several human and animal studies demonstrated the improvement in ECM quantity and quality.^{3-6,8} However, from animal and human studies, it is known that an absence of normal joint loading causes a reduction in PG content, a decreased PG synthesis, and thinning of the (calcified) cartilage.³⁹⁻⁴¹ Additionally, in rabbits, a 9-week distraction period of healthy knee joints resulted in degenerative changes in the articular cartilage similar to those in early OA.42 Nonetheless, Wiegant et al.18 employed the groove canine model in a similar fashion as in the present study and demonstrated 25 weeks after KJD a significant improvement of the histological OARSI grade compared to the untreated OA knee joints.

Hypothetically, the dominating catabolic stimuli during the unloading of the joint that deplete the ECM may allow for remodeling with healthy cartilaginous matrix on the long term. For example, if aggrecan molecules are enzymatically truncated, but not removed from the hyaluronic acid core in the process of OA, and with that from the matrix, that aggrecan molecule cannot be replaced, leaving an impaired aggrecan complex.⁴³ Only upon further degradation is the truncated molecule removed and fully replaced. Within this context, even though matrix catabolism seems to predominate at the transcriptional and protein level halfway the distraction phase, there are some important differences in the transcriptional response of the distracted compared to the OA cartilage. These differences are discussed below and could provide insight into several suitable regenerative mechanisms by which distraction could stimulate an intrinsic cartilage repair at a later stage.

One of these proposed mechanisms is the involvement of stem cells. There is increasing evidence for the presence of chondroprogenitor cells in cartilage, even in OA.²⁸ In this study, several markers were upregulated in distraction versus OA, such as CD105, CD166, Notch-1, CD90, and CD146, that are associated with chondroprogenitor cells.44-48 Whether this is because of an increased amount and/or activity of progenitor cells in the cartilage or because of the attraction and retention of mesenchymal stromal cells (MSC) from surrounding tissues remains to be further clarified. The latter has been proposed to be initiated by the intermittent fluid oscillations in the joint during distraction.⁴⁹ Recently, it was shown that unloading of the joint by KJD resulted in a significant increase in synovial fluid MSC (SF-MSC) colony size and density.⁵⁰ Furthermore, after 3 weeks of KJD treatment many transcriptional changes were found in these SF-MSC compared to baseline. These changes included a sustained upregulation of ACAN and a significant increase of the MSC chondrogenic commitment markers gremlin 1, and growth differentiation factor 5 (GDF5), markers associated with cartilage homeostasis and OA, among others.⁵⁰⁻⁵² In addition, the joint environment during KJD is changed, favoring attachment of MSC.²¹

Another proposed mechanism is the effect of the periarticular bone turnover on the cartilage, as a decrease in subchondral bone sclerosis has been reported in humans and rats after KJD, which was directly associated with the reported clinical improvement.^{20,23} The present study further corroborates these findings: in the distracted subchondral bone compared to the OA control reduced transcription of the majority of the investigated matrix genes was observed together with decreased *OPG* and *RANKL*, representatives of the RANKL/RANK/OPG pathway involved in bone remodeling. Noteworthy, and in line with concepts from the rheumatoid arthritis field,⁵³ *RANKL* was profoundly upregulated in the distracted cartilage and may mediate subchondral bone remodeling upon its diffusion.

Other pathways that emerged, and could provide clues for further research, were the IGF pathway, the notch pathway and the TGF/BMP pathway. TGF- β mediated signal transduction via the Smad2/3 pathway is generally thought to be a protective factor for cartilage,²⁴ while an increased ALK1/ALK5 ratio, promoting Smad1/5/8 pathway signal transduction, is associated with increased *MMP13* expression, an OA hallmark.⁵⁴ In the distracted cartilage, *ALK1* expression was upregulated compared to the OA control, corresponding with the upregulation of *MMP13* in the distracted joint. However, *PAI1*, the downstream mediator of the Smad2/3 pathway, and thereby the PAI1/ID1 ratio, was upregulated in the distracted cartilage. Furthermore, *TGF* β *RII* was upregulated in the tibial plateaus. This receptor has been associated with a chondroprotective role, as its expression is downregulated in human OA chondrocytes.²⁴ These findings coincide with the finding of the study of Watt *et al.*, which found an upregulation of TGF- β 1 in the synovial fluid of human patients after 6 weeks of KJD (directly after treatment) compared to baseline.⁵⁵ In this study, *TGF*- β 1 was also upregulated, though not statistically significant.

Finally, inflammatory processes during joint distraction may also be at play. IL-6, IL4 receptor (IL4R), IL-10, IL-10R were significant upregulated in the cartilage, and IL-6 was 4-fold upregulated in the synovial tissue of distracted joints compared to the OA joints. In line with these findings, an upregulation of IL-6 and CCL2, also referred to as monocyte chemoattractant protein 1 (MCP1), was also found in the synovial fluid of human patients after the 6-week KJD treatment compared to baseline.55 Together with the upregulation of IL-10, and the IL4- and 10 receptor, shown to have a chondroprotective and anti-inflammatory role,^{56,57} there seems to be involvement of multiple anabolic pathways that might initiate the reparative response generated by KJD on top of the (initial) clear catabolic activity. Upregulation of IL-10 in the blood and synovial fluid was also found in rabbits treated for 4 weeks with joint distraction and excercise.⁵⁸ Finally, a downregulation of *IL-1*β was found in the distracted cartilage and subchondral bone compared to the OA joints. Downregulation of IL-1ß was also reported by studies that investigated the effects of joint distraction in rats and rabbits,^{20,58} and also points toward a chondroprotective effect of joint distraction.

Some caution is warranted interpreting these results. This study contained only a small number of dogs per group and is therefore clearly exploratory. Dogs have been used before to study KJD,^{18,59} and provide many advantages. The anatomy of the dog knee, as well as the biochemical and histological characteristics of their cartilage and subchondral bone is similar to that of humans.¹⁹ In addition, canine OA progresses similar to that of humans and they do not show the spontaneous cartilage repair that is reported in rabbits.^{19,60} However, weight bearing of the limb during the distraction period could have been suboptimal as quadruped dogs are easily able to walk on 3 limbs. This could diminish the effect of the intermittent fluid changes proposed to elicit a beneficial effect on joint health.¹⁸ Importantly, KJD is a dynamic process and the information provided by transcriptional profiling is a static representation of a single time point without guaranteed translation to the protein level. An interesting approach for the future would be to investigate multiple follow-up time points to identify the catabolic-toanabolic turning point in the distracted joint.

Conclusion

This study showed for the first time that treatment of knee OA with joint distraction initiates catabolic as well as anabolic transcriptional responses. This results in a catabolic joint environment halfway during joint distraction, with aggravation of OA at the histological and transcriptional levels. This explorative study provides clues for future studies that focus on elucidating the mechanisms behind joint distraction, including the involvement of progenitor cells and the cross-talk between subchondral bone and cartilage, and the role of pro- and anti-inflammatory cytokines.

Acknowledgments and Funding

The authors would like to thank P. M. Roermund for his assistance during application of the distraction device in the *in vivo* experiment. The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This project was financially supported by the Dutch Arthritis Society (LLP9 and LLP22) and NWO Applied and Engineering Sciences (P15-23). There is no further involvement in the present work of the above-mentioned sources.

Declaration of Conflicting Interests

The author(s) declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: FPL is consultant as UMCU employee for Synerkine Pharma BV and cofounder of ArthroSave BV without further relations. The other authors declare that there is no conflict of interest.

Ethical Approval

Ethical approval was provided by the local ethics committee on animal experimentation (2013.III.08.054).

Animal Welfare

The present study followed international, national, and/or institutional guidelines for humane animal treatment and complied with relevant legislation.

ORCID iDs

Michelle Teunissen (iD) https://orcid.org/0000-0003-0119-3257

Alberto Miranda Bedate D https://orcid.org/0000-0003-2623 -2857

References

- March L, Cross M, Lo C, Arden N, Gates L, Leyland K, et al. Osteoarthritis: a serious disease. Published December 1, 2016. Accessed February 1, 2021. https://oarsi.org/sites/ default/files/library/2018/pdf/oarsi_white_paper_oa_serious_ disease121416_1.pdf
- Hawker GA. Osteoarthritis is a serious disease. Clin Exp Rheumatol. 2019;37(suppl 120):3-6.

- Cross M, Smith E, Hoy D, Nolte S, Ackerman I, Fransen M, *et al.* The global burden of hip and knee osteoarthritis: estimates from the Global Burden of Disease 2010 study. Ann Rheum Dis. 2014;73:1323-30. doi:10.1136/annrheum-dis-2013-204763
- Ackerman IN, Kemp JL, Crossley KM, Culvenor AG, Hinman RS. Hip and knee osteoarthritis affects younger people, too. J Orthop Sports Phys Ther. 2017;47:67-79. doi:10.2519/ jospt.2017.7286
- Bayliss LE, Culliford D, Monk AP, Glyn-Jones S, Prieto-Alhambra D, Judge A, *et al.* The effect of patient age at intervention on risk of implant revision after total replacement of the hip or knee: a population-based cohort study. Lancet. 2017;389:1424-30. doi:10.1016/S0140-6736(17) 30059-4
- Goh EL, Lou WCN, Chidambaram S, Ma S. The role of joint distraction in the treatment of knee osteoarthritis: a systematic review and quantitative analysis. Orthop Res Rev. 2019;11:79-92. doi:10.2147/ORR.S211060
- Takahashi T, Baboolal TG, Lamb J, Hamilton TW, Pandit HG. Is knee joint distraction a viable treatment option for knee OA?—A literature review and meta-analysis. J Knee Surg. 2019;32:788-95. doi:10.1055/s-0038-1669447
- Mastbergen SC, Saris DBF, Lafeber FPJG. Functional articular cartilage repair: here, near, or is the best approach not yet clear? Nat Rev Rheumatol. 2013;9:277-90. doi:10.1038/ nrrheum.2013.29
- Flouzat-Lachaniette CH, Roubineau F, Heyberger C, Bouthors C. Distraction to treat knee osteoarthritis. Joint Bone Spine. 2017;84:141-4. doi:10.1016/j.jbspin.2016.03.004
- 10. Jansen MP, Boymans TAEJ, Custers RJH, Van Geenen RCI, Van Heerwaarden RJ, Huizinga MR, *et al*. Kneejoint distraction as treatment for osteoarthritis results in clinical and structural benefit: a systematic review and meta-analysis of the limited number of studies and patients available. Cartilage. Published online July 22, 2020. doi:10.1177/1947603520942945
- Intema F, Van Roermund PM, Marijnissen ACA, Cotofana S, Eckstein F, Castelein RM, *et al.* Tissue structure modification in knee osteoarthritis by use of joint distraction: an open 1-year pilot study. Ann Rheum Dis. 2011;70:1441-6. doi:10.1136/ard.2010.142364
- Wiegant K, Van Roermund PM, Intema F, Cotofana S, Eckstein F, Mastbergen SC, *et al.* Sustained clinical and structural benefit after joint distraction in the treatment of severe knee osteoarthritis. Osteoarthritis Cartilage. 2013;21:1660-7. doi:10.1016/j.joca.2013.08.006
- van der Woude JTAD, Wiegant K, van Roermund PM, Intema F, Custers RJH, Eckstein F, *et al.* Five-year follow-up of knee joint distraction: clinical benefit and cartilaginous tissue repair in an open uncontrolled prospective study. Cartilage. 2017;8:263-71. doi:10.1177/1947603516665442
- Jansen MP, van der Weiden GS, Van Roermund PM, Custers RJH, Mastbergen SC, Lafeber FPJG. Initial tissue repair predicts long-term clinical success of knee joint distraction as treatment for knee osteoarthritis. Osteoarthritis Cartilage. 2018;26:1604-8. doi:10.1016/j.joca.2018.08.004
- 15. Besselink NJ, Vincken KL, Bartels LW, van Heerwaarden RJ, Concepcion AN, Marijnissen ACA, et al. Cartilage

quality (dGEMRIC Index) following knee joint distraction or high tibial osteotomy. Cartilage. 2020;11:19-31. doi:10. 1177/1947603518777578

- 16. Jansen MP, Besselink NJ, van Heerwaarden RJ, Custers RJH, Emans PJ, Spruijt S, *et al.* Knee joint distraction compared with high tibial osteotomy and total knee arthroplasty: twoyear clinical, radiographic, and biochemical marker outcomes of two randomized controlled trials. Cartilage. 2021;12:181-91. doi:10.1177/1947603519828432
- Mastbergen SC, Marijnissen AC, Vianen ME, van Roermund PM, Bijlsma JW, Lafeber FP. The canine "groove" model of osteoarthritis is more than simply the expression of surgically applied damage. Osteoarthritis Cartilage. 2006;14:39-46. doi:10.1016/j.joca.2004.07.009
- Wiegant K, Intema F, Van Roermund PM, Barten-Van Rijbroek AD, Doornebal A, Hazewinkel HAW, *et al.* Evidence of cartilage repair by joint distraction in a canine model of osteoarthritis. Arthritis Rheumatol. 2015;67:465-74. doi:10.1002/art.38906
- Cook JL, Kuroki K, Visco D, Pelletier JP, Schulz L, Lafeber FPJG. The OARSI histopathology initiative—recommendations for histological assessments of osteoarthritis in the dog. Osteoarthritis Cartilage. 2010;18(suppl 3):S66-S79. doi:10.1016/j.joca.2010.04.017
- Chen Y, Sun Y, Pan X, Ho K, Li G. Joint distraction attenuates osteoarthritis by reducing secondary inflammation, cartilage degeneration and subchondral bone aberrant change. Osteoarthritis Cartilage. 2015;23:1728-35. doi:10.1016/j.joca .2015.05.018
- Baboolal TG, Mastbergen SC, Jones E, Calder SJ, Lafeber FPJG, McGonagle D. Synovial fluid hyaluronan mediates MSC attachment to cartilage, a potential novel mechanism contributing to cartilage repair in osteoarthritis using knee joint distraction. Ann Rheum Dis. 2016;75:908-15. doi:10.1136/annrheumdis-2014-206847
- 22. van Valburg AA, Van Roy HLAM, Lafeber FPJG, Bijlsma JWJ. Beneficial effects of intermittent fluid pressure of low physiological magnitude on cartilage and inflammation in osteoarthritis. An in vitro study. J Rheumatol. 1998;25: 515-20.
- Intema F, Thomas TP, Anderson DD, Elkins JM, Brown TD, Amendola A, *et al.* Subchondral bone remodeling is related to clinical improvement after joint distraction in the treatment of ankle osteoarthritis. Osteoarthritis Cartilage. 2011;19:668-75. doi:10.1016/j.joca.2011.02.005
- Zhai G, Doré J, Rahman P. TGF-β signal transduction pathways and osteoarthritis. Rheumatol Int. 2015;35:1283-92. doi:10.1007/s00296-015-3251-z
- Martel-Pelletier J, Di Battista JA, Lajeunesse D, Pelletier JP. IGF/IGFBP axis in cartilage and bone in osteoarthritis pathogenesis. Inflamm Res. 1998;47:90-100. doi:10.1007/ s000110050288
- Saito T, Tanaka S. Molecular mechanisms underlying osteoarthritis development: notch and NF-κB. Arthritis Res Ther. 2017;19:94. doi:10.1186/s13075-017-1296-y
- Huang J, Zhao L, Chen D. Growth factor signalling in osteoarthritis. Growth Factors. 2018;36:187-95. doi:10.1080/0897 7194.2018.1548444

- Jiang Y, Tuan RS. Origin and function of cartilage stem/ progenitor cells in osteoarthritis. Nat Rev Rheumatol. 2015; 11:206-12. doi:10.1038/nrrheum.2014.200
- Kapoor M, Martel-Pelletier J, Lajeunesse D, Pelletier JP, Fahmi H. Role of proinflammatory cytokines in the pathophysiology of osteoarthritis. Nat Rev Rheumatol. 2011;7: 33-42. doi:10.1038/nrrheum.2010.196
- R Core Team. R: a language and environment for statistical computing. Published online 2020. Accessed February 1, 2021. https://www.r-project.org/
- 31. RStudio Team. RStudio: Integrated Development for R. RStudio; 2020.
- Marijnissen ACA, Van Roermund PM, TeKoppele JM, Bijlsma JWJ, Lafeber FPJG. The canine "groove" model, compared with the ACLT model of osteoarthritis. Osteoarthritis Cartilage. 2002;10:145-55. doi:10.1053/ joca.2001.0491
- Marijnissen ACA, Van Roermund PM, Verzijl N, Tekoppele JM, Bijlsma JWJ, Lafeber FPJG. Steady progression of osteoarthritic features in the canine groove model. Osteoarthritis Cartilage. 2002;10:282-9. doi:10.1053/joca .2001.0507
- van der Kraan PM, van den Berg WB. Chondrocyte hypertrophy and osteoarthritis: role in initiation and progression of cartilage degeneration? Osteoarthritis Cartilage. 2012;20:223-32. doi:10.1016/j.Joca.2011.12.003
- Miosge N, Hartmann M, Maelicke C, Herken R. Expression of collagen type I and type II in consecutive stages of human osteoarthritis. Histochem Cell Biol. 2004;122:229-36. doi:10.1007/s00418-004-0697-6
- 36. Matyas JR, Huang D, Chung M, Adams ME. Regional quantification of cartilage type II collagen and aggrecan messenger RNA in joints with early experimental osteoarthritis. Arthritis Rheum. 2002;46:1536-43. doi:10.1002/art.10331
- 37. Lorenz H, Wenz W, Ivancic M, Steck E, Richter W. Early and stable upregulation of collagen type II, collagen type I and YKL₄₀ expression levels in cartilage during early experimental osteoarthritis occurs independent of joint location and histological grading. Arthritis Res Ther. 2005;7:R156-R165. doi:10.1186/ar1471
- Lafeber FPG, Van Roy H, Wilbrink B, Huber-Bruning O, Bijlsma JWJ. Human osteoarthritic cartilage is synthetically more active but in culture less vital than normal cartilage. J Rheumatol. 1992;19:123-9.
- Kiviranta I, Jurvelin J, Tammi M, SääMäunen AM, Helminen HJ. Weight bearing controls glycosaminoglycan concentration and articualr cartilage thickness in the knee joints of young beagle dogs. Arthritis Rheum. 1987;30:801-9. doi:10.1002/art.1780300710
- Palmoski MJ, Colyer RA, Brandt KD. Joint motion in the absence of normal loading does not maintain normal articular cartilage. Arthritis Rheum. 1980;23:325-34. doi:10.1002/ art.1780230310
- Vanwanseele B, Eckstein F, Knecht H, Stüssi E, Spaepen A. Knee cartilage of spinal cord-injured patients displays progressive thinning in the absence of normal joint loading and movement. Arthritis Rheum. 2002;46:2073-78. doi:10.1002/ art.10462

- 42. Hung SC, Nakamura K, Shiro R, Tanaka K, Kawahara H, Kurokawa T. Effects of continuous distraction on cartilage in a moving joint: an investigation on adult rabbits. J Orthop Res. 1997;15:381-90. doi:10.1002/jor.1100150310
- Roughley PJ, Mort JS. The role of aggrecan in normal and osteoarthritic cartilage. J Exp Orthop. 2014;1:8. doi:10.1186/ s40634-014-0008-7
- Alsalameh S, Amin R, Gemba T, Lotz M. Identification of mesenchymal progenitor cells in normal and osteoarthritic human articular cartilage. Arthritis Rheum. 2004;50:1522-32. doi:10.1002/art.20269
- Dowthwaite GP, Bishop JC, Redman SN, Khan IM, Rooney P, Evans DJR, *et al.* The surface of articular cartilage contains a progenitor cell populations. J Cell Sci. 2004;117(pt 6):889-97. doi:10.1242/jcs.00912
- 46. Fickert S, Fiedler J, Brenner RE. Identification of subpopulations with characteristics of mesenchymal progenitor cells from human osteoarthritic cartilage using triple staining for cell surface markers. Arthritis Res Ther. 2004;6:R422-R432. doi:10.1186/ar1210
- 47. Grogan SP, Miyaki S, Asahara H, D'Lima DD, Lotz MK. Mesenchymal progenitor cell markers in human articular cartilage: normal distribution and changes in osteoarthritis. Arthritis Res Ther. 2009;11:R85. doi:10.1186/ar2719
- Su X, Wu Z, Chen J, Wu N, Ma P, Xia Z, *et al.* CD146 as a new marker for an increased chondroprogenitor cell subpopulation in the later stages of osteoarthritis. J Orthop Res. 2015;33:84-91. doi:10.1002/jor.22731
- Richter W. Mesenchymal stem cells and cartilage in situ regeneration. J Intern Med. 2009;266:390-405. doi:10.1111/ j.1365-2796.2009.02153.x
- 50. Sanjurjo-Rodriguez C, Altaie A, Mastbergen S, Baboolal T, Welting T, Lafeber F, *et al.* Gene expression signatures of synovial fluid multipotent stromal cells in advanced knee osteoarthritis and following knee joint distraction. Front Bioeng Biotechnol. 2020;8:579751. doi:10.3389/fbioe.2020 .579751
- Kania K, Colella F, Riemen AHK, Wang H, Howard KA, Aigner T, *et al.* Regulation of Gdf5 expression in joint remodelling, repair and osteoarthritis. Sci Rep. 2020;10:157. doi:10.1038/s41598-019-57011-8
- 52. Leijten JCH, Bos SD, Landman EBM, Georgi N, Jahr H, Meulenbelt I, et al. GREM₁, FRZB and DKK1 mRNA levels correlate with osteoarthritis and are regulated by osteoarthritis-associated factors. Arthritis Res Ther. 2013;15:R126. doi:10.1186/ar4306
- Buckland J. Experimental arthritis: RANKL from cartilage linked to subchondral bone loss. Nat Rev Rheumatol. 2012;8:439. doi:10.1038/nrrheum.2012.110
- van der Kraan PM. Age-related alterations in TGF beta signaling as a causal factor of cartilage degeneration in osteoarthritis. Biomed Mater Eng. 2014;24(1 suppl):75-80. doi:10.3233/ BME-140976
- 55. Watt FE, Hamid B, Garriga C, Judge A, Hrusecka R, Custers RJH, *et al.* The molecular profile of synovial fluid changes upon joint distraction and is associated with clinical response in knee osteoarthritis. Osteoarthritis Cartilage. 2020;28:324-33. doi:10.1016/j.joca.2019.12.005

- 56. van Helvoort EM, Popov-Celeketic J, Eijkelkamp N, Coeleveld K, Tryfonidou MA, Wijne CD, *et al.* Canine IL4-10 fusion protein provides disease modifying activity in a canine model of OA; an exploratory study. PLoS One. 2019;14:e0219587. doi:10.1371/journal.pone.0219587
- 57. Steen-Louws C, Popov-Celeketic J, Mastbergen SC, Coeleveld K, Hack CE, Eijkelkamp N, *et al.* IL4-10 fusion protein has chondroprotective, anti-inflammatory and potentially analgesic effects in the treatment of osteoarthritis. Osteoarthritis Cartilage. 2018;26:1127-35. doi:10.1016/j. joca.2018.05.005
- Sun ZB, Peng H. Experimental study on the prevention of posttraumatic osteoarthritis in the rabbit knee using a hinged external fixator in combination with exercises. J Investig Surg. 2019;32:552-9. doi:10.1080/08941939.2018.1543483
- Van Valburg AA, Van Roermund PM, Marijnissen ACA, Wenting MJG, Verbout AJ, Lafeber FPJG, *et al.* Joint distraction in treatment of osteoarthritis (II): effects on cartilage in a canine model. Osteoarthritis Cartilage. 2000;8:1-8. doi:10.1053/joca.1999.0263
- McCoy AM. Animal models of osteoarthritis. Vet Pathol. 2015;52:803-18. doi:10.1177/0300985815588611