REVIEW ARTICLE

The Utility of Circulating and Imaging Biomarkers Alone and in Combination in Heart Failure

Biyanka Jaltotage¹, Girish Dwivedi^{1,2,3,4}, Daryl Eng Lee Ooi^{5,6} and Gnanadevan Mahadavan^{5,6,7,*}

¹Department of Cardiology, Fiona Stanley Hospital, Murdoch, Western Australia, Australia, ²Division of Cardiology, Faculty of Medicine, University of Ottawa Heart Institute, University of Ottawa, Ottawa, Ontario, Canada; ³School of Medicine, The University of Western Australia, Perth, Australia; ⁴Harry Perkins Institute of Medical Research, Nedlands, Western Australia, Australia; ⁵Discipline of Medicine, University of Adelaide, Adelaide, South Australia, Australia; ⁶Cardiology Unit, Lyell McEwin Hospital, Northern Adelaide Local Health Network, Haydown Road, Elizabeth Vale, SA 5112, Australia; ⁷Cardiology Unit, The Queen Elizabeth Hospital, Central Adelaide Local Health Network, 28 Woodville Road, Woodville South, SA 5011, Australia

	Abstract: Clinical trials in the treatment of heart failure have relied on the use of a composite of
ARTICLE HISTORY	hard clinical endpoints to evaluate the efficacy of the treatment arm. This has led to prolonged trials requiring large patient cohorts and extensive funding to reach statistical significance.
eceived: August 13, 2020 evised: December 17, 2020 cccepted: March 01, 2021	In this paper, we have explored the potential of currently available circulating and imaging bio- markers associated with heart failure as a surrogate for hard clinical end points in clinical trials.
01: 0.2174/1573403X17666210525103512	This would be expected to result in shorter trials, smaller patient cohorts and limited funding re- quired. We have subsequently theorized on combining circulating and imaging biomarkers as a sur- rogate for clinical end points such as hospitalization from heart failure and cardiac mortality.

Keywords: Circulating and imaging biomarkers, heart failure, cardiac mortality, myocardial injury, CMR, ECG.

1. INTRODUCTION

Heart failure (HF) is a global pandemic affecting at least 26 million people worldwide [1] and is associated with high morbidity and mortality [2]. It is a clinical syndrome characterised by signs and symptoms of dyspnoea, fatigue, oedema and pulmonary rales. The syndrome may represent either systolic dysfunction causing HF with reduced ejection fraction (HFrEF) or diastolic dysfunction resulting in HF with preserved ejection fraction (HFpEF). No single diagnostic test exists, which is why diagnosis is made with a combination of history, examination, laboratory testing and imaging [3]. Biomarkers are now frequently relied upon to aid diagnosis, monitor treatment and identify those at the highest risk of deterioration [4]. A number of circulating and imaging biomarkers for HF exist but alone have limited prognostic power [5]. A combination of biomarkers typically yields the best results, and although limited, there is some evidence to suggest this may also be true in HF [6].

This review will discuss available circulating and imaging (i.e., echocardiographic and Cardiovascular Magnetic Resonance imaging (CMR)) biomarkers used in the assessment and prognostication of HF patients. The review will also discuss the available evidence for combining these biomarkers with a focus on their utility in describing cardiac inflam-

* Address correspondence to this author at the Discipline of Medicine, University of Adelaide, Adelaide, SA 5005, Australia;

Tel: 0408410370; E-mail: devan.mahadavan@gmail.com

mation and fibrosis. Lastly, mortality and HF admissions are used as endpoints in pharmacological and device trials in HF. They require large study cohorts over an extended trial duration. The review will allude to the unique potential of combining these biomarkers as a surrogate for outcome assessments.

2. BIOMARKERS

A biomarker can be defined as a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacological responses to therapeutic interventions [7]. Biomarkers are utilised frequently to improve patient care; this is most prominently seen in the field of medical oncology. Biomarkers such as oestrogen receptor status in breast cancer directly dictate the use of tamoxifen treatment to improve outcomes. This direct relationship between biological processes and targeted treatment is not well-established in cardiology [8]. This individualised precise medical management is attractive, and there have been large volumes of research into biomarkers in HF. Some such as B-type Natriuretic Peptide (B-NP) or Ejection Fraction (EF) are routine in clinical practice, while others are regularly employed in research settings. For an HF biomarker to be clinically useful, it should fulfill a number of suggested criteria: 1 -) testing should be low cost, 2 -) assays used to detect that the biomarker must be robust with quick turnaround, 3 -) the biomarker should reflect an important pathophysiological pathway involved in the HF disease process, 4 -) the biomarker should provide information beyond what is available from routine examination and laboratory evaluation, and 5 -) it should add to clinical judgement for understanding diagnosis, prognosis and management of HF [9, 10].

3. METHODS

We completed a review of the literature for articles discussing biomarkers in HF. We searched the PubMed database in March 2020 for studies published between January 2010 and February 2020. We used a number of search terms, including free-text terms such as heart failure biomarkers, circulating biomarkers in heart failure, imaging biomarkers in heart failure and combining biomarkers in heart failure. Article references were also searched further for additional relevant studies.

4. CIRCULATING BIOMARKERS

4.1. Myocardial Stretch

The first natriuretic peptide introduced as a marker of myocardial stretch was Atrial Natriuretic Peptide (ANP) [11]. ANP was a marker of elevated cardiac filling pressures [11], but its use was limited by instability [12]. As a result, ANP was replaced by the largely ventricular-derived BNP and its amino-terminal propeptide equivalent N-terminal pro-BNP (NT-proBNP). Their use is now widespread in the management of HF [9]. Ventricular wall stress is the most significant trigger for the induction of the BNP gene, which releases the prepeptide proBNP. This is cleaved both within the cardiomyocyte and in peripheral sites to the biologically active BNP and inactive NT-proBNP [9]. BNP binds to natriuretic peptide receptors, stimulating natriuresis, vasodilation, lusitropy and reducing cardiac remodelling [9]. BNP and NT-proBNP play a role in diagnosis with values less than 35 pg/ml and 100 pg/ml, respectively, effective in excluding HF, while values of less than 100pg/ml and 300pg/ml, respectively, are appropriate cut-offs in acute presentations [2, 13]. Both BNP and NT-proBNP are established markers predicting prognosis in HF, and their raised levels are independently associated with mortality and other adverse outcomes [14-16]. The 2017 American College of Cardiology (ACC) guidelines for the management of heart failure endorse the use of BNP and NT-proBNP, with class I recommendations for its use in diagnosis and prognosis [17]. The use of both markers alone to guide therapy in HF is less clear and controversial due to conflicting results. Few studies have shown significant survival benefits with natriuretic peptide-guided therapy, while others have only shown positive trends or even neutral results [18-24].

Despite ANP's lack of stability, its precursor protein mid-regional-proANP (MR-proANP) is stable and can be measured [9]. Data suggests that MR-proANP is a robust marker in HF [25]. It is non-inferior to BNP and NT-proB-NP in the diagnosis of HF and can even reclassify patients with BNP results that are difficult to interpret [26]. In addition, MR-proANP has prognostic power and can predict mortality in patients with chronic HF [27].

4.2. Myocardial Injury

Troponin was first identified as an integral protein within the cardiac muscle in 1963 [28]. It is superior to traditional markers of injury such as myoglobin and creatine kinase-MB due to its clinical sensitivity and tissue specificity. It has formed the basis of acute myocardial infarction diagnosis for many years [29]. High sensitivity assays now exist measuring troponin T (TnT) and troponin I (TnI), two distinct troponin subunits. Both TnT and TnI are found in individual isoforms encoded in three genes, slow skeletal, fast skeletal and cardiac muscle [30]. These high sensitivity assays detect the cardiac-specific isoforms. TnT and TnI assays are often used interchangeably, and their use clinically is often determined by local biochemistry laboratory supply. Comparisons between the assays in certain populations have shown only a modest correlation (r=0.54) between TnT and TnI, suggesting that some care is required in interpreting these assays [31].

Cardiac troponin is almost completely found bound to the sarcomere, but 5% can be found in the cytoplasm [32]. During episodes of ischaemia, this cytoplasmic troponin is released first and causes the initial rapid rise in serum troponin levels [32]. However, rises in detectable troponin occur most commonly in the absence of ischaemia [33]. This is now termed myocardial injury and occurs through a number of mechanisms, including HF [33]. Myocardial injury in HF arises as a result of a number of factors, including subendocardial stress and myocyte degeneration [10]. A number of studies have confirmed that elevation of high sensitivity troponin T (despite undetectable levels on conventional troponin assays) in patients with acute HF predicts mortality [34, 35]. High sensitivity troponin T remains prognostic in patients with chronic stable HF, with elevated levels predicting adverse cardiovascular outcomes, hospitalisation for cardiovascular causes and mortality [36, 37]. The use of troponin as a prognostic marker in HF has a class I recommendation from the ACC guidelines [17].

4.3. Cardiac Inflammation

The myocardium is sensitive to inflammatory cytokines, which can promote inflammation and cardiac injury [38, 39]. The resulting damage can impair heart function and cause HF. The most prominent and frequently used marker for the overall systemic burden of inflammation is the acute phase reactant C-Reactive Protein (CRP), which is produced by hepatocytes and stimulated by the inflammatory cytokine interleukin IL-6 [40]. Interest in inflammation and HF outcomes began as early as 1956 when it was found that chronic HF patients with elevated levels of CRP had more severe cardiac dysfunction [41]. More recently, high sensitivity CRP (hsCRP) has become available to detect low-grade inflammation in conditions such as HF [42]. Elevated hsCRP in chronic HF is now an established marker of poor prognosis, predicting more severe disease and increased morbidity and mortality [43-45]. For patients presenting with acute HF, the role of hsCRP as a marker of prognosis is less clear and is not recommended [38].

Tumour necrosis factor-alpha (TNF α) is a proinflammatory cytokine that forms part of the innate immune system's inflammatory response [46]. It was in 1990 that it was first identified that circulating levels of TNF- α were elevated in patients with chronic HF [47]. Further research into the association between TNF α and chronic HF demonstrates that raised levels can predict the development of HF in healthy individuals [48], predict symptom severity [49] and predict mortality [50, 51]. These findings are explained by TNF α downregulating myocardial sarcoplasmic reticulum Ca²⁺ AT-Pase and promoting cardiac remodelling resulting in impaired cardiac function [38].

4.4. Cardiac Fibrosis

Fibrotic diseases, including HF, are a proven cause of morbidity and mortality, accounting for over 800,000 deaths around the world each year [52]. Cardiac fibrosis is triggered following an insult, most frequently ischaemia. It is a protective mechanism but, over time, leads to irreversible ventricular remodelling and cardiac dysfunction [52]. The fibrosis-related dysfunction is a result of increased ventricular stiffness and compromised electrical conduction [53]. Given the impact of fibrosis on the heart, there is great interest in novel markers of cardiac fibrosis [54].

The suppression of the tumorigenicity-2 (ST2) network plays a critical role in mediating fibrosis and myocardial and vascular remodelling. Usually, IL-33 binds to the ST2 receptor causing the downstream reduction in programmed cell death and activation of profibrotic pathways. Soluble ST2 (sST2) acts as a decoy receptor for IL-33, preventing binding to the ST2 receptor promoting myocardial cell death and fibrosis [9]. Levels of sST2 may be elevated in the absence of heart failure in 10-18% of men and 2-8% of women, but measurement still has value in both acute and chronic HF [55]. Serial measurements of sST2 in the acute setting predicts mortality [56], and in chronic HF, it is superior to BNP and NT-proBNP in predicting worsening HF, rehospitalisation, heart transplantation and death [57]. In contrast to the natriuretic peptides, sST2 concentrations are also unchanged by obesity, age, atrial fibrillation or renal function [9].

Galectin-3 is produced by activated macrophages and stimulates macrophage migration and fibroblast proliferation-inducing cardiac fibrosis [58]. The concentration of galectin-3 is maximal during peak fibrosis and is almost absent after recovery, making it a dynamic marker of fibrosis [59]. It has been shown to be a superior predictor to NTproBNP in patients with acute HF in predicting episodes of recurrent HF and death [60]. Despite being a useful marker in acute HF, its role in diagnosis is limited, and it is inferior to the natriuretic peptides in this regard [61]. In chronic HF, galectin-3 is effective in predicting mortality [62], and the use of serial levels has been shown to predict the first morbid event, hospitalisation for HF and mortality [63]. In addition, elevated levels of galectin-3 in healthy individuals predict the development of new-onset HF and also mortality [64]. Galectin-3 is an effective marker in HFpEF, elevated levels correspond to more severe diastolic dysfunction [65,

66], and it is the most accurate marker of hospitalisation and mortality in these patients [67].

Despite the available evidence for cardiac fibrosis biomarkers, their utilisation in clinical settings remains limited. At this time, a class IIb recommendation by the ACC exists for their use in HF, and it has been suggested that more benefit may be derived through the combination with other biomarkers (circulating or imaging) [17].

4.5. Future Directions

The search continues for novel circulating biomarkers in heart failure despite the wide range currently available. Recent work has identified the potential of MicroRNAs (miR-NAs) and metabolomics as biomarkers.

MiRNAs are endogenous, conserved, single-stranded, small (~22 nucleotides) non-coding ribonucleic acid (RNA) with critical roles in cardiovascular biology. They have shown promise in challenging clinical settings where currently established biomarkers perform poorly, such as atrial fibrillation or obesity, discriminating between HFrEF and HFpEF, and determining heart failure aetiology [68-70]. Further investigation is still required in validating miRNA panels and clarifying their role in prognosis, diagnosis and management.

The heart is a highly metabolically active organ that is capable of generating adenosine triphosphate (ATP), the energy-carrying molecule in cells, from a diverse range of sources, including carbohydrates, lipids, lactate, amino acids and ketones. Despite the heart's adaptability, it remains susceptible to disruptions in cardiac metabolism, frequently seen in most cardiovascular diseases. Metabolomics investigates metabolites affecting genetic, epigenetic, transcription and protein factor variation that remain responsive to environmental exposures, dietary intake and the gut microbiome [71]. Measurement of these circulating metabolites exposes changes in both cardiac and systemic metabolism [72]. Studies have already identified metabolites such as longchain acylcarnitines levels that are related to heart failure severity and are sensitive to treatment [73, 74]. Metabolomics has also revealed that patients with HFrEF and HFpEF, currently thought to be unique clinical entities, share common metabolic derangements with raised levels of long-chain acylcarnitines [75]. Despite currently being in its infancy, metabolomics has the potential to identify clinically relevant biomarkers that add to our understanding of heart failure pathophysiology and may lead to the development of targeted metabolic therapies.

5. IMAGING BIOMARKERS

5.1. Echocardiogram

Echocardiogram is a safe and available resource for cardiac imaging. The use of transthoracic echocardiogram is the mainstay of HF assessment and cardiac function [2].

The EF is the basis of ventricular systolic function and can be defined as the percentage of blood ejected in systole in relation to the volume of blood in the ventricle at the end of diastole. It can be calculated by a number of methods, most frequently by using a biplane technique [76]. The Left Ventricular EF [LVEF] is a strong predictor of clinical outcomes in patients with HF. Reduction in LVEF strongly predicts the severity of symptoms and all-cause mortality [77]. Furthermore, a declining trajectory of LVEF also predicts mortality [78]. In addition to its prognostic power, LVEF is vital in guiding medical and device therapy [2].

Increases in Left Ventricular Mass (LVM) are caused by cardiac remodelling, and it is associated with high blood pressure, increased body mass index, smoking status and diabetes mellitus. LVM can be estimated by using either two-dimensional (2D) or three-dimensional (3D) echocardiography. The LVM is based on the myocardial density (1.05g/m-l) multiplied by the myocardial volume (left ventricular (LV) volume subtracted from the volume enclosed by the epicardium). Echocardiogram-derived LVM has been shown to be a reliable predictor of adverse cardiovascular events [79].

The term strain in echocardiography describes local shortening, lengthening and thickening of the myocardium to evaluate regional LV function. Strain is frequently calculated using speckle-tracking echocardiography. This technique employs the speckles in myocardial tissue caused by acoustic markers on ultrasound. The speckles throughout the myocardium are stable over a short time period, and their 2D displacement can be calculated. With these results, strain can be calculated for the LV in circumferential, longitudinal and radial directions. In 2D echocardiography, strain can be only be calculated in two axes, whilst strain in all three axes can be recorded with 3D echocardiography. Typically to measure global LV function, strain is recorded as Global Longitudinal Strain (GLS), the average strain of all speckles in the longitudinal direction [80]. In acute HF, GLS predicts HF readmission [81], adverse cardiac events [82], and it has been shown to be superior to LVEF in predicting mortality [83]. In chronic HF, GLS effectively predicts HF exacerbation, ventricular assist device placement, cardiac transplantation and all-cause mortality [84, 85]. Furthermore, in asymptomatic individuals, GLS can predict the development of new-onset HF [86]. This has led to increasing utility within the cardio-oncology specialty for monitoring LV function in patients at risk of chemotherapy-related cardiac toxicity [87]. This allows timely initiation of HF therapy to avoid cessation of potential lifesaving cancer therapy, an area with further studies underway [88].

Myocardial Work (MW) is a novel non-invasive echocardiographic approach that assesses regional myocardial work by LV Pressure-Strain Loop (PSL) analysis *via* echocardiographic software, thus incorporating both strain and afterload with non-invasively estimated pressure from brachial cuff blood pressure. This technique has been validated with invasive LV Pressure-Volume Loops (PVL) and regional myocardial metabolism with glucose turnover measured by positron emission tomography, providing a robust method for LV performance assessment taking loading conditions into account [89]. The association between non-invasive derived global MW by PSL and favourable response to Cardiac Resynchronisation Therapy (CRT) was demonstrated in patients with HF of both ischaemic and non-ischaemic aetiology, predicting subsequent reverse remodelling [90, 91]. MW efficiency is distinctly reduced in HFrEF patients [92]. A Global Work Index (GWI) of <500 mmHg% was shown to be a predictor of poor prognosis with established prognostic parameters of HF [94]. Non-invasive myocardial work is still at an early developmental stage. The published results to date are promising whilst requiring further validation to reach routine clinical practice.

Diastolic dysfunction of the LV or HFpEF may be challenging to detect with echocardiography. During diastole, the left atrium is exposed to increasing LV pressures. Subsequently, left atrial pressures rise to maintain appropriate filling. This sustained increase in pressure leads to dilatation and stretching of the atrial myocardium. Therefore, left atrial volume is an established marker of the severity of diastolic dysfunction [94]. Despite this clear association, up to one-third of patients with diastolic dysfunction have normal left atrial volume [94, 95]. This triggered an investigation into assessing left atrial function by evaluating parameters such as left atrial strain. Work in this area has identified that left atrial strain is a marker of diastolic dysfunction, worsening atrial fibrillation, stroke and may be a predictor of adverse cardiovascular events [96, 97].

The Right Ventricle (RV) can be defined as low pressure, high volume pump in contrast to the LV, which is a high pressure, high volume pump [98]. Consequently, due to exposure to lower pressures, the overall mass of the RV is approximately one-sixth of the LV. The RV and LV are interconnected by networks of muscle fibres and are functionally interdependent. As a result, impairment of RV function is detrimental to overall cardiac function [99]. Assessment of the RV, although not the gold standard, is possible with the use of echocardiography through a number of methods [5]. RV fractional area change is a percentage change in cavity area from end-diastole to end-systole and is a predictor of stroke, HF, cardiovascular death and all-cause mortality [100]. Tricuspid Annular Plane Systolic Excursion (TAPSE) is a relatively simple measure of RV function. It is calculated with the use of M-mode in the apical four-chamber view measuring the displacement of the tricuspid ring in the longitudinal plane of the RV. TAPSE has been shown in healthy individuals to predict the development of cardiovascular disease [101]. Given the success of strain measurements in the LV, RV strain has also been investigated. The results show that RV strain in chronic HF patients is a powerful predictor of admission for HF, cardiac transplantation, emergency ventricular assist device implantation and death, and may even be superior to other measurements of RV and LV function [102].

5.2. CMR

CMR is the gold standard cardiac imaging modality for the measurement of volumes, mass, and EF. It also plays a pivotal role in determining HF aetiology, visualising myocardial fibrosis and assessing viability. Weaknesses include availability, cost, limited use of gadolinium-based contrast in renal impairment due to the risk of nephrogenic systemic fibrosis and patient compliance factors such as orthopnoea and claustrophobia [2]. Detailed images produced by CMR rely on hydrogen nuclei which are abundant in water and fat. The use of a strong magnetic field followed by a radiofrequency wave causes the randomly spinning nuclei with their own magnetic vector to initially align and subsequently resonate. The magnetic field and radiofrequency wave are then terminated, causing the nuclei to emit a signal which is used to create the images seen in MRI. Multiple radiofrequency pulses can be used in sequence to emphasise different mediums. Each medium has a different rate of relaxation and can be measured in two ways, the first is T1 relaxation which is the time for the nuclei to return to the resting magnetic vector and T2 relaxation is the time taken for the nuclei to return to their resting spin [103]. The use of gadolinium-based contrast is commonplace. The contrast reduces the T1 relaxation time of tissues, and the factors contributing to the greatest changes are local perfusion, the extracellular volume of distribution and water exchange rates (between vascular, interstitial and cellular spaces) [104, 105].

As discussed earlier, myocardial fibrosis has a significant impact on heart function. CMR is an effective tool in measuring fibrosis with the use of Late Gadolinium Enhancement (LGE) to measure focal localised fibrosis and T1 mapping to measure diffuse interstitial fibrosis. The use of LGE to measure cardiac fibrosis was first described in animal models in 1984 [106]. Since this time, it has been increasingly used to accurately measure cardiac fibrosis in a wide range of conditions, including HF, myocardial infarction, hypertrophic cardiomyopathy, aortic valve disease, sarcoidosis, amyloidosis, hypertensive cardiomyopathy and diabetic cardiomyopathy [107]. LGE has also been shown to predict adverse outcomes in many of these conditions [108-112] and more recently has been proposed to guide implantable cardioverter defibrillator therapy [113]. Native T1 mapping to measure fibrosis and inflammation is more novel and could be used in the absence of gadolinium without limitation by renal impairment. There are a number of different techniques to acquire a T1 map, the more common ones being a Modified Look-Locker inversion recovery (MOLLI) and the Shortened Modified Look-Locker inversion recovery (SchMOLLI), which have been shown to accurately measure fibrosis both in the intracellular and extracellular space, in myocardial infarction [114, 115], amyloidosis [116], systemic sclerosis [117], diabetic cardiomyopathy [118], hypertrophic cardiomyopathy [119] and chronic HF [120, 121]. The map is an image of each pixel which is colour-coded according to the absolute T1 time. There is also evidence now demonstrating that fibrosis detected by T1 mapping predicts adverse outcomes [122]. Increased TI times may also indicate inflammation and, therefore, will need to be interpreted in conjunction with T2 mapping described in a later section.

The other method of evaluating extracellular volume (ECV) is by creating pre and post-contrast T1 maps (ECV

mapping). The extracellular volume is a marker of volume expansion, which can be due to diffuse myocardial fibrosis or myocardial inflammation. There was a prospective observational multi-centre longitudinal study in 637 consecutive patients with dilated non-ischemic cardiomyopathy undergoing CMR with T1 mapping and LGE at 1.5-T and 3.0-T [123]. The primary endpoint was all-cause mortality, and a composite of HF mortality and hospitalization was a secondary endpoint. During a median follow-up period of 22 months (interquartile range: 19 to 25 months), a total of 28 deaths (22 cardiac) and 68 composite HF events were observed. T1 mapping indices (native T1 and extracellular volume fraction), as well as the presence and extent of LGE, were predictive of all-cause mortality and HF endpoint (p <0.001 for all). In multivariable analyses, native T1 was the sole independent predictor of all-cause mortality and HF composite endpoints (hazard ratio: 1.1; 95% confidence interval: 1.06 to 1.15; hazard ratio: 1.1; 95% confidence interval: 1.05 to 1.1; p < 0.001 for both), followed by the models including the extent of LGE and RV EF, respectively. Noninvasive measures of diffuse myocardial disease by T1 mapping are significantly predictive of all-cause mortality and HF events in non-ischaemic dilated cardiomyopathy. The average LVEF in the initial cohort was about 47%, suggesting that the sensitivity of detecting changes is very high using these methods in a cohort with early disease.

The combination of other poor prognostic markers such as LVEF <35% or LGE with native T1 did not improve the predictive value of native T1 values alone, indicating the independent pathophysiological role of a diffuse myocardial disease as indicated by an elevated native T1 value [124]. Vita *et al.* further refined the assessment of diffuse myocardial disease by mapping 6 anatomical locations using all 4 CMR tissue-characterizing methods (native T1, extracellular volume mapping, partition coefficient (λ GD) and late gadolinium enhancement) associating this with outcome [125]. The authors performed T1 mapping of the myocardium and the blood pool, before and serially after contrast injection, using a Look-Locker cine gradient-echo technique to obtain T1 and the corresponding reciprocal R1(1/T1) values. XGd values were derived from the slopes of the least-squares regression lines for myocardial versus blood R1, then adjusted to serum haematocrit to yield ECV.

After a median of 3.8 years, 36 (15%) experienced major adverse cardiac events (MACE), including 22 HF hospitalizations and 14 deaths. Non-ischemic LGE was detected in 34%, whereas ECV was elevated (in more than 1 location) in 58%. Comparing the 4 methods, mean ECV and λ Gd both demonstrated a strong association with MACE (both p < 0.001). In contrast to native T1 and LGE, ECV values from all 6 locations were associated with MACE and death, with the anteroseptum being the most significant (p < 0.0001). The number of abnormal ECV locations correlated linearly with annual MACE rates (p = 0.0003). Mean ECV was the only predictor to enter a prognostic model that contained age, sex, New York Heart Association functional class, and LVEF. For every 10% increase, mean ECV portended to 2.8-fold adjusted increase risk to MACE (p < 0.001). These newer non-invasive imaging techniques, namely serial T1 mapping imaging characterizing ECV fraction, have been validated against diffuse interstitial fibrosis by histology in non-ischaemic dilated cardiomyopathy [126, 127].

Detection of myocardial inflammation-causing cardiac dysfunction is significant as it is treatable with a number of management options available [128, 129]. The gold standard in diagnosis is the endomyocardial biopsy, but it is prone to sampling errors, is invasive and has risks [130]. The use of CMR with T2 mapping presents an opportunity for non-invasive detection of myocardial inflammation. Research comparing the use of T2 mapping to endomyocardial biopsy suggests that it may be a reasonable alternative [131], and increased T2 has also been shown to be a predictor of MACE [132].

As with any diagnostic test, standardization of data acquisition and post-processing, as well as predefined reference ranges, are a prerequisite for the application of quantifiable imaging biomarkers in clinical routine. Achieving this and standardizing the various vendor platforms remain the main impediments to this being used in routine clinical practice, but in the setting of clinical trials, these obstacles may be overcome, especially with the use of a core lab to acquire and process the images.

6. COMBINING CIRCULATING AND IMAGING BIO-MARKERS

An extensive range of biomarkers is now available in HF [5, 133]. Both circulating and imaging biomarkers have much to offer, but most research has investigated their utility in isolation. Risk models do exist but largely only incorporate circulating biomarkers [134]. The limited work available combining circulating and imaging biomarkers has been confined to mostly natriuretic peptides and echocardiogram [5].

Natriuretic peptides have been considered the gold standard biomarker in HF and are routinely used in its management [135]. However, as discussed earlier, the benefits of using natriuretic peptides alone to guide HF therapy are unclear. The work by Simioniuc et al. highlights the potential benefits of combining circulating and imaging biomarkers. The authors compared clinically guided HF therapy and BNP combined with echocardiogram-derived measures of increased LV pressure (E wave deceleration time as a surrogate of pulmonary capillary wedge pressure and for patients in atrial fibrillation deceleration time of mitral flow velocity) guided HF therapy. Patients were not randomised, and propensity score matching of confounding baseline variables was utilized to minimise bias. The results demonstrated that combining these biomarkers reduced rates of acute kidney injury (9.8% vs. 21.4%, p<0.0001) and death (hazard ratio: 0.45; 95% confidence interval: 0.30-0.67, p<0.0001) [6] (Fig. 1). Furthermore, Bajraktari et al. compared 794 outpatients with heart failure treated in three groups: group I with BNP combined with echocardiogram (E/e' and E wave deceleration time as surrogates of increased LV pressure together with lung ultrasound to assess B lines) guided therapy, group II with clinically guided therapy and group III with those managed with no specific specialist follow up. They found a 60 months survival of 88% in group I, 75% in group II and 54% in group III (p<0.0001) [136]. These results highlight a clinical role for the combination of BNP and echocardiogram in improving HF outcomes.

BNP: B-type natriuretic peptide; HR: Hazard ratio; CI: Confidence interval. Figure used with permission from Simioniuc *et al.* [6].

As discussed previously, circulating biomarkers of fibrosis exist, such as galectin-3, which have been proposed to not be just a by-product but an active culprit in the development of myocardial fibrosis [137]. CMR and possible myocardial works with echocardiography is an effective tool for visualising both focal and diffuse myocardial fibrosis. The combination of CMR and markers of fibrosis presents an opportunity to directly visualise the underlying pathological process. There is some evidence to suggest that galectin-3 and Matrix Metalloproteinase-2 (MMP-2), a marker of extracellular matrix remodelling, may best correlate with the level of fibrosis seen on CMR with the use of T1 mapping and LGE [138]. Further work is needed to develop models combining CMR and circulating fibrosis markers to predict outcomes.

Multiple circulating biomarkers of myocardial inflammation also exist. There has been work performed to investigate combining T1 mapping and LGE findings of fibrosis together with circulating inflammatory biomarkers. Mateus et al. investigated a population of 1345 patients from the Multi--Ethnic Study of Atherosclerosis (MESA), a multicentre prospective cohort study. These patients had CMR with T1 mapping using the MOLLI recovery sequence. Patients were excluded if they self-reported medical conditions that could elevate non-specific inflammatory markers. 772 participants remained in the final study population. The results showed that in men, elevated IL-6 was associated with a 0.4% higher ECV (p=0.05), and elevated CRP was associated with a 4.9ms higher native T1 (p=0.03). However, no such correlation between inflammatory markers and CMR detected fibrosis was present in women [139]. Markers of inflammation and CMR were also explored by Wu et al. to identify HF patients at the highest risk of malignant cardiac rhythms. In total, 235 patients with chronic ischaemic and non-ischaemic cardiomyopathy with an LVEF <35% undergoing insertion of an Implantable Cardioverter-Defibrillator (ICD) had hsCRP and CMR to assess grey zone (scar related heterogenous myocardium via LGE) performed and the median follow-up was 3.6 years. The primary end point was appropriate ICD shock for ventricular tachycardia/fibrillation or cardiac death. The adjusted hazard ratio for the primary end point for hsCRP was up to 2.8 (95% confidence interval: 1.1-7.1, p=0.03), and for the grey zone, it was 4.6 (95% confidence interval: 1.4-15.4, p<0.01). Significantly, the combination of hsCRP and grey zone was associated with a hazard ratio of up to 24.0 (95% confidence interval: 3.1-184, p=0.002), suggesting the combination of hsCRP and CMR may assist in identifying low-risk patients, with the least poe160721193557

tential benefit, meeting current guidelines for ICD insertion [140]. Ongoing work in this area may continue to identify further prognostic indicators.

CMR with T2 mapping provides an alternative to endomyocardial biopsy to evaluate myocardial inflammation [131]. However, to our knowledge, no studies exist that combine circulating biomarkers of inflammation and T2 mapping, providing an opportunity for future studies.

7. UTILITY OF BIOMARKER(S) AS A SURROGATE FOR AN OUTCOME MEASURE

The selection of the primary 'endpoint' or 'outcome measure' has a major impact on the reliability and interpretability of clinical trials designed to study the effect of an intervention. As HF studies, especially interventional studies, with hard outcome measures such as mortality, are difficult to conduct, the use of biomarkers as a surrogate measure becomes attractive for a number of reasons [7]. Biomarkers are far cheaper, easier and faster to measure than the actual outcome measure and also require smaller sample sizes which can subsequently reduce costs and the time for trial completion. However, to prevent confounding factors from nullifying the value of surrogate outcome measures, a thorough knowledge of the pathophysiology of the disease and the intervention being performed is required [141]. While HF biomarkers strongly correlate with clinical efficacy measures in natural history observations, they are not causal in the pathway of the disease process and may provide misleading information about clinical efficacy.

HF is a complex heterogenous condition, and in some respects, such as HFpEF, it is still poorly understood [142]. In addition, our understanding of biomarkers is still not comprehensive, the timing of measuring biomarkers to confirm benefit is uncertain, and we have limited knowledge in regard to how significant a change in biomarker levels is needed before there is any benefit [143]. Vaduganathan et al. evaluated the use of ANP, BNP and NT-proBNP biomarkers known to be associated with mortality as a surrogate endpoint for HF therapy. The authors found that the changes in natriuretic peptides levels were associated with HF hospitalisation but not mortality [144]. These findings highlight that we still lack a complete understanding of the relationships between HF and its biomarkers. It would be reasonable to conclude that despite an array of promising biomarkers available to us, we are not yet in a position to completely replace mortality as a primary endpoint, but considering biomarkers as a composite of surrogate outcomes may have potential [145].



Fig. (1). (A) Kaplan-Meier curves for all-cause death in patients of the echo-BNP guided and clinically guided groups before (left) and after (right) propensity score matching. (B) Kaplan-Meier curves for the combined end point of and worsening renal function in echo-BNP guided and clinically guided groups before (left) and after (right) propensity score matching. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

Fibrosis, a marker of cell death, represents the last stage of the pathophysiological process of myocardial injury from a range of disease processes [146]. We posit that targeting cardiac fibrosis as the surrogate endpoint for HF therapy with the use of a combination of circulating and imaging biomarkers is a future research area. Based on available evidence from previous studies, the combination of Galectin-3 and either myocardial works with echocardiography or T1 mapping with CMR holds the most promise as they represent the closest surrogate for fibrotic replacement of the myocardium [59, 107, 147]. Future studies should evaluate a combination of fibrosis biomarkers as a composite end point that would also include HF hospitalization as the latter has been shown to result in poorer quality of life, prognosis, as well as represents a huge economic burden on the health system [148].

CONCLUSION

A vast range of circulating and imaging biomarkers are used for HF diagnosis, monitoring treatment and also identifying patients who are at the highest risk of deterioration. Preliminary studies indicate that a strategy based on the combination of circulating and imaging biomarkers is superior to when either one is used alone for such purposes. Larger studies are, however, needed before such approaches can be adopted into clinical practices.

CONSENT FOR PUBLICATION

Not applicable.

FUNDING

None.

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

ACKNOWLEDGEMENTS

Declared none.

REFERENCES

[1] Savarese G, Lund LH. Global public health burden of heart failure. Card Fail Rev 2017; 3(1): 7-11.

http://dx.doi.org/10.15420/cfr.2016:25:2 PMID: 28785469
[2] Ponikowski P, Voors AA, Anker SD, *et al.* Authors/Task Force Members; Document Reviewers. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC). Developed with the special contribution of the Heart Failure Association (H-FA) of the ESC. Eur J Heart Fail 2016; 18(8): 891-975. http://dx.doi.org/10.1002/ejhf.592 PMID: 27207191

- [3] King M, Kingery J, Casey B. Diagnosis and evaluation of heart failure. Am Fam Physician 2012; 85(12): 1161-8. PMID: 22962896
- [4] Suzuki T, Bossone E. Biomarkers of heart failure: past, present, and future. Heart Fail Clin 2018; 14(1): ix-x.
- http://dx.doi.org/10.1016/j.hfc.2017.08.012 PMID: 29153205
 [5] Salzano A, Marra AM, D'Assante R, *et al.* Biomarkers and imaging: complementary or subtractive? Heart Fail Clin 2019; 15(2):

321-31.

http://dx.doi.org/10.1016/j.hfc.2018.12.008 PMID: 30832821

[6] Simioniuc A, Čarluccio E, Ghio S, *et al.* Investigators of the Network Labs Ultrasound (NEBULA) in Heart Failure Study Group. Echo and natriuretic peptide guided therapy improves outcome and reduces worsening renal function in systolic heart failure: an observational study of 1137 outpatients. Int J Cardiol 2016; 224: 416-23.

http://dx.doi.org/10.1016/j.ijcard.2016.09.034 PMID: 27690339

- [7] Group BDW. Biomarkers Definitions Working Group. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. Clin Pharmacol Ther 2001; 69(3): 89-95. http://dx.doi.org/10.1067/mcp.2001.113989 PMID: 11240971
- [8] Zanad F. What is measured by cardiac fibrosis biomarkers and imaging? Circ Heart Fail 2014; 7(2): 239-42. http://dx.doi.org/10.1161/CIRCHEARTFAILURE.114.001156
 PMID: 24643887
- Ibrahim NE, Januzzi JL Jr. Established and emerging roles of biomarkers in heart failure. Circ Res 2018; 123(5): 614-29. http://dx.doi.org/10.1161/CIRCRESAHA.118.312706 PMID: 30355136
- Ibrahim NE, Januzzi JL Jr. Beyond natriuretic peptides for diagnosis and management of heart failure. Clin Chem 2017; 63(1): 211-22. http://dx.doi.org/10.1373/clinchem.2016.259564 PMID:

http://dx.doi.org/10.13/3/clinchem.2016.259564 PMID 28062619

[11] Burnett JC Jr, Kao PC, Hu DC, et al. Atrial natriuretic peptide elevation in congestive heart failure in the human. Science 1986; 231(4742): 1145-7.

 http://dx.doi.org/10.1126/science.2935937 PMID: 2935937

 [12]
 Braunwald E. Heart failure. JACC Heart Fail 2013; 1(1): 1-20.

- http://dx.doi.org/10.1016/j.jchf.2012.10.002 PMID: 24621794 [13] Januzzi JL Jr, Camargo CA, Anwaruddin S, *et al.* The N-terminal
- Pro-BNP investigation of dyspnea in the emergency department (PRIDE) study. Am J Cardiol 2005; 95(8): 948-54. http://dx.doi.org/10.1016/j.amjcard.2004.12.032 PMID: 15820160
- [14] Fonarow GC, Peacock WF, Phillips CO, Givertz MM, Lopatin M. ADHERE Scientific Advisory Committee and Investigators. Admission B-type natriuretic peptide levels and in-hospital mortality in acute decompensated heart failure. J Am Coll Cardiol 2007; 49(19): 1943-50.

http://dx.doi.org/10.1016/j.jacc.2007.02.037 PMID: 17498579

[15] Cheng V, Kazanagra R, Garcia A, et al. A rapid bedside test for B-type peptide predicts treatment outcomes in patients admitted for decompensated heart failure: A pilot study. J Am Coll Cardiol 2001; 37(2): 386-91. http://dx.doi.org/10.1016/S0735-1097(00)01157-8 PMID:

11216951

[16] O'Brien RJ, Squire IB, Demme B, Davies JE, Ng LL. Predischarge, but not admission, levels of NT-proBNP predict adverse prognosis following acute LVF. Eur J Heart Fail 2003; 5(4): 499-506.

http://dx.doi.org/10.1016/S1388-9842(03)00098-9 PMID: 12921811

- [17] Yancy CW, Jessup M, Bozkurt B, et al. 2017 ACC/AHA/HFSA Focused Update of the 2013 ACCF/AHA Guideline for the Management of Heart Failure: A report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines and the Heart Failure Society of America. J Card Fail 2017; 23(8): 628-51.
- http://dx.doi.org/10.1016/j.cardfail.2017.04.014 PMID: 28461259
 Jourdain P, Jondeau G, Funck F, *et al.* Plasma brain natriuretic
- peptide-guided therapy to improve outcome in heart failure: The STARS-BNP multicenter study. J Am Coll Cardiol 2007; 49(16): 1733-9.

http://dx.doi.org/10.1016/j.jacc.2006.10.081 PMID: 17448376

[19] Januzzi JL Jr, Rehman SU, Mohammed AA, et al. Use of amino-terminal pro-B-type natriuretic peptide to guide outpatient therapy of patients with chronic left ventricular systolic dysfunction. J Am Coll Cardiol 2011; 58(18): 1881-9.

http://dx.doi.org/10.1016/j.jacc.2011.03.072 PMID: 22018299

[20] Eurlings LW, van Pol PE, Kok WE, *et al.* Management of chronic heart failure guided by individual N-terminal pro-B-type natriuret-

ic peptide targets: Results of the PRIMA (Can PRo-brain-natriuretic peptide guided therapy of chronic heart failure IMprove heart fAilure morbidity and mortality?) study. J Am Coll Cardiol 2010; 56(25): 2090-100.

http://dx.doi.org/10.1016/j.jacc.2010.07.030 PMID: 21144969

- [21] Shah MR, Califf RM, Nohria A, et al. The STARBRITE trial: A randomized, pilot study of B-type natriuretic peptide-guided therapy in patients with advanced heart failure. J Card Fail 2011; 17(8): 613-21.
- http://dx.doi.org/10.1016/j.cardfail.2011.04.012 PMID: 21807321
 Pfisterer M, Buser P, Rickli H, *et al.* TIME-CHF Investigators. BNP-guided *vs.* symptom-guided heart failure therapy: The Trial of Intensified *vs.* Standard Medical Therapy in Elderly Patients with Congestive Heart Failure (TIME-CHF) randomized trial. JA-MA 2009; 301(4): 383-92.
- http://dx.doi.org/10.1001/jama.2009.2 PMID: 19176440
 [23] Lainchbury JG, Troughton RW, Strangman KM, *et al.* N-terminal pro-B-type natriuretic peptide-guided treatment for chronic heart failure: Results from the BATTLESCARRED (NT-proBNP-Assisted Treatment To Lessen Serial Cardiac Readmissions and Death) trial. J Am Coll Cardiol 2009; 55(1): 53-60.
- http://dx.doi.org/10.1016/j.jacc.2009.02.095 PMID: 20117364
 [24] Gaggin HK, Mohammed AA, Bhardwaj A, et al. Heart failure outcomes and benefits of NT-proBNP-guided management in the elderly: Results from the prospective, randomized ProBNP outpatient tailored chronic heart failure therapy (PROTECT) study. J Card Fail 2012; 18(8): 626-34.

http://dx.doi.org/10.1016/j.cardfail.2012.05.005 PMID: 22858078

- [25] Mueller C, McDonald K, de Boer RA, et al. Heart Failure Association of the European Society of Cardiology. Heart Failure Association of the European Society of Cardiology practical guidance on the use of natriuretic peptide concentrations. Eur J Heart Fail 2019; 21(6): 715-31.
 - http://dx.doi.org/10.1002/ejhf.1494 PMID: 31222929
- [26] Maisel A, Mueller C, Nowak R, et al. Mid-region pro-hormone markers for diagnosis and prognosis in acute dyspnea: Results from the BACH (Biomarkers in Acute Heart Failure) trial. J Am Coll Cardiol 2010; 55(19): 2062-76. http://dx.doi.org/10.1016/j.jacc.2010.02.025 PMID: 20447528
- [27] Masson S, Latini R, Carbonieri E, *et al.* GISSI-HF Investigators. The predictive value of stable precursor fragments of vasoactive peptides in patients with chronic heart failure: Data from the GIS-SI-heart failure (GISSI-HF) trial. Eur J Heart Fail 2010; 12(4): 338-47.
- http://dx.doi.org/10.1093/eurjhf/hfp206 PMID: 20097683
 [28] Ebashi S. Third component participating in the super precipitation of 'Natural Actomyosin'. Nature 1963; 200(4910): 1010. http://dx.doi.org/10.1038/2001010a0 PMID: 14097720
- [29] Wu AH, Feng YJ, Contois JH, Pervaiz S. Comparison of myoglobin, creatine kinase-MB, and cardiac troponin I for diagnosis of acute myocardial infarction. Ann Clin Lab Sci 1996; 26(4): 291-300.
- PMID: 8800429
 [30] Sheng J-J, Jin J-P. TNNI1, TNNI2 and TNNI3: Evolution, regulation and protein structure-function relationships. Gene. 2016.

tion, and protein structure-function relationships. Gene 2016; 576(1 Pt 3): 385-94. http://dx.doi.org/10.1016/j.gene.2015.10.052 PMID: 26526134

[31] Kimenai DM, Henry RM, van der Kallen CJ, et al. Direct comparison of clinical decision limits for cardiac troponin T and I. Heart 2016; 102(8): 610-6.

http://dx.doi.org/10.1136/heartjnl-2015-308917 PMID: 26794233

- [32] Katus HA, Remppis A, Scheffold T, Diederich KW, Kuebler W. Intracellular compartmentation of cardiac troponin T and its release kinetics in patients with reperfused and nonreperfused myocardial infarction. Am J Cardiol 1991; 67(16): 1360-7. http://dx.doi.org/10.1016/0002-9149(91)90466-X PMID: 1904190
- http://dx.doi.org/10.1016/0002-9149(91)90466-X PMID: 1904190
 [33] McCarthy CP, Raber I, Chapman AR, *et al.* Myocardial injury in the era of high-sensitivity cardiac troponin assays: a practical approach for clinicians. JAMA Cardiol 2019; 4(10): 1034-42.
- http://dx.doi.org/10.1001/jamacardio.2019.2724 PMID: 31389986
 [34] Pascual-Figal DA, Casas T, Ordonez-Llanos J, *et al.* Highly sensitive troponin T for risk stratification of acutely destabilized heart failure. Am Heart J 2012; 163(6): 1002-10.

http://dx.doi.org/10.1016/j.ahj.2012.03.015 PMID: 22709753

- [35] Parissis JT, Papadakis J, Kadoglou NP, et al. Prognostic value of high sensitivity troponin T in patients with acutely decompensated heart failure and non-detectable conventional troponin T levels. Int J Cardiol 2013; 168(4): 3609-12.
- http://dx.doi.org/10.1016/j.ijcard.2013.05.056 PMID: 23711451 Nagarajan V, Hernandez AV, Tang WH. Prognostic value of cardi-
- [36] Nagarajan V, Hernandez AV, Tang WH. Prognostic value of cardiac troponin in chronic stable heart failure: A systematic review. Heart 2012; 98(24): 1778-86.
- http://dx.doi.org/10.1136/heartjnl-2012-301779 PMID: 23118345
 [37] Aimo A, Januzzi JL Jr, Vergaro G, *et al.* Prognostic value of high-sensitivity troponin T in chronic heart failure: an individual patient data meta-analysis. Circulation 2018; 137(3): 286-97. http://dx.doi.org/10.1161/CIRCULATIONAHA.117.031560
 PMID: 29335288
- [38] Chaikijurajai T, Tang WHW. Reappraisal of inflammatory biomarkers in heart failure. Curr Heart Fail Rep 2020; 17(1): 9-19. http://dx.doi.org/10.1007/s11897-019-00450-1 PMID: 31916187
- [39] Braunwald E. Biomarkers in heart failure. N Engl J Med 2008; 358(20): 2148-59.

http://dx.doi.org/10.1056/NEJMra0800239 PMID: 18480207

[40] Castell JV, Gómez-Lechón MJ, David M, Fabra R, Trullenque R, Heinrich PC. Acute-phase response of human hepatocytes: regulation of acute-phase protein synthesis by interleukin-6. Hepatology 1990; 12(5): 1179-86.

http://dx.doi.org/10.1002/hep.1840120517 PMID: 1699862

- [41] Elster SK, Braunwald E, Wood HF. A study of C-reactive protein in the serum of patients with congestive heart failure. Am Heart J 1956; 51(4): 533-41. http://dx.doi.org/10.1016/0002-8703(56)90099-0 PMID: 13302128
- [42] Araújo JP, Lourenço P, Azevedo A, et al. Prognostic value of highsensitivity C-reactive protein in heart failure: A systematic review. J Card Fail 2009; 15(3): 256-66.

http://dx.doi.org/10.1016/j.cardfail.2008.10.030 PMID: 19327628

- [43] Anand IS, Latini R, Florea VG, et al. Val-HeFT Investigators. Creactive protein in heart failure: prognostic value and the effect of valsartan. Circulation 2005; 112(10): 1428-34. http://dx.doi.org/10.1161/CIRCULATIONAHA.104.508465 PMID: 16129801
- [44] Lamblin N, Mouquet F, Hennache B, *et al.* High-sensitivity C-reactive protein: potential adjunct for risk stratification in patients with stable congestive heart failure. Eur Heart J 2005; 26(21): 2245-50.

http://dx.doi.org/10.1093/eurheartj/ehi501 PMID: 16183690

- [45] Windram JD, Loh PH, Rigby AS, Hanning I, Clark AL, Cleland JG. Relationship of high-sensitivity C-reactive protein to prognosis and other prognostic markers in outpatients with heart failure. Am Heart J 2007; 153(6): 1048-55.
- http://dx.doi.org/10.1016/j.ahj.2007.03.044 PMID: 17540208
- [46] Zelová H, Hošek J. TNF-α signalling and inflammation: interactions between old acquaintances. Inflamm Res 2013; 62(7): 641-51. http://dx.doi.org/10.1007/c00011.012.0622.0.DMID: 22685857.

http://dx.doi.org/10.1007/s00011-013-0633-0 PMID: 23685857

- [47] Levine B, Kalman J, Mayer L, Fillit HM, Packer M. Elevated circulating levels of tumor necrosis factor in severe chronic heart failure. N Engl J Med 1990; 323(4): 236-41. http://dx.doi.org/10.1056/NEJM199007263230405 PMID: 2195340
- [48] Kalogeropoulos A, Georgiopoulou V, Psaty BM, et al. Health ABC Study Investigators. Inflammatory markers and incident heart failure risk in older adults: The Health ABC (Health, Aging, and Body Composition) study. J Am Coll Cardiol 2010; 55(19): 2129-37.

http://dx.doi.org/10.1016/j.jacc.2009.12.045 PMID: 20447537

- [49] Testa M, Yeh M, Lee P, et al. Circulating levels of cytokines and their endogenous modulators in patients with mild to severe congestive heart failure due to coronary artery disease or hypertension. J Am Coll Cardiol 1996; 28(4): 964-71. http://dx.doi.org/10.1016/S0735-1097(96)00268-9 PMID: 8837575
- [50] Deswal A, Petersen NJ, Feldman AM, Young JB, White BG, Mann DL. Cytokines and cytokine receptors in advanced heart fail-

[52] Murtha LA, Schuliga MJ, Mabotuwana NS, *et al.* The processes and mechanisms of cardiac and pulmonary fibrosis. Front Physiol 2017; 8(777): 777.

http://dx.doi.org/10.3389/fphys.2017.00777 PMID: 29075197
 [53] Hinderer S, Schenke-Layland K. Cardiac fibrosis - A short review of causes and therapeutic strategies. Adv Drug Deliv Rev 2019; 14: 77-92

- 146: 77-82. http://dx.doi.org/10.1016/j.addr.2019.05.011 PMID: 31158407
- [54] Passino C, Barison A, Vergaro G, et al. Markers of fibrosis, inflammation, and remodeling pathways in heart failure. Clin Chim Acta 2015; 443: 29-38.
- http://dx.doi.org/10.1016/j.cca.2014.09.006 PMID: 25218738
 [55] Biaggi P, Ammann C, Imperiali M, *et al.* Soluble ST2-a new biomarker in heart failure. Cardiovasc Med 2019; 22: w02008. http://dx.doi.org/10.4414/cvm.2019.02008
- [56] Boisot S, Beede J, Isakson S, *et al.* Serial sampling of ST2 predicts 90-day mortality following destabilized heart failure. J Card Fail 2008; 14(9): 732-8.
- http://dx.doi.org/10.1016/j.cardfail.2008.06.415 PMID: 18995177 [57] Gaggin HK, Szymonifka J, Bhardwaj A, *et al.* Head-to-head com-
- [57] Gaggin FR, Szynonika J, Bhardwaj A, et al. Fread-to-head comparison of serial soluble ST2, growth differentiation factor-15, and highly-sensitive troponin T measurements in patients with chronic heart failure. JACC Heart Fail 2014; 2(1): 65-72. http://dx.doi.org/10.1016/j.jchf.2013.10.005 PMID: 24622120
- [58] Sharma UC, Pokharel S, van Brakel TJ, et al. Galectin-3 marks activated macrophages in failure-prone hypertrophied hearts and contributes to cardiac dysfunction. Circulation 2004; 110(19): 3121-8. http://dx.doi.org/10.1161/01.CIR.0000147181.65298.4D PMID: 15520318
- [59] de Boer RA, Voors AA, Muntendam P, van Gilst WH, van Veldhuisen DJ. Galectin-3: a novel mediator of heart failure development and progression. Eur J Heart Fail 2009; 11(9): 811-7. http://dx.doi.org/10.1093/eurjhf/hfp097 PMID: 19648160
- [60] van Kimmenade RR, Januzzi JL Jr, Ellinor PT, *et al.* Utility of amino-terminal pro-brain natriuretic peptide, galectin-3, and apelin for the evaluation of patients with acute heart failure. J Am Coll Cardiol 2006; 48(6): 1217-24.
- http://dx.doi.org/10.1016/j.jacc.2006.03.061 PMID: 16979009
 [61] Gehlken C, Suthahar N, Meijers WC, de Boer RA. Galectin-3 in heart failure: an update of the last 3 years. Heart Fail Clin 2018; 14(1): 75-92.
 - http://dx.doi.org/10.1016/j.hfc.2017.08.009 PMID: 29153203 [2] Lok DJ, Van Der Meer P, de la Porte PW, *et al.* Prognostic value
- [62] Lok DJ, Van Der Meer P, de la Porte PW, et al. Prognostic value of galectin-3, a novel marker of fibrosis, in patients with chronic heart failure: Data from the DEAL-HF study. Clin Res Cardiol 2010; 99(5): 323-8. http://dx.doi.org/10.1007/s00392-010-0125-y PMID: 20130888
- [63] Anand IS, Rector TS, Kuskowski M, Adourian A, Muntendam P, Cohn JN. Baseline and serial measurements of galectin-3 in patients with heart failure: relationship to prognosis and effect of treatment with valsartan in the Val-HeFT. Eur J Heart Fail 2013; 15(5): 511-8.
- http://dx.doi.org/10.1093/eurjhf/hfs205 PMID: 23291728 [64] Ho JE, Liu C, Lyass A, *et al.* Galectin-3, a marker of cardiac fibro-
- Cardiol 2012; 60(14): 1249-56. http://dx.doi.org/10.1016/j.jacc.2012.04.053 PMID: 22939561
- [65] Wu CK, Su MY, Lee JK, *et al.* Galectin-3 level and the severity of cardiac diastolic dysfunction using cellular and animal models and clinical indices. Sci Rep 2015; 5: 17007. http://dx.doi.org/10.1038/srep17007.PMID: 26582585
- http://dx.doi.org/10.1038/srep17007 PMID: 26582585
 [66] Edelmann F, Holzendorf V, Wachter R, *et al.* Galectin-3 in patients with heart failure with preserved ejection fraction: results from the Aldo-DHF trial. Eur J Heart Fail 2015; 17(2): 214-23. http://dx.doi.org/10.1002/ejhf.203 PMID: 25418979

- [67] de Boer RA, Lok DJ, Jaarsma T, *et al.* Predictive value of plasma galectin-3 levels in heart failure with reduced and preserved ejection fraction. Ann Med 2011; 43(1): 60-8. http://dx.doi.org/10.3109/07853890.2010.538080 PMID: 21189092
- [68] Zhou SS, Jin JP, Wang JQ, et al. miRNAS in cardiovascular diseases: Potential biomarkers, therapeutic targets and challenges. Acta Pharmacol Sin 2018; 39(7): 1073-84. http://dx.doi.org/10.1038/aps.2018.30 PMID: 29877320
- [69] Watson CJ, Gupta SK, O'Connell E, *et al.* MicroRNA signatures differentiate preserved from reduced ejection fraction heart failure. Eur J Heart Fail 2015; 17(4): 405-15. http://dx.doi.org/10.1002/ejhf.244 PMID: 25739750
- [70] Wong LL, Zou R, Zhou L, *et al.* Combining circulating MicroR-NA and NT-proBNP to detect and categorize heart failure subtypes. J Am Coll Cardiol 2019; 73(11): 1300-13. http://dx.doi.org/10.1016/j.jacc.2018.11.060 PMID: 30898206
- [71] McGarrah RW, Crown SB, Zhang GF, Shah SH, Newgard CB. Cardiovascular metabolomics. Circ Res 2018; 122(9): 1238-58. http://dx.doi.org/10.1161/CIRCRESAHA.117.311002 PMID: 29700070
- [72] Hunter WG, Kelly JP, McGarrah RW III, Kraus WE, Shah SH. Metabolic dysfunction in heart failure: diagnostic, prognostic, and pathophysiologic insights from metabolomic profiling. Curr Heart Fail Rep 2016; 13(3): 119-31. http://dx.doi.org/10.1007/s11897-016-0289-5 PMID: 27216948
- [73] Cheng ML, Wang CH, Shiao MS, et al. Metabolic disturbances identified in plasma are associated with outcomes in patients with heart failure: diagnostic and prognostic value of metabolomics. J Am Coll Cardiol 2015; 65(15): 1509-20. http://dx.doi.org/10.1016/j.jacc.2015.02.018 PMID: 25881932
- [74] Ahmad T, Kelly JP, McGarrah RW, *et al.* Prognostic implications of long-chain acylcarnitines in heart failure and reversibility with mechanical circulatory support. J Am Coll Cardiol 2016; 67(3): 291-9.
- http://dx.doi.org/10.1016/j.jacc.2015.10.079 PMID: 26796394
- [75] Hunter WG, Kelly JP, McGarrah RW III, et al. Metabolomic profiling identifies novel circulating biomarkers of mitochondrial dysfunction differentially elevated in heart failure with preserved versus reduced ejection fraction: evidence for shared metabolic impairments in clinical heart failure. J Am Heart Assoc 2016; 5(8): e003190.

http://dx.doi.org/10.1161/JAHA.115.003190 PMID: 27473038

- [76] Bristow MR, Kao DP, Breathett KK, et al. Structural and functional phenotyping of the failing heart: is the left ventricular ejection fraction obsolete? JACC Heart Fail 2017; 5(11): 772-81. http://dx.doi.org/10.1016/j.jchf.2017.09.009 PMID: 29096787
- [77] Solomon SD, Anavekar N, Skali H, et al. Candesartan in Heart Failure Reduction in Mortality (CHARM) Investigators. Influence of ejection fraction on cardiovascular outcomes in a broad spectrum of heart failure patients. Circulation 2005; 112(24): 3738-44. http://dx.doi.org/10.1161/CIRCULATIONAHA.105.561423 PMID: 16330684
- [78] Lupón J, Gavidia-Bovadilla G, Ferrer E, et al. Dynamic trajectories of left ventricular ejection fraction in heart failure. J Am Coll Cardiol 2018; 72(6): 591-601.
- http://dx.doi.org/10.1016/j.jacc.2018.05.042 PMID: 30071987
 [79] Armstrong AC, Gidding S, Gjesdal O, Wu C, Bluemke DA, Lima JA. LV mass assessed by echocardiography and CMR, cardiovascular outcomes, and medical practice. JACC Cardiovasc Imaging 2012; 5(8): 837-48.
- http://dx.doi.org/10.1016/j.jcmg.2012.06.003 PMID: 22897998
 [80] Smiseth OA, Torp H, Opdahl A, Haugaa KH, Urheim S. Myocardial strain imaging: How useful is it in clinical decision making? Eur Heart J 2016; 37(15): 1196-207.
- http://dx.doi.org/10.1093/eurheartj/ehv529 PMID: 26508168
 [81] Romano S, Mansour IN, Kansal M, *et al.* Left Ventricular global longitudinal strain predicts heart failure readmission in acute decompensated heart failure. Cardiovasc Ultrasound 2017; 15(1): 6. http://dx.doi.org/10.1186/s12947-017-0098-3 PMID: 28298230
- [82] Cho GY, Marwick TH, Kim HS, Kim MK, Hong KS, Oh DJ. Global 2-dimensional strain as a new prognosticator in patients with heart failure. J Am Coll Cardiol 2009; 54(7): 618-24.

http://dx.doi.org/10.1016/j.jacc.2009.04.061 PMID: 19660692

- [83] Park JJ, Park J-B, Park J-H, Cho G-Y. Global longitudinal strain to predict mortality in Patients With Acute Heart Failure. J Am Coll Cardiol 2018; 71(18): 1947-57. http://dx.doi.org/10.1016/j.jacc.2018.02.064 PMID: 29724346
- [84] Kaufmann D, Szwoch M, Kwiatkowska J, Raczak G, Daniłowicz-Szymanowicz L. Global longitudinal strain can predict heart failure exacerbation in stable outpatients with ischemic left ventricular systolic dysfunction. PLoS Ône 2019; 14(12): e0225829. http://dx.doi.org/10.1371/journal.pone.0225829 PMID: 31790492
- [85] Zhang KW, French B, May Khan A, et al. Strain improves risk prediction beyond ejection fraction in chronic systolic heart failure. J Am Heart Assoc 2014; 3(1): e000550. http://dx.doi.org/10.1161/JAHA.113.000550 PMID: 24419736
- Yang H, Negishi K, Wang Y, Nolan M, Saito M, Marwick TH. Echocardiographic screening for non-ischaemic stage B heart fail-[86] ure in the community. Eur J Heart Fail 2016; 18(11): 1331-9. http://dx.doi.org/10.1002/ejhf.643 PMID: 27813300
- [87] Plana JC, Galderisi M, Barac A, et al. Expert consensus for multimodality imaging evaluation of adult patients during and after cancer therapy: a report from the American Society of Echocardiography and the European Association of Cardiovascular Imaging. J Am Soc Echocardiogr 2014; 27(9): 911-39. http://dx.doi.org/10.1016/j.echo.2014.07.012 PMID: 25172399
- [88] Negishi T, Thavendiranathan P, Negishi K, Marwick TH. SUC-COUR investigators. Rationale and design of the strain surveillance of chemotherapy for improving cardiovascular outcomes: the SUCCOUR Trial. JACC Cardiovasc Imaging 2018; 11(8): 1098-105.
- http://dx.doi.org/10.1016/j.jcmg.2018.03.019 PMID: 29909105 [89] Russell K, Eriksen M, Aaberge L, et al. A novel clinical method for quantification of regional left ventricular pressure-strain loop area: A non-invasive index of myocardial work. Eur Heart J 2012; 33(6): 724-33.
 - http://dx.doi.org/10.1093/eurheartj/ehs016 PMID: 22315346
- [90] Galli E, Leclercq C, Hubert A, et al. Role of myocardial constructive work in the identification of responders to CRT. Eur Heart J Cardiovasc Imaging 2018; 19(9): 1010-8. http://dx.doi.org/10.1093/ehjci/jex191 PMID: 28954293
- van der Bijl P, Vo NM, Kostyukevich MV, et al. Prognostic impli-[91] cations of global, left ventricular myocardial work efficiency before cardiac resynchronization therapy. Eur Heart J Cardiovasc Imaging 2019; 20(12): 1388-94. http://dx.doi.org/10.1093/ehjci/jez095 PMID: 31131394
- El Mahdiui M, van der Bijl P, Abou R, Marsan NA, Delgado V, Bax JJ. Global left ventricular myocardial work efficiency in [92] healthy individuals and patients with cardiovascular disease. J Am Soc Echocardiogr 2019; 32(9): 1120-7. http://dx.doi.org/10.1016/j.echo.2019.05.002 PMID: 31279618
- [93] Hedwig F, Soltani S, Stein J, et al. Global work index correlates with established prognostic parameters of heart failure. Echocardiography 2020; 37(3): 412-20.
- http://dx.doi.org/10.1111/echo.14612 PMID: 32077524 Tsang TS, Barnes ME, Gersh BJ, Bailey KR, Seward JB. Left [94] atrial volume as a morphophysiologic expression of left ventricular diastolic dysfunction and relation to cardiovascular risk burden. Am J Cardiol 2002; 90(12): 1284-9 http://dx.doi.org/10.1016/S0002-9149(02)02864-3 PMID: 12480035
- [95] Persson H, Lonn E, Edner M, et al. Investigators of the CHARM Echocardiographic Substudy-CHARMES. Diastolic dysfunction in heart failure with preserved systolic function: need for objective evidence: Results from the CHARM Echocardiographic Substudy-CHARMES. J Am Coll Cardiol 2007; 49(6): 687-94. http://dx.doi.org/10.1016/j.jacc.2006.08.062 PMID: 17291934
- [96] Santos AB, Kraigher-Krainer E, Gupta DK, et al. PARAMOUNT Investigators. Impaired left atrial function in heart failure with preserved ejection fraction. Eur J Heart Fail 2014; 16(10): 1096-103. http://dx.doi.org/10.1002/ejhf.147 PMID: 25138249
- Gan GCH, Ferkh A, Boyd A, Thomas L. Left atrial function: eval-[97] uation by strain analysis. Cardiovasc Diagn Ther 2018; 8(1): 29-46

http://dx.doi.org/10.21037/cdt.2017.06.08 PMID: 29541609

- [98] Dell'Italia LJ. Anatomy and physiology of the right ventricle. Cardiol Clin 2012; 30(2): 167-87
- http://dx.doi.org/10.1016/j.ccl.2012.03.009 PMID: 22548810 [99] Pleister A, Kahwash R, Haas G, Ghio S, Cittadini A, Baliga RR. Echocardiography and heart failure: A glimpse of the right heart. Echocardiography 2015; 32 (Suppl. 1): S95-S107. http://dx.doi.org/10.1111/echo.12678 PMID: 25234293
- [100] Anavekar NS, Skali H, Bourgoun M, et al. Usefulness of right ventricular fractional area change to predict death, heart failure, and stroke following myocardial infarction (from the VALIANT ECHO Study). Am J Cardiol 2008; 101(5): 607-12 http://dx.doi.org/10.1016/j.amjcard.2007.09.115 PMID: 18308007
- Modin D, Møgelvang R, Andersen DM, Biering-Sørensen T. [101] Right ventricular function evaluated by tricuspid annular plane systolic excursion predicts cardiovascular death in the general population. J Am Heart Assoc 2019; 8(10): e012197. http://dx.doi.org/10.1161/JAHA.119.012197 PMID: 31088196
- [102] Guendouz S, Rappeneau S, Nahum J, et al. Prognostic significance and normal values of 2D strain to assess right ventricular systolic function in chronic heart failure. Circ J 2012; 76(1): 127-36. http://dx.doi.org/10.1253/circj.CJ-11-0778 PMID: 22033348
- [103] Berger A. Magnetic resonance imaging. BMJ 2002; 324(7328): 35

http://dx.doi.org/10.1136/bmj.324.7328.35 PMID: 11777806

- [104] Croisille P, Revel D, Saeed M. Contrast agents and cardiac MR imaging of myocardial ischemia: from bench to bedside. Eur Radiol 2006; 16(9): 1951-63. http://dx.doi.org/10.1007/s00330-006-0244-z PMID: 16633792
- [105] Judd RM, Atalay MK, Rottman GA, Zerhouni EA. Effects of myocardial water exchange on T1 enhancement during bolus administration of MR contrast agents. Magn Reson Med 1995; 33(2): 215-23
- http://dx.doi.org/10.1002/mrm.1910330211 PMID: 7707912 [106] Wesbey GE, Higgins CB, McNamara MT, et al. Effect of gadolinium-DTPA on the magnetic relaxation times of normal and infarcted myocardium. Radiology 1984; 153(1): 165-9. http://dx.doi.org/10.1148/radiology.153.1.6473778 PMID: 6473778
- Mewton N, Liu CY, Croisille P, Bluemke D, Lima JA. Assess-[107] ment of myocardial fibrosis with cardiovascular magnetic resonance. J Am Coll Cardiol 2011; 57(8): 891-903. http://dx.doi.org/10.1016/j.jacc.2010.11.013 PMID: 21329834
- [108] Assomull RG, Prasad SK, Lyne J, et al. Cardiovascular magnetic resonance, fibrosis, and prognosis in dilated cardiomyopathy. J Am Coll Cardiol 2006; 48(10): 1977-85. http://dx.doi.org/10.1016/j.jacc.2006.07.049 PMID: 17112987
- [109] Rubinshtein R, Glockner JF, Ommen SR, et al. Characteristics and clinical significance of late gadolinium enhancement by contrast-enhanced magnetic resonance imaging in patients with hypertrophic cardiomyopathy. Circ Heart Fail 2010; 3(1): 51-8. http://dx.doi.org/10.1161/CIRCHEARTFAILURE.109.854026 PMID: 19850699
- [110] Austin BA, Tang WH, Rodriguez ER, et al. Delayed hyper-enhancement magnetic resonance imaging provides incremental diagnostic and prognostic utility in suspected cardiac amyloidosis. JACC Cardiovasc Imaging 2009; 2(12): 1369-77. http://dx.doi.org/10.1016/j.jcmg.2009.08.008 PMID: 20083070
- [111] Krittayaphong R, Boonyasirinant T, Chaithiraphan V, et al. Prognostic value of late gadolinium enhancement in hypertensive patients with known or suspected coronary artery disease. Int J Cardiovasc Imaging 2010; 26 (Suppl. 1): 123-31 http://dx.doi.org/10.1007/s10554-009-9574-7 PMID: 20049536
- Kwong RY, Sattar H, Wu H, et al. Incidence and prognostic impli-[112] cation of unrecognized myocardial scar characterized by cardiac magnetic resonance in diabetic patients without clinical evidence of myocardial infarction. Circulation 2008; 118(10): 1011-20. http://dx.doi.org/10.1161/CIRCULATIONAHA.107.727826 PMID: 18725488
- Barison A, Aimo A, Mirizzi G, et al. The extent and location of [113] late gadolinium enhancement predict defibrillator shock and cardiac mortality in patients with non-ischaemic dilated cardiomyopathy. Int J Cardiol 2020; 307: 180-6.

http://dx.doi.org/10.1016/j.ijcard.2020.02.028 PMID: 32067833

- [114] Messroghli DR, Niendorf T, Schulz-Menger J, Dietz R, Friedrich MG. T1 mapping in patients with acute myocardial infarction. J Cardiovasc Magn Reson 2003; 5(2): 353-9. http://dx.doi.org/10.1081/JCMR-120019418 PMID: 12765114
- [115] Messroghli DR, Walters K, Plein S, et al. Myocardial T1 mapping: application to patients with acute and chronic myocardial infarction. Magn Reson Med 2007; 58(1): 34-40. http://dx.doi.org/10.1002/mrm.21272 PMID: 17659622
- [116] Maceira AM, Joshi J, Prasad SK, *et al.* Cardiovascular magnetic resonance in cardiac amyloidosis. Circulation 2005; 111(2): 186-93.

http://dx.doi.org/10.1161/01.CIR.0000152819.97857.9D PMID: 15630027

- [117] Thuny F, Potton L, Rapacchi S, et al. Myocardial T1-mapping for early detection of left ventricular myocardial fibrosis in systemic sclerosis. J Cardiovasc Magn Reson 2011; 13(1): M10. http://dx.doi.org/10.1186/1532-429X-13-S1-M10
- [118] Thibault H, Ernande L, Rapacchi S, et al. Early detection of myocardial fibrosis in type II diabetic patients using MR T1-mapping. J Cardiovasc Magn Reson 2011; 13(1): O110. http://dx.doi.org/10.1186/1532-429X-13-S1-O110
- [119] Ellims AH, Iles LM, Ling LH, et al. A comprehensive evaluation of myocardial fibrosis in hypertrophic cardiomyopathy with cardiac magnetic resonance imaging: linking genotype with fibrotic phenotype. Eur Heart J Cardiovasc Imaging 2014; 15(10): 1108-16.

http://dx.doi.org/10.1093/ehjci/jeu077 PMID: 24819852

- [120] Iles L, Pfluger H, Phrommintikul A, et al. Evaluation of diffuse myocardial fibrosis in heart failure with cardiac magnetic resonance contrast-enhanced T1 mapping. J Am Coll Cardiol 2008; 52(19): 1574-80.
- http://dx.doi.org/10.1016/j.jacc.2008.06.049 PMID: 19007595
 [121] Sibley CT, Noureldin RA, Gai N, *et al.* T1 Mapping in cardiomyopathy at cardiac MR: comparison with endomyocardial biopsy. Radiology 2012; 265(3): 724-32. http://dx.doi.org/10.1148/radiol.12112721 PMID: 23091172
- [122] Zhao L, Li S, Ma X, *et al.* Prognostic significance of left ventricular fibrosis assessed by T1 mapping in patients with atrial fibrillation and heart failure. Sci Rep 2019; 9(1): 13374. http://dx.doi.org/10.1038/s41598-019-49793-8 PMID: 31527757
- [123] Puntmann VO, Carr-White G, Jabbour A, et al. International T1 Multicentre CMR Outcome Study. T1-mapping and outcome in nonischemic cardiomyopathy: all-cause mortality and heart failure. JACC Cardiovasc Imaging 2016; 9(1): 40-50. http://dx.doi.org/10.1016/j.jcmg.2015.12.001 PMID: 26762873
- [124] Weber KT, Sun Y, Bhattacharya SK, Ahokas RA, Gerling IC. Myofibroblast-mediated mechanisms of pathological remodelling of the heart. Nat Rev Cardiol 2013; 10(1): 15-26. http://dx.doi.org/10.1038/nrcardio.2012.158 PMID: 23207731
- [125] Vita T, Gräni C, Abbasi SA, et al. Comparing CMR mapping methods and myocardial patterns toward heart failure outcomes in nonischemic dilated cardiomyopathy. JACC Cardiovasc Imaging 2019; 12(8 Pt 2): 1659-69.
- http://dx.doi.org/10.1016/j.jcmg.2018.08.021 PMID: 30448130
 [126] Kehr E, Sono M, Chugh SS, Jerosch-Herold M. Gadolinium-enhanced magnetic resonance imaging for detection and quantification of fibrosis in human myocardium *in vitro*. Int J Cardiovasc Imaging 2008; 24(1): 61-8. http://dx.doi.org/10.1007/s10554-007-9223-y PMID: 17429755
- [127] Coelho-Filho OR, Mongeon FP, Mitchell R, et al. Role of transcytolemmal water-exchange in magnetic resonance measurements of diffuse myocardial fibrosis in hypertensive heart disease. Circ Cardiovasc Imaging 2013; 6(1): 134-41. http://dx.doi.org/10.1161/CIRCIMAGING.112.979815 PMID: 23159497
- [128] Lu C, Qin F, Yan Y, Liu T, Li J, Chen H. Immunosuppressive treatment for myocarditis: a meta-analysis of randomized controlled trials. J Cardiovasc Med (Hagerstown) 2016; 17(8): 631-7. http://dx.doi.org/10.2459/JCM.00000000000134 PMID: 25003999
- [129] Wojnicz R, Nowalany-Kozielska E, Wojciechowska C, et al. Randomized, placebo-controlled study for immunosuppressive treat-

ment of inflammatory dilated cardiomyopathy: two-year follow-up results. Circulation 2001; 104(1): 39-45. http://dx.doi.org/10.1161/01.CIR.104.1.39 PMID: 11435335

- [130] Caforio AL, Pankuweit S, Arbustini E, et al. Current state of knowledge on aetiology, diagnosis, management, and therapy of myocarditis: a position statement of the European Society of Cardiology Working Group on Myocardial and Pericardial Diseases. Eur Heart J 2013; 34(33): 2636-48. http://dx.doi.org/10.1093/eurheartj/eht210
- [131] Spieker M, Katsianos E, Gastl M, et al. T2 mapping cardiovascular magnetic resonance identifies the presence of myocardial inflammation in patients with dilated cardiomyopathy as compared to endomyocardial biopsy. Eur Heart J Cardiovasc Imaging 2018; 19(5): 574-82.

http://dx.doi.org/10.1093/ehjci/jex230 PMID: 29136120

- [132] Spieker M, Haberkorn S, Gastl M, et al. Abnormal T2 mapping cardiovascular magnetic resonance correlates with adverse clinical outcome in patients with suspected acute myocarditis. J Cardiovasc Magn Reson 2017; 19(1): 38.
- http://dx.doi.org/10.1186/s12968-017-0350-x PMID: 28351402
 Chiotoroiu A-L, Buicu C-F, Neagu C, Benedek T. Recent advances in biomarker discovery-from serum to imaging-based biomarkers for a complex assessment of heart failure patients. J Inter-
- discip Med 2016; 1(2): 125.
 [134] Doumouras BS, Lee DS, Levy WC, Alba AC. An appraisal of biomarker-based risk-scoring models in chronic heart failure: which one is best? Curr Heart Fail Rep 2018; 15(1): 24-36.
- http://dx.doi.org/10.1007/s11897-018-0375-y PMID: 29404976
 [135] McKie PM, Burnett JC Jr. NT-proBNP: the gold standard biomarker in heart failure. J Am Coll Cardiol 2016; 68(22): 2437-9.
- http://dx.doi.org/10.1016/j.jacc.2016.10.001 PMID: 27908348 [136] Bajraktari G, Pugliese NR, D'Agostino A, *et al.* Echo- and B-
- [136] Bajraktari G, Pugliese NR, D'Agostino A, et al. Echo- and B--Type natriuretic peptide-guided follow-up versus symptom-guided follow-up: comparison of the outcome in ambulatory heart failure patients. Cardiol Res Pract 2018; 2018: 3139861. http://dx.doi.org/10.1155/2018/3139861 PMID: 30363950
- [137] Yu L, Ruifrok WP, Meissner M, *et al.* Genetic and pharmacological inhibition of galectin-3 prevents cardiac remodeling by interfering with myocardial fibrogenesis. Circ Heart Fail 2013; 6(1): 107-17. http://dx.doi.org/10.1161/CIRCHEARTFAILURE.112.971168

PMID: 23230309

- [138] Wu CK, Su MM, Wu YF, Hwang JJ, Lin LY. Combination of plasma biomarkers and clinical data for the detection of myocardial fibrosis or aggravation of heart failure symptoms in heart failure with preserved ejection fraction patients. J Clin Med 2018; 7(11): E427. http://dx.doi.org/10.3390/jcm7110427 PMID; 30413105
- [139] Marques MD, Nauffal V, Ambale-Venkatesh B, et al. Association between inflammatory markers and myocardial fibrosis. Hypertension 2018; 72(4): 902-8. http://dx.doi.org/10.1161/HYPERTENSIONAHA.118.11463 PMID: 30354713
- [140] Wu KC, Gerstenblith G, Guallar E, et al. Combined cardiac magnetic resonance imaging and C-reactive protein levels identify a cohort at low risk for defibrillator firings and death. Circ Cardiovasc Imaging 2012; 5(2): 178-86. http://dx.doi.org/10.1161/CIRCIMAGING.111.968024 PMID: 22267750
- Aronson JK. Biomarkers and surrogate endpoints. Br J Clin Pharmacol 2005; 59(5): 491-4. http://dx.doi.org/10.1111/j.1365-2125.2005.02435.x PMID: 15842546
- [142] Bianco CM, Farjo PD, Ghaffar YA, Sengupta PP. Myocardial mechanics in patients with normal LVEF and diastolic dysfunction. JACC Cardiovasc Imaging 2020; 13(1 Pt 2): 258-71. http://dx.doi.org/10.1016/j.jcmg.2018.12.035 PMID: 31202770
- [143] Januzzi JL Jr. Will biomarkers succeed as a surrogate endpoint in heart failure trials? JACC Heart Fail 2018; 6(7): 570-2. http://dx.doi.org/10.1016/j.jchf.2018.02.008 PMID: 29501808
- [144] Vaduganathan M, Claggett B, Packer M, et al. Natriuretic peptides as biomarkers of treatment response in clinical trials of heart failure. JACC Heart Fail 2018; 6(7): 564-9.

http://dx.doi.org/10.1016/j.jchf.2018.02.007 PMID: 29501807

- Anand IS, Florea VG, Fisher L. Surrogate end points in heart failure. J Am Coll Cardiol 2002; 39(9): 1414-21. http://dx.doi.org/10.1016/S0735-1097(02)01773-4 PMID: 11985901
- [146] Piek A, de Boer RA, Silljé HH. The fibrosis-cell death axis in heart failure. Heart Fail Rev 2016; 21(2): 199-211. http://dx.doi.org/10.1007/s10741-016-9536-9 PMID: 26883434
- [147] Galli E, Vitel E, Schnell F, et al. Myocardial constructive work is impaired in hypertrophic cardiomyopathy and predicts left ventricular fibrosis. Echocardiography 2019; 36(1): 74-82. http://dx.doi.org/10.1111/echo.14210 PMID: 30488501
- [148] Corrao G, Ghirardi A, Ibrahim B, Merlino L, Maggioni AP. Burden of new hospitalization for heart failure: A population-based investigation from Italy. Eur J Heart Fail 2014; 16(7): 729-36. http://dx.doi.org/10.1002/ejhf.105 PMID: 24806352