RESEARCH ARTICLE

Revised: 31 May 2022

eJHaem

British Society for Haematology

NTBI levels in C282Y homozygotes after therapeutic phlebotomy

¹Liver Centre, Mater Misericordiae University Hospital, Dublin, Ireland

²Department of Biochemistry and Diagnostic Endocrinology, Mater Misericordiae University Hospital, Dublin, Ireland

³Laboratory of Genetic, Endocrine and Metabolic Diseases, Department of Laboratory Medicine, Radboud University Medical Centre, Nijmegen, The Netherlands

Correspondence

Stephen Stewart, Liver Centre, Mater Misericordiae University Hospital, Dublin, Ireland. Email: sstewart@mater.ie

Abstract

C282Y homozygotes exposed to sustained elevated transferrin saturation (TS) may develop worsening clinical symptoms. This might be related to the appearance of nontransferrin bound iron (NTBI) when TS≥50% and labile plasma iron (LPI) when TS levels reach 75-80%. In this study, NTBI levels were examined in 219 randomly selected untreated and treated C282Y homozygotes. Overall, 161 of 219 had TS \geq 50%, 124 of whom had detectable NTBI (\geq 0.47 µM, 1.81 µM [0.92–2.46 µM]) with a median serum ferritin 320 μ g/L (226-442 μ g/L). Ninety of 219 homozygotes had TS > 75%, and all had detectable NTBI (2.21 µM [1.53-2.59 µM] with a median ferritin 338 µg/L [230-447 μ g/L]). Of 125 homozygotes who last had phlebotomy >12 months ago (42 months [25-74 months], 92 had TS levels > 50%, and 70 of these had NTBI > 0.47 μ M (2.06 μ M [1.23-2.61µM]). Twenty-six of these 70 had a normal ferritin. Fifty-five of 125 had TS \geq 75%, and NTBI was detected in all of these (2.32 μ M [1.57–2.77 μ M]) with a median ferritin 344 µg/L (255–418 µg/L). Eighteen of these 55 had a normal ferritin. In summary, NTBI is frequently found in C282Y homozygotes with TS \geq 50%. Furthermore, C282Y homozygotes in the maintenance phase often have TS \geq 50% together with a normal ferritin. Therefore, monitoring the TS level during the maintenance phase is recommended as an accessible clinical marker of the presence of NTBI.

KEYWORDS

ferritin, hereditary haemochromatosis, non-transferrin bound iron, phlebotomy, transferrin saturation

1 | INTRODUCTION

In Ireland, 93% of individuals with hereditary haemochromatosis (HH) are homozygous for the C282Y mutation in the *HFE* gene [1, 2]. C282Y homozygotes are deficient in the liver-derived hormone hepcidin, leading to increased intestinal iron absorption and systemic iron overload [3–6].

Phlebotomy treatment lowers hepcidin by normalising serum ferritin levels, but phlebotomy can also increase iron absorption, further lowering hepcidin via erythropoiesis [4, 7–9]. However, normalising the ferritin level does not guarantee that transferrin saturation (TS) levels are under control as TS levels are frequently elevated in the maintenance phase despite a normal ferritin [10, 11]. Sustained exposure to TS \geq 50% is associated with a worsening of arthritis in HH

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. eJHaem published by British Society for Haematology and John Wiley & Sons Ltd.

individuals on long-term maintenance and may be related to increased levels of non-transferrin bound iron (NTBI) [12–15].

When TS levels exceed 75%–80%, the redox active component of NTBI, labile plasma iron (LPI) is found [16, 17]. LPI is highly reactive and damages organs through the formation of free radicals and lipid peroxidation [11, 15]. NTBI induced by sustained exposure to high TS levels may also predispose individuals to morbidities associated with aging such as stroke, diabetes and joint problems [18].

We have previously examined hepcidin and NTBI in a cohort comprising untreated non-iron loaded and iron loaded C282Y homozygotes, compound heterozygotes (heterozygous for C282Y and H63D) and others with less at risk of iron overload genotypes. We found that C282Y homozygotes had significantly lower hepcidin levels and significantly higher NTBI levels compared to those with the 'less at risk of iron overload' genotypes [6]. We also reported a strong correlation of TS with NTBI [6].

While studies have shown that TS levels are often high in the maintenance phase even in the presence of a normal ferritin, there is a lack of information regarding NTBI levels following therapeutic phlebotomy. Consequently, the aim of this study was to examine NTBI in untreated and treated C282Y homozygotes and to look at its relationship with TS, hepcidin and other iron indices. We also sought to explore the relationship of NTBI with obesity, alcohol and joint pain.

2 | MATERIALS AND METHODS

2.1 | Participants

Fasting serum samples were randomly collected (8.30 am to 11 am) between February 2017 and November 2019, from 136 male and 83 female C282Y homozygous patients attending the Liver Centre and stored at -80° C. Patients were identified via family screening or presented with symptoms/raised iron indices and were diagnosed from 1986 to 2019. Phlebotomy treatment was initiated when the serum ferritin was raised (\geq 300 µg/l for males and \geq 200µg for females) and entailed weekly venesection until the ferritin was in the normal range (<300 µg/l for males and <200 µg for females, [16]) with a target of \leq 100 µg/L. Once depleted, ferritin levels are monitored to ensure they remain within the normal reference range before phlebotomy recommences (maintenance phase). The study received approval from the ethics committee of the Mater Misericordiae University Hospital, and informed consent was obtained from all participants.

Body mass index (BMI) was measured and participants completed a self-administered questionnaire, from which information regarding alcohol consumption, joint pain and phlebotomy history were obtained. In Ireland, the low risk alcohol limit is <17 and <11 standard drinks per week for men and women, respectively (one standard drink contains 10 g pure alcohol). Additional phlebotomy information was obtained from the hospital record system.

2.2 | Laboratory measurements

Serum iron, transferrin, glucose, C-reactive protein (CRP), liver function tests, lipids, haemoglobin (Hgb) and serum ferritin were measured using routine methods. *HFE* genotyping was performed using LightCycler technology (Roche Diagnostics) with Genes-4U toolsets (Ratiogen, AG).

Serum hepcidin was measured using the high sensitivity enzyme immunoassay ELISA kit (Hepcidin 25 (bioactive), EIA-5782, DRG Diagnostics, Marburg, Germany). Thawed samples (one thaw only) were analysed for hepcidin between April and September 2019. Low and high controls provided in the kit were run with each calibration curve, and results were considered invalid if the assay did not fit the acceptable ranges of the low and high controls. The assay range is between 0.153 ng/ml and 81 ng/ml. The limit of detection is 0.304 ng/ml. NTBI was measured using a nitrilotriacetic (NTA) assay [19]. The lower limit of detection was <0.47 μ M. For those samples that received a <0.47 μ M value, a value of 0.46 was entered onto the SPSS database. NTBI was measured from October to December 2020.

2.3 Statistical analyses

All data were tested for normality using the Kolmogorov-Smimov test. Non-normal data are presented as the median (interquartile range [25th-75th percentile]), while normal data are presented as means \pm standard deviation. Differences between groups were calculated using the non-parametric Mann-Whitney *U* test. Relationships between variables were assessed using Spearman r. Statistical evaluation was carried out using IBM SPSS for Windows, version 25 (IBM Corp., Armonk, NY, USA).

3 | RESULTS

3.1 | Baseline characteristics

The main characteristics of the study cohort are presented in Table 1. The 219 C282Y homozygotes were diagnosed between 1986 and 2019. For those who underwent phlebotomy treatment, the median end of treatment ferritin was 131 µg/L (96–168 µg/L) and 165 µg/L (130-200 µg/L) for males and females, respectively. Twenty-seven males and 34 females never had phlebotomy treatment, while 21 males and 12 females had phlebotomy within the 12 months prior to the study, while 88 males and 37 females last had phlebotomy \geq 12 months prior to the study. The 12-month cut off point was chosen because iron absorption has been shown to be increased in the 12 months following phlebotomy treatment, and phlebotomy can further lower hepcidin via erythropoiesis [4, 7–9, 20].

⁶⁴⁶ WILEY-

TABLE 1 Characteristics of male and female C282Y homozygotes

| Variables* | Male | Female | Mann-Whitney U test p-value |
|--|---------------------------|---------------------------|-----------------------------|
| Age (years) | 53.8 ± 14.1 | 52.7 ± 14.0 | 0.742 |
| Serum iron (µmol/l) | 33.9 ± 8.9 | 29.0 ± 8.4 | <0.001 |
| Transferrin (g/l) | 1.92 ± 0.27 | 1.91 ± 0.37 | 0.658 |
| Transferrin saturation (%) | 75.4 (53.2-85.9) | 59.6 (42.6-75.2) | <0.001 |
| Hepcidin (ng/ml) | 10.2 (7.1–14.9) | 12.0 (6.3-14.9) | 0.422 |
| Serum ferritin (µg/l) | 271 (160-397) | 230 (73-333) | 0.024 |
| Hepcidin:ferritin ratio | 0.042 (0.027-0.063) | 0.059 (0.039-0.103) | <0.001 |
| ΝΤΒΙ (μΜ) | 0.95 (0.46-2.25) | 0.53 (0.46-1.32) | 0.009 |
| Hgb (g/dl) | 15.6 (14.6-16.2) | 14.3 (13.7-14.9) | <0.001 |
| Bilirubin (µmol/L) | 13 (10-17) | 10 (8-13) | <0.001 |
| ALT (I.U./L) | 28.0 (21-39.3) | 20 (16-30) | <0.001 |
| γGT (I.U./L) | 30 (22-44.0) | 20 (15-27) | <0.001 |
| AST (I.U./L) | 29 (25-36) | 26 (22-31) | 0.001 |
| Alkp (I.U./L) | 74 (64-86) | 70 (59-90) | 0.326 |
| Glucose (mmol/L) | 4.9 (4.6-5.3) | 4.8 (4.4–5.2) | 0.148 |
| C-reactive protein (CRP) (mg/L) | 2.0 (1.0-3.0) | 2.0 (1.0-4.0) | 0.344 |
| Cholesterol (mmol/L) | 4.8 ± 0.9 | 5.0 ± 0.8 | 0.132 |
| Triglycerides (mmol/L) | 1.31 (0.91-1.77) | 1.18 (0.84–1.57) | 0.059 |
| HDL (mmol/L) | 1.21 (1.06-1.44) | 1.50 (1.31-1.81) | <0.001 |
| LDL (mmol/L) | 2.90 (2.20-3.40) | 2.85 (2.20-3.50) | 0.892 |
| Joint pain (no/yes) | 31/41 | 19/20 | |
| Body mass index (BMI) | 27.0 (24.5-29.6) (n = 84) | 26.3 (23.5-29.3) (n = 52) | 0.306 |
| Alcohol (std drinks/week) | 7.5 (2–15) (n = 110) | 3.5 (0.75-7.3) (n = 66) | 0.001 |
| Phlebotomy | (<i>n</i>) | (n) | |
| Never had phlebotomy | 27 | 34 | |
| <12 months since last phlebotomy | 21 | 12 | |
| \geq 12 months ago since last phlebotomy | 88 | 37 | |

*All values are those determined at the time the sample was taken and are not diagnostic values. Data were tested for normality using the Kolmogorov-Smimov test. Non-normal data are presented as the median (25th–75th percentile), while normal data are presented as means \pm standard deviation. Differences between groups were calculated using the non-parametric Mann-Whitney *U* test. Data refer to the date on which the blood sample was taken. Note: 49 males and 40 females had NTBI <0.47 μ M (not detected). A value of 0.46 μ M represents undetected NTBI.

3.2 | Hepcidin levels in non-iron loaded and iron loaded male and female C282Y homozygotes

C282Y homozygotes with a raised ferritin regardless of phlebotomy status had higher hepcidin levels and lower hepcidin : ferritin ratios compared to those with a normal ferritin (Figures 1 and 2).

Overall, hepcidin was significantly higher in C282Y homozygotes with a raised ferritin (for their gender) (n = 109), 13.10 ng/ml (10.00–18.22 ng/ml) compared to those with a normal ferritin (n = 110) 8.30 ng/ml (4.82–11.86 ng/ml), p < 0.001. The hepcidin : ferritin ratio was significantly lower in C282Y homozygotes with a raised ferritin, 0.032 (0.021–0.048) compared to those with a normal ferritin (0.068 [0.047–0.109]), p < 0.001.

3.3 | TS and NTBI levels in male and female C282Y homozygotes

C282Y homozygotes with a raised ferritin regardless of phlebotomy status had higher TS levels compared to those with a normal ferritin (Figure 3).

Male and female C282Y homozygotes with NTBI \geq 0.47 μM (the level at which NTBI becomes detectable), regardless of phlebotomy status, or whether the ferritin was normal or raised, had higher TS levels (Tables 2 and 3).

Overall, TS levels were significantly higher in those with NTBI \ge 0.47 μ M (n = 130), 82.3% (72.9%-87.0%) compared to those with NTBI < 0.47 μ M (n = 89), 47.6% (40.4%-56.4%), p < 0.001.

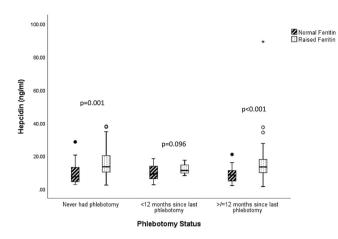


FIGURE 1 Boxplots of hepcidin levels in C282Y homozygotes according to phlebotomy status. A normal serum ferritin was defined as a serum ferritin $<300 \,\mu$ g/l for males and $<200 \,\mu$ g for females. The line in the middle of the boxplot is the median. The top and bottom box lines show the 75th percentile and 25th percentile. The whiskers show the maximum and minimum values. Outliers (circles) are at least 1.5 box lengths from the median, while extremes (asterisks) are at least 3 box lengths from the median

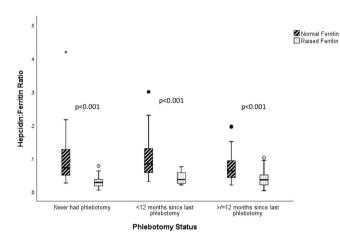


FIGURE 2 Boxplots of hepcidin:ferritin ratios in C282Y homozygotes according to phlebotomy status. A normal serum ferritin was defined as a serum ferritin <300 µg/l for males and <200 µg for females. The line in the middle of the boxplot is the median. The top and bottom box lines show the 75th percentile and 25th percentile. The whiskers show the maximum and minimum values. Outliers (circles) are at least 1.5 box lengths from the median, while extremes (asterisks) are at least 3 box lengths from the median

In the cohort overall (n = 219), 161/219 (74%, 105 males and 56 females) had TS \geq 50%, 124 of whom (77%) had NTBI \geq 0.47 μ M (1.81 μM [0.92–2.46 μM]) with a median serum ferritin 320 $\mu g/L$ (226–442 μ g/L). Ninety of 219 (41%, 68 males and 22 females) had TS \geq 75% with a median serum ferritin 338 $\mu g/L$ (230–447 $\mu g/L),$ and all had NTBI \geq 0.47 μ M (2.21 μ M [1.53–2.59 μ M]). Ten per cent of those with a TS <50% (6 of 58) and 31% of those with a TS < 75% (40 of 129) had NTBI \geq 0.47 μ M.

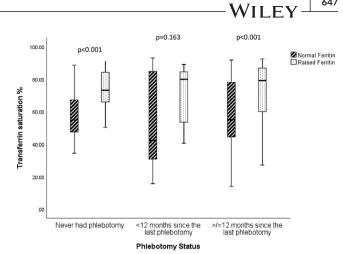


FIGURE 3 Boxplots of transferrin saturation levels in C282Y homozygotes according to phlebotomy status. A normal serum ferritin was defined as a serum ferritin $<300 \,\mu$ g/l for males and $<200 \,\mu$ g for females. The line in the middle of the boxplot is the median. The top and bottom box lines show the 75th percentile and 25th percentile. The whiskers show the maximum and minimum values

3.4 \mid TS levels of >50% and >75% and the presence of NTBI in those who last had phlebotomy \geq 12 months ago

In the cohort of 125 male and female C282Y homozygotes who last had phlebotomy \geq 12 months ago (42.0 months [25–74 months]), 92 (74%, 67 males and 25 females) had TS levels \geq 50% and 70 of these (76%) had NTBI \geq 0.47 μ M (2.06 μ M [1.23–2.61 μ M]). Thirty-seven per cent of these (26/70, 24 males and two females) had a normal ferritin. Fortyfour per cent (55/125, 44 males and 11 females) had TS \geq 75%, and all had NTBI \geq 0.47 µM (2.32µM [1.57–2.77 µM]). Thirty-three per cent (18/55) of these (17 males and one female) had a normal ferritin.

3.5 Iron parameters, obesity, alcohol and joint pain

BMI was available for 84 males and 52 females. Seventeen of 84 males (20%) and 10 of 52 females (19%) were obese (BMI \geq 30). No significant differences in TS, hepcidin, ferritin or hepcidin : ferritin ratio were noted between those who were obese and those who were not obese nor when analysed according to phlebotomy status. But, TS levels tended to be lower in obese homozygotes (males, p = 0.070; females, p = 0.057). Forty-seven per cent (8/17) obese males had NTBI \geq 0.47 μ M compared to 67.2% (45/67) of the non-obese cohort, while 25% (4/10) obese females had NTBI \geq 0.47 μ M compared to 60% (25/42) of the non-obese cohort.

110 males and 66 females responded to the questionnaire regarding alcohol intake. Twenty-one of 110 males (19%) and eight of 66 females (12%) had an alcohol intake above that recommended for their gender. Serum iron levels were significantly higher in males who drank \geq 17 units alcohol per week (37.6 μ mol/l \pm 7.6 μ mol/l) compared to those

647

| | | Never had phlebotomy | otomy | | <12 months since last phlebotomy | last phlebotomy | | ≥12 months since last phlebotomy | last phlebotomy | |
|------------------|----------------------------|-----------------------------------|-----------------------------------|------------------|----------------------------------|------------------|------------------|----------------------------------|------------------|-------------------|
| | | Overall | Normal Ferritin | Raised Ferritin | Overall | Normal Ferritin | Raised Ferritin | Overall | Normal Ferritin | Raised Ferritin |
| | | n = 7 | n = 4 | $n = 3^{*}$ | n = 9 | n = 9 | n = 0 | n = 33 | n = 26 | n = 7 |
| NTBI < 0.47 | Age (years) | 56.1 ± 15.9 | 55.8 ± 15.1 | 56.7 ± 20.3 | 53.9 ± 15.4 | 53.9 ± 15.4 | | 58.0 ± 12.7 | 55.4 ± 12.4 | 66.6 ± 10.8 |
| (μM) (n = 49) | Serum iron (µmol/l) | 30.1 ± 5.5 | 27.7±5.2 | 33.3 ± 4.8 | 19.0 ± 6.2 | 19.0 ± 6.2 | | 26.7 ± 6.6 | 25.8 ± 6.6 | 30.1 ± 6.31 |
| | Trans Sat (%) | 61.3 (54.1-68.6) | 61.3 (54.1–68.6) 54.5 (41.6–65.3) | 66.7 (61.3- | 38.0 (23.0-44.0) | 38.0 (23.0-44.0) | | 48.3 (43.5-56.7) | 47.1 (41.7-54.0) | 57.1 (53.1-68.9) |
| | Transferrin (g/l) | 1.98 ± 0.28 | 2.01 ± 0.35 | 1.94 ± 0.20 | 2.15 ± 0.25 | 2.15 ± 0.25 | | 2.07 ± 0.26 | 2.10 ± 0.26 | 1.94 ± 0.21 |
| | Hepcidin (ng/ml) | 13.4 (4.9–20.6) | 9.2 (3.3-18.8) | 19.1 (8.5– | 9.2 (4.1–13.5) | 9.2 (4.1-13.5) | | 9.2 (4.6-11.8) | 7.1 (4.5–10.2) | 16.8 (11.4-18.2) |
| | Ferritin (µg/I) | 273 (74–922) | 139 (61–256) | 922 (418– | 158 (38-218) | 158 (38-218) | | 147 (63–258) | 95 (55-186) | 398 (313-536) |
| | Hepcidin:Ferritin Ratio | 0.05 (0.02-0.07) | 0.05 (0.02-0.07) 0.06 (0.05-0.09) | 0.02 (0.02- | 0.07 (0.06-0.12) | 0.07 (0.06–0.12) | | 0.06 (0.04-0.10) | 0.07 (0.04-0.11) | 0.04 (0.03-0.04) |
| | | n = 20 | n = 4 | n = 16 | n = 12 | n = 7 | n = 5 | n = 55 | n = 26 | n = 29 |
| NTBI ≥ 0.47 | Age (years) | 44.1 ± 14.0 | 39.5 ± 21.5 | 45.3 ± 12.2 | 53.4 ± 16.1 | 48.1 ± 14.6 | 60.8 ± 16.5 | 54.6 ± 13.0 | 55.3 ± 11.4 | 54.1 ± 11.4 |
| (μM) (n = 87) | Serum iron (µmol/l) | 38.8±6.8 | 39.9±6.4 | 38.5 ± 7.1 | 39.6±5.9 | 40.3 ± 7.2 | 38.5 ± 3.8 | 38.0±5.8 | 36.1 ± 5.7 | 39.8 ± 5.4 |
| | Trans Sat (%) | 82.9 (73.5-88.5) | 83.7 (76.7-87.5) | 82.7 (72.5-89.1) | 85.5 (82.6-88.2) | 85.9 (83.6-88.3) | 85.0 (70.9-87.5) | 82.2 (77.3-88.2) | 78.7 (64.6-86.1) | 85.8 (79.8-88.9) |
| | Transferrin (g/l) | 1.84 ± 0.19 | 1.84 ± 0.21 | 1.85 ± 0.19 | 1.80 ± 0.22 | 1.78 ± 0.28 | 1.84 ± 0.12 | 1.83 ± 0.26 | 1.84 ± 0.30 | 1.83 ± 0.23 |
| | Hepcidin (ng/ml) | 13.2 (9.8–22.2) | 7.6 (6.0–11.0) | 16.2 (10.6–29.3) | 9.9 (8.2-13.2) | 9.9 (7.2-13.9) | 10.0 (9.4–14.5) | 10.2 (7.5-14.8) | 9.6 (7.1–11.8) | 11.8 (8.2-18.0) |
| | Ferritin (µg/I) | 533 (313-976) | 225 (206-275) | 676 (382-1110) | 263 (143-416) | 207 (76-244) | 439 (335-590) | 310 (228-389) | 224 (161–257) | 388 (349-554) |
| | Hepcidin:Ferritin Ratio | 0.03 (0.01-0.04) 0.03 (0.03-0.04) | 0.03 (0.03-0.04) | 0.03 (0.01-0.04) | 0.04 (0.03-0.06) | 0.06 (0.05-0.10) | 0.03 (0.02-0.03) | 0.04 (0.02-0.05) | 0.04 (0.03-0.06) | 0.03 (0.02-0.05) |
| | NTBI (µM) | 1.38 (0.77-2.05) | 1.38 (0.77-2.05) 1.25 (0.65-1.89) | 1.44 (0.77-2.19) | 2.31 (1.70-2.85) | 2.37 (1.63-2.74) | 2.26 (1.47–2.89) | 2.06 (1.26-2.60) | 1.62 (0.61–2.40) | 2.44 (1.59-2.83) |

TABLE 2 Characteristics and serum iron indices according to a normal or elevated ferritin, phlebotomy status and NTBI level in male C282Y homozygotes

⁶⁴⁸ │ WILEY

| | | Never had phlebotomy | tomy | | <12 months since | <12 months since last phlebotomy | | ≥12 months since last phlebotomy | last phlebotomy | |
|------------------|----------------------------|----------------------|-----------------------------------|------------------|------------------|----------------------------------|------------------|----------------------------------|------------------|------------------|
| | | Overall | Normal ferritin | Raised ferritin | Overall | Normal ferritin | Raised ferritin | Overall | Normal ferritin | Raised ferritin |
| | | n = 16 | n = 12 | n = 4 | n=6 | $n = 3^{*}$ | n = 3 | n = 18 | n = 7 | n = 11 |
| NTBI < 0.47 | Age (years) | 46.7 ± 12.7 | 46.5 ± 14.6 | 47.3 ± 5.0 | 55.3 ± 11.8 | 45.3 ± 8.3 | 61.3 ± 9.4 | 59.9 ± 10.1 | 60.1 ± 7.8 | 59.7 ± 11.6 |
| (μM) (n = 40) | Serum iron (µmol/1) | 27.6 ± 6.2 | 26.5 ± 7.0 | 27.4±3.4 | 20.3±3.6 | 19.2 ± 4.6 | 21.4 ± 2.9 | 23.5 ± 6.3 | 21.7±6.6 | 24.8 ± 6.1 |
| | Trans Sat (%) | 51.5 (42.9-59.6) | 51.5 (42.9-59.6) 48.5 (39.4-56.1) | 64.2 (53.3-66.8) | 40.4 (28.9-41.4) | 30.8 (23.1–) | 41.0 (40.5–) | 46.8 (38.4-54.7) | 45.0 (27.8-52.7) | 50.4 (38.6-60.9) |
| | Transferrin (g/l) | 2.02 ± 0.43 | 2.12 ± 0.45 | 1.73 ± 0.21 | 2.18 ± 0.36 | 2.38 ± 0.38 | 1.98 ± 0.21 | 1.95 ± 0.24 | 1.96 ± 0.28 | 1.95 ± 0.23 |
| | Hepcidin (ng/ml) | 11.1 (5.3–15.9) | 9.6 (4.5-15.9) | 12.3 (10.5–17.9) | 13.7 (6.3–17.3) | 6.3 (6.2–) | 14.9 (12.5–) | 13.1 (8.6-17.1) | 7.9 (4.7-11.3) | 14.6 (13.0–27.6) |
| | Serum iron (µmol/l) | 27.6 ± 6.2 | 26.5 ± 7.0 | 27.4±3.4 | 20.3±3.6 | 19.2 ± 4.6 | 21.4 ± 2.9 | 23.5 ± 6.3 | 21.7 ± 6.6 | 24.8 ± 6.1 |
| | Ferritin (µg/l) | 155 (64–235) | 132 (42-163) | 336 (262-534) | 157 (35–290) | 40 (21-) | 289 (49-) | 256 (103-321) | 68 (65-121) | 291 (260-379) |
| | Hepcidin:Ferritin Ratio | 0.09 (0.05-0.13) | 0.11 (0.07-0.14) | 0.04 (0.03-0.05) | 0.11 (0.06-0.19) | 0.16 (0.15-) | 0.06 (0.05–) | 0.07 (0.05-0.11) | 0.09 (0.07-0.12) | 0.05 (0.04–0.10) |
| | | n = 18 | n = 7 | n = 11 | n = 6 | $n = 2^{*}$ | n = 4 | n = 19 | n = 3 | n = 16 |
| NTBI ≥ 0.47 | Age (years) | 44.4 ± 14.8 | 36.1 ± 13.4 | 49.6 ± 13.6 | 57.0 ± 8.2 | 58.0 ± 1.4 | 56.5 ± 10.5 | 57.3 ± 14.8 | 66.3 ± 11.6 | 55.6 ± 15.0 |
| (μM) (n = 43) | Serum iron (µmol/1) | 30.8 ± 4.6 | 30.3±6.0 | 31.1 ± 3.8 | 33.6 ±9.2 | 29.7 ± 16.9 | 35.6 ± 5.6 | 35.5 ± 9.5 | 33.7 ± 16.0 | 35.8 ± 8.5 |
| | Trans Sat (%) | 66.2 (62.2-84.7) | 64.3 (50.7–86.2) | 73.1 (62.4-84.2) | 81.3 (67.2-85.1) | 59.5 (30.4–) | 81.3(79.6-83.5) | 79.7 (64.3-86.6) | 70.4 (28.2-) | 80.0 (65.1-87.6) |
| | Transferrin (g/l) | 1.73 ± 0.33 | 1.84 ± 0.40 | 1.67 ± 0.28 | 1.78 ± 0.30 | 2.01 ± 0.30 | 1.67 ± 0.25 | 1.92 ± 0.41 | 2.22 ± 0.49 | 1.86 ± 0.39 |
| | Hepcidin (ng/ml) | 10.5 (4.5-13.7) | 4.7 (3.6-10.1) | 12.8 (8.1–14.9) | 10.5 (6.8-14.8) | 9.4 (2.6–) | 10.5 (8.6–13.5) | 12.6 (9.1–16.9) | 5.8 (3.9-) | 13.0 (10.2-17.0) |
| | Ferritin (μ g/l) | 242 (66-371) | 62 (24-71) | 335 (253-443) | 208 (60-250) | 49 (26-) | 218 (205-305) | 295 (248-424) | 68 (20-) | 333 (273-441) |
| | Hepcidin:Ferritin Ratio | 0.05 (0.03-0.08) | 0.09 (0.05-0.22) | 0.03 (0.02-0.05) | 0.06 (0.04-0.13) | 0.17 (0.10–) | 0.05 (0.03-0.06) | 0.04 (0.02-0.08) | 0.20 (0.09–) | 0.04 (0.02-0.06) |
| | NTBI (µM) | 0.96 (0.80-1.37) | 0.96 (0.80-1.37) 0.83 (0.61-1.17) | 1.00 (0.83-1.81) | 2.21 (1.73-2.54) | 1.86 (0.66–) | 2.21 (2.11-2.34) | 1.34 (0.69-2.60) | 0.69 (0.47–) | 1.87 (0.76-2.75) |

TABLE 3 Characteristics and serum iron indices according to a normal or elevated ferritin, phlebotomy status and NTBI level in female C282Y homozygotes

RYAN ET AL.

the non-parametric Mann-Whitney U test. * SPSS does not generate 75th percentile for cohorts of 2 or 3 individuals.

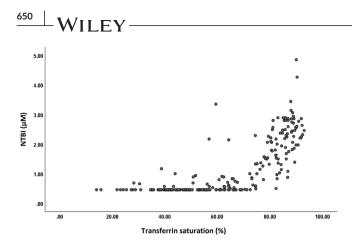


FIGURE 4 Scatterplot of NTBI versus transferrin saturation for the entire cohort. Spearman correlation r = 0.839, p < 0.001, n = 219

who drank <17 units per week (33.0 µmol/l ± 8.6 µmol/l), *p* = 0.009. No other significant differences were noted. But, TS levels tended to be higher in those who drank alcohol at levels beyond that recommended (males, *p* = 0.065; females, *p* = 0.377). Eighty-one per cent (17/21) of males who drank ≥17 units per week had NTBI ≥ 0.47 µM compared to 61% (54/89) of those who drank <17 units per week. Fifty per cent (4/8) of females who drank ≥11 units per week had NTBI ≥ 0.47 µM compared to 52% (30/58) of females who drank <11 alcohol units per week.

One hundred and eleven individuals responded to the questionnaire regarding joint pain, 41 of 72 (57%) of males and 20 of 39 (51%) of females responded yes to the query about whether they had joint pain (Table 1). Those who reported joint pain were older (55.5 \pm 11.8 years vs. 50.0 \pm 14.9 years, p = 0.059 for males; 60.7 \pm 10.3 years vs. 46.0 \pm 16.2 years, p = 0.004 for females). No other statistical differences were noted for serum iron, TS, ferritin or hepcidin levels in those males/females who reported joint pain compared to those who did not. Twenty-six of 41 (63%) males and 10 of 20 (50%) females who reported having joint pain had NTBI \geq 0.47µM.

3.6 Correlations

In the cohort overall, NTBI correlated strongly with TS (Spearman r = 0.839, p < 0.001; Figure 4) and serum iron (Spearman r = 0.747, p < 0.001) and less strongly with ferritin (Spearman r = 0.429, p < 0.001). Ferritin also correlated with hepcidin (Spearman r = 0.541, p < 0.001), TS (Spearman r = 0.512, p < 0.001) and serum iron (Spearman r = 0.426, p < 0.001).

4 DISCUSSION

In this study of C282Y homozygotes, NTBI was present in 77% of patients who had TS \geq 50% and in 100% of patients with TS \geq 75% irrespective of treatment status. Previous studies have suggested that it can be taken for granted that the potentially harmful component of

NTBI, LPI is present at TS levels of 70%–90%, so LPI is also most likely present in the sera of our C282Y homozygotes [15–17, 21].

Thirty-one per cent of those with a TS level < 75% and 10% of those with a TS level < 50% had NTBI \geq 0.47 μ M demonstrating that NTBI can appear in the absence of fully saturated transferrin, which is consistent with other studies [22–24].

In agreement with other reports, TS was found to be frequently elevated in the maintenance phase [17, 18]. In the cohort who last had phlebotomy at least a year prior to the study, 74% had TS \geq 50%, and 76% of these had detectable NTBI, while 44% had TS \geq 75%, and all had detectable NTBI. This represents a significant proportion of C282Y homozygotes who may have been exposed to sustained elevated TS and NTBI, and at levels of TS \geq 75%, prolonged exposure to LPI. Sustained exposure to TS \geq 50% for more than 6 years and to TS \geq 75% for more than 8 months has been associated with a worsening of metacarpophalangeal and proximal interphalangeal joint symptoms, which are specifically associated with HH [12]. The current study did not look at sustained exposure to high TS levels but it is hoped that further studies will address this.

Thirty-seven per cent and 33% of those with TS \geq 50% and TS \geq 75%, respectively, who last had phlebotomy at least a year prior to the study had a normal ferritin for their gender demonstrating that a normal ferritin is no guarantee that the TS level is under control, which is in agreement with other studies that have found elevated TS levels in the maintenance phase despite normal body iron stores [10, 11].

Our data might suggest that because of the risk of a sustained exposure to high TS levels and thus the probability of the LPI being present, that phlebotomy treatment should ensure that TS is maintained < 50%. However, this must be tempered with the fact that excessive phlebotomy treatment may result in iron deficiency [18, 25]. An alternative approach may involve the use of therapies that correct hepcidin deficiency, which would be beneficial in the maintenance phase to control TS and thus NTBI and consequently ameliorate the long-term complications in HH patients [11, 18, 26].

Our data confirm that ferritin levels play a significant role in determining hepcidin levels regardless of phlebotomy status demonstrating that C282Y homozygotes retain some ability to up-regulate hepcidin in response to iron overload [4, 9]. However, the significantly lower hepcidin : ferritin ratios in the iron loaded cohorts show that this response is entirely inadequate. Clearly, while phlebotomy results in the removal of excess iron, this does not restore the inadequate physiological response of hepcidin to iron overload in C282Y homozygotes [11].

Obese individuals had higher hepcidin and lower TS levels (although not significantly), which may be consistent with low grade inflammation that is associated with a high BMI [27–30]. We have previously found significantly higher hepcidin levels in overweight male C282Y homozygotes indicating perhaps that hepcidin is up-regulated in response to low grade inflammation, which may partly protect against the inadequate production of hepcidin (arising from the mutation in HFE) and thereby modulating phenotypic expression of HH [6]. The observation that a lower percentage of obese homozygotes had detectable NTBI is also in keeping with the lower TS noted in

 $/\text{ILEY}^{\perp 651}$

this cohort and a previously described decreased iron burden in overweight female C282Y homozygotes [28, 29].

Males whose alcohol intake was \geq 17 standard units per week had higher TS and NTBI levels (although again not significantly) which have previously been reported in individuals who drink alcohol in excess and are associated with a greater incidence of liver cirrhosis [23, 31]. Ferritin levels also tended to be higher in those who drank alcohol to excess in agreement with previous reports [32]. In view of the double assault of C282Y homozygosity and alcohol on the liver, further investigations to clarify the pathogenic role of NTBI are warranted in this cohort.

Those who reported joint pain were significantly older, but our study of joint pain in HH is limited because of our reliance on self-administered questionnaires and our lack of information on the type of joint pain as has been previously highlighted [12, 33]. In addition, we have only a one point measurement of NTBI, and the development of joint pain may arise from long-term exposure to high iron levels.

While our data demonstrate that NTBI is always present at TS levels \geq 75%, the clinical relevance of this is largely unknown. A single measurement of NTBI only represents the previous 24–48 h so repeated measurements are required to ascertain toxicity potential and indeed the exposure time to NTBI [17, 34]. Furthermore, the application of clinical assays for NTBI is limited because of the lack of standardisation between the methods used. However, high TS levels can be used as a surrogate marker for the presence of NTBI at TS 70%–90% [16, 17].

Despite the fact that high TS levels are frequently found in the maintenance phase, international guidelines do not currently recommend its measurement in the follow-up maintenance phase [35, 36]. However, in light of the data presented in this study demonstrating that high TS is associated with the appearance of NTBI, together with data that have shown that sustained exposure to high TS levels is associated with increased reports of fatigue and joint symptoms, it may be advisable to include the measurement of TS during the maintenance phase as has recently been recommended [37].

In conclusion, we have found raised NTBI levels in HH patients with raised TS and normal ferritins, the impact of which is unclear. Further studies are required to delineate the clinical significance of NTBI (in particular the clinical significance of sustained exposure to LPI) as the presence of NTBI has been associated with cardiovascular disease, diabetes, arthritis and an increased risk of cirrhosis, which are all elements of the HH clinical picture.

ACKNOWLEDGEMENTS

This work was supported by a grant from the Mater Foundation (the official fundraising body of the Mater Misericordiae University Hospital, Dublin) (grant number MG157). The grant covered the cost of purchasing the Elisa kits used for the measurement of hepcidin. We thank Deirdre Mazzone, Eva Vaughan and Caroline Walsh for help with the collection of blood samples and recruiting of patients.

CONFLICT OF INTEREST

The authors declare no competing financial interest. DWS is an employee of Radboudumc that offers analysis of iron biomarkers and

reference material for a fee for service via its hepcidinanalysis.com initiative.

ETHICS STATEMENT

The study received approval from the ethics committee of the Mater Misericordiae University Hospital, and informed written consent was obtained from all participants.

AUTHOR CONTRIBUTIONS

E R designed the study, performed the hepcidin assay, analysed the data and wrote the paper. K M performed the hepcidin assay. E W performed NTBI measurements. J R performed HFE testing. S S and D W S reviewed the data and critically revised the paper.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ORCID

Eleanor Ryan ^b https://orcid.org/0000-0002-2383-5866 Dorine W. Swinkels ^b https://orcid.org/0000-0002-1040-9446 Stephen Stewart ^b https://orcid.org/0000-0002-0865-2930

REFERENCES

- Feder JN, Gnirke A, Thomas W, Tsuchihashi DA, Ruddy A, Basava F, et al. (1996). A novel MHC class I-like gene is mutated in patients with Hereditary Hemochromatosis. Nat Genet, 13, 399–408.
- 2. Ryan E, O'Keane C, & Crowe J (1998). Hemochromatosis in Ireland and HFE. Blood Cells Mol Dis, 24, 428–432.
- Bridle KR, Frazer DM, Wilkins SJ, Dixon JL, Purdie DM, Crawford DH, et al. (2003). Disrupted hepcidin regulation in *HFE*-associated Hemochromatosis and the liver as a regulator of body iron homeostasis. Lancet, 361, 669–673.
- van Dijk BAC, Laarakkers CMM, Klaver SM, Jacobs EMG, van Tits LJH, Janssen MCH, et al. (2008). Serum hepcidin levels are innately low in *HFE* related Hemochromatosis but differ between C282Yhomozygotes with elevated and normal ferritin levels. Br J Haematol, 142, 979–985
- Nemeth E, Tuttle MS, Powelson J, Vaughan MB, Donovan A, Ward DM, et al. (2004). Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. Science, 306, 2090– 2093.
- Ryan E, Ryan JD, Russell J, Coughlan B, Tjalsma H, Swinkels DW, et al. (2015). Correlates of hepcidin and NTBI according to HFE status in patients referred to a liver centre. Acta Haematol, 133, 155–161.
- Girelli D, Trombini P, Busti F, Campostrini N, Sandri M, Pelucchi S, et al. (2011). A time course of hepcidin response to iron challenge in patients with HFE and TFR2 hemochromatosis. Haematologica, 96, 500–506.
- Mast AE, Schlumpf KS, Wright DJ, Johnson B, Glynn SA, Busch MP, et al., for the NHLBI Retrovirus Epidemiology Donor Study II (REDS-II).(2013). Hepcidin level predicts haemoglobin concentration in individuals undergoing repeated phlebotomy. Haematologica, 98, 1324–1330.
- Piperno A, Girelli D, Nemeth E, Trombini P, Bozzini C, Poggiali E, et al. (2007). Blunted response to oral iron challenge in *HFE*-related hemochromatosis. Blood, 110, 4096–4100.
- Brissot P, Cavey T, Ropert M, Guggenbuhl P, & Loréal O (2017). Clinical management of hemochromatosis: Current perspectives. Int J Clin Transfus Med, 5, 1–7.

⁶⁵² WILEY

- Loréal O, Cavey T, Robin F, Kenawi M, Guggenbuhl P, & Brissot P (2018). Iron as a therapeutic target in HFE-related hemochromatosis: Usual and novel aspects. Pharmaceuticals (Basel), 11, 131.
- Bardou-Jacquet E, Lainé F, Guggenbuhl P, Morcet J, Jezequel C, Guyader D, et al. (2017). Worse outcomes of patients with HFE hemochromatosis with persistent increases in TS during maintenance therapy. Clin Gastroenterol Hepatol, 15, 1620–1627.
- Brissot P, Ropert M, Le Lan C, & Loréal O (2012). Non-transferrin bound iron: A key role in iron overload and iron toxicity. Biochim Biophys Acta, 1820, 403–410.
- Gutteridge JM, Rowley DA, Griffiths E, & Halliwell B (1985). Lowmolecular-weight iron complexes and oxygen radical reactions in idiopathic haemochromatosis. Clin Sci (Lond), 68, 463–467.
- Le Lan C, Loréal O, Cohen T, Ropert M, Glickenstein H, Lainé F, et al. (2005). Redox active plasma in C282Y/C282Y haemochromatosis. Blood, 105, 4527-4531.
- Brissot P, Brissot E, Loréal O, & Ropert M (2020). Laboratory medicine and iron overload: Diagnostic and therapeutic aspects. J Lab Precis Med, 5, 25.
- de Swart L, Hendriks JC, van der Vorm LN, Cabantchik ZI, Evans PJ, Hod EA, et al. (2016). Second international round robin for the quantification of serum non-transferrin-bound iron and labile plasma iron in patients with iron-overload disorders. Haematologica, 101, 38–45.
- Smit SL, Peters TMA, Gisbertz IAM, Moolenaar W, Hendriks Y, Vincent HH, et al. (2018). Variable workup calls for guideline development for type 2A hereditary haemochromatosis. Neth J Med, 76, 365–373.
- Zhang D, Okada S, Kawabata T, & Yasuda T (1995). An improved simple colorimetric method for quantitation of non-transferrin-bound iron in serum. Biochem Mol Biol Int, 35, 635–641.
- Williams R, Manenti F, Williams HS, & Pitcher CS (1966). Iron absorption in idiopathic haemochromatosis before, during and after venesection therapy. Br Med J (Clin Res Ed), 2, 78–81.
- Pootrakul P, Breuer W, Sametband M, Sirankapracha P, Hershko C, & Cabantchik ZI (2004). Labile plasma iron (LPI) as an indicator of chelatable plasma redox activity in iron-overloaded beta-thalassemia/HbE patients treated with an oral chelator. Blood, 104, 1504–1510.
- Breuer W, Hershko C, & Cabantchik ZI (2000). The importance of nontransferrin bound iron in disorders of iron metabolism. Transfus Sci, 23, 185–192.
- De Feo TM, Fargion S, Duca L, Cesana BM, Boncinelli L, Lozza P, et al. (2001). Non-transferrin-bound iron in alcohol abusers. Alcohol: Clin Exp Res, 25, 1494–1499.
- de Valk B, Addicks MA, Gosriwatana I, Lu S, Hider RC, & Marx JJ (2000). Non-transferrin-bound iron is present in serum of hereditary haemochromatosis heterozygotes. Eur J Clin Invest, 30, 248–251.
- Brissot P, Pietrangelo A, Adams PC, De Graaff B, McLaren CE, & Loréal O (2018). Haemochromatosis. Nat Rev Dis Primers, 4, 1–15.

- 26. Casu C, Nemeth E, & Rivella S (2018). Hepcidin agonists as therapeutic tools. Blood, 131, 1790–1794.
- Cheng HL, Bryant C, Cook R, O'Connor H, Rooney K, & Steinbeck K (2012). The relationship between obesity and hypoferraemia in adults: A systematic review. Obes Rev, 13, 150–161.
- Desgrippes R, Lainé F, Morcet J, Perrin M, Manet G, Jezequel C, et al. (2013). Decreased iron burden in overweight C282Y homozygous women: Putative role of increased hepcidin production. Hepatology, 57, 1784–1792.
- Lainé F, Jouannolle A-M, Morcet J, Brigand A, Pouchard M, Lafraise B, et al. (2005). Phenotypic expression in detected C282Y homozygous women depends on body mass index. J Hepatol, 43, 1055–1059.
- Marmur J, Beshara S, Eggertsen G, Onelöv L, Albiin N, Danielsson O, et al. (2018). Hepcidin levels correlate to liver iron content, but not steatohepatitis, in non-alcoholic fatty liver disease. BMC Gastroenterol, 18, 78.
- Ioannou GN, Weiss NS, & Kowdley KV (2007). Relationship between transferrin-iron saturation, alcohol consumption, and the incidence of cirrhosis and liver cancer. Clin Gastroenterol Hepatol, 5, 624– 629.
- 32. Harrison-Findik DD (2007). Role of alcohol in the regulation of iron metabolism. World J Gastroenterol, 13, 4925–4930.
- Whalen NL, & Olynyk JK (2017). Association of TS with the arthropathy of hereditary haemochromatosis. Clin Gastroenterol Hepatol, 15, 1507–1508.
- Wood JC (2014). Guidelines for quantifying iron overload. Hematology 2014, the American Society of Hematology Education Program Book, 1, 210–215.
- Bacon BR, Adams PC, Kowdley KV, Powell LW, & Tavill AS & American Association for the Study of Liver Diseases. (2011) Diagnosis and management of hemochromatosis: 2011 practice guideline by the American Association for the Study of Liver Diseases. Hepatology, 54, 328–43.
- Pietrangelo A, Deugnier Y, Dooley J, Erhardt A, Zoller H, & Safadi R (2010). EASL clinical practice guidelines for HFE Haemochromatosis. J Hepatol, 53, 3–22.
- Brissot P, & Brissot E (2020). What's important and new in Hemochromatosis? Clin Hemat Int, 2, 143–148.

How to cite this article: Ryan E, Mulready K, Wiegerinck E, Russell J, Swinkels DW, Stewart S. NTBI levels in C282Y homozygotes after therapeutic phlebotomy. eJHaem. 2022;3:644–652. https://doi.org/10.1002/jha2.507