



Genome Sequence of *Microbacterium* sp. Strain R1, Isolated from a *Synechococcus* Culture

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ABSTRACT Synechococcus cultures in the laboratory are often associated with heterotrophic bacteria. Here, we report the genome sequence of the bacterium *Microbacterium* sp. strain R1, isolated from a culture of the estuarine *Synechococcus* strain CBW1107. Several secondary metabolites and transporter-related genes were identified in the genome of *Microbacterium* sp. strain R1.

S ynechococcus spp. are important primary producers in the marine environment, and heterotrophic bacteria can utilize dissolved organic matter released from *Synechococcus* spp. in the surrounding waters (1). Meanwhile, *Synechococcus* spp. also depend on the remineralization of dissolved organic matter (DOM) by heterotrophic bacteria to provide essential nutrients (2, 3). Therefore, interactions between *Synechococcus* and associated bacteria play important roles in the ocean biogeochemical cycle (4). The heterotrophic bacterial strain R1 was isolated from a culture of *Synechococcus* sp. strain CBW1107 (5). To obtain strain R1, CBW1107 was grown in SN15 medium (6) to the exponential phase; then, 100 μ l of the algal solution was spread onto agar rich organic (RO) plates and incubated at 28°C for 1 to 2 days. A single colony was transferred to a new agar plate for further purification. Phylogenetic analysis based on 16S rRNA gene sequencing revealed that strain R1 belongs to the genus *Microbacterium*, which is widely distributed in both seawater and deep-sea sediment (7, 8). Here, we present the draft genome sequence of *Microbacterium* sp. strain R1.

Microbacterium sp. strain R1 was grown in RO medium at 28°C to the exponential phase, and bacterial cells were collected by centrifugation (10,000 \times g, 10 min); then, bacterial genomic DNA was obtained from the cells following the phenol-chloroform method (9). A library was prepared using the TruSeq DNA library prep kit (Illumina, San Diego, CA, USA). Whole-genome sequencing was performed on the Illumina HiSeq 4000 sequencing system using HiSeq 3000/4000 sequencing-by-synthesis (SBS) kits (Illumina), following the format for an average 2 \times 250-bp paired-end library. Low-quality (score, \leq 20) raw reads, N nucleotide-containing reads, and filtered sequences less than 100 bp were removed to obtain clean reads using Trimmomatic V0.33 software. A total of 2,972.45 Mbp reads (clean data) were assembled using the IDBA algorithm (10). The assembled draft genome sequence has a total size of 3,986,645 bp in 21 contigs and a GC content of 68.20%. The N_{50} value and the average coverage (BBMap V38.92) for the contigs are 2,408,454 bp and $967.483 \times$, respectively. Open reading frame (ORF) prediction and genome annotation were performed using the RAST server (http://rast.nmpdr.org) (11). A total of 3,929 protein-coding genes, 47 tRNAcoding genes, and 4 rRNA genes were predicted in the draft genome of Microbacterium sp. strain R1. Of the 3,929 ORFs, 2,369 (60.30%) have predicted functions. Default parameters were used for all software unless otherwise noted.

Potential secondary metabolites of *Microbacterium* sp. strain R1 were predicted using antiSMASH bacterial version. Four genes encoding nonribosomal peptide synthase, polyketide synthase type III, terpenes, and β -lactones were identified (12, 13). The genome of this strain contains 278 genes related to ABC transporters.

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Received 31 May 2021 Accepted 16 August 2021 Published 9 September 2021 **Data availability.** The genome sequence of *Microbacterium* sp. strain R1 has been deposited in GenBank under accession no. JADDUD000000000, where the Prokaryotic Genome Annotation Pipeline (PGAP) annotation is also available. The raw sequence data are available under SRA accession no. SRR14683051. The BioProject accession no. is PRJNA667551, and the BioSample accession no. is SAMN16377710.

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REFERENCES

- Azam F, Fenchel T, Field JG, Gray JS, Meyer-Reil LA, Thingstad F. 1983. The ecological role of water-column microbes in the sea. Mar Ecol Prog Ser 10:257–263. https://doi.org/10.3354/meps010257.
- Christie-Oleza JA, Sousoni D, Lloyd M, Armengaud J, Scanlan DJ. 2017. Nutrient recycling facilitates long-term stability of marine microbial phototroph-heterotroph interactions. Nat Microbiol 2:17100. https://doi.org/10 .1038/nmicrobiol.2017.100.
- Christie-Oleza JA, Scanlan DJ, Armengaud J. 2015. "You produce while I clean up," a strategy revealed by exoproteomics during *Synechococcus– Roseobacter* interactions. Proteomics 15:3454–3462. https://doi.org/10.1002/ pmic.201400562.
- Zheng Q, Wang Y, Lu J, Lin W, Chen F, Jiao N. 2020. Metagenomic and metaproteomic insights into photoautotrophic and heterotrophic interactions in a *Synechococcus* culture. mBio 11:e03261-19. https://doi.org/10 .1128/mBio.03261-19.
- Fucich D, Xu Y, Sosa A, Zhang R, Jiao N, Chen F. 2021. Complete genome sequence of Chesapeake Bay winter *Synechococcus* sp. strain CBW1107, a member of subalpine cluster II. Microbiol Resour Announc 10:e01399-20. https://doi.org/10.1128/MRA.01399-20.
- Xu Y, Jiao N, Chen F. 2015. Novel psychrotolerant picocyanobacteria isolated from Chesapeake Bay in the winter. J Phycol 51:782–790. https://doi .org/10.1111/jpy.12318.
- Wu Y-H, Zhou P, Cheng H, Wang C-S, Wu M, Xu X-W. 2015. Draft genome sequence of Microbacterium profundi Shh49^T, an actinobacterium isolated

from deep-sea sediment of a polymetallic nodule environment. Genome Announc 3:e00642-15. https://doi.org/10.1128/genomeA.00642-15.

- Kim KK, Lee KC, Oh H-M, Lee J-S. 2008. *Microbacterium aquimaris* sp. nov., isolated from seawater. Int J Syst Evol Microbiol 58:1616–1620. https://doi .org/10.1099/ijs.0.65763-0.
- Kan J, Wang K, Chen F. 2006. Temporal variation and detection limit of an estuarine bacterioplankton community analyzed by denaturing gradient gel electrophoresis (DGGE). Aquat Microb Ecol 42:7–18. https://doi.org/10 .3354/ame042007.
- Peng Y, Leung HCM, Yiu SM, Chin FYL. 2012. IDBA-UD: a de novo assembler for single-cell and metagenomic sequencing data with highly uneven depth. Bioinformatics 28:1420–1428. https://doi.org/10.1093/bioinformatics/bts174.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olsen R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Yue C, Liu N, Liu M, Lü Y, Shao M, Wang M, Ai G, Huang Y. 2015. Tandem expression in E. coli of type III PKS and P450 genes from marine *Streptomyces olivaceus* FXJ 7.023 gives production of phenol and indole. World J Microbiol Biotechnol 31:541–548. https://doi.org/10.1007/s11274-015-1825-2.
- Ansari MZ, Yadav G, Gokhale RS, Mohanty D. 2004. NRPS-PKS: a knowledgebased resource for analysis of NRPS/PKS megasynthases. Nucleic Acids Res 32: W405–W413. https://doi.org/10.1093/nar/gkh359.