# **CASE REPORT**

**Open Access** 

# Identification of a novel *KAT6A* variant in an infant presenting with facial dysmorphism and developmental delay: a case report and literature review

Soyoung Bae<sup>1†</sup>, Aram Yang<sup>2†</sup>, Jinsup Kim<sup>1\*</sup>, Hyun Ju Lee<sup>1</sup> and Hyun Kyung Park<sup>1</sup>

# Abstract

**Background:** Arboleda-Tham syndrome (ARTHS), caused by a pathogenic variant of *KAT6A*, is an autosomal dominant inherited genetic disorder characterized by various degrees of developmental delay, dysmorphic facial appearance, cardiac anomalies, and gastrointestinal problems.

**Case presentation:** A baby presented multiple facial deformities including a high arched and cleft palate, with philtral ridge and vermilion indentation, a prominent nasal bridge, a thin upper lip, low-set ears, an epicanthal fold, and cardiac malformations. Whole exome sequencing (WES) revealed a heterozygous nonsense mutation in exon 8 of the *KAT6A* gene (c.1312C>T, p.[Arg438\*]) at 2 months of age. After a diagnosis of ARTHS, an expressive language delay was observed during serial assessments of developmental milestones.

**Conclusions:** In this study, we describe a case with a novel *KAT6A* variant first identified in Korea. This case broadens the scope of clinical features of ARTHS and emphasizes that WES is necessary for early diagnosis in patients with dysmorphic facial appearances, developmental delay, and other congenital abnormalities.

Keywords: Case report, Facial dysmorphism, Cleft palate, KAT6A, Arboleda-Tham syndrome

# Background

Arboleda-Tham syndrome (ARTHS, MIM 616268) is a newly defined, rare genetic disease caused by a pathogenic variant of *KAT6A* (MIM 601408) with autosomal dominant inheritance [1, 2]. Exome sequencing in patients with syndromic features associated with developmental delay and intellectual disability has revealed several causative pathogenic variants of *KAT6A* that are involved in the regulation of transcriptional activity and

<sup>1</sup> Department of Pediatrics, Hanyang University Medical Center, Hanyang University College of Medicine, 222-1, Wangshimri-ro, Sungdong-gu, Seoul 04763, Republic of Korea

Full list of author information is available at the end of the article



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.gr/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.gr/licenses/by/4.0/. The CreativeCommons Public Domain Dedication waiver (http://creativecommons.gr/licenses/by/4.0/. The CreativeCommons.gr/licenses/by/4.0/. The CreativeCommons Public Domain Dedication waiver (h

gene expression [1, 3–10]. The *KAT6A* gene, located on chromosome 8p11.21 and composed of 18 exons (KAT6A-201; ENST00000396930.3), encodes lysine (K) acetyltransferase 6A (KAT6A), which participates in chromatin remodeling, transcriptional regulation, and cellular replication [11, 12]. The major features of this syndrome include developmental delay, facial dysmorphism, microcephaly, cardiac anomalies, and gastrointestinal problems [5]. The frequent dysmorphic facial appearances in ARTHS include a prominent nasal bridge with a broad nasal tip, a thin-tented upper lip, and lowset ears [5, 7]. Most patients with pathogenic *KAT6A* variants have intellectual disabilities and speech delays that range from mild to severe. The genotype–phenotype analysis demonstrated that the truncating frameshift or

<sup>\*</sup>Correspondence: pinees@gmail.com

<sup>&</sup>lt;sup>†</sup>Soyoung Bae and Aram Yang contributed equally to this work

nonsense variant within the penultimate and ultimate exons (16 and 17, respectively) is associated with severe developmental delay [5, 7, 13].

Here, we identified a novel likely pathogenic variant of *KAT6A* using whole exome sequencing (WES) analysis in an infant with a dysmorphic facial appearance, cleft palate, and minor cardiac malformations at the age of two months before a developmental delay was apparent. Early diagnosis with ARTHS such a young age, in which only facial dysmorphism was apparent before presenting with developmental delay, improved her cognitive outcome with intensive rehabilitation therapy.

# Case presentation Methods

## Whole exome sequencing and Sanger sequencing

WES was performed in 3billion, Inc. (Seoul, South Korea) using a NovaSeq platform (Illumina, San Diego, CA, USA). Exome capture was performed with a Twist Human Core Exome Kit (Twist Bioscience, San Francisco, CA, USA). Raw genome sequencing data analyses included alignment to the reference sequence (National Center for Biotechnology Information genome assembly GRCh37; accessed in February of 2009). The mean read depth was 100-fold and 99.2% of the target region was covered. Variant calling, annotation, and prioritization were performed as previously described [14]. Clinical symptoms of cleft lip and palate (HP:0000202), facial dysmorphism (HP:0001999), and cardiac anomalies (HP:0001627) chosen for variant filtering of causative genes [15]. The identified variant was validated by Sander sequencing.

# Case report and result

A newborn girl from healthy and non-consanguineous Korean parents was transferred to the neonatal intensive care unit at birth because of respiratory distress and desaturation. The infant was delivered at 40+2 weeks. The birth weight of the child was 3140 g (- 0.14 standard deviation [SD]), length was 52.3 cm (1.68 SD), and head circumference was 32.5 cm (-1.17)SD). At birth, her mother and father were both 37 years old. Prenatal examinations, including non-invasive screening for aneuploidies, did not reveal abnormalities. During a one-month hospitalization for respiratory distress syndrome and persistent pulmonary hypertension, she presented with craniofacial deformities, including a high arched and cleft palate with philtral ridge and vermilion indentation, and a subtle dysmorphic facial appearance with a prominent nasal bridge, a thin-tented upper lip, low-set ears, and epicanthal fold (Fig. 1d). She showed no neurological abnormalities, such as hypotonia or spasticity, during hospitalization. Results of laboratory tests, including newborn screening tests for inborn errors of metabolism using tandem mass spectrometry, were normal. Although a thyroid hormone profile showed mild elevation of thyroidstimulating hormone and free thyroxine, both recovered within one month. Minimal cleft lip with alveolar cleft was confirmed with facial computed tomography at age of three weeks (Fig. 1a-c). Depending on the size of the defect, the patient may require cheiloplasty at around age seven for cleft palate, which was classified as a unilateral incomplete form. Atrial septal defect (ASD) and small size (0.77 mm) patent ductus arteriosus (PDA) were observed via echocardiography three days after birth. The ASD closed on its own within one month. The device or surgical closure of PDA was not considered because of its small size requiring regular echocardiography. Results of the automated auditory brainstem response test, abdominal ultrasound, and ophthalmic examination were normal. Her brain magnetic resonance imaging, performed at three weeks of age, showed no structural abnormalities.

She had a normal karyotype of 46, XX by high resolution (550-band level) karyotyping. WES was conducted when the patient was two months of age to search for pathogenic variants of genes causative of dysmorphic facial appearance, cleft palate, and cardiac anomalies (see Additional file 1). The analysis showed a novel de novo heterozygous nonsense variant, Chr8:41834577G>A, KAT6A:NM\_001305878.1:c.1312 C>T:p.(Arg438\*), in exon 8 of the *KAT6A* gene (Fig. 2a). The KAT6A variant was confirmed by Sanger sequencing. This nonsense was expected to cause protein truncation through nonsense-mediated mRNA, resulting in the loss of normal protein function. This variant was not detected in the several public genomic databases, such as The Genome Aggregation Database, the Exome Aggregation Consortium, 1000 Genomes, and the Exome Sequencing Project. Altogether, this variant was annotated as "likely pathogenic" according to the 2015 American College of Medical Genetics guidelines [16].

After diagnosis of ARTHS, developmental screening tests at the age of four months (Korean developmental screening test for infants and children; The Bayley Infant Neurodevelopmental Screener, 3<sup>rd</sup> edition) did not indicate any developmental delays. However, a comprehensive developmental evaluation at the age of eight months with the Bayley Scales of Infant and Tod-dler Development, 3<sup>rd</sup> edition (Bayley-III) found a mild language delay. Currently, the patient received a variety of intensive rehabilitation therapies, twice a week.



Fig. 1 Facial computed tomography (CT) and photograph of the patient. a Axial, b coronal, and c three-dimensional reconstruction non-contrast CT images at three-weeks-of-age show unilateral bony defect (arrow) of the maxilla hard palate and mixed dentition. d Photograph of the patient

# **Discussion and conclusions**

Lysine acetyltransferase 6A, KAT6A, a member of the MYST family of proteins, forms a histone acetyltransferase complex and plays an essential role in regulating transcriptional activity and gene expression [17]. KAT6A is also involved in senescence regulation, neural stem cell proliferation, and cardiac septum development [12, 18]. Therefore, misregulation of KAT6A may result in tumorigenesis, intellectual disability, or congenital heart disease [18]. Since a recurrent translocation was noted in acute monocytic leukemia [19], pathogenic variants and misregulation of the human *KAT6A* gene have been identified in solid tumors and patients with syndromic disease with developmental disorders of variable severity [1, 2, 18, 20]. As WES has become a powerful means of investigating de novo pathogenic variants in neurodevelopmental disorders, the number of patients identified with novel genetic *KAT6A* variants has gradually increased [5, 7, 8, 13, 21, 22].



Since patients with syndromic developmental delay due to pathogenic variants of KAT6A were first reported in 2015, nearly 100 cases of ARTHS had delineated variable presentation, with a continuously expanding extent of symptoms [1, 2, 5, 7, 13]. We performed a literature review of 89 patients carrying de novo variants in KAT6A, including 34 frameshift mutations, 41 nonsense mutations, 9 missense mutations, 4 splicing mutations, and 1 deletion mutation (Table 1) [3-10]. Most of the reported mutations are loss of function and more than half of variants are located in the acidic domain (Fig. 2c) [5]. While the patients with late-truncating variants (defects in exon 16 or 17) displayed moderate-to-severe intellectual disabilities, early- truncations (exons 1-15) of KAT6A were mild [4, 5, 7]. The variant of this case is located in the HAT domain (Fig. 2b), which corresponds to an early truncating pathogenic variant. The phenotypic manifestations of ARTHS, including microcephaly, hypotonia, gastrointestinal problems and congenital heart defects, might be subtle in patients with early-truncating variants [5, 7].

The spectrum of phenotypes with *KAT6A* variants includes neurological, craniofacial, gastrointestinal, cardiac, and ocular features. Neurological manifestations, such as developmental delay, hypotonia, microcephaly, and seizure, are main characteristic features of patients with ARTHS [4, 7]. All reported cases showed various degrees of developmental delays with frequent involvement of speech and expressive language (Table 1). In addition to speech delay, microcephaly, facial dysmorphism with a cleft palate, and cardiac problems were evident in this case. Microcephaly has been reported in 41.0% of patients (Table 1); however, it was less common in early-truncating variants [5, 7]. Most patients with ARTHS have a prominent nasal bridge with a broad nasal tip, a thin tented upper lip, and low-set, posteriorly

	Kennedy et al. [5] (total 76)	Urreizti et al. [7]. (total 5)	Trinh et al. [4] (total 2)	Marji et al. [8] (total 1)	Alkhateeb et al. [6] (total 1)	Efthymiou et al. [3] (total 1)	Lin et al. [ <mark>9</mark> ] (total 1)	Jiang et al. [10] (total 1)	Patient	Overall (total 89)
			Number of su	ıbjects (%)						
Genotype										
Frameshift										
Early truncating	10	0	0	0	0	0	0	0	0	10/89 (11.2%)
Late truncating	19	2	0	<i>—</i>	_	0	0	-	0	24/89 (27.0%)
Nonsense										
Early truncating	7	0	0	0	0	0	<del>,</del>	0	-	9/89 (10.1%)
Late truncating	29	2	0	0	0	-	0	0	0	32/89 (36.0%)
Missense	9	-	2	0	0	0	0	0	0	9/89 (10.1%)
Splicing, late truncating	4	0	0	0	0	0	0	0	0	4/89 (4.5%)
Deletion, late truncating	-	0	0	0	0	0	0	0	0	1/89 (1.1%)
Phenotype										
Neurological problems										
Microcephaly	22	5	2	<i>—</i>	0	-	<i>—</i>	1	-	34/83 (41.0%)
Developmental delay (speech)	71	5	2	<del>,</del>	-	-	<del>,</del>	1	-	84/84 (100%)
Neonatal hypotonia	57	2	-	QN	-	-	<del>.                                    </del>	0	0	63/86 (73.4%)
Craniofacial problems										
Broad/prominent nasal tip	57	4	2	QN	0	-	<i>—</i>	0	-	66/79 (83.5%)
Low-set ear	17	4	0	QN	0	0	<i>—</i>	0	-	23/40 (57.5%)
Cleft palate	e	0	0	ND	0	0	0	0	-	4/40 (10.0%)
Congenital heart problem										
ASD	25	2	0	ND	0	0	0	0	-	28/86 (32.6%)
VSD	9	0	0	ND	0	0	0	0	0	6/86 (7.0%)
PDA	14	0	0	ND	0	0	0	0	-	15/86 (17.4%)
Other problems										
Feeding difficulty	56	5	-	ND	-	ND	QN	ND	0	63/80 (78.7%)
Skeletal anomaly <sup>*</sup>	22	4	2	<del>,</del>	DN	ND	ND	ND	0	29/31 (93.5%)
* Skeletal anomalies include clinodactvi	v. camptodactvlv. bra	schvdactalv. arthro	avposis, scoliosis,	torticollis. kvph	osis. craniosynostos	is. and congenital hi	o dvsplasia			

 Table 1
 The genotypes and phenotypes of Arboleba-Tham syndrome [3–10]

Percentages were calculated excluding unknown numbers. ND, not described; ASD, atrial septal defect, VSD, ventricular septal defect; PDA, patent ductus arteriosus

rotated ears (Table 1). Other craniofacial features may be present, including frontal bossing, bitemporal narrowing, epicanthic folds, short and flat philtrums, and mild micrognathia [5, 7, 13]. Among craniofacial deformities, craniosynostosis was reported in 10% of patients with ARTHS, and cleft palate was noted in only three patients (Table 1) [5, 8]. Feeding difficulties were observed in 78.7% of patients with ARTHS (Table 1). Reflux, food allergies, and intestinal malrotation can also occur [5, 7, 22]. However, in patients with the early-truncating variant, gastrointestinal symptoms are less frequent [7]. Nearly half of patients with KAT6A variants have congenital heart disease, with ASD noted most frequently, followed by PDA (Table 1). The absence of gastrointestinal symptoms and minor cardiac anomaly shown in this case might be associated with the location of the variant. Other previously reported clinical features are strabismus, visual defects, joint hypermobility, syndactyly, supernumerary nipples, and cryptorchidism [7, 8, 20]. As more symptoms and comorbidities of ARTHS are discovered each time a new case is reported, it will be important to verify the genetic background of each case and to rule out the existence of other mild causal variants.

The salient features of this case included dysmorphic facial appearance, cleft palate, and congenital heart disease, which are major features that strongly suggest multiple congenital abnormalities [23-26]. As WES successfully identifies de novo, rare, and truncating genomic variants of several genes in undiagnosed cases with neurodevelopmental disorders or multiple congenital anomalies [21, 24, 27, 28]. Based on the cases reported and the database of the Deciphering Developmental Disorders Study, as many as 1% of undiagnosed syndromic developmental delays involve pathogenic KAT6A variants [1, 13]. WES also unravels the molecular basis of syndromic developmental delay associated with congenital neutropenia, food allergies, or multiple pituitary hormone deficiencies with malformation of the pituitary gland in patients with KAT6A variants [29-31]. Because of its broad phenotype spectrum, it is difficult to diagnose ARTHS based on clinical symptoms alone.

Genetic diagnosis at a younger age is important in syndromic cases because special treatments or surveillance programs are available for major symptoms. Most patients diagnosed with ARTHS have speech delays and intellectual disabilities, and early assessment of developmental delays can improve prognoses through proper treatment according to patient age [3-6]. In the present case, contrary to previous reports of patients with ARTHS, the genetic origin of the infant's syndromic features was identified before an expressive language delay of mild degree developed at the age of eight months. At the time of diagnosis, the infant was only two months old, which is the youngest age reported to date, and developmental delays were not discernible at that age. She currently receives rehabilitation treatments twice a week and regular check-ups with Bayley-III at fourmonth intervals, which are expected to improve her cognitive impairment. As over 50% of patients experienced ophthalmologic abnormalities, including strabismus and visual abnormalities, regular visual assessments are required [5, 7]. Skeletal surveys are also needed because skeletal anomalies, including scoliosis, torticollis, and kyphosis, were found in 29 patients (Table 1).

Even though the role of KAT6A has not been clearly identified, the number of patients found to have de novo pathogenic variants of KAT6A continues to increase. Therefore, clinicians must be familiar with the various phenotypic features of ARTHS for the diagnosis by exome sequencing. In our case, because a genetic diagnosis was confirmed at an early age, the unexpressed features that may develop later in life can be monitored. Moreover, further study of the variant is important since there had been no functional studies to confirm that this variant was truly deleterious. This report broadens the scope of clinical features of ARTHS that might be an unrevealed genetic cause of neurodevelopmental disorder and facial dysmorphism. WES is useful to identify any associated genetic disorders and provide personalized care for patients with facial dysmorphism and other congenital anomalies possibly associated with developmental delay.

#### Abbreviations

ARTHS: Arboleda-Tham syndrome; ASD: Atrial septal defect; Bayley-III: Bayley Scales of Infant and Toddler Development, 3rd edition; CT: Computed tomography; IRB: Institutional Review Board; KAT6A: Lysine acetyltransferase 6A; ND: Not described; PCR: Polymerase chain reaction; PDA: Patent ductus arteriosus; VSD: Ventricular septal defect; WES: Whole exome sequencing.

# Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s12920-021-01148-x.

Additional file 1. Candidate variants identified by whole exome sequencing

#### Acknowledgements

Hanyang Medicine-Engineering-Biology Center.

#### Authors' contributions

JK substantially contributed to the conceptualization of the manuscript. JK, SB, HL and HP contributed to the acquisition and interpretation of the clinical data. SB, AY and JK drafted and revised the manuscript. All authors read and approved the final manuscript.

#### Funding

The analysis costs of Sanger sequencing were supported by a grant from the Hanyang Medicine-Engineering-Biology Center. The analysis cost of whole exome sequencing was supported by 3billion, Inc. (Seoul, South Korea). The

funding source has not been involved in study design, collection, analysis and interpretation of data, writing of the report or in the decision to submit the article for publication.

## Availability of data and materials

The details of the variant analyzed during the current study are available in the ClinVar database repository (https://www.ncbi.nlm.nih.gov/clinvar/varia tion/1325410/). The raw datasets of participant generated during the current study are available in the Sequence Read Archive (SRA) database repository (https://www.ncbi.nlm.nih.gov/sra/?term=PRJNA788062/). Public databases used in this study included 1000genomes (http://www.1000genomes.org/), the Genome Aggregation Database (http://gomad.broadinstitute.org), the Exome Aggregation Consortium (http://exac.broadinstitute.org/), the Exome Aggregation Consortium (http://exac.broadinstitute.org/), the Exome Sequencing Project (http://evs.gs.washington.edu/), the Ensemble Project (http://ensemble.org/), the Human Phenotype Ontology (http://hpo.jax.org/), assembly/GCF\_000001405.13/), the DECIPHER database (https://www.decip hergenomics.org/), and OMIM (https://www.omim.org/).

## Declarations

#### Ethics approval and consent to participate

The current study was approved by the Institutional Review Board (IRB) of Hanyang University Medical Center, Seoul, Korea. (IRB No. 2019-11-040). A written informed consent was obtained from both parents of the participant.

#### **Consent for publication**

Written informed consent for the publication of clinical details and any accompanying images was obtained from the both parents of the participant.

# **Competing interests**

The authors declare that they have no potential conflict of interest relevant to this article.

#### Author details

<sup>1</sup>Department of Pediatrics, Hanyang University Medical Center, Hanyang University College of Medicine, 222-1, Wangshimri-ro, Sungdong-gu, Seoul 04763, Republic of Korea. <sup>2</sup>Department of Pediatrics, Kangbuk Samsung Hospital, Sungkyunkwan University School of Medicine, Seoul, Republic of Korea.

Received: 16 May 2021 Accepted: 13 December 2021 Published online: 20 December 2021

### References

- Arboleda VA, Lee H, Dorrani N, Zadeh N, Willis M, Macmurdo CF, et al. De novo nonsense mutations in KAT6A, a lysine acetyl-transferase gene, cause a syndrome including microcephaly and global developmental delay. Am J Hum Genet. 2015;96(3):498–506.
- Tham E, Lindstrand A, Santani A, Malmgren H, Nesbitt A, Dubbs HA, et al. Dominant mutations in KAT6A cause intellectual disability with recognizable syndromic features. Am J Hum Genet. 2015;96(3):507–13.
- 3. Efthymiou S, Salpietro V, Bettencourt C, Houlden H. Paroxysmal movement disorder and epilepsy caused by a de novo truncating mutation in KAT6A. J Pediatr Genet. 2018;7(3):114–6.
- Trinh J, Hüning I, Yüksel Z, Baalmann N, Imhoff S, Klein C, et al. A KAT6A variant in a family with autosomal dominantly inherited microcephaly and developmental delay. J Hum Genet. 2018;63(9):997–1001.
- Kennedy J, Goudie D, Blair E, Chandler K, Joss S, McKay V, et al. KAT6A Syndrome: genotype-phenotype correlation in 76 patients with pathogenic KAT6A variants. Genet Med. 2019;21(4):850–60.
- Alkhateeb A, Alazaizeh W. A novel de novo frameshift mutation in KAT6A identified by whole exome sequencing. J Pediatr Genet. 2019;8(1):10–4.
- Urreizti R, Lopez-Martin E, Martinez-Monseny A, Pujadas M, Castilla-Vallmanya L, Pérez-Jurado LA, et al. Five new cases of syndromic intellectual disability due to KAT6A mutations: widening the molecular and clinical spectrum. Orphanet J Rare Dis. 2020;15(1):44.

- Marji FP, Hall JA, Anstadt E, Madan-Khetarpal S, Goldstein JA, Losee JE. A novel frameshift mutation in KAT6A Is associated with pancraniosynostosis. J Pediatr Genet. 2021;10(1):81–4.
- 9. Lin YF, Lin TC, Kirby R, et al. Diagnosis of Arboleda-Tham syndrome by whole genome sequencing in an Asian boy with severe developmental delay. Mol Genet Metab Rep. 2020;25:100686.
- Jiang M, Yang L, Wu J, Xiong F, Li J. A de novo heterozygous variant in KAT6A is associated with a newly named neurodevelopmental disorder Arboleda-Tham syndrome-a case report. Transl Pediatr. 2021;10(6):1748–54.
- Voss AK, Collin C, Dixon MP, Thomas T. Moz and retinoic acid coordinately regulate H3K9 acetylation, Hox gene expression, and segment identity. Dev Cell. 2009;17(5):674–86.
- Rokudai S, Laptenko O, Arnal SM, Taya Y, Kitabayashi I, Prives C. MOZ increases p53 acetylation and premature senescence through its complex formation with PML. Proc Natl Acad Sci USA. 2013;110(10):3895–900.
- Wright CF, Fitzgerald TW, Jones WD, Clayton S, McRae JF, van Kogelenberg M, et al. Genetic diagnosis of developmental disorders in the DDD study: a scalable analysis of genome-wide research data. Lancet. 2015;385(9975):1305–14.
- Seo GH, Kim T, Park J, Lee J, Kim S, Won D, et al. Pilot study of EVIDENCE: high diagnostic yield and clinical utility of whole exome sequencing using an automated interpretation system for patients with suspected genetic disorders. bioRxiv. 2019:628438.
- Köhler S, Gargano M, Matentzoglu N, et al. The human phenotype ontology in 2021. Nucleic Acids Res. 2021;49(D1):D1207–17.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17(5):405–24.
- 17. Avvakumov N, Cote J. The MYST family of histone acetyltransferases and their intimate links to cancer. Oncogene. 2007;26(37):5395–407.
- Huang F, Abmayr SM, Workman JL. Regulation of KAT6 acetyltransferases and their roles in cell cycle progression, stem cell maintenance, and human disease. Mol Cell Biol. 2016;36(14):1900–7.
- Borrow J, Stanton VP Jr, Andresen JM, Becher R, Behm FG, Chaganti RS, et al. The translocation t(8;16)(p11;p13) of acute myeloid leukaemia fuses a putative acetyltransferase to the CREB-binding protein. Nat Genet. 1996;14(1):33–41.
- Millan F, Cho MT, Retterer K, Monaghan KG, Bai R, Vitazka P, et al. Whole exome sequencing reveals de novo pathogenic variants in KAT6A as a cause of a neurodevelopmental disorder. Am J Med Genet A. 2016;170(7):1791–8.
- Srivastava S, Love-Nichols JA, Dies KA, Ledbetter DH, Martin CL, Chung WK, et al. Meta-analysis and multidisciplinary consensus statement: exome sequencing is a first-tier clinical diagnostic test for individuals with neurodevelopmental disorders. Genet Med. 2019;21(11):2413–21.
- Han JY, Lee IG. Genetic tests by next-generation sequencing in children with developmental delay and/or intellectual disability. Clin Exp Pediatr. 2020;63(6):195–202.
- 23. Jones KL, Adam MP. Evaluation and diagnosis of the dysmorphic infant. Clin Perinatol. 2015;42(2):243-viii.
- Smigiel R, Demkow U. Next generation sequencing in dysmorphology. In: Demkow U, Płoski R, editors. Clinical applications for next-generation sequencing. Cambridge: Academic Press; 2016. p. 137–51.
- Bu H, Liu L, Hu S, Tan Z, Zhao T. Targeted nextgeneration sequencing for research and diagnostics in congenital heart disease, and cleft lip and/or palate. Mol Med Rep. 2019;19(5):3831–40.
- Munabi NCO, Swanson J, Auslander A, Sanchez-Lara PA, Davidson Ward SL, Magee WP 3rd. The prevalence of congenital heart disease in nonsyndromic cleft lip and/or palate: a systematic review of the literature. Ann Plast Surg. 2017;79(2):214–20.
- Nambot S, Thevenon J, Kuentz P, Duffourd Y, Tisserant E, Bruel AL, et al. Clinical whole-exome sequencing for the diagnosis of rare disorders with congenital anomalies and/or intellectual disability: substantial interest of prospective annual reanalysis. Genet Med. 2018;20(6):645–54.
- Dechiphering Developmental Disorders Study. Prevalence and architecture of de novo mutations in developmental disorders. Nature. 2017;542(7642):433–8.

- Elenius V, Lähdesmäki T, Hietala M, Jartti T. Food allergy in a child with de novo KAT6A mutation. Clin Transl Allergy. 2017;7:19.
- Gauthier-Vasserot A, Thauvin-Robinet C, Bruel AL, Duffourd Y, St-Onge J, Jouan T, et al. Application of whole-exome sequencing to unravel the molecular basis of undiagnosed syndromic congenital neutropenia with intellectual disability. Am J Med Genet A. 2017;173(1):62–71.
- Zwaveling-Soonawala N, Maas SM, Alders M, Majoie CB, Fliers E, van Trotsenburg ASP, et al. Variants in KAT6A and pituitary anomalies. Am J Med Genet A. 2017;173(9):2562–5.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

### Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

#### At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

