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REVIEW



Targeting colorectal cancer using dietary flavonols

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Abstract

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Colorectal cancer is among the well-known forms of cancer and a prominent cause of cancer demises worldwide. In vitro experiments reinforced by animal studies, as well as epidemiological studies of human colorectal cancer propose that the growth of this disease can be moderated by eating aspects. Dietary intake including green vegetables and fruits may result in the reduction of colon cancer chances. The finding suggests that the combinations of dietary nutrients may deliver additive or synergistic effects and might be a powerful method to avoid or eradicate colon cancer beginning and/or development. Flavonols are one of the most widespread dietary nutrients of the polyphenolsflavonoids and major constituent of Allium and Brassicaceae vegetables. Flavonols present in vegetables of Allium and Brassicaceae family are kaempferol, myricetin, quercetin, and isorhamnetin. These flavonols are claimed to have antiproliferative activity in vivo and in vitro against colorectal cancer. The objective of this review is to summarize the role of flavonols obtained from dietary sources in the prevention and treatment of colorectal cancer.

K E Y W O R D S isorhamnetin, kaempferol, myricetin, quercetin

1 | INTRODUCTION

Colorectal cancer is one of the most fatal forms of cancer; it is among the commonly diagnosed nature of cancer. A combination of various factors such as the lack of physical exercise, oxidative stress, mutation of oncogenes, environmental factors, dietary factors, and hereditary factors leads to the development of the diseases. A cascade of reaction due to persistent oxidative stress results in DNA damage, which leads to the formation of precancer lesions in mucosal cells and also inhibit the activity of the tumor suppressor gene

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Abbreviations: 5-FU, fluorouracil; AIN, American Institute of Nutrition; AMPK, 5' adenosine monophosphate-activated protein kinase; AOM, azoxymethane; ATM, ataxia telangiectasia mutated; CRC, colorectal cancer; CSK, C-terminal Src kinase; CY, cytochrome; CYP, cytochrome P450; DNA, deoxyribonucleic acid; DSS, dextran sodium sulfate; HCT-15, human colorectal carcinoma; HT-29, human colorectal adenocarcinoma cell line; IC50, half-maximal inhibitory concentration; MAPK, mitogen-activated protein kinase; MCF, Michigan cancer foundation human breast cancer cells; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide); PUMA, p53-upregulated modulator of apoptosis; TBARS, thiobarbituric acid reactive substances; TCF, T cell transcription factor; TRAIL, tumor necrosis factor-related apoptosis-inducing ligand.

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further resulting in one of the most common and fatal colorectal cancer. Here, antioxidants may play a critical role. Fortunately, a number of food articles are rich sources of such phytochemicals. Recently various in vitro studies and animal experimentation have suggested that dietary factors may play a significant part in the initiation or control of human colorectal cancer. Studies have shown that individuals who ingest regimes rich in phytochemicals particularly polyphenolic compounds with strong antioxidant properties have poorer degrees of colorectal cancer (Figure 1). A repeated acquaintance of colonic cells to these polyphenolic compounds present in the diet not only helps in inhibiting the progress of cancer cells but may be efficacious in preventing colon cancer [1-4].

Flavonoids are widely distributed polyphenolic compounds in nature and a prevalent component of the human diet. Many food flavonoids influence different cellular processes to reduce the changes in colon epithelia, inhibit tumor formation, and act as chemopreventive blocker agents or, chemopreventive suppressor agents using different biological mechanisms of action. However, their exact mechanism of action in vivo is still not established; it may be due to inhibition of angiogenesis, metastasis, initiation of apoptosis, cell cycle arrest, antisurvival, anti-inflammatory, and antiproliferative effects. Flavanoids are supposed to have CYP1 inhibitory action by a significant interaction with cytochrome P450 CYP1 enzymes, thereby resulting in carcinogenesis blockage at the initiation stage. Researchers have tried to derive structure-activity relationships for these potential therapeutic agents which might lead to new drug discovery. Further, as a lot of flavonoids are present in our diet, a greater understanding of their anticancer properties might also help us modify our dietary habits and develop an effective supplement

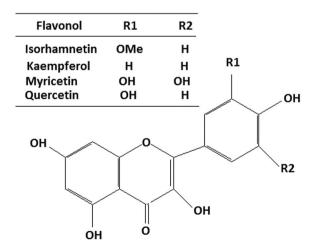


FIGURE 1 Chemical structure of different flavonols.

therapy that may provide additive or synergistic effects and could be a potent way to prevent or eliminate colon cancer initiation and/or progression [5–13].

Chemical classification of flavonoids includes isoflavones, flavanones, flavanols, flavonols, flavones, and anthocyanidins. They are abundantly found in Allium and Brassicaceae vegetables. Chive (Allium schoenoprasum L.), leek (Allium porrum L.), shallot (Allium ascalonicum Hort.), scallion (Allium fistulosum L.), and above all garlic and onion are popular and routine parts of dietary intake. Brassicaceae vegetables are also considered an integral part of the human diet chart. Cauliflower, broccoli, brussels sprouts, cabbage, and kale, represent popular B. oleracea, a class of vegetables (Table 1). Various clinical studies have found the reduced risk of colon cancer associated with higher intake of vegetables of these classes. Flavonoids, subclass flavonols, and organosulfur compounds are among the chief chemical constituents present in these vegetables. Kaempferol, myricetin, quercetin, and isorhamnetin are the flavonols present in the above vegetables which have shown some promising in vivo and in vitro antiproliferative action against colorectal cancer (Table 2). This review emphasizes the pronounced prospect of flavonols in cancer prevention and therapy. It is apprehended to give an inclusive summary of the existing knowledge and integral modes of action, concentrating on the flavonols present in the Allium and Brassica family [14–17].

2 QUERCETIN

Quercetin (3,3',4',5,7-pentahydroxyflavone) is the aglycone form of a flavonol, widely distributed in dietary sources. It combines with many flavonoid glycosides such as rutin and quercetin. It is mainly distributed in the perinuclear and nucleoli areas of the cell. It is an antioxidant, like many other phenolic heterocyclic compounds. Quercetin has potential chemopreventive activity. Another observation suggested that the effects of dietary quercetin on colon cancer risk may vary with varying intake of fruit or tea. It was also concluded that quercetin has a defensive effect only on proximal colon cancer [18, 19]. There are several reports on the activity of quercetin in colorectal cancer both in vivo and in vitro [20-27]. Quercetin prevents p21-RAS expression in human colon cancer cell lines and in primary colorectal tumors. These effects of quercetin recommend a conceivable chemopreventive role for this molecule in colorectal carcinogenesis. It is reported to initiate autophagy in Ha-RAS-transformed human colon cells by modulating deprivation of oncogenic Ras. Quercetin facilitates frequent mutations of RAS genes by downregulating

TABLE 1 Distribution of polyphenols in dietary sources.

No	Class	Subclass	Polyphenols	Food source
1	Phenolic acids	Hydroxy-benzoic acid	Gallic acid	Tea, blackcurrent
			Protocatechuic acid	Rasberry, mushroom
			Syringic acid	Acai palm
		Hydroxy-cinnamic acid	Ferulic acid	Cereal grains
			Caffeic acid	Kiwi
			Coumaric acid	Plum
			Sinapic acid	Potato, coffee, chicory, pear, flour
2	Flavonoids	Flavonols	Galangin	Lesser galangal (Alpinia officinarum)
			Kaempferride	Onion, olives, lettuce, parsley
			Kaempferol	Broccoli, leek
			Myricetin	Broccoli, tomato, cherry, blueberry, fruit peels
			Quercetin	Onions, tea, curly kale, bark, and rind of numerous plants
		Flavones	Apigenin	Celery, apple skin
			Bacalein	Roots of Scutellaria baicalensis
			Chrysin	Berries
			Diosmetin	Oregano, spearmint
		Isoflavones	Daidzein	Soy food, soybeans, legumes
			Genistein	Miso
		Flavanones	Eriodictyol	Citrus fruits
			Hesperidin	Lemon juice, citrus fruits
			Naringenin	Citrus fruits
			Silymarin	Coriander seeds, turmeric root
		Flavanols (catechins)	Catechin	Red wine, red grape, tea
			Gallocatechin	Pomegranate, bananas
			Epicatechin	Grapes, peach, apricot, apple, red wine
			Epigallocatechin 3-gallate	Tea
		Anthocyanidins	Cyanidin	Blackberry, berries
			Delphinidin	Black grapes, cherries
			Pelargonidin	Raspberry
			Peonidin	Blueberry, red grapes
			Malvidin	Strawberry, red wine, plum, cherry
		Chalcones	Hop chalcones	Beer, hops
3	Stilbenes		Resveratrol	Grapes
4	Lignans		Secoisolariciresinol	Linseed

the concentration of oncogenic *RAS* genes in affected cells and acts as a chemopreventive agent [20]. Quercetin prevents human DLD-1 colon cancer cell progression and polyamine biosynthesis by affecting the polyamines and ornithine decarboxylase. Both of them are convoluted in cell progress and differentiation. It initiates apoptosis and

decreases the polyamine biosynthesis [21]. Quercetin also inhibits human SW480 colon cancer growth in association with the inhibition of cyclin D1 and survivin expression at the molecular level through the Wnt/betacatenin signaling pathway. The expression of cyclin D(1) and survivin is inhibited via the Wnt/beta-catenin

	Content (mg/100 g)			
Names of dietary sources	Quercitin	Kaempferol	Myricetin	Isorhamnetin
Onion	20.30	0.65	0.03	5.01
Garlic	1.74	0.26	1.61	-
Leek	0.09	2.67	0.22	_
Broccoli	3.26	7.84	0.06	-
Cabbage	0.28	0.18	_	_
Cauliflower	0.54	0.36	0.00	-

signaling pathway attributing to its antitumor effect [22]. Dietary quercetin modulated exposed confluent Caco-2 monolayers to exhibit a decrease in cell differentiation. The biphasic effect on cell proliferation was also observed in human colon cancer cell line Caco-2 [23]. It was reported that quercetin boosted TRAIL-tempted apoptosis by initiating the redistribution of DR4 and DR5 into lipid rafts. It was projected that quercetin, through its capacity to redistribute death receptors at the cell surface, facilitates death-inducing signaling complex formation, and activation of caspases in response to death receptor stimulation [24].

The treatment of HT-29 colon cancer demonstrated the mechanism of its action. Quercetin was found to activate AMPK to modulate apoptosis via p53-dependent apoptotic cell death leading to a significant reduction in tumor in HT-29 colon cancer cells [25]. Quercetin decreases the expression of ErbB2 and ErbB3 proteins in HT-29 human colon cancer cells. In vitro studies indicated the cytotoxic nature of quercetin, as it initiated the differentiation in undifferentiated cancer cell lines. It also inhibited the active proliferation of cells [26]. Another study indicated its inhibitory activity over cell viability by inducing mutation in RAS genes. Levels of Ras proteins and half-life of oncogenic Ras were found to be decreased. Autophagic responses were also observed [27]. Another finding suggested the role of quercetin and luteolin in a decrease in drug resistance leading to increased 5-FU effects in CO115 p53 WT tumors using microsatellite instability therapy. The combination therapy increased apoptosis by enhancing p53 expression [28]. The effect of quercetin was also evaluated using Caco-2 cells. It involved the study of its expression over 4000 human genes. It was found to positively influence the activity of tumor suppressor genes using signal transduction pathways such as MAPK and TCF [29]. Sulphated derivative of quercetin, quercetin-5', 8-disulfonate was found to influence the ROSdependent apoptosis pathway to inhibit cell proliferation in the S phase. The study involved LoVo cells and MCF-7

cells. The effects were dose-dependent [30]. Another study reported that curcumin and quercetin caused a dose-dependent enhancement of tumors induced by AOM in the colon model, upon azoxymethane-induced colon cancer. A significant decrease in cell viability was evident in a dose-dependent manner. In vivo experiments revealed promising preventive or therapeutic efficacy [31]. Quercetin, was also found to reduce colorectal carcinogenesis of azoxymethane-treated rats in high dosage; however, rutin was not found to be effective possibly due to its low bioavailability [32]. Similarly, azoxymethane-treated mice were kept on a standard AIN-76A diet and investigated for focal areas of dysplasia and cyclin D(1) expression. Quercetin was found to decrease not only the total number of focal areas of dysplasia but also the number of mice exhibiting the same [33, 34]. The synergistic effect of quercetin and kaempferol was evident in studies conducted using HuTu-80, Caco-2, and PMC42. A significant reduction in total cell proliferation was seen possibly owing to reduced nuclear proliferation antigen Ki67 expression [35-37]. In the year 2019, Yang et al. [38] induced apoptosis in colorectal cancer cells via JNK signaling pathways. Recent studies themselves depicted that the use of quercetin restricts colon cancer via modulating antiaging protein expression [39].

3 | KAEMPFEROL

Kaempferol is a polyphenol antioxidant (3,5,7-trihydroxy-2-(4-hydroxyphenyl)-4H-1-benzopyran-4-one) under the subclass flavonols, dispersed extensively in *Allium* and *Brassica* vegetables and fruits. It is widely distributed in many edible plants. Its daily dietary intake was estimated to be up to approximately 10 mg/day. Kaempferol has established a prodigious deal of consideration in research, due to its certain part in tumbling many diseases such as osteoporosis, diabetes, neurodegenerative diseases, anxiety, infectious diseases, allergies, inflammation, and pain. Various in vitro and in vivo studies have also testified it to display anticancer, anti-inflammatory, anti-allergic, antiasthmatic, antimicrobial, and antioxidant activities. It is also used in traditional folk medicines. Further, several epidemiological lessons evaluated the positive interaction between the consumption of kaempferol-rich foods and its role in sinking many forms of distortions such as lung, ovarian, gastric, and pancreatic cancers in the human population. Numerous clinical investigations have demonstrated its therapeutic and anticancer effects. Kaempferol can be used as an adjuvant with chemotherapeutic medications in cancer treatment plans to make cells more susceptible to cytotoxicity. Kaempferol scavenges free radicals and inhibits the neoplastic process [40]. A study over male Wistar rats was performed to reveal the effect of kaempferol on tissue lipid peroxidation, its efficacy with irinotecan, and its antioxidant properties against 1,2dimethyl hydrazine-induced colorectal cancer. Kaempferol was found to lower liver thiobarbituric acid reactive substances level and erythrocyte lysate. It rejuvenated various antioxidant enzymes such as catalase, superoxide dismutase, and glutathione peroxidase. The outcomes supported the role of kaempferol as a chemopreventive agent in colorectal cancer [41]. An experiment over human HCT116 colon cancer cells revealed its mechanism of action. It activates caspase-3 cleavage and induces cytochrome *c* release from mitochondria using Bcl-2 family proteins (i.e., PUMA). It was found to affect the Ataxia-Telangiectasia Mutated-p53 pathway. The p53 upregulated modulator was also involved in apoptosis. It was also found to induce ATM and H2AX phosphorylation in cancer cells. Inhibition of ATM helps to control cell proliferation leading to cell cycle arrest [42]. Another study examined the effect of kaempferol on HT-29 human colon cancer cells. Cells were treated with $0-60 \,\mu mol/L$ concentration of kaempferol. MTT assay was used to evaluate its effect on cell proliferation and [(3)H]thymidine incorporation assay determined its effect on DNA synthesis. Cell cycle phase distribution was also studied using fluorescence-activated cell sorting analyses. The activity of cyclin-dependent kinase (CDK)s was measured using in vitro kinase assays and western blot analyses revealed the expression of proteins in various stages of the cell cycle. It was found that the kaempferol reduced the number of viable cells and brought the dose-dependent incorporation of [(3)H]thymidine in HT29 cell DNA. Cell cycle was arrested at the G1 stage within 6 h, and G2/M arrest required 12 h. The protein expression of CDK2, CDK4, cyclins D1, cyclin E, and cyclin A and the activity of CDK2 and CDK4 was inhibited. The phosphorylation retinoblastoma protein was also suppressed. Further, it reduced the concentration of Cdc25C, Cdc2, and cyclin B1 proteins. Cdc2 activity was similarly decreased. The study found that kaempferol suppresses CDK2, CDK4, and Cdc2 activity, causing cell cycle arrest at the G1 and G2/M stages. Kaempferol's ability to suppress colon cancer growth may be linked to its ability to induce cell cycle arrest in colon cancer cells [43, 44].

Another study conducted to study the activity of curcumin and kaempferol on the DLD-1 colon cancer cell line of epithelial origin indicated the inhibitory role of both phytochemicals during cell proliferation [45]. Significant antiproliferative activity of kaempferol was observed at high levels, while curcumin was effective in relatively low concentrations. Synergistic antiproliferative effects were evident. Kaempferol in low concentration was very effective when combined with curcumin at 12.5 µM (IC50). This combination of kaempferol and curcumin showed significant antiproliferative activity against colon cancer cells (HCT-15) and human normal lymphocytes [46, 47]. Recent studies in the literature clarified that kaempferol is a promising phytopharmaceutical used in the management of colon cancer via mediating inflammation and signal transduction pathways [48].

4 | ISORHAMNETIN

Isorhamnetin is widely distributed in fruits (i.e., Hippophae rhamnoides L). Isorhamnetin, a 3'-O-methylated metabolite of quercetin, has lethal effects on human colon cancer cells via affecting cell cycle, cell death, and proliferation in human colon carcinoma (HCT-116) cells. The number of cells was discovered to rise during the G2/ M phase. With an IC50 of 72 M, the MTT assay demonstrated its effectiveness in a dose- and timedependent manner. It initiates apoptosis and necrosis as suggested by flow cytometry and fluorescence microscopy. Low serum levels initiate cell cycle progression to the G0/ G1 phase and positively influence cell death leading to cell necrosis or apoptotic processes. Here, the G2/M stage may be a target to inhibit HCT-116 cell growth. The above activities suggest the potential chemopreventive property of isorhamnetin [49]. Another report suggests oncogenic Src and β -catenin as potential mediators for their chemopreventive action in colorectal cancer. Advanced adenoma relapse could be prevented using an isorhamnetin-rich diet in a poly-prevention trial. Dietary isorhamnetin was found to be effective in controlling the tumor burden, and tumor number by 59% and 35%. The mortality rates were also decreased to 62% in a study involving colorectal tumorigenesis of FVB/N mice. The study used azoxymethane-treated cells, which were further treated with dextran sodium sulfate (DSS). DSS is a colonic irritant and azoxymethane is a known

chemical carcinogen. It was observed using immune histochemical analysis and histopathology studies that dietary isorhamnetin controlled DSS-induced inflammation faster than the control diet. This response was mediated by stimulation of C-terminal Src kinase (CSK) and inhibition of β -catenin nuclear translocation. c-Src activation, which was induced by DSS was also suppressed. CSK is a negative regulator of of Src family of tyrosine kinases [50]. A similar mechanism was observed in another study using HT-29 colon cancer cells, where RNA interference helped to stimulate CSK expression. As evident in studies conducted over human CRC such as HT-29, HCT116, and SW480; it may be concluded that dietary isorhamnetin mediates its activity by inhibiting PI3K-Akt-mTOR pathway and anti-inflammatory mechanisms and positively modulating the expression of CSK and negatively influencing the oncogenic Src activity and expression of cyclin B1 protein. It also modulated the phosphorylation levels of phosph-p70S6 kinase, phosph-4E-BP1 (t37/46), and protein Akt (ser473) [51]. Isorhamnetin is identified to encourage vital anticancer activity via numerous signaling pathways (PI3K/AKT, and NF-kB) and has shown promising activity against cancer [52, 53]. Recent data itself supports the fact that isorhamnetin has anti-colon rectal cancer activity. This has proven that this may be due to upregulating factors responsible for cell cycle arrest or cell death.

5 | MYRECITIN

Myricetin (3,3',4',5,5,7'-hexahydroxyflavone) is widely distributed in berries, tea, and red wine. It exhibits structural similarity with quercetin. Studies conducted over HT-29 cells, Caco-2, SW480, COLO 205, VACO-235, and T84, show its inhibitory effect on cell growth, induction of apoptosis, inhibition of cell multiplication, and antimetastatic properties. It is believed to act via inhibitory effects over EGFR kinases, matrix metalloproteinase isoform 2 activity modulation of apopain activity, and auto-oxidation process [54, 55]. Its effects on rat colon carcinogenesis were investigated using 1,2 dimethyl hydrazine as an inducer. Overall it reduced the number of tumors and the progress of the disease. Liver TBARS were significantly reduced. The bacterial enzyme activity was significantly reduced at a dose of 50 and 100 mg/kg. Glutathione peroxidase, GSH, and catalase activity were all considerably increased in a dose-dependent manner [56]. In HCT-15 cells, myricetin was discovered to cause apoptosis. To increase the BCL2-associated X protein/Bcell lymphoma 2 ratio, it likely functions by releasing the positive apoptosis influencer from the mitochondrial membrane. However, the cleavage of caspase-3 and

caspase-9 was unaffected [57]. Additional in vivo research may shed light on specific pathways [58–61]. Myricetin's anticancer activity and ability to operate on S4-10 have been demonstrated in vitro and in vivo during experiments involving the proliferation of A549 cells [62, 63].

6 | CONCLUSION AND RELEVANCE TO FUTURE RESEARCH

Flavonols are new hope, a gift by nature, not only because this class assists in normal cell growth by apoptosis, metastasis, and antiproliferative activity using various molecular cascade of events, but it seems to elevate general well-being silently because they are consumed widely without prescription as a dietary ingredient. However, a cautious approach is recommended as these studies have not been proven through clinical trials. These studies have used a concentration range which is difficult to achieve through dietary sources, as the free form of many flavonols shows low aqueous solubility and high molecular weight leading to low bioavailability, extensive metabolism, and rapid elimination. Poor vascular/oral bioavailability and bitter taste further create a challenge for formulators. The clinical relevance of conjugated forms of their metabolites is required to be investigated as they may not be as effective as compared to their precursors. The stability of these compounds is another concern. Flavonols are sensitive to various environmental conditions such as light and heat and rapidly undergo oxidation and hydrolysis. The exposure time is still to be manipulated and understood before utilizing their potential as colorectal cancer preventive agents. The effect of dose with reference to the treatment period is required to be established. The question remains whether their dietary concentration will help or only therapeutic formulation will be effective. The synergistic effects of these compounds are required to be further explored. Whether its bioavailability will improve after targeted intestinal absorption and metabolism, is still to be answered. Their safety and toxicity-related concerns are required to be addressed as this class utilizes various critical biochemical and molecular cascades of events to express their therapeutic efficacy. The modulation of these pathways may interfere with the metabolism of other compounds, hampering other required events, and antagonizing systemic drug effects. Pro-drug design (i.e., methoxylated flavonoids) may also be helpful to achieve some goals. Methoxylated flavonoids are promising chemopreventive compounds, which act as prodrugs in events associated with CYP1A1 and CYP1 metabolism.

Sufficient in vitro studies have reported their chemopreventive properties. If, flavonols and their prodrugs could be further explored in vivo, a new arena for research may emerge.

AUTHOR CONTRIBUTIONS

Nitin Dubey: Investigation (equal), methodology (equal), writing—original draft (equal), writing—review and editing (equal). Nidhi Dubey: Conceptualization (equal), supervision (equal), writing—original draft (equal), writing—review and editing (equal). Upendra Bhadoria: Methodology (equal), writing—original draft (equal), writing—review and editing (equal). Kamal Shah: Investigation (equal), methodology (equal), writing—original draft (equal), writing—review and editing (equal). Nagendra Singh Chauhan: Investigation (equal), supervision (equal), writing—review and editing (equal).

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CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no data sets were generated or analyzed during the current study.

ETHICS STATEMENT

Not applicable.

INFORMED CONSENT

Not applicable.

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REFERENCES

- Lipkin M, Reddy B, Newmark H, Lamprecht SA. Dietary factors in human colorectal cancer. Annu Rev Nutr. 1999;19: 545–86. https://doi.org/10.1146/annurev.nutr.19.1.545
- Araújo JR, Gonçalves P, Martel F. Chemopreventive effect of dietary polyphenols in colorectal cancer cell lines. Nutr Res. 2011;31(2):77–87. https://doi.org/10.1016/j.nutres.2011.01.006
- Pampaloni B, Palmini G, Mavilia C, Zonefrati R, Tanini A, Brandi ML. In vitro effects of polyphenols on colorectal cancer cells. World J Gastrointest Oncol. 2014;6(8):289–300. https:// doi.org/10.4251/wjgo.v6.i8.289
- 4. Kuntz S, Wenzel U, Daniel H. Comparative analysis of the effects of flavonoids on proliferation, cytotoxicity and

apoptosis in human colon cancer cell lines. Eur J Nutr. 1999;38(3):133–42. https://doi.org/10.1007/s003940050054

- Sak K. Cytotoxicity of dietary flavonoids on different human cancer types. Phcog Rev. 2014;8(16):122–46. https://doi.org/ 10.4103/0973-7847.134247
- Ramos S. Effects of dietary flavonoids on apoptotic pathways related to cancer chemoprevention. J Nutr Biochem. 2007;18(7): 427–42. https://doi.org/10.1016/j.jnutbio.2006.11.004
- Venturelli S, Burkard M, Biendl M, Lauer UM, Frank J, Busch C. Prenylated chalcones and flavonoids for the prevention and treatment of cancer. Nutrition. 2016;32(11–12):1171–8. https://doi.org/10.1016/j.nut.2016.03.020
- Lopez-Lazaro M. Flavonoids as anticancer agents: structureactivity relationship study. Curr Med Chem-Anti-Cancer Agents. 2002;2(6):691–714. https://doi.org/10.2174/1568011023353714
- Park EJ, Pezzuto JM. Flavonoids in cancer prevention. Anti Cancer Agents Med Chem. 2012;12(8):836–51. https://doi.org/ 10.2174/187152012802650075
- Kuo SM. Antiproliferative potency of structurally distinct dietary flavonoids on human colon cancer cells. Cancer Lett. 1996; 110(1–2):41–8. https://doi.org/10.1016/s0304-3835(96)04458-8
- Androutsopoulos VP, Papakyriakou A, Vourloumis D, Tsatsakis AM, Spandidos DA. Dietary flavonoids in cancer therapy and prevention: substrates and inhibitors of cytochrome P450 CYP1 enzymes. Pharmacol Ther. 2010;126(1): 9–20. https://doi.org/10.1016/j.pharmthera.2010.01.009
- Galati G, O'brien PJ. Potential toxicity of flavonoids and other dietary phenolics: significance for their chemopreventive and anticancer properties. Free Radic Biol Med. 2004;37(3):287– 303. https://doi.org/10.1016/j.freeradbiomed.2004.04.034
- Batra P, Sharma AK. Anti-cancer potential of flavonoids: recent trends and future perspectives. 3 Biotech. 2013;3(6): 439–59. https://doi.org/10.1007/s13205-013-0117-5
- Cartea ME, Francisco M, Soengas P, Velasco P. Phenolic compounds in brassica vegetables. Molecules. 2010;16(1):251– 80. https://doi.org/10.3390/molecules16010251
- Bianchini F, Vainio H. Allium vegetables and organosulfur compounds: do they help prevent cancer? Environ Health Perspect. 2001;109(9):893–902. https://doi.org/10.1289/ehp. 01109893
- Bhagwat S, Haytowitz DB, Holden JM. USDA database for the flavonoid content of selected foods release 3.1. Nutrient Data Laboratory, Beltsville Human Nutrition Research Center Agricultural Research Service, US Department of Agriculture; 2014.
- Lanzotti V. The analysis of onion and garlic. J Chromatogr A. 2006;1112(1-2):3-22. https://doi.org/10.1016/j.chroma.2005. 12.016
- Murakami A, Ashida H, Terao J. Multitargeted cancer prevention by quercetin. Cancer Lett. 2008;269(2):315–25. https://doi.org/10.1016/j.canlet.2008.03.046
- Djuric Z, Severson RK, Kato I. Association of dietary quercetin with reduced risk of proximal colon cancer. Nutr Cancer. 2012;64(3):351–60. https://doi.org/10.1080/01635581.2012.658950
- Ranelletti FO, Maggiano N, Serra FG, Ricci R, Larocca LM, Lanza P, et al. Quercetin inhibits p21-RAS expression in human colon cancer cell lines and in primary colorectal tumors. Int J Cancer. 2000;85(3):438–45. https://doi.org/10.1002/(SICI)1097-0215(20000201)85:3<438::AID-IJC22>3.0.CO;2-F

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- Linsalata M, Orlando A, Messa C, Refolo MG, Russo F. Quercetin inhibits human DLD-1 colon cancer cell growth and polyamine biosynthesis. Anticancer Res. 2010;30(9):3501–7. https://pubmed.ncbi.nlm.nih.gov/20944129/
- Mouat MF, Kolli K, Orlando R, Hargrove JL, Grider A. The effects of quercetin on SW480 human colon carcinoma cells: a proteomic study. Nutr J. 2005;4:11. https://doi.org/10.1186/ 1475-2891-4-11
- Dihal AA, Woutersen RA, van Ommen B, Rietjens IMCM, Stierum RH. Modulatory effects of quercetin on proliferation and differentiation of the human colorectal cell line Caco-2. Cancer Lett. 2006;238(2):248–59. https://doi.org/10.1016/j. canlet.2005.07.007
- Psahoulia FH, Drosopoulos KG, Doubravska L, Andera L, Pintzas A. Quercetin enhances TRAIL-mediated apoptosis in colon cancer cells by inducing the accumulation of death receptors in lipid rafts. Mol Cancer Ther. 2007;6(9):2591–9. https://doi.org/10.1158/1535-7163.MCT-07-0001
- Kim HJ, Kim SK, Kim BS, Lee SH, Park YS, Park BK, et al. Apoptotic effect of quercetin on HT-29 colon cancer cells via the AMPK signaling pathway. J Agricult Food Chem. 2010; 58(15):8643–50. https://doi.org/10.1021/jf101510z
- Kim WK, Bang MH, Kim ES, Kang NE, Jung KC, Cho HJ, et al. Quercetin decreases the expression of erbb2 and erbb3 proteins in HT-29 human colon cancer cells. J Nutr Biochem. 2005;16(3):155– 62. https://doi.org/10.1016/j.jnutbio.2004.10.010
- Psahoulia FH, Moumtzi S, Roberts ML, Sasazuki T, Shirasawa S, Pintzas A. Quercetin mediates preferential degradation of oncogenic Ras and causes autophagy in Ha-RAS-transformed human colon cells. Carcinogenesis. 2007;28(5):1021–31. https:// doi.org/10.1093/carcin/bgl232
- Xavier CPR, Lima CF, Rohde M, Pereira-Wilson C. Quercetin enhances 5-fluorouracil-induced apoptosis in MSI colorectal cancer cells through p53 modulation. Cancer Chemother Pharmacol. 2011;68(6):1449–57. https://doi.org/10.1007/ s00280-011-1641-9
- van Erk MJ, Roepman P, van der Lende TR, Stierum RH, Aarts JMMJG, van Bladeren PJ, et al. Integrated assessment by multiple gene expression analysis of quercetin bioactivity on anticancer-related mechanisms in colon cancer cells in vitro. Eur J Nutr. 2005;44(3):143–56. https://doi.org/10.1007/s00394-004-0503-1
- Zhang HS, Zhang M, Yu LH, Zhao Y, He NW, Yang XB. Antitumor activities of quercetin and quercetin-5',8disulfonate in human colon and breast cancer cell lines. Food Chem Toxicol. 2012;50(5):1589–99. https://doi.org/10. 1016/j.fct.2012.01.025
- Pereira MA, Grubbs CJ, Barnes LH, Li H, Olson GR, Eto I, et al. Effects of the phytochemicals, curcumin and quercetin, upon azoxymethane-induced colon cancer and 7,12-dimethylbenz[a]anthracene-induced mammary cancer in rats. Carcinogenesis. 1996;17(6):1305–11. https://doi.org/ 10.1093/carcin/17.6.1305
- 32. Dihal AA, de Boer VCJ, van der Woude H, Tilburgs C, Bruijntjes JP, Alink GM, et al. Quercetin, but not its glycosidated conjugate rutin, inhibits azoxymethane-induced colorectal carcinogenesis in F344 rats. J Nutr. 2006;136(11): 2862–7. https://doi.org/10.1093/jn/136.11.2862

- Yang K, Lamprecht SA, Liu Y, Shinozaki H, Fan K, Leung D, et al. Chemoprevention studies of the flavonoids quercetin and rutin in normal and azoxymethane-treated mouse colon. Carcinogenesis. 2000;21(9):1655–60. https://doi.org/10.1093/ carcin/21.9.1655
- Ackland ML, van de Waarsenburg S, Jones R. Synergistic antiproliferative action of the flavonols quercetin and kaempferol in cultured human cancer cell lines. In Vivo. 2005;19(1): 69–76.
- Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics. CA Cancer J Clin. 2022;72(1):7–33. https://doi.org/10.3322/ caac.21708
- Jiang Y, Lin WF, Zhu LY. Targeted drug delivery for the treatment of blood cancers. Molecules. 2022;27(4):1310. https://doi.org/10.3390/molecules27041310
- Wang Q, Chen YK, Lu HJ, Wang HJ, Feng H, Xu JF, et al. Quercetin radiosensitizes non-small cell lung cancer cells through the regulation of miR-16-5p/WEE1 axis. IUBMB Life. 2020;72(5):1012–22. https://doi.org/10.1002/iub.2242
- Yang YW, Wang T, Chen DG, Ma QZ, Zheng YJ, Liao SP, et al. Quercetin preferentially induces apoptosis in KRAS-mutant colorectal cancer cells via JNK signaling pathways. Cell Biol Int. 2019;43(2):117–24. https://doi.org/10.1002/cbin.11055
- Bhatiya M, Pathak S, Jothimani G, Duttaroy AK, Banerjee A. A comprehensive study on the anti-cancer effects of quercetin and its epigenetic modifications in arresting progression of colon cancer cell proliferation. Arch Immunol Ther Exp. 2023; 71(1):6. https://doi.org/10.1007/s00005-023-00669-w
- Calderón-Montaño JM, Burgos-Morón E, Pérez-Guerrero C, López-Lázaro M. A review on the dietary flavonoid kaempferol. Mini-Rev Med Chem. 2011;11(4):298–344. https://doi.org/ 10.2174/138955711795305335
- Nirmala P, Ramanathan M. Effect of kaempferol on lipid peroxidation and antioxidant status in 1,2-dimethyl hydrazine induced colorectal carcinoma in rats. Eur J Pharmacol. 2011; 654(1):75–9. https://doi.org/10.1016/j.ejphar.2010.11.034
- 42. Li W, Du BN, Wang TY, Wang SL, Zhang JH. Kaempferol induces apoptosis in human HCT116 colon cancer cells via the Ataxia-Telangiectasia Mutated-p53 pathway with the involvement of p53 upregulated modulator of apoptosis. Chemico-Biolog Interactions. 2009;177(2):121–7. https:// doi.org/10.1016/j.cbi.2008.10.048
- Lee HS, Cho HJ, Yu RN, Lee KW, Chun HS, Park HJY. Mechanisms underlying apoptosis-inducing effects of kaempferol in HT-29 human colon cancer cells. Int J Mol Sci. 2014;15(2):2722–37. https://doi.org/10.3390/ijms15022722
- Kalyani C, Narasu ML, Devi YP. Effect of phytochemical kaempferol on HCT-15 and lymphocytes. Biology. 2015;5(10): 452–4.
- Kaverimanian A, Khatwani N, Ezekiel U. Antiproliferative effect of curcumin and kaempferol on colon cancer cells. FASEB J. 2016;30(S1):841.5. https://doi.org/10.1096/fasebj.30. 1_supplement.841.5
- 46. Afzal M, Alarifi A, Karami AM, Ayub R, Abduh NAY, Saeed WS, et al. Antiproliferative mechanisms of a polyphenolic combination of kaempferol and fisetin in triplenegative breast cancer cells. Int J Mol Sci. 2023;24(7):6393. https://doi.org/10.3390/ijms24076393

- Radziejewska I, Supruniuk K, Tomczyk M, Izdebska W, Borzym-Kluczyk M, Bielawska A, et al. p-coumaric acid, kaempferol, astragalin and tiliroside influence the expression of glycoforms in AGS gastric cancer cells. Int J Mol Sci. 2022;23(15):8602. https://doi.org/10.3390/ijms23158602
- Almatroudi A, Allemailem KS, Alwanian WM, Alharbi BF, Alrumaihi F, Khan AA, et al. Effects and mechanisms of kaempferol in the management of cancers through modulation of inflammation and signal transduction pathways. Int J Mol Sci. 2023;24(10):8630. https://doi.org/10.3390/ijms24108630
- Jaramillo S, Lopez S, Varela LM, Rodriguez-Arcos R, Jimenez A, Abia R, et al. The flavonol isorhamnetin exhibits cytotoxic effects on human colon cancer cells. J Agricult Food Chem. 2010;58(20):10869–75. https://doi.org/10.1021/jf102669p
- Saud SM, Young MR, Jones-Hall YL, Ileva L, Evbuomwan MO, Wise J, et al. Chemopreventive activity of plant flavonoid isorhamnetin in colorectal cancer is mediated by oncogenic Src and β-catenin. Cancer Res. 2013;73(17):5473–84. https://doi. org/10.1158/0008-5472.CAN-13-0525
- Li C, Yang X, Chen C, Cai SX, Hu JB. Isorhamnetin suppresses colon cancer cell growth through the PI3K-Akt-mtor pathway. Mol Med Rep. 2014;9(3):935–40. https://doi.org/10.3892/mmr. 2014.1886
- Ishola IO, Osele MO, Chijioke MC, Adeyemi OO. Isorhamnetin enhanced cortico-hippocampal learning and memory capability in mice with scopolamine-induced amnesia: role of antioxidant defense, cholinergic and BDNF signaling. Brain Res. 2019;1712: 188–96. https://doi.org/10.1016/j.brainres.2019.02.017
- Jiayi C, Tianyi N, Dan T, Tingguo K, Qingfeng W, Qianqian Z. Isorhamnetin protects endothelial cells model CRL1730 from oxidative injury by hydrogen peroxide. Pak J Pharm Sci. 2019;32(1):131–6. https://pubmed.ncbi.nlm.nih.gov/30772801/
- De S, Paul S, Manna A, Majumder C, Pal K, Casarcia N, et al. Phenolic phytochemicals for prevention and treatment of colorectal cancer: a critical evaluation of in vivo studies. Cancers. 2023;15(3):993. https://doi.org/10.3390/cancers15030993
- Ko CH, Shen SC, Lee TJF, Chen YC. Myricetin inhibits matrix metalloproteinase 2 protein expression and enzyme activity in colorectal carcinoma cells. Mol Cancer Ther. 2005;4(2):281– 90. https://doi.org/10.1158/1535-7163.281.4.2

- Nirmala P, Ramanathan M. Effect of myricetin on 1,2 dimethylhydrazine induced rat colon carcinogenesis. J Exp Ther Oncol. 2011;9(2):101–8. https://pubmed.ncbi.nlm.nih. gov/21699017/
- 57. Kim ME, Ha TK, Yoon JH, Lee JS. Myricetin induces cell death of human colon cancer cells via BAX/BCL2-dependent pathway. Anticancer Res. 2014;34(2):701–6. https://pubmed. ncbi.nlm.nih.gov/24511002/
- Gulbake A, Jain A, Jain A, Jain A, Jain SK. Insight to drug delivery aspects for colorectal cancer. World J Gastroenterol. 2016;22(2):582–99. https://doi.org/10.3748/wjg.v22.i2.582
- Xu GY, Shi HS, Ren LB, Gou HF, Gong DY, Gao X, et al. Enhancing the anti-colon cancer activity of quercetin by selfassembled micelles. Int J Nanomedicine. 2015;10:2051–63. https://doi.org/10.2147/IJN.S75550
- Gao S, Hu M. Bioavailability challenges associated with development of anti-cancer phenolics. Mini-Rev Med Chem. 2010;10(6):550–67. https://doi.org/10.2174/138955710791384081
- Parisi OI, Puoci F, Restuccia D, Farina G, Iemma F, Picci N. Polyphenols and their formulations: different strategies to overcome the drawbacks associated with their poor stability and bioavailability. Polyphenols Human Health Dis. 2014;1: 29–45. https://doi.org/10.1016/B978-0-12-398456-2.00004-9
- Zhou HT, Xu LL, Shi Y, Gu SH, Wu N, Liu F, et al. A novel myricetin derivative with anti-cancer properties induces cell cycle arrest and apoptosis in A549 cells. Biol Pharm Bull. 2023;46(1):42–51. https://doi.org/10.1248/bpb.b22-00483
- Zhang JF, Aray B, Zhang Y, Bai YL, Yuan T, Ding SL, et al. Synergistic effect of cucurbitacin E and myricetin on anti-nonsmall cell lung cancer: molecular mechanism and therapeutic potential. Phytomedicine. 2023;111:154619. https://doi.org/10. 1016/j.phymed.2022.154619

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