

RESEARCH ARTICLE

# $\beta$ 7 integrins contribute to intestinal tumor growth in mice

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## Abstract

The gut homing receptor integrin  $\alpha$ 4 $\beta$ 7 is essential for the migration of pro-inflammatory T cells into the gut mucosa. Since intestinal neoplasia has been associated with chronic inflammation, we investigated whether interfering with gut-homing affects intestinal tumorigenesis. Using chemically induced and spontaneous intestinal tumor models we showed that lack of  $\beta$ 7 integrin significantly impairs tumor growth without affecting tumor frequencies, with a mild translatable effect on overall survival. This correlates with human data showing lower MAdCAM-1 expression and disease-free survival in colorectal cancer patients. Thus, paradoxically in contrast to extra-intestinal tumors, blocking migration of immune cells into the gut might have a positive therapeutic effect on intestinal neoplasia.

## Introduction

Colorectal cancer (CRC) is the second most common cancer in women and the third most common in men around the world [1] [2]. Among potential causes, intestinal inflammation is associated with a significantly increased risk of developing colorectal cancer [3], and non-steroidal anti-inflammatory drugs decrease the incidence of intestinal cancer in mice and humans [4–6]. In agreement, inflammatory bowel disease (IBD) patients have an increased risk to develop CRC, likely due to elevated inflammatory cytokines [7]. In addition, CD25<sup>+</sup> Foxp3<sup>+</sup> regulatory T cells (T<sub>REG</sub>) can prevent the development of inflammation and intestinal cancer in two murine models [8, 9], suggesting that intestinal inflammation plays a key role in the pathogenesis of intestinal neoplasia. In agreement, tumor infiltrating FOXP3<sup>+</sup> Treg density in human colorectal cancer was shown to be associated with improved survival compared to CD8<sup>+</sup> and CD45RO<sup>+</sup> lymphocytes [10].

T effector cells (T<sub>EFF</sub>) continuously monitor the body to detect and destroy neoplastic cells, a process known as tumor immunosurveillance. However, while T<sub>EFF</sub> are typically protective against tumors in peripheral tissues, they paradoxically promote inflammation-driven neoplasia in the gut [11, 12]. On the other hand, T<sub>REG</sub> which are conventionally immunosuppressive

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T cells, by virtue of their anti-inflammatory properties, might prevent and even reverse tumorigenesis in the gut, presumably by dampening inflammation. Thus, the gut mucosa represents a paradox regarding tumor immunosurveillance, as observed in IBD patients who exhibit higher risk of developing colorectal cancer [3, 13].

Given that pro-inflammatory T<sub>EFF</sub> cells (Th<sub>1</sub> and Th<sub>17</sub>) require the gut-homing receptor  $\alpha 4\beta 7$  to migrate to the gut mucosa [14], we sought to investigate whether  $\alpha 4\beta 7$  is required for tumorigenesis. Using two intestinal tumor models we show that  $\beta 7$  integrins are not required for tumorigenesis, but lack of these integrins have a clear impact on tumor size. The involvement of  $\beta 7$  integrins in intestinal tumor growth suggests that blocking gut-homing receptors could be an adjuvant strategy to prevent and/or treat intestinal neoplasia in the context of chronic gut inflammation, as in IBD patients.

## Materials and methods

### Mice

Itgb7<sup>-/-</sup>, C57BL/6, and Apc<sup>Min/+</sup> (C57BL/6J-APC<sup>Min/J</sup>) mice were purchased from Jackson Laboratories. Apc<sup>Min/+</sup>;Itgb7<sup>-/-</sup> were generated by breeding Apc<sup>Min/+</sup> and Itgb7<sup>-/-</sup>. Mice were maintained in SPF/VAF animal facilities at Massachusetts General Hospital (MGH) and University of Gothenburg. This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the Subcommittee on Research Animal Care at MGH and Harvard Medical School and the government animal ethics committee at the University of Gothenburg, respectively. The protocol was approved by the Committee on the Ethics of Animal Experiments of the MGH (Ethical permit number 2007N000115) and University of Gothenburg (Ethical permit number 84–2014). All efforts were made to minimize animal suffering.

### DSS-induced colon tumor progression

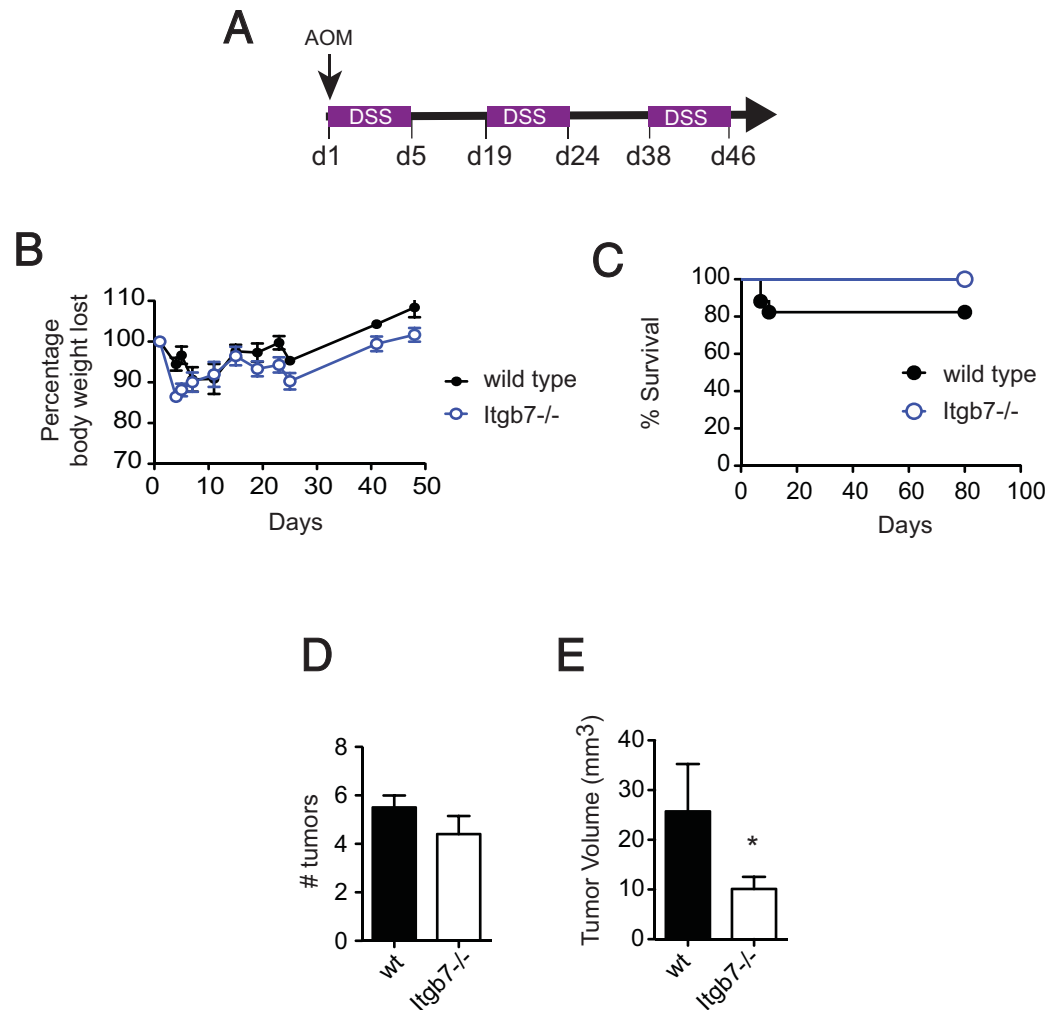
Wild type or Itgb7<sup>-/-</sup> mice received one intraperitoneal injection of AOM (Sigma Aldrich) at 10 mg/kg body weight followed by 3 cycles of DSS treatments (Fig 1). Mice were sacrificed and tumor analysis were done 10 weeks after AOM injection, corresponding to day 10 after 3 cycles of DSS treatment or after initial signs of distress, such as more than 15% of body weight loss, dehydration or hunched posture.

### Competitive homing experiments

Total T cells isolated from spleen of wild type (CD45.1<sup>het</sup>) were mixed with the congenic Itgb7<sup>-/-</sup> counterpart (CD45.1<sup>homo</sup>) in a 1:1 ratio and then adoptively transferred intravenously into 10 weeks old Apc<sup>Min/+</sup> recipients (CD45.2). Mice were euthanized after 12 hours and single cell suspensions from the spleen, small intestine lamina propria (SILP) and intestinal tumors were obtained for analysis. Cell samples were stained to detect T cells (CD3), live cells and congenic markers (CD45.1 and CD45.2) and analyzed on a fluorescence-activated cell sorter (FACS) LSRII (BD Biosciences). Analysis and homing index calculation was performed as described elsewhere [15].

### Transcript analysis in CRC patients

To investigate the relation between MAdCAM-1 mRNA expression and CRC patient survival we used the “R2: Genomics Analysis and Visualization Platform” (<http://r2.amc.nl>), which contains genome-wide gene expression data from human diseases. The Kaplan curve with gene expression was automatically displayed using the “Tumor Colon CIT (Combat)–Marisa



**Fig 1. Mice lacking  $\beta 7$  integrins ( $Itgb7^{-/-}$ ) exhibit impaired tumor development in a chemical/inflammation-driven colon tumorigenesis model.** (A) Colitis-induced tumorigenesis: AOM (10 mg/kg body weight) was injected i.p. at day 1 followed by 3 cycles of 2.5% DSS. (B) Weight loss of wild type (wt) and  $Itgb7^{-/-}$  mice ( $n = 8-14$ ). (C) Kaplan-Meier plot of wt and  $Itgb7^{-/-}$  mice survival ( $n = 8-14$ ). Macroscopic tumors were counted (D) and tumor volume measured (E) after 10 weeks ( $n = 8$ ). Mean  $\pm$  SEM, \* $p < 0.05$ .

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566 rma-u133p2” dataset and the MADCAM1 (208037\_s\_at) probe with an expression cutoff of 36.2 (min.grp = 8).

### Statistical analysis

Data are presented as mean  $\pm$  SEM and were analyzed using GraphPad Prism Software 5.0b. Statistics were calculated using either unpaired  $t$  test when comparing two groups or ANOVA with Dunnett’s or Bonferroni’s post-hoc test when comparing more than 2 groups. Significance was set at  $p < 0.05$ .

### Results and discussion

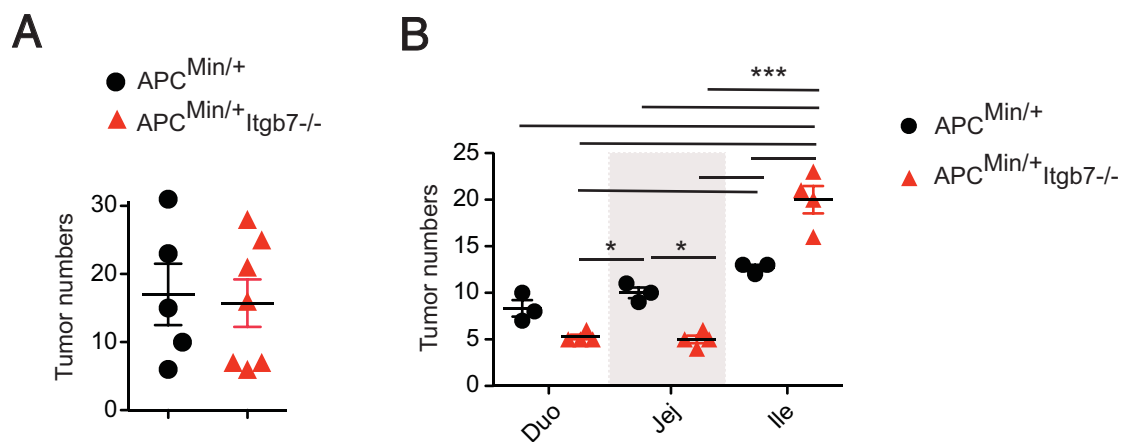
To investigate whether intestinal tumor development requires gut-tropic immune cells we performed the inflammation-driven AOM/DSS model of colon tumorigenesis (Fig 1A) [16] in

wild type (wt) or  $\beta 7$  integrin chain (*Itgb7*)-deficient mice, which lacks both  $\beta 7$  integrins ( $\alpha 4\beta 7$  and  $\alpha E\beta 7$ ) [17, 18]. AOM/DSS-induced tumors exhibit high levels of CD3<sup>+</sup> cell infiltration [19], suggesting that lymphocyte migration to the gut might play a role in tumor progression. Both wt and *Itgb7*<sup>-/-</sup> mice showed a similar decrease in body weight upon AOM/DSS treatment and repeated DSS exposure (Fig 1B). Of note, body weight recovery was similar between wt and *Itgb7*<sup>-/-</sup> mice during the water cycles, and although *Itgb7*<sup>-/-</sup> showed a tendency to lose more weight upon DSS exposure, all *Itgb7*<sup>-/-</sup> mice survived for the length of the study. By contrast, 20% of wt mice were euthanized due to signs of marked distress (Fig 1C).

Treatment with AOM/DSS triggered tumorigenesis in all wt and *Itgb7*<sup>-/-</sup> mice analyzed ( $n = 8-14$ ), and by week 10 tumor numbers were comparable in wt ( $5.5 \pm 0.7$ ) and *Itgb7*<sup>-/-</sup> ( $4.4 \pm 1.6$ ) mice (Fig 1D). However, tumor mass was significantly less in *Itgb7*<sup>-/-</sup> compared to wt mice (Fig 1E), and the average tumor volume per mouse was  $25 \pm 10.7 \text{ mm}^3$  in wt versus  $9.9 \pm 6.1$  in *Itgb7*<sup>-/-</sup> mice. Taken together, our data suggest that gut-tropic immune cells are not required for tumor initiation, but they might promote tumor growth in the context of inflammation-driven intestinal tumorigenesis.

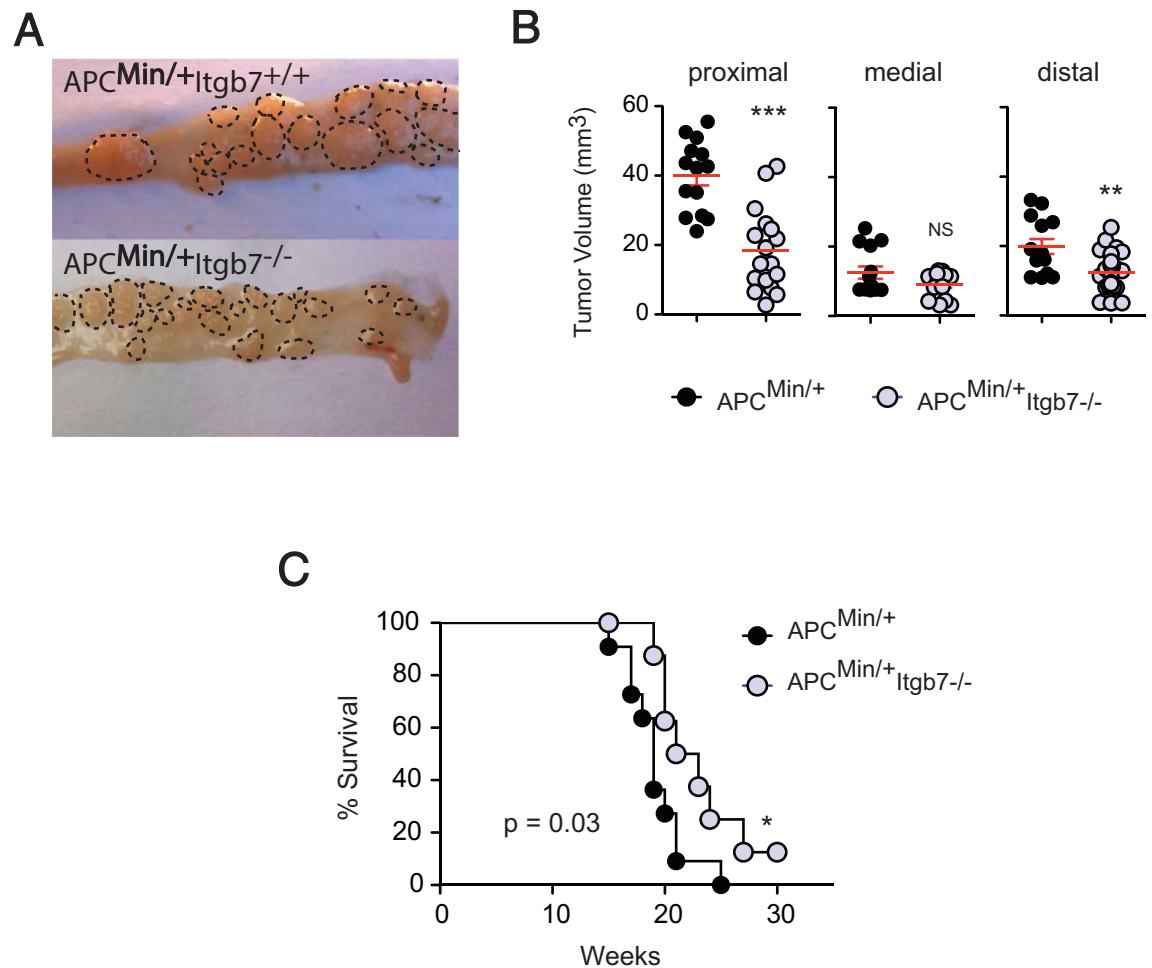
*Apc* is the most commonly mutated gene in both familial and sporadic human colorectal cancer [20], making *Apc*<sup>Min/+</sup> mice a relevant translational model to study intestinal tumorigenesis. We then asked whether lack of  $\beta 7$  integrins affect tumor development in the *Apc*<sup>Min/+</sup> spontaneous model of intestinal tumorigenesis. Consistent with our previous results using a chemically-induced inflammation-driven tumorigenesis model, we observed that *Apc*<sup>Min/+</sup>; *Itgb7*<sup>-/-</sup> mice exhibit similar numbers of intestinal tumors in the whole small bowel when compared to *Apc*<sup>Min/+</sup> mice (Fig 2A). However, we observed some regional differences, with lower tumor numbers in the duodenum and jejunum of *Apc*<sup>Min/+</sup>; *Itgb7*<sup>-/-</sup> mice, but increased tumor numbers in the ileum when compared to *Apc*<sup>Min/+</sup> mice (Fig 2B).

Moreover, consistent with our results in the AOM/DSS model, we observed smaller tumors in *Apc*<sup>Min/+</sup>; *Itgb7*<sup>-/-</sup> compared to *Apc*<sup>Min/+</sup> mice (Fig 3A and 3B), which correlated with a slightly better survival in *Apc*<sup>Min/+</sup>; *Itgb7*<sup>-/-</sup> mice (Fig 3C). Thus, similar to AOM/DSS model, our data suggest that gut-tropic immune cells also promote tumor growth in the spontaneous *Apc*<sup>Min/+</sup> intestinal tumor model.



**Fig 2. Altered tumor distribution in *Apc*<sup>Min/+</sup>; *Itgb7*<sup>-/-</sup> mice.** (A) *Apc*<sup>Min/+</sup> and *Apc*<sup>Min/+</sup>; *Itgb7*<sup>-/-</sup> mice were euthanized between 20–25 weeks, the small bowel was dissected, open longitudinally, and mounted for macroscopic tumor analysis. (A) Number of tumors throughout the small bowel length ( $n = 3-4$  mice). (B) The small bowel was divided into duodenum (Duo), jejunum (Jej) and ileum (Ile) and macroscopic tumors were counted ( $n = 3-4$ ). Mean  $\pm$  SEM, ANOVA with Bonferroni as post-test \* $p < 0.05$ , \*\*\* $p < 0.001$ .

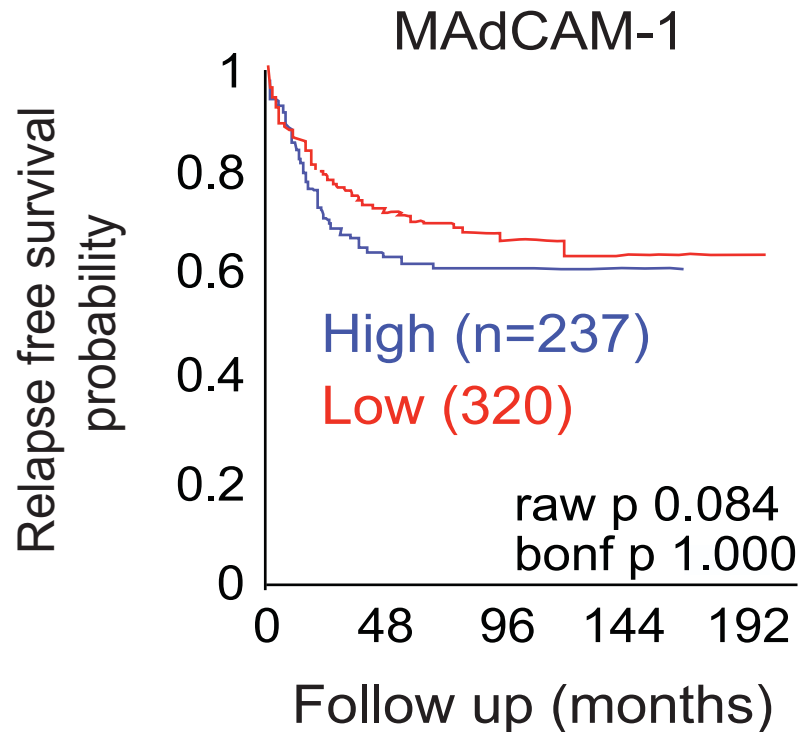
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**Fig 3. Decreased tumor size in *Apc*<sup>Min/+</sup>;Itgb7<sup>-/-</sup> mice.** *Apc*<sup>Min/+</sup> and *Apc*<sup>Min/+</sup>;Itgb7<sup>-/-</sup> mice were euthanized between 20–25 weeks and the small bowel was dissected and open longitudinally for macroscopic tumor analysis. (A) Dashed line delineates single tumors within the small bowel (smaller tumor areas in *Apc*<sup>Min/+</sup>;Itgb7<sup>-/-</sup> mice). Pictures are representative of two independent experiments. (B) Tumor volume measured in the duodenum (proximal), jejunum (medial) and ileum (distal). (C) Kaplan-Meier plot of *Apc*<sup>Min/+</sup> and *Apc*<sup>Min/+</sup>;Itgb7<sup>-/-</sup> mice survival (*n* = 10 mice/group, 3 experiments). Data are from three independent experiments. Mean ± SEM. Unpaired t-test, \*\**p* < 0.01, \*\*\**p* < 0.001.

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Survival analysis of publicly available datasets in human CRCs suggests that patients expressing lower levels of MAdCAM-1, the ligand of  $\alpha 4\beta 7$ , in the tumor have better survival compared to patients expressing higher levels (Fig 4). This suggests that decreased  $\alpha 4\beta 7^+$  cell might be associated with decreased CRC progression in humans. Although the impact of the immune system in colorectal cancer progression is yet poorly understood, it is well appreciated that combination of cell types and immune environment may result in either promotion or inhibition of tumor growth [21]. For example, patients with high expression of Th1 gene cluster had a better prognosis compared to high expression of Th17 cluster which instead had poor prognosis using an unsupervised hierarchical clustering of a set of genes associated with different T cell subsets [22]. Likewise, even if CD4<sup>+</sup> T cells have been implicated in tumor surveillance, IL-22-producer CD4<sup>+</sup> T cells (Th22) migrate and infiltrate the tumor to eventually promote stemness and tumorigenic potential [23]. We next attempt to investigate the mechanisms by which Itgb7 may promote tumor growth. Hence, we next sought to investigate whether the reduced tumor volume observed in *Apc*<sup>Min/+</sup>;Itgb7<sup>-/-</sup> mice was associated with



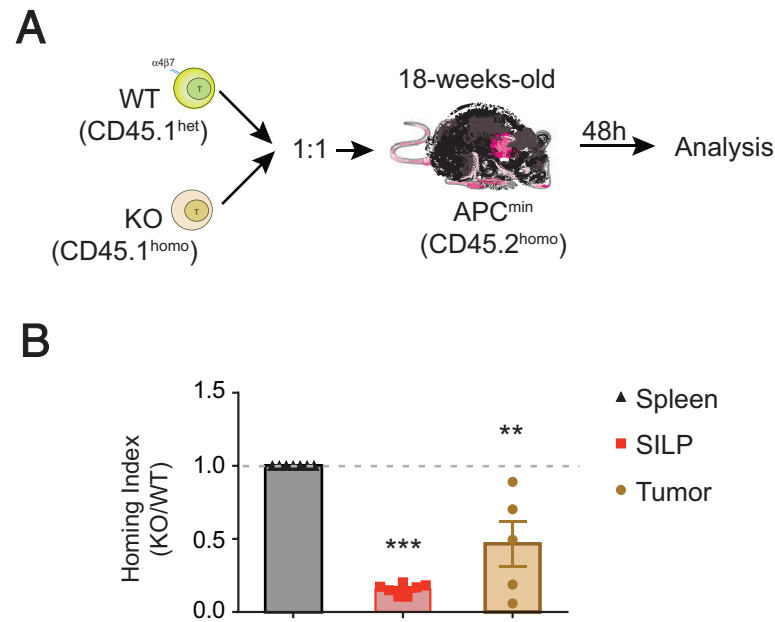
**Fig 4. Lower MAdCAM-1 expression is associated with increase survival in CRC patients.** Kaplan-Meier analysis showing disease free survival of patients with CRC ( $n = 557$ ) on the basis of *madcam-1* mRNA expression using publicly available data sets (source: AMC onco genomics).

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decrease T cell infiltration within the tumor. To assess whether T cells require Itgb7 to infiltrate the tumor, we performed homing competitive experiments in which congenic total WT and Itgb7<sup>-/-</sup> T cells were adoptively transferred in a 1:1 ratio into 10 weeks old Apc<sup>Min/+</sup> mice (Fig 5A). Twelve hours after the injection the Apc<sup>Min/+</sup> mice were sacrificed and T cells ratios were analyzed. The homing index (the ratio of KO to WT T cells, corrected by the input) was one in the spleen indicating that either WT or Itgb7<sup>-/-</sup> preferentially populate this organ. As expected, Itgb7<sup>-/-</sup> T cells home poorly to the SILP compared to WT T cells as seen by a homing index of ~0.2 (Fig 5B). Analysis of T cell infiltrating the tumors revealed that Itgb7<sup>-/-</sup> T cells were at a disadvantage to WT T cells in homing to the tumors as seen by a homing index of ~0.5 (Fig 5B), indicating that Itgb7 provides T cells a higher capacity to home to intestinal tumors.

Our data suggest that immune cells expressing  $\beta 7$  integrins are not required for tumor initiation in the gut but might promote tumor growth. Therefore, the earliest molecular events initiating the appearance of intestinal tumors might be independent of cell migration to the intestine, at least in a  $\beta 7$  integrin-dependent manner. Supporting this hypothesis, it has been shown that alteration of inflammasome or MyD88 signaling pathways in the intestinal epithelium, but not in hematopoietic-derived cells, results in a higher susceptibility to develop colon cancer [24], and that specific *Apc* mutations in intestinal stem cells (using Lgr5-cre mice) are sufficient to induce rapid tumor development [25].

On the other hand, gut-tropic immune cells are believed to drive intestinal tumor progression by promoting angiogenesis, inflammation and/or proliferation of cancer cells [3, 23, 26]. Therefore, the decreased tumor sizes in Apc<sup>Min/+</sup>;Itgb7<sup>-/-</sup> as well as in AOM/DSS-treated



**Fig 5. T cells require Itgb7 to home to the intestinal tumors.** (A) Congenic WT (CD45.1<sup>het</sup>) and Itgb7<sup>-/-</sup> (CD45.1<sup>homo</sup>) total T cells were isolated from the spleen and injected intravenously in a 1:1 ratio into 10-week-old (ongoing tumor development) Apc<sup>Min/+</sup> mice. Twelve hours after the injection, T cell ratios were analyzed in the spleen, small intestine lamina propria (SILP) and tumors. (B) The homing index was calculated as the ratio KO to WT divided by the ratio KO to WT of the input. Data represent two different experiments with 3–4 mice each. Mean  $\pm$  SEM, ANOVA with Dunnett as post-test, \*\* $p < 0.01$ , \*\*\* $p < 0.001$ .

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Itgb7<sup>-/-</sup> mice could be explained by impaired homing of pro-inflammatory T<sub>EFF</sub> to the gut. An apparent discrepancy with this hypothesis is the fact that higher numbers of Th1 cells as well as cytotoxic CD8<sup>+</sup> T cells within tumors correlate with a better prognosis [3]. However, a recent study showed comparable numbers of cytotoxic- and regulatory- CD8<sup>+</sup> T cells infiltrating the tumor [3], suggesting that regulatory rather than cytotoxic-CD8 T cells account for a better prognosis factor. Indeed, it has been proposed that T<sub>REG</sub> rather than CD8<sup>+</sup>, and CD45RO<sup>+</sup> T cell density in the tumor tissues as a stronger prognosis factor in CRC [3]. Of note, location of Foxp3<sup>+</sup> T<sub>REG</sub> in the gut might not critically rely on gut-homing receptors [27]. Therefore, the net effect of lacking  $\alpha$ 4 $\beta$ 7 would be an increased T<sub>REG</sub> / T<sub>EFF</sub> ratio, which might contribute to decreased inflammation and subsequent intestinal tumorigenesis. In the same line, CD4<sup>+</sup> T helper subsets might differentially contribute to either tumor initiation, progression or regression. For instance, Th1 seems to be protective as seen by a better survival in patients with higher Th1 numbers [28]. Whereas Th22 cells might promote tumor growth [29]. Hence, it would be interesting to determine whether specific T helper subsets (e.g. Th1) require Itgb7 to modulate tumorigenesis. Experiments in which WT or Itgb7<sup>-/-</sup> *in vitro* differentiated T helper subsets transferred in mice undergoing intestinal tumorigenesis might provide important insights into this question.

In addition to lymphocytes, some innate immune cells also express  $\alpha$ 4 $\beta$ 7 [30], and future studies will be aimed to dissect the relative contributions of the specific gut-tropic adaptive and innate immune cells subsets in intestinal tumorigenesis. Our findings might have translational implications, as drugs blocking gut-homing receptors such as vedolizumab (anti- $\alpha$ 4 $\beta$ 7) are currently widely used in IBD patients to ameliorate inflammation, and colon cancer is a potential complication of chronic intestinal inflammation in these patients.

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## Author Contributions

**Conceptualization:** Srustidhar Das, Marianne Quiding-Järbrink, J. Rodrigo Mora, Eduardo J. Villablanca.

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**Funding acquisition:** Marianne Quiding-Järbrink, J. Rodrigo Mora, Eduardo J. Villablanca.

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**Resources:** Eduardo J. Villablanca.

**Supervision:** J. Rodrigo Mora, Eduardo J. Villablanca.

**Writing – review & editing:** Srustidhar Das, Cristian Doñas, Paulina Akeus, Marianne Quiding-Järbrink, J. Rodrigo Mora, Eduardo J. Villablanca.

## References

1. Siegel RL, Miller KD, Jemal A. Cancer Statistics, 2017. *CA Cancer J Clin.* 2017; 67(1):7–30. Epub 2017/01/06. <https://doi.org/10.3322/caac.21387> PMID: 28055103.
2. Jemal A, Siegel R, Ward E, Murray T, Xu J, Thun MJ. Cancer statistics, 2007. *CA Cancer J Clin.* 2007; 57(1):43–66. Epub 2007/01/24. PMID: 17237035.
3. West NR, McCuaig S, Franchini F, Powrie F. Emerging cytokine networks in colorectal cancer. *Nat Rev Immunol.* 2015; 15(10):615–29. Epub 2015/09/12. <https://doi.org/10.1038/nri3896> PMID: 26358393.
4. Corpet DE, Pierre F. Point: From animal models to prevention of colon cancer. Systematic review of chemoprevention in min mice and choice of the model system. *Cancer Epidemiol Biomarkers Prev.* 2003; 12(5):391–400. Epub 2003/05/17. PMID: 12750232; PubMed Central PMCID: PMC2797538.
5. Garcia-Rodriguez LA, Huerta-Alvarez C. Reduced risk of colorectal cancer among long-term users of aspirin and nonaspirin nonsteroidal antiinflammatory drugs. *Epidemiology.* 2001; 12(1):88–93. PMID: 11138826.
6. Sansbury LB, Millikan RC, Schroeder JC, Moorman PG, North KE, Sandler RS. Use of nonsteroidal antiinflammatory drugs and risk of colon cancer in a population-based, case-control study of African Americans and Whites. *Am J Epidemiol.* 2005; 162(6):548–58. <https://doi.org/10.1093/aje/kwi248> PMID: 16093288.
7. Dahlhamer JM, Zammitti EP, Ward BW, Wheaton AG, Croft JB. Prevalence of Inflammatory Bowel Disease Among Adults Aged  $\geq 18$  Years—United States, 2015. *MMWR Morb Mortal Wkly Rep.* 2016; 65(42):1166–9. Epub 2016/10/28. <https://doi.org/10.15585/mmwr.mm6542a3> PMID: 27787492.
8. Erdman SE, Rao VP, Poutahidis T, Ihrig MM, Ge Z, Feng Y, et al. CD4(+)CD25(+) regulatory lymphocytes require interleukin 10 to interrupt colon carcinogenesis in mice. *Cancer Res.* 2003; 63(18):6042–50. PMID: 14522933.
9. Erdman SE, Sohn JJ, Rao VP, Nambiar PR, Ge Z, Fox JG, et al. CD4+CD25+ regulatory lymphocytes induce regression of intestinal tumors in ApcMin/+ mice. *Cancer Res.* 2005; 65(10):3998–4004. <https://doi.org/10.1158/0008-5472.CAN-04-3104> PMID: 15899788.
10. Salama P, Phillips M, Grieu F, Morris M, Zeps N, Joseph D, et al. Tumor-infiltrating FOXP3+ T regulatory cells show strong prognostic significance in colorectal cancer. *J Clin Oncol.* 2009; 27(2):186–92. Epub 2008/12/10. <https://doi.org/10.1200/JCO.2008.18.7229> PMID: 19064967.
11. Chae WJ, Gibson TF, Zelterman D, Hao L, Henegariu O, Bothwell AL. Ablation of IL-17A abrogates progression of spontaneous intestinal tumorigenesis. *Proc Natl Acad Sci U S A.* 2010; 107(12):5540–4. <https://doi.org/10.1073/pnas.0912675107> PMID: 20212110; PubMed Central PMCID: PMC2851824.
12. Schoonjans K, Dubuquoy L, Mebis J, Fayard E, Wendling O, Haby C, et al. Liver receptor homolog 1 contributes to intestinal tumor formation through effects on cell cycle and inflammation. *Proc Natl Acad*



- Sci U S A. 2005; 102(6):2058–62. <https://doi.org/10.1073/pnas.0409756102> PMID: 15684064; PubMed Central PMCID: PMCPMC548586.
13. Feagins LA, Souza RF, Spechler SJ. Carcinogenesis in IBD: potential targets for the prevention of colorectal cancer. *Nature reviews Gastroenterology & hepatology*. 2009; 6(5):297–305. Epub 2009/05/01. <https://doi.org/10.1038/nrgastro.2009.44> PMID: 19404270.
  14. Do JS, Visperas A, Freeman ML, Iwakura Y, Oukka M, Min B. Colitogenic effector T cells: roles of gut-homing integrin, gut antigen specificity and gammadelta T cells. *Immunol Cell Biol*. 2014; 92(1):90–8. Epub 2013/11/06. <https://doi.org/10.1038/icb.2013.70> PMID: 24189163; PubMed Central PMCID: PMCPMC3947309.
  15. Villablanca EJ, Mora JR. Competitive homing assays to study gut-tropic t cell migration. *J Vis Exp*. 2011;(49). Epub 2011/03/16. doi: 2619 [pii] <https://doi.org/10.3791/2619>. PMID: 21403633; PubMed Central PMCID: PMC3197317.
  16. Neufert C, Becker C, Neurath MF. An inducible mouse model of colon carcinogenesis for the analysis of sporadic and inflammation-driven tumor progression. *Nat Protoc*. 2007; 2(8):1998–2004. Epub 2007/08/19. <https://doi.org/10.1038/nprot.2007.279> PMID: 17703211.
  17. Park EJ, Mora JR, Carman CV, Chen J, Sasaki Y, Cheng G, et al. Aberrant activation of integrin alpha4-beta7 suppresses lymphocyte migration to the gut. *J Clin Invest*. 2007; 117(9):2526–38. Epub 2007/09/06. <https://doi.org/10.1172/JCI31570> PMID: 17786243; PubMed Central PMCID: PMCPMC1952632.
  18. Gorfu G, Rivera-Nieves J, Ley K. Role of beta7 integrins in intestinal lymphocyte homing and retention. *Curr Mol Med*. 2009; 9(7):836–50. Epub 2009/10/29. PMID: 19860663; PubMed Central PMCID: PMCPMC2770881.
  19. Barrett CW, Fingleton B, Williams A, Ning W, Fischer MA, Washington MK, et al. MTGR1 is required for tumorigenesis in the murine AOM/DSS colitis-associated carcinoma model. *Cancer Res*. 2011; 71(4):1302–12. Epub 2011/02/10. <https://doi.org/10.1158/0008-5472.CAN-10-3317> PMID: 21303973; PubMed Central PMCID: PMCPMC3150168.
  20. Nagamine CM, Sohn JJ, Rickman BH, Rogers AB, Fox JG, Schauer DB. Helicobacter hepaticus infection promotes colon tumorigenesis in the BALB/c-Rag2(-/-) Apc(Min/+) mouse. *Infect Immun*. 2008; 76(6):2758–66. <https://doi.org/10.1128/IAI.01604-07> PMID: 18411292; PubMed Central PMCID: PMCPMC2423080.
  21. Norton SE, Ward-Hartstonge KA, Taylor ES, Kemp RA. Immune cell interplay in colorectal cancer prognosis. *World J Gastrointest Oncol*. 2015; 7(10):221–32. Epub 2015/10/21. <https://doi.org/10.4251/wjgo.v7.i10.221> PMID: 26483876; PubMed Central PMCID: PMCPMC4606176.
  22. Tosolini M, Kirilovsky A, Mlecnik B, Fredriksen T, Mauger S, Bindea G, et al. Clinical impact of different classes of infiltrating T cytotoxic and helper cells (Th1, th2, treg, th17) in patients with colorectal cancer. *Cancer Res*. 2011; 71(4):1263–71. Epub 2011/02/10. <https://doi.org/10.1158/0008-5472.CAN-10-2907> PMID: 21303976.
  23. Kryczek I, Lin Y, Nagarsheth N, Peng D, Zhao L, Zhao E, et al. IL-22(+)CD4(+) T cells promote colorectal cancer stemness via STAT3 transcription factor activation and induction of the methyltransferase DOT1L. *Immunity*. 2014; 40(5):772–84. Epub 2014/05/13. <https://doi.org/10.1016/j.immuni.2014.03.010> PMID: 24816405; PubMed Central PMCID: PMCPMC4032366.
  24. Saleh M, Trinchieri G. Innate immune mechanisms of colitis and colitis-associated colorectal cancer. *Nat Rev Immunol*. 2011; 11(1):9–20. Epub 2010/12/15. <https://doi.org/10.1038/nri2891> PMID: 21151034.
  25. Barker N, Ridgway RA, van Es JH, van de Wetering M, Begthel H, van den Born M, et al. Crypt stem cells as the cells-of-origin of intestinal cancer. *Nature*. 2009; 457(7229):608–11. Epub 2008/12/19. <https://doi.org/10.1038/nature07602> PMID: 19092804.
  26. Elinav E, Nowarski R, Thaiss CA, Hu B, Jin C, Flavell RA. Inflammation-induced cancer: crosstalk between tumours, immune cells and microorganisms. *Nat Rev Cancer*. 2013; 13(11):759–71. Epub 2013/10/25. <https://doi.org/10.1038/nrc3611> PMID: 24154716.
  27. Wang C, Kang SG, HogenEsch H, Love PE, Kim CH. Retinoic acid determines the precise tissue tropism of inflammatory Th17 cells in the intestine. *J Immunol*. 2010; 184(10):5519–26. Epub 2010/04/20. <https://doi.org/10.4049/jimmunol.0903942> PMID: 20400707; PubMed Central PMCID: PMCPMC3009589.
  28. Muzes G, Molnar B, Sipos F. Regulatory T cells in inflammatory bowel diseases and colorectal cancer. *World J Gastroenterol*. 2012; 18(40):5688–94. Epub 2012/11/17. <https://doi.org/10.3748/wjg.v18.i40.5688> PMID: 23155308; PubMed Central PMCID: PMCPMC3484336.
  29. Huber S, Gagliani N, Zenewicz LA, Huber FJ, Bosurgi L, Hu B, et al. IL-22BP is regulated by the inflammasome and modulates tumorigenesis in the intestine. *Nature*. 2012; 491(7423):259–63. <https://doi.org/10.1038/nature11535> PMID: 23075849; PubMed Central PMCID: PMC3493690.

30. Villablanca EJ, De Calisto J, Torregrosa Paredes P, Cassani B, Nguyen DD, Gabrielsson S, et al. beta7 integrins are required to give rise to intestinal mononuclear phagocytes with tolerogenic potential. *Gut*. 2014; 63(9):1431–40. <https://doi.org/10.1136/gutjnl-2013-305386> PMID: 24030488.