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# Chromatography/Mass Spectrometry-Based Biomarkers in the Field of Obstructive Sleep Apnea

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**Abstract:** Biomarker assessment is based on quantifying several proteins and metabolites. Recent developments in proteomics and metabolomics have enabled detection of these small molecules in biological samples and exploration of the underlying disease mechanisms in obstructive sleep apnea (OSA). This systemic review was performed to identify biomarkers, which were only detected by chromatography and/or mass spectrometry (MS) and to discuss the role of these biomarkers in the field of OSA.

We systemically reviewed relevant articles from PubMed and EMBASE referring to proteins and metabolite profiles of biological samples in patients with OSA. The analytical platforms in this review were focused on chromatography and/or MS.

In total, 30 studies evaluating biomarkers in patients with OSA using chromatography and/or MS methods were included. Numerous proteins and metabolites, including lipid profiles, adrenergic/dopaminergic biomarkers and derivatives, amino acids, oxidative stress biomarkers, and other micromolecules were identified in patients with OSA.

Applying chromatography and/or MS methods to detect biomarkers helps develop an understanding of OSA mechanisms. More proteomic and metabolomic studies are warranted to develop potential diagnostic and clinical monitoring methods for OSA.

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**Abbreviations:** <sup>1</sup>NMR = proton nuclear magnetic resonance spectroscopy, 2D-DIGE-MS = 2-dimensional polyacrylamide gel electrophoresis mass spectrometry, 5-HIAA = 5-hydroxyindoleacetic acid, AA = arachidonic acid, ADMA = asymmetric dimethylarginine, AEA = anandamide, AG = arachidonoylglycerols, AHSG = alpha-2-HS-glycoprotein, ApoB = apolipoprotein B, CO = carbon monoxide, CPAP = continuous positive airway pressure, CVD = cardiovascular disease, DA =

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dopamine, DOPAC = 34-dihydroxyphenylacetic acid, Eepinephrine, ELISA = enzyme-linked immunosorbent assay, FGA = fibrinogen alpha chain, GC = gas chromatography, GC/ MS = gas chromatography-mass spectrometry, GLC = gas-liquid chromatography, Hcy = homocysteine, HPLC = high-pressure or high-performance liquid chromatography, HVA = homovanillic acid, HVA = homovanillic acid, IH = intermittent hypoxia, LBP = lipopolysaccharide-binding protein, LC = liquid chromatography, LC/MS = liquid chromatography-mass spectrometry, LC-ESI-MS/ MS = liquid chromatography coupled with electrospray source ionization-tandem mass spectrometry, LC-MS/MS = liquid chromatography-tandem mass spectrometry, LC-QTOF-MS = liquid chromatography-hybrid quadrupole time-of-flight mass spectrometry, LPA = lysophosphatidic acids, LPC = lysophosphatidylcholines, LPE = lysophosphatidylethanolamine, MALDI-TOF-MS = matrix-assisted laser desorption/ionization time-of-flight mass spectrometry, MDA = malondialdehyde, MHPG = 3-methoxy-4-hydroxyphenyl glycol, MN = metanephrine, MS = mass spectrometry, NE = norepinephrine, NMN = normetanephrine, OEA = oleoyl ethanolamide, OSA = obstructive sleep apnea, PC = phosphatidylcholine, PE = phosphoethanolamine, PRISMA = Preferred Reporting Items for Systematic Reviews and Meta-Analyses, PS = phosphoserine, PSG = polysomnography, T2DM = type 2 diabetes, TUBA4A = tubulin alpha-4A chain, U-LTE(4) = leukotriene E(4).

## INTRODUCTION

O bstructive sleep apnea (OSA) is one of the most common sleep disorders, affecting about 4% of middle-aged males and 2% of middle-aged females.<sup>1</sup> Moreover, the frequency of OSA has increased along with the obesity epidemic and the changes in lifestyle that have occurred over the last 2 decades.<sup>2</sup> OSA is presently characterized as recurrent episodes of a completely or partly obstructed upper airway during sleep.<sup>3</sup> The immediate effects caused by episodes of apnea and hypopnea include large intra-thoracic pressure swings, intermittent hypoxia (IH), frequent micro-arousals, and fragmented sleep.<sup>4</sup> The ensuing sympathetic activation, oxidative stress, and inflammatory cytokines are correlated closely with a range of sequelae (ie, hypertension, stroke, cardiovascular disease (CVD), diabetes mellitus, and cancer).<sup>5–11</sup> Moreover, OSA is associated with all-cause mortality.<sup>12</sup> Thus, OSA has become a major challenge for global public healthcare.

To date, the gold standard tool to diagnose OSA is overnight polysomnography (PSG).<sup>13</sup> However, PSG itself can lead to sleep disturbance. Moreover, PSG is expensive and uncomfortable for patients with suspected OSA. In addition, it is laborintensive for the polysomnographic technologist. Thus, more comfortable and easier tools as surrogates for PSG monitoring, or alternative tests, should be developed. Identifying potential biomarkers would facilitate the clinical decision-making process. Unfortunately, no ideal biomarkers for early diagnosis,

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severity, prognosis, or response to OSA treatment were found by reviewing the literature.<sup>14</sup>

Systems biology, including genomics, transcriptomics, proteomics, and metabolomics, have been extensively applied to various sleep diseases.<sup>15,16</sup> Proteomics and metabolomics have been widely used to discover differences in small molecules downstream of genetic or environmental variations in chronic kidney disease,<sup>17,18</sup> polycystic ovary syndrome,<sup>19,20</sup> cancers,<sup>21,22</sup> diabetes mellitus,<sup>23,24</sup> and pulmonary diseases,<sup>25,26</sup> and have shown promising initial results.

Proteomics is the large-scale study of the proteome, particularly analysis of its structure and function.<sup>27</sup> Metabolomics refers to the study of complete sets of metabolites, which are context-dependent and vary depending on physiology and the developmental or pathological status of cells, tissues, and organisms.<sup>28</sup> Perturbations in biological pathways and changes in protein and metabolite concentration occur when an organism or cell is in an abnormal state. Understanding altered proteomic and metabolic profiles in patients with OSA could improve diagnostic tests as well as uncover new approaches to treat or even prevent OSA.

To date, few studies have used proteomics and metabolomics to reveal differences in small molecules in patients with OSA.<sup>29–36</sup> Other studies have identified several proteins and metabolites using chromatography and/or mass spectrometry (MS) methods. The purpose of this review is to summarize all identified biomarkers associated with OSA.

## **METHODS**

This systematic review was conducted according to the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) statement.<sup>37</sup> Ethical approval was not necessary for this meta-analysis as each included study has already received ethics approval.

## Search Strategy

A comprehensive search was performed in December 2014 using the PubMed and EMBASE electronic databases to identify relevant studies. The following Mesh terms and keywords were used: "sleep disordered breathing," "SDB," "obstructive sleep apnea," "OSA," and "sleep apnea" combined with relevant terms, such as "chromatography" and "MS." In addition, the references of the included studies were also reviewed to increase our search yield. Only studies published in English were included. This search was performed by 2 authors separately.

Two reviewers (HX and XZ) screened titles and abstracts of studies identified through the search, respectively. Full-text articles of studies evaluating metabolic and protein profiles of plasma, urine, adipose tissue, saliva, and tonsillar tissue, among others, from patients with OSA using chromatography and/or MS were included. Exclusion criteria included: not a human study; and did not use overnight PSG to diagnose OSA.

Two reviewers (HX and XZ) independently extracted data from eligible studies using a standard form. The following information was collected: first author, publication year, types of analytical techniques, specimen types, participants, and main biomarkers. Discrepancies were resolved by group discussion. If the protein and metabolite chemical nomenclature differed among studies, their common names were described.

#### RESULTS

### Search Results

We identified 350 citations (113 from PubMed and 237 from EMBASE) following the initial search. Of the 350 references, 71 were removed due to duplication. After screening titles and abstracts, 36 full-text articles were selected for further consideration. Six articles were further excluded for the following reasons: overlapping population,<sup>38,39</sup> animal study,<sup>40–42</sup> and book review.<sup>43</sup> Thus, 30 studies that evaluated biomarkers using chromatography and/or MS in patients with OSA were included in this systematic review (Figure 1).

## **Study Characteristics**

Of the 30 studies, 7 evaluated proteins and 23 evaluated metabolites in patients with OSA. The basic characteristics are shown in Tables 1 and 2.

The analytical platforms used for detecting proteins and metabolites included gas chromatography (GC) (n = 2), liquid



FIGURE 1. Flow chart of the studies included and excluded in this systemic review.

TABLE 1. Totents identified in rationis with Obstituctive sleep Aprica (OSA)								
References	Analytical Technique	Sample	Number of Participants	Proteins				
Krishna et al <sup>30</sup>	MALDI-TOF MS	Urine	11 Pediatric OSA and 11 controls	Gelsolin, perlecan (a heparan sulfate proteoglycan), albumin, and immunoglobulin				
Shah et al <sup>31</sup>	SELDI-TOF MS	Serum	20 Pediatric OSA and 20 controls	Three differentially regulated proteins with molecular masses of 5896, 3306, and 6068 Da				
Gozal et al <sup>32</sup>	2D-DIGE-MS	Urine	30 Pediatric OSA and 30 controls	Uromodulin urocortin-3 orosomucoid-1 kallikrein-1				
Becker et al <sup>33</sup>	LC-ESI-MS/MS	Urine	14 Pediatric OSA and 13 controls	$\sim$ 30-fold more candidate biomarkers				
Jurado-Gamez et al <sup>34</sup>	MALDI-TOF MS	Serum	30 OSA and 10 controls	103 Proteins				
Seetho et al <sup>35</sup>	CE-MS	Urine	61 OSA and 31 controls	24 Polypeptides				
Zheng et al <sup>36</sup>	MALDI-TOF MS	Saliva	20 Non-CVD OSA and 18 CVD OSA	FGA, AHSG, TUBA4A				

2D-DIGE-MS = 2-dimensional polyacrylamide gel electrophoresis mass spectrometry; AHSG = alpha-2-HS-glycoprotein; CE-MS = capillary electrophoresis-mass spectrometry; CVD = cardiovascular disease; FGA = fibrinogen alpha chain; LC-ESI-MS/MS = liquid chromatography-electrospray ionization-tandem mass spectrometry; MALDI-TOF MS = matrix-assisted laser desorption/ionization time-of-flight mass spectrometry; OSA = obstructive sleep apnea; SELDI-TOF MS = surface-enhanced laser desorption/ionization time-of-flight mass spectrometry; TUBA4A = tubulin alpha-4A chain.

chromatography (LC) (n = 2), high-pressure or high-performance liquid chromatography (HPLC) (n = 8), reverse-phase HPLC (n = 1), gas-liquid chromatography (GLC) (n = 1), gas chromatography-mass spectrometry (GC/MS) (n = 4), liquid chromatography-mass spectrometry (LC/MS) (n = 2), liquid chromatography-tandem mass spectrometry (LC-MS/ MS) (n = 2), liquid chromatography coupled with electrospray source ionization-tandem mass spectrometry (LC-ESI-MS/MS) (n = 2), liquid chromatography-hybrid quadrupole time-offlight mass spectrometry (LC-QTOF-MS) (n = 1), matrixassisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF-MS) (n = 3), surface-enhanced laser desorption/ionization time-of-flight mass spectrometry (n = 1), and 2-dimensional polyacrylamide gel electrophoresis mass spectrometry (2D-DIGE-MS) (n = 1).

The biological samples utilized for the protein and metabolite analyses included urine (n = 13), saliva (n = 2), plasma (n = 11), tonsillar tissue (n = 1), adipose tissue (n = 1), venous blood (n = 1), cerebrospinal fluid (n = 1), and serum (n = 2).

## **Protein Profiles in Patients With OSA**

Krishna et al<sup>30</sup> performed the first study on urinary protein expression in children with OSA and revealed that the levels of particular proteins (gelsolin, perlecan, albumin, and immunoglobulin) were different from those of primary snorers. This preliminary study suggested that altered renal glomerular/tubular permeability and increased catabolism of certain proteins occur in patients with OSA.<sup>30</sup> Almost at the same time, Shah et al<sup>31</sup> determined that levels of 3 serum proteins with masses of 5896, 3306, and 6068 kDa in children with OSA were different from those of controls, and that these proteomic biomarkers could be used to diagnose OSA with 93% sensitivity and 90% specificity. However, identification of other related proteins may have been precluded due to a methodological deficiency in both studies. Gozal et al used 2D-DIGE-MS and found 16 differentially expressed proteins in patients with OSA. Seven of these were validated by either Western blot or enzyme-linked immunosorbent assay (ELISA).<sup>32</sup> In addition, sensitivity was 95% and specificity was 100% using a combination of urinary uromodulin, urocortin-3, orosomucoid-1, and kallikrein.<sup>32</sup> Becker et al developed a rigorous and reproducible proteomics workflow method using LC-MS/MS to explore deeper in the proteome and to detect proteins of lower abundance. As a result, more than 30 candidate biomarkers were identified in children with OSA.33 Two other studies used MALDI-TOF-MS and CE-MS analyses to reveal differentially expressed serum and urine proteins in adult patients with OSA and control subjects, respectively. Jurado-Gamez et al reported differential expression of 103 proteins between patients with OSA and controls. The differentially expressed proteins were associated with derangements in lipid and vascular metabolic pathways.<sup>34</sup> See tho et al profiled the expression of urinary peptides in OSAobese and non-OSA obese subjects. As a result, 24 candidate polypeptides were identified, of which 8 were derived from collagen and fibrinogen alpha; however, no significant differences were obtained.<sup>35</sup> Zheng et al identified 5 upregulated mass peaks and 6 downregulated mass peaks in patients with OSA and CVD and verified that the salivary alpha-2-HSglycoprotein (AHSG) level was lower in patients with OSA and CVD compared with that in patients with OSA without CVD.36

## Metabolite Profiles in Patients With OSA

Only 1 pilot metabolomics study explored and assessed potential plasma biomarkers in patients with nonsevere and severe OSA using LC-QTOF-MS. Fourteen statistically significant features were identified and highlighted.<sup>29</sup> The biochemical categories of these metabolites included phosphatidylcholine (PC), lysophosphatidylcholines (LPE), phosphoethanolamine (PE), lysophosphatidylethanolamine (LPA), phosphoserine (PS), lysophosphatidic acids, fatty acids, eicosanoids, amino acid metabolites and derivatives, and peptides.<sup>29</sup> Besides these metabolites were detected in metabolomics studies, metabolic biomarkers in patients with OSA detected by MS or chromatography alone should also be addressed.

References	Analytical Technique	Sample	Number of Participants	Metabolites		
Ferrarini et al <sup>29</sup> LC-QTOF-MS P		Plasma	18 Severe OSA and 15 non-severe OSA	PC, PS, LPC, LPE, LPA, PE methyl- hydroperoxyoctadecatrienoate, PGF2-alpha diethyl amide, ninecolic acid Arg Phe His		
Kawai et al <sup>44</sup>	LC-MS/MS	Saliva	20 Male OSA	PC		
Engeli et al <sup>45</sup>	LC-MS	Plasma	29 OSA, 26 OSA with T2DM, and 21 obese non-OSA	AEA, AG, OEA, AA		
Ezzedini et al <sup>46</sup>	GLC	Tonsillar tissue	114 Pediatric OSA and 92 recurrent tonsillitis	Palmitoleic acid, oleic acid, stearic acid		
Papandreou <sup>47</sup>	GC	Adipose tissue	63 OSA	Myristic, palmitic, stearic, oleic acid,		
Fletcher et al <sup>48</sup>	LC	Urine	8 Severe OSA and 5 obese hypertensive non-OSA	E, NE, MN, NMN		
Findley et al49	HPLC	Plasma	12 OSA and 6 controls	E		
Paci et al <sup>50</sup>	HPLC	Plasma	10 Male OSA and 11 age-matched controls	NE, E, DA		
O'Driscoll et al <sup>51</sup>	HPLC	Urine	70 Pediatric snorers and 26 controls	E, NE, DA		
Paik et al <sup>52</sup>	GC/MS	Urine	49 OSA (23 nonsleepy, 26 sleepy)	HVA, DOPAC		
Gislason et al <sup>53</sup>	Reverse-phase HPLC	Cerebrospinal fluid	15 OSA and 18 controls	5-HIAA, HVA, MHPG		
Grunstein et al54	HPLC and LC	Plasma and urine	8 OSA	NE, E		
Minemura et al <sup>55</sup>	HPLC	Urine and plasma	26 Male OSA	NE, E		
Monneret et al <sup>56</sup>	LC-MS/MS	Urine	26 OSA with MetS; 9 MetS	Hcy, 15-F2t-isoprostane		
Jordan et al <sup>57</sup>	HPLC	Plasma	16 OSA	Hey		
Ozkan et al <sup>58</sup>	HPLC	Plasma	34 OSA and 15 controls	ADMA, Hey		
Svatikova et al <sup>59</sup>	LC-MS/MS	Plasma	10 Male OSA and 10 controls	Nitrotyrosine		
Monneret et al <sup>60</sup>	LC-ESI-MS/MS	Urine	31 Nonobese OSA and 10 lean non- OSA	15-F2t-isoprostane		
Dikmenoğlu et al <sup>61</sup>	HPLC	Plasma	11 OSA and 11 matched controls	MDA		
Montgomery-Downs et al <sup>62</sup>	GC/MS	Urine	14 pediatric OSA and 33 non-OSA	15-F2t-isoprostane		
Stanke-Labesque et al <sup>63</sup>	LC-MS	Urine	40 nonobese OSA and 20 matched controls	U-LTE (4)		
Day et al <sup>64</sup>	HPLC and GC/MS	Plasma	9 OSA and 9 matched controls	Retinoids, carotenoids, and		
Kobayashi et al <sup>65</sup>	GC	Venous blood	35 OSA and 17 matched controls	CO		

TABLE 2.	Metabolites	Identified i	in	Patients	With	Obstructive	Sleep	Apnea	(OSA)
									(

5-HIAA = 5-hydroxyindoleacetic acid; AA = arachidonic acid; ADMA = asymmetric dimethylarginine; AEA = anandamide; AG = arachidonoylglycerols; CO = carbon monoxide; DA = dopamine; DOPAC = 3,4-dihydroxyphenylacetic acid; E = epinephrine; GC = gas chromatography; GC/MS = gas chromatography-mass spectrometry; GLC = gas-liquid chromatography; Hcy = homocysteine; HPLC = high-persource or high-performance liquid chromatography; HVA = homovanillic acid; LC = liquid chromatography; LC-ESI-MS/MS = liquid chromatography coupled to electrospray source ionization-tandem mass spectrometry; U-LTE(4) = leukotriene E(4); LC-MS/MS = liquid chromatography-tandem mass spectrometry; LC-QTOF-MS = liquid chromatography-bybrid quadrupole time-of-flight mass spectrometry; LPC = lysophosphatilylcholines; LPE = lysophosphatidylethanolamine; MDA = malondialdehyde; MHPG = 3-methoxy-4-hydroxyphenyl glycol; MN = metanephrine; NE = norepinephrine; NMN = normetanephrine; OEA = oleoyl ethanolamide; OSA = obstructive sleep apnea; PA = lysophosphatidic acids; PC = phosphatidylcholine; PE = phosphoethanolamine; PS = phosphoserine; T2DM = type 2 diabetes.

## Lipid Metabolites

Salivary PC concentrations increased in patients with OSA when detected by the LC-MS/MS method, and lower levels of PC in patients with OSA may decrease upper airway patency by reducing surface tension.<sup>44</sup> Circulating anandamide (AEA), 1/ 2-arachidonoylglycerols (AG), and oleoyl ethanolamide (OEA) levels were higher in plasma of patients with OSA compared with controls, whereas arachidonic acid concentrations were similar between the 2 groups, suggesting the important role of the endocannabinoid system in blood pressure regulation in patients with OSA at high risk for hypertension and CVD.45 Ezzedini et al<sup>46</sup> used GC-LC and found that palmitoleic and oleic acid levels were lower, whereas the stearic acid level was higher, in tonsillitis tissue than in hyperplastic tonsillar tissue, suggesting that fatty acid composition is altered in tonsillar tissue of children with OSA. In addition, Papandreou et al<sup>38,47</sup> reported that saturated fatty acids (ie, myristic, palmitic, stearic, and oleic acids) and n-6 fatty acids were correlated with sleep parameters in adipose tissue of patients with OSA using GC.

AEA), 1/ e (OEA) Fletcher et al<sup>48</sup> found that norepinephrine (NE) and normetanephrine levels were significantly higher in 8 patients with

metanephrine levels were significantly higher in 8 patients with OSA compared with those in 5 obese hypertensive controls. In another study, 12 patients with OSA had higher plasma epinephrine (E) levels than those of 6 normal volunteers.<sup>49</sup> Paci et al examined E, NE, and dopamine (DA) levels in 10 male patients with OSA and 11 age-matched controls. The results showed that plasma NE levels in the patients with OSA were higher than those in controls.<sup>50</sup> O'Driscoll et al<sup>51</sup> used HPLC to reveal that all urinary catecholamines increase significantly in children with OSA, and that levels of full-night NE and E are related to the severity of OSA. Paik et al used GC-MS to detect urinary neurotransmitter metabolites and showed that the dopamine metabolites homovanillic acid (HVA) and

After controlling for confounding factors, the results revealed

an independent association between n-3 fatty acids and sleep quality in patients with OSA.<sup>38,47</sup>

3,4-dihydroxyphenylacetic acid (DOPAC) increased in sleepy patients with OSA, indicating that excessive daytime sleepiness (EDS) in patients with OSA is probably caused by overnight upregulation of the dopaminergic system.<sup>52</sup> These results indicate that increased sympathetic tone occurs in patients with OSA.

However, findings of several other studies are inconsistent with these aforementioned studies. E and metanephrine (MN) levels were not different between 8 patients with OSA and 5 obese hypertensive controls.48 Paci et al50 reported that E and DA levels were in the normal range in both patients with OSA and control subjects. Gislason et al<sup>53</sup> detected 5-hydroxyindoleacetic acid (5-HIAA), HVA, and 3-methoxy-4-hydroxyphenyl glycol (MHPG) in the cerebrospinal fluid of 15 patients with OSA and 18 controls; however, levels of all of the aforementioned biomarkers were similar in patients with OSA and control subjects. Two studies evaluated the effects of continuous positive airway pressure (CPAP) treatment on urinary E and NE levels, which were confirmed in plasma. One of the studies found that withdrawing CPAP treatment led to no change in NE level.54 The other study detected no correlation among PSG parameters, but CPAP treatment reduced E levels 55

The inconsistency of catecholamine metabolites in patients with OSA may be due in part to use of different analytical platforms, various biological samples, small sample size, and different sample collection protocols. In addition, different study populations and unadjusted confounding factors would also result in inconsistencies.

## **Amino Acid Metabolites**

Serum homocysteine (Hcy) levels in patients with OSA are significantly higher than those in controls,<sup>56</sup> and CPAP effectively decreases Hcy levels in patients with OSA by almost 30%.<sup>57</sup> Another study showed that plasma Hcy level is higher in patients with OSA, while asymmetric dimethylarginine (ADMA) not.<sup>58</sup> OSA is not accompanied by increased circulating free nitrotyrosine either before or after sleep.<sup>59</sup> This result suggests that chronic nocturnal IH does not increase systemic nitric oxide-mediated oxidative/nitrosative burden in patients with OSA.<sup>59</sup>

## **Oxidative Stress Metabolites**

Urinary 15-F2t-isoprostane, one of the most sensitive and specific metabolites of lipid peroxidation, is positively correlated with carotid intima-media thickness and IH in nonobese patients with OSA,<sup>60</sup> and both 15-F2t-isoprostane and the apnea-hypopnea index are independently associated with Hcy levels in patients with OSA and metabolic syndrome.<sup>56</sup> Malondialdehyde (MDA) is another important oxidative stress biomarker that is significantly higher in patients with OSA than that in controls.<sup>61</sup> However, 15-F2t-isoprostane is not a significant feature in children with OSA.<sup>62</sup>

## **Other Metabolites**

Leukotriene E4 (U-LTE4) is a marker of proinflammatory cysteinyl leukotriene production, and increased urinary U-LTE4 has been demonstrated in patients with OSA.<sup>63</sup> CPAP treatment reduces U-LTE4 concentration by 22% only in patients with OSA and a normal body mass index (BMI).<sup>63</sup> Patients with mild OSA exhibit altered levels of specific retinoids, carotenoids, and tocopherols, which may be markers and/or mediators of increased susceptibility to vascular

disease.<sup>64</sup> Circulating carbon monoxide levels are enable assessment of the clinical severity of OSA.<sup>65</sup>

## DISCUSSION

This systemic review pooled available studies that applied chromatography and/or MS-based methods to detect biomarkers in patients with OSA. Numerous proteins and metabolites, including lipid profiles, adrenergic/dopaminergic biomarkers and their derivatives, amino acids, and oxidative stress biomarkers in patients with OSA have been identified. However, the results were not always consistent and far from clinical application. These differences may have been caused by: the source, storage, handling, and preparation of the biological sample; interindividual differences due to the highly transient and sensitive nature of metabolic fluxes; the close interaction between the micro and macroenvironments, including diet and microflora; and lack of uniformity in the technology and data analysis across different analytical platforms. Further studies with larger populations grouped by different phenotypes for subclass analyses are necessary to validate these results. Additionally, there is lack of a clear understanding of the mechanisms of these biomarkers. More organized investigations in cell and animal models are necessary to determine the molecular mechanisms.

Many novel small metabolites and proteins are identified and quantified in biological samples from patients with specific human diseases, including OSA, with development of the bioanalytical approach. The most-used samples include bio-fluids, such as plasma, serum, saliva, and urine, as well as tissues. Two major high-throughput analytical spectroscopic approaches (MS and proton nuclear magnetic resonance spectroscopy [<sup>1</sup>NMR]) are commonly used and have become core technologies for metabolomics and proteomics to determine protein metabolic profiles. <sup>1</sup>NMR spectroscopy-based methods provide high resolution but low sensitivity. To date, no study has used this analytical platform to detect biomarkers in biological samples from patients with OSA. MS-based methods have higher sensitivity and can simultaneously analyze a broader range of molecules.<sup>66,67</sup> MS-based analyses focus on detecting ionized molecules and measuring their mass-to-charge ratio. The combination of MS and chromatography (GC or LC) is commonly used for metabolomics and proteomics. GC requires volatile and thermally stable analytics, and nonvolatile compounds must be derivatized before a GC-MS analysis.<sup>69</sup> Public libraries have enabled reliable identification of compounds from GC-MS data. LC-MS-based methods are more universal techniques and enable separation of different types of compounds with the appropriate column and solvents.<sup>69,70</sup> Thus, each platform has its own strengths and limitations in terms of assessing samples and identifying compounds. A combination of different analytical platforms would be an ideal approach to OSA biomarker research.

The characteristics of OSA include chronic IH and sleep fragmentation, both of which can activate the sympathetic nervous system, followed by oxidative stress/systemic inflammation. Levels of various biomarkers fluctuate during this progression. Proteic biomarkers: in pediatric OSA, gelsolin, perlecan, albumin, immunoglobulin, uromodulin, urocortin-3, orosomucoid-1 and kallikrein alter in urine.<sup>30,32</sup> In rats exposed to IH, reduced kallikrein may play an important role in preservation of renal function.<sup>71</sup> Becker et al map identified urinary proteins to specific functional pathways (ie, acute-phase proteins, angiogenesis, hemostasis, leukocyte immunity, and

lipid binding).<sup>33</sup> In adult OSA, the identified proteins in serum are primarily associated with derangements in lipid and vascular metabolic pathways.<sup>34</sup> In obese OSA, some of the identified peptides in urine comprise collagen alpha chain subtypes and fibrinogen; these peptides seem to be associated with increased metabolic syndrome.<sup>35</sup> In OSA with CVD, the salivary AHSG decreases in CVD group.<sup>36</sup> AHSG plays an important role protecting against vascular calcification, as vascular calcification is a strong predictor of CVD.<sup>72,73</sup> Thus, we speculate that proteic biomarkers in pediatric OSA may be associated with both kidney dysfunction (ie, altered glomerulotubular permeability) and disease-related pathological processes, while the proteic biomarkers in adult OSA seem to be associated with cardiovascular and metabolic alterations. Two proteomic studies use immunoblotting/ELISA or Western blot method to further verified the identified peptides, the results are consistent with proteomic approaches.<sup>32,36</sup> Only 1 proteomic study explores differential serum protein expression and OSA severity in adults.<sup>34</sup> The underexpressed complement component 4binding alpha and thrombospondin and overexpressed fibronectin, apolipoprotein B (ApoB)-100 and apolipoprotein D alter with OSA severity.<sup>34</sup> Lipidic biomarkers: Compared with recurrent tonsillitis, hyperplastic tonsillar tissue in pediatric OSA contains more proportions of monounsaturated fatty acids and fewer proportions of saturated fatty acids.<sup>46</sup> In adult OSA, significant positive correlations are found between sleep parameters and lipid (ie, myristic, palmitic, stearic, saturated fatty, oleic, and polyunsaturated fatty acids) metabolism in buttock subcutaneous tissue.<sup>38</sup> Plasma PC, LPE, PE, LPA, PS, AEA, 1/2-AG, and OEA are higher in adult OSA.<sup>29,45</sup> Salivary PC is also higher in patients with OSA.<sup>44</sup> Altered state of lipid metabolism in tonsillar tissue in pediatric OSA may be linked to glucose metabolism and membrane permeability.46 Thus the lipid metabolism disorder might be correlated with pediatric OSA occurrence. In adult OSA, the lipid metabolism disorder is simply a sign of metabolic disorders in general condition. Catecholamine metabolites and derivatives: In pediatric OSA, increased sympathetic tone can increasingly release of catecholamines (eg, E, NE, and DA).<sup>51</sup> And, the E. NE, and DA are all related to the severity of OSA in children. In earlier studies, catecholamines and dopamine metabolites are similar between adult OSA and controls. That may partly be explained by small enrolled subjects and strict case selection. In later studies, similar to results in children, E, NE, and DA in adult OSA are all higher than those in controls. Thus, catecholamines seem to be ideal biomarkers reflecting sympathetic overactivity.<sup>74</sup> *Amino acid metabolites*: Hcy level increased in adult OSA,<sup>56,58</sup> and CPAP treatment could effectively lower serum Hcy.<sup>57</sup> However, no significant change of nitrotyrosine and ADMA was witnessed in OSA. These might indicate that reactive oxygen species, not reactive nitrogen species is the most important free radicals in adult OSA. Recent meta-analyses also reveal the similar results of Hcy level in OSA and effect of CPAP therapy despite various methods employed in included studies.<sup>75,76</sup> Significantly increased Hcy level is observed in the severe OSA group, thus the Hcy could be recognized as a representative biomarker in severe OSA. Markers of oxidative stress: In pediatric OSA, Montgomery-Downs et al find the 15-F2t-isoprostane is not elevated in children with OSA.<sup>62</sup> The authors ascribe this phenomenon to intrinsic antioxidant capacity and small sample size in this study. However, the metabolic intermediates of enhanced oxidative stress (eg, 15-F2t-isoprostane and MDA) are elevated in adult OSA.<sup>56,60,61</sup> The 15-F2t-isoprostane elevates with OSA

severity in adults and could add the notion that oxidative stress may contribute to OSA-related cardiovascular complications.<sup>60</sup> The MDA, which measures lipid peroxidation, is also verified to be elevated in OSA by numerous methods.<sup>77</sup> For quantifying isoprostane, the ELISA method has often been used. However, due to cross-reactivity interference, it gives only an approximate estimation.<sup>60</sup> Thus, the ELISA overestimates by twofold of the isoprostane level when compared with GC/MS.<sup>78</sup>

Interestingly, gut microbiota participate in regulating many metabolic pathways, and the comprehensive interaction between the host and microbiota and metabolism has been hotly debated.<sup>79,80</sup> One study found that IH alters the composition and diversity of fecal microbiota in a murine model.<sup>\$1</sup> Levels of Prevotella, Paraprevotella, Desulfovibrio, and Lachnospiraceae increase, whereas those of Bacteroides, Odoribacter, Turicibacter, Peptococcaceae, and Erysipelotrichaceae decrease.<sup>81</sup> Evidence from another murine model initially demonstrated that sleep fragmentation also has a significant impact on microbial community structure in mice.<sup>82</sup> <sup>2</sup> Lipopolysaccharide-binding protein (LBP) is a biomarker of potential low-cascade endotoxemia, formed in response to microbial translocation; LBP is elevated in children with OSA, including those who are not obese.83 All of these studies directly or indirectly reported that OSA influenced the gut microbiome composition. Interestingly, several biomarkers discussed in this systemic review associated with an altered gut microbiome are noteworthy. An aromatic amino acid (ie, tryptophan and tyrosine) metabolic disorder is always persistent in intestinal dysbacteriosis, so the metabolites of tryptophan (5-HIAA and HVA) and tyrosine (DA and NE) are altered due to the imbalance in the intestinal flora. Endocannabinoid system activity is involved in the control of glucose and energy metabolism, and endocannabinoids (ie, AEA, 1/2-AG, and OEA) can be up- or down-regulated by specific gut microbes, such as *Akkermansia muciniphila*.<sup>84,85</sup> Thus, we infer that alterations in particular gut microflora species in patients with OSA affect downstream metabolites. Alterations in these metabolites could cause low-grade systemic inflammation, resulting in insulin resistance and increased CVD risk. Integrating metabolomics and metagenomics could facilitate development of potential preventive and therapeutic interventions for OSA-related complications.

We must recognize some inevitable limitations in our systemic review. Eligible studies that were not indexed or published, and so were not included, may have resulted in publication bias. The marked heterogeneity in the included studies was due to relatively small sample sizes, use of different techniques, and different biological samples. Thus, a metaanalysis of the results of these studies was not feasible. Additionally, not all of the studies controlled for confounding factors, such as age and BMI. Last, few metabolomic studies have been performed to identify diagnostic biomarkers in adult patients with OSA, although some studies speculated that a particular protein might be a predictor of pediatric OSA. Additionally, the results of these proteomics studies are preliminary and should be validated in larger independent cohorts. Despite these limitations, advances in quantitative proteomics and metabolomics will enable more in-depth analyses of cellular systems. There are a number of advantages of quantitative proteins and metabolites compared with traditional chemometric approaches, including improved statistical robustness, simpler biological interpretation, and detailed information on specific protein and metabolite identities; hence, they show great potential for clinical application.

In summary, many novel OSA-related biomarkers have been identified by chromatography and/or MS-based methods. In addition, applying the proteomics and metabolomics approach helps us better understand OSA pathophysiology, and will enable early detection and development of personalized treatment strategies. However, well-matched case–control proteomic and metabolomic studies in community-based populations are warranted to confirm these findings. Then, proteins and metabolites identified using these translational and personalized approaches will be considered biomarkers for OSA screening.

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