

Review Article

# A central role for amyloid fibrin microclots in long COVID/PASC: origins and therapeutic implications

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Post-acute sequelae of COVID (PASC), usually referred to as ‘Long COVID’ (a phenotype of COVID-19), is a relatively frequent consequence of SARS-CoV-2 infection, in which symptoms such as breathlessness, fatigue, ‘brain fog’, tissue damage, inflammation, and coagulopathies (dysfunctions of the blood coagulation system) persist long after the initial infection. It bears similarities to other post-viral syndromes, and to myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS). Many regulatory health bodies still do not recognize this syndrome as a separate disease entity, and refer to it under the broad terminology of ‘COVID’, although its demographics are quite different from those of acute COVID-19. A few years ago, we discovered that fibrinogen in blood can clot into an anomalous ‘amyloid’ form of fibrin that (like other  $\beta$ -rich amyloids and prions) is relatively resistant to proteolysis (fibrinolysis). The result, as is strongly manifested in platelet-poor plasma (PPP) of individuals with Long COVID, is extensive fibrin amyloid microclots that can persist, can entrap other proteins, and that may lead to the production of various autoantibodies. These microclots are more-or-less easily measured in PPP with the stain thioflavin T and a simple fluorescence microscope. Although the symptoms of Long COVID are multifarious, we here argue that the ability of these fibrin amyloid microclots (fibrinaloids) to block up capillaries, and thus to limit the passage of red blood cells and hence  $O_2$  exchange, can actually underpin the majority of these symptoms. Consistent with this, in a preliminary report, it has been shown that suitable and closely monitored ‘triple’ anticoagulant therapy that leads to the removal of the microclots also removes the other symptoms. Fibrin amyloid microclots represent a novel and potentially important target for both the understanding and treatment of Long COVID and related disorders.

## Introduction

In many cases, individuals infected with the SARS-CoV-2 virus and suffering from COVID-19 continue (or in some case begin) to display symptoms long after the acute phase. Depending on the demographics of the hosts and variant of SARS-CoV-2, these post-acute sequelae of COVID-19 (PASC [1]) may affect ~30% of all infected individuals [2,3], are starting to be observed even in children [4,5], and are commonly known as long COVID [6–10]. Many regulatory health bodies still do not recognize this syndrome as a separate disease entity, and refers to it under the broad terminology of ‘COVID’. The symptoms of long COVID are multifarious [11,12] and include breathlessness, fatigue, chest pain, myalgia, cognitive dysfunction, innate immune responses coupled to inflammatory cytokine production, and a pro-coagulant state. Our focus is on the latter.

At present there is no established treatment for long COVID (e.g. [13,14]), so from a systems point of view it is important to understand which symptoms are ‘primary’, and which are simply secondary effects of the primary symptoms themselves. This would allow treatment strategies to focus on the primary symptoms and their causes. We here make the case (with evidence) that much of the

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aetiology of long COVID can be attributed to the formation of aberrant amyloid fibrin microclots, triggered in particular by the SARS-Cov-2 spike protein, and that by inhibiting the transport of erythrocytes to capillaries, and hence O<sub>2</sub> transfer, it is these amyloid microclots that are mainly responsible for the various long COVID symptoms observed. The microclots may also present novel antigens that lead to the production of autoantibodies, that can exacerbate symptoms further. This understanding of the role of such microclots may be expected to lead to an effective strategy for treating long COVID (and probably for other, related conditions such as myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS)).

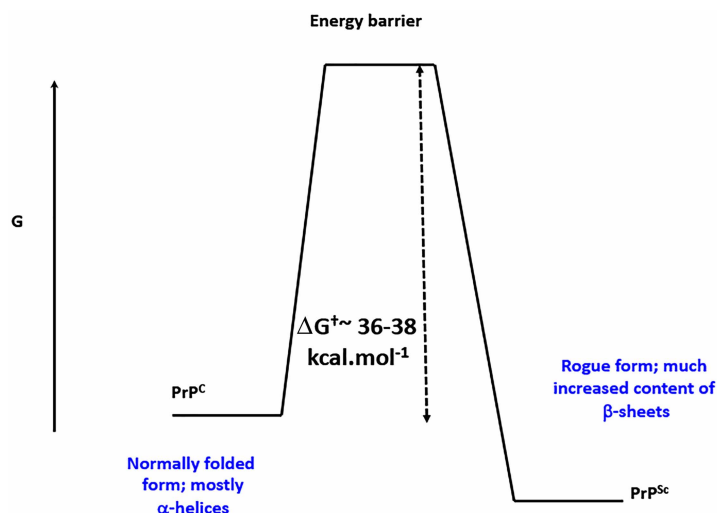
## Protein conformation, prions, and amyloid structures

Classically, it was assumed that proteins folded into their conformational states of lowest free energy, because unfolded proteins typically refolded into their original forms [15]. However, the discovery of prion proteins in particular showed that this was not always the case; the more common and native form of the prion protein PrP<sup>C</sup> could be converted — with no change in primary sequence — into a thermodynamically more stable, ‘rogue’ version referred to as PrP<sup>Sc</sup> (Figure 1), and this transformation was itself catalysed by PrP<sup>Sc</sup> (e.g. [16,17]). These ‘rogue’ versions commonly contain a large number of ordered β-sheets in a cross-β architecture [17–23], and are referred to generally as amyloid forms. A great many proteins are now known to be able to adopt such amyloid forms, and over 50 have been implicated in a variety of diseases (‘amyloidoses’) [24–26]. These kinds of amyloid structures may be stained with fluorogenic dyes [27,28] such as congo red [29], thioflavin T [30–35], which is much more visible than congo red, or a variety of conjugated oligothiophenes [36] commercialized as Amytracker™ dyes (e.g. [37,38]).

## Amyloidogenic blood clotting

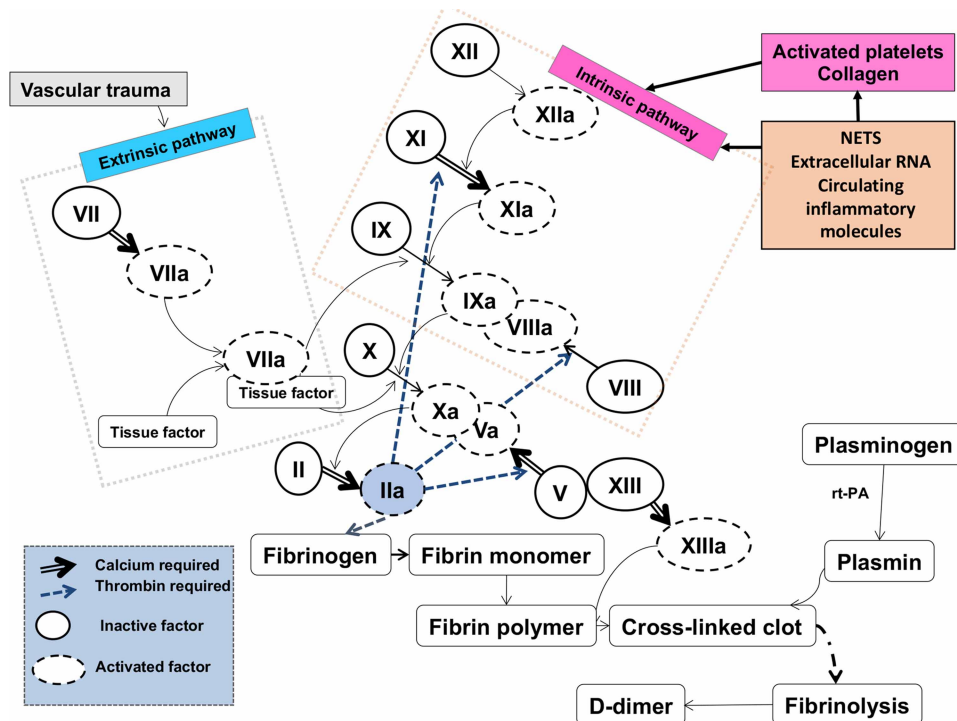
The normal blood clotting cascade is well established (e.g. [39–41]) (Figure 2), with the terminal stages of both ‘intrinsic’ and ‘extrinsic’ pathways involving the self-organised polymerization of soluble fibrinogen to insoluble fibrin, catalysed by thrombin [42]. Fibrinogen, a cigar-shaped molecule of some 5 × 45 nm, is one of the most abundant proteins in plasma (commonly present at 1.5–3.5 g L<sup>-1</sup> [41]).

The action of thrombin on fibrinogen leads to the removal of two small fibrinopeptides (Figure 3), and this initiates the thermodynamically favorable conversion to fibrin, first as small protofibrils and then as long fibrils, typically with a diameter of 50–100 nm. Factor XIII is a transglutaminase which inserts cross-links into the



**Figure 1. Many proteins can adopt more than one thermodynamically stable microstate with no change in primary structure (sequence), in which the more stable contains an ordered β-sheet ‘amyloid’ structure.**

Normally, however, it is present in a less stable state that is kinetically more accessible during and following its synthesis. The more stable (labeled PrP<sup>Sc</sup>) is separated from the initial state (PrP<sup>C</sup>) via a large energy barrier. This is true for amyloid proteins generally, and is illustrated here for classical prion proteins. Redrawn from a CC-BY publication at [26].



**Figure 2. Representation of the classical blood-clotting cascades, ending in the removal of two fibrinopeptides from fibrinogen, and its self organization to produce fibrin (that may then be cross-linked).**

The eventual result of fibrinolysis, that acts as a record of its extent, is D-dimer. Redrawn from a CC-BY publication at [26].

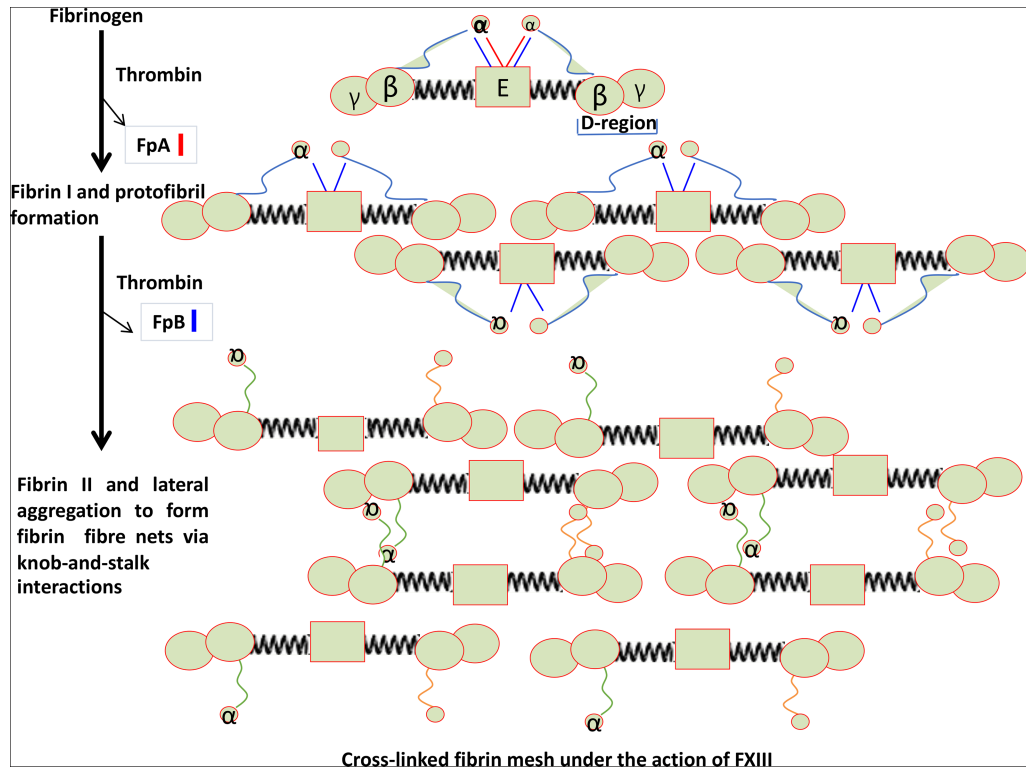
developing clot, and also serves to cross-link  $\alpha 2$ -antiplasmin ( $\alpha 2$ -AP) to fibrin.  $\alpha 2$ -AP and other similar molecules are major inhibitors of plasmin, the enzyme which is mainly responsible for clot proteolysis (fibrinolysis).

The structure of fibrin clots is typically characterized by the diameter of the spaghetti-like fibres and by the size of the residual pores [43]. However, in some cases these spaghetti-like structures with pores are absent. We initially characterized this phenomenon, observed in the electron microscope, as ‘dense matted deposits’ (e.g. [44–46]). We later discovered that such structures could be induced by highly substoichiometric amounts of bacterial lipopolysaccharide (LPS) (1 molecule LPS per 100,000,000 fibrinogen molecules), and it emerged that they too were, in fact, amyloid in character [26,47–50]. This highly substoichiometric induction (of a thermodynamically favourable process) is very important, as it effectively provides a means for a massive amplification of what may be a tiny amount of trigger material.

The list of molecules that could induce this anomalous clotting, additional to the bacterial LPS that was then our main focus, included iron ions [51–56], oestrogens [46,57], lipoteichoic acid [49,52], and serum amyloid A [58].

While the list of the many other molecules that can plausibly effect this anomalous amyloid-type clotting is unclear, such clots may also be observed in the blood of individuals with inflammatory diseases such as Alzheimer’s [37,50,59–61], Parkinson’s [37,48], type 2 diabetes [37,38,62–64], and rheumatoid arthritis [65–68]. Similar phenomena have also been observed in the pregnancy disorder pre-eclampsia [69], where there is also strong evidence for a microbial component [70,71].

Although this was a novel finding for ‘normal’ blood, it had previously been shown that  $\beta$ -sheet structures could be induced in fibrin artificially by mechanical means [72]. There have also been rare reports (see [26,73,74]) of amyloidogenic mutations (alleles) in the fibrinogen  $\alpha$  chain. Additionally, it has long been known that aged fibrin deposits can bind amyloid dyes in tissue sections [75] and that fibrin-derived peptides can form cross-beta structures [76]. The amyloid form of the prion protein is highly resistant to proteolysis (resistance to proteolysis by proteinase K is used in an assay for PrP<sup>Sc</sup> [77–79]), and so this provided a ready explanation for the different nature and persistence of the amyloid fibrin clots [52].



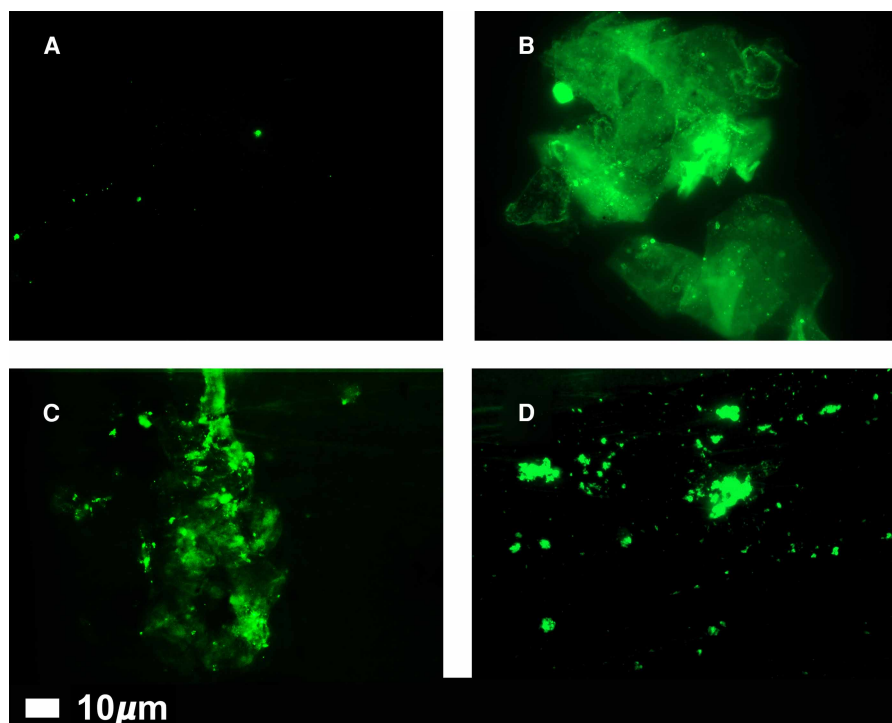
**Figure 3.** Self organization of fibrinogen into fibrin under the action of thrombin in removing two fibrinopeptides. Redrawn from a CC-BY publication at [26].

### A note on ‘amyloid’ terminology

We recognize that there has occasionally been confusion in our use of the term ‘amyloid’ to describe amyloidogenic blood clotting, as blood is not seen as a source of the classical amyloidoses. While we could have used another term, the structures and staining of these blood clot fibres, presumably consisting of the ordered  $\beta$ -sheet architectures necessary (i) to bind fluorogenic amyloid stains and (ii) to be resistant to the normal sources of proteolysis, do reflect the well-established term ‘amyloid’. In addition, there is a very simple discrimination between ‘classical’ amyloids and those fibrin-rich amyloids that are our focus here: this is that the amyloid fibrils seen in classical amyloidoses tend to be ca 5–25 nm or so in diameter [21,22,80,81] whereas those in fibrin amyloid microclots tend to be in the range 50–150 nm or even more. They are thus easily distinguished microscopically, even without the greater analytic power afforded by proteomics [82], antibody staining [83], and so on. However, to avoid such ambiguity in the future, we consider it sensible to refer to the kinds of fibrin-based amyloids we are speaking about as fibrinaloids.

## Amyloid fibrin microclots (fibrinaloids) in COVID-19 and long COVID

Coagulopathies [84–102], and especially the formation of extensive microclots *in vivo*, are a hallmark of both COVID [85,103–115] and long COVID [116,117], and we have demonstrated that these microclots too are amyloid in character [108,109,116]. Importantly, the addition of purified, recombinant SARS-CoV-2 S1 spike protein to coagulation-competent normal plasma is sufficient to induce the formation of anomalous clots [118] that adopt amyloid states that are also resistant to fibrinolysis [108]. Note that the observations of the microclots in (platelet-poor) plasma are performed without the addition of exogenous thrombin; they are naturally there in the circulation of patients with both acute and long COVID. The size of these amyloid microclots, that can be observed microscopically and stained e.g. with thioflavin T [108,109,116] (Figure 4; control vs LC plasma), are typically anywhere from 1–200  $\mu\text{m}$ ; this means that they can effectively block up, and inhibit



**Figure 4. Fluorescence microscopy of representative micrographs showing microclots in the circulation of controls (A) and in patients with Long COVID (B–D).**

Absence of significant amyloid microclots in the plasma of ‘normal’ individuals, and their significant presence in the plasma of individuals with long COVID. Platelet-poor plasma was produced by centrifugation at 3000×g for 15 min, stained with 5 μM thioflavin T, and imaged in a fluorescence microscope (Zeiss Axio Observer 7 with a Plan-Apochromat 63×/1.4 Oil DIC M27 objective (Carl Zeiss Microscopy, Munich, Germany). Wavelengths were Exc 450–488 nm/emission 499–529 nm, all as in [108].

blood flow through, all kinds of microcapillaries, thereby strongly lowering the availability of oxygen in tissues. As expected, they consist mainly of fibrin, but also contain many other proteins, including  $\alpha$ 2-antiplasmin [108] (and even the virus itself [119]). They also have heightened pro-inflammatory activity and elicit fibrin autoantibodies [118] (and maybe others). Elements of at least some spike protein variants of SARS-CoV-2 can also stimulate (*in silico*) the amyloidogenic aggregation of serum amyloid A [120]. More generally, there is considerable evidence for the induction of amyloid production by amyloidogenic proteins by viruses such as those of the Herpesviridae family, e.g. in Alzheimer’s disease [121–125].

### ‘Ground-glass’ opacities in COVID and long COVID

One of the early hallmarks of acute COVID-19 was the observation of extensive, diffuse opacities in high-resolution computed tomography (HRCT) scans of the lungs of patients, consistent with the breathing difficulties widely observed. These opacities resembled ground glass [126], and indeed are commonly referred to as ‘ground-glass opacities’ (GGO) [127–132]. They are largely formed of fibrin, though we are not aware that anyone has yet stained them for amyloid structures (which is what we believe they must be, and there is ample precedent for this in other diseases (e.g. [133–136])). In COVID patients they persist well beyond the acute phase [137].

### More on the size and properties of microclots

As mentioned, the fibrinoid microclots that we observe are typically in the range 1–200 μm on their longest axis. This is consistent with the ‘ground glass’ appearance. It is also true for ‘artificial’ amyloid-type protein structures [28], and for the kinds of amyloid seen as deposits in thrombotic microangiopathies [138–142]. However, the size distribution differs markedly between individuals (Figure 4), and while hard to pin down it



does provide a ready general explanation for the very different manifestations of Long COVID (including suggestions that the term covers ‘multiple’ diseases). This is also true for prion diseases, where specific ‘strains’ based on particular conformations can propagate in the same form. In some cases, there can be differential staining of individual amyloids by different dyes [26,49,143–145], which also allows a certain degree of differentiation of the structure of the clots. In the case of Long COVID, we are probably not quite ready for such subtleties.

## Role of D-dimer

A major marker for fibrinolytic activity (Figure 2) is a polypeptide referred to as D-dimer Figure 5. It is a strong prognostic indicator for disease outcome (survival) in acute COVID [85,146–154]. It is, in effect, a composite measure of how much fibrinogen there was, how much was converted to fibrin clots (whether normal or fibrinolytic), and how much these then went on to be lysed. Each of these general steps may of course be regulated independently (i.e. proceed at a ‘fast’ or slow’ rate, as encoded simplistically in Figure 6), so the analysis of D-dimer measurements must take all of these issues into account. Thus high levels of D-dimer must of necessity represent the production of many more clots (and sufficient fibrinolysis to be occurring) but cannot of

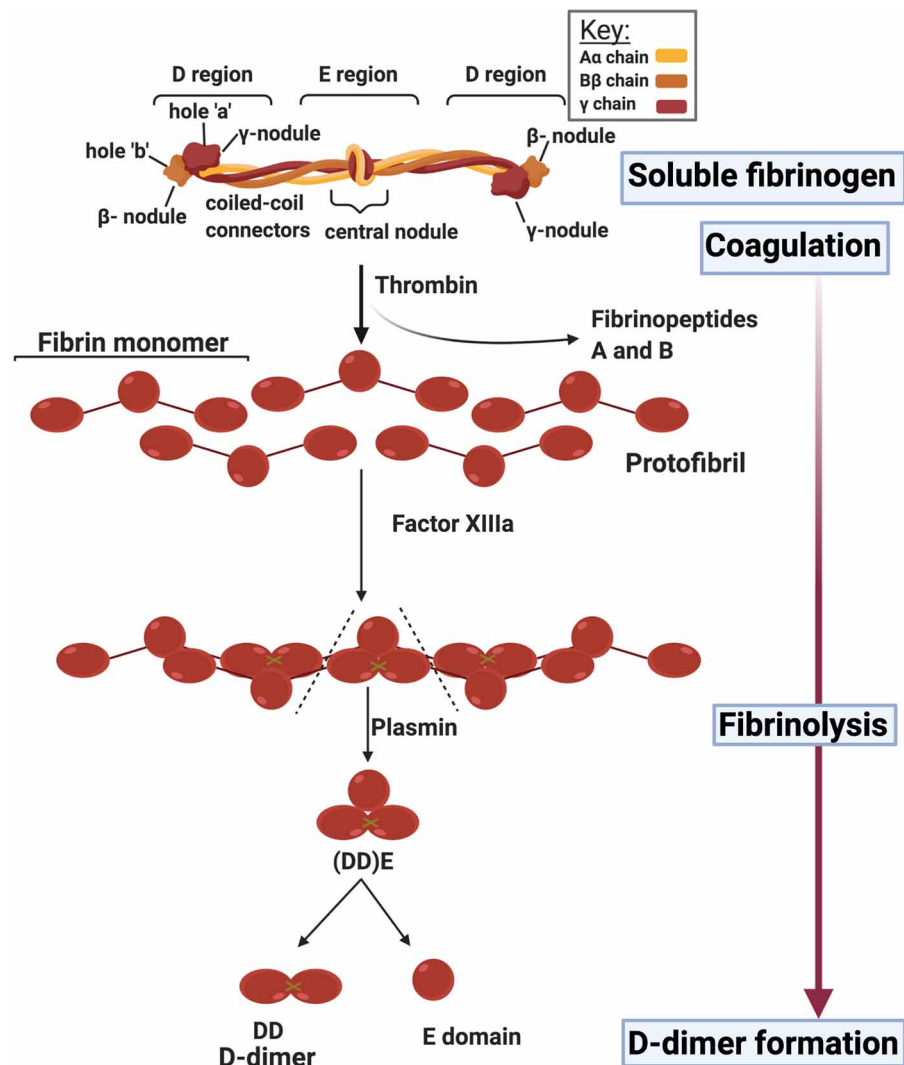
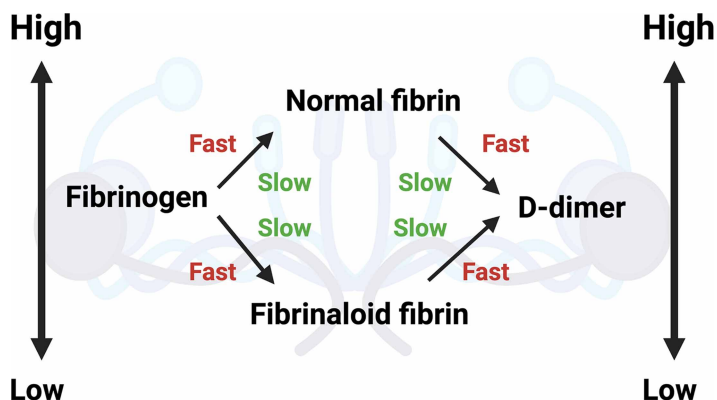


Figure 5. D-dimer production by fibrinolysis.

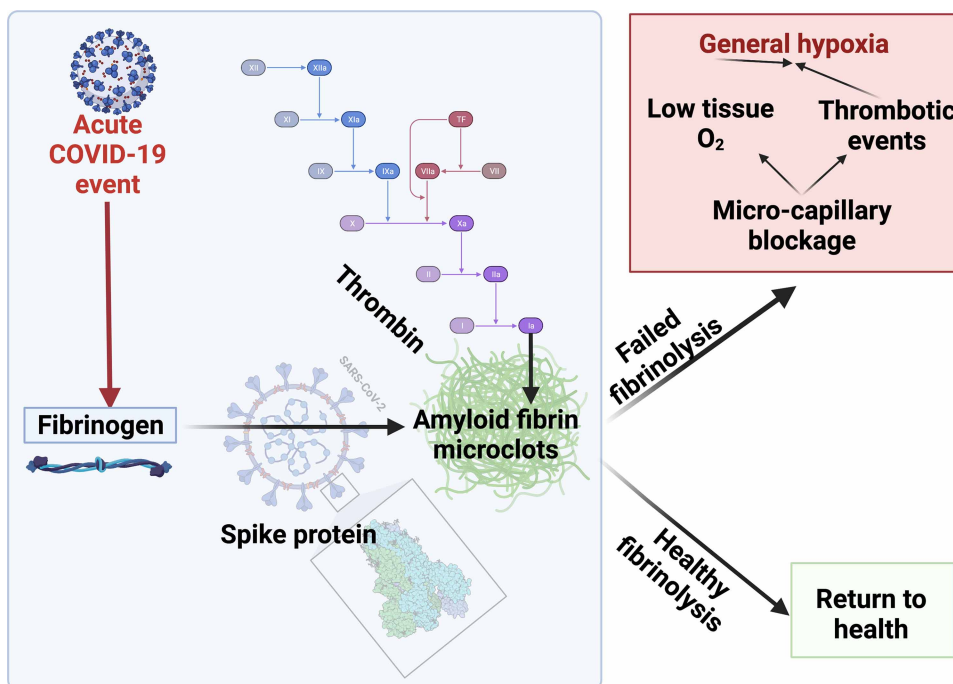
It is believed to be similar when produced from fibrinolytic clots, but the rate is considered to be slower. Image created with BioRender (<https://biorender.com/>).



**Figure 6.** D-dimer levels reflect both the rate of production and rate of degradation of clots, whether the clots are ‘normal’ or fibrinoid in nature.

Image created with BioRender (<https://biorender.com/>).

themselves reflect how many remained after their lysis nor which of them were ‘normal’ and which amyloid (see Figure 7 for a simplified pathway explanation). This explains why D-dimer levels are significantly raised following infection with many of the earlier variants of SARS-CoV-2, which produce multiple fibrinoid microclots and Long COVID. However, D-dimer is massively increased in the case of the omicron variant where much clotting takes place but seemingly not to a fibrinoid form, and thus effective clot fibrinolysis takes place. That there may be strong dependencies on the precise variant is not really surprising (omicron differs from both alpha and delta variants by more than 20 mutations [155], more than enough to vary a protein’s activity 1000-fold in typical directed evolution experiments [156]). High D-dimer thus may, but does not have



**Figure 7.** A simplified diagram to explain microclot formation that might either be resolved via fibrinolytic processes after acute COVID-19 or, in some patients, result in a failed fibrinolytic process.

Image created with BioRender (<https://biorender.com/>).

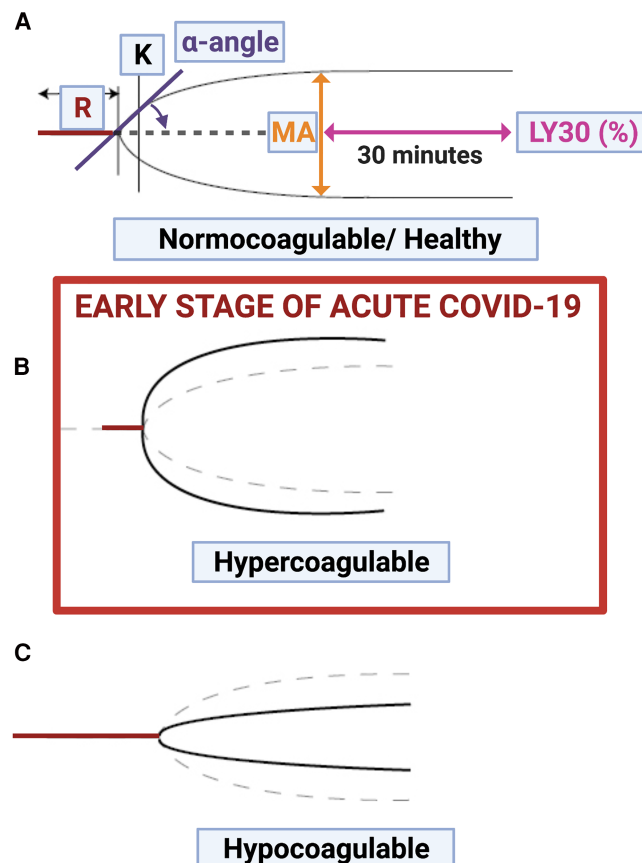
to [157], reflect disease severity given a particular SARS-CoV-2 variant; it depends on the context, and in particular the type of fibrin that can serve as the substrate for its production.

## Assessment of clotting

### Why TEG rather than APTT, INR and other clotting measures?

A variety of optical clotting tests are in more-or-less common usage [158]. These include the aPTT (activated partial thromboplastin clotting time), prothrombin time (PT) and International normalized ratio (INR), but they are considered to have limited predictive value for bleeding and are not cost-effective [159,160]. INR and APTT are of limited value to predict clotting (and identifying patients that will bleed) in patients with acute and Long COVID. The test uses platelet poor plasma after discarding the cellular component of blood (including the platelets).

As blood clots, the viscosity of the clot increases, and measuring the time-dependent rate and extent of this viscoelasticity change directly in whole blood provides a more comprehensive and convenient *in vitro* assessment of the clotting behavior of the blood at the time. Note that clots of fibrinoid that have already formed are effectively inert and invisible to such methods. Here the method of choice is known as Thromboelastography (TEG) [48,52,56,58,59,65,66,108,109,161–165] (or a related version known as ROTEM [166–173]). Figure 8 shows stylized traces of TEG, as typically found in health and disease. There are three main generalized traces: healthy (normocoagulable), hypercoagulable (as commonly seen in acute COVID-19) and hypocoagulable (during bleeding risk).



**Figure 8. TEG® traces with the main parameters visualized.**

(A) Healthy (normocoagulable) trace; (B) Hypercoagulable trace (seen during early stages of acute COVID-19) and (C) Hypocoagulable trace. Image created with BioRender (<https://biorender.com/>). In B and C the dotted lines represent the normocoagulable case. R represents the time taken to initiate clot formation,  $\alpha$  and K reflect the rate and extent of clot formation (e.g. [52]).



The TEG is a well-known point-of-care method, lately suggested as a good method for following blood clotting status in acute COVID-19, especially when anticoagulation regimes are prescribed to patients [109]. However, in our experience, it is not useful for determining (hyper)coagulation status in Long COVID, and should be reserved for those Long COVID patients where there is either a bleeding concern or actual bleeding. TEG should then be used in conjunction with platelet and microclot mapping, to follow patients closely when bleeding might be expected. Because microclots already contain clotted plasma proteins, and TEG only measures unclotted (soluble) plasma proteins, the TEG might not be suitable to determine the actual existing coagulation status (consisting of both clotted and soluble plasma proteins).

TEG trace ranges might in fact be within normal ranges, and our take on this is that the fibrin inside the microclots has already been coagulated, and trapped inside the microclots, leaving only a lower amount of soluble fibrinogen that might partake in the TEG assay. Therefore both the already-coagulated fibrin in the microclots and the soluble fibrinogen levels should in fact be considered, and preferably measured. Unfortunately the TEG cannot do that. However, we consider that TEG is vital for assessing the potential for coagulation when Long COVID patients are treated with anticoagulation therapy, or in any individual undergoing treatment designed to lower the levels of fibrinoid microclots. In our view, TEG is therefore a much more true evaluation of the patient's coagulation state, as it uses whole blood, thus bringing in the very important role played by platelets.

INR, APTT D-dimer, fibrinogen levels and platelet count may be of help in the very late event of end stage acute COVID-19, where acute disseminated intravascular coagulopathy (DIC)-coagulopathy is seen. Interestingly, it is now accepted that DIC-bleeding events are uncommon in acute COVID-19 [111]. Even in this event, TEG is still of more use to manage this situation [109].

## Mechanical properties of amyloid microclots

While we do not yet know the details of the mechanical properties of the fibrin amyloid microclots, it is known that amyloid fibrils can typically exhibit unusually high mechanical strength and resistance to deformation (e.g. [174–177]). The stiffness also increases with the thickness of the fibres [178]. This implies strongly that the fibrinoids are likely to be more prone to 'getting stuck' in capillaries.

## Bradford Hill criteria and microclots

The Bradford Hill criteria for causation of a disease Y by an environmental factor X [70,179] represent a useful framework for assessing the role of fibrin amyloid microclots in Long COVID, and are as follows: (1) strength of association between X and Y, (2) consistency of association between X and Y, (3) specificity of association between X and Y, (4) experiments verify the relationship between X and Y, (5) modification of X alters the occurrence of Y, and (6) biologically plausible cause and effect relationship. We think that the evidence is very strongly consistent with these criteria when X is represented by microclots and Y by Long COVID and potentially a variety of other conditions.

## Ability of amyloid microclots to explain the symptoms of long COVID

While long COVID is a multi-system disorder with multiple symptoms of varying severity, it remains possible that there is in fact a particular major underlying cause (or that a very small number are the main contributors). Our view is as follows: given that amyloid microclots can clog up capillaries and inhibit the transport of O<sub>2</sub> to tissues, this alone can in fact more or less self-evidently serve to explain a great many observations, and in particular the symptoms of both acute and long COVID. These obviously include breathlessness due to low O<sub>2</sub> directly, and thrombotic events such as acute myocardial infarction [180,181], stroke [181–183], etc due to the microclots. The lack of O<sub>2</sub> transport to tissues explains straightforwardly how a nominally respiratory disease also leads to the dysfunction of organs such as the kidney [184,185], PoTS (postural tachycardia syndrome [186]), myalgia in skeletal muscle [187,188], neurological disorders [189], and lactic acidosis [190,191] (the mass of lactate is too low to have been detected in our own COVID untargeted metabolomics experiments [192]), and potentially the benefits of hyperbaric O<sub>2</sub> therapy [193]. It is also worth stressing that amyloid structures themselves tend to be cytotoxic, often via membrane disruption [194–198].

## What makes long COVID long?

For any deterministic system to change its behavior there has to be a change of its parameters [199]. In the case of long COVID we need mechanisms that can explain how something that was initiated a long time ago can somehow persist. One source of the continuous production of a stimulus is represented by microbes, including virions, that persist in a largely dormant state (often in intracellular reservoirs) but can occasionally continue to replicate [1,200]. There is now considerable evidence for the persistence of SARS-CoV-2 [201]. Another is the continued release of sequestered microbially derived substances that can act as stimuli for continuing microclot formation. Here, the finding [202] that S1 spike protein can itself persist in CD16<sup>+</sup> Monocytes in PASC for up to 15 months post-infection is highly relevant, as the amplification of trigger proteins to make microclots as part of the clotting mechanism means that miniscule (and highly substoichiometric) amounts of suitable triggers can suffice [26,47]. This alone is sufficient to account for the chronic nature of such diseases.

## Microclot sequestration of biomarkers

An extra consequence of the production of microclots that sequester other proteins whose concentration would otherwise appear elevated is that those proteins do not then appear in plasma from which the microclots have been removed (e.g. by centrifugation), and thus they do not manifest, and cannot usefully be used, as biomarkers. Thus, our experience [108,116] is (i) that many such proteins including  $\alpha$ 2-AP and autoantibodies are so sequestered, and (ii) that the degradation of these protease-resistant microclots for classical proteomic analyses requires multiple rounds of trypsinization.

## The role of autoantibodies and biomimicry

Autoantibodies are a feature of many chronic, inflammatory diseases, such as rheumatoid arthritis [68] (where *Proteus* spp are strongly implicated and where the cross-reactive epitopes leading to ‘mimicry’ by and of host protein targets are understood [68,203,204]). Importantly, it has already been shown that both acute [205,206] and Long COVID [207] are accompanied by immunological dysfunction, and by novel antibodies [1,206], including in the latter case to (an ‘abnormal’ but unspecified form of) fibrin [118]. We recognise that any change in the conformation of a protein can result in the generation of novel epitopes that can thereby lead to the production of novel antibodies; indeed the use of secondary antibodies in the detection of small molecules [208] relies precisely on this fact. Although we consider that the more primary event is the generation of fibrinoid microclots, we also recognise that they are likely to be able to change the natural conformation of many proteins that might normally present as harmless (seen as ‘self’). However, the detailed nature of these autoantibodies is not known (but a subject of considerable present interest [1]).

## Epidemiological aspects

One approach to understanding the mechanisms of long COVID is to analyse epidemiological data, since the characteristics of those experiencing acute COVID differ markedly from those with long COVID [11]. Among these (and in significant contrast with the case of acute COVID [209]), is a striking over-representation of long COVID in women, and especially younger women [3,10,210]. From this point of view, it is of considerable interest that female sex hormones can induce anomalous blood clotting [46,57,211].

Haemophiliacs (and others with hypocoagulatory disorders) represent a complex test case, because depending on the stage of the disease [85], COVID can be both hyper- and hypocoagulatory [212]. However, we are not presently aware of any analyses of fibrinoid microclots in such individuals.

## Strain differences

While it has been somewhat slowed by the generally sluggish recognition of Long COVID, it is to be expected that different strains of SARS-CoV-2 may have different tendencies to induce it. To this end, it is reasonable that if microclots are important to LC their prevalence should also vary with the severity or frequency of LC induced by different strains of SARS-CoV-2. In a sense this would provide a very important kind of ‘control’, since the only thing varying as the stimulus is the strain of SARS-CoV-2.

## Other causes of amyloid microclots

While we here focus on SARS-CoV-2, we note that all kinds of molecules have been shown to affect the extent of fibrinoid clot formation, including iron, other amyloids, bacterial cell wall components, etc., and that we have observed them in a variety of chronic, inflammatory diseases including Alzheimer's [37,50,60–62], Parkinson's disease [37,48], Type 2 diabetes [37,38,62,63,213,214] (where the amyloid protein amylin [215–217] is of course a well-known player) and rheumatoid arthritis [65,67]. Although not yet tested directly, we consider it likely that this will also be true for infectious diseases known to be causing similar post-infection syndromes, such as Dengue [218–220], Ebola [221–223], Lyme [224], Zika and others where viruses persist and can cause microangiopathies [139] that we suspect are also amyloid in character. There is also likely to be a role for molecules raised in pre-existing inflammatory diseases, as well as substances produced by dysregulation microbiomes [225], and Leiden factor V [226,227].

## Similarities to ME/CFS

As well as the post-infection diseases referred to above, the emergence of long COVID has brought to the fore its similarities to other even more widely established syndromes such as myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS) [1,228], and full details are available in these recent reviews [1,228]. Berg and colleagues have also highlighted the role of coagulopathies in ME/CFS [229]. We note, too, that other 'anomalous' diseases bearing symptoms that overlap with those of ME/CFS include Gulf war syndrome [230], where again we would hazard that the analysis of fibrinoid microclot formation would have a positive outcome. Thus, while we consider it likely that the phenomena we describe will also broadly be true for ME/CFS, we focus here on PASC/Long COVID.

## Endotheliopathy

An important component of severe acute COVID-19 disease accompanying pathological clotting, is virus-induced endotheliopathy resulting in systemic endotheliitis [111,231–235]. Endotheliitis is central to initiating a state of failing normal clotting physiology and it is also known to be significantly linked to coagulopathies, as it activates microthrombotic pathways and initiates endotheliopathy-associated intravascular microthrombi [236].

## A role for iron dysregulation

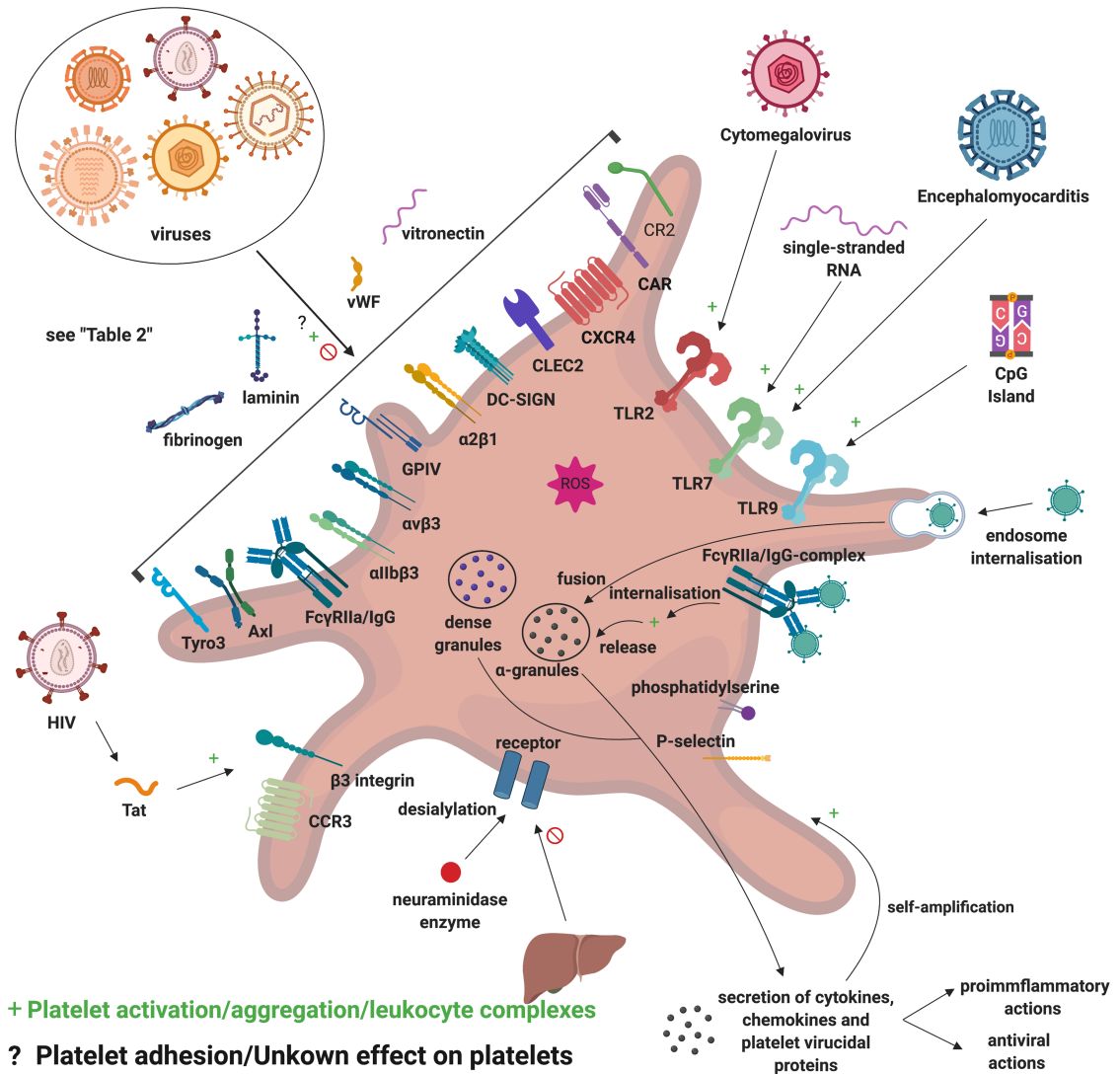
A consequence of cell death, such as occurs in endotheliopathy, is the release in COVID patients of the normally intracellular iron storage protein ferritin [237,238], which can then release free iron [239]. Iron dysregulation is an accompaniment to a huge number of chronic, inflammatory diseases [200,240,241], and iron dysregulation is a significant accompaniment in COVID [238,242–245]. Indeed, there is evidence that SARS-CoV-2 can release iron from haemoglobin directly [246]. The involvement of iron dysregulation would be consistent with the potentially protective effects of chelating it, e.g. with lactoferrin [247–250] or other chelators [251–254].

## Role of platelets

The important role that platelets play in acute COVID-19 (Figure 9), has been discussed in the context of disease severity, the development of endotheliopathy and as general drivers of coagulation pathology [232,233,255–257]. Platelets interact with circulating inflammatory molecules, the (damaged) endothelium itself, and also with immune cells, resulting in platelet/cellular and platelet/molecule complexes [258,259]. Platelet complexes are mediated by membrane-membrane interactions via receptor binding. Two of the molecules central in this discussion are fibrinogen and VWF, and platelets form significant complexes with them both [260]. Ultimately, platelet–cell and platelet–coagulation molecule protein complexes form part of platelet activation mechanisms and vascular remodeling and these complexes drive granule secretion, surface glycoprotein expression, and molecular activation platelet hyperactivation pathways [258,259].

## Consequences for treatment protocols

Given that there is a definite background removal rate of fibrinoid microclots, albeit slower than that of normal non-amyloid clots, it is then mostly necessary to ensure that they do not form further. Consequently, we do not see a role for 'clot-busters' as have occasionally been used when acute COVID-19 is accompanied by



**Figure 9. Platelet interactions with viruses.**

Figure adapted from [259]. Various platelet receptors can mediate binding to viral particles, where pattern recognition receptors recognize viral signals, viral products and also modulate platelet function. Platelets mediate viral attack by secreting virucidal proteins and by engulfing viral particles, as well as by interacting with immune cells and enhancing the immune response. Virus-platelet aggregates and platelets with a viral load are targeted by leukocytes, and platelets are ultimately cleared from the circulation (Figure created with <https://biorender.com/>).

Acute Respiratory Distress Syndrome [261]. However, addressing only one of the pathologies (either clot pathologies or platelet hyperactivation) will always tend to fail, as was also noted in recent clinical trials where just one of the two therapies was trialed in Acute COVID-19. Lawler and co-workers [262] investigated the use of therapeutic anticoagulation with heparin in noncritically ill patients with COVID-19. (We note too that heparin, like lactoferrin [250], may have antiviral properties [263–266].) Lopes and coworkers [267] also investigated the use of therapeutic versus prophylactic anticoagulation for patients admitted to hospital with COVID-19. Earlier, Viecca and colleagues reported [268] on a single-center, investigator-initiated, proof-of-concept, case control, phase IIb study. The study [268] explored the effects of anti-platelet therapy on arterial oxygenation and on clinical outcomes in patients with severe COVID-19 with hypercoagulability. Outcomes were not significantly positive in any of the trials mentioned.

However, it must be noted that the trials were done on acute COVID-19 patients. This has led us to suggest a multipronged approach and a regime of triple anticoagulation treatment [231], where Long COVID patients might be treated by one month of dual antiplatelet therapy (DAPT) (Clopidogrel 75 mg/Aspirin 75 mg) once a day, as well as a direct oral anticoagulant (DOAC) (Apixiban) 5 mg twice a day, together with a proton pump inhibitor (PPI) (e.g. pantoprazole 40 mg/day for gastric protection). Such a treatment regime showed promise under condition of clinical practice (there were no treatment-free ‘controls’) [231]. However, especially because of the potential for hypocoagulation (bleeding), it must only be followed under strict and qualified medical guidance in which we recommend the regular assessment of coagulation status before and during treatment. It (as do any other methods [269]) now needs to be studied in a proper randomized controlled trial.

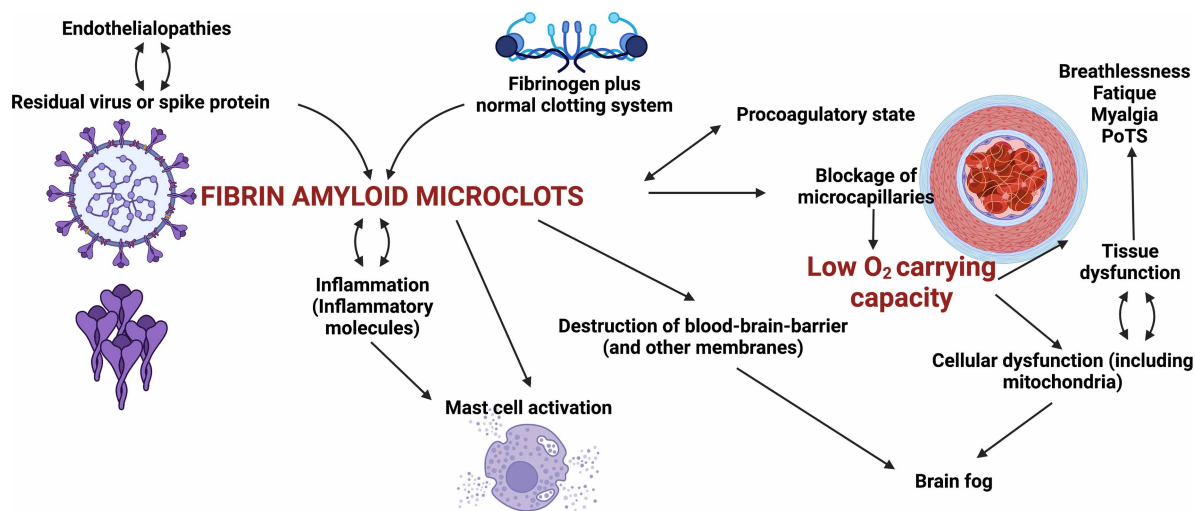
We do note that the use of triple therapy is not new in clinical practice. Its uses predate COVID-19, where it is successfully prescribed in patients where thromboses (which may or may not involve fibrinoid clots) are particularly worrisome and in various coronary diseases accompanied by atrial fibrillation. Here strong anti-coagulant treatments tailored to the needs of the individual patient are recommended [270], commonly involving dual or triple treatments with various anticoagulant agents. The present authors have focused on the ‘triple treatment’ for anticoagulation that involves an oral anticoagulant plus two drugs designed to decrease platelet activation (usually the P2Y<sub>12</sub> inhibitor clopidogrel, plus low-dose aspirin). (In addition, a gastric proton pump inhibitor is also given to reduce the likelihood of gastric bleeding.)

## What is needed next?

Many pieces of research-level evidence (especially [85,101,106,108,109,116,231,271]) suggest strongly that fibrin amyloid microclots, driven by the presence of the SARS-CoV-2 spike protein, are an inevitable accompaniment to (and a likely cause of) Long COVID. A biologically coherent explanation [272,273] can link such observations with the other observable symptoms of Long COVID, and thus serve to satisfy the logic normally required [179,274,275] to provide a causative explanation.

However, we are still potentially far from translating this kind of understanding into both diagnostics and therapeutics. Some of the elements that are needed both to make the evidence even more robust and to pass regulatory approval more widely include the following:

- Our fibrin amyloid diagnostic methods, presently semi-quantitative, must be subject to further optimization, made robust and quantitative, and reproduced more widely
- Work is required to provide a simple, cheap and widely available point-of-care instrument to effect the necessary measurements of the number, size and nature of fibrinoid microclots



**Figure 10. Some of the sequelae of fibrinoid microclot formation in the symptomatology of Long COVID.**

Many others, such as a role for auto-antibodies, are not shown.



- Longitudinal studies, and those relating fibrinolytic presence to the severity of Long COVID, will help recognise the role of different fibrinolytics in causing different symptoms; these may vary predictively between SARS-CoV-2 strains
- There would be value in assessing other blood parameters simultaneously (e.g. fibrinogen, D-dimer, von Willebrand Factor) plus suitable inflammatory cytokines
- Armed with the above, we should reasonably expect clinicians to be able to obtain ethics for a suitable designed Randomized Controlled Trial of anticoagulant and platelet inhibitor regimes, coupled to the assessment, using viscoelastic assays such as Thromboelastography (TEG or ROTEM or Sonoclot), before and during such treatments of coagulation potential so as to guard against any hypocoagulation and the dangers of bleeding.
- Such individuals should be followed for a period after the end of treatment to assess the degree of permanence

Of course it is easy to design detailed and expensive trials, accompanied by a great many other measurements of covariates, but the above sets out what we would consider as the minimum necessary to strengthen the claims that the analysis and removal of fibrin amyloid microclots should have major utility in providing substantial benefits to those with Long COVID, and likely in related conditions as well.

## Conclusions

Here we have argued, and focus on the fact, that Long COVID is characterized by the presence of persistent fibrin amyloid microclots that might block capillaries and inhibit the transport of O<sub>2</sub> to tissues, entrapping numerous inflammatory molecules, including those that prevent clot breakdown (as we have indeed recently shown) (Figure 10).

In addition to microclot formation, significant platelet dysfunction and a systemic endotheliitis drive systemic cellular hypoxia. These pathologies can explain most, if not all, of the lingering symptoms to which individuals with long COVID refer. We have noted that amyloid microclots, platelet hyperactivation and endothelial dysfunction, might form a suitable set of foci for the clinical treatment of the symptoms of long COVID [231]. Therefore, if fibrinolytic microclots are largely responsible for the symptoms of Long COVID, their removal is to be seen as paramount for relieving these symptoms and allowing the body to repair itself.

## Competing Interests

E.P. is a named inventor on a patent application covering the use of fluorescence methods for microclot detection in Long COVID.

## Open Access Statement

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## Abbreviations

DAPT, dual antiplatelet therapy; DIC, disseminated intravascular coagulopathy; INR, International normalized ratio; LPS, lipopolysaccharide; PASC, Post-acute sequelae of COVID; PPP, platelet-poor plasma; TEG, Thromboelastography.

## References

- 1 Proal, A.D. and VanElzakker, M.B. (2021) Long COVID or post-acute sequelae of COVID-19 (PASC): an overview of biological factors that may contribute to persistent symptoms. *Front. Microbiol.* **12**, 698169 <https://doi.org/10.3389/fmicb.2021.698169>
- 2 Logue, J.K., Franko, N.M., McCulloch, D.J., McDonald, D., Magedson, A., Wolf, C.R. et al. (2021) Sequelae in adults at 6 months after COVID-19 infection. *JAMA Netw. Open.* **4**, e210830 <https://doi.org/10.1001/jamanetworkopen.2021.0830>
- 3 Nalbandian, A., Sehgal, K., Gupta, A., Madhavan, M.V., McGroder, C., Stevens, J.S. et al. (2021) Post-acute COVID-19 syndrome. *Nat. Med.* **27**, 601–615 <https://doi.org/10.1038/s41591-021-01283-z>
- 4 Asadi-Pooya, A.A., Nemat, H., Shahisavandi, M., Akbari, A., Emami, A., Lotfi, M. et al. (2021) Long COVID in children and adolescents. *World J. Pediatr.* **17**, 495–499 <https://doi.org/10.1007/s12519-021-00457-6>



- 5 Gurdasani, D., Akrami, A., Bradley, V.C., Costello, A., Greenhalgh, T., Flaxman, S. et al. (2022) Long COVID in children. *Lancet Child Adolesc. Health* **6**, e2 [https://doi.org/10.1016/S2352-4642\(21\)00342-4](https://doi.org/10.1016/S2352-4642(21)00342-4)
- 6 Alwan, N.A. (2021) The road to addressing long COVID. *Science* **373**, 491–493 <https://doi.org/10.1126/science.abg7113>
- 7 Di Toro, A., Bozzani, A., Tavazzi, G., Urtis, M., Giuliani, L., Pizzoccheri, R. et al. (2021) Long COVID: long-term effects? *Eur. Heart J. Suppl.* **23**, E1–E5 <https://doi.org/10.1093/eurheartj/suab080>
- 8 Marshall, M. (2021) The four most urgent questions about long COVID. *Nature* **594**, 168–170 <https://doi.org/10.1038/d41586-021-01511-z>
- 9 Michelen, M., Manoharan, L., Elkheir, N., Cheng, V., Dagens, A., Hastie, C. et al. (2021) Characterising long COVID: a living systematic review. *BMJ Glob. Health* **6**, e005427 <https://doi.org/10.1136/bmjgh-2021-005427>
- 10 Sigfrid, L., Drake, T.M., Pauley, E., Jesudason, E.C., Olliaro, P., Lim, W.S. et al. (2021) Long COVID in adults discharged from UK hospitals after COVID-19: a prospective, multicentre cohort study using the ISARIC WHO clinical characterisation protocol. *Lancet Reg. Health Eur.* **8**, 100186 <https://doi.org/10.1016/j.lanepc.2021.100186>
- 11 Davis, H.E., Assaf, G.S., McCorkell, L., Wei, H., Low, R.J., Re'em, Y. et al. (2021) Characterizing long COVID in an international cohort: 7 months of symptoms and their impact. *EClinicalMedicine* **38**, 101019 <https://doi.org/10.1016/j.eclinm.2021.101019>
- 12 Deer, R.R., Rock, M.A., Vasilevsky, N., Carmody, L., Rando, H., Anzalone, A.J. et al. (2021) Characterizing long COVID: deep phenotype of a complex condition. *EBioMedicine* **74**, 103722 <https://doi.org/10.1016/j.ebiom.2021.103722>
- 13 Décarry, S., Gaboury, I., Poirier, S., Garcia, C., Simpson, S., Bull, M. et al. (2021) Humility and acceptance: working within Our limits With long COVID and myalgic encephalomyelitis/chronic fatigue syndrome. *J. Orthop. Sports Phys. Ther.* **51**, 197–200 <https://doi.org/10.2519/jospt.2021.0106>
- 14 Greenhalgh, T., Knight, M., A'Court, C., Buxton, M. and Husain, L. (2020) Management of post-acute COVID-19 in primary care. *BMJ* **370**, m3026 <https://doi.org/10.1136/bmj.m3026>
- 15 Anfinsen, C.B. (1973) Principles that govern the folding of protein chains. *Science* **181**, 223–230 <https://doi.org/10.1126/science.181.4096.223>
- 16 Prusiner, S.B. (1998) Prions. *Proc. Natl Acad. Sci. U.S.A.* **95**, 13363–13383 <https://doi.org/10.1073/pnas.95.23.13363>
- 17 Baral, P.K., Yin, J., Aguzzi, A. and James, M.N.G. (2019) Transition of the prion protein from a structured cellular form (PrP<sup>C</sup>) to the infectious scrapie agent (PrP<sup>Sc</sup>). *Protein Sci.* **28**, 2055–2063 <https://doi.org/10.1002/pro.3735>
- 18 Tycko, R. (2016) Structure of aggregates revealed. *Nature* **537**, 492–493 <https://doi.org/10.1038/nature19470>
- 19 Ke, P.C., Zhou, R., Serpell, L.C., Riek, R., Knowles, T.P.J., Lashuel, H.A. et al. (2020) Half a century of amyloids: past, present and future. *Chem. Soc. Rev.* **49**, 5473–5509 <https://doi.org/10.1039/C9CS00199A>
- 20 Collinge, J. (2016) Mammalian prions and their wider relevance in neurodegenerative diseases. *Nature* **539**, 217–226 <https://doi.org/10.1038/nature20415>
- 21 Iadanza, M.G., Jackson, M.P., Hewitt, E.W., Ranson, N.A. and Radford, S.E. (2018) A new era for understanding amyloid structures and disease. *Nat. Rev. Mol. Cell Biol.* **19**, 755–773 <https://doi.org/10.1038/s41580-018-0060-8>
- 22 Fitzpatrick, A.W. and Saibil, H.R. (2019) Cryo-EM of amyloid fibrils and cellular aggregates. *Curr. Opin. Struct. Biol.* **58**, 34–42 <https://doi.org/10.1016/j.sbi.2019.05.003>
- 23 Carta, M. and Aguzzi, A. (2021) Molecular foundations of prion strain diversity. *Curr. Opin. Neurobiol.* **72**, 22–31 <https://doi.org/10.1016/j.conb.2021.07.010>
- 24 Blancas-Mejía, L.M. and Ramirez-Alvarado, M. (2013) Systemic amyloidoses. *Annu. Rev. Biochem.* **82**, 745–774 <https://doi.org/10.1146/annurev-biochem-072611-130030>
- 25 Palladini, G. and Merlini, G. (2013) Systemic amyloidoses: what an internist should know. *Eur. J. Intern. Med.* **24**, 729–739 <https://doi.org/10.1016/j.ejim.2013.10.007>
- 26 Kell, D.B. and Pretorius, E. (2017) Proteins behaving badly. Substoichiometric molecular control and amplification of the initiation and nature of amyloid fibril formation: lessons from and for blood clotting. *Progr. Biophys. Mol. Biol.* **123**, 16–41 <https://doi.org/10.1016/j.pbiomolbio.2016.08.006>
- 27 Cao, K.J. and Yang, J. (2018) Translational opportunities for amyloid-targeting fluorophores. *Chem. Commun (Camb)* **54**, 9107–9118 <https://doi.org/10.1039/C8CC03619E>
- 28 Apter, B., Lapshina, N., Barhom, H., Fainberg, B., Handelman, A., Accardo, A. et al. (2020) Fluorescence phenomena in amyloid and amyloidogenic bionanostructures. *Crystals* **10**, 668 <https://doi.org/10.3390/cryst10080668>
- 29 Howie, A.J. and Brewer, D.B. (2009) Optical properties of amyloid stained by Congo red: history and mechanisms. *Micron* **40**, 285–301 <https://doi.org/10.1016/j.micron.2008.10.002>
- 30 Peccati, F., Pantalone, S., Riffet, V., Solans-Monfort, X., Contreras-Garcia, J., Guallar, V. et al. (2017) Binding of thioflavin T and related probes to polymorphic models of amyloid-beta fibrils. *J. Phys. Chem. B* **121**, 8926–8934 <https://doi.org/10.1021/acs.jpcc.7b06675>
- 31 Sulatskaya, A.I., Kuznetsova, I.M. and Turoverov, K.K. (2012) Interaction of thioflavin T with amyloid fibrils: fluorescence quantum yield of bound dye. *J. Phys. Chem. B* **116**, 2538–2544 <https://doi.org/10.1021/jp2083055>
- 32 Sulatskaya, A.I., Rodina, N.P., Sulatsky, M.I., Povarova, O.I., Antifeeva, I.A., Kuznetsova, I.M. et al. (2018) Investigation of alpha-Synuclein amyloid fibrils using the fluorescent probe thioflavin T. *Int. J. Mol. Sci.* **19**, 2486 <https://doi.org/10.3390/ijms19092486>
- 33 Khurana, R., Coleman, C., Ionescu-Zanetti, C., Carter, S.A., Krishna, V., Grover, R.K. et al. (2005) Mechanism of thioflavin T binding to amyloid fibrils. *J. Struct. Biol.* **151**, 229–238 <https://doi.org/10.1016/j.jsb.2005.06.006>
- 34 Amdursky, N., Erez, Y. and Huppert, D. (2012) Molecular rotors: what lies behind the high sensitivity of the thioflavin-T fluorescent marker. *Acc. Chem. Res.* **45**, 1548–1557 <https://doi.org/10.1021/ar300053p>
- 35 Biancalana, M. and Koide, S. (2010) Molecular mechanism of Thioflavin-T binding to amyloid fibrils. *Biochim. Biophys. Acta* **1804**, 1405–1412 <https://doi.org/10.1016/j.bbapap.2010.04.001>
- 36 Klingstedt, T. and Nilsson, K.P.R. (2012) Luminescent conjugated poly- and oligo-thiophenes: optical ligands for spectral assignment of a plethora of protein aggregates. *Biochem. Soc. Trans.* **40**, 704–710 <https://doi.org/10.1042/BST20120009>
- 37 de Waal, G.M., Engelbrecht, L., Davis, T., de Villiers, W.J.S., Kell, D.B. and Pretorius, E. (2018) Correlative light-electron microscopy detects lipopolysaccharide and its association with fibrin fibres in Parkinson's disease, Alzheimer's disease and type 2 diabetes mellitus. *Sci. Rep.* **8**, 16798 <https://doi.org/10.1038/s41598-018-35009-y>

- 38 Pretorius, E., Page, M.J., Engelbrecht, L., Ellis, G.C. and Kell, D.B. (2017) Substantial fibrin amyloidogenesis in type 2 diabetes assessed using amyloid-selective fluorescent stains. *Cardiovasc. Diabetol.* **16**, 141 <https://doi.org/10.1186/s12933-017-0624-5>
- 39 Baskurt, O.K., Hardeman, M.R., Rampling, M.W. and Meiselman, H.J. (2007) *Handbook of Hemorheology and Hemodynamics*, IOS Press, Amsterdam
- 40 Cilia La Corte, A.L., Philippou, H. and Ariens, R.A.S. (2011) Role of fibrin structure in thrombosis and vascular disease. *Adv. Protein Chem. Struct. Biol.* **83**, 75–127 <https://doi.org/10.1016/B978-0-12-381262-9.00003-3>
- 41 Litvinov, R.I., Pieters, M., de Lange-Loots, Z. and Weisel, J.W. (2021) Fibrinogen and fibrin. *Subcell. Biochem.* **96**, 471–501 [https://doi.org/10.1007/978-3-030-58971-4\\_15](https://doi.org/10.1007/978-3-030-58971-4_15)
- 42 Weisel, J.W. and Litvinov, R.I. (2017) Fibrin formation, structure and properties. *Subcell. Biochem.* **82**, 405–456 [https://doi.org/10.1007/978-3-319-49674-0\\_13](https://doi.org/10.1007/978-3-319-49674-0_13)
- 43 Undas, A. (2016) How to assess fibrinogen levels and fibrin clot properties in clinical practice? *Semin. Thromb. Hemost.* **42**, 381–388 <https://doi.org/10.1055/s-0036-1579636>
- 44 Pretorius, E., Briedenhann, S., Marx, J. and Franz, R.C. (2006) Structural changes in the fibrin network of a Pretoria family with dysfibrinogenemia: a scanning electron microscopical study. *Ultrastruct. Pathol.* **30**, 167–176 <https://doi.org/10.1080/01913120600689772>
- 45 Pretorius, E. and Lipinski, B. (2013) Differences in morphology of fibrin clots induced with thrombin and ferric ions and its pathophysiological consequences. *Heart Lung Circ.* **22**, 447–449 <https://doi.org/10.1016/j.hlc.2012.10.010>
- 46 Swanepoel, A.C., Visagie, A., de Lange, Z., Emmerson, O., Nielsen, V.G. and Pretorius, E. (2016) The clinical relevance of altered fibrinogen packaging in the presence of 17beta-estradiol and progesterone. *Thromb. Res.* **146**, 23–34 <https://doi.org/10.1016/j.thromres.2016.08.022>
- 47 Pretorius, E., Mbotwe, S., Bester, J., Robinson, C.J. and Kell, D.B. (2016) Acute induction of anomalous and amyloidogenic blood clotting by molecular amplification of highly substoichiometric levels of bacterial lipopolysaccharide. *J. R. Soc. Interface.* **123**, 20160539 <https://doi.org/10.1098/rsif.2016.0539>
- 48 Pretorius, E., Page, M.J., Mbotwe, S. and Kell, D.B. (2018) Lipopolysaccharide-binding protein (LBP) can reverse the amyloid state of fibrin seen or induced in Parkinson's disease. *PLoS ONE* **13**, e0192121 <https://doi.org/10.1371/journal.pone.0192121>
- 49 Pretorius, E., Page, M.J., Hendricks, L., Nkosi, N.B., Benson, S.R. and Kell, D.B. (2018) Both lipopolysaccharide and lipoteichoic acids potently induce anomalous fibrin amyloid formation: assessment with novel amytracker™ stains. *J. R. Soc. Interface* **15**, 20170941 <https://doi.org/10.1098/rsif.2017.0941>
- 50 Pretorius, E., Bester, J., Page, M.J. and Kell, D.B. (2018) The potential of LPS-binding protein to reverse amyloid formation in plasma fibrin of individuals with Alzheimer-type dementia. *Front. Aging Neurosci.* **10**, 257 <https://doi.org/10.3389/fnagi.2018.00257>
- 51 Jankun, J., Landeta, P., Pretorius, E., Skrzypczak-Jankun, E. and Lipinski, B. (2014) Unusual clotting dynamics of plasma supplemented with iron(III). *Int. J. Mol. Med.* **33**, 367–372 <https://doi.org/10.3892/ijmm.2013.1585>
- 52 Kell, D.B. and Pretorius, E. (2015) The simultaneous occurrence of both hypercoagulability and hypofibrinolysis in blood and serum during systemic inflammation, and the roles of iron and fibrin(ogen). *Integr. Biol.* **7**, 24–52 <https://doi.org/10.1039/c4ib00173g>
- 53 Lipinski, B., Pretorius, E., Oberholzer, H.M. and Van Der Spuy, W.J. (2012) Iron enhances generation of fibrin fibers in human blood: implications for pathogenesis of stroke. *Microsc. Res. Tech.* **75**, 1185–1190 <https://doi.org/10.1002/jemt.22047>
- 54 Pretorius, E., Vermeulen, N., Bester, J. and Lipinski, B. (2013) Novel use of scanning electron microscopy for detection of iron-induced morphological changes in human blood. *Microsc. Res. Tech.* **76**, 268–271 <https://doi.org/10.1002/jemt.22163>
- 55 Pretorius, E., Bester, J., Vermeulen, N., Lipinski, B., Gericke, G.S. and Kell, D.B. (2014) Profound morphological changes in the erythrocytes and fibrin networks of patients with hemochromatosis or with hyperferritinemia, and their normalization by iron chelators and other agents. *PLoS ONE* **9**, e85271 <https://doi.org/10.1371/journal.pone.0085271>
- 56 Pretorius, E. and Kell, D.B. (2014) Diagnostic morphology: biophysical indicators for iron-driven inflammatory diseases. *Integr. Biol.* **6**, 486–510 <https://doi.org/10.1039/C4IB00025K>
- 57 Swanepoel, A.C., Lindeque, B.G., Swart, P.J., Abdool, Z. and Pretorius, E. (2014) Estrogen causes ultrastructural changes of fibrin networks during the menstrual cycle: a qualitative investigation. *Microsc. Res. Tech.* **77**, 594–601 <https://doi.org/10.1002/jemt.22378>
- 58 Page, M.J., Thomson, G.J.A., Nunes, J.M., Engelbrecht, A.M., Nell, T.A., de Villiers, W.J.S. et al. (2019) Serum amyloid A binds to fibrin(ogen), promoting fibrin amyloid formation. *Sci. Rep.* **9**, 3102 <https://doi.org/10.1038/s41598-019-39056-x>
- 59 Bester, J., Soma, P., Kell, D.B. and Pretorius, E. (2015) Viscoelastic and ultrastructural characteristics of whole blood and plasma in Alzheimer-type dementia, and the possible role of bacterial lipopolysaccharides (LPS). *Oncotarget Gerontol.* **6**, 35284–35303 <https://doi.org/10.18632/oncotarget.6074>
- 60 Lipinski, B. and Pretorius, E. (2013) The role of iron-induced fibrin in the pathogenesis of Alzheimer's disease and the protective role of magnesium. *Front. Hum. Neurosci.* **7**, 735 <https://doi.org/10.3389/fnhum.2013.00735>
- 61 Lipinski, B. and Pretorius, W. (2014) Iron-induced fibrin formation may explain vascular pathology in Alzheimer's disease. *Folia Neuropathol.* **52**, 205 <https://doi.org/10.5114/fn.2014.43792>
- 62 Pretorius, E., Oberholzer, H.M., van der Spuy, W.J., Swanepoel, A.C. and Soma, P. (2011) Qualitative scanning electron microscopy analysis of fibrin networks and platelet abnormalities in diabetes. *Blood Coagul. Fibrinol.* **22**, 463–467 <https://doi.org/10.1097/MBC.0b013e3283468a0d>
- 63 Pretorius, E., Bester, J., Vermeulen, N., Alummoottil, S., Soma, P., Buys, A.V. et al. (2015) Poorly controlled type 2 diabetes is accompanied by significant morphological and ultrastructural changes in both erythrocytes and in thrombin-generated fibrin: implications for diagnostics. *Cardiovasc. Diabetol.* **13**, 30 <https://doi.org/10.1186/s12933-015-0192-5>
- 64 Soma, P. and Pretorius, E. (2015) Interplay between ultrastructural findings and atherothrombotic complications in type 2 diabetes mellitus. *Cardiovasc. Diabetol.* **14**, 96 <https://doi.org/10.1186/s12933-015-0261-9>
- 65 Bezuidenhout, J., Venter, C., Roberts, T., Tarr, G., Kell, D. and Pretorius, E. (2020) The atypical fibrin fibre network in rheumatoid arthritis and its relation to autoimmunity, inflammation and thrombosis. *bioRxiv* 2020.05.28.121301v1 <https://doi.org/10.1101/2020.05.28.121301>
- 66 Bezuidenhout, J., Venter, C., Roberts, T., Tarr, G., Kell, D. and Pretorius, E. (2020) Detection of citrullinated fibrin in plasma clots of RA patients and its relation to altered structural clot properties, disease-related inflammation and prothrombotic tendency. *Front. Immunol.* **11**, 577523 <https://doi.org/10.3389/fimmu.2020.577523>
- 67 Pretorius, E., Oberholzer, H.M., van der Spuy, W.J., Swanepoel, A.C. and Soma, P. (2012) Scanning electron microscopy of fibrin networks in rheumatoid arthritis: a qualitative analysis. *Rheumatol. Int.* **32**, 1611–1615 <https://doi.org/10.1007/s00296-011-1805-2>

- 68 Pretorius, E., Akeredolu, O.-O., Soma, P. and Kell, D.B. (2017) Major involvement of bacterial components in rheumatoid arthritis and its accompanying oxidative stress, systemic inflammation and hypercoagulability. *Exp. Biol. Med.* **242**, 355–373 <https://doi.org/10.1177/1535370216681549>
- 69 Buhimschi, I.A., Nayeri, U.A., Zhao, G., Shook, L.L., Pensalfini, A., Funai, E.F. et al. (2014) Protein misfolding, congophilia, oligomerization, and defective amyloid processing in preeclampsia. *Sci. Transl. Med.* **6**, 245ra292 <https://doi.org/10.1126/scitranslmed.3008808>
- 70 Kell, D.B. and Kenny, L.C. (2016) A dormant microbial component in the development of pre-eclampsia. *Front. Med. Obs. Gynecol.* **3**, 60 <https://doi.org/10.3389/fmed.2016.00060>
- 71 Kenny, L.C. and Kell, D.B. (2018) Immunological tolerance, pregnancy and pre-eclampsia: the roles of semen microbes and the father. *Front. Med. Obs. Gynecol.* **4**, 239 <https://doi.org/10.3389/fmed.2017.00239>
- 72 Litvinov, R.I., Faizullin, D.A., Zuev, Y.F. and Weisel, J.W. (2012) The alpha-helix to beta-sheet transition in stretched and compressed hydrated fibrin clots. *Biophys. J.* **103**, 1020–1027 <https://doi.org/10.1016/j.bpj.2012.07.046>
- 73 Benson, M.D., Liepnieks, J., Uemichi, T., Wheeler, G. and Correa, R. (1993) Hereditary renal amyloidosis associated with a mutant fibrinogen alpha-chain. *Nat. Genet.* **3**, 252–255 <https://doi.org/10.1038/ng0393-252>
- 74 Haidinger, M., Werzowa, J., Kain, R., Antlanger, M., Hecking, M., Pfaffenberger, S. et al. (2013) Hereditary amyloidosis caused by R554L fibrinogen aalpha-chain mutation in a spanish family and review of the literature. *Amyloid* **20**, 72–79 <https://doi.org/10.3109/13506129.2013.781998>
- 75 Lendrum, A.C., Slidders, W. and Fraser, D.S. (1972) Renal hyalin. A study of amyloidosis and diabetic fibrinous vasculosis with new staining methods. *J. Clin. Pathol.* **25**, 373–396 <https://doi.org/10.1136/jcp.25.5.373>
- 76 Kranenburg, O., Bouma, B., Kroon-Batenburg, L.M.J., Reijkerker, A., Wu, Y.P., Voest, E.E. et al. (2002) Tissue-type plasminogen activator is a multiligand cross-beta structure receptor. *Curr. Biol.* **12**, 1833–1839 [https://doi.org/10.1016/S0960-9822\(02\)01224-1](https://doi.org/10.1016/S0960-9822(02)01224-1)
- 77 Aguzzi, A. and Calella, A.M. (2009) Prions: protein aggregation and infectious diseases. *Physiol. Rev.* **89**, 1105–1152 <https://doi.org/10.1152/physrev.00006.2009>
- 78 Silva, C.J., Vazquez-Fernández, E., Onisko, B. and Requena, J.R. (2015) Proteinase K and the structure of PrP<sup>Sc</sup>: the good, the bad and the ugly. *Virus Res.* **207**, 120–126 <https://doi.org/10.1016/j.virusres.2015.03.008>
- 79 Wang, F., Wang, X., Abskharon, R. and Ma, J. (2018) Prion infectivity is encoded exclusively within the structure of proteinase K-resistant fragments of synthetically generated recombinant PrP(Sc). *Acta Neuropathol. Commun.* **6**, 30 <https://doi.org/10.1186/s40478-018-0534-0>
- 80 Fitzpatrick, A.W.P., Falcon, B., He, S., Murzin, A.G., Murshudov, G., Garringer, H.J. et al. (2017) Cryo-EM structures of tau filaments from Alzheimer's disease. *Nature* **547**, 185–190 <https://doi.org/10.1038/nature23002>
- 81 Li, D. and Liu, C. (2020) Structural diversity of amyloid fibrils and advances in their structure determination. *Biochemistry* **59**, 639–646 <https://doi.org/10.1021/acs.biochem.9b01069>
- 82 Lavatelli, F., di Fonzo, A., Palladini, G. and Merlini, G. (2016) Systemic amyloidoses and proteomics: the state of the art. *EuPA Open Proteom.* **11**, 4–10 <https://doi.org/10.1016/j.euprot.2016.02.003>
- 83 Linke, R.P. (2012) On typing amyloidosis using immunohistochemistry. Detailed illustrations, review and a note on mass spectrometry. *Prog. Histochem. Cytochem.* **47**, 61–132 <https://doi.org/10.1016/j.proghi.2012.03.001>
- 84 Altschul, D.J., Unda, S.R., de La Garza Ramos, R., Zampolin, R., Benton, J., Holland, R. et al. (2020) Hemorrhagic presentations of COVID-19: risk factors for mortality. *Clin. Neurol. Neurosurg.* **198**, 106112 <https://doi.org/10.1016/j.clineuro.2020.106112>
- 85 Grobler, C., Maphumulo, S.C., Grobbelaar, L.M., Bredenkamp, J., Laubscher, J., Lourens, P.J. et al. (2020) COVID-19: the rollercoaster of fibrin(ogen), D-dimer, von willebrand factor, P-selectin and their interactions with endothelial cells, platelets and erythrocytes. *Int. J. Mol. Sci.* **21**, 5168 <https://doi.org/10.3390/ijms21145168>
- 86 Iba, T., Levy, J.H., Levi, M. and Thachil, J. (2020) Coagulopathy in COVID-19. *J. Thromb. Haemost.* **18**, 2103–2109 <https://doi.org/10.1111/jth.14975>
- 87 Klok, F.A., Kruip, M., van der Meer, N.J.M., Arbous, M.S., Gommers, D., Kant, K.M. et al. (2020) Confirmation of the high cumulative incidence of thrombotic complications in critically ill ICU patients with COVID-19: an updated analysis. *Thromb. Res.* **191**, 148–150 <https://doi.org/10.1016/j.thromres.2020.04.041>
- 88 Malas, M.B., Naazie, I.N., Elsayed, N., Mathlouthi, A., Marmor, R. and Clary, B. (2020) Thromboembolism risk of COVID-19 is high and associated with a higher risk of mortality: a systematic review and meta-analysis. *EClinicalMedicine* **29**, 100639 <https://doi.org/10.1016/j.eclinm.2020.100639>
- 89 Thachil, J., Tang, N., Gando, S., Falanga, A., Cattaneo, M., Levi, M. et al. (2020) ISTH interim guidance on recognition and management of coagulopathy in COVID-19. *J. Thromb. Haemost.* **18**, 1023–1026 <https://doi.org/10.1111/jth.14810>
- 90 Gómez-Mesa, J.E., Galindo-Coral, S., Montes, M.C. and Muñoz Martin, A.J. (2021) Thrombosis and coagulopathy in COVID-19. *Curr. Probl. Cardiol.* **46**, 100742 <https://doi.org/10.1016/j.cpcardiol.2020.100742>
- 91 Leentjens, J., van Haaps, T.F., Wessels, P.F., Schutgens, R.E.G. and Middeldorp, S. (2021) COVID-19-associated coagulopathy and antithrombotic agents-lessons after 1 year. *Lancet Haematol.* **8**, e524–e533 [https://doi.org/10.1016/S2352-3026\(21\)00105-8](https://doi.org/10.1016/S2352-3026(21)00105-8)
- 92 Lorini, F.L., Di Matteo, M., Gritti, P., Grazioli, L., Benigni, A., Zacchetti, L. et al. (2021) Coagulopathy and COVID-19. *Eur. Heart J. Suppl.* **23**, E95–E98 <https://doi.org/10.1093/eurheartj/uaab100>
- 93 Polimeni, A., Leo, I., Spaccarotella, C., Mongiardo, A., Sorrentino, S., Sabatino, J. et al. (2021) Differences in coagulopathy indices in patients with severe versus non-severe COVID-19: a meta-analysis of 35 studies and 6427 patients. *Sci. Rep.* **11**, 10464 <https://doi.org/10.1038/s41598-021-89967-x>
- 94 Sarkar, M., Madabhavi, I.V., Quy, P.N. and Govindagoudar, M.B. (2021) COVID-19 and coagulopathy. *Clin. Respir. J.* **15**, 1259–1274 <https://doi.org/10.1111/crj.13438>
- 95 Vincent, J.L., Levi, M. and Hunt, B.J. (2021) Prevention and management of thrombosis in hospitalised patients with COVID-19 pneumonia. *Lancet Respir. Med.* **10**, 214–220 [https://doi.org/10.1016/S2213-2600\(21\)00455-0](https://doi.org/10.1016/S2213-2600(21)00455-0)
- 96 Ortega-Paz, L., Capodanno, D., Montalescot, G. and Angiolillo, D.J. (2021) Coronavirus disease 2019-associated thrombosis and coagulopathy: review of the pathophysiological characteristics and implications for antithrombotic management. *J. Am Heart Assoc.* **10**, e019650. <https://doi.org/10.1161/JAHA.120.019650>
- 97 Wang, Z., Gao, X., Miao, H., Ma, X. and Ding, R. (2021) Understanding COVID-19-associated coagulopathy: from PIC to SIC or DIC. *J. Intens. Med.* **1**, 35–41 <https://doi.org/10.1016/j.jointm.2021.03.002>

- 98 Douillet, D., Riou, J., Penalzoza, A., Moumneh, T., Soulie, C., Savary, D. et al. (2021) Risk of symptomatic venous thromboembolism in mild and moderate COVID-19: a comparison of two prospective European cohorts. *Thromb. Res.* **208**, 4–10 <https://doi.org/10.1016/j.thromres.2021.10.001>
- 99 Levi, M. and Thachil, J. (2020) Coronavirus disease 2019 coagulopathy: disseminated intravascular coagulation and thrombotic microangiopathy—Either, neither, or both. *Semin. Thromb. Hemost.* **46**, 781–784 <https://doi.org/10.1055/s-0040-1712156>
- 100 Thachil, J. (2021) Lessons learnt from COVID-19 coagulopathy. *EJHaem* **2**, 577–584 <https://doi.org/10.1002/jha2.228>
- 101 Venter, C., Bezuidenhout, J.A., Laubscher, G.J., Lourens, P.J., Steenkamp, J., Kell, D.B. et al. (2020) Erythrocyte, platelet, serum ferritin and P-selectin pathophysiology implicated in severe hypercoagulation and vascular complications in COVID-19. *Int. J. Mol. Sci.* **21**, 8234 <https://doi.org/10.3390/ijms21218234>
- 102 Fogarty, H., Townsend, L., Ni Cheallaigh, C., Bergin, C., Martin-Loeches, I., Browne, P. et al. (2020) COVID-19 coagulopathy in Caucasian patients. *Br. J. Haematol.* **189**, 1044–1049 <https://doi.org/10.1111/bjh.16749>
- 103 Bray, M.A., Sartain, S.E., Gollamudi, J. and Rumbaut, R.E. (2020) Microvascular thrombosis: experimental and clinical implications. *Transl. Res.* **225**, 105–130 <https://doi.org/10.1016/j.trsl.2020.05.006>
- 104 Levi, M., Thachil, J., Iba, T. and Levy, J.H. (2020) Coagulation abnormalities and thrombosis in patients with COVID-19. *Lancet Haematol.* **7**, e438–e440 [https://doi.org/10.1016/S2352-3026\(20\)30145-9](https://doi.org/10.1016/S2352-3026(20)30145-9)
- 105 Willyard, C. (2020) Coronavirus blood-clot mystery intensifies. *Nature* **581**, 250 <https://doi.org/10.1038/d41586-020-01403-8>
- 106 Pretorius, E., Venter, C., Laubscher, G.J., Lourens, P.J., Steenkamp, J. and Kell, D.B. (2020) Prevalence of amyloid blood clots in COVID-19 plasma. *medRxiv* <https://doi.org/10.1101/2020.07.28.20163543>
- 107 Asakura, H. and Ogawa, H. (2021) COVID-19-associated coagulopathy and disseminated intravascular coagulation. *Int. J. Hematol.* **113**, 45–57 <https://doi.org/10.1007/s12185-020-03029-y>
- 108 Grobelaar, L.M., Venter, C., Vlok, M., Ngoepe, M., Laubscher, G.J., Lourens, P.J. et al. (2021) SARS-CoV-2 spike protein S1 induces fibrin(ogen) resistant to fibrinolysis: implications for microclot formation in COVID-19. *Biosci. Rep.* **41**, BSR20210611 <https://doi.org/10.1042/BSR20210611>
- 109 Laubscher, G.J., Lourens, P.J., Venter, C., Kell, D.B. and Pretorius, E. (2021) TEG®, microclot and platelet mapping for guiding early management of severe COVID-19 coagulopathy. *J. Clin. Med.* **10**, 5381 <https://doi.org/10.3390/jcm10225381>
- 110 Wygrecka, M., Birnhuber, A., Seeliger, B., Michalick, L., Pak, O., Schultz, A.S. et al. (2021) Altered fibrin clot structure and dysregulated fibrinolysis contribute to thrombosis risk in severe COVID-19. *Blood Adv.* **6**, 1074–1087 <https://doi.org/10.1182/bloodadvances.2021004816>
- 111 Wool, G.D. and Miller, J.L. (2021) The impact of COVID-19 disease on platelets and coagulation. *Pathobiology* **88**, 15–27 <https://doi.org/10.1159/000512007>
- 112 Iba, T., Levy, J.H., Connors, J.M., Warkentin, T.E., Thachil, J. and Levi, M. (2020) The unique characteristics of COVID-19 coagulopathy. *Crit. Care* **24**, 360 <https://doi.org/10.1186/s13054-020-03077-0>
- 113 Loo, J., Spittle, D.A. and Newnham, M. (2021) COVID-19, immunothrombosis and venous thromboembolism: biological mechanisms. *Thorax* **76**, 412–420 <https://doi.org/10.1136/thoraxjnl-2020-216243>
- 114 Manolis, A.S., Manolis, T.A., Manolis, A.A., Papatheou, D. and Melita, H. (2021) COVID-19 Infection: viral macro- and micro-Vascular coagulopathy and thromboembolism/Prophylactic and therapeutic management. *J. Cardiovasc. Pharmacol. Ther.* **26**, 12–24 <https://doi.org/10.1177/1074248420958973>
- 115 Poor, H.D. (2021) Pulmonary thrombosis and thromboembolism in COVID-19. *Chest* **160**, 1471–1480 <https://doi.org/10.1016/j.chest.2021.06.016>
- 116 Pretorius, E., Vlok, M., Venter, C., Bezuidenhout, J.A., Laubscher, G.J., Steenkamp, J. et al. (2021) Persistent clotting protein pathology in long COVID/post-Acute sequelae of COVID-19 (PASC) is accompanied by increased levels of antiplasmin. *Cardiovasc. Diabetol.* **20**, 172 <https://doi.org/10.1186/s12933-021-01359-7>
- 117 Zuin, M., Engelen, M.M., Barco, S., Spyropoulos, A.C., Vanassche, T., Hunt, B.J. et al. (2021) Incidence of venous thromboembolic events in COVID-19 patients after hospital discharge: a systematic review and meta-analysis. *Thromb. Res.* **209**, 94–98 <https://doi.org/10.1016/j.thromres.2021.11.029>
- 118 Ryu, J.K., Sozmen, E.G., Dixit, K., Montano, M., Matsui, Y., Liu, Y. et al. (2021) SARS-CoV-2 spike protein induces abnormal inflammatory blood clots neutralized by fibrin immunotherapy. *bioRxiv* 2021.10.12.464152 <https://doi.org/10.1101/2021.10.12.464152>
- 119 Marfella, R., Paolisso, P., Sardu, C., Palomba, L., D'Onofrio, N., Cesaro, A. et al. (2021) SARS-CoV-2 colonizes coronary thrombus and impairs heart microcirculation bed in asymptomatic SARS-CoV-2 positive subjects with acute myocardial infarction. *Crit. Care* **25**, 217 <https://doi.org/10.1186/s13054-021-03643-0>
- 120 Jana, A.K., Greenwood, A.B. and Hansmann, U.H.E. (2021) Presence of a SARS-CoV-2 protein enhances amyloid formation of serum amyloid A. *J. Phys. Chem. B* **125**, 9155–9167 <https://doi.org/10.1021/acs.jpcc.1c04871>
- 121 Itzhaki, R.F., Lathe, R., Balin, B.J., Ball, M.J., Braak, H., Bearer, E.L. et al. (2016) Microbes and Alzheimer's disease. *J. Alzheimers Dis.* **51**, 979–984 <https://doi.org/10.3233/JAD-160152>
- 122 Brothers, H.M., Gosztyla, M.L. and Robinson, S.R. (2018) The physiological roles of amyloid-beta peptide hint at New ways to treat Alzheimer's disease. *Front. Aging Neurosci.* **10**, 118 <https://doi.org/10.3389/fnagi.2018.00118>
- 123 Eimer, W.A., Vijaya Kumar, D.K., Navalpur Shanmugam, N.K., Rodriguez, A.S., Mitchell, T., Washicosky, K.J. et al. (2018) Alzheimer's disease-Associated beta-Amyloid Is rapidly seeded by herpesviridae to protect against brain infection. *Neuron* **99**, 56–63.e53 <https://doi.org/10.1016/j.neuron.2018.06.030>
- 124 Bortolotti, D., Gentili, V., Rotola, A., Caselli, E. and Rizzo, R. (2019) HHV-6A infection induces amyloid-beta expression and activation of microglial cells. *Alzheimers Res. Ther.* **11**, 104 <https://doi.org/10.1186/s13195-019-0552-6>
- 125 Sait, A., Angeli, C., Doig, A.J. and Day, P.J.R. (2021) Viral involvement in Alzheimer's disease. *ACS Chem. Neurosci.* **12**, 1049–1060 <https://doi.org/10.1021/acschemneuro.0c00719>
- 126 Hansell, D.M., Goldin, J.G., King, Jr, T.E., Lynch, D.A., Richeldi, L. and Wells, A.U. (2015) CT staging and monitoring of fibrotic interstitial lung diseases in clinical practice and treatment trials: a position paper from the fleischner society. *Lancet Respir. Med.* **3**, 483–496 [https://doi.org/10.1016/S2213-2600\(15\)00096-X](https://doi.org/10.1016/S2213-2600(15)00096-X)
- 127 Parekh, M., Donuru, A., Balasubramanya, R. and Kapur, S. (2020) Review of the chest CT differential diagnosis of ground-Glass opacities in the COVID Era. *Radiology* **297**, E289–E302 <https://doi.org/10.1148/radiol.2020202504>
- 128 Warman, A., Warman, P., Sharma, A., Parikh, P., Warman, R., Viswanadhan, N. et al. (2020) Interpretable artificial intelligence for COVID-19 diagnosis from chest CT reveals specificity of ground-Glass opacities. *medRxiv* <https://doi.org/10.1101/2020.05.16.20103408>



- 129 Zhao, W., Zhong, Z., Xie, X., Yu, Q. and Liu, J. (2020) Relation between chest CT findings and clinical conditions of coronavirus disease (COVID-19) pneumonia: a multicenter study. *AJR Am. J. Roentgenol.* **214**, 1072–1077 <https://doi.org/10.2214/AJR.20.22976>
- 130 Cozzi, D., Cavigli, E., Moroni, C., Smorchkova, O., Zantonelli, G., Pradella, S. et al. (2021) Ground-glass opacity (GGO): a review of the differential diagnosis in the era of COVID-19. *Jpn J. Radiol.* **39**, 721–732 <https://doi.org/10.1007/s11604-021-01120-w>
- 131 Roig-Marín, N. and Roig-Rico, P. (2021) Ground-glass opacity on emergency department chest X-ray: a risk factor for in-hospital mortality and organ failure in elderly admitted for COVID-19. *Postgrad. Med.* <https://doi.org/10.1080/00325481.2021.2021741>
- 132 Wang, D., Shang, Y., Chen, Y., Xia, J., Tian, W., Zhang, T. et al. (2021) Clinical value of COVID-19 chest radiography and high-resolution CT examination. *Curr. Med. Imaging* <https://doi.org/10.2174/1573405618666211229143121>
- 133 Pickford, H.A., Swensen, S.J. and Utz, J.P. (1997) Thoracic cross-sectional imaging of amyloidosis. *AJR Am. J. Roentgenol.* **168**, 351–355 <https://doi.org/10.2214/ajr.168.2.9016204>
- 134 Milani, P., Basset, M., Russo, F., Foli, A., Palladini, G. and Merlini, G. (2017) The lung in amyloidosis. *Eur. Respir. Rev.* **26**, 170046 <https://doi.org/10.1183/16000617.0046-2017>
- 135 Liu, Y., Jin, Z., Zhang, H., Zhang, Y., Shi, M., Meng, F. et al. (2018) Diffuse parenchymal pulmonary amyloidosis associated with multiple myeloma: a case report and systematic review of the literature. *BMC Cancer* **18**, 802 <https://doi.org/10.1186/s12885-018-4565-5>
- 136 Baumgart, J.V., Stuhlmann-Laeisz, C., Hegenbart, U., Nattenmüller, J., Schönland, S., Krüger, S. et al. (2018) Local vs. systemic pulmonary amyloidosis-impact on diagnostics and clinical management. *Virchows. Arch.* **473**, 627–637 <https://doi.org/10.1007/s00428-018-2442-x>
- 137 Frija-Masson, J., Debray, M.P., Boussouar, S., Khalil, A., Bancal, C., Motiejunaite, J. et al. (2021) Residual ground glass opacities three months after COVID-19 pneumonia correlate to alteration of respiratory function: the post COVID M3 study. *Respir. Med.* **184**, 106435 <https://doi.org/10.1016/j.rmed.2021.106435>
- 138 Zheng, X.L. and Sadler, J.E. (2008) Pathogenesis of thrombotic microangiopathies. *Annu. Rev. Pathol.* **3**, 249–277 <https://doi.org/10.1146/annurev.pathmechdis.3.121806.154311>
- 139 Lopes da Silva, R. (2011) Viral-associated thrombotic microangiopathies. *Hematol. Oncol. Stem Cell Ther.* **4**, 51–59 <https://doi.org/10.5144/1658-3876.2011.51>
- 140 Bommer, M., Wolffe-Guter, M., Bohl, S. and Kuchenbauer, F. (2018) The differential diagnosis and treatment of thrombotic microangiopathies. *Dtsch. Arztebl. Int.* **115**, 327–334 <https://doi.org/10.3238/arztebl.2018.0327>
- 141 Fakhouri, F. (2016) Pregnancy-related thrombotic microangiopathies: clues from complement biology. *Transfus. Apher. Sci.* **54**, 199–202 <https://doi.org/10.1016/j.transci.2016.04.009>
- 142 Rosove, M.H. (2014) Thrombotic microangiopathies. *Semin. Arthritis Rheum.* **43**, 797–805 <https://doi.org/10.1016/j.semarthrit.2013.11.004>
- 143 Magnusson, K., Simon, R., Sjölander, D., Sigurdson, C.J., Hammarström, P. and Nilsson, K.P.R. (2014) Multimodal fluorescence microscopy of prion strain specific PrP deposits stained by thiophene-based amyloid ligands. *Prion* **8**, 319–329 <https://doi.org/10.4161/pri.29239>
- 144 Magnusson, K., Appelqvist, H., Cieslar-Pobuda, A., Back, M., Kågedal, B., Jonasson, J.A. et al. (2015) An imidazole functionalized pentameric thiophene displays different staining patterns in normal and malignant cells. *Front. Chem.* **3**, 58 <https://doi.org/10.3389/fchem.2015.00058>
- 145 Wegenast-Braun, B.M., Skodras, A., Bayraktar, G., Mahler, J., Fritschi, S.K., Klingstedt, T. et al. (2012) Spectral discrimination of cerebral amyloid lesions after peripheral application of luminescent conjugated oligothiophenes. *Am. J. Pathol.* **181**, 1953–1960 <https://doi.org/10.1016/j.ajpath.2012.08.031>
- 146 Düz, M.E., Balci, A. and Menekşe, E. (2020) D-dimer levels and COVID-19 severity: systematic review and meta-Analysis. *Tuberk. Toraks.* **68**, 353–360 <https://doi.org/10.5578/t.70351>
- 147 Favalaro, E.J. and Thachil, J. (2020) Reporting of D-dimer data in COVID-19: some confusion and potential for misinformation. *Clin. Chem. Lab. Med.* **58**, 1191–1199 <https://doi.org/10.1515/cclm-2020-0573>
- 148 Gerber, G.F. and Chaturvedi, S. (2021) How to recognize and manage COVID-19-associated coagulopathy. *Hematol. Am. Soc. Hematol. Educ. Program* **2021**, 614–620 <https://doi.org/10.1182/hematology.2021000297>
- 149 Mehrdad, R., Zahra, K. and Mansouritorghabeh, H. (2021) Hemostatic system (Fibrinogen level, D-Dimer, and FDP) in severe and Non-Severe patients With COVID-19: a systematic review and meta-Analysis. *Clin. Appl. Thromb. Hemost.* **27**, 10760296211010973 <https://doi.org/10.1177/10760296211010973>
- 150 Paliogiannis, P., Mangoni, A.A., Dettori, P., Nasrallah, G.K., Pintus, G. and Zinellu, A. (2020) D-Dimer concentrations and COVID-19 severity: a systematic review and meta-Analysis. *Front. Public Health* **8**, 432 <https://doi.org/10.3389/fpubh.2020.00432>
- 151 Poudel, A., Poudel, Y., Adhikari, A., Aryal, B.B., Dangol, D., Bajracharya, T. et al. (2021) D-dimer as a biomarker for assessment of COVID-19 prognosis: D-dimer levels on admission and its role in predicting disease outcome in hospitalized patients with COVID-19. *PLoS ONE* **16**, e0256744 <https://doi.org/10.1371/journal.pone.0256744>
- 152 Rostami, M. and Mansouritorghabeh, H. (2020) D-dimer level in COVID-19 infection: a systematic review. *Expert Rev. Hematol.* **13**, 1265–1275 <https://doi.org/10.1080/17474086.2020.1831383>
- 153 Shah, S., Shah, K., Patel, S.B., Patel, F.S., Osman, M., Velagapudi, P. et al. (2020) Elevated D-Dimer levels are associated with increased risk of mortality in COVID-19: a systematic review and meta-Analysis. *Cardiol. Rev.* **28**, 295–302 <https://doi.org/10.1097/CRD.0000000000000330>
- 154 Wang, Q., Cheng, J., Shang, J., Wang, Y., Wan, J., Yan, Y.Q. et al. (2021) Clinical value of laboratory indicators for predicting disease progression and death in patients with COVID-19: a retrospective cohort study. *BMJ Open* **11**, e043790 <https://doi.org/10.1136/bmjopen-2020-043790>
- 155 Kupferschmidt, K. (2021) Startling new variant raises urgent questions. *Science* **374**, 1178–1180 <https://doi.org/10.1126/science.acx9737>
- 156 Currin, A., Swainston, N., Day, P.J. and Kell, D.B. (2015) Synthetic biology for the directed evolution of protein biocatalysts: navigating sequence space intelligently. *Chem. Soc. Rev.* **44**, 1172–1239 <https://doi.org/10.1039/C4CS00351A>
- 157 Townsend, L., Fogarty, H., Dyer, A., Martin-Loeches, I., Bannan, C., Nadarajan, P. et al. (2021) Prolonged elevation of D-dimer levels in convalescent COVID-19 patients is independent of the acute phase response. *J. Thromb. Haemost.* **19**, 1064–1070 <https://doi.org/10.1111/jth.15267>
- 158 Perry, D.J. (2021) A pratial guide to haemostasis
- 159 Weil, I.A., Seicean, S., Neuhauser, D., Schiltz, N.K. and Seicean, A. (2015) Use and utility of hemostatic screening in adults undergoing elective, non-cardiac surgery. *PLoS ONE* **10**, e0139139 <https://doi.org/10.1371/journal.pone.0139139>

- 160 Loizou, E., Mayhew, D.J., Martlew, V. and Murthy, B.V.S. (2018) Implications of deranged activated partial thromboplastin time for anaesthesia and surgery. *Anaesthesia* **73**, 1557–1563 <https://doi.org/10.1111/anae.14344>
- 161 Chitlur, M., Sorensen, B., Rivard, G.E., Young, G., Ingerslev, J., Othman, M. et al. (2011) Standardization of thromboelastography: a report from the TEG-ROTEM working group. *Haemophilia* **17**, 532–537 <https://doi.org/10.1111/j.1365-2516.2010.02451.x>
- 162 Nielsen, V.G. and Pretorius, E. (2014) Iron-enhanced coagulation is attenuated by chelation: thromboelastographic and ultrastructural analysis. *Blood Coagul. Fibrinol.* **25**, 845–850 <https://doi.org/10.1097/MBC.0000000000000160>
- 163 de Villiers, S., Bester, J., Kell, D.B. and Pretorius, E. (2019) Erythrocyte health and the possible role of amyloidogenic blood clotting in the evolving haemodynamics of female migraine-with-aura pathophysiology: results from a pilot study. *Front. Neurol.* **10**, 1262 <https://doi.org/10.3389/fneur.2019.01262>
- 164 Kell, D.B. and Pretorius, E. (2015) On the translocation of bacteria and their lipopolysaccharides between blood and peripheral locations in chronic, inflammatory diseases: the central roles of LPS and LPS-induced cell death. *Integr. Biol.* **7**, 1339–1377 <https://doi.org/10.1039/c5ib00158g>
- 165 Erdoes, G., Schloer, H., Eberle, B. and Nagler, M. (2018) Next generation viscoelasticity assays in cardiothoracic surgery: feasibility of the TEG6s system. *PLoS ONE* **13**, e0209360 <https://doi.org/10.1371/journal.pone.0209360>
- 166 Da Luz, L.T., Nascimento, B., Shankarakutty, A.K., Rizoli, S. and Adhikari, N.K. (2014) Effect of thromboelastography (TEG(R)) and rotational thromboelastometry (ROTEM(R)) on diagnosis of coagulopathy, transfusion guidance and mortality in trauma: descriptive systematic review. *Crit. Care* **18**, 518 <https://doi.org/10.1186/s13054-014-0518-9>
- 167 Franz, R.C. (2009) ROTEM analysis: a significant advance in the field of rotational thromboelastography. *S. Afr. J. Surg.* **47**, 2–6
- 168 Jackson, G.N.B., Ashpole, K.J. and Yentis, S.M. (2009) The TEG(R) vs the ROTEM(R) thromboelastography/thromboelastometry systems. *Anaesthesia* **64**, 212–215 <https://doi.org/10.1111/j.1365-2044.2008.05752.x>
- 169 Lang, T., Bauters, A., Braun, S.L., Potzsch, B., von Pape, K.W., Kolde, H.J. et al. (2005) Multi-centre investigation on reference ranges for ROTEM thromboelastometry. *Blood Coagul. Fibrinol.* **16**, 301–310 <https://doi.org/10.1097/01.mbc.0000169225.31173.19>
- 170 Nielsen, V.G. (2007) A comparison of the Thromboelastograph and the ROTEM. *Blood Coagul. Fibrinol.* **18**, 247–252 <https://doi.org/10.1097/MBC.0b013e328092ee05>
- 171 Tanaka, K.A., Bolliger, D., Vadlamudi, R. and Nimmo, A. (2012) Rotational thromboelastometry (ROTEM)-based coagulation management in cardiac surgery and major trauma. *J. Cardiothorac. Vasc. Anesth.* **26**, 1083–1093 <https://doi.org/10.1053/j.jvca.2012.06.015>
- 172 Theusinger, O.M., Schroder, C.M., Eismont, J., Emmert, M.Y., Seifert, B., Spahn, D.R. et al. (2013) The influence of laboratory coagulation tests and clotting factor levels on Rotation Thromboelastometry (ROTEM(R)) during major surgery with hemorrhage. *Anesth. Analg.* **117**, 314–321 <https://doi.org/10.1213/ANE.0b013e31829569ac>
- 173 Wikkelsø, A., Wetterslev, J., Møller, A.M. and Afshari, A. (2016) Thromboelastography (TEG) or thromboelastometry (ROTEM) to monitor haemostatic treatment versus usual care in adults or children with bleeding. *Cochrane Database Syst. Rev.* **2016**, CD007871 <https://doi.org/10.1002/14651858.CD007871.pub3>
- 174 Smith, J.F., Knowles, T.P., Dobson, C.M., Macphree, C.E. and Welland, M.E. (2006) Characterization of the nanoscale properties of individual amyloid fibrils. *Proc. Natl Acad. Sci. U.S.A.* **103**, 15806–15811 <https://doi.org/10.1073/pnas.0604035103>
- 175 Knowles, T.P., Fitzpatrick, A.W., Meehan, S., Mott, H.R., Vendruscolo, M., Dobson, C.M. et al. (2007) Role of intermolecular forces in defining material properties of protein nanofibrils. *Science* **318**, 1900–1903 <https://doi.org/10.1126/science.1150057>
- 176 Paul, T.J., Hoffmann, Z., Wang, C., Shanmugasundaram, M., DeJonnas, J., Shekhtman, A. et al. (2016) Structural and mechanical properties of amyloid beta fibrils: a combined experimental and theoretical approach. *J. Phys. Chem. Lett.* **7**, 2758–2764 <https://doi.org/10.1021/acs.jpclett.6b01066>
- 177 Lamour, G., Nassar, R., Chan, P.H.W., Bozkurt, G., Li, J., Bui, J.M. et al. (2017) Mapping the broad structural and mechanical properties of amyloid fibrils. *Biophys. J.* **112**, 584–594 <https://doi.org/10.1016/j.bpj.2016.12.036>
- 178 Yoon, G., Lee, M., Kim, K., Kim, J.L., Chang, H.J., Baek, I. et al. (2015) Morphology and mechanical properties of multi-stranded amyloid fibrils probed by atomistic and coarse-grained simulations. *Phys. Biol.* **12**, 066021 <https://doi.org/10.1088/1478-3975/12/6/066021>
- 179 Bradford Hill, A. (1965) Environment and disease: association or causation? *Proc. R. Soc. Med.* **58**, 295–300
- 180 Fanaroff, A.C., Garcia, S. and Giri, J. (2021) Myocardial infarction during the COVID-19 pandemic. *JAMA* **326**, 1916–1918 <https://doi.org/10.1001/jama.2021.19608>
- 181 Katsoularis, I., Fonseca-Rodríguez, O., Farrington, P., Lindmark, K. and Fors Connolly, A.M. (2021) Risk of acute myocardial infarction and ischaemic stroke following COVID-19 in Sweden: a self-controlled case series and matched cohort study. *Lancet* **398**, 599–607 [https://doi.org/10.1016/S0140-6736\(21\)00896-5](https://doi.org/10.1016/S0140-6736(21)00896-5)
- 182 Ntaios, G., Michel, P., Georgiopoulos, G., Guo, Y., Li, W., Xiong, J. et al. (2020) Characteristics and outcomes in patients With COVID-19 and acute ischemic stroke: the global COVID-19 stroke registry. *Stroke* **51**, e254–e258 <https://doi.org/10.1161/STROKEAHA.119.026907>
- 183 Perry, R.J., Smith, C.J., Roffe, C., Simister, R., Narayanamoorthi, S., Marigold, R. et al. (2021) Characteristics and outcomes of COVID-19 associated stroke: a UK multicentre case-control study. *J. Neurol. Neurosurg. Psychiatry* **92**, 242–248 <https://doi.org/10.1136/jnnp-2020-324927>
- 184 Bowe, B., Xie, Y., Xu, E. and Al-Aly, Z. (2021) Kidney outcomes in long COVID. *J. Am. Soc. Nephrol.* **32**, 2851–2862 <https://doi.org/10.1681/ASN.2021060734>
- 185 Yende, S. and Parikh, C.R. (2021) Long COVID and kidney disease. *Nat. Rev. Nephrol.* **17**, 792–793 <https://doi.org/10.1038/s41581-021-00487-3>
- 186 Raj, S.R., Arnold, A.C., Barboi, A., Claydon, V.E., Limberg, J.K., Lucci, V.M. et al. (2021) Long-COVID postural tachycardia syndrome: an American autonomic society statement. *Clin. Auton. Res.* **31**, 365–368 <https://doi.org/10.1007/s10286-021-00798-2>
- 187 Agergaard, J., Leth, S., Pedersen, T.H., Harbo, T., Blicher, J.U., Karlsson, P. et al. (2021) Myopathic changes in patients with long-term fatigue after COVID-19. *Clin. Neurophysiol.* **132**, 1974–1981 <https://doi.org/10.1016/j.clinph.2021.04.009>
- 188 Soares, M.N., Eggelbusch, M., Naddaf, E., Gerrits, K.H.L., van der Schaaf, M., van den Borst, B. et al. (2021) Skeletal muscle alterations in patients with acute COVID-19 and post-acute sequelae of COVID-19. *J. Cachexia Sarcopenia Muscle* **13**, 11–22 <https://doi.org/10.1002/jcsm.12896>
- 189 Huang, C., Huang, L., Wang, Y., Li, X., Ren, L., Gu, X. et al. (2021) 6-month consequences of COVID-19 in patients discharged from hospital: a cohort study. *Lancet* **397**, 220–232 [https://doi.org/10.1016/S0140-6736\(20\)32656-8](https://doi.org/10.1016/S0140-6736(20)32656-8)



- 190 Vassiliou, A.G., Jahaj, E., Ilias, I., Markaki, V., Malachias, S., Vrettou, C. et al. (2020) Lactate kinetics reflect organ dysfunction and are associated with adverse outcomes in intensive care unit patients with COVID-19 pneumonia: preliminary results from a GREEK single-Centre study. *Metabolites* **10**, 386 <https://doi.org/10.3390/metabo10100386>
- 191 Nechipurenko, Y.D., Semyonov, D.A., Lavrinenko, I.A., Lagutkin, D.A., Generalov, E.A., Zaitceva, A.Y. et al. (2021) The role of acidosis in the pathogenesis of severe forms of COVID-19. *Biology (Basel)* **10**, 852 <https://doi.org/10.3390/biology10090852>
- 192 Roberts, I., Wright Muelas, M., Taylor, J.M., Davison, A.S., Xu, Y., Grixti, J.M. et al. (2022) Untargeted metabolomics of COVID-19 patient serum reveals potential prognostic markers of both severity and outcome. *Metabolomics* **18**, 6 <https://doi.org/10.1007/s11306-021-01859-3>
- 193 Robbins, T., Gonevski, M., Clark, C., Baitule, S., Sharma, K., Magar, A. et al. (2021) Hyperbaric oxygen therapy for the treatment of long COVID: early evaluation of a highly promising intervention. *Clin. Med. (Lond)* **21**, e629–e632 <https://doi.org/10.7861/clinmed.2021-0462>
- 194 Chen, G.F., Xu, T.H., Yan, Y., Zhou, Y.R., Jiang, Y., Melcher, K. et al. (2017) Amyloid beta: structure, biology and structure-based therapeutic development. *Acta Pharmacol. Sin.* **38**, 1205–1235 <https://doi.org/10.1038/aps.2017.28>
- 195 Drabik, D., Chodaczek, G. and Kraszewski, S. (2020) Effect of amyloid-beta monomers on lipid membrane mechanical parameters-Potential implications for mechanically driven neurodegeneration in Alzheimer's disease. *Int. J. Mol. Sci.* **22**, 18 <https://doi.org/10.3390/ijms22010018>
- 196 Milanesi, L., Sheynis, T., Xue, W.F., Orlova, E.V., Hellewell, A.L., Jelinek, R. et al. (2012) Direct three-dimensional visualization of membrane disruption by amyloid fibrils. *Proc. Natl Acad. Sci. U.S.A.* **109**, 20455–20460 <https://doi.org/10.1073/pnas.1206325109>
- 197 Fusco, G., Chen, S.W., Williamson, P.T.F., Cascella, R., Perni, M., Jarvis, J.A. et al. (2017) Structural basis of membrane disruption and cellular toxicity by alpha-synuclein oligomers. *Science* **358**, 1440–1443 <https://doi.org/10.1126/science.aan6160>
- 198 Scheckel, C. and Aguzzi, A. (2018) Prions, prionoids and protein misfolding disorders. *Nat. Rev. Genet.* **19**, 405–418 <https://doi.org/10.1038/s41576-018-0011-4>
- 199 Kell, D.B. and Knowles, J.D. (2006) The role of modeling in systems biology. In *System Modeling in Cellular Biology: From Concepts to Nuts and Bolts* (Szallasi, Z., Stelling, J. and Periwé, V., eds) pp. 3–18, MIT Press, Cambridge
- 200 Kell, D.B. and Pretorius, E. (2018) No effects without causes. The iron dysregulation and dormant microbes hypothesis for chronic, inflammatory diseases. *Biol. Rev.* **93**, 1518–1557 <https://doi.org/10.1111/brv.12407>
- 201 Stein, S.R., Ramelli, S.C., Grazioli, A., Chung, J.-Y., Singh, M., Yinda, C.K. et al. (2021) SARS-CoV-2 infection and persistence throughout the human body and brain. *Res. Square* <https://doi.org/10.21203/rs.3.rs-1139035/v1>
- 202 Patterson, B.K., Francisco, E.B., Yogendra, R., Long, E., Pise, A., Rodrigues, H. et al. (2022) Persistence of SARS CoV-2 S1 protein in CD16+ monocytes in post-Acute sequelae of COVID-19 (PASC) Up to 15 months post-Infection. *Front. Immunol.* **12**, 746021 <https://doi.org/10.3389/fimmu.2021.746021>
- 203 Ebringer, A., Rashid, T. and Wilson, C. (2010) Rheumatoid arthritis, proteus, anti-CCP antibodies and karl popper. *Autoimmun. Rev.* **9**, 216–223 <https://doi.org/10.1016/j.autrev.2009.10.006>
- 204 Ebringer, A. (2012) *Rheumatoid Arthritis and Proteus*, Springer, London
- 205 Leonardi, A.J. and Proenca, R.B. (2020) Akt-Fas to quell aberrant T cell differentiation and apoptosis in COVID-19. *Front. Immunol.* **11**, 600405 <https://doi.org/10.3389/fimmu.2020.600405>
- 206 Woodruff, M.C., Ramonell, R.P., Lee, F.E.-H. and Sanz, I. (2020) Clinically identifiable autoreactivity is common in severe SARS-CoV-2 infection. *medRxiv* <https://doi.org/10.1101/2020.10.21.20216192>
- 207 Phetsouphanh, C., Darley, D.R., Wilson, D.B., Howe, A., Munier, C.M.L., Patel, S.K. et al. (2022) Immunological dysfunction persists for 8 months following initial mild-to-moderate SARS-CoV-2 infection. *Nat. Immunol.* **23**, 210–216 <https://doi.org/10.1038/s41590-021-01113-x>
- 208 Guntas, G., Mansell, T.J., Kim, J.R. and Ostermeier, M. (2005) Directed evolution of protein switches and their application to the creation of ligand-binding proteins. *Proc. Natl Acad. Sci. U.S.A.* **102**, 11224–11229 <https://doi.org/10.1073/pnas.0502673102>
- 209 Vizcaychipi, M.P., Shovlin, C.L., McCarthy, A., Howard, A., Brown, A., Hayes, M. et al. (2020) Development and implementation of a COVID-19 near real-time traffic light system in an acute hospital setting. *Emerg. Med. J.* **37**, 630–636 <https://doi.org/10.1136/emered-2020-210199>
- 210 Stewart, S., Newton, L., Briggs, T.A., Grammatopoulos, D., Young, L. and Gill, P. (2021) Long COVID risk - a signal to address sex hormones and women's health. *Lancet Reg. Health Eur.* **11**, 100242 <https://doi.org/10.1016/j.lanep.2021.100242>
- 211 Swanepoel, A.C., Lindeque, B.G., Swart, P.J., Abdool, Z. and Pretorius, E. (2014) Ultrastructural changes of fibrin networks during three phases of pregnancy: a qualitative investigation. *Microsc. Res. Tech.* **77**, 602–608 <https://doi.org/10.1002/jemt.22385>
- 212 Pipe, S.W., Kaczmarek, R., Srivastava, A., Pierce, G.F., Makris, M. and Hermans, C. and Interim Guidance Coagulation Products Safety Supply Access Committee of the World Federation of H. (2021) Management of COVID-19-associated coagulopathy in persons with haemophilia. *Haemophilia* **27**, 41–48 <https://doi.org/10.1111/hae.14191>
- 213 Pretorius, E., Lipinski, B., Bester, J., Vermeulen, N. and Soma, P. (2013) Albumin stabilizes fibrin fiber ultrastructure in low serum albumin type 2 diabetes. *Ultrastruct. Pathol.* **37**, 254–257 <https://doi.org/10.3109/01913123.2013.778929>
- 214 Pretorius, E., Mbotwe, S. and Kell, D.B. (2017) Lipopolysaccharide-binding protein (LBP) reverses the amyloid state of fibrin seen in plasma of type 2 diabetics with cardiovascular comorbidities. *Sci. Rep.* **7**, 9680 <https://doi.org/10.1038/s41598-017-09860-4>
- 215 Cooper, G.J.S., Leighton, B., Dimitriadis, G.D., Parrybillings, M., Kowalchuk, J.M., Howland, K. et al. (1988) Amylin found in amyloid deposits in human type-2 diabetes-Mellitus May Be a hormone that regulates glycogen-Metabolism in skeletal-Muscle. *Proc. Natl Acad. Sci. U.S.A.* **85**, 7763–7766 <https://doi.org/10.1073/pnas.85.20.7763>
- 216 Roberts, A.N., Leighton, B., Todd, J.A., Cockburn, D., Schofield, P.N., Sutton, R. et al. (1989) Molecular and functional characterization of amylin, a peptide associated with type 2 diabetes mellitus. *Proc. Natl Acad. Sci. U.S.A.* **86**, 9662–9666 <https://doi.org/10.1073/pnas.86.24.9662>
- 217 Konarkowska, B., Aitken, J.F., Kistler, J., Zhang, S. and Cooper, G.J.S. (2006) The aggregation potential of human amylin determines its cytotoxicity towards islet beta-cells. *FEBS J.* **273**, 3614–3624 <https://doi.org/10.1111/j.1742-4658.2006.05367.x>
- 218 Seet, R.C.S., Quek, A.M.L. and Lim, E.C.H. (2007) Post-infectious fatigue syndrome in dengue infection. *J. Clin. Virol.* **38**, 1–6 <https://doi.org/10.1016/j.jcv.2006.10.011>
- 219 Hung, T.M., Wills, B., Clapham, H.E., Yacoub, S. and Turner, H.C. (2019) The uncertainty surrounding the burden of post-acute consequences of dengue infection. *Trends Parasitol.* **35**, 673–676 <https://doi.org/10.1016/j.pt.2019.06.004>

- 220 Umakanth, M. (2018) Post dengue fatigue syndrome (PDFS) among dengue IgM-Antibody positive patients at batticaloa teaching hospital, Sri Lanka. *Open Acc. Lib. J.* **5**, e4798 <https://doi.org/10.4236/oalib.1104798>
- 221 Bond, N.G., Grant, D.S., Himmelfarb, S.T., Engel, E.J., Al-Hasan, F., Gbokie, M. et al. (2021) Post-Ebola syndrome presents With multiple overlapping symptom clusters: evidence from an ongoing cohort study in eastern Sierra Leone. *Clin. Infect. Dis.* **73**, 1046–1054 <https://doi.org/10.1093/cid/ciab267>
- 222 Burki, T.K. (2016) Post-Ebola syndrome. *Lancet Infect. Dis.* **16**, 780–781 [https://doi.org/10.1016/S1473-3099\(15\)00259-5](https://doi.org/10.1016/S1473-3099(15)00259-5)
- 223 Scott, J.T., Sesay, F.R., Massaquoi, T.A., Idriss, B.R., Sahr, F. and Semple, M.G. (2016) Post-Ebola syndrome, Sierra Leone. *Emerg. Infect. Dis.* **22**, 641–646 <https://doi.org/10.3201/eid2204.151302>
- 224 Rebman, A.W. and Aucott, J.N. (2020) Post-treatment lyme disease as a model for persistent symptoms in lyme disease. *Front. Med. (Lausanne)* **7**, 57 <https://doi.org/10.3389/fmed.2020.00057>
- 225 Potgieter, M., Bester, J., Kell, D.B. and Pretorius, E. (2015) The dormant blood microbiome in chronic, inflammatory diseases. *FEMS Microbiol. Rev.* **39**, 567–591 <https://doi.org/10.1093/femsre/fuv013>
- 226 Bots, M.L., Breteler, M.M., van Kooten, F., Haverkate, F., Meijer, P., Koudstaal, P.J. et al. (1998) Coagulation and fibrinolysis markers and risk of dementia. The Dutch vascular factors in dementia study. *Haemostasis* **28**, 216–222
- 227 Cortes-Canteli, M., Paul, J., Norris, E.H., Bronstein, R., Ahn, H.J., Zamolodchikov, D. et al. (2010) Fibrinogen and beta-amyloid association alters thrombosis and fibrinolysis: a possible contributing factor to Alzheimer's disease. *Neuron* **66**, 695–709 <https://doi.org/10.1016/j.neuron.2010.05.014>
- 228 Wong, T.L. and Weitzer, D.J. (2021) Long COVID and myalgic encephalomyelitis/Chronic fatigue syndrome (ME/CFS)-A systemic review and comparison of clinical presentation and symptomatology. *Medicina (Kaunas)* **57**, 418 <https://doi.org/10.3390/medicina57050418>
- 229 Berg, D., Berg, L.H., Couvaras, J. and Harrison, H. (1999) Chronic fatigue syndrome and/or fibromyalgia as a variation of antiphospholipid antibody syndrome: an explanatory model and approach to laboratory diagnosis. *Blood Coagul. Fibrinol.* **10**, 435–438 <https://doi.org/10.1097/00001721-199910000-00006>
- 230 Hannan, K.L., Berg, D.E., Baumzweiger, W., Harrison, H.H., Berg, L.H., Ramirez, R. et al. (2000) Activation of the coagulation system in gulf War illness: a potential pathophysiologic link with chronic fatigue syndrome. A laboratory approach to diagnosis. *Blood Coagul. Fibrinol.* **11**, 673–678 <https://doi.org/10.1097/00001721-200010000-00013>
- 231 Pretorius, E., Venter, C., Laubsher, G.J., Kotze, M.J., Moremi, K., Oladejo, S. et al. (2021) Combined triple treatment of fibrin amyloid microclots and platelet pathology in individuals with long COVID/Post-Acute sequelae of COVID-19 (PASC) can resolve their persistent symptoms. *Res. Square* <https://doi.org/10.21203/rs.3.rs-1205453/v1>
- 232 Barrett, T.J., Cornwell, M., Myndzar, K., Rolling, C.C., Xia, Y., Drenkova, K. et al. (2021) Platelets amplify endotheliopathy in COVID-19. *Sci. Adv.* **7**, eabh2434 <https://doi.org/10.1126/sciadv.abh2434>
- 233 Barrett, T.J., Bilaloglu, S., Cornwell, M., Burgess, H.M., Virginio, V.W., Drenkova, K. et al. (2021) Platelets contribute to disease severity in COVID-19. *J. Thromb. Haemost.* **19**, 3139–3153 <https://doi.org/10.1111/jth.15534>
- 234 Nishikawa, M., Kanno, H., Zhou, Y., Xiao, T.H., Suzuki, T., Ibayashi, Y. et al. (2021) Massive image-based single-cell profiling reveals high levels of circulating platelet aggregates in patients with COVID-19. *Nat. Commun.* **12**, 7135 <https://doi.org/10.1038/s41467-021-27378-2>
- 235 Zhang, S., Liu, Y., Wang, X., Yang, L., Li, H., Wang, Y. et al. (2020) SARS-CoV-2 binds platelet ACE2 to enhance thrombosis in COVID-19. *J. Hematol. Oncol.* **13**, 120 <https://doi.org/10.1186/s13045-020-00954-7>
- 236 Chang, J.C. (2018) Disseminated intravascular coagulation: is it fact or fancy? *Blood Coagul. Fibrinolysis* **29**, 330–337 <https://doi.org/10.1097/MBC.0000000000000727>
- 237 Perricone, C., Bartoloni, E., Bursi, R., Cafaro, G., Guidelli, G.M., Shoenfeld, Y. et al. (2020) COVID-19 as part of the hyperferritinemic syndromes: the role of iron depletion therapy. *Immunol. Res.* **68**, 213–224 <https://doi.org/10.1007/s12026-020-09145-5>
- 238 Vargas-Vargas, M. and Cortés-Rojo, C. (2020) Ferritin levels and COVID-19. *Rev. Panam. Salud. Publica* **44**, e72 <https://doi.org/10.26633/RPSP.2020.72>
- 239 Kell, D.B. and Pretorius, E. (2014) Serum ferritin is an important disease marker, and is mainly a leakage product from damaged cells. *Metallomics* **6**, 748–773 <https://doi.org/10.1039/C3MT00347G>
- 240 Kell, D.B. (2009) Iron behaving badly: inappropriate iron chelation as a major contributor to the aetiology of vascular and other progressive inflammatory and degenerative diseases. *BMC Med. Genom.* **2**, 2 <https://doi.org/10.1186/1755-8794-2-2>
- 241 Kell, D.B. (2010) Towards a unifying, systems biology understanding of large-scale cellular death and destruction caused by poorly liganded iron: Parkinson's, huntington's, Alzheimer's, prions, bactericides, chemical toxicology and others as examples. *Arch. Toxicol.* **577**, 825–889 <https://doi.org/10.1007/s00204-010-0577-x>
- 242 Cavezzi, A., Troiani, E. and Corrao, S. (2020) COVID-19: hemoglobin, iron, and hypoxia beyond inflammation. A narrative review. *Clin. Pract.* **10**, 1271 <https://doi.org/10.4081/cp.2020.1271>
- 243 Edeas, M., Saleh, J. and Peyssonnaud, C. (2020) Iron: innocent bystander or vicious culprit in COVID-19 pathogenesis? *Int. J. Infect. Dis.* **97**, 303–305 <https://doi.org/10.1016/j.ijid.2020.05.110>
- 244 McLaughlin, K.-M., Bechtel, M., Bojkova, D., Münch, C., Ciesek, S., Wass, M.N. et al. (2020) COVID-19-Related coagulopathy—Is transferrin a missing link? *Diagnostics* **10**, 539 <https://doi.org/10.3390/diagnostics10080539>
- 245 Chakurkar, V., Rajapurkar, M., Lele, S., Mukhopadhyay, B., Lobo, V., Injarapu, R. et al. (2021) Increased serum catalytic iron may mediate tissue injury and death in patients with COVID-19. *Sci. Rep.* **11**, 19618 <https://doi.org/10.1038/s41598-021-99142-x>
- 246 Liu, W. and Li, H. (2022) COVID-19: attacks the 1-beta chain of hemoglobin to disrupt respiratory function and escape immunity. *chemRxiv* <https://doi.org/10.26434/chemrxiv-2021-dtpv3-v>
- 247 Campione, E., Cosio, T., Rosa, L., Lanna, C., Di Girolamo, S., Gaziano, R. et al. (2020) Lactoferrin as protective natural barrier of respiratory and intestinal mucosa against coronavirus infection and inflammation. *Int. J. Mol. Sci.* **21**, 4903 <https://doi.org/10.3390/ijms21144903>
- 248 Campione, E., Lanna, C., Cosio, T., Rosa, L., Conte, M.P., Iacovelli, F. et al. (2020) Pleiotropic effect of Lactoferrin in the prevention and treatment of COVID-19 infection: *in vivo*, *in silico* and *in vitro* preliminary evidences. *bioRxiv* 2020.08.11.244996 <https://doi.org/10.1101/2020.08.11.244996>
- 249 Dalamaga, M., Karampela, I. and Mantzoros, C.S. (2020) Commentary: could iron chelators prove to be useful as an adjunct to COVID-19 treatment regimens? *Metabolism* **108**, 154260 <https://doi.org/10.1016/j.metabol.2020.154260>

- 250 Kell, D.B., Heyden, E.L. and Pretorius, E. (2020) The biology of lactoferrin, an iron-binding protein that can help defend against viruses and bacteria. *Front. Immunol.* **11**, 1221 <https://doi.org/10.3389/fimmu.2020.01221>
- 251 Abobaker, A. (2020) Can iron chelation as an adjunct treatment of COVID-19 improve the clinical outcome? *Eur. J. Clin. Pharmacol.* **76**, 1619–1620 <https://doi.org/10.1007/s00228-020-02942-9>
- 252 Carota, G., Ronsisvalle, S., Panarello, F., Tibullo, D., Nicolosi, A. and Li Volti, G. (2021) Role of iron chelation and protease inhibition of natural products on COVID-19 infection. *J. Clin. Med.* **10**, 2306 <https://doi.org/10.3390/jcm10112306>
- 253 Naseef, P.P., Elayadeth-Meethal, M., Salim, K.T.M., Anjana, A., Muhas, C., Vajid, K.A. et al. (2022) Therapeutic potential of induced iron depletion using iron chelators in COVID-19. *Saudi J. Biol. Sci.* <https://doi.org/10.1016/j.sjbs.2021.11.061>
- 254 Vlahakos, V.D., Marathias, K.P., Arkadopoulos, N. and Vlahakos, D.V. (2021) Hyperferritinemia in patients with COVID-19: an opportunity for iron chelation? *Artif. Organs.* **45**, 163–167 <https://doi.org/10.1111/aor.13812>
- 255 Fogarty, H., Townsend, L., Morrin, H., Ahmad, A., Comerford, C., Karampini, E. et al. (2021) Persistent endotheliopathy in the pathogenesis of long COVID syndrome. *J. Thromb. Haemost.* **19**, 2546–2553 <https://doi.org/10.1111/jth.15490>
- 256 Goshua, G., Pine, A.B., Meizlish, M.L., Chang, C.H., Zhang, H., Bahel, P. et al. (2020) Endotheliopathy in COVID-19-associated coagulopathy: evidence from a single-centre, cross-sectional study. *Lancet Haematol.* **7**, e575–e582 [https://doi.org/10.1016/S2352-3026\(20\)30216-7](https://doi.org/10.1016/S2352-3026(20)30216-7)
- 257 Showlin, C.L. and Vizcaychipi, M.P. (2020) Vascular inflammation and endothelial injury in SARS-CoV-2 infection: the overlooked regulatory cascades implicated by the ACE2 gene cluster. *QJM* <https://doi.org/10.1093/qjmed/hcaa241>
- 258 Gremmel, T., Frelinger, III, A.L. and Michelson, A.D. (2016) Platelet physiology. *Semin Thromb Hemost.* **42**, 191–204 <https://doi.org/10.1055/s-0035-1564835>
- 259 Page, M.J. and Pretorius, E. (2020) A champion of host defense: a generic large-Scale cause for platelet dysfunction and depletion in infection. *Semin. Thromb. Hemost.* **46**, 302–319 <https://doi.org/10.1055/s-0040-1708827>
- 260 Rubenstein, D.A. and Yin, W. (2018) Platelet-Activation mechanisms and vascular remodeling. *Compr. Physiol.* **8**, 1117–1156 <https://doi.org/10.1002/cphy.c170049>
- 261 Wang, J., Hajizadeh, N., Moore, E.E., McIntyre, R.C., Moore, P.K., Veress, L.A. et al. (2020) Tissue plasminogen activator (tPA) treatment for COVID-19 associated acute respiratory distress syndrome (ARDS): a case series. *J. Thromb. Haemost.* **18**, 1752–1755 <https://doi.org/10.1111/jth.14828>
- 262 ATTACC Investigators, ACTIV-4a Investigators, REMAP-CAP Investigators, Lawler, P.R., Goligher, E.C., Berger, J.S. et al. (2021) Therapeutic anticoagulation with heparin in noncritically ill patients with COVID-19. *N. Engl. J. Med.* **385**, 790–802 <https://doi.org/10.1056/NEJMoa2105911>
- 263 Mycroft-West, C.J., Su, D., Pagani, I., Rudd, T.R., Elli, S., Gandhi, N.S. et al. (2020) Heparin inhibits cellular invasion by SARS-CoV-2: structural dependence of the interaction of the spike S1 receptor-Binding domain with heparin. *Thromb. Haemost.* **120**, 1700–1715 <https://doi.org/10.1055/s-0040-1721319>
- 264 Tandon, R., Sharp, J.S., Zhang, F., Pomin, V.H., Ashpole, N.M., Mitra, D. et al. (2021) Effective inhibition of SARS-CoV-2 entry by heparin and enoxaparin derivatives. *J. Virol.* **95**, e01987-20 <https://doi.org/10.1128/JVI.01987-20>
- 265 Tree, J.A., Turnbull, J.E., Buttigieg, K.R., Elmore, M.J., Coombes, N., Hogwood, J. et al. (2021) Unfractionated heparin inhibits live wild type SARS-CoV-2 cell infectivity at therapeutically relevant concentrations. *Br. J. Pharmacol.* **178**, 626–635 <https://doi.org/10.1111/bph.15304>
- 266 Paiardi, G., Richter, S., Oreste, P., Urbinati, C., Rusnati, M. and Wade, R.C. (2022) The binding of heparin to spike glycoprotein inhibits SARS-CoV-2 infection by three mechanisms. *J. Biol. Chem.* **298**, 101507 <https://doi.org/10.1016/j.jbc.2021.101507>
- 267 Lopes, R.D., de Barros, E.S.P.G.M., Furtado, R.H.M., Macedo, A.V.S., Bronhara, B., Damiani, L.P. et al. (2021) Therapeutic versus prophylactic anticoagulation for patients admitted to hospital with COVID-19 and elevated D-dimer concentration (ACTION): an open-label, multicentre, randomised, controlled trial. *Lancet* **397**, 2253–2263 [https://doi.org/10.1016/S0140-6736\(21\)01203-4](https://doi.org/10.1016/S0140-6736(21)01203-4)
- 268 Viecca, M., Radovanovic, D., Forleo, G.B. and Santus, P. (2020) Enhanced platelet inhibition treatment improves hypoxemia in patients with severe COVID-19 and hypercoagulability. A case control, proof of concept study. *Pharmacol. Res.* **158**, 104950 <https://doi.org/10.1016/j.phrs.2020.104950>
- 269 van der Togt, V., McFarland, S., Esperti, M., Semper, E., Citrenbaum, S. and Rossman, J.S. (2021) Promotion of non-evidence-based therapeutics within patient-led long COVID support groups. *Nat. Med.* **27**, 2068–2069 <https://doi.org/10.1038/s41591-021-01589-y>
- 270 Koziel, M., Potpara, T.S. and Lip, G.Y.H. (2020) Triple therapy in patients with atrial fibrillation and acute coronary syndrome or percutaneous coronary intervention/stenting. *Res. Pract. Thromb. Haemost.* **4**, 357–365 <https://doi.org/10.1002/rth2.12319>
- 271 Pretorius, E., Venter, C., Laubscher, G.J., Lourens, P.J., Steenkamp, J. and Kell, D.B. (2020) Prevalence of readily detected amyloid blood clots in ‘unclotted’ type 2 diabetes mellitus and COVID-19 plasma: a preliminary report. *Cardiovasc. Diabetol.* **19**, 193 <https://doi.org/10.1186/s12933-020-01165-7>
- 272 Thagard, P. (2007) Coherence, truth, and the development of scientific knowledge. *Philos. Sci.* **74**, 28–47 <https://doi.org/10.1086/520941>
- 273 Thagard, P. (2008) Explanatory Coherence. Reasoning: Studies of Human Inference and Its Foundations, 471–513
- 274 Bradford Hill, A. and Hill, I.D. (1991) *Bradford Hill's Principles of Medical Statistics*, Edward Arnold, London
- 275 Schaffner, K.F. (1993) *Discovery and Explanation in Biology and Medicine*, University of Chicago Press, Chicago