

Sequence analysis of two F1 mycobacteriophages, Deb65 and DocMcStuffins

Marcus O. Royster,¹ Victoria Figgins,¹ Vera Pande,¹ Jason D. Robinson,¹ Deeka S. Abdi,¹ Ali Amin,¹ Zephaniah Ansah,¹ Ethan W. Bomersheim,¹ Gianna Dunn,¹ Ali A. Elfaki,¹ Jordyn Foulk,¹ Kate C. Ingle,¹ Avi D. Lavu,¹ Ved Pande,¹ Priya T. Shan,¹ Marie P. Smithbey,¹ Gunnar R. Ternstrom,¹ Olivia S. Trager,¹ David A. Washington,¹ Monica Xu,¹ Margaret S. Saha¹

AUTHOR AFFILIATION See affiliation list on p. 3.

ABSTRACT Isolated from wetland soil, Deb65 and DocMcStuffins are bacteriophages with a siphoviral morphology that infect *Mycobacterium smegmatis*. Deb65 and DocMcStuffins encode 97 and 91 putative genes, 41 of which are shared. Based on gene content similarity to actinobacteriophages more broadly, both phages are assigned to subcluster F1.

KEYWORDS *Mycobacterium smegmatis*, mycobacteriophage

Advancing our knowledge of mycobacteriophage diversity is essential for understanding the abundance, community dynamics, and evolution of *Mycobacteria*, an environmentally and clinically important genus (1–5). We report the sequence of two genetically distinct mycobacteriophages, Deb65 and DocMcStuffins.

Both phages were isolated from wet, silty soil samples at the College of William & Mary in Williamsburg, VA, USA, using a standard enrichment procedure (Table 1) (6). Briefly, 5 g of each soil sample was suspended in 50 mL of 7H9 media, inoculated with *Mycobacterium smegmatis* mc² 155, and incubated in a 37°C shaker (250 rpm) for 2 days. The resulting cultures were filtered through a 0.22 µm filter, and the filtrates were plated with *M. smegmatis* in 7H9 top agar, yielding clear plaques for both phages, Deb65 and DocMcStuffins, after 24–48 h. Phages were then purified through three rounds of plating before being imaged by negative stain (1% uranyl acetate) transmission electron microscopy to reveal siphoviral morphologies for both phages (Fig. 1).

Phage DNA was extracted from a lysate using the phenol-chloroform-isoamyl alcohol method and ethanol precipitated (7). DNA was prepared for sequencing using the NEB Ultra II Library Kit and sequenced using an Illumina MiSeq Sequencer (v3 reagents, single-end, 150 base read). Newbler (version 2.9) was then used to assemble the genome and Consed (version 29) to check for completeness and reveal 3′ single-stranded genome termini (8). Sequencing data, overhangs, and genome characteristics are presented in Table 1.

The genomes were annotated using DNA Master (version 5.23.6) and PECAAN (version 20221109) (9). Translational start sites were verified using the coding potential predicted by GeneMark and Glimmer (10–12), and the similarity of start sites in homologs was identified using Starterator (<http://phages.wustl.edu/starterator/>) and BLASTp against the Actinobacteriophage and NCBI non-redundant protein databases (6, 13). No tRNAs were identified using Aragorn version 1.2.41 (14) and tRNAscan (15).

Putative gene functions were assigned based on predictions from HHPred (using the PDB_mmCIF70, NCBI_CD, SCOPe70, and pFAM-A as databases), BLASTp, and Phamerator (Actino_draft database) for highly similar genes (16, 17). Using the gene content similarity (GCS) tool at the Actinobacteriophage database, phagesDB (<https://phagesdb.org/>) and clustering parameters of at least 35% GCS to actinobacteriophages, both

Editor John J. Dennehy, Department of Biology, Queens College, New York, USA

Address correspondence to Margaret S. Saha, mssaha@wm.edu.

The authors declare no conflict of interest

See the funding table on p. 3.

Received 18 December 2024

Accepted 24 March 2025

Published 14 April 2025

Copyright © 2025 Royster et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

TABLE 1 Genome and sequencing information for DocMcStuffins and Deb65

Phage	DocMcStuffins	Deb65
Isolation GPS	37°16′08.4″N 76°43′08.4″W	37°16′15.1″N 76°42′56.7″W
Morphology	Siphovirus	Siphovirus
Capsid diameter	50 ± 3 nm (<i>n</i> = 5)	50 ± 2 nm (<i>n</i> = 5)
Tail length	~180 ± 5 nm (<i>n</i> = 7)	~190 ± 5 nm (<i>n</i> = 5)
Sequencing reads	453,180	440,517
Sequencing coverage, fold	1,122	1,119
Genome length (bp)	58,159	55,767
Genome end sequence	5′ CCGAAGGCAT	5′ CGGACGGCGC
Number of open reading frames	91	97
GC content (%)	62.7	61.6
Accession number	PQ184804	PQ184836
SRA	SRX26785850	SRX26785849

phages are assigned to cluster F, subcluster F1 (18, 19). Default settings were used for all software.

Both phages share 41 GCS, which are primarily in the first half of the genome and include genes encoding functions in virion structure, assembly, lysis, and lysogeny, the latter consistent with the temperate lifecycle for F cluster phages (Fig. 1). Noteworthy here is that while both phages encode homologous tyrosine integrases with 90% amino acid identity (AAI), their immunity repressors only share 39% AAI. Within this genomic region, and consistent with many phages of the F1 subcluster, both Deb65 and DocMcStuffins encode anti-repressor and Cro proteins. Within the second half of the genome, where gene conservation is lower, both phages encode

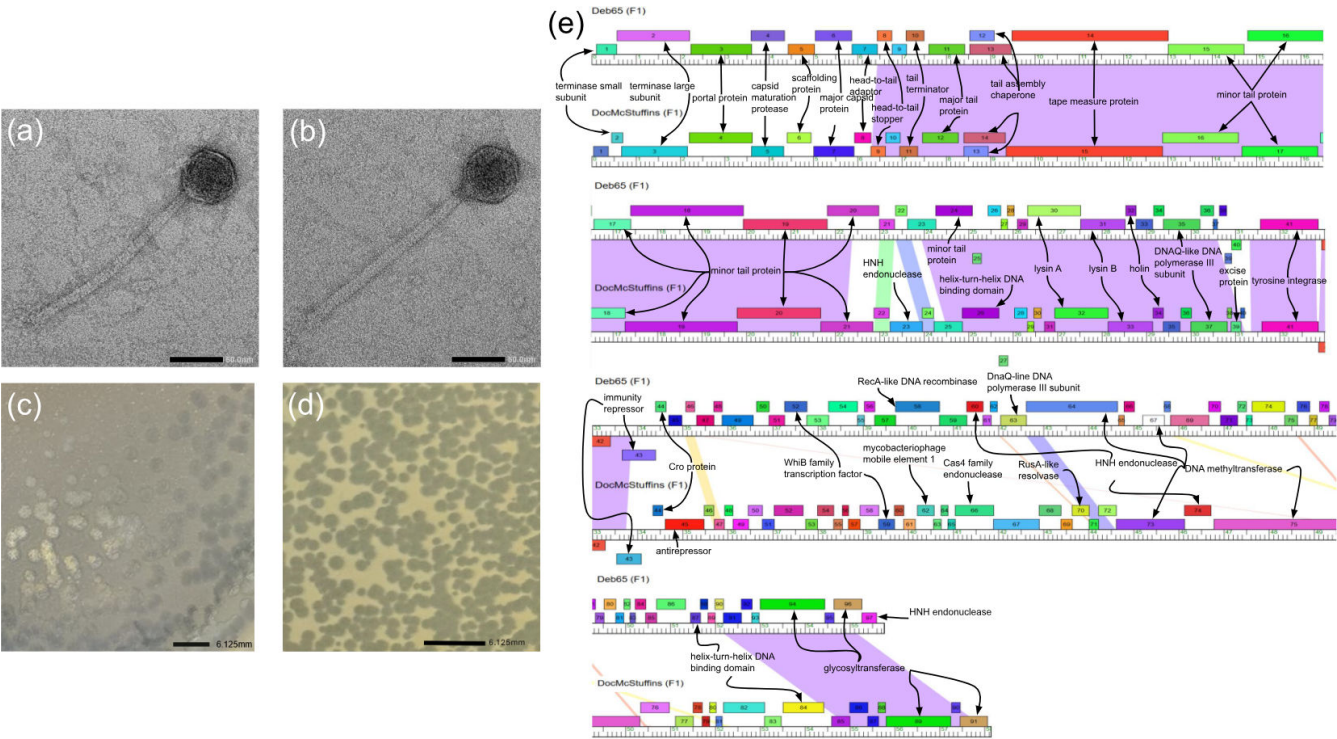


FIG 1 (a) Negative stain (uranyl acetate, 1%) transmission electron microscopy of DocMcStuffins revealing siphoviral morphology. Scale bar is 50 nm. (b) Negative stain (uranyl acetate, 1%) transmission electron microscopy of Deb65 revealing siphoviral morphology. Scale bar is 50 nm. (c) Plaque image of DocMcStuffins showing clear plaques. Scale bar is 6.125 mm. (d) Plaque image of Deb65 showing clear plaques. Scale bar is 6.125 mm. (e) Phamerator comparative alignment of Deb65 and DocMcStuffins.

two glycosyltransferases that are highly conserved across the F1 subcluster. Within this region, Deb65 encodes a unique DNA methyltransferase for which no homolog exists in the Actinobacteriophage database.

ACKNOWLEDGMENTS

We thank the University of Pittsburgh for genome sequencing, the Hatfull lab, and the entire SEA-PHAGES program for their support. We thank Old Dominion University Applied Research Center for the TEM images.

This work was supported in part by NIH grant 1R15HD096415-01 to M.S.S.

AUTHOR AFFILIATION

¹Department of Biology, William and Mary, Williamsburg, Virginia, USA

AUTHOR ORCIDs

Marcus O. Royster  <http://orcid.org/0009-0002-9577-1184>

Victoria Figgins  <http://orcid.org/0009-0008-4392-9852>

Margaret S. Saha  <http://orcid.org/0000-0003-0096-2667>

FUNDING

Funder	Grant(s)	Author(s)
HHS NIH Eunice Kennedy Shriver National Institute of Child Health and Human Development (NICHD)	1R15HD114135-01	Margaret Somosi Saha

AUTHOR CONTRIBUTIONS

Marcus O. Royster, Conceptualization, Formal analysis, Investigation, Methodology, Software, Supervision, Validation, Writing – original draft, Writing – review and editing | Victoria Figgins, Formal analysis, Investigation, Software, Supervision, Writing – original draft, Writing – review and editing | Vera Pande, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Jason D. Robinson, Investigation, Methodology, Supervision, Writing – original draft, Writing – review and editing | Ali Amin, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Zephaniah Ansah, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Ethan W. Bomersheim, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Gianna Dunn, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Ali A. Elfaki, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Jordyn Foulk, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Kate C. Ingle, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Avi D. Lavu, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Ved Pande, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Priya T. Shan, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Marie P. Smithbey, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Gunnar R. Ternstrom, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Olivia S. Trager, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | David A. Washington, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Monica Xu, Formal analysis, Investigation, Software, Writing – original draft, Writing – review and editing | Margaret S. Saha, Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Writing – original draft, Writing – review and editing.

DATA AVAILABILITY

Deb65 and DocMcStuffs are available at GenBank with accession nos. [PQ184836](#) and [PQ184804](#), and Sequence Read Archive (SRA) nos. [SRX26785849](#) and [SRX26785850](#).

REFERENCES

- Jacobs-Sera D, Marinelli LJ, Bowman C, Broussard GW, Guerrero Bustamante C, Boyle MM, Petrova ZO, Dedrick RM, Pope WH, Modlin RL, Hendrix RW, Hatfull GF. Science Education Alliance Phage Hunters Advancing Genomics And Evolutionary Science Sea-Phages Program. 2012. On the nature of mycobacteriophage diversity and host preference. Edited by R. L. Modlin, R. W. Hendrix, and G. F. Hatfull. *Virology (Auckl)* 434:187–201. <https://doi.org/10.1016/j.virol.2012.09.026>
- Esposito LA, Gupta S, Streiter F, Prasad A, Dennehy JJ. 2016. Evolutionary interpretations of mycobacteriophage biodiversity and host-range through the analysis of codon usage bias. *Microb Genom* 2:e000079. <https://doi.org/10.1099/mgen.0.000079>
- Walsh CM, Gebert MJ, Delgado-Baquerizo M, Maestre FT, Fierer N. 2019. A global survey of mycobacterial diversity in soil. *Appl Environ Microbiol* 85:e01180–19. <https://doi.org/10.1128/AEM.01180-19>
- Hatfull GF. 2022. Mycobacteriophages: from petri dish to patient. *PLoS Pathog* 18:e1010602. <https://doi.org/10.1371/journal.ppat.1010602>
- Papke RT, Doolittle WF. 2003. Phage evolution: New worlds of genomic diversity. *Curr Biol* 13:R606–R607. [https://doi.org/10.1016/S0960-9822\(03\)00527-X](https://doi.org/10.1016/S0960-9822(03)00527-X)
- Poxleitner M, Pope W, Jacobs-Sera D, Sivanathan V, Hatfull G. 2018. Phage discovery guide. Howard Hughes Medical Institute.
- Sambrook J, Russell DW. 2006. Purification of nucleic acids by extraction with phenol:chloroform. *Cold Spring Harb Protoc* 2006. <https://doi.org/10.1101/pdb.prot4455>
- Russell D. A. 2018. Sequencing, assembling, and finishing complete bacteriophage genomes. *Methods Mol Biol* 1681:109–125. https://doi.org/10.1007/978-1-4939-7343-9_9
- Pope WH, Jacobs-Sera D, Russell DA, Hatfull GF. 2017. SEA-PHAGES bioinformatics guide. Howard Hughes Medical Institute.
- Besemer J, Borodovsky M. 2005. GeneMark: web software for gene finding in prokaryotes, eukaryotes and viruses. *Nucleic Acids Res* 33:W451–454. <https://doi.org/10.1093/nar/gki487>
- Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. *Nucleic Acids Res* 29:2607–2618. <https://doi.org/10.1093/nar/29.12.2607>
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25:955–964. <https://doi.org/10.1093/nar/25.5.955>
- Zimmermann L, Stephens A, Nam SZ, Rau D, Kübler J, Lozajic M, Gabler F, Söding J, Lupas AN, Alva V. 2018. A completely reimplemented MPI bioinformatics toolkit with a new HHpred server at its core. *J Mol Biol* 430:2237–2243. <https://doi.org/10.1016/j.jmb.2017.12.007>
- Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. *BMC Bioinformatics* 12:395. <https://doi.org/10.1186/1471-2105-12-395>
- Russell DA, Hatfull GF. 2017. PhagesDB: the actinobacteriophage database. *Bioinformatics* 33:784–786. <https://doi.org/10.1093/bioinformatics/btw711>
- Pope WH, Mavrich TN, Garlena RA, Guerrero-Bustamante CA, Jacobs-Sera D, Montgomery MT, Russell DA, Warner MH, Hatfull GF, Science Education Alliance-Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES). 2017. Bacteriophages of *Gordonia* spp. display a spectrum of diversity and genetic relationships. *mBio* 8:e01069–17. <https://doi.org/10.1128/mBio.01069-17>