



Effect of Epigallocatechin Gallate on Viability of *Kudoa septempunctata*

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Abstract: *Kudoa septempunctata* have been reported as a causative agent for acute transient gastrointestinal troubles after eating raw olive flounder (*Paralichthys olivaceus*). It raised public health concerns and quarantine control in several countries. Quantitative evaluation on viability of *K. septempunctata* is crucial to develop effective chemotherapeutics against it. A cytometry using fluorescent stains was employed to assess effect of three compounds on viability of *K. septempunctata*. Epigallocatechin gallate reduced markedly viability of *K. septempunctata* at 0.5 mM or more, and damaged *K. septempunctata* spores by producing cracks.

Key words: *Kudoa septempunctata*, olive flounder, flow cytometry, epigallocatechin gallate

Myxozoa (Cnidaria) include more than 2,100 species, most of which are coelozoic or histozoic parasites in fish [1]. Although most myxozoans do not cause a serious problem in fish, several species *Enteromyxum leei*, *E. scophthalmi*, and *Myxobolus cerebralis* cause mass mortality [2-4], and *Kudoa septempunctata*, *K. iwatai*, *K. hexapunctata*, and *Umicapsula seriolae* have been reported to cause fish-born illness in Korea and Japan [5-8]. In particular, *K. septempunctata* has been reported in olive flounder *Paralichthys olivaceus*, which is the most reared fish species in Korea. This species provoked a conflict in fish trade and economic loss in fish farming [9].

Based on the differences in fluorescent stains and morphological changes of the myxozoa, several studies have been conducted to find chemotherapeutic agents that affect the viability of *K. septempunctata* [10-12]. However, the quantification and determination of accurate effective doses of chemicals based on microscopic observation can lead to inconsistency and misinterpretations, particularly due to subjective visual criteria. As an alternative, flow cytometry allows rapid, accurate, and quantitative analysis on cell morphology and viability. Flow cytometry has been applied to determine viability of the protozoans such as *Cryptosporidium parvum* and *Perkinsus marinus*

[13,14]. However, any assay on the viability of the myxozoans was performed using flow cytometry. Aim of present study is to assay on effect of three chemicals compounds to the viability of *K. septempunctata* spores using a simple and objective method of flow cytometry.

Olive flounders (n=20) were obtained from an olive flounder farm that suffered annually from *K. septempunctata* infection. Squash preparations of the fish samples were examined microscopically to detect *K. septempunctata* spores. The spores were purified by a modified Percoll (Sigma-Aldrich, St. Louis, Missouri, USA) density-gradient centrifugation method, as previously described [15]. The *K. septempunctata* spores were exposed to three kinds of chemicals including amprolium hydrochloride (AH; Dr. Ehrenstorfer, Augsburg, Germany), chlorogenic acid (CA; Carl Roth, Karlsruhe, Germany), and epigallocatechin gallate (EGCG; Sigma-Aldrich), which have been reported to be effective against the myxospores [11,12]. Fifty microliters of chemicals (1 and 10 mM) were added to 450 μ l of phosphate-buffered saline (PBS) containing 3.2×10^5 spores, and the mixtures were incubated at room temperature (approximately 20-25°C) for 30 min. The spores were then stained with 50 μ l of 20 μ g/ml propidium iodide (PI) for 10 min. The proportion of dead and live spores was calculated to estimate viability based on 3,000 spores counted by the red fluorescence detector of the flow cytometer (CytoFlex, Beckman Coulter, Brea, California, USA).

To determine the criteria of viability of *K. septempunctata*, the spores (8.5×10^3 spores/500 μ l) were exposed to PBS, 1% for-

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malin, and 0.01 and 0.1 mM final concentrations of EGCG. It determined that the PBS exposed group 100% viability while the 1% formalin exposed group has 0% viability. We investigated the effect of EGCG at concentrations, ranging from 0.01 to 1 mM. The flow cytometry assay was performed with 1.7×10^4 spores/500 μ l exposed to 0.01, 0.05, 0.1, and 1 mM final concentrations of EGCG, and PBS. For scanning electron microscopy (SEM), the purified spores were divided into 3 groups that exposed to 0.1 and 1 mM EGCG, and PBS for 30 min, respectively. The spores were immediately rinsed twice with PBS and fixed in 2% glutaraldehyde in 0.1 M phosphate buffer (pH 7.4) at room temperature for 2 hr. The spores were post-fixed in 1% osmium tetroxide in the same buffer and dehydrated through 50, 70, 90, 95, and 100% ethanol each for 10 min. The spores were dried in an atmosphere and then dried using a CO₂ critical point dryer. Finally, the spores were sputter coated with platinum, and observed using a JSM-6700F SEM at 10 kV.

Since PI is permeable only through the membrane of dead *K. septempunctata* spores, intensity of the stain was counted as dead spores. The spores exposed to 0.1 and 1 mM of CA and AM were unstained with PI since there was no damage in the membrane (Fig 1A). On the other hand, the spores treated with 0.1 and 1 mM EGCG showed strong red fluorescence, indicating that the spores lost membrane integrity and were stained with PI (Fig. 1A). The efficacy of EGCG was found at 0.1 mM, but not at 0.01 mM. The finding/efficacy was similar to that of 1% formalin (Fig. 1B). We observed that more than 90% of the spores were dead in EGCG concentrations ≥ 0.5 mM. The effect of EGCG on the viability of *K. septempunctata* was depended on EGCG concentration (Fig. 2A-E). The evaluation of viability using flow cytometer was completed under 5 min.

It was reported several chemicals affecting the viability of *K. septempunctata* using approximately 100 spores with two kinds of fluorescent stains to identify the dead/live spores [11,12]. In our study, the viability of 5,000 spores was determined using one fluorescent stain. Ahn et al. [11] reported that AH had an anti-*Kudooa* effect at higher concentrations ≥ 1 mM and at longer treatment time (48 hr). The candidate chemicals should have anti-*Kudooa* effect at lower concentrations and in a shorter time since it will be used in fish farms. Ohnishi et al. [12] reported the anti-*Kudooa* effect of extract of coffee and tea, caffeine, and EGCG. The caffeine did not show any effect at 1 mM concentration, whereas 100% of *K. septempunctata* spores were inactivated at 0.1 mM EGCG. They proposed that the EGCG is an excellent candidate for the treatment and prevention of *K. sep-*

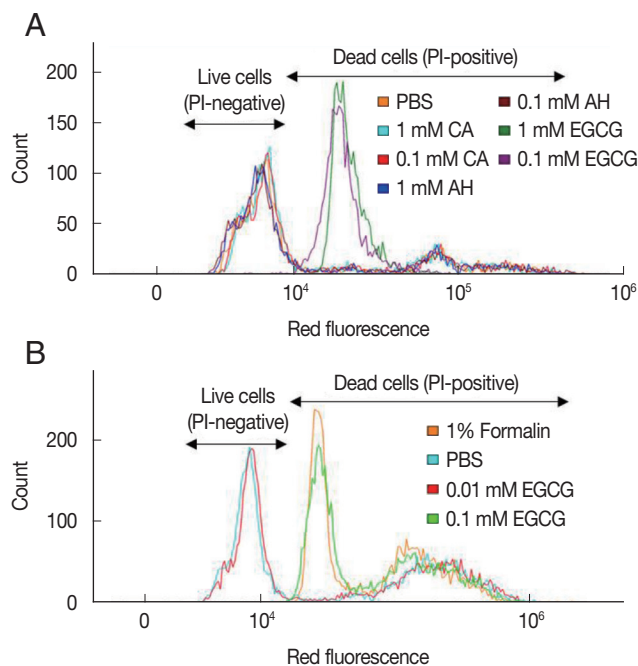


Fig. 1. Assessment of viability of *Kudooa septempunctata* myxospores using flow cytometry and propidium iodide staining. (A) The myxospores were treated with PBS, and 0.1 and 1 mM of amprolium hydrochloride (AH), chlorogenic acid (CA), and epigallocatechin gallate (EGCG). (B) The myxospores were treated with PBS, 1% formalin, and 0.01 and 0.1 mM EGCG.

tempunctata infection and emphasized the need to investigate other chemicals, such as chlorogenic acid (coffee ingredient). The present study also showed a complete anti-*Kudooa* effect of EGCG at 0.1 mM or more. There was, however, a difference of anti-*Kudooa* effect (36.9% vs. 80.6% dead spores) in the 0.01 mM EGCG groups different treatment time (30 min vs. 4 hr). In addition, we did not find any anti-*Kudooa* effect when spores were treated with CA. Consequently, we suggest that the EGCG has an anti-*Kudooa* effect at low concentrations and short treatment times as well as it is more efficient than CA in affecting the viability of *K. septempunctata* *in vitro*.

We speculated that EGCG effects on the integrity of shell valves of *K. septempunctata* spores, and it was confirmed by SEM. The spores in the PBS group showed intact surface feature (Fig. 3A and D). The spores treated with 0.1 mM EGCG showed partial cracks on the shell valves (Fig. 3B and E), cracks on whole shell valves in the 1 mM EGCG group (Fig. 3C and F). Previous studies reported that EGCG had potential to bind and damage to the gram-positive bacterial cell membrane which contain polysaccharides and glycocalyx [16,17]. This effect is of particular importance because carbohydrate residues are

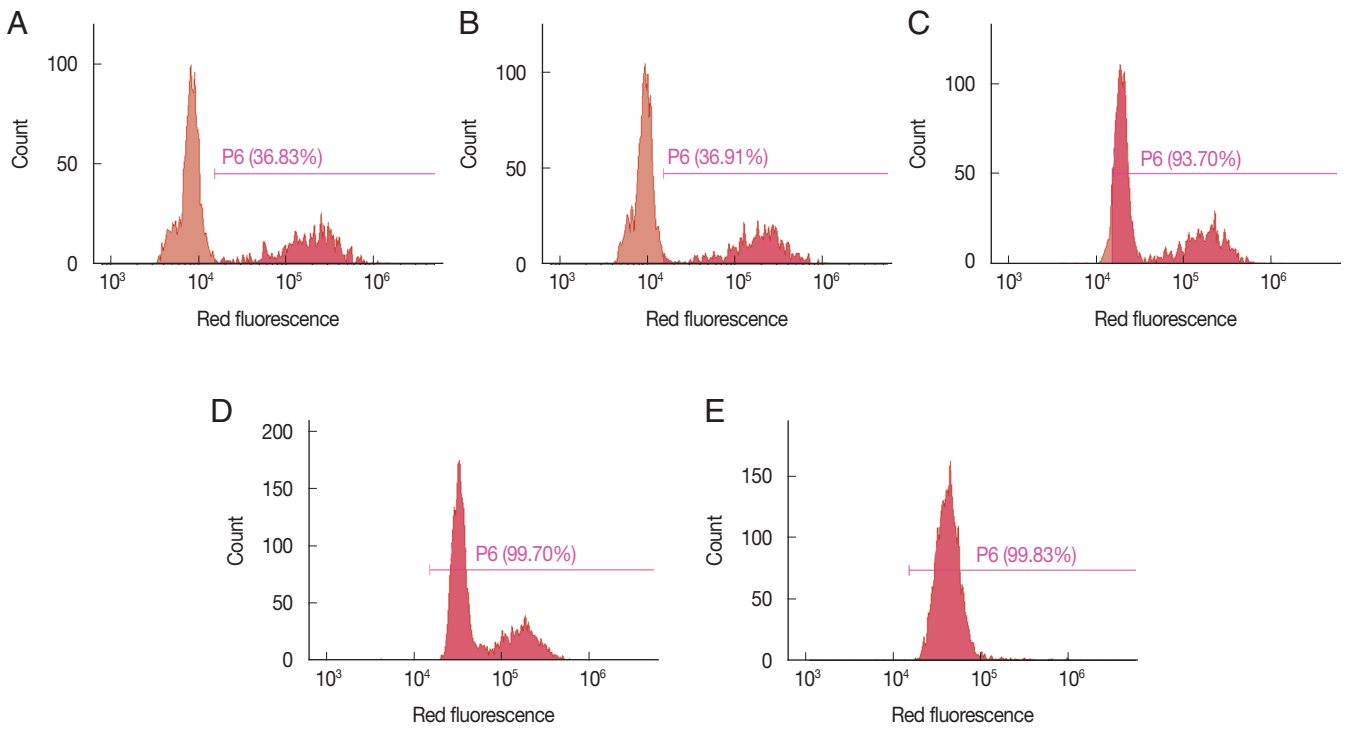


Fig. 2. Assessment of viability of *K. septempunctata* myxospores depends on the concentration of epigallocatechin gallate. Myxospores were treated with (A) phosphate-buffered saline, and (B) 0.01, (C) 0.05, (D) 0.1, and (E) 1 mM epigallocatechin gallate.

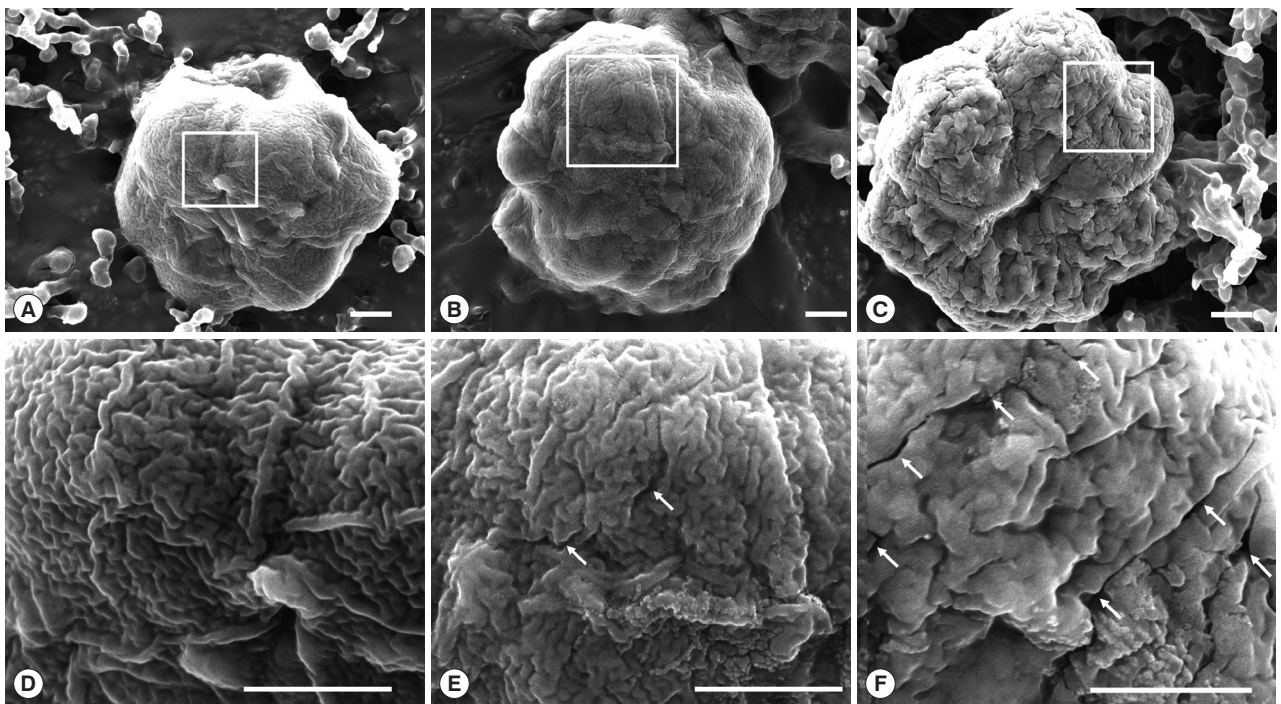


Fig. 3. Scanning electron micrographs of the spores of *K. septempunctata* treated with epigallocatechin gallate. (A) The spores were treated with phosphate buffered saline, (B) 0.1 EGCG, and (C) 1 mM EGCG. (D-F) Magnification of the square regions. Arrow indicates cracks on the shell valve. Scale bar = 1 μ m.

present in the spore and plasmodial membrane of *Kudoa* species [18,19]. In addition, the carbohydrate residues are participatory in the host-parasite relationships and immune responses in myxosporeans [19,20]. It was speculated that EGCG binds to carbohydrate or glycosylated components of the shell valve and damages integrity of the spores. However, further studies are needed to determine the type of cell wall constituents that EGCG binds to and the mechanism of crack formation on the shell valve of *K. septempunctata*.

Application of herbal remedies including EGCG were reported to control disease in the aquaculture, which had low toxicity and minimal environmental impacts [21]. EGCG inhibited grass carp reovirus, white spot syndrome virus, and *Vibrio alginolyticus* infection in fish, crab, and shrimp [22-24]. However, EGCG to crab caused the hemocyte damages and the administration of EGCG into mouse peritoneum produced severe hepatotoxicity [23,25]. We speculate that the sharp increase of EGCG concentration in plasma might cause adverse effects. It is suggested to administer EGCG to fish by adding to aquaculture feed.

In conclusion, EGCG markedly decreased viability of *K. septempunctata* spores and affected integrity of the shell valves of spores. This finding will contribute to the development of chemotherapeutic agents against the infection of *K. septempunctata* on olive flounders. In addition, flow cytometry could be used to evaluate and quantify the efficacy of chemicals on other myxosporeans. Further studies need to determine an optimal mixing ratio of EGCG to the feed by identifying pharmacokinetics and toxicity of EGCG after oral administering to fish.

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CONFLICT OF INTEREST

The authors declare no conflict of interest related to this study.

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