Contents lists available at ScienceDirect

Heliyon



journal homepage: www.cell.com/heliyon

Review article

5²CelPress

Emulsification and stabilisation technologies used for the inclusion of lipophilic functional ingredients in food systems

Alejandra Henao-Ardila^{a, b}, María Ximena Quintanilla-Carvajal^b, Fabián Leonardo Moreno^{b,*}

^a Doctorate in Biosciences, Faculty of Engineering, Universidad de La Sabana, Campus Universitario del Puente del Común, Km7 Autopista Norte de Bogotá, Chía, Cundinamarca, Colombia

^b Grupo de Investigación en Procesos Agroindustriales, Faculty of Engineering, Universidad de La Sabana, Campus Universitario del Puente del Común, Km7 Autopista Norte de Bogotá, Chía, Cundinamarca, Colombia

ARTICLE INFO

Keywords: Food science Encapsulation Food Oil

ABSTRACT

Food industry is increasingly using functional ingredients to improve the food product quality. Lipid-containing functional ingredients are important sources of nutrients. This review examines the current state of emulsification and stabilisation technologies for incorporating lipophilic functional ingredients into food systems. Lipophilic functional ingredients, such as omega-3 fatty acids, carotenoids, and fat-soluble vitamins, offer numerous health benefits but present challenges due to their limited solubility in water-based food matrices. Emulsification techniques enable the dispersion of these ingredients in aqueous environments, facilitating their inclusion in a variety of food products. This review highlights recent advances in food emulsion formulation, emulsification methods and stabilisation techniques which, together, improve the stability and bioavailability of lipophilic compounds. The role of various emulsifiers, stabilizers, and encapsulation materials in enhancing the functionality of these ingredients is also explored. Furthermore, the review discusses different stabilisation techniques which can yield in emulsion in a solid or liquid state. By providing a comprehensive overview of current technologies, this review aims to guide future research and application in the development of functional foods enriched with lipophilic ingredients.

1. Introduction

Global population is estimated to reach approximately 8.5 billion by 2030, according to the [1]. However, the expansion of agricultural land may not keep pace with such rapid growth. As daily food requirements increase, meeting the rising demand has become a challenge. A potential solution to this dilemma is the development of healthy and more nutritionally dense food products. This approach allows individuals to fulfil their nutritional needs while consuming smaller portions of food, as proposed by Santos-Buelga et al., in 2019 [2]. These foods are commonly referred to as functional foods. Functional foods represent a category of food that has the potential to enhance various physiological aspects of human well-being. They are designed not only to provide essential nutrients but also to positively impact specific functions within the human body. Functional foods play a crucial role in improving human vitality by promoting the overall health and preventing or managing diverse metabolic disorders and illnesses [3,4].

* Corresponding author. *E-mail address:* leonardo.moreno@unisabana.edu.co (F.L. Moreno).

https://doi.org/10.1016/j.heliyon.2024.e32150

Received 2 March 2024; Received in revised form 24 May 2024; Accepted 29 May 2024

Available online 29 May 2024

^{2405-8440/© 2024} The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC license (http://creativecommons.org/licenses/by-nc/4.0/).

Functional ingredients that confer specific benefits to foods are distinct from conventional food additives. These ingredients, which are extracted from bioresources, retain their inherent properties after extraction [5]. Lipophilic functional ingredients such as vitamins, essential oils, and fatty acids play a crucial role in human health. For instance, provitamin A carotenoids like beta-carotene convert in the body to retinal and retinoic acid, supporting vision, immune function, and growth [6,7]. Consuming polyunsaturated fatty acids (PUFA) offers antiarrhythmic effects, prevents platelet aggregation, relaxes arterial smooth muscle, and reduces plasma triglycerides and blood pressure. Increased intake of omega-3 fatty acids may also protect against chronic and inflammatory disorders like rheumatoid arthritis and asthma [8–10]. Vitamin E is a fat-soluble antioxidant that prevents the production of reactive oxygen species during fat oxidation, aiding muscle preservation and brain health. It protects fatty acids in lipoproteins and cell membranes, reducing oxidative damage to DNA and RNA, and boosts the immune system, potentially lowering the risk of cancer and arthritis [11–15].

Incorporating these compounds directly into food systems is challenging because of various factors, such as limited water solubility, susceptibility to chemical or enzymatic degradation, and incompatibility with the surrounding food matrix, which collectively hinder their optimal oral bioavailability [16]. Consequently, protection of lipophilic functional ingredients is essential. Protecting of lipophilic functional ingredients via emulsification plays a vital role in increasing the oral bioavailability and providing stability and protection to these compounds [16–18].

Emulsions are colloidal systems made up of at least two immiscible liquids in which one of them if finely dispersed in the other with the help of surfactants and homogenization technologies. The dispersed liquid is known as the dispersed phase, and the other liquid is known as the continuous phase [19]. Oil and water are the most common liquids used to prepare food-grade emulsions; therefore, there are two basic types of emulsions: oil-in-water emulsions (O/W), where oil is the dispersed phase, and water-in-oil emulsions (W/O), where water is the dispersed phase. O/W and W/O emulsions are the most common types of colloidal dispersions used for the encapsulation and controlled release of lipid components such as lipophilic vitamins and essential fatty acids [20,21].

Such systems are thermodynamically unstable because the contact between water and oil is unfavourable; therefore, they break out over time, affecting the shelf-life and protective capacity of the emulsions [18,21]. Several mechanisms can cause the destabilization of an emulsion, among which creaming, flocculation, and coalescence are the most common [22]. Different stabilisation techniques such as drying, bead encapsulation, or mixtures of different emulsifiers can be used to guarantee the stabilization of emulsions and prevent their separation over time. Spray drying is one of the most commonly used techniques for the stabilisation of emulsions [23–25] however, several emerging and traditional technologies are being studied as alternatives to stabilise these emulsions and confer additional protection against adverse conditions in the gastrointestinal tract.

The study of the fate of emulsions in the gastrointestinal tract is important because food digestion is a complex and multi-stage process that has a direct link between the food ingested and the health or disease of a person [6,18,19]. During human digestion, two main processes occur simultaneously: (i) mechanical transformations that reduce the size of food particles and (ii) enzymatic transformations, where macromolecules are hydrolysed into smaller constituents that have the potential to interact and be absorbed by the organism to reach the bloodstream and become bio-accessible [19,21,26,27]. The digestibility of the fundamental components of the emulsions was measured separately to observe the behaviour of the compounds and verify their digestibility. It is also important to evaluate the behaviour of emulsions after passage through the gastrointestinal tract to assess their protective effect on the compound of interest. A study on the behaviour of the stabilized emulsion is also required to determine whether the stabilisation process can provide an additional protective effect to the emulsions. Finally, analysis of the behaviour of the emulsion in the final matrix is essential to determine whether it will reach the final consumer or if there is some interaction between the functional ingredient and food.

Publications on the encapsulation and stabilisation of food-grade functional oils via emulsification or other technologies and the study of their functional activities have increased since 2015. A total of 157 research articles on this topic were published between



Fig. 1. Publications on encapsulation and emulsification of food grade functional lipophilic ingredients.

2015 and 2023, with the number increasing in recent years, showing the growing research interest in this area (Fig. 1).

The process of developing, stabilising, and protecting lipophilic functional compounds involves several steps Fig. 2. This figure also describes the organisation of topics presented in this review. The development of functional emulsions begins with knowledge of the food matrix in which the encapsulated functional ingredients are included. The type of emulsion to be prepared depends on the characteristics of the food system and the raw materials required for the preparation and stabilisation of the emulsions. Once the formulation is established, an emulsification process is performed in which various emulsification techniques are used to generate stable food systems. However, in many cases, these systems are not stable, and it is necessary to use stabilisation technologies to increase the shelf-life and achieve greater protection of lipophilic functional compounds. The selection of these technologies is important for product development. The functionally encapsulated and stable lipophilic compound is then included in the food system to reach the final consumer. Finally, evaluation of the behaviour of lipophilic functional ingredients during their movement through the gastrointestinal tract at each stage of the encapsulation process is important to ensure the bootability of functional ingredients susceptible to degradation. The aim of this review was to provide insights into the emulsification and stabilisation technologies used for the inclusion of lipophilic functional ingredients in food systems. This review also discusses various raw materials, formulations of emulsions, and emulsification and encapsulation technologies for the release of compounds during passage through the gastrointestinal tract.

For the preparation of this review, articles, including both original research and review papers, were sourced from the Science-Direct database. The selected articles contained the keywords "emulsion," "encapsulation," "food," "oil," and "functional" in their titles, keywords, and abstracts and the time period ranged between 2015 and 2023.

2. Food emulsion formulation

2.1. Emulsions

An emulsion is a fine dispersion system in which a discontinuous phase is homogeneously distributed as small spherical droplets in a continuous phase, with the emulsion droplets stabilized by an amphiphilic surface-active surfactant [16,19]. Emulsions can be classified according to the spatial organisation of their phases. A system consisting of oil droplets dispersed in an aqueous phase is called an oil-in-water emulsion (O/W), and a system in which water is dispersed in an oil phase is called a water-in-oil emulsion (W/O) [17,21,28]. The choice of emulsion system for the protection of functional ingredients depends on the nature of the compound to be protected and the matrix in which the emulsion is to be incorporated.

The oil-in-water emulsion consisted of small oil droplets dispersed in an aqueous medium surrounded by a thin interfacial layer consisting of emulsifier molecules. The advantages of these systems include their relative ease of preparation and low cost [29]. Regarding the protection of lipophilic functional ingredients, oil-in-water (O/W) emulsions are very common; however, O/W



Fig. 2. Steps for the development and evaluation of stable functional emulsions.

Table 1

Main research articles on the use of functional lipophilic ingredients as active ingredients for the formulation of emulsions.

Lipophilic functional ingredient	Disperse phase	Continuous phase	Main findings	Emulsification technology	Reference
Curcumin	Rapeseed oil	Chitosan (CS) in acetic acid solution and gum Arabic (GA) in ultrapure water	Curcumin emulsions showed a half-life of 144 h, and approximately 50 % of curcumin was degraded. Pickering emulsion stabilized by CS-GA nanoparticles significantly improved the binavailability of curcumin	Pickering emulsion. High- speed homogenizer at 12,000 rpm for 8 min	(Han et al., 2020) [91]
	Medium chain triglyceride (Miglyol 812)	Octenyl succinic anhydride (OSA)-modified quinoa starch in phosphate buffer solution	Pickering emulsion preserved up to 95.3 % of curcumin after simulated oral digestion and 86.2 % after simulated gastric digestion, reaching the intestine, where most nutrient absorption occurs, ultimately retaining up to 86.8 % of encapsulated curcumin.	High sheer homogenizer at 22,000 rpm for 30 s	(Marefati et al., 2017) [171]
	Curcumin and lecithin diluted in warm absolute ethanol	Debranched starch dispersed in distilled water	Curcumin was successfully encapsulated using debranched starch but the stability of the emulsions was not good, consistent with the low zeta potential values. Results also showed that water bridges between the debranched starch and curcumin may contribute to the complexing processes for encapsulation.	High pressure homogenizer at 500 bar for 2 cycles	(Feng et al., 2020) [185]
	Medium chain triglycerides	Modified kaolinite with phosphatidylcholine dispersed in deionized	-	high-shear homogenizer at 8000 rpm for 30 s	
Carotenoids	Medium Chain Triglycerides oil	pullulan and whey protein isolate dissolved in deionized water	Concentration of beta carotene went from 0 %, after 7 d of storage without wall material, to 40 % after 7 d of storage with the wall material mixture showing an improvement in beta carotene stability due to emulsion formulation	Ultrasonication at 0, 120, 240, 360, and 480 W	(Niu et al., 2020) [63]
	Corn oil	Wheat gluten and Xantan gum dissolved in deionized water	Presence of Xantan gum in the formulation increased the stability of beta carotene by 39 % after storage at 60 °C for 14 d.	High-shear mixer at 13,500 rpm for 2.5 min	(Fu et al., 2019) [64]
	Soybean oil	Whey protein concentrate dissolved in distilled water and Zein from corn dissolved in a water-ethanol mixture (20:80 v/v)	Emulsion exhibited high beta carotene concentration, up to 74 %. Encapsulated beta carotene showed a higher bioaccessibility after in-vitro digestion, which was negligible in its free form.	High-shear mixer at 6000 rpm for 5 min and ultrasonication at an amplitude of 10 % and frequency of 20 kHz for 2 min in pulse mode.	(Gómez- Mascaraque et al., 2017) [51]
	Medium chain triglycerides	OSA-starch (Hi-cap 100 ®) dissolved in distilled water and chitosan dissolved in acetic acid solution (1 % v/v)	OSA-starch and chitosan interface in Layer-by-Layer emulsion showed a higher stability of beta carotene against heating than the single layer emulsion	High-pressure homogenizer at 600 bar for 3 cycles for OSA-starch primary emulsion and 300 bar for 4 cycles for OSA-starch- chitosan emulsion	(S. Fang et al., 2019) [65]
	Corn oil	β-lactoglobulin and tween 20 dispersed in phosphate buffer	Vitamin E and CoEnzyme Q10 were used as antioxidant agents to prevent beta carotene oxidation during emulsion storage. Vitamin E, along with the wall materials, effectively retarded beta	Microfluidizer at 9000 psi for 3 cycles	(Qian et al., 2012a) [66]

(continued on next page)

Vegetable oil Maltodextrin and sodium caseinate Vitamin E Sunflower oil Maltodextrin, acacia gum β-glucans-rich extracts obtained from <i>P. ostreatus</i> powder in distilled water	carotene degradation in emulsion systems. Lower temperature and feed rate conditions showed a higher beta carotene preservation than higher conditions. Storage temperature had influence on the degradation of beta carotene since higher loss was observed at 45 °C. , and Samples with 10 % oil and β -glucan extracts had an α -tocopherol retention up to 95.7 %, which was 68 % higher than that of the control	High pressure homogenizer at 68.95 MPa for 1 cycle Rotor-stator homogenization at 3900 rpm for 10 min	(Coronel-Aguilera and San Martín- González, 2015) [67] (Gallotti et al., 2020) [75]
Vegetable oil Maltodextrin and sodium caseinate Vitamin E Sunflower oil Maltodextrin, acacia gum β-glucans-rich extracts obtained from <i>P. ostreatus</i> powder in distilled water	Lower temperature and feed rate conditions showed a higher beta carotene preservation than higher conditions. Storage temperature had influence on the degradation of beta carotene since higher loss was observed at 45 °C. , and Samples with 10 % oil and β -glucan extracts had an α -tocopherol retention up to 95.7 %, which was 68 % higher than that of the control	High pressure homogenizer at 68.95 MPa for 1 cycle Rotor-stator homogenization at 3900 rpm for 10 min	(Coronel-Aguilera and San Martín- González, 2015) [67] (Gallotti et al., 2020) [75]
Vitamin E Sunflower oil Maltodextrin, acacia gum β -glucans-rich extracts obtained from <i>P. ostreatus</i> powder in distilled water	, and Samples with 10 % oil and β-glucan extracts had an α-tocopherol retention up to 95.7 %, which was 68 % higher than that of the control	Rotor-stator homogenization at 3900 rpm for 10 min	(Gallotti et al., 2020) [75]
	with no β-glucan extracts, which have antioxidant properties.		
Medium chain OSA-modified starches triacylglycerol oil (Capsul, Capsul TA, and HICAP100) dissolved in distilled water	Vitamin E retention ranged from 79.16 to 71.46 %. Dried emulsions containing Capsul were able to retain up to 50 % of the encapsulated vitamin E after 60 d storage at 4–35 °C. HICAP showed a reduced retention and shorter half-life (35 d).	Rotor-stator homogenizer at 14000 rpm for 5 min	(Hategekimana et al., 2015) [76]
Canola and Ora-pro-nobis mucilage a coconut oils whey protein isolate dispo in distilled water	nd α-Tocopherol retention and degradation kinetics were greater affected when canola oil was used as a dispersed phase compared to coconut oil, where more than 85 % was retained. Wall materials improved the chemical stability and bioaccessibility by increasing its value by 5.8- times.	High-pressure homogenizer at 50 MPa and 10 cycles	(Neves et al., 2020) [78]
Orange oil Whey protein isolate, suc stevia, and citric acid dissolved in distilled wate	rose, Vitamin E retention reached levels up to 85 % for most er. emulsion formulations, which indicates that emulsions show great potential as delivery systems of lipophilic bioactive compounds.	Single stage valve homogenizer at 25 MPa for the first cycle and at 8 MPa for the second cycle	(Raikos, 2017) [77]
Medium Chain Citric Buffer triglycerides (Miglyol 812)	Surfactants present in the nanoemulsions led to the formation of colloidal structures in the mixed micelle phase that were able to encapsulate vitamin E acetate, thereby achieving a bioavailability value of 98 %.	Microfluidizer at 12,000 psi for 4 cycles	(Mayer et al., 2013) [69]
Omega 3 Sacha inchi oil Xantan gum, pectin, and ovalbumin	Pectin had a greater efficiency to hold Sacha Inchi Oil than Xantan Gum. Procedure did not affect the omega 3 content in the emulsion. Microcapsules showed a very slow omega-3 release profile, which is good because it prevents omega-3 loss before reaching the intestine.	Sonication at a frequency of 30 KHz and amplitude of 100 % for 12 min	(Vicente et al., 2017) [84]
Fish oil (extracted from Carp viscera)Chitosan dissolved in ace acid (1 % v/v)	tic Encapsulation process had no impact on the oil quality. They also showed a potential	Mechanical stirrer at 10,000 rpm for 10 and 20 min	(Esquerdo et al., 2015) [186]

A. Henao-Ardila et al. Table 1 (continued)

Lipophilic functional	Disperse phase	Continuous phase	Main findings	Emulsification technology	Reference
ingredient					
	Fish and garlic essential oils	Persian gum dissolved in distilled water and chitosan dissolved in glacial acetic acid (1 % v/v)	to retard the primary oxidation of unsaturated fatty acid contained in the Carp viscera oil Increasing Persian gum concentration improved the stability of emulsions. Capsules were stable at temperatures under 250 °C, indicating that wall material may protect the encapsulated oil in food processing methods involving high temperatures	Rotor-stator homogenizer at 4000 and 10,000 rpm for 3 and 1 min, respectively, followed by sonification at an amplitude of 100 % for 4 min for the first layer; rotor- stator homogenizer at 3000 rpm for 0.5 min and 10,000 rpm for 0.5 min followed by sonication at an amplitude of 80 % for 0.5 min	(Raeisi et al., 2019) [83]
	Fish oil	Whey protein isolate (WPI) and fish protein hydrolysate (FPH) dissolved in distilled water	WPI exhibited a higher emulsifying property than FPH and led to more physically stable blend with smaller droplet size. Although most of the fish oil was encapsulated inside the fibers as small droplets, the fibers presented poor oxidative stability	Microfluidizer at 9000 psi for 3 cycles	(García-Moreno et al., 2016) [187]
Vitamin D	Corn oil, fish oil, mineral oil, orange oil, or Miglyol 812	Q-Naturale® diluted in 10 mM sodium phosphate buffer	Bioaccessibility was highly dependent on the type of carrier oil present in the nanoemulsions. It was higher for corn and fish oils due to the higher solubilization capacity for vitamin D3 of mixed micelles formed by long chain free fatty acids. Long chain triglyceride nanoemulsions were the most suitable for increasing the bioaccessibility of vitamin D3.	High-pressure homogenizer at 12,000 for 3 cycles	(Ozturk et al., 2015) [188]
	Soybean oil	Nanofibrillated cellulose (NFC) or WPI diluted in 10 mM sodium phosphate buffer	Residual vitamin D in emulsions after passage through simulated conditions of the gastrointestinal tract decreased when the NFC content increased because the cellulose fibres did not allow lipase to access the oil droplets present in the emulsion. The cellulose fibres did not have a major impact on vitamin bioaccessibility and stability, and led to higher values for whey protein when used in low concentrations (1:10 NFC-to-oil).	High pressure homogenizer at 12,000 psi for 3 cycles	(Winuprasith et al., 2018) [55]

A. Henao-Ardila et al.

Table 1 (continued)

emulsions are not only designed to protect lipid compounds, Essential oils [30–34] and microorganisms [35–37] can also be protected in O/W emulsions.

W/O emulsions are usually present in the solid or semi-solid state because they are very unstable in the liquid state owing to the high mobility of water droplets [38,39]. Water-in-oil emulsions can also be used as a tool to reduce the saturated and *trans*-fat content of processed foods [40]; Luo et al. (2019) fabricated a water-in-oil emulsion using tea polyphenol palmitate as a stabiliser to substitute for transfat-rich solid or semi-solid fats. They found that the palmitate particles produced stable, surfactant-free, gel-like W/O emulsions. The emulsion had a high slip melting point, comparable to that of margarine, providing a potential alternative to solid fats [41].

Oil-in-water or water-in-oil emulsions are not the only possibilities; it is also possible to create several types of emulsions, such as oil-in-water or water-in-oil (O/W/O) or water-in-oil-in-water (W/O/W) emulsions [42]. Water-in-oil-in-water (W/O/W) emulsions consist of small water droplets contained within larger oil droplets dispersed in an aqueous continuous phase, whereas oil-in-water-in-oil emulsions consist of small oil droplets contained within larger water droplets dispersed in an oil continuous phase. In the food industry, double emulsions are used in the preparation of low-calorie and reduced-fat-content products to encapsulate and protect bioactive ingredients (vitamins, minerals, enzymes, proteins, polysaccharides, etc.), antioxidants, and flavour compounds and to control the release of bioactive compounds during digestion [42–45]. Double emulsions are more accurately designated as W1/O/W2 or O1/W/O2 emulsions, where W1 or O1 is the inner phase and W2 or O2 is the outer phase, which may have different compositions. Chouaibi et al. (2019) used W/O/W emulsions to encapsulate quercetin using different oil phases (olive, red pepper seed and sunflower oils) finding that double emulsions were stable when stored at refrigerated (5 °C) and room temperature, they also concluded that the release behaviour was strongly influenced by the composition of the oil phase, being sunflower oil the one that showed a greater release rate and red pepper seed oil the one with the smallest release rate [46].

2.2. Lipophilic functional ingredients

There has been growing interest in the food industry to encapsulate, protect, and release various lipophilic food components [47]. Functional ingredients such as β -carotene [48–51], tocopherols [52–54], vitamine D₃ [55,56], etc. have been encapsulated successfully, improving the bioavailability, and stability of functional ingredients. Table 1 lists some of the most common lipophilic ingredients encapsulated using the emulsification technique.

Carotenoids: Carotenoids are a large and diverse group of yellow-to-red lipophilic compounds that consist of 3–13 conjugated double bonds and, in some cases 6 carbon ring structures at one or both ends of the molecule [57,58]. Carotenoids containing oxygen are known as xanthophylls while those without oxygen are known as carotenes, where β and α -carotene are found [59]. Carotenoids, particularly beta-carotene, can be metabolized within the body to form retinal and retinoic acid, compounds vital for vision, immune function, normal growth, and development. Retinoic acid has demonstrated the ability to inhibit the development of various cancers in both animal and in vitro studies. Additionally, the accumulation of lutein and zeaxanthin in the eye has been linked to a decreased risk of age-related macular degeneration, a major cause of blindness in older adults. These compounds help protect against light-induced damage, a key risk factor for macular degeneration, by filtering high-energy blue light and neutralizing reactive oxidants [60,61].

Owing to their importance in the food industry and their role in promoting human health and reducing the risk of chronic diseases due to vitamin A deficiency, carotenoids, specifically beta-carotene, are lipophilic ingredients that have drawn the attention of the food industry [58]. Carotenoids that are naturally present in foods are generally stable; however, when used as food additives, they are relatively unstable because they are sensitive to pH, light, oxygen, and temperature and are susceptible to oxidation, isomerisation, and photosensitization during production and storage, in addition to their low water solubility [16]. Therefore, the use of carotenoids as additives in food matrices may result in degradation and poor solubility [62]. To overcome these difficulties, several researchers (Table 1) have included beta carotene in emulsion systems. All results showed how beta-carotene stability improved [63–67] and how bioavailability increased as a result of water solubility improvement [51].

Vitamin E: Vitamin E is a yellow viscous oil that is readily oxidised when exposed to light and oxygen [13]. Vitamin E is made up of out of eight lipophilic compounds; four tocopherols and four tocotrienols designated as α , β , γ and δ , with α -tocopherol being the most abundant and bioactive form [68,69]. Tocopherols differ from tocotrienols in that they contain three double bonds at carbons 3, 7, and 11 [70]. Vitamin E plays a crucial role in human health, offering a range of benefits supported by substantial research. It helps to strengthen the immune system and prevent inflammation, which can reduce the risk of diseases such as cancer and arthritis. By inhibiting the production of reactive oxygen species during fat oxidation, vitamin E may also play a key role in preventing muscle degeneration associated with aging. Additionally, vitamin E contributes to brain health by combating oxidative stress, a major factor in neurodegenerative disorders such as Alzheimer's disease [68,71,72].

Vitamin E has low water solubility and poor chemical stability (to oxygen, light, and heat). Food processing may lead to substantial changes in the vitamin E content that is naturally present in food, leading to variable bioavailability [73,74]. Suitable delivery systems are required to incorporate Vitamin E into aqueous products to improve their chemical stability [75–77] and bioavailability [69,78].

Omega-3 fatty acids: Omega-3 fatty acids are unsaturated fatty acids with a double bond that is 3 carbon atoms from the methyl end of the molecule. The most common ω -3 fatty acids are α -linolenic acid, eicosapentaenoic acid, and docosahexaenoic acid [59]. The literature indicates that omega-3 fatty acids are essential for normal growth and development, as well as for their positive effects on the heart, brain, eyes, joints, skin, mood, and behaviour [79,80]. Had also shown to possess antiarrhythmic actions, prevention of platelet aggregation, relaxing of arterial smooth muscle, lowering of plasma triglycerides and blood pressure, and a modest antithrombotic effect [81]. Fatty acids are highly susceptible to lipid oxidation and have low water solubility. Therefore, omega-3 fatty acids are usually incorporated into colloidal delivery systems to protect them during processing, storage, and transport [82]. The main food source of Omega-3 fatty acids is fish oil [83], which is why most authors shown in Table 1 use fish oil as the dispersed phase in the formulation of emulsions. However, new sources of omega 3 fatty acids, such as Sacha Inchi oil, have been explored [84].

Vitamin D: Vitamin D is a liposoluble micronutrient found in two different chemical forms, vitamin D_2 (ergocalciferol) and vitamin D_3 (cholecalciferol). In humans, vitamin D_3 has greater biological potency than vitamin D_2 and is typically synthesised by the skin after exposure to light [55,85]. Vitamin D plays a crucial role in maintaining the health of older adults by preventing osteoporosis and reducing the risk of cancer. A deficiency in this nutrient has been linked to heart disease, including hypertension, type 1 and 2 heart failure, and ischemic heart disease. Long recognized for its role in bone health, insufficient vitamin D can lead to rickets, a condition characterized by soft bones and skeletal deformities. Adequate dietary intake of vitamin D is essential for supporting overall health and

A. Henao-Ardila et al.

mitigating these health risks [26,86,87].

Fortification with vitamin D has been a challenge to the food industry because of its instability under various conditions (temperature, iodine, acidic conditions, and oxidation) and heterogeneous distribution in food [87]; therefore, its incorporation into colloidal delivery systems is an alternative to overcome these difficulties. The results presented in Table 1 show that bioaccessibility increased when vitamin D was included in colloidal delivery systems. However, it is also important to optimise the encapsulation conditions because excessive use of stabilisers can prevent the release of the oil included within the emulsions, limiting the bioavailability and bioaccessibility of functional oils even more than when they are not included in the colloidal matrices.

Other lipophilic functional ingredients: Curcumin, a non-polar polyphenol and the main component of turmeric roots, is often used as a natural food pigment due to its bright yellow colour [88,89]. Its use in the food industry as a pigment is well known, but curcumin has also aroused great interest in the functional food industry owing to its health benefits, including antioxidant, anti-inflammatory, and anticancer activities might be found. Therefore, there has been great interest in including this ingredient in different food products. However, the poor physicochemical stability and bioavailability of curcumin severely limit its application, and therefore, its encapsulation has attracted great attention [90]. The emulsification technique improved the half-life and bioavailability of curcumin under storage conditions [91].

Another lipophilic functional ingredient of interest is resveratrol, a natural polyphenolic compound found in various plants, including grapes, red wine, and some berries. It is well-known for its potential health benefits, including antioxidant and antiinflammatory properties [23]. The incorporation of resveratrol into functional food matrices can be challenging because of its poor water solubility, low bioavailability, and chemical instability [92]. Emulsions can provide a protective environment for resveratrol by enhancing its stability, solubility, and bioavailability [93]. Several studies have shown that the use of emulsions improves resveratrol bioavailability [17,18,92,94].

Coenzyme Q10 (CoQ10), another lipophilic functional ingredient of interest, is an endogenous compound found in human cells that plays a pivotal role in cellular energy production and functions as an antioxidant [95,96]. This molecule plays an important role in the electron transport chain and is responsible for generating adenosine triphosphate (ATP), the primary cellular energy source. Notably, CoQ10 contributes to cell well-being by supporting the maintenance of mitochondrial function. While the body has the capacity to synthesise CoQ10 naturally, its production can diminish with advancing age or specific medical conditions [97]. CoQ10, being fat-soluble, holds the potential to undergo degradation and oxidative stress. Thus, emulsions have emerged as valuable tools. Emulsions provide an ingenious means of encapsulating and protecting CoQ10 from the detrimental effects of oxidation, light, and heat. This encapsulation strategy enhanced the stability and bioavailability of CoQ10, thereby amplifying its effectiveness. Moreover, the oil-in-water (O/W) structure of emulsions promotes enhanced solubility and controlled release of CoQ10 in the digestive system, thereby optimising its absorption [98].

2.3. Solid particle stabilizers

Solid particles can adhere to the interface between immiscible phases, forming a tangible barrier that effectively halts droplet merging, a phenomenon known as coalescence. These particles, which can originate from natural sources such as clays or proteins, as well as synthetic sources such as polymers and nanoparticles, act as emulsion stabilisers. When these solid particles were introduced, they occupied the interfacial region, establishing a robust and intricate lattice-like structure. This framework exerts its influence by acting as a shield, shielding droplets from coalescence [99]. This stabilisation mechanism involving solid particles enables the creation of more stable emulsions. The same solid particles used to stabilise emulsions can also be employed to encapsulate lipophilic ingredients within an emulsion system. These particles serve a dual purpose: they provide stability to the emulsion, while also acting as carriers for the encapsulated lipophilic components [63,100].

Solid Particle selection is crucial during emulsion preparation as it influences the rheological properties of the emulsion, the mechanical stability, encapsulation efficiency and has an important effect on the shelf-life of the final product. Solid particles used in functional food-based emulsions should have good water solubility, film-forming properties, and emulsifying properties [99,101]. The



Fig. 3. Wall material classification. Adapted from: (Lekshmi et al., 2021a; Sobel et al., 2014).

solid particles are mainly natural polymers. These can be classified, as shown in Fig. 3. Their selection depends on the characteristics of the lipophilic ingredient to be encapsulated and the desired characteristics of the final product.

Proteins and polysaccharides are widely used as solid particle stabilisers [102]. Proteins can be divided into animal or vegetal proteins depending on their origin. Common animal proteins include milk-derived proteins (whey protein, milk protein concentrate, and different forms of caseinates, among others), ovalbumin, and gelatine. These proteins have unique properties, including biode-gradability, biocompatibility, and a high degree of structural flexibility, making them attractive materials for encapsulation. Whey protein, a byproduct of cheese production, has been studied for the encapsulation of various oils, including pomegranate seed oil [103], sunflower oil [104,105], as well as functional ingredients such as iron [21], resveratrol [106], and tocopherol [107]. The encapsulation efficiency of whey protein was high, and the coated oils showed improved oxidative stability [108]. Gelatine, a protein obtained from collagen, has also been studied for encapsulation of edible oils and functional lipophilic ingredients [109–112]. Casein, a milk protein, forms stable emulsions with various oils [113,114].

Among vegetable-based proteins, legumes, nuts, tubers, and cereal proteins are used for the encapsulation of lipophilic active ingredients. These natural polymers have good emulsifying and amphiphilic properties, which provide the necessary physical, chemical, and functional properties for the encapsulation of these ingredients. Legume-based proteins are the group with the greatest variety and include soybeans, peanuts, peas, chickpeas, beans, and lentils. Among the legume groups, soy protein is the most commonly used protein in the emulsification of bioactive compounds [115]. Vegetable proteins are often combined with natural polymers to create stable emulsions. Yang et al. (2020) studied the interaction between soy protein isolate (SPI) and chitosan (CS) and found that SPI and CS produced very stable emulsions that could be stored for 20 days, showing little change in appearance or droplet size. Wu et al. (2022) prepared emulsions based on soy protein isolates and mixed them with tannic acid to improve the antioxidant properties of the emulsion and protect essential oil properties. The results showed that nanoparticles also maintain the aroma of beverages and lengthen their shelf-life.

There are several advantages of using vegetable-based proteins for emulsification over animal-based proteins, including Allergenfree: Vegetable-based proteins that are free from common allergens such as milk, egg, and wheat, making them suitable for consumers with food allergies or intolerances [118]; Vegan-friendly: Vegetable-based proteins are suitable for vegan diets, which are becoming increasingly popular owing to ethical and environmental concerns and Health benefits: Some vegetable-based proteins, such as soy protein, have been associated with health benefits such as improved cholesterol levels and reduced risk of heart disease [35,119].

Polysaccharides are commercially used in the food industry because of their low toxicity, high biocompatibility, and biodegradability. Polysaccharides can reduce the Brownian movement of the system, maintain the stability of the emulsified components for a longer time, and can be used as thickeners or stabilisers to form a network structure to increase the viscosity of the continuous phase. Finally, polysaccharides can modify the rheological properties and increase the stability of emulsions [120]. Among polysaccharides, gums, starches, and chitosan are the most commonly used.

Natural gums are widely used in emulsification processes owing to their ability to stabilise emulsions and improve their texture and stability. One commonly used natural gum in emulsification is xanthan gum, a polysaccharide produced by Xanthomonas campestris [121]. Guar gum, derived from the seeds of the guar plant, *Cyamopsis tetragonoloba*. It is a galactomannan gum that has excellent water-binding properties [120]. Pectin is a heteropolysaccharide composed mainly of galacturonic acid units found in the cell walls of plants [120], and Gum Arabic: A natural exudate obtained from the Acacia tree. It is a complex mixture of polysaccharides and glycoproteins [31]; Carrageenan: Extracted from red marine algae, is an anionic polysaccharide consisting of p-galactose chain bearing one sulphate group every two of units. Commonly used stabilisers and emulsifiers for emulsions [122].

Starch is an abundant natural polysaccharide found in plant cells that serves as an energy storage molecule. Traditionally, they are recognized for their thickening and gelling properties. Starches act as emulsifying agents through several mechanisms including steric stabilisation, electrostatic interactions, and film formation [123]. When starches are dispersed in water, they form a colloidal system that can coat oil droplets, preventing their aggregation and subsequent phase separation [124]. The use of starch in emulsification is of growing interest given its abundance, feasibility of extraction, and potential to be modified to increase its hydrophobicity through esterification, such as hydrophobisation with octenyl succinic anhydride (n-OSA) which confers amphiphilic properties to the starch molecule [124,125].

Moreover, chitosan is a polymer obtained from the alkaline deacetylation of chitin, which naturally occurs in the cell walls of some fungi but mainly in the insect cuticles and exoskeletons of crustaceans [126,127]. Owing to its film-forming properties and antimicrobial activity, chitosan has been used as a wall material to carry lycopene [128,129] and essential oils, such as clove [130], cinnamon [33], garlic [32] and allspice [131]. The results showed that chitosan nanocarriers loaded with different pro-vitamin A showed improved thermal stability, dispersibility, and controlled release, indicating that it is a good alternative for the delivery of several bioactive compounds [129]. Essential oils retain their antimicrobial and fungicidal properties and enhance food and plant preservation [32,33,130,131].

Proteins and polysaccharides are commonly used to produce O/W emulsions. Proteins can stabilise emulsions based on their isoelectric points (Ips) [132]. However, aggregation can occur at high pH, Ip, and ionic strength. Therefore, the addition of polysaccharides can increase the stability of aggregation because polysaccharides form a thick charged interface layer that increases steric and electrostatic repulsion between droplet surfaces [133]. Polysaccharides are hydrophilic molecules that can be adsorbed onto the surface of protein-coated oil droplets through electrostatic interactions. Therefore, the stability of emulsions can be improved by forming complexes between the proteins and polysaccharides [120]. Several authors have reported the use of protein-polysaccharide complexes to encapsulate lipophilic ingredients such as clove essential oil [31]soybean oil [123], medium-chain triglycerides [63,65, 120], curcumin [91], fish oil [83], and Sacha inchi [84].

2.4. Surfactants

A surfactant, short for surface active agent, is a compound that has both hydrophilic and hydrophobic properties. This unique chemical structure allows surfactants to lower the surface tension between two immiscible phases by adsorption at the interface between them [134]. Therefore, surfactants are used in a wide range of applications, including emulsification, foaming, wetting, and dispersion. Surfactants contain polar head groups that can interact with water molecules via hydrogen bonding or electrostatic interactions. The head group can be a variety of chemical groups such as carboxylate, sulphate, amine, or quaternary ammonium. In addition, they have a nonpolar tail group, mainly composed of hydrocarbon chains of varying lengths and saturation levels, or fluorinated chains [135].

Surfactants produce more stable emulsions by reducing the interfacial tension between the two immiscible phases in the emulsion. This interfacial tension is the force that opposes the formation of an interface between the two phases and can lead to the formation of unstable emulsions, such as oil droplets coalescing or settling. Surfactants are commonly used in the food industry to emulsify oils and water-based ingredients and create stable matrices. Different types of surfactants, including natural and synthetic, can be used for food oil emulsification. An important consideration when selecting surfactants for food oil emulsification is their safety and potential impact on human health. Some surfactants may have negative effects on human health, such as allergic reactions or disruption of hormonal balance. Table 2 lists the most common surfactants for food-oil emulsification is their accepted daily intake per kg of body weight. Another consideration when selecting surfactants for food-oil emulsification is their able emulsions. Surfactants that are too weak may not be effective in creating stable emulsions, whereas surfactants that are too strong may cause emulsions to break

Table 2

Common surfactants used in the food industr	y, their descript	tion, and European Food Safety	Authority (EFSA)) accepted dail	v intake (ADI).
---	-------------------	--------------------------------	------------------	-----------------	-----------------

Surfactant	Description	International numbering system (INS)	EFSA	Reference
Lecithin	Lecithin is a naturally occurring phospholipid, a type of fat molecule, found in many plant and animal tissues. It is commonly extracted from sources such as soybeans, egg yolks, and sunflower seeds	E322	No safety concern for the general population from more than 1 year of age	(Mortensen et al., 2017b) [189]
Polysorbates	Polysorbates, also known as tweens, are non- ionic surfactants derived from polyethylene glycol (PEG) and sorbitan. The number in the name (20, 40, 60, 65, and 80) refers to the number of ethylene oxide units in the PEG chain	E432, E434, E435, E436, and E433	ADI of 25 mg/kg body weight/day. There are no concerns of genotoxicity, carcinogenicity, or developmental toxicity if the ADI dosage is followed.	(EFSA Panel of Food Additives and Nutrient Sources, 2015) [190]
Salts of fatty acids	Salts of fatty acids, also known as soap salts, are compounds formed from the reaction of a fatty acid with an alkali metal hydroxide or alkaline earth metal hydroxide. This reaction produces a salt, with fatty acid as the anion (negatively charged ion) and metal ion as the cation (positively charged ion). The most commonly used salts of fatty acids are sodium and potassium salts	E470	There were no data on subchronic toxicity, chronic toxicity, and reproductive and developmental toxicity of the salts of fatty acids. There was no concern for mutagenicity. No ADI was needed.	(Younes et al., 2018) [191]
Mono- and diglycerides	derived from natural sources such as vegetable oils and animal fats. Mono- and diglycerides are composed of one or two fatty acid chains attached to a glycerol backbone. They are soluble in both water and oil, which makes them ideal for stabilising emulsions	E471	No evidence for adverse effects was reported in short-term, subchronic, chronic, and reproductive and developmental toxicity studies. Neither carcinogenic potential nor a promotion effect in initiation/promotion was reported. No ADI was needed.	(Younes et al., 2017) [192]
Sorbitan fatty acid esters	Sorbitan monostearate (span 60), sorbitan tristearate (Span 65), sorbitan monolaurate (Span 20), sorbitan monooleate (Span 80), and sorbitan monopalmitate (Span 40) are non-ionic surfactants often used as food additives	E491, E492, E493, E494, and E495	ADI of 25 mg/kg body weight/day for E491, E492, and E495 and ADI of 5 mg/kg body weight/day for E493 and E494. Span may have side effects on the kidneys. No indication of carcinogenicity was observed if ADI was represented	(Mortensen et al., 2017a) [193]
Ammonium phosphatides	They are derived from a mixture of phospholipids, primarily derived from soybean oil, and are composed of phosphatidylcholine, phosphatidylethanolamine, and phosphatidylinositol among others.	E442	ADI of 30 mg/kg body weight/day. Usage as a food additive at the permitted ADI dosage is not of safety concern.	(Mortensen et al., 2016) [194]
Sucrose acetate isobutyrate	It is a complex mixture of esters formed by the reaction of sucrose with acetic and isobutyric acids	E444	ADI of 20 mg/kg body weight/day. High dosage may cause hepatic effects. Usage as a food additive at the permitted ADI dosage is not of safety concern.	(EFSA ANS Panel (EFSA Panel on Food Additives and Nutrient Sources added to Food, 2016) [195]

Table 3

Description of main emulsification technologies.

Emulsification technologies	Description	Equipment	Example	Reference
Mechanical agitation	Mechanical agitation is a common method used to create emulsions by applying mechanical energy to the mixture. This method involves the use of an external device, such as a mixer, blender, or homogenizer. The process involves subjecting the mixture to high shear forces, which help to break up the droplets and disperse them throughout the continuous phase	Stirring or mixing: a mixing device, such as a paddle, propeller, or anchor, is rotated in a vessel or tank. Rotational movement creates turbulence and shear forces. High-Speed Homogenization: rotor- stator mixers or high-shear mixers generate high shear forces by rapidly rotating a rotor within a stationary stator. Colloid Mills: They consist of a rotor and a stator with a narrow gap between them. The product is forced through the gap, where high shear forces and friction occur.	Fish oil loaded gelatine/surfactant- stabilized emulsions were obtained by homogenizing the disperse phase into the continuous phase using an ultra- turrax homogenizer at a speed of 8000 rpm and homogenizing time of 2 min. Emulsions with droplet sizes ranging from 4.02 to 15.51 µm were obtained.	(Zhang et al., 2020) [138]
High pressure homogenization	It is a mechanical process used to reduce the particle size and create a stable dispersion of components in a fluid, typically in the form of an emulsion. It involves subjecting the sample to high pressure and forcing it through a narrow gap or valve, resulting in intense shear forces and turbulence. This process facilitates the breakup of larger particles or droplets, leading to a reduction in size and improved homogenization	Piston-gap homogenizer: It consists of a reciprocating piston that forces the sample through a narrow gap or valve, generating high shear forces and turbulence. Valve homogenizer: The sample is pumped into the homogenizer and forced through a small orifice or series of valves. The sudden reduction in cross-sectional area at the valve creates high shear forces.	Clove essential oil-loaded sodium caseinate and gum Arabic emulsions were obtained via high pressure homogenization. Coarse emulsion was subjected to homogenization under different pressures (200, 400, 600, and 800 bar) and homogenization times (2.5, 5, 7.5, and 10 min). Droplet size ranged between 200 and 260 nm. Average droplet size decreased as homogenization pressure increased, reaching a maximum stability of 60 d	(Yin et al., 2023) [31]
Microfluidization	In this method, the emulsion is forced through microfluidic channels under high pressure, which causes the droplets to break up and disperse into the continuous phase. The microfluidic channels are designed with narrow and precise dimensions to provide a controlled and uniform distribution of shear forces, resulting in the formation of droplets with a narrow size distribution. The emulsion is usually passed through the microfluidic channels multiple times to further reduce the droplet size and improve the stability of the emulsion	Microfluidizer: It uses microchannels or chambers with micron-sized dimensions to exert intense shear and impact forces on the sample.	Most favourable conditions for the microfluidization of high oleic palm oil emulsion were studied. Pressure varied from 10000 psi to 20000 psi for 1, 2 and 3 cycles. Results showed that 20000 psi for 2 cycles avoided the coalescence effect, producing stable nanoemulsions. Average droplet size ranged between 163 and 2268 nm	(Ricaurte et al., 2016) [54]
Ultrasonication	Ultrasonication is a technology that involves the use of sound waves in a specific frequency range between 20 kHz and 10 MHz (above the range of human hearing) to produce a shear force and break up the droplets in the emulsion. Microbubbles will form, expand, and eventually collapse in the medium during this process, which is caused by alternating pressure changes, known as acoustic cavitation.	Ultrasonic bath: consists of a tank or bath filled with a liquid sample. It contains ultrasonic transducers that emit high-frequency sound waves into the liquid. The sample is immersed in the bath, and the ultrasonic waves cause cavitation and acoustic streaming. Ultrasonic Probes/Horns: They are handheld or mountable devices that are directly inserted into the liquid sample. When the generator is activated, the probe vibrates at a high frequency, producing ultrasonic waves that propagate through the liquid. The intense cavitation and shear forces generated by the probe result in particle cirae raduction	Different homogenization techniques were evaluated in the production of high oleic palm oil nanoliposomes. Coarse emulsion was homogenized using ultrasound (20 kHz), microfluidizer (500–1500 bar), and rotor stator (16000–26000 rpm). Results showed that ultrasound and microfluidization nanoliposomes presented the highest stability when loaded oil was maximized.	(Beltrán et al., 2020) [167]
Membrane emulsification	Membrane emulsification is a specialized emulsification technique that involves the use of porous membranes to create small and uniform droplets in the emulsion. It offers advantages, such as precise control over droplet size, high production efficiency, ability to work	A suitable membrane is chosen based on factors such as membrane material, pore size, and surface properties. The membrane should have a pore size smaller than the desired droplet size to ensure efficient droplet formation (Konovalova et al., 2023)	Influence of the emulsification techniques (membrane and ultrasound) on microparticles properties was evaluated. Droplet size obtained by membranes ranged between 154 and 52 μ m, while ultrasound produced 0.16 μ m size droplets. Even though ultrasound particles were larger than membrane	(Consoli et al., 2023) [23]

(continued on next page)

Table 3 (continued)

Emulsification technologies	Description	Equipment	Example	Reference
	with a wide range of materials, and protection of heat and shear sensitive ingredients (Konovalova et al., 2023)		emulsion particles, membrane technique had the highest encapsulation efficiency and resveratrol retention.	

down over time. The properties of surfactants can be adjusted by varying their concentration and type.

2.4.1. Natural surfactants

Natural surfactants are compounds derived from natural sources, such as plant extracts, proteins, and lipids, and are commonly used as emulsifiers in the food industry. They are preferred to synthetic surfactants because of their perceived safety, natural origin, and environmental sustainability [136].

Lecithin, a phospholipid found in egg yolk and soybeans, is one of the most commonly used natural surfactants in the food industry. Lecithin is primarily used to stabilise emulsions, particularly oil-in-water emulsions, by reducing the interfacial tension between the oil and water phases, showing good results compared to synthetic surfactants such as Tween 80 and Span 80 [137,138]. Several studies have used lecithin as a surfactant to stabilise emulsions [54,139–141].

Protein-based surfactants, such as casein and whey protein, are also widely used as emulsifiers in the food industry. Casein, a milk protein, is commonly used in dairy products, such as cheese and yoghurt. Both proteins exhibit excellent emulsifying properties and are often used to improve the texture and stability of oil-loaded emulsions [108,128,142–144].

Plant-based extracts, such as saponins, which are found in plants such as quinoa and spinach, are also used as natural surfactants in the food industry. Other plant-based surfactants include gum arabic (GA) [105,145,146] and pectin [120,147].

2.4.2. Synthetic surfactants

Synthetic surfactants are widely used as emulsifiers in the food industry owing to their high efficiency, versatility, and costeffectiveness. Synthetic surfactants are often produced from petroleum-based sources via chemical synthesis and can be tailored for specific applications and performance requirements [148]. Examples of the synthetic surfactants commonly used in the food industry are polysorbates, sorbitan esters, and polyglycerol esters [149]. Polysorbates, such as polysorbate 80, and sorbitan esters, such as Span 80 are widely used as emulsifiers [19,149–151]. Although synthetic surfactants are highly effective emulsifiers, their use in the food industry has raised concerns regarding their safety and environmental impact. Some synthetic surfactants exert adverse health effects, such as disruption of the endocrine system and allergic reactions. Additionally, synthetic surfactants are not biodegradable and can accumulate in the environment, posing a threat to ecosystems.

2.4.3. Recent advances in surfactants

New surfactants are constantly being developed and introduced into various industries. Recent examples include the following: Biosurfactants: Surfactants are produced by microorganisms, such as bacteria, fungi, and yeasts. Biosurfactants have attracted considerable attention because of their biodegradability, low toxicity, and unique properties. Biosurfactants are widely seen as a favourable replacement for their synthetically produced counterparts; as such, their applications and the proportion of the overall surfactant market they occupy have been increasing over the past 15 years [152]. Similar to their synthetic counterparts, biosurfactants have various chemical structures. They are produced by microorganisms grown on insoluble (oils, fossils, and hydrocarbons) and soluble (carbohydrates) substrates [153,154].

Gemini surfactants contain two hydrophilic head groups and two hydrophobic tail groups covalently attached by spacer groups [155,156]. Gemini surfactants have been found to have superior emulsification and wetting properties compared to conventional surfactants. They are used for applications such as enhanced oil recovery, detergency, and drug delivery. Different types of classifications for GS based on physicochemical aspects of the spacer and tail groups. One of the key features of gemini surfactants is their low critical micelle concentration (CMC), which is 100 times lower than that of single-chain surfactants. It is also more effective at reducing the interface/surface tension. GS was first used as a catalyst; however, new applications of this technology include the cosmetic industry and drug delivery systems [155].

Sustainable surfactants: With the increasing demand for sustainable products, there has been a focus on developing surfactants that are derived from renewable sources and have a low environmental impact. Examples include surfactants produced from vegetable oils, sugars, and proteins [31,123,125,132].

3. Emulsification technologies

3.1. Main emulsification technologies

Emulsification technologies play a crucial role in various industries by enabling the production of stable homogeneous emulsions with finely dispersed droplets. These technologies have advanced significantly and offer efficient and scalable methods for emulsion formation. Table 3 shows some of the most commonly used emulsification technologies. The emulsification technologies and their operational ranges are listed in Table 1.

Each emulsification technology has its advantages and limitations, and the choice depends on factors such as the desired droplet size, stability requirements, scalability, and properties of the materials involved. Understanding the principles and capabilities of these emulsification technologies allows the selection of the most suitable method for specific applications, leading to efficient and optimised emulsion production and the conservation of lipophilic functional ingredients.

3.2. Emulsion characterisation and stabilisation mechanisms

Emulsion characterisation involves the measurement and analysis of various physical and chemical properties such as droplet size, charge, stability, and composition. The techniques for emulsion characterisation included Microscopy, Particle size analysis, rheological analysis, electrical conductivity, chemical analysis, and ultrasonic attenuation spectroscopy (Table 4). These techniques can be used in combination to comprehensively characterise emulsions and are essential for understanding and optimising the performance of emulsions in various applications.

Multiple factors influence emulsion destabilization, such as temperature, pH, ionic strength, and emulsifier concentration, thorough emulsion characterization can provide valuable insights into potential stability issues and help guide adjustments to maintain emulsion integrity across different environments and conditions.

Emulsion destabilization is influenced by temperature, which plays a crucial role in reducing interfacial tension between phases and accelerating processes such as drop coalescence and flocculation. Increased temperature can lead to phase inversion, making the emulsion more sensitive to mechanical degradation and potentially causing the emulsion to transition to a microemulsion state. Additionally, pH level can impact interfacial tension and charge characteristics, affecting the stability of emulsifier films and increasing susceptibility to flocculation and coalescence. Ionic strength also plays a part in stabilising or destabilizing emulsions, depending on the types of surfactants used, while emulsifier concentration must be carefully managed to prevent excessive micelle formation and potential coalescence.

Finally, factors such as creaming, flocculation, Ostwald ripening, and phase separation may occur due to changes in these variables, affecting the stability of oil-in-water emulsions [120,122,157]. The first step in the separation of emulsions is the rising of the dispersed phase through the continuous phase. This is known as creaming and is a result of the density differential between the two phases. There are typically three mechanisms associated with creaming, but the process is usually governed by one. These mechanisms are gravitational drainage, packing and shrinkage of the droplet layer, and flocculation during the creaming process [158–160]. Flocculation is a process whereby droplets come together to form loose clumps or aggregates. These aggregates will then interact to form a more rigid, compact structure with a higher packing density than the primary droplet structure. The interaction between droplets is due to an attractive interaction potential, which is always a result of a free energy increase in the system [158–160]. Flocculation is driven by many mechanisms, including van der Waals forces, coalescence, bridging flocculation, and hydrophobic interaction. Ostwald ripening is a slow process that is driven by differences in droplet size. Due to Laplace's law, smaller droplets will have a higher solubility, resulting in the diffusion of material through the droplet surface into the continuous phase, and the diffusion of material from the continuous phase into the droplet. This leads to smaller droplets dissolving and the material precipitating in the region of the droplet

Table 4

Technique	Description
Microscopy	A technique used to observe the droplet size and morphology of emulsions under a microscope. The most used technologies include optical, electron, and atomic force microscopy. These techniques often provide information about structurally complex
	systems in a way that is easily understood by humans. Each microscopic technique operates on different physicochemical principles and can be used to examine different levels and types of structural organization [196].
Particle size analysis	This technique is used to measure the size distribution of the droplets in an emulsion. It can be performed using laser diffraction,
	the particle size in emulsions, and it can be used for a wide range of particle sizes. In laser diffraction, the sample is first dispersed
	in a liquid medium and then introduced into the instrument. Intensity of the scattered light at each angle is then measured and
	used to calculate the particle size distribution and stability of the sample. Nanoparticle tracking analysis is also used to measure and analyse the nanoparticles in various liquid samples. It is particularly valuable for characterizing and quantifying
	nanoparticles in the size range of approximately 10 nm to 1 μ m. It operates based on the principles of light scattering and
	Brownian motion similar to dynamic light scattering technique [197].
Rheological analysis	Rheology delves into the behaviours of material flow and deformation. This field explores viscosity, elasticity, and various
	attributes within emulsions. To maintain physical stability, interfacial rheology emerges as a crucial factor. Traditionally assessed
	through dilatational or shearing deformations, interfacial rheology wields significant influence. Dilatational measurements are
	more relevant for short-term stability, whereas interfacial shear rheology provides more valuable information on middle or long- term stability [198]
Electrical conductivity	Electrical conductivity measurements can be used to estimate the stability of an emulsion by monitoring the changes in
	conductivity due to changes in droplet size, concentration, and composition [54,167].
Chemical analysis	Chemical analysis can be used to determine the composition of emulsions, such as the concentration of oil, water, and surfactants,
	and to monitor changes in the composition of emulsions over time [196].
Ultrasonic attenuation	It is an interesting method to measure the distribution of droplet sizes in emulsions due to its ability to analyse viscous and
spectroscopy	optically transparent emuisions. This method depends on the ultrasound acoustic speed and viscosity coefficient in emuision stability and abanges in emulation dependent size (Alchaeff, Depletch, and Mercer 2020). [100]. In ultrasound accosities and the second s
	stability and changes in emusion in optic size (Assinati, Frankasi, and Mercer 2020) [199], in unasonic spectrum analysis,
	the concentration and size distribution of the colloidal particles [200]
	are concontration and once doubled of the contrating particles [200].

with a higher curvature. This results in larger droplets growing and smaller droplets shrinking. The difference in the rate of material movement in and out of droplets is what separates this process from coalescence [158–160]. Coalescence is a dynamic process influenced by many factors. It is generally agreed that, before two droplets can coalesce, they must come into close contact with one another. The force that brings droplets together is the disjoining pressure [158–160].

4. Emulsion stabilisation technologies

Several emulsion stabilisation techniques have been used to prevent phase separation and maintain the stability of the dispersed phase within the continuous phase. Common stabilisation techniques include surfactants, emulsifiers, rheological modifications, Pickering emulsions, and encapsulation technologies. Surfactants and emulsifiers, both known as emulsifiers, form a protective interfacial layer that is adsorbed on the surface of the droplets (reviewed in previous sections), whereas stabilisers tend to maintain the physicochemical state of the emulsion by modifying its rheology. By modifying the viscosity and rheological properties of the continuous phase, emulsions can be stabilized [161]. Thickening agents such as gums, starches, or cellulose derivatives can be added to increase the viscosity and provide resistance to droplet movement and coalescence [157].

Pickering emulsion stabilisation involves the adsorption of fine insoluble solid particles at the oil–water interface [161]. The stabilisation of pickering emulsions occurs by reducing the interfacial energy between two immiscible phases through the adsorption of solid particles on the surface of the droplets, providing excellent stability to the emulsions [126]. Good Pickering stabilisation can be achieved when the average size of the adsorbed solid particles is at least one order of magnitude smaller than that of a dispersed-phase droplet [161]. The particle-based stabilisation of fluid-in-fluid dispersions has been actively studied as an alternative to surfactant-based stabilisation [112,114,116,124,162–166]. Matos et al. (2018) compared the use of OSA-modified quinoa starch as a solid particle stabiliser with that of Tween 20, a common non-ionic surfactant. The results showed that Pickering emulsions were more stable against creaming than Tween 20 emulsions. Additionally, the Pickering emulsions exhibited higher encapsulation efficiency, reaching values of up to 98 % [162]. More than one solid particle source can be utilised to obtain better results, H. Yang et al. (2020) used a mixture of soy protein isolate and chitosan to stabilise emulsions, obtaining stable emulsions up to 20 days, stored at 4 °C. emulsions showed good storage stability and anti-coalescence ability [116]. Pickering emulsions are promising candidates for overcoming the adverse effects of classical emulsifiers on environment and human health [117].

In emulsion stabilisation, encapsulation technologies can be classified into two distinct categories: those that yield liquid-state products, such as micelles and liposomes, and those that result in solid-state presentations, such as drying technologies. First, liposomes are spherical vesicles characterised by a lipid bilayer encapsulating an aqueous core and hold a significant position in colloidal



Fig. 4. Microphotography of emulsions stabilized via A) spray drying (Used as per permission from Elsevier [23], B) freeze drying (Used as per permission from Elsevier [80]) C) refractance window drying (Used as per permission from Elsevier [141]), and D) pickering (Used as per permission from Springer Nature [166]).

systems. Their unique structure offers remarkable versatility for various applications, with a particular focus on the encapsulation of water-soluble hydrophilic substances [167]. The encapsulation process is facilitated by an aqueous core that serves as an ideal environment for housing such substances. Simultaneously, the lipid bilayer enveloping the aqueous core acts as a protective barrier, effectively segregating aqueous content from the external environment. In this context, liposomes have emerged as valuable tools for stabilising oil-in-water (O/W) emulsions [168]. Because of their lipid bilayers, liposomes can interact with and envelop oil droplets dispersed in an aqueous medium. This enveloping action effectively hindered the coalescence and merging of oil droplets, thereby contributing to the stability of the O/W emulsions. Secondly, the micelles exhibit a spherical or cylindrical morphology, comprising amphiphilic molecules. These molecules manifest as a hydrophobic moiety that congeals at the micelle core and a hydrophilic segment that resides on its surface. Micelles demonstrate enhanced suitability for stabilising oil-in-water (O/W) emulsions, primarily because the hydrophilic region of amphiphilic molecules is oriented towards the aqueous phase, while the hydrophobic component resides within the micelles, protecting the dispersed water droplets within the oil phase [26].

Encapsulation technologies such as drying can be employed to stabilise emulsions by removing the liquid phase and converting the emulsion into a solid or powdered form. This can enhance the stability, shelf-life, and ease of handling of the emulsions. Common drying techniques used for emulsion stabilisation include spray drying, freeze drying, and refractance window drying.

Spray drying is a widely used technique for converting liquid emulsions into powder form. Spray drying offers advantages, such as functional lipophilic ingredient protection, shelf-life extension, ease of storage or transport, and easy reconstitution with water or other suitable solvents when needed [23]. Several researchers have explored the use of spray drying as a stabilisation technique [23,65,67, 169]. When using spray drying to encapsulate and seal an emulsion, temperature is a very important factor to consider. Temperature might vary between 160 °C and 185 °C [65] and it will depend on the emulsion properties. X. H. Zhao & Tang. (2016) used a pea protein isolate, soy protein isolate, defatted milk powder, OSA starch, and spray drying to stabilise and encapsulate CoQ_{10} . Good encapsulation efficiency and redispersion behaviour were obtained [170].

Freeze-drying involves freezing the emulsion, followed by sublimation of the frozen water directly into vapour. This method preserves the structure of the emulsion while maintaining its original particle size and distribution. Freeze-dried emulsions exhibit improved stability, prolonged shelf-life, and the ability to be reconstituted by adding a suitable liquid. Marefati et al. (2015) [171] used a freeze-drying technique to dry multiple emulsions and obtained food-grade oil-filled powders from OSA-modified starch Pickering emulsions with an oil content of approximately 70 wt %. When working with multiple emulsions, heat might cause emulsion destabilization, which is why freeze-drying is a good alternative for drying these emulsions.

Although spray drying and freeze drying are the most commonly used technologies for stabilising emulsions through drying, in recent years, technologies that allow for cost reduction and drying time optimisation have emerged. Refractive window drying is based on the evaporation of moisture through the transfer of thermal energy from hot water to the material being dried, primarily through infrared radiation [172]. To date, few studies have been conducted on the use of refractance window technology to stabilise oil-in-water emulsions. In 2010, Cadwallader et al. [173], the use of a refractance window and spray drying were compared as technologies for the encapsulation of orange oil. The authors found that, for spray drying, the surface oil content increased with increasing inlet temperature, whereas for refractance window drying. Research on the stabilisation of high-oleic palm oil nanoemulsions using the refractance window method has been conducted by Hernandez-Carrión et al. [174], and Henao-Ardila et al. [141], from the same research group. High-oleic palm oil was encapsulated in whey protein and starch (cornstarch and OSA modifies starch, respectively). The first work focused on the design of nanoemulsions suitable for refractance window drying, obtaining optimal whey protein and corn starch concentrations. The second work focused on the effect of the wall material on lipophilic functional compounds, finding the concentration that maximized their conservation.

Emulsion electrospinning is a novel and straightforward technique for creating multifunctional packaging materials capable of effectively protecting loaded active substances during the production process. This method allows the preparation of emulsion-based gelatine edible films that can encapsulate, protect, and release bioactive compounds for food packaging applications. The process demonstrates significant potential for stabilising and delivering lipophilic functional ingredients in emulsions, offering enhanced



Fig. 5. Typical conditions for bioaccessibility evaluation of lipophilic functional ingredients.

Main evaluation results of the digestibility of lipophilic functional ingredients.

Lipophilic functional ingredient	Main results	Reference
β-carotene	After in vitro digestion, the bioaccessibility of β -carotene increased from 3.1 to 35.6 % after nanoencapsulation. Moreover, the modified starch used as a wall material led to higher retention but lower bioaccessibility of β -carotene.	(Liang et al., 2013) [57]
Tocopherol and total phenolic content	Digested nanoemulsions showed high lipid digestion (85.25 %) and bioaccessibility of antioxidants and a lower rate of phytosterol degradation compared to the digested untreated oil.	(Cheong et al., 2016) [184]
β-carotene	Bioaccessibility of β -carotene decreased in the order of LCT > MCT > orange oil. Bioaccessibility of β -carotene was very low (0 %) in orange oil nanoemulsions because mixed micelles were not formed to solubilize β -carotene but high in LCT nanoemulsions (66 %).	(Qian et al., 2012b) [66]
Astaxanthin	Encapsulated astaxanthin was more stable than non-encapsulated astaxanthin.	(Acevedo et al., 2014). [183]
β -carotene	All encapsulation structures, except those obtained from high-speed homogenization of whey protein concentrate emulsions, increased the bioaccessibility of β -carotene after in vitro digestion, which was insignificant in its free form.	(Gómez-Mascaraque et al., 2017) [51].

performance and versatility in food packaging. The use of edible packaging materials filled with functional lipophilic ingredients have shown to effectively inhibit the growth of bacteria in packed food [175], produce packaging bio films with antioxidant activity [111] and inhibit the growth of microorganism and prolong the shelf lives of foods [176].

Drying technologies are often used for the encapsulation of sensitive ingredients, but drying may also be considered a stabilisation technique because it removes the liquid phase, which can be a potential source of instability, and converts the emulsion into a more stable solid or powdered form [23]. Drying can also enhance the long-term storage stability of emulsions by reducing the water activity and inhibiting microbial growth. However, it is important to note that the choice of drying technology should consider the specific properties of the emulsion, such as its composition, viscosity, and thermal sensitivity. Optimisation of the drying process parameters, including temperature, residence time, and drying medium, is crucial for maintaining emulsion quality and stability during the drying process.

The final particle size and shape depend on the stabilisation technique. Micrographs of the final stabilized and encapsulated emulsions obtained using the four different technologies are shown in Fig. 4. Fig. 4A shows the resveratrol emulsion powder obtained by spray drying. Rounded soft particles of several sizes were obtained. Fig. 4B shows the freeze-dried emulsions. Several additional porous structures were obtained. The size depended on the post-milling operation. Fig. 4C shows an emulsion of highly oleic palm oil that was dried using a refractance window. Flakes of several sizes with homogeneous structures were obtained using this technique. Fig. 4D shows an O/W Pickering emulsion stabilized with cellulose. This is a liquid with an emulsion stabilized by solid particles. The drop size depended on the emulsification technique used. These stabilisation techniques can be used individually or in combination, depending on the specific emulsion system and desired properties. The choice of technique depends on factors such as the nature of the components, application, and desired shelf-life of the emulsion.

5. Digestibility of lipophilic functional ingredients

Food digestion is a complex and multistage process that has recently attracted the interest of the food industry owing to the development of links between food and health or diseases [177]. During human digestion, two main processes occur simultaneously: (i) mechanical transformations that reduce the size of food particles, and (ii) enzymatic transformations where macromolecules are hydrolysed into smaller constituents that are released from the food matrix during digestion to become available for absorption (bioaccessibility) and are therefore absorbed from the digestive tract and utilised by the body (bioavailability) [178].

To evaluate the bioavailability of phytonutrients of interest, food digestion can be studied using in vitro or in vivo models. In vitro digestion assays simulate the physiological conditions of the human gut and are useful tools for studying and understanding the changes, interactions, and bioavailability of nutrients. In vivo models refer to the process of food digestion that occurs within a living organism. Ideally, the study of food digestion should be conducted in vivo, considering the complexity of the gastrointestinal tract. However, obtaining data on the human gut is challenging. The variations observed from one individual to another, invasive techniques required, and ethical and financial aspects involved make in vivo testing almost impossible [179]. In vitro digestion helps overcome these drawbacks. Human gastrointestinal tract can be seen as 4 in series reactors, each with specific conditions of pH, residence time, enzymes and fluids which can be replicated in vitro models [180], Fig. 5 shows the relevant regions and their conditions typically simulated *in vitro* models.

In vitro models can be static, semidynamic, or dynamic. Static models simulate the stages of digestion using different solutions that mimic the physiological conditions of the gastrointestinal tract. Specific digestive enzymes such as amylase, pepsin, and pancreatic enzymes are used, and parameters such as pH and temperature are controlled. Food was mixed with the digestive solutions and incubated for specific periods to simulate oral, gastric, and intestinal digestion [180,181]. is the most widely used approach for assessing the bioaccessibility of compounds. They have simplified the highly complex processes of human digestion. Consequently, they can be used in most laboratories as they do not require complex equipment or analyst training. Examples of the static in vitro digestion methods used to evaluate the bioaccessibility of different compounds are presented inTable 5.

Although the simplicity of the static approach is a major advantage, it fails to capture the dynamic nature of in vivo digestive

processes. Various efforts have been made to study digestion under physiologically relevant conditions. The goal of this study was to provide physiologically relevant kinetic information on nutrient partitioning patterns and structural changes in food [180]. Therefore, the semi-dynamic in vitro technique involves a gradual pH change in the digestive tract, especially in the gastric phase, secretion of enzymes and digestive fluids, and gastric emptying, making the semi-dynamic technique suitable for investigating the structural changes in food during digestion, such as the effect on the breakdown of the food matrix and absorption of nutrients [179].

Although static models are simple and easy to use, they are inaccurate. It is difficult to recreate the entire dynamic environment of the human body and the flow of food through the different compartments in a real digestive system. Several dynamic models, such as the dynamic gastric model (DGM), human gastric simulator (HGS), gastric digestion simulator (GDS), dynamic gastrointestinal simulator (SIMGI) dynamic gastrointestinal digester (DIDGI), TIM, and simulator of the human intestinal microbial ecosystem (SHIME), have been explored [182]. Such models are much more complex and expensive, require large amounts of enzymes and samples, and necessitate the use of specific equipment that is not normally available in the laboratory.

6. Final remarks

The use of emulsions offers an effective approach for incorporating functional lipophilic compounds into food matrices, addressing the challenges posed by their sensitivity to environmental factors, processing conditions, and gastrointestinal environments. By encapsulating these compounds in emulsions, their protection from external stressors is enhanced, and their passage through the gastrointestinal tract is improved. This method increases solubility, facilitates controlled release, and ultimately boosts the bio-accessibility and bioavailability of these beneficial compounds. Emulsions therefore represent a promising strategy for maximizing the health benefits of lipophilic functional ingredients in food products. Future trends in the field may involve the development of more advanced and sustainable emulsification technologies, such as the use of natural emulsifiers and innovative encapsulation materials. Additionally, the integration of emulsified lipophilic compounds into novel food formats and delivery systems may expand their applicability and impact on consumer health. Further research into the optimisation of emulsion-based delivery systems will likely lead to improved stability, targeted release, and enhanced functionality, opening new opportunities for the incorporation of lipophilic functional ingredients in a wider range of food products.

Formulations should be considered to achieve an efficient emulsification process. Regarding wall materials, plant-based proteins have gained increasing interest in recent years because of their advantages over animal-based proteins. Additionally, it is recommended to employ more than one natural polymer in emulsion formulations, as they enhance emulsion stability by forming complexes among themselves. Blends of proteins and polysaccharides have shown positive results when functional lipophilic ingredients were encapsulated in emulsions.

Inclusion of digestion studies for emulsions is essential for developing effective functional encapsulation systems. It is crucial that protection systems for functional lipophilic ingredients not only safeguard them during processing but also ensure their resilience against the stomach environment. This allows the ingredients to reach areas where they can be absorbed by the digestive system and become bioavailable for use in various metabolic pathways within the human body. Conducting in vitro digestion studies on emulsions is a growing trend that can provide valuable insights into their stability and performance. This type of research is especially important for emulsions stabilized by drying techniques, offering a clearer understanding of how these systems function during digestion. Future research may focus on refining in vitro digestion models to better mimic human digestive processes and improving the predictability of in vivo outcomes. Additionally, combining in vitro digestion studies with other characterization techniques may provide a more comprehensive understanding of emulsion stability and behaviour in the digestive tract. Such advances will facilitate the development of more robust and effective encapsulation systems for functional lipophilic ingredients.

Emulsions are inherently thermodynamically unstable, making the study of technologies for their stabilisation a critical area of research. While the use of insoluble solid particles (Pickering emulsions) has been extensively explored, drying technologies offer an alternative approach by removing the aqueous phase, effectively preventing the movement of oil droplets and avoiding emulsion destabilization. Drying processes, which encapsulate emulsions by removing the liquid phase and preventing oil droplet coalescence, can also be considered as stabilisation methods. The exploration of innovative drying technologies, such as electrohydrodynamic drying, microwave-assisted drying, and infrared drying which offer rapid and uniform drying while maintaining product quality, holds considerable promise in emulsion stabilisation. An interesting option to stabilized and carrying lipophilic functional ingredients are the edible films. This application can be improved with technologies such as electro spinning.

Modifications to conventional drying methods such as the application of vacuum pressures in refractance window drying, the combination of different drying techniques such as convective drying and microwave drying can offer improved preservation of emulsion structure and bioactive content during drying. Given the central role of emulsions in various sectors such as food, cosmetics, and pharmaceuticals, improving their stability and shelf life remains a critical focus. These advanced drying techniques may contribute to achieving these objectives by minimizing heat damage and preserving sensitive bioactive ingredients. Their application in emulsion processing could result in safer, more effective, and more sustainable products. Continued investigation into these and other novel drying methods will play a key role in advancing the use of emulsions across industries and enhancing their overall performance.

Moreover, optimising operating conditions to minimize the degradation of functional compounds of interest will be crucial to ensuring the quality and efficacy of the final product. The challenge lies in balancing efficiency and cost-effectiveness with the preservation of the nutritional and bioactive properties of emulsions. As technology advances, there is potential for the emergence of new drying methods that meet these objectives and enhance the stability and functionality of emulsions for various applications.

Future research should prioritize exploring novel natural ingredients for encapsulating functional oils through emulsification, as this is increasingly important in the modern food and pharmaceutical industries. With consumer preferences trending towards clean

A. Henao-Ardila et al.

labels and sustainable practices, the search for effective nature-derived encapsulants has gained significant importance. These ingredients not only provide enhanced stability and controlled release of bioactive compounds but also align with the rising demand for healthier and more eco-friendly products.

CRediT authorship contribution statement

Alejandra Henao-Ardila: Writing – original draft, Investigation, Formal analysis. María Ximena Quintanilla-Carvajal: Writing – review & editing, Formal analysis, Conceptualization. Fabián Leonardo Moreno: Writing – review & editing, Supervision, Funding acquisition, Formal analysis, Conceptualization.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This research was funded by the Universidad de La Sabana under project ING-206-2018. The first author thanks Universidad de La Sabana for the grant awarded for her doctoral studies.

References

- United Nations Department of Economic and Social Affairs Population Division, World population prospects 2019. http://www.ncbi.nlm.nih.gov/pubmed/ 12283219, 2019.
- [2] C. Santos-Buelga, A.M. González-Paramás, T. Oludemi, B. Ayuda-Durán, S. González-Manzano, Plant Phenolics as Functional Food Ingredients, first ed., Elsevier Inc., 2019 https://doi.org/10.1016/bs.afnr.2019.02.012.
- [3] M. Ashwell, Concepts of functional foods, in: IISI Europe Concise Monograph Series, Belgium, 2002, pp. 1–35. http://fst.sagepub.com/content/3/4/306.2.full. pdf. (Accessed 14 September 2019).
- [4] O.F. Vázquez-Vuelvas, F.A. Chávez-Camacho, J.A. Meza-Velázquez, E. Mendez-Merino, M.M. Ríos-Licea, J.C. Contreras-Esquivel, A comparative FTIR study for supplemented agavin as functional food, Food Hydrocolloids 103 (2020), https://doi.org/10.1016/j.foodhyd.2020.105642.
- [5] S.S. Fernandes, M.S. Coelho, M. de las, M. Salas-Mellado, Bioactive Compounds as Ingredients of Functional Foods, Elsevier Inc., 2019, https://doi.org/ 10.1016/b978-0-12-814774-0.00007-4.
- [6] K. Szabo, B. Emőke Teleky, F. Ranga, E. Simon, O. Lelia Pop, V. Babalau-Fuss, N. Kapsalis, D. Cristian Vodnar, Bioaccessibility of microencapsulated carotenoids, recovered from tomato processing industrial by-products, using in vitro digestion model, Lebensm. Wiss. Technol. 152 (2021) 112285, https:// doi.org/10.1016/J.LWT.2021.112285.
- [7] K. Joshi, P. Kumar, R. Kataria, Microbial carotenoid production and their potential applications as antioxidants: a current update, Process Biochem. 128 (2023) 190–205, https://doi.org/10.1016/j.procbio.2023.02.020.
- [8] U. Akbar, M. Yang, D. Kurian, C. Mohan, Omega-3 fatty acids in rheumatic diseases a critical review, J. Clin. Rheumatol. 23 (2017) 330–339, https://doi.org/ 10.1097/RHU.000000000000563.
- [9] C. Parolini, The role of marine n-3 polyunsaturated fatty acids in inflammatory-based disease: the case of rheumatoid arthritis, Mar. Drugs 22 (2024) 17, https://doi.org/10.3390/MD22010017, 22 (2023) 17.
- [10] U.K. Barua, P. Kar-Maker, A.K. Saha, M.M. Mostofa, D.K. Ghosh, K.K. Biswas, The effects of omega-3 fatty acids supplementation in bronchial asthma, Journal of Biosciences and Medicines How To 11 (2023) 208–219, https://doi.org/10.4236/jbm.2023.114015.
- [11] A. Azzi, Many tocopherols, one vitamin E, Mol. Aspect. Med. 61 (2018) 92-103, https://doi.org/10.1016/j.mam.2017.06.004.
- [12] A. Reddy Konda, M. Gelli, C. Pedersen, R.E. Cahoon, C. Zhang, T. Obata, E.B. Cahoon, Vitamin E biofortification: maximizing oilseed tocotrienol and total vitamin E tocochromanol production by use of metabolic bypass combinations, Metab. Eng. 79 (2023) 1096–7176, https://doi.org/10.1016/j. ymben.2023.06.011.
- [13] H.Y. Peh, W.S.D.D. Tan, W. Liao, W.S.F.F. Wong, H. Yong, W.S.D.D. Tan, W. Liao, W.S.F.F. Wong, H.Y. Peh, W.S.D.D. Tan, W. Liao, W.S.F.F. Wong, Vitamin E therapy beyond cancer: tocopherol versus tocotrienol, Pharmacol. Ther. 162 (2016) 152–169, https://doi.org/10.1016/j.pharmthera.2015.12.003.
- [14] A. Azzi, Tocopherols, tocotrienols and tocomonoenols: many similar molecules but only one vitamin E, Redox Biol. 26 (2019), https://doi.org/10.1016/j. redox.2019.101259.
- [15] A.M. Ribeiro, B.N. Estevinho, F. Rocha, Improvement of vitamin E microencapsulation and release using different biopolymers as encapsulating agents, Food Bioprod. Process. 130 (2021) 23–33, https://doi.org/10.1016/J.FBP.2021.08.008.
- [16] J. Zhu, Q. Huang, Nanoencapsulation of Functional Food Ingredients, first ed., Elsevier Inc., 2019 https://doi.org/10.1016/bs.afnr.2019.03.005.
- [17] A. Shi, J. Wang, R. Guo, X. Feng, Y. Ge, H. Liu, D. Agyei, Q. Wang, Improving resveratrol bioavailability using water-in-oil-in-water (W/O/W) emulsion: physicochemical stability, in vitro digestion resistivity and transport properties, J. Funct.Foods 87 (2021) 1756–4646, https://doi.org/10.1016/j. jff.2021.104717.
- [18] Z. Ren, Z. Chen, Y. Zhang, X. Lin, W. Weng, B. Li, Characteristics and in Vitro Digestion of Resveratrol Encapsulated in Pickering Emulsions Stabilized by Tea Water-Insoluble Protein Nanoparticles, 2023, https://doi.org/10.1016/j.fochx.2023.100642.
- [19] N. Luo, A. Ye, F.M. Wolber, H. Singh, Digestion behaviour of capsaicinoid-loaded emulsion gels and bioaccessibility of capsaicinoids: effect of emulsifier type, Curr. Res. Food Sci. 6 (2023) 100473, https://doi.org/10.1016/J.CRFS.2023.100473.
- [20] D.T. Piorkowski, D.J. McClements, Beverage emulsions: recent developments in formulation, production, and applications, Food Hydrocolloids 42 (2014) 5–41, https://doi.org/10.1016/j.foodhyd.2013.07.009.
- [21] B.S.T. Barbosa, E.E. Garcia-Rojas, Double emulsions as delivery systems for iron: stability kinetics and improved bioaccessibility in infants and adults, Curr. Res. Food Sci. 5 (2022) 718–725, https://doi.org/10.1016/J.CRFS.2022.04.003.
- [22] S. Sevdin, B. Ozel, U. Yucel, M.H. Oztop, H. Alpas, High hydrostatic pressure induced changes on palm stearin emulsions, J. Food Eng. 229 (2018) 65–71, https://doi.org/10.1016/j.jfoodeng.2017.10.007.
- [23] L. Consoli, M. Dupas Hubinger, M.M. Dragosavac, Encapsulation of resveratrol via spray-drying of oil-in-water emulsions produced by ultrasound or membrane emulsification, J. Food Eng. 350 (2023) 111488, https://doi.org/10.1016/j.jfoodeng.2023.111488.
- [24] M. Balakrishnan, S. Gayathiri, P. Preetha, R. Pandiselvam, G. Jeevarathinam, D.S.A. Delfiya, A. Kothakota, Microencapsulation of bixin pigment by spray drying: evaluation of characteristics, LWT–Food Sci. Technol. 145 (2021) 111343, https://doi.org/10.1016/j.lwt.2021.111343.
- [25] D. Nkurunziza, S.P. Sivagnanam, J.-S. Park, Y.-J. Cho, B.S. Chun, Effect of wall materials on the spray drying encapsulation of brown seaweed.pdf, Algal Res. 102381 (2021) 1–10.

- [26] S.L. Mulrooney, G.J. O'neill, D.F. Brougham, D. O'riordan, Enhancing the bioaccessibility of vitamin D using mixed micelles-An in vitro study. https://doi.org/ 10.1016/j.foodchem.2022.133634, 2022.
- [27] M. Iddir, C. Degerli, G. Dingeo, C. Desmarchelier, T. Schleeh, P. Borel, Y. Larondelle, T. Bohn, Whey protein isolate modulates beta-carotene bioaccessibility depending on gastro-intestinal digestion conditions, Food Chem. (2019) 157–166, https://doi.org/10.1016/j.foodchem.2019.04.003.
- [28] M. Saifullah, M.R.I. Shishir, R. Ferdowsi, M.R. Tanver Rahman, Q. Van Vuong, Micro and nano encapsulation, retention and controlled release of flavor and aroma compounds: a critical review, Trends Food Sci. Technol. 86 (2019) 230–251, https://doi.org/10.1016/j.tifs.2019.02.030.
- [29] A.M. Bakry, S. Abbas, B. Ali, H. Majeed, M.Y. Abouelwafa, A. Mousa, L. Liang, Microencapsulation of oils: a comprehensive review of benefits, techniques, and applications, Compr. Rev. Food Sci. Food Saf. 15 (2016) 143–182, https://doi.org/10.1111/1541-4337.12179.
- [30] Q.-Q. Yang, Z. Sui, W. Lu, H. Corke, Soybean lecithin-stabilized oil-in-water (O/W) emulsions increase the stability and in vitro bioaccessibility of bioactive nutrients, Food Chem. 338 (2021) 128071, https://doi.org/10.1016/i.foodchem.2020.128071.
- [31] X. Yin, Q. Hu, X. Chen, S. Tan, A. Niu, W. Qiu, G. Wang, Inclusion complexes of clove essential oil with sodium caseinate and gum Arabic prepared by highpressure homogenization: characterization and non-contact antimicrobial activity, Food Control 150 (2023) 109765, https://doi.org/10.1016/J. FOODCONT.2023.109765.
- [32] M. Mondéjar-López, A. Rubio-Moraga, A. José López-Jimenez, J.C. García Martínez, O. Ahrazem, L. Gómez-Gómez, E. Niza, Chitosan nanoparticles loaded with garlic essential oil: a new alternative to tebuconazole as seed dressing agent, Carbohydr. Polym. 277 (2022) 1–10, https://doi.org/10.1016/j. carbool.2021.118815.
- [33] J. Liu, F. Song, R. Chen, G. Deng, Y. Chao, Z. Yang, H. Wu, M. Bai, P. Zhang, Y. Hu, Effect of cellulose nanocrystal-stabilized cinnamon essential oil Pickering emulsions on structure and properties of chitosan composite films, Carbohydr. Polym. 275 (2022), https://doi.org/10.1016/j.carbpol.2021.118704.
- [34] M. Hadidi, A. Motamedzadegan, A.Z. Jelyani, S. Khashadeh, Nanoencapsulation of hyssop essential oil in chitosan-pea protein isolate, LWT Food Sci. Technol. (Lebensmittel-Wissenschaft -Technol.) 144 (2021) 1–9.
- [35] Y. Premjit, J. Mitra, Synthesis, characterization, and in vitro digestion of electrosprayed and freeze-dried probiotics encapsulated in soy protein isolatesunflower oil emulsions, Food Biosci. 53 (2023) 2212–4292, https://doi.org/10.1016/j.fbio.2023.102532.
- [36] P.A. Loyeau, M.J. Spotti, G. Vinderola, C.R. Carrara, Encapsulation of potential probiotic and canola oil through emulsification and ionotropic gelation, using protein/polysaccharides Maillard conjugates as emulsifiers, Lebensm. Wiss. Technol. 150 (2021) 111980, https://doi.org/10.1016/J.LWT.2021.111980.
- [37] H. Gao, L. Ma, W. Sun, D.J. McClements, C. Cheng, H. Zeng, L. Zou, W. Liu, Impact of encapsulation of probiotics in oil-in-water high internal phase emulsions on their thermostability and gastrointestinal survival, Food Hydrocolloids 126 (2022) 107478, https://doi.org/10.1016/J.FOODHYD.2021.107478.
 [38] F.Y. Ushikubo, R.L. Cunha, Stability mechanisms of liquid water-in-oil emulsions, Food Hydrocolloids 34 (2014) 145–153, https://doi.org/10.1016/J.
- FOODHYD.2012.11.016.
 [39] D. Amadei, M.D. Chatzidaki, J. Devienne, J. Monteil, M. Cansell, A. Xenakis, F. Leal-Calderon, Low shear-rate process to obtain transparent W/O fine
- [39] D. Amadei, M.D. Chatzidaki, J. Devienne, J. Monteil, M. Cansell, A. Xenakis, F. Leal-Calderon, Low shear-rate process to obtain transparent W/O fine emulsions as functional foods, Food Res. Int. 62 (2014) 533–540, https://doi.org/10.1016/j.foodres.2014.03.069.
- [40] K. Wijarnprecha, A. de Vries, P. Santiwattana, S. Sonwai, D. Rousseau, Rheology and structure of oleogelled water-in-oil emulsions containing dispersed aqueous droplets as inactive fillers, Lwt 115 (2019), https://doi.org/10.1016/j.lwt.2019.04.068.
- [41] S.Z. Luo, X.F. Hu, L.H. Pan, Z. Zheng, Y.Y. Zhao, L.L. Cao, M. Pang, Z.G. Hou, S.T. Jiang, Preparation of camellia oil-based W/O emulsions stabilized by tea polyphenol palmitate: structuring camellia oil as a potential solid fat replacer, Food Chem. 276 (2019) 209–217, https://doi.org/10.1016/j. foodchem.2018.09.161.
- [42] J. Liu, M. Kharat, Y. Tan, H. Zhou, J.L. Muriel Mundo, D.J. McClements, Impact of fat crystallization on the resistance of W/O/W emulsions to osmotic stress: potential for temperature-triggered release, Food Res. Int. 134 (2020) 109273, https://doi.org/10.1016/J.FOODRES.2020.109273.
- [43] C. Dima, S. Dima, Bioaccessibility study of calcium and vitamin D3 co-microencapsulated in water-in-oil-in-water double emulsions, Food Chem. 303 (2020), https://doi.org/10.1016/j.foodchem.2019.125416.
- [44] J. He, X. Wu, Y. Xie, Y. Gao, D.J. McClements, L. Zhang, L. Zou, W. Liu, Capsaicin encapsulated in W/O/W double emulsions fabricated via ethanol-induced pectin gelling: improvement of bioaccessibility and reduction of irritation, Int. J. Biol. Macromol. 235 (2023) 123899, https://doi.org/10.1016/J. LJBIOMAC.2023.123899.
- [45] D. Li, M. Hu, L. Hou, Y. Gao, Z. Tian, W. Wen, B. Fan, S. Li, F. Wang, The structural and functional properties of soybean protein-polyglutamic acid complex effected the stability of W/O/W emulsion encapsulated Nattokinase, Food Chem. 414 (2023) 135724, https://doi.org/10.1016/J.FOODCHEM.2023.135724.
- [46] M. Chouaibi, J. Mejri, L. Rezig, K. Abdelli, S. Hamdi, Experimental study of quercetin microencapsulation using water-in-oil-in-water (W1/O/W2) double emulsion, J. Mol. Liq. 273 (2019) 183–191, https://doi.org/10.1016/j.molliq.2018.10.030.
- [47] A. Matalanis, O.G. Jones, D.J. McClements, Structured biopolymer-based delivery systems for encapsulation, protection, and release of lipophilic compounds, Food Hydrocolloids 25 (2011) 1865–1880, https://doi.org/10.1016/j.foodhyd.2011.04.014.
- [48] Z. Fang, Wusigale, H. Bao, Y. Ni, N. Choijilsuren, L. Liang, Partition and digestive stability of α-tocopherol and resveratrol/naringenin in whey protein isolate emulsions, Int. Dairy J. 93 (2019) 116–123, https://doi.org/10.1016/j.idairyj.2019.01.017.
- [49] Y.J. Jo, Y.J. Kwon, Characterization of β-carotene nanoemulsions prepared by microfluidization technique, Food Sci. Biotechnol. 23 (2014) 107–113, https:// doi.org/10.1007/s10068-014-0014-7.
- [50] L. Zhong, N. Ma, Y. Wu, L. Zhao, G. Ma, F. Pei, Q. Hu, Gastrointestinal fate and antioxidation of β-carotene emulsion prepared by oat protein isolate-Pleurotus ostreatus β-glucan conjugate, Carbohydr. Polym. 221 (2019) 10–20, https://doi.org/10.1016/j.carbpol.2019.05.085.
- [51] L.G. Gómez-Mascaraque, R. Perez-Masiá, R. González-Barrio, M.J. Periago, A. López-Rubio, Potential of microencapsulation through emulsion-electrospraying to improve the bioaccesibility of β-carotene, Food Hydrocolloids 73 (2017) 1–12, https://doi.org/10.1016/j.foodhyd.2017.06.019.
- [52] A.H. Saberi, Y. Fang, D.J. McClements, Fabrication of vitamin E-enriched nanoemulsions by spontaneous emulsification: effect of propylene glycol and ethanol on formation, stability, and properties, Food Res. Int. 54 (2013) 812–820, https://doi.org/10.1016/j.foodres.2013.08.028.
- [53] Y. Yang, D.J. McClements, Encapsulation of vitamin E in edible emulsions fabricated using a natural surfactant, Food Hydrocolloids 30 (2013) 712–720, https://doi.org/10.1016/j.foodhyd.2012.09.003.
- [54] L. Ricaurte, M.D.J. Perea-Flores, A. Martinez, M.X. Quintanilla-Carvajal, Production of high-oleic palm oil nanoemulsions by high-shear homogenization (microfluidization), Innovative Food Sci. Emerging Technol. 35 (2016) 75–85, https://doi.org/10.1016/j.ifset.2016.04.004.
- [55] T. Winuprasith, P. Khomein, W. Mitbumrung, M. Suphantharika, A. Nitithamyong, D.J. McClements, Encapsulation of vitamin D3 in pickering emulsions stabilized by nanofibrillated mangosteen cellulose: impact on in vitro digestion and bioaccessibility, Food Hydrocolloids 83 (2018) 153–164, https://doi.org/ 10.1016/j.foodhyd.2018.04.047.
- [56] K. Ziani, Y. Fang, D.J. McClements, Encapsulation of functional lipophilic components in surfactant-based colloidal delivery systems: vitamin E, vitamin D, and lemon oil, Food Chem. 134 (2012) 1106–1112, https://doi.org/10.1016/j.foodchem.2012.03.027.
- [57] R. Liang, C.F. Shoemaker, X. Yang, F. Zhong, Q. Huang, Stability and bioaccessibility of β-carotene in nanoemulsions stabilized by modified starches, J. Agric. Food Chem. 61 (2013) 1249–1257, https://doi.org/10.1021/if303967f.
- [58] E.S.M. Abdel-Aal, J.C. Young, Carotenoids, in: HEALTHGRAIN Methods: Analysis of Bioactive Components in Small Grain Cereals, Elsevier Inc., 2009, pp. 69–94, https://doi.org/10.1016/B978-1-891127-70-0.50011-X.
- [59] D.J. Mcclements, E.A. Decker, Y. Park, J. Weiss, Structural design principles for delivery of bioactive components in nutraceuticals and functional foods. https://doi.org/10.1080/10408390902841529, 2009.
- [60] V.O. Hunsu, C.O.B. Facey, J.Z. Fields, B.M. Boman, Retinoids as chemo-preventive and molecular-targeted anti-cancer therapies, Int. J. Mol. Sci. 22 (2021) 7731, https://doi.org/10.3390/IJMS22147731, 22 (2021) 7731.
- [61] B. Ramchatesingh, A. Martínez Villarreal, D. Arcuri, F. Lagacé, S.A. Setah, F. Touma, F. Al-Badarin, I.V. Litvinov, The use of retinoids for the prevention and treatment of skin cancers: an updated review, Int. J. Mol. Sci. 23 (2022) 12622, https://doi.org/10.3390/IJMS232012622, 23 (2022) 12622.
- [62] G.F. Combs, J.P. McClung, Vitamin A. https://doi.org/10.1016/B978-0-12-802965-7.00006-X, 2017.

- [63] B. Niu, P. Shao, P. Sun, Ultrasound-assisted emulsion electrosprayed particles for the stabilization of β-carotene and its nutritional supplement potential, Food Hydrocolloids 102 (2020), https://doi.org/10.1016/j.foodhyd.2019.105634.
- [64] D. Fu, S. Deng, D.J. McClements, L. Zhou, L. Zou, J. Yi, C. Liu, W. Liu, Encapsulation of β-carotene in wheat gluten nanoparticle-xanthan gum-stabilized Pickering emulsions: enhancement of carotenoid stability and bioaccessibility, Food Hydrocolloids 89 (2019) 80–89, https://doi.org/10.1016/j. foodhyd.2018.10.032.
- [65] S. Fang, X. Zhao, Y. Liu, X. Liang, Y. Yang, Fabricating multilayer emulsions by using OSA starch and chitosan suitable for spray drying: application in the encapsulation of β-carotene, Food Hydrocolloids 93 (2019) 102–110, https://doi.org/10.1016/j.foodhyd.2019.02.024.
- [66] C. Qian, E.A. Decker, H. Xiao, D.J. McClements, Inhibition of β-carotene degradation in oil-in-water nanoemulsions: influence of oil-soluble and water-soluble antioxidants, Food Chem. 135 (2012) 1036–1043, https://doi.org/10.1016/j.foodchem.2012.05.085.
- [67] C.P. Coronel-Aguilera, M.F. San Martín-González, Encapsulation of spray dried b-carotene emulsion by fluidized bed coating technology, Food Sci. Technol. (2015) 187–193, https://doi.org/10.1016/j.lwt.2014.12.036.
- [68] D.J. Mustacich, R.S. Bruno, M.G. Traber, Vitamin E, in: G. Litwack (Ed.), Vitam. Horm. (2007) 1-21.
- [69] S. Mayer, J. Weiss, D.J. McClements, Behavior of vitamin E acetate delivery systems under simulated gastrointestinal conditions: lipid digestion and bioaccessibility of low-energy nanoemulsions, J. Colloid Interface Sci. 404 (2013) 215–222, https://doi.org/10.1016/j.jcis.2013.04.048.
- [70] J. Lachman, A. Hejtmánková, M. Orsák, M. Popov, P. Martinek, Tocotrienols and tocopherols in colored-grain wheat, tritordeum and barley Jaromír, Food Chem. 240 (2017) 670–678, https://doi.org/10.1016/j.foodchem.2017.07.151.
- [71] G.F. Combs, J.P. McClung, Vitamin E. https://doi.org/10.1016/B978-0-12-802965-7.00008-3, 2017.
- [72] R. Lakhan, M. Sharma, K. Batra, F.B. Beatty, The role of vitamin E in slowing down mild cognitive impairment: a narrative review, Healthcare 9 (2021), https://doi.org/10.3390/HEALTHCARE9111573.
- [73] R.G. Saratale, H.S. Lee, Y.E. Koo, G.D. Saratale, Y.J. Kim, J.Y. Imm, Y. Park, Absorption kinetics of vitamin E nanoemulsion and green tea microstructures by intestinal in situ single perfusion technique in rats, Food Res. Int. 106 (2018) 149–155, https://doi.org/10.1016/j.foodres.2017.12.076.
- [74] I. Sundl, M. Murkovic, D. Bandoniene, B.M. Winklhofer-Roob, Vitamin E content of foods: comparison of results obtained from food composition tables and HPLC analysis, Clin. Nutr. 26 (2007) 145–153, https://doi.org/10.1016/j.clnu.2006.06.003.
- [75] F. Gallotti, V. Lavelli, C. Turchiuli, Application of Pleurotus ostreatus β-glucans for oil-in-water emulsions encapsulation in powder, Food Hydrocolloids 105 (2020), https://doi.org/10.1016/j.foodhyd.2020.105841.
- [76] J. Hategekimana, K.G. Masamba, J. Ma, F. Zhong, Encapsulation of vitamin E: effect of physicochemical properties of wall material on retention and stability, Carbohydr. Polym. 124 (2015) 172–179, https://doi.org/10.1016/j.carboh.2015.01.060.
- [77] V. Raikos, Encapsulation of vitamin E in edible orange oil-in-water emulsion beverages: influence of heating temperature on physicochemical stability during chilled storage, Food Hydrocolloids 72 (2017) 155–162, https://doi.org/10.1016/j.foodhyd.2017.05.027.
- [78] I.C.O. Neves, S.H. Silva, N.L. Oliveira, A.M.T. Lago, N. Ng, A. Sultani, P.H. Campelo, L.A.A. Veríssimo, J.V. de Resende, M.A. Rogers, Effect of carrier oil on α-tocopherol encapsulation in ora-pro-nobis (Pereskia aculeata Miller) mucilage-whey protein isolate microparticles, Food Hydrocolloids 105 (2020), https:// doi.org/10.1016/j.foodhyd.2020.105716.
- [79] P. Kaushik, K. Dowling, C.J. Barrow, B. Adhikari, Microencapsulation of omega-3 fatty acids: a review of microencapsulation and characterization methods, J. Funct.Foods 19 (2015) 868–881, https://doi.org/10.1016/j.jff.2014.06.029.
- [80] A.G. Abdel-Razek, M.M. Hassanein, B. Ozçelik, D.A. Baranenko, T.M. El-Messery, Omega fatty acid-balanced oil formula and enhancing its oxidative stability by encapsulation with whey protein concentrate. https://doi.org/10.1016/j.fbio.2022.101975, 2022.
- [81] M.J. MacArtney, G.E. Peoples, P.L. McLennan, Cardiac arrhythmia prevention in ischemia and reperfusion by low-dose dietary fish oil supplementation in rats, J. Nutr. 150 (2020) 3086–3093, https://doi.org/10.1093/JN/NXAA256.
- [82] C.E. Gumus, E.A. Decker, D.J. McClements, Gastrointestinal fate of emulsion-based ω-3 oil delivery systems stabilized by plant proteins: lentil, pea, and faba bean proteins, J. Food Eng. 207 (2017) 90–98, https://doi.org/10.1016/j.jfoodeng.2017.03.019.
- [83] S. Raeisi, S.M. Ojagh, S.Y. Quek, P. Pourashouri, F. Salaün, Nano-encapsulation of fish oil and garlic essential oil by a novel composition of wall material: Persian gum-chitosan, LWT - Food Sci. Technol. (Lebensmittel-Wissenschaft -Technol.) 116 (2019), https://doi.org/10.1016/j.lwt.2019.108494.
- [84] J. Vicente, T. de Souza Cezarino, L.J.B. Pereira, E.P. da Rocha, G.R. Sá, O.D. Gamallo, M.G. de Carvalho, E.E. Garcia-Rojas, Microencapsulation of sacha inchi oil using emulsion-based delivery systems, Food Res. Int. 99 (2017) 612–622, https://doi.org/10.1016/j.foodres.2017.06.039.
- [85] Y. Zhang, F. Zhou, X. Zeng, P. Shen, D. Yuan, M. Zhong, Q. Zhao, M. Zhao, pH-driven-assembled soy peptide nanoparticles as particulate emulsifier for oil-inwater Pickering emulsion and their potential for encapsulation of vitamin D3, Food Chem. 383 (2022) 132489, https://doi.org/10.1016/J. FOODCHEM 2022 132489
- [86] H. Kweder, H. Eidi, Vitamin D deficiency in elderly: risk factors and drugs impact on vitamin D status, Avicenna J Med 8 (2018) 139, https://doi.org/10.4103/ AJM.AJM 20 18.
- [87] V.K. Maurya, K. Bashir, M. Aggarwal, Vitamin D microencapsulation and fortification: trends and technologies, J. Steroid Biochem. Mol. Biol. 196 (2020), https://doi.org/10.1016/j.jsbmb.2019.105489.
- [88] X. Luo, L.T. Lim, Curcumin-loaded electrospun nonwoven as a colorimetric indicator for volatile amines, Lwt 128 (2020), https://doi.org/10.1016/j. lwt.2020.109493.
- [89] M. Kharat, M. Skrzynski, E.A. Decker, D.J. McClements, Enhancement of chemical stability of curcumin-enriched oil-in-water emulsions: impact of antioxidant type and concentration, Food Chem. 320 (2020), https://doi.org/10.1016/j.foodchem.2020.126653.
- [90] K. Tai, M. Rappolt, X. He, Y. Wei, S. Zhu, J. Zhang, L. Mao, Y. Gao, F. Yuan, Effect of β-sitosterol on the curcumin-loaded liposomes: vesicle characteristics, physicochemical stability, in vitro release and bioavailability, Food Chem. 293 (2019) 92–102, https://doi.org/10.1016/j.foodchem.2019.04.077.
- [91] J. Han, F. Chen, C. Gao, Y. Zhang, X. Tang, Environmental stability and curcumin release properties of Pickering emulsion stabilized by chitosan/gum Arabic nanoparticles, Int. J. Biol. Macromol. 157 (2020) 202–211, https://doi.org/10.1016/j.ijbiomac.2020.04.177.
- [92] H. Cheng, X. Chang, H. Luo, H. Tang, L. Chen, L. Liang, Co-encapsulation of resveratrol in fish oil microcapsules optimally stabilized by enzyme-crosslinked whey protein with gum Arabic. https://doi.org/10.1016/j.colsurfb.2023.113172, 2023.
- [93] T. Xu, Z. Gu, L. Cheng, C. Li, Z. Li, Y. Hong, Stability, oxidizability, and topical delivery of resveratrol encapsulated in octenyl succinic anhydride starch/ chitosan complex-stabilized high internal phase Pickering emulsions, Carbohydr. Polym. 305 (2023), https://doi.org/10.1016/j.carbpol.2023.120566.
- [94] M. Sessa, M.L. Balestrieri, G. Ferrari, L. Servillo, D. Castaldo, N. D'onofrio, F. Donsì, R. Tsao, Bioavailability of encapsulated resveratrol into nanoemulsionbased delivery systems. https://doi.org/10.1016/j.foodchem.2013.09.088, 2013.
- [95] I. Cirilli, E. Damiani, P.V. Dludla, I. Hargreaves, F. Marcheggiani, L.E. Millichap, P. Orlando, S. Silvestri, L. Tiano, Role of coenzyme q10 in health and disease: an update on the last 10 years (2010–2020), Antioxidants 10 (2021), https://doi.org/10.3390/antiox10081325.
- [96] S. Sifuentes-Franco, D.C. Sánchez-Macías, S. Carrillo-Ibarra, J.J. Rivera-Valdés, L.Y. Zuñiga, V.A. Sánchez-López, Antioxidant and anti-inflammatory effects of coenzyme Q10 supplementation on infectious diseases, Healthcare (Switzerland) 10 (2022), https://doi.org/10.3390/healthcare10030487.
- [97] A. Hidalgo-Gutiérrez, P. González-García, M.E. Díaz-Casado, E. Barriocanal-Casado, S. López-Herrador, C.M. Quinzii, L.C. López, Metabolic targets of coenzyme q10 in mitochondria, Antioxidants 10 (2021), https://doi.org/10.3390/antiox10040520.
- [98] Y. Sato, S. Yokoyama, Y. Yamaki, Y. Nishimura, M. Miyashita, S. Maruyama, Y. Takekuma, M. Sugawara, Enhancement of Intestinal Absorption of Coenzyme Q10 Using Emulsions Containing Oleyl Polyethylene Acetic Acids, 2019, https://doi.org/10.1016/j.ejps.2019.105144.
- [99] R.G.K. Lekshmi, C.S.S. Tejpal, K.K.K. Anas, N.S.S. Chatterjee, S. Mathew, C.N.N. Ravishankar, R.G. Kumar Lekshmi, C.S.S. Tejpal, K.K.K. Anas, N.S. S. Chatterjee, M. Suseela, C.N.N. Ravishankar, Binary blend of maltodextrin and whey protein outperforms gum Arabic as superior wall material for squalene encapsulation, Food Hydrocolloids 106976 (2021) 1–8, https://doi.org/10.1016/J.FOODHYD.2021.106976.
- [100] E. Morales, C. Burgos-Díaz, R.N. Zúñiga, J. Jorkowski, M. Quilaqueo, M. Rubilar, Influence of O/W emulsion interfacial ionic membranes on the encapsulation efficiency and storage stability of powder microencapsulated astaxanthin, Food Bioprod. Process. 126 (2021) 143–154, https://doi.org/10.1016/j. fbp.2020.12.014.

- [101] Y. Guo, D. Qiao, S. Zhao, B. Zhang, F. Xie, Starch-based materials encapsulating food ingredients: recent advances in fabrication methods and applications, Carbohydr. Polym. 270 (2021) 118358, https://doi.org/10.1016/J.CARBPOL.2021.118358.
- [102] S. Vijeth, G.B. Heggannavar, M.Y. Kariduraganavar, Encapsulating Wall Materials for Micro-/Nanocapsules, Microencapsulation Processes, Technologies and Industrial Applications, 2019, https://doi.org/10.5772/INTECHOPEN.82014.
- [103] T.A. Comunian, G. Grassmann Roschel, A.G. da Silva Anthero, I.A. de Castro, M. Dupas Hubinger, Influence of heated, unheated whey protein isolate and its combination with modified starch on improvement of encapsulated pomegranate seed oil oxidative stability, Food Chem. 326 (2020) 126995, https://doi.org/ 10.1016/J.FOODCHEM.2020.126995.
- [104] E. Hebishy, M. Buffa, B. Guamis, A. Blasco-Moreno, A.J. Trujillo, Physical and oxidative stability of whey protein oil-in-water emulsions produced by conventional and ultra high-pressure homogenization: effects of pressure and protein concentration on emulsion characteristics, Innovat. Food Sci. Emerg. Technol. 32 (2015) 79–90, https://doi.org/10.1016/J.IFSET.2015.09.013.
- [105] H. Zhang, Q. Fan, D. Li, X. Chen, L. Liang, Impact of gum Arabic on the partition and stability of resveratrol in sunflower oil emulsions stabilized by whey protein isolate, Colloids Surf. B Biointerfaces 181 (2019) 749–755, https://doi.org/10.1016/J.COLSURFB.2019.06.034.
- [106] H. Cheng, H. Zhang, D. Li, H. Duan, L. Liang, Impact of oil type on the location, partition and chemical stability of resveratrol in oil-in-water emulsions stabilized by whey protein isolate plus gum Arabic, Food Hydrocolloids 109 (2020) 106119, https://doi.org/10.1016/J.FOODHYD.2020.106119.
- [107] X. Chang, W. Feng, L. He, X. Chen, L. Liang, Fabrication and characterisation of whey protein isolate–propolis–alginate complex particles for stabilising α-tocopherol-contained emulsions, Int. Dairy J. 109 (2020) 104756, https://doi.org/10.1016/J.IDAIRYJ.2020.104756.
- [108] M. Silva, T.H. Anh Bui, D. Dharmadana, B. Zisu, J. Chandrapala, Ultrasound-assisted formation of double emulsions stabilized by casein-whey protein mixtures, Food Hydrocolloids 109 (2020) 106143, https://doi.org/10.1016/J.FOODHYD.2020.106143.
- [109] H. Tan, L. Han, C. Yang, Effect of oil type and β-carotene incorporation on the properties of gelatin nanoparticle-stabilized pickering emulsions, Lebensm. Wiss. Technol. 141 (2021) 110903, https://doi.org/10.1016/J.LWT.2021.110903.
- [110] J. Zhang, R. Zhang, Y. Zhang, Y. Pan, H.C. Shum, Z. Jiang, Alginate-gelatin emulsion droplets for encapsulation of vitamin A by 3D printed microfluidics, Particuology 64 (2022) 164–170, https://doi.org/10.1016/J.PARTIC.2021.09.004.
- [111] K. Ganeson, M.R. Razifah, A. Mubarak, A. Kam, S. Vigneswari, S. Ramakrishna, Improved functionality of cinnamon oil emulsion-based gelatin films as potential edible packaging film for wax apple, Food Biosci. 47 (2022) 101638, https://doi.org/10.1016/J.FBIO.2022.101638.
- [112] G. Kan, Y. Zi, L. Li, H. Gong, J. Peng, X. Wang, J. Zhong, Curcumin-encapsulated hydrophilic gelatin nanoparticle to stabilize fish oil-loaded Pickering emulsion, Food Chem. X 17 (2023) 100590, https://doi.org/10.1016/J.FOCHX.2023.100590.
- [113] A.Q. Bi, X.B. Xu, Y. Guo, M. Du, C.P. Yu, C. Wu, Ultrasound pre-fractured casein and in-situ formation of high internal phase emulsions, Ultrason. Sonochem. 64 (2020) 104916, https://doi.org/10.1016/J.ULTSONCH.2019.104916.
- [114] T. Zhang, K.W. Seah, T. Ngai, Single and double Pickering emulsions stabilized by sodium caseinate: effect of crosslinking density, Food Hydrocolloids 139 (2023) 108483, https://doi.org/10.1016/J.FOODHYD.2023.108483.
- [115] A. Can Karaca, E. Assadpour, S.M. Jafari, Historical Perspective Plant protein-based emulsions for the delivery of bioactive compounds. https://doi.org/10. 1016/j.cis.2023.102918, 2023.
- [116] H. Yang, Z. Su, X. Meng, X. Zhang, J.F. Kennedy, B. Liu, Fabrication and characterization of Pickering emulsion stabilized by soy protein isolate-chitosan nanoparticles, Carbohydr. Polym. 247 (2020) 116712, https://doi.org/10.1016/J.CARBPOL.2020.116712.
- [117] K. Wu, Z. Shi, C. Liu, C. Su, S. Zhang, F. Yi, Preparation of Pickering emulsions based on soy protein isolate-tannic acid for protecting aroma compounds and their application in beverages, Food Chem. (2022), https://doi.org/10.1016/j.foodchem.2022.133182.
- [118] H. Li, K. Zhu, H. Zhou, W. Peng, Effects of high hydrostatic pressure treatment on allergenicity and structural properties of soybean protein isolate for infant formula, Food Chem. 132 (2012) 808–814, https://doi.org/10.1016/J.FOODCHEM.2011.11.040.
- [119] O. Mileti, N. Baldino, F.R. Lupi, D. Gabriele, Interfacial Behavior of Vegetable Protein Isolates at Sunflower Oil/Water Interface, 2022, https://doi.org/ 10.1016/j.colsurfb.2022.113035.
- [120] Z. Ren, X. Li, F. Ma, Y. Zhang, W. Hu, M.Z.H. Khan, X. Liu, Oil-in-water emulsions prepared using high-pressure homogenisation with Dioscorea opposita mucilage and food-grade polysaccharides: guar gum, xanthan gum, and pectin, Lebensm. Wiss. Technol. 162 (2022) 113468, https://doi.org/10.1016/J. LWT.2022.113468.
- [121] C. Liu, Y. Li, R. Liang, H. Sun, L. Wu, C. Yang, Y. Liu, Development and characterization of ultrastable emulsion gels based on synergistic interactions of xanthan and sodium stearoyl lactylate, Food Chem. 400 (2023) 133957, https://doi.org/10.1016/J.FOODCHEM.2022.133957.
- [122] H.-P. Lim, S.-S.D. Ng, D.B. Dasa, S.A. Adnan, B.-T. Tey, E.-S. Chan, K.-W. Ho, C.-W. Ooi, Dual (pH and thermal) stimuli-responsive Pickering emulsion stabilized by chitosan-carrageenan composite microgels, Int. J. Biol. Macromol. 232 (2023) 123461, https://doi.org/10.1016/j.ijbiomac.2023.123461.
- [123] Y. Guan, G. Zhao, S. Thaiudom, Evaluation of the physico-chemical properties of potato starch-based foods and their interactions with milk protein and soybean oil, Food Chem. X 16 (2022) 100495, https://doi.org/10.1016/j.fochx.2022.100495.
- [124] E.J. Leal-Casta, Y. García-Tejeda, H. Hern Andez-S Anchez, L. Alamilla-Beltr, D.I. T Ellez-Medina, G. Calder On-Domínguez, H.S. García, G.F. Guti Errez-L Opez, Pickering emulsions stabilized with native and lauroylated amaranth starch, Food Hydrocolloids (2018) 177–185, https://doi.org/10.1016/j. foodhyd.2018.01.043.
- [125] R.E. López-Hernández, S.E. García-Solís, I. Monroy-Rodríguez, M. Cornejo-Mazón, G. Calderón-Domínguez, L. Alamilla-Beltrán, H. Hernández-Sánchez, G. F. Gutiérrez-López, Preparation and characterization of canola oil-in-water Pickering emulsions stabilized by barley starch nanocrystals, J. Food Eng. 326 (2022) 111037, https://doi.org/10.1016/J.JFOODENG.2022.111037.
- [126] A. Sharkawy, M. Filomena Barreiro, A.E. Rodrigues, Chitosan-based Pickering emulsions and their applications: a review, Carbohydr. Polym. 250 (2020) 1–18, https://doi.org/10.1016/j.carbpol.2020.116885.
- [127] A. Muxika, A. Etxabide, J. Uranga, P. Guerrero, K. de la Caba, Chitosan as a bioactive polymer: processing, properties and applications, Int. J. Biol. Macromol. 105 (2017) 1358–1368, https://doi.org/10.1016/J.IJBIOMAC.2017.07.087.
- [128] P. Lv, D. Wang, R. Liang, J. Liu, J. Li, Y. Gao, J. Zhang, F. Yuan, Lycopene-loaded bilayer emulsions stabilized by whey protein isolate and chitosan, Lebensm. Wiss. Technol. 151 (2021) 112122, https://doi.org/10.1016/J.LWT.2021.112122.
- [129] C. Tan, B. Feng, X. Zhang, W. Xia, S. Xia, Biopolymer-coated liposomes by electrostatic adsorption of chitosan (chitosomes) as novel delivery systems for carotenoids, Food Hydrocolloids 52 (2016) 774–784, https://doi.org/10.1016/j.foodhyd.2015.08.016.
- [130] L. Hua, J. Deng, Z. Wang, Y. Wang, B. Chen, Y. Ma, X. Li, B. Xu, Improving the functionality of chitosan-based packaging films by crosslinking with nanoencapsulated clove essential oil, Int. J. Biol. Macromol. 192 (2021) 627–634, https://doi.org/10.1016/j.ijbiomac.2021.09.197.
- [131] A. Kumar Chaudhari, V. Kumar Singh, S. Das, N. Deepika, Kishore Dubey, Fabrication, characterization, and bioactivity assessment of chitosan nanoemulsion containing allspice essential oil to mitigate Aspergillus flavus contamination and aflatoxin B1 production in maize, Food Chem. 372 (2022) 1–11, https://doi. org/10.1016/j.foodchem.2021.131221.
- [132] J. Zhao, T. Wei, Z. Wei, F. Yuan, Y. Gao, Influence of soybean soluble polysaccharides and beet pectin on the physicochemical properties of lactoferrin-coated orange oil emulsion, Food Hydrocolloids 44 (2015) 443–452, https://doi.org/10.1016/j.foodhyd.2014.10.025.
- [133] E. Dickinson, Interfacial structure and stability of food emulsions as affected byprotein-polysaccharide interactions, Soft Matter 4 (2008) 932–942, https://doi. org/10.1039/b800106e.
- [134] A. Deshpande, M. Mohamed, S.B. Daftardar, M. Patel, S.H.S. Boddu, J. Nesamony, Solid lipid nanoparticles in drug delivery: opportunities and challenges, in: Emerging Nanotechnologies for Diagnostics, Drug Delivery and Medical Devices, Elsevier Inc., 2017, pp. 291–330, https://doi.org/10.1016/B978-0-323-42978-8.00012-7.
- [135] M. Aguirre-Ramírez, H. Silva-Jiménez, I.M. Banat, M.A. Díaz De Rienzo, Surfactants: physicochemical interactions with biological macromolecules, Biotechnol. Lett. 43 (2021) 523–535, https://doi.org/10.1007/s10529-020-03054-1.
- [136] T.F. Passos, P. de Camargo Bertuso, M. Nitschke, Biosurfactants as food additives: new trends and applications, Biosurfactants (2023) 85–106, https://doi.org/ 10.1016/B978-0-323-91697-4.00005-3.

- [137] Y. Zi, J. Xu, S. Huang, Y. Zheng, J. Peng, M. Yang, X. Wang, J. Zhong, Effects of octenyl succinic anhydride chemical modification and surfactant physical modification of bovine bone gelatin on the stabilization of fish oil-loaded emulsions, Food Chem. X 16 (2022) 100517, https://doi.org/10.1016/J. FOCHX.2022.100517.
- [138] T. Zhang, M. Ding, H. Zhang, N. Tao, X. Wang, J. Zhong, Fish oil-loaded emulsions stabilized by synergetic or competitive adsorption of gelatin and surfactants on oil/water interfaces, Food Chem. 308 (2020) 125597, https://doi.org/10.1016/J.FOODCHEM.2019.125597.
- [139] L. Ricaurte, M. Hernández-Carrión, M. Moyano-Molano, A. Clavijo-Romero, M.X. Quintanilla-Carvajal, Physical, thermal and thermodynamical study of high oleic palm oil nanoemulsions, Food Chem. 256 (2018) 62–70, https://doi.org/10.1016/j.foodchem.2018.02.102.
- [140] M. Hernández-Carrión, M. Moyano-Molano, L. Ricaurte, A. Clavijo-Romero, M.X. Quintanilla-Carvajal, The effect of process variables on the physical properties and microstructure of HOPO nanoemulsion flakes obtained by refractance window, Sci. Rep. 11 (2021) 1–14.
- [141] A. Henao-Ardila, M. Ximena Quintanilla-Carvajal, P. Román, S. Miguel Caldas-Abril, V. Bonilla-Bravo, F. Leonardo, Effect of wall material on lipophilic functional compounds of high oleic palm oil emulsions encapsulated by Refractance Window drying, Available at: SSRN: https://ssrn.com/abstract=4441392, 2023. (Accessed 3 October 2023), 1, 18.
- [142] S. Sadeghi, A. Madadlou, M. Yarmand, Microemulsification-cold gelation of whey proteins for nanoencapsulation of date palm pit extract, Food Hydrocolloids 35 (2014) 590–596, https://doi.org/10.1016/j.foodhyd.2013.07.021.
- [143] B.G. Erdem, S. Kaya, Production and application of freeze dried biocomposite coating powders from sunflower oil and soy protein or whey protein isolates, Food Chem. 339 (2021), https://doi.org/10.1016/j.foodchem.2020.127976.
- [144] L. Mahalakshmi, P. Choudhary, J.A. Moses, C. Anandharamakrishnan, Emulsion electrospraying and spray drying of whey protein nano and microparticles with curcumin, Food Hydrocolloids for Health 3 (2023) 100122, https://doi.org/10.1016/J.FHFH.2023.100122.
- [145] J.M. Rodríguez-Bernal, E. Tello, E. Flores-Andrade, M. de Jesús Perea-Flores, A.A. Vallejo-Cardona, G.F. Gutiérrez-López, M.X. Quintanilla-Carvajal, Effect of borojo (Borojoa patinoi Cuatrecasas) three-phase composition and gum Arabic on the glass transition temperature, J. Sci. Food Agric. 96 (2016) 1027–1036, https://doi.org/10.1002/jsfa.7190.
- [146] S.F. Subtil, G.A. Rocha-Selmi, M. Thomazini, M.A. Trindade, F.M. Netto, C.S. Favaro-Trindade, Effect of spray drying on the sensory and physical properties of hydrolysed casein using gum Arabic as the carrier, J. Food Sci. Technol. 51 (2014) 2014–2021, https://doi.org/10.1007/s13197-012-0722-z.
- [147] J. Guo, M.M. Giusti, G. Kaletunç, Encapsulation of purple corn and blueberry extracts in alginate- pectin hydrogel particles: impact of processing and storage parameters on encapsulation efficiency, Food Res. Int. (2018), https://doi.org/10.1016/j.foodres.2018.02.035.
- [148] J. Guo, L. Sun, F. Zhang, B. Sun, B. Xu, Y. Zhou, Review: progress in synthesis, properties and application of amino acid surfactants, Chem. Phys. Lett. 794 (2022) 139499, https://doi.org/10.1016/J.CPLETT.2022.139499.
- [149] S. Yousefi, P. Rajaei, L. Nateghi, H.R. Nodeh, L. Rashidi, Encapsulation of sesamol and retinol using alginate and chitosan-coated W/O/W multiple emulsions containing Tween 80 and Span 80, Int. J. Biol. Macromol. 242 (2023) 124766, https://doi.org/10.1016/j.ijbiomac.2023.124766.
- [150] J.N. Naso, F.A. Bellesi, V.M. Pizones Ruiz-Henestrosa, A.M.R. Pilosof, Solubilization of lipolysis products in mixed micelles is enhanced in presence of bile salts and Tween 80 as revealed by a model study (oleic acid) and emulsified chia-oil, Food Res. Int. 161 (2022) 111804, https://doi.org/10.1016/j. foodres.2022.111804.
- [151] M. Espert, A. Salvador, T. Sanz, Rheological and microstructural behaviour of xanthan gum and xanthan gum-Tween 80 emulsions during in vitro digestion, Food Hydrocolloids 95 (2019) 454–461, https://doi.org/10.1016/J.FOODHYD.2019.05.004.
- [152] P.J. Naughton, R. Marchant, V. Naughton, I.M. Banat, Microbial biosurfactants: current trends and applications in agricultural and biomedical industries. https://doi.org/10.1111/jam.14243, 2018.
- [153] K.G.O. Bezerra, R.D. Rufino, J.M. Luna, L.A. Sarubbo, Saponins and microbial biosurfactants: potential raw materials for the formulation of cosmetics, Biotechnol. Prog. 34 (2018) 1482–1493, https://doi.org/10.1002/BTPR.2682.
- [154] R. De Cássia, F.S. Silva, D.G. Almeida, R.D. Rufino, J.M. Luna, V.A. Santos, L.A. Sarubbo, Applications of biosurfactants in the petroleum industry and the remediation of oil spills, OPEN ACCESS Int. J. Mol. Sci 15 (2014) 15, https://doi.org/10.3390/ijms150712523.
- [155] X. Deng, Y. Yang, Y. Ma, X. Sun, G. Zhou, H. Wu, G. Lu, Self-assembled structure of sulfonic gemini surfactant solution, AIP Adv. 8 (2018) 075003, https://doi. org/10.1063/1.5040684.
- [156] M.H. Mondal, S. Malik, A. Roy, R. Saha, B. Saha, Modernization of surfactant chemistry in the age of gemini and bio-surfactants: a review, RSC Adv. 5 (2015) 92707–92718, https://doi.org/10.1039/C5RA18462B.
- [157] B. Cilek Tatar, G. Sumnu, S. Sahin, Emulsion rheology, in: Advances in Food Rheology and its Applications, Elsevier, 2023, pp. 637–659, https://doi.org/ 10.1016/B978-0-12-823983-4.00014-5.
- [158] X. Wang, F. Wang, M.A.M. Taleb, Z. Wen, X. Chen, A review of the seepage mechanisms of heavy oil emulsions during chemical flooding, Energies 15 (2022) 8397, https://doi.org/10.3390/EN15228397, 15 (2022) 8397.
- [159] F. Heidari, S.M. Jafari, A.M. Ziaiifar, N. Malekjani, Stability and release mechanisms of double emulsions loaded with bioactive compounds; a critical review, Adv. Colloid Interface Sci. 299 (2022) 102567, https://doi.org/10.1016/J.CIS.2021.102567.
- [160] S. Akbari, A.H. Nour, Emulsion types, stability mechanisms and rheology: a review, International Journal of Innovative Research and Scientific Studies 1 (2018) 11–17, https://doi.org/10.53894/ijirss.v1i1.4.
- [161] M. Zembyla, B.S. Murray, A. Sarkar, Water-in-oil emulsions stabilized by surfactants, biopolymers and/or particles: a review, Trends Food Sci. Technol. 104 (2020) 49–59, https://doi.org/10.1016/j.tifs.2020.07.028.
- [162] M. Matos, A. Laca, F. Rea, O. Iglesias, M. Rayner, G. Gutiérrez, O/W emulsions stabilized by OSA-modified starch granules versus non-ionic surfactant: stability, rheological behaviour and resveratrol encapsulation, J. Food Eng. 222 (2018) 207-217, https://doi.org/10.1016/j.jfoodeng.2017.11.009.
- [163] J. Cui, M. Hossain, Z. Wang, C. Chang, Enzymatically produced nanocellulose as emulsifier for Pickering emulsion, Food Hydrocolloids (2023), https://doi. org/10.1016/j.foodhyd.2023.108997.
- [164] K. Kamwilaisak, K. Rittiwut, P. Jutakridsada, W. Iamamorphanth, N. Pimsawat, J.T.N. Knijnenburg, S. Theerakulpisut, Rheology, stability, antioxidant properties, and curcumin release of oil-in-water Pickering emulsions stabilized by rice starch nanoparticles, Int. J. Biol. Macromol. 214 (2022) 370–380, https://doi.org/10.1016/j.ijbiomac.2022.06.032.
- [165] H.A. Fonseca-Florido, H.G. Vázquez-García, G. Méndez-Montealvo, U.A. Basilio-Cortés, R. Navarro-Cortés, M.L. Rodríguez-Marín, J. Castro-Rosas, C.A. Gómez-Aldapa, Effect of acid hydrolysis and OSA esterification of waxy cassava starch on emulsifying properties in Pickering-type emulsions, Food Sci. Technol. 91 (2018) 258–264, https://doi.org/10.1016/j.lwt.2018.01.057.
- [166] S. Liu, Y. Zhu, Y. Wu, A. Lue, C. Zhang, Hydrophobic modification of regenerated cellulose microparticles with enhanced emulsifying capacity for O/W Pickering emulsion, Cellulose 26 (2019) 6215–6228, https://doi.org/10.1007/s10570-019-02538-2.
- [167] J.D. Beltrán, L. Ricaurte, K.B. Estrada, M.X. Quintanilla-Carvajal, Effect of homogenization methods on the physical stability of nutrition grade nanoliposomes used for encapsulating high oleic palm oil, Lwt 118 (2020), https://doi.org/10.1016/j.lwt.2019.108801.
- [168] J.D. Beltrán, C.E. Sandoval-Cuellar, K. Bauer, M.X. Quintanilla-Carvajal, In-vitro digestion of high-oleic palm oil nanoliposomes prepared with unpurified soy lecithin: physical stability and nano-liposome digestibility, Colloids Surf. A Physicochem. Eng. Asp. 578 (2019), https://doi.org/10.1016/j. colsurfa.2019.123603.
- [169] M.L.F.F. Ribeiro, Y.H. Roos, A.P.B. Ribeiro, V.R. Nicoletti, Effects of maltodextrin content in double-layer emulsion for production and storage of spray-dried carotenoid-rich microcapsules, Food Bioprod. Process. 124 (2020) 208–221, https://doi.org/10.1016/j.fbp.2020.09.004.
- [170] X.H. Zhao, C.H. Tang, Spray-drying microencapsulation of CoQ10 in olive oil for enhanced water dispersion, stability and bioaccessibility: influence of type of emulsifiers and/or wall materials, Food Hydrocolloids 61 (2016) 20–30, https://doi.org/10.1016/j.foodhyd.2016.04.045.
- [171] A. Marefati, M. Sj, A. Timgren, P. Dejmek, M. Rayner, Fabrication of encapsulated oil powders from starch granule stabilized W/O/W Pickering emulsions by freeze-drying. https://doi.org/10.1016/j.foodhyd.2015.04.022, 2015.
- [172] C.I. Ochoa-Martínez, P.T. Quintero, A.A. Ayala, M.J. Ortiz, Drying characteristics of mango slices using the Refractance WindowTM technique, J. Food Eng. 109 (2012) 69–75, https://doi.org/10.1016/j.jfoodeng.2011.09.032.

- [173] K.R. Cadwallader, J.J. Moore, Z. Zhang, S.J. Schmidt, Comparison of spray drying and refractance window drying tchnologies for the encapsulation of Orange oil, in: C.T. Ho, C.J. Mussinan, F. Shahidi, E. Tratas Contis (Eds.), Recent Advances in Food and Flavour Chemestry, RSCPublishing, 2010, pp. 256–264.
- [174] M. Hernandez-Carrión, M. Moyano, M.X. Quintanilla-Carvajal, Design of high-oleic palm oil nanoemulsions suitable for drying in refractance windowTM, J. Food Process. Preserv. 45 (2021) 1–2, https://doi.org/10.1111/jfpp.15076.
- [175] Q. Li, W. Liang, L. Lv, Z. Fang, D. Xu, J. Liao, Y. Liu, Preparation of PCL/lecithin/bacteriocin CAMT6 antimicrobial and antioxidant nanofiber films using emulsion electrospinning: characteristics and application in chilled salmon preservation, Food Res. Int. 175 (2024) 113747, https://doi.org/10.1016/J. FOODRES.2023.113747.
- [176] M. Wu, Z. Zhou, J. Yang, M. Zhang, F. Cai, P. Lu, ZnO nanoparticles stabilized oregano essential oil Pickering emulsion for functional cellulose nanofibrils packaging films with antimicrobial and antioxidant activity, Int. J. Biol. Macromol. 190 (2021) 433–440, https://doi.org/10.1016/J.IJBIOMAC.2021.08.210.
- [177] G.M. Bornhorst, O. Gouseti, M.S.J. Wickham, S. Bakalis, Engineering digestion: multiscale processes of food digestion, J. Food Sci. 81 (2016) R534–R543, https://doi.org/10.1111/1750-3841.13216.
- [178] A. Guerra, L. Etienne-Mesmin, V. Livrelli, S. Denis, S. Blanquet-Diot, M. Alric, Relevance and challenges in modeling human gastric and small intestinal digestion, Trends Biotechnol. 30 (2010) 591–600. https://ac.els-cdn.com/S016777991200128X/1-s2.0-S016777991200128X-main.pdf?_tid=64b48f38-c2c8-4415-96df-33c6d60ac2ab&acdnat=1521224971_ef5e816a53c22c68f0fb57f470f6a158. (Accessed 15 March 2018).
- [179] A.A. Xavier, L.R. Mariutti, Static and semi-dynamic in vitro digestion methods: state of the art and recent achievements towards standardization. https://doi. org/10.1016/j.cofs.2021.08.002, 2021.
- [180] D. Duijsens, K. Pälchen, J.M. Guevara-Zambrano, S.H.E. Verkempinck, M.R. Infantes-Garcia, M.E. Hendrickx, A.M. Van Loey, T. Grauwet, Strategic choices for in vitro food digestion methodologies enabling food digestion design. https://doi.org/10.1016/j.tifs.2022.06.017, 2022.
- [181] C. Li, W. Yu, P. Wu, X.D. Chen, Current in vitro digestion systems for understanding food digestion in human upper gastrointestinal tract. https://doi.org/10. 1016/j.tifs.2019.12.015, 2019.
- [182] W. Liu, Y. Kong, A. Ye, P. Shen, L. Dong, X. Xu, Y. Hou, Y. Wang, Y. Jin, J. Han, Preparation, formation mechanism and in vitro dynamic digestion behavior of quercetin-loaded liposomes in hydrogels, Food Hydrocolloids 104 (2020) 105743, https://doi.org/10.1016/J.FOODHYD.2020.105743.
- [183] F. Acevedo, M. Rubilar, I. Jofré, M. Villarroel, P. Navarrete, M. Esparza, F. Romero, E.A. Vilches, V. Acevedo, C. Shene, Oil bodies as a potential microencapsulation carrier for astaxanthin stabilisation and safe delivery, J Microencapsul 31 (2014) 488–500. https://doi.org/10.3109/02652048.2013. 879931.
- [184] A.M. Cheong, C.P. Tan, K.L. Nyam, In vitro evaluation of the structural and bioaccessibility of kenaf seed oil nanoemulsions stabilised by binary emulsifiers and β-cyclodextrin complexes, J Food Eng 189 (2016) 90–98. https://doi.org/10.1016/j.jfoodeng.2016.06.002.
- [185] T. Feng, Z. Hu, K. Wang, X. Zhu, D. Chen, H. Zhuang, L. Yao, S. Song, H. Wang, M. Sun, Emulsion-based delivery systems for curcumin: Encapsulation and interaction mechanism between debranched starch and curcumin, Int J Biol Macromol 161 (2020) 746–754. https://doi.org/10.1016/j.ijbiomac.2020.06.088.
- [186] V.M. Esquerdo, G.L. Dotto, L.A.A. Pinto, Preparation of nanoemulsions containing unsaturated fatty acid concentrate-chitosan capsules, J Colloid Interface Sci 445 (2015) 137–142. https://doi.org/10.1016/j.jcis.2014.12.094.
- [187] P.J. García-Moreno, K. Stephansen, J. Van Der Kruijs, A. Guadix, E.M. Guadix, I.S. Chronakis, C. Jacobsen, Encapsulation of fish oil in nanofibers by emulsion electrospinning: Physical characterization and oxidative stability, J Food Eng 183 (2016) 39–49. https://doi.org/10.1016/J.JFOODENG.2016.03.015.
- [188] B. Ozturk, S. Argin, M. Ozilgen, D.J. McClements, Nanoemulsion delivery systems for oil-soluble vitamins: Influence of carrier oil type on lipid digestion and vitamin D3 bioaccessibility, Food Chem 187 (2015) 499–506. https://doi.org/10.1016/j.foodchem.2015.04.065.
- [189] A. Mortensen, F. Aguilar, R. Crebelli, A. Di Domenico, M.J. Frutos, P. Galtier, D. Gott, U. Gundert-Remy, C. Lambré, J. Leblanc, O. Lindtner, P. Moldeus, P. Mosesso, A. Oskarsson, D. Parent-Massin, I. Stankovic, I. Waalkens-Berendsen, R.A. Woutersen, M. Wright, B. Dusemund, Re-evaluation of lecithins (E 322) as a food additive, EFSA Journal 15 (4) (2017). https://doi.org/10.2903/j.efsa.2017.4742.
- [190] EFSA Panel on Food Additives and Nutrient Sources added to Food (2015). Scientific Opinion on the re-evaluation of polyoxyethylene sorbitan monolaurate (E 432), polyoxyethylene sorbitan monoleate (E 433), polyoxyethylene sorbitan monostearate (E 435) and polyoxyethylene sorbitan tristearate (E 436) as food additives. EFSA Journal, 13(7). https://doi.org/10.2903/j.efsa.2015.4152.
- [191] M. Younes, P. Aggett, F. Aguilar, R. Crebelli, B. Dusemund, M. Filipic, M.J. Frutos, P. Galtier, D. Gott, U. Gundert-Remy, G.G. Kuhnle, J.C. Leblanc, I. T. Lillegaard, P. Moldeus, A. Mortensen, A. Oskarsson, I. Stankovic, I. Waalkens-Berendsen, R.A. Woutersen, C. Lambré, Re-evaluation of sodium, potassium and calcium salts of fatty acids (E 470a) and magnesium salts of fatty acids (E 470b) as food additives, EFSA Journal 16 (3) (2018). https://doi.org/10.2903/j. efsa.2018.5180.
- [192] M. Younes, P. Aggett, F. Aguilar, R. Crebelli, B. Dusemund, M. Filipic, M.J. Frutos, P. Galtier, D. Gott, U. Gundert-Remy, G.G. Kuhnle, J.C. Leblanc, I. T. Lillegaard, P. Moldeus, A. Mortensen, A. Oskarsson, I. Stankovic, I. Waalkens-Berendsen, R.A. Woutersen, C. Lambre, Re-evaluation of mono-and diglycerides of fatty acids (E 471) as food additives, EFSA Journal 15 (11) (2017). https://doi.org/10.2903/J.EFSA.2017.5045.
- [193] A. Mortensen, F. Aguilar, R. Crebelli, A. Di Domenico, B. Dusemund, M.J. Frutos, P. Galtier, D. Gott, U. Gundert-Remy, J. Leblanc, O. Lindtner, P. Moldeus, P. Mosesso, D. Parent-Massin, A. Oskarsson, I. Stankovic, I. Waalkens-Berendsen, R.A. Woutersen, M. Wright, C. Lambré, Re-evaluation of sorbitan monostearate (E 491), sorbitan tristearate (E 492), sorbitan monolaurate (E 493), sorbitan monooleate (E 494) and sorbitan monopalmitate (E 495) when used as food additives, EFSA Journal 15 (5) (2017). https://doi.org/10.2903/j.efsa.2017.4788.
- [194] Mortensen, A., Aguilar, F., Crebelli, R., Domenico, A. Di, Dusemund, B., Frutos, M. J., Galtier, P., Gott, D., Gundert-Remy, U., Leblanc, J., Lindtner, O., Moldeus, P., Mosesso, P., Parent-Massin, D., Oskarsson, A., Stankovic, I., Waalkens-Berendsen, I., Woutersen, R. A., Wright, M., ... Lambré, C. (2016). Reevaluation of ammonium phosphatides (E 442) as a food additive. *EFSA Journal*, 14(11). https://doi.org/10.2903/j.efsa.2016.4597.
- [195] EFSA ANS Panel (EFSA Panel on Food Additives and Nutrient Sources added toFood). (2016). Scientific Opinion on the re-evaluation of sucrose acetate isobutyrate (E 444) as a foodadditive. EFSA Journal, 14(5), 1–39. https://doi.org/10.2903/j.efsa.2016.4489.
- [196] D.J. McClements, Critical review of techniques and methodologies for characterization of emulsion stability, Critical Reviews in Food Science and Nutrition 47 (7) (2007) 611–649. https://doi.org/10.1080/10408390701289292.
- [197] S. Schuster, R. Bernewitz, G. Guthausen, J. Zapp, A.M. Greiner, K. Köhler, H.P. Schuchmann, Analysis of W1/O/W2 double emulsions with CLSM: Statistical image processing for droplet size distribution, Chemical Engineering Science 81 (2012) 84–90. https://doi.org/10.1016/J.CES.2012.06.059.
- [198] M. Félix, C. Carrera, A. Romero, C. Bengoechea, A. Guerrero, Rheological approaches as a tool for the development and stability behaviour of protein-stabilized emulsions, Food Hydrocolloids 104 (2020) 105719. https://doi.org/10.1016/J.FOODHYD.2020.105719.
- [199] E.A. Alshaafi, A. Prakash, S.M. Mercer, Ultrasonic based methods to characterize stability of water-in-crude oil emulsions. https://doi.org/10.1016/j.colsurfa.2020. 125900.
- [200] M. Su, M. Xue, X. Cai, Z. Shang, F. Xu, Particle size characterization by ultrasonic attenuation spectra, Particulogy 6 (2008) 276–281. https://doi.org/10. 1016/j.partic.2008.02.001.