Received: 23 March 2022

Revised: 19 May 2022

(wileyonlinelibrary.com) DOI 10.1002/ps.7006

Change-point analysis of lambda-cyhalothrin efficacy against soybean aphid (*Aphis glycines* Matsumura): identifying practical resistance from field efficacy trials

James P. Menger,^a <a>> Arthur V. Ribeiro,^a Bruce D. Potter^b and Robert L. Koch^{a*}

Abstract

Background: Soybean aphid (*Aphis glycines* Matsumura) remains the most economically important arthropod pest of soybean in the Upper Midwest Region of the USA. Soybean aphid resistance to the pyrethroid insecticides emerged in 2015; however, the reduction in the efficacy of field applications of pyrethroid insecticides has not been quantified. Based on time-series data from insecticide efficacy trials at two locations, a novel approach of continuous two-phase change point-regression models was used to indicate whether a change in percent control had occurred, and to provide an indication of when and to what degree the percent control had changed.

Results: At both locations examined in this study, a significant change point for percent control of λ -cyhalothrin was detected in 2014, thus marking the onset of practical resistance in the soybean aphid. Percent control decreased at a rate of 4.30% and 19.90% per year at these locations. By contrast, percent control for chlorpyrifos remained high over time with no significant change point.

Conclusion: This research demonstrates that retrospective time-series analysis of insecticide efficacy data can identify the onset and magnitude of practical resistance in the field. This further validates and compliments the other lines of evidence related to pyrethroid resistance in soybean aphid.

© 2022 The Authors. Pest Management Science published by John Wiley & Sons Ltd on behalf of Society of Chemical Industry.

Keywords: practical resistance; insecticide; pest management; pyrethroid; percent control; efficacy

1 INTRODUCTION

Soybean aphid (*Aphis glycines* Matsumura) (Hemiptera: Aphididae) persists as a significant pest in the Midwest United States (US)¹ more than 20 years after its initial discovery in North America.² Despite extensive research into several alternative management tactics, including host-plant resistance and biological control,³ the use of foliar insecticides has remained the most effective and economical strategy for managing soybean aphid.^{4–6} These foliar applications of insecticides on soybean have relied heavily on organophosphates (Group 1B) and pyrethroids (Group 3A).⁵ Insecticide use on soybean in the Midwest increased dramatically after the invasion by soybean aphid.^{3.7}

An overreliance on insecticides can lead to ecological backlash in the form of insecticide resistance. This evolutionary response is frequently associated with the repeated use of insecticides, and resistance typically surfaces as a delayed response after years of seemingly good control.⁸ The emergence of insecticide resistance in soybean aphid has posed a significant challenge to soybean production. Reports of field applications of pyrethroid insecticides failing to control soybean aphid were made by growers and consultants in Minnesota in 2015, and in Minnesota and Iowa in 2016.⁹ In the following years, reports of such control failures continued in Minnesota and expanded to South Dakota and North Dakota.^{10,11} In response to these reports, laboratory bioassays were performed and confirmed reduced susceptibility in some field-collected populations of soybean aphid from Minnesota, North Dakota, South Dakota, Iowa and Manitoba to the pyrethroids λ -cyhalothrin and bifenthrin, relative to a laboratory-susceptible population.^{9,11}

More recent work has begun to identify the mechanisms of resistance present in the soybean aphid. Phenotypically resistant

b University of Minnesota Southwest Research and Outreach Center, Lamberton, MN, 56152

© 2022 The Authors. *Pest Management Science* published by John Wiley & Sons Ltd on behalf of Society of Chemical Industry. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

^{*} Correspondence to: Robert L. Koch, Department of Entomology, University of Minnesota, 1980 Folwell Avenue, Saint Paul, MN 55108, USA. E-mail: koch0125@umn.edu

a Department of Entomology, University of Minnesota, Saint Paul, MN, 55108

populations of soybean aphid with both induced and constitutive overexpression of detoxifying enzyme genes have been reported.¹² Furthermore, point mutations in the voltage-gated sodium channel genes associated with *skdr* and *kdr* have been identified in soybean aphid populations.^{13,14}

It is important to note that such documentation of resistance in the laboratory does not always equate to a reduction in the field efficacy of insecticide applications against the pest.¹⁵ Potential differences in efficacy of insecticides under laboratory and field conditions have been documented elsewhere.^{16–19} Practical resistance is defined as "field-evolved resistance that reduces pesticide efficacy and has practical consequences for pest control".²⁰ More specifically, the Insecticide Resistance Action Committee's definition of resistance specifies "the repeated failure of a product to achieve the expected level of control when used according to the label recommendation for that pest species."²¹

Reductions in the efficacy of field applications of pyrethroid insecticides over time for sovbean aphid have not been quantified. However, data to perform such an examination are available from replicated and controlled field experiments (efficacy trials), performed by university researchers at multiple locations and over several years to evaluate the efficacy of various insecticides against soybean aphid. Time-series analysis can be useful in elucidating underlying trends and systematic patterns in data over time. A class of regression models in which predictors are associated with the outcome in a threshold-dependent manner by introducing a "change point" provide a simple way to interpret certain types of nonlinear relationships.²² Change-point analysis is a distinct form of threshold analysis that is specifically concerned with finding structural changes within a "natural axis" such as time.²³ Data from these insecticide efficacy trials can provide insight into the onset and magnitude of practical resistance of soybean aphid to insecticides.

Here, time series were created of data from insecticide efficacy trials performed at two locations in Minnesota spanning 2005–2020. For each location, percent control relative to the untreated control was calculated for a common pyrethroid (λ -cyhalothrin), an insecticide that soybean aphids have documented resistance towards in laboratory bioassays. In addition, for one location, percent control was calculated for a common organophosphate (chlorpyrifos), an insecticide for which there have been no reports of resistance for soybean aphid. These analyses can indicate whether a change point in percent control has occurred and provide an indication of when and to what degree the percent control has changed.

2 MATERIALS AND METHODS

2.1 Data collection and summary

Data were compiled from insecticide efficacy trials conducted from 2005 to 2020 at the University of Minnesota's Southwest Research and Outreach Center (SWROC) in Lamberton, MN and the University of Minnesota Outreach, Research and Education (UMORE) Park in Rosemount, MN. All the efficacy trials were performed using standard agronomic practices with aphidsusceptible soybean varieties (that is, not containing *Rag* genes) adapted to the region (Table 1). Each efficacy trial was conducted as a randomized complete block design with four blocks and multiple insecticide treatments, including foliar application of the pyrethroid λ -cyhalothrin (Warrior or Warrior II; Syngenta Corp.) applied at a high label rate using standard application practices, and an untreated control treatment (Table 1). In addition, several of the efficacy trials conducted at SWROC included the organophosphate chlorpyrifos (Lorsban 4e or Lorsban Advanced; Dow AgroSciences) applied at a high label rate (Table 1), which were also evaluated as a positive control for insecticide efficacy over time. Over the period that the data were collected, the efficacy trials were conducted by a limited number of people, which minimizes variability among trials. At SWROC, all trials were conducted by B.D. Potter. At UMORE, trials conducted prior to 2013 were performed by D.W. Ragsdale, and trials in 2013 and later were performed by R.L. Koch following the methods used previously at this location.

In insecticide efficacy trials, insecticide applications are not always triggered by densities of pests that would typically require treatment. Sometimes, low pest densities are treated to ensure at least some data will be collected. However, to minimize potential effects of variable aphid densities on the results, data sets were filtered to include only those with pretreatment densities of approximately 100 aphids per plant or more. From each field trial, the efficacy of the targeted insecticides was estimated as percent control measured at approximately 2 weeks after insecticide application (Table 1). Two weeks after treatment falls within the typical range of data collection for soybean aphid efficacy trials, and is a period that has historically represented good control for soybean aphid with foliar applied λ -cyhalothrin across the region.^{24–30}

Percent control was calculated as $100 \times (C - T)/C$, where C is the mean number of aphids per plant in the untreated control plots, and T is the mean number of aphids per plant in the insecticidetreated plots.^{31,32} The efficacy trials conducted at SWROC in 2004 and 2006 had two control treatments per block whose data were averaged prior to calculation of percent control. In addition, the efficacy trial conducted at SWROC in 2018 had two control treatments and two λ -cyhalothrin treatments per block that were averaged prior to calculation of percent control. One efficacy trial was performed at each location per year, except in 2020 at SWROC and in 2014 at UMORE, where two separate efficacy trials occurred. Percent control was calculated for each trial independently, and individual trial percent control values were averaged to produce a single percent control value in these instances of two trials within a single year at a location, due to trial proximity and the potential lack of independence between aphid populations. These instances of multiple trials at a given location within a year were accounted for by weighting in the analysis (see below).

2.2 Data analysis

The resulting time series of percent control for λ -cyhalothrin at SWROC and UMORE, and for chlorpyrifos at SWROC, were analyzed in R version 4.0.3 (ref.³³) and R Studio version 1.3.1093.³⁴ Each time series was analyzed separately using the chngpt package²³ (code: chngptm) with continuous two-phase regression models with percent control as the dependent variable and year as the independent variable. In the models, years were weighted for the number of trials within a location. Models were selected by first fitting a "segmented" model (slopes before and after threshold); however, in all cases, the slope before the threshold was not significantly different from 0, which allowed the use of a "hinge" model (slope of 0 before threshold and decreasing slope after threshold). Use of a hinge model is preferred because the model can be estimated with substantially higher precision than that of a segmented model.^{23,35} Model significance was evaluated using a likelihood ratio test (package: Imtest, code: Irtest) comparing the hinge model to the null (intercept-only linear) model. Root mean square errors (RMSE) were manually calculated, and change-point (breakpoint) significance was tested using the chngpt.test

ω
Ō
4
0

wileyonlinelibrary.com/journal/ps	© 2022 The Authors.	Pest Manag Sci 2022; 78 : 3638–3643								
Pest Management Science published by John Wiley & Sons Ltd on behalf of Society of Chemical Industry.										

meets	business															~~~		.50	CI.C	'ng												51 101
		Reference	f	f		ų لو	f	f	f			f	f			f			f			f	52	52	53	48	54	48	f	49	51	; (Lorsban 4e
		Pressure (kPa)	275.8	275.8		275.8	275.8	275.8	275.8			241.3	241.3			241.3			241.3			241.3	241.3	241.3	б	275.8	275.8	275.8	275.8	206.8	206.8	r II), chlorpyrifos
:005 to 2020		Volume (L ha ⁻¹)	187.08	187.08		187.08	187.08	187.08	187.08			140.31	140.31			140.31			140.31			140.31	140.31	140.31	187.08	187.08	187.08	187.08	187.08	140.31	140.31	arrior or Warrio
10RE), MN from 2		Rate (L ha ⁻¹)	0.23	0.23	1.17	0.23	0.12	0.12	0.12	1.17		0.12	0.12	1.17		0.14	1.17		0.14	1.17		0.14	0.14	0.12	0.18	0.12	0.12	0.12	0.12	0.12	0.12	λ-cyhalothrin (W
C) and Rosemount (UN		Formulation	Warrior	Warrior	Lorsban 4e	Warrior	Warrior II	Warrior II	Warrior II	Lorsban	Advanced	Warrior II	Warrior II	Lorsban	Advanced	Warrior II	Lorsban	Advanced	Warrior II	Lorsban	Advanced	Warrior II	Warrior II	Warrior II	Warrior	Warrior II	ng foliar application of					
mberton (SWROC		Treated density ^e	0	0	0		4	0	0	0		-	502	m		12	12		110	9		158	37	48	39	26	11	21	375	102	478	eatments includi
erformed at Lai	ost-treatment	Control density ^d	145	233		300	189	92	243			56	3742			153			376			850	157	163	1784	744	448	526	977	83	634	and multiple tre rpyrifos.
oybean aphid p	Pc	Sample date	17 Aug	12 Sep		24 Aug	12 Aug	19 Aug	17 Aug			26 Aug	28 Aug			6 Sep			29 Aug			3 Sep	25 Aug	25 Aug	3 Aug	27 Aug	18 Aug	26 Aug	20 Aug	30 Aug	5 Sep	with four blocks alothrin or chlo
cy trials ^a against s		Treatment date ^c	2 Aug	29 Aug		9 Aug	30 Jul	4 Aug	3 Aug			13 Aug	14 Aug			24 Aug			16 Aug			20 Aug	11 Aug	11Aug	23 Jul	13 Aug	3 Aug	12 Aug	4 Aug	17 Aug	22 Aug	ete block designs mental plots. rrifos. tred control plots reated with λ-cy ^t ta.
isecticide efficad	tment	Overall density ^b	117	146		284	956	151	272			123	468			284			389			179	152	202	215	382	95	328	323	181	407	domized comple ced control. cross the experi sthrin or chlorpy cross the untree cross the plots t unpublished dat
tal details for in	Pretrea	Sample date	2 Aug	28 Aug		9 Aug	29 Jul	3 Aug	3 Aug			13 Aug	14 Aug			23 Aug			15 Aug			19 Aug	11 Aug	11 Aug	22 Jul	12 Aug	1 Aug	8 Aug	31 Jul	16 Aug	21 Aug	rformed as ranc and an untreat ids per plant a tion of <i>i</i> -cyhalc ids per plant a obert L. Koch, u cation.
. Experiment		Year	2005	2006		2007	2008	2010	2011			2014	2015			2017			2018			2019	2020	2020	2009	2013	2014	2014	2015	2018	2019	nents were per in Advanced), number of aph foliar applicat number of aph number of aph Potter and R
TABLE 1.		Location	SWROC																						UMORE							^a Experir or Lorsbé b Mean r c Date of d Mean r f Bruce D g Not rep



FIGURE 1. Efficacy of λ -cyhalothrin in field trials in southwest Minnesota. Percent control of soybean aphid populations by foliar application of λ -cyhalothrin in insecticide efficacy field trials conducted at the University of Minnesota's Southwest Research and Outreach Center (SWROC) in Lamberton, MN from 2005 to 2020. Percent control was calculated relative to the untreated control at approximately 2 weeks after insecticide application. Analysis identified 2014 as the change point in λ -cyhalothrin efficacy at this location.

function. Estimations of slopes after threshold were derived from the lincomb function.

2.3 Literature search

To assess the novelty of using this approach for examining practical resistance, a literature search was conducted on 14 November 2021 to search for the use of similar statistical methods in evaluating field-evolved resistance and the efficacy of pesticides with the search string ("piecewise regression" OR "threshold regression" OR "broken stick" OR "hockey stick" OR "changepoint" OR "change point") AND ("insecticide" OR "entomology" OR "insect" OR "resistance" OR "pesticide" OR "fungicide" OR "antibiotic" OR "herbicide") in Web of Science, CAB Abstracts and Agricola.

3 RESULTS

3.1 SWROC

The time series of percent control for λ -cyhalothrin at SWROC is presented in Figure 1. For λ -cyhalothrin, the hinge model provided a better fit than the linear null model ($\chi^2 = 21.03$, d.f. = 2, p < 0.001). A significant change point in percent control was identified at 2014 (95% confidence interval: 2007, 2017) (RMSE = 4.50) ($\chi^2 = 22.81$, d.f. = 1, p < 0.001). The pre-change point intercept was 98.57% (95% confidence interval: 96.85, 100.76), for the years 2005–2014. The slope after the change point was -4.30% (95% confidence interval: -25.62, -1.68) per year.

The time series of percent control for chlorpyrifos at SWROC is presented in Figure 2. For chlorpyrifos, a candidate change point for percent control was detected in 2011; however, it was found to be nonsignificant ($\chi^2 = 2.12$, d.f. = 1, p = 0.269), and the fit of the hinge model did not differ from that of the null model ($\chi^2 = 2.12$, d.f. = 2, p = 0.3466). An intercept-only linear model revealed an average (\pm SEM) percent control of 98.1% \pm 1.46%.

3.2 UMORE

The time series of percent control for λ -cyhalothrin at UMORE is presented in Figure 3. For λ -cyhalothrin, the hinge model provided a better fit than the linear null model ($\chi^2 = 10.27$, d.f. = 2,



FIGURE 2. Efficacy of chlorpyrifos in field trials in southwest Minnesota. Percent control of soybean aphid populations by foliar application of chlorpyrifos in insecticide efficacy field trials conducted at the University of Minnesota's Southwest Research and Outreach Center (SWROC) in Lamberton, MN from 2005 to 2020. Percent control was calculated relative to the untreated control at approximately 2 weeks after insecticide application. Analysis did not identify a significant change point in chlorpyrifos efficacy at this location.



FIGURE 3. Efficacy of λ -cyhalothrin in field trials in southeast Minnesota. Percent control of soybean aphid populations by foliar application of λ -cyhalothrin in insecticide efficacy field trials conducted at the University of Minnesota's Outreach, Research and Education (UMORE) Park in Rose-mount, MN from 2005 to 2020. Percent control was calculated relative to the untreated control at approximately 2 weeks after insecticide application. Analysis identified 2014 as the change point in λ -cyhalothrin efficacy at this location.

p = 0.006). A significant change point in percent control was identified at 2014 (95% confidence interval: 2009, 2015) (RMSE = 20.33) ($\chi^2 = 10.15$, d.f. = 1, p = 0.003). The pre-change point intercept was 92.90% (95% confidence interval: 85.72, 102.72) for the years 2005–2014, and the slope after the changepoint was –19.90% (95% confidence interval: –35.44, –8.52) per year.

3.3 Literature search

Our literature search resulted in 97, 47 and 27 publications from Web of Science, CAB Abstracts and Agricola, respectively. Of the 171 total results, 120 unique publications were found. Within these unique publications, 16 were related to entomology, and 21 involved the use of pesticides. Within our literature search, examples were found of change point and similar statistical methods used in entomology when modeling population dynamics in monarch butterflies (*Danaus plexippus*),³⁶ pupation success and behavior of western flower thrips (*Frankliniella occidentalis*),³⁷ and the residual efficacy of pesticide-impregnated ear tags for cattle.³⁸ Also, similar methods have been used when evaluating the dissipation of pesticides used as seed treatment in soils,³⁹ examining the behavior and environmental fate of glyphosate in water and sediments,⁴⁰ and for determining the effects of defoliation on yield in field beans.⁴¹ However, no examples were found of previous research using change point or similar types of analyses to model time-series data on insecticide efficacy related to resistance.

4 DISCUSSION AND CONCLUSION

For both locations examined in this study, a significant change point in percent control for λ -cyhalothrin was found at 2014; thus, signifying the last year prior to the onset of decreasing control. In the next soybean growing season (2015), reports of control failures and laboratory confirmations of reduced susceptibility began.^{9,10} In this research, a retrospective time-series analysis revealed the onset of practical resistance in the field, further validating and complimenting other lines of evidence demonstrating the presence of pyrethroid resistance in soybean aphid. Furthermore, this may be the first time this type of change-point analysis has been used for retrospective assessment of the onset of practical resistance to insecticides.

Insecticides are designed to deliver high levels of consistent control, as seen in the current study from 2005 to 2014. Therefore, the use of hinge modeling (no initial slope) in this system would be expected, which is corroborated by the lack of a significant pre-change point slope indicated in the segmented models. It is important to note that the pre-change point intercept does not necessarily represent the average of the pre-change point observations. For SWROC and UMORE, the pre-change point intercepts were numerically less than the mean of the pre-change point observations (SWROC = 99.4%, UMORE = 97.0%). This may occur because the estimated change point candidate is selected considering the overall model with the highest likelihood.²³ A similar effect can also be seen in other publications that use similar analysis.^{42,43}

The post-change point slopes for the two locations examined in this study demonstrate the importance of conducting pesticide efficacy trials over time using consistent methods to provide such long-term data sets. At UMORE, the estimated slope of percent control after the change point was approximately five times that of SWROC, although not statistically different because of the wide confidence intervals. The continued evaluation of these insecticides at these locations may refine these estimates as trajectories of change near the terminus of a time series may be difficult to determine.³⁶

Resistance to pesticides is an ever-increasing challenge to global agriculture with nearly a 1000 species of pests, including approximately 600 arthropod species, resistant to one or more pesticides.²⁰ Resistance to a pesticide can be directly documented by demonstrating a reduction in susceptibility over time within a population.⁴⁴ However, the impact of resistance on the practical control of a pest can vary because of several factors including frequency of resistance, population density and geographic distribution.^{45,46}

The broader impact of field-evolved resistance on pest control can vary from insignificant to severe depending on the level of practical resistance realized in the field and the availability of alternative control measures.²⁰ The current state of pyrethroid resistance in the soybean aphid was likely accelerated through regularly applying a limited number of insecticidal modes of action to manage the insect, and the lack of adoption of other integrated pest management strategies such as host-plant resistance and biological control.¹⁰

Despite the practical resistance observed for the pyrethroids, the organophosphate chlorpyrifos has remained highly effective. However, on 18 August 2021, the US Environmental Protection Agency released their Final Tolerance Rule for chlorpyrifos revoking all tolerances for the insecticide on food products nationwide,⁴⁷ further limiting the products available for the control of soybean aphid. Although some newer, more selective insecticides, such as sulfoxaflor (group 4C), flupyradifurone (group 4D), and afidopyropen (group 9D) are labeled for and effective against soybean aphid,^{48–51} caution should be taken to preserve their efficacy through the use of insecticide resistance management tactics. Regular efficacy monitoring of these alternative chemistries, both in the laboratory and in the field, is necessary to quantify of the onset and magnitude of practical resistance, and to avoid a similar fate as that of the pyrethroids for soybean aphid.

ACKNOWLEDGMENTS

We thank Drs. David Andow and Seth Naeve for providing comments on an earlier version of this manuscript, Shannon Farrell for assistance with the literature search, the North Central Soybean Research Program for financial support of this work, and the entities that supported the insecticide efficacy trials cited in this study.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

REFERENCES

- 1 Hurley T and Mitchell P, Value of neonicotinoid seed treatments to US soybean farmers. *Pest Manag Sci* **73**:102–112 (2017).
- 2 Alleman RJ, Grau CR and Hogg DB, Soybean aphid host range and virus transmission efficiency. Proceedings of the Wisconsin Fertilizer, Agline Pest Management Conference. (2002). https://extension.soils.wisc. edu/wp-content/uploads/sites/68/2016/07/Alleman-Conf-2002.pdf [accessed 6 March 2022].
- 3 Ragsdale DW, Landis DA, Brodeur J, Heimpel GE and Desneux N, Ecology and management of the soybean aphid in North America. *Annu Rev Entomol* **56**:375–399 (2011).
- 4 Johnson KD, O'Neal ME, Ragsdale DW, Difonzo CD, Swinton SM, Dixon PM et al., Probability of cost-effective management of soybean aphid (Hemiptera: Aphididae) in North America. J Econ Entomol **102**:2101–2108 (2009).
- 5 Hodgson EW, McCornack BP, Tilmon K and Knodel JJ, Management recommendations for soybean aphid (Hemiptera: Aphididae) in the United States. *J Integr Pest Manag* **3**:E1–E10 (2012).
- 6 Krupke CH, Alford AM, Cullen EM, Hodgson EW, Knodel JJ, McCornack B et al., Assessing the value and pest management window provided by neonicotinoid seed treatments for management of soybean aphid (*Aphis glycines* Matsumura) in the upper midwestern United States. *Pest Manag Sci* **73**:2184–2193 (2017).
- 7 Coupe RH and Capel PD, Trends in pesticide use on soybean, corn and cotton since the introduction of major genetically modified crops in the United States. *Pest Manag Sci* **72**:1013–1022 (2016).



- 8 Pedigo LP, Rice ME and Krell RK, *Entomology and Pest Management*, Seventh edn. Waveland Press, Long Grove, (2021).
- 9 Hanson AA, Menger-Anderson J, Silverstein C, Potter BD, MacRae IV, Hodgson EW *et al.*, Evidence for soybean aphid (Hemiptera: Aphididae) resistance to pyrethroid insecticides in the upper midwestern United States. *J Econ Entomol* **110**:2235–2246 (2017).
- 10 Koch RL, Hodgson EW, Knodel JJ, Varenhorst AJ and Potter BD, Management of insecticide-resistant soybean aphids in the upper Midwest of the United States. *J Integr Pest Manag* **9**:23 (2018).
- 11 Menger J, Beauzay P, Chirumamilla A, Dierks C, Gavloski J, Glogoza P et al., Implementation of a diagnostic-concentration bioassay for detection of susceptibility to pyrethroids in soybean aphid (Hemiptera: Aphididae). J Econ Entomol **113**:932–939 (2020).
- 12 Paula DP, Menger JP, Andow DA and Koch RL, Diverse patterns of constitutive and inducible overexpression of detoxifying enzyme genes among resistant *Aphis glycines* populations. *Pestic Biochem Physiol* **164**:100–114 (2020).
- 13 Paula DP, Lozano RE, Menger JP, Andow DA and Koch RL, Identification of point mutations related to pyrethroid resistance in voltage-gated sodium channel genes in *Aphis glycines. Entomol Gen* **41**:243–255 (2021).
- 14 Valmorbida I, Hohenstein JD, Coates BS, Bevilagua JG, Menger JP, Hodgson EW *et al.*, Association of voltage-gated sodium channel mutations with field-evolved pyrethroid resistant phenotypes in soybean aphid and genetic markers for their detection. *Sci Rep* (in press).
- 15 [FAO] Food and Agriculture Organization, Rome, Italy (1979).
- 16 Hafez AM, Mota-Sanchez D, Vandervoort C, and Wise JC, Resistance affects the field performance of insecticides used for control of *Choristoneura rosaceana* in Michigan apples and cherries, *Insects* 12:846, Multidisciplinary Digital Publishing Institute (2021).
- 17 Mota-Sanchez D, Wise JC, Poppen RV, Gut LJ and Hollingworth RM, Resistance of codling moth, *Cydia pomonella* (L.) (Lepidoptera: Tortricidae), larvae in Michigan to insecticides with different modes of action and the impact on field residual activity. *Pest Manag Sci* 64: 881–890 (2008).
- 18 Yee WL and Alston DG, Effects of spinosad, spinosad bait, and chloronicotinyl insecticides on mortality and control of adult and larval western cherry fruit fly (Diptera: Tephritidae). J Econ Entomol 99: 1722–1732 (2006).
- 19 Reissig WH, Field and laboratory tests of new insecticides against the apple maggot, *Rhagoletis pomonella* (Walsh) (Diptera: Tephritidae). *J Econ Entomol* **96**:1463–1472 (2003).
- 20 Tabashnik BE, Mota-Sanchez D, Whalon ME, Hollingworth RM and Carrière Y, Defining terms for proactive management of resistance to Bt crops and pesticides. J Econ Entomol 107:496–507 (2014).
- 21 [IRAC] Insecticide Resistance Action Committee, Resistance Basics: Resistance definition, IRAC, 2013. https://irac-online.org/about/ resistance/ [accessed 3 March 2022].
- 22 Fong Y, Di C, Huang Y and Gilbert PB, Model-robust inference for continuous threshold regression models. *Biometrics* 73:452–462 (2017).
- 23 Fong Y, Huang Y, Gilbert PB and Permar SR, Chngpt: threshold regression model estimation and inference. *BMC Bioinformatics* 18:454 (2017).
- 24 McManus BL and Fuller BW, Soybean aphid management using foliar applied insecticides in South Dakota, 2011. Arthropod Manag Tests 38:F65 (2013).
- 25 McManus BL, Fuller BW and Giddings LG, Soybean aphid management in South Dakota with foliar applied insecticdes, 2014. *Arthropod Manag Tests* **41**:tsw094 (2016).
- 26 Dana LA, Hunt TE and Zahn DK, Control of soybean aphid with foliar insecticides in soybean, 2011. Arthropod Manag Tests 37:F72 (2012).
- 27 Dana LA and Hunt TE, Control of soybean aphid with foliar insecticdes in soybeans, 2013. Arthropod Manag Tests **39**:F12 (2014).
- 28 Knodel J and Bradley C, Efficacy of foliar applied insecticidesfungicides against soybean aphids on soybeans, 2006. Arthropod Manag Tests 32:F54 (2007).
- 29 Hodgson EW and VanNostrand G, Evaluation of insecticides for control of soybean aphid, 2009. Arthropod Manag Tests 36:F87 (2011).
- 30 Hodgson EW and VanNostrand G, Soybean aphid efficacy screening program, 2011. Arthropod Manag Tests 37:F80 (2012).
- 31 Tabashnik BE, Patin AL, Dennehy TJ, Liu Y-B, Carrière Y, Sims MA et al., Frequency of resistance to Bacillus thuringiensis in field populations of pink bollworm. Proc Natl Acad Sci U S A 97:12980–12984 (2000).
- 32 Burkness EC, Hutchison WD, Bolin PC, Bartels DW, Warnock DF and Davis DW, Field efficacy of sweet corn hybrids expressing a bacillus

thuringiensis toxin for management of *Ostrinia nubilalis* (Lepidoptera: Crambidae) and *Helicoverpa zea* (Lepidoptera: Noctuidae). *J Econ Entomol* **94**:197–203 (2001).

- 33 R Core Team, *R: A Language and Environment for Statistical Computing.* R Foundation for Statistical Computing, Vienna, Austria (2020).
- 34 R Studio Team, *RStudio: Intergrated Development for R*. RStudio, Boston, MA (2020).
- 35 Elder A and Fong Y, Estimation and inference for upper hinge regression models. *Environ Ecol Stat* **26**:287–302 (2019).
- 36 Thogmartin WE, Szymanski JA and Weiser EL, Evidence for a growing population of eastern migratory monarch butterflies is currently insufficient. *Front Ecol Evol* **8**:43 (2020).
- 37 Steiner MY, Spohr LJ and Goodwin S, Relative humidity controls pupation success and dropping behaviour of western flower thrips, *Frankliniella occidentalis* (Pergande) (Thysanoptera: Thripidae). Aust J Entomol 50:179–186 (2011).
- 38 Lysyk TJ and Colwell DD, Duration of efficacy of diazinon ear tags and ivermectin pour-on for control of horn fly (Diptera: Muscidae). J Econ Entomol 89:1513–1520 (1996).
- 39 Wayment DarceyG, Ledet HJ, Torres KA, and White PM, Soil dissipation of sugarcane billet seed treatment fungicides and insecticide using QuEChERS and HPLC, J Environ Sci Health Part B 56:188–196, Taylor & Francis (2021).
- 40 Maqueda C, Undabeytia T, Villaverde J and Morillo E, Behaviour of glyphosate in a reservoir and the surrounding agricultural soils. *Sci Total Environ* **593–594**:787–795 (2017).
- 41 Capinera JL, Horton DR, Epsky ND and Chapman PL, Effects of plant density and late-season defoliation on yield of field beans. *Environ Entomol* **16**:274–280 (1987).
- 42 Chen, CH, Climate warming and the effectiveness of restoring longitudinal river connectivity for endangered non-anadromous Taiwan salmon (Oncorhynchus formosanus). Doctoral dissertation, University of Washington. (2020). https://digital.lib.washington.edu/research works/handle/1773/45498
- 43 Qi, M, Berry, JC, Veley KW, O'Connor L, Finkel OM, Salas-Gonz'lez I et al., Identification of beneficial and detrimental bacteria impacting sorghum responses to drought using multi-scale and multi-system microbiome comparisons. *ISME J* (2022). https://doi.org/10.1038/ s41396-022-01245-4
- 44 Tabashnik BE, Evolution of resistance to *Bacillus thuringiensis*. Annu Rev Entomol **39**:47–79 (1994).
- 45 Tabashnik BE, Van Rensburg JBJ and Carrière Y, Field-evolved insect resistance to Bt crops: definition, theory, and data. J Econ Entomol 102:2011–2025 (2009).
- 46 Tabashnik BE, Brévault T, and Carrière Y, Insect resistance to Bt crops: lessons from the first billion acres, *Nat Biotechnol* **31**:510–521, Nature Publishing Group (2013).
- 47 [EPA] Messina, E., Chlorpyrifos; Tolerance Revocations. https://www. regulations.gov/document/EPA-HQ-OPP-2021-0523-0001 [accessed 3 March 2022].
- 48 Tran AK, Alves TM and Koch RL, Potential for sulfoxaflor to improve conservation biological control of *Aphis glycines* (Hemiptera: Aphididae) in soybean. *J Econ Entomol* **109**:2105–2114 (2016).
- 49 da Silva Queiroz O, Aita RC and Koch RL, Efficacy of foliar insecticides for management of soybean aphid, 2018. Arthropod Manag Tests 44:tsz040 (2019).
- 50 Koch RL, da Silva Queiroz O, Aita RC, Hodgson EW, Potter BD, Nyoike T et al., Efficacy of afidopyropen against soybean aphid (Hemiptera: Aphididae) and toxicity to natural enemies. *Pest Manag Sci* 76:375– 383 (2020).
- 51 Aita RC, Vieira Ribeiro A and Koch RL, Efficacy of foliar insecticides for management of soybean aphid, 2019. Arthropod Manag Tests 44: tsz089 (2019).
- 52 Potter BD and Vollmer T, 2020 Aphid Insecticide report. University of Minnesota Southwest Research and Outreach Center. https:// swroc.cfans.umn.edu/research/ipm/research-results [accessed 3 March 2022].
- 53 Heidel TT and Ragsdale DW, Efficacy of foliar insecticides and host plant resistance on soybean aphid management, 2009. Arthropod Manag Tests **37**:F76 (2012).
- 54 Koch RL, Alves TM and Hanson AA, Efficacy of foliar insecticides for management of soybean aphid, 2014. *Arthropod Manag Tests* **41**: tsw044 (2016).