# WILEY

# Angiotensin-converting enzyme 2: The old door for new severe acute respiratory syndrome coronavirus 2 infection

Heng Wee Tan 🖻 | Yan-Ming Xu 🖻 | Andy T. Y. Lau 🖻

Laboratory of Cancer Biology and Epigenetics, Department of Cell Biology and Genetics, Shantou University Medical College, Shantou, Guangdong, People's Republic of China

#### Correspondence

Andy T. Y. Lau and Yan-Ming Xu, Laboratory of Cancer Biology and Epigenetics, Department of Cell Biology and Genetics, Shantou University Medical College, 22 Xinling Road, Shantou, Guangdong 515041, People's Republic of China. Email: andytylau@stu.edu.cn (A. T. Y. L.) amyymxu@stu.edu.cn (Y. M. X.)

#### **Funding information**

"Thousand, Hundred, and Ten" project of the Department of Education of Guangdong Province of China; "Yang Fan" Project of Guangdong Province of China, Grant/Award Numbers: Andy T. Y. Lau-2016, Yan-Ming Xu-2015; "Young Innovative Talents" Project of Guangdong Province of China, Grant/Award Number: 2019KQNCX034; Basic and Applied Research Major Projects of Guangdong Province of China, Grant/Award Numbers: 2017KZDXM035, 2018KZDXM036; National Natural Science Foundation of China, Grant/ Award Numbers: 31271445, 31771582; Natural Science Foundation of Guangdong Province, Grant/Award Number: 2017A030313131

#### Summary

Coronavirus (CoV) disease 2019 (COVID-19) is an ongoing pandemic caused by severe acute respiratory syndrome CoV 2 (SARS-CoV-2). The highly contagious SARS-CoV-2 belongs to the genus *Betacoronavirus*, and it is phylogenetically closely related to SARS-CoV, a human CoV that caused an outbreak back in 2002 to 2003. Both SARS-CoV-2 and SARS-CoV enter human cells via the interactions between viral crown-like spike protein and human angiotensin-converting enzyme 2 (ACE2) receptor. Here, we aim to review the involvement of ACE2 in human CoV infections by discussing the roles of ACE2 in CoV evolution, cross-species transmissibility, and COVID-19 susceptibility. We also provide our perspectives on COVID-19 treatment and prevention.

### KEYWORDS

ACE2, COVID-19, SARS-CoV-2

#### 1 | INTRODUCTION

The recent outbreak of a novel coronavirus (CoV) disease 2019 (COVID-19) caused by severe acute respiratory syndrome CoV 2 (SARS-CoV-2; formally 2019-nCoV) has emerged as a global health and economic crisis. The World Health Organization has officially declared COVID-19 a pandemic on March 11, 2020. As of April 28, 2020, the extremely contagious SARS-CoV-2 had already spread

to all continents except Antarctica, infected more than 3 million people and caused at least 210 000 deaths worldwide.

The CoVs are enveloped, positive-sense single-stranded RNA viruses with genomes ranging from 26 to 32 kb, the largest among all known RNA viruses.<sup>1</sup> They belong to the subfamily Orthocoronavirinae (a.k.a. Coronavirinae) in the family Coronaviridae, and can be classified into four genera: *Alphacoronavirus, Betacoronavirus, Gammacoronavirus,* and *Deltacoronavirus.*<sup>2</sup> CoVs are commonly found in mammals, vertebrates, and other wild animals.<sup>3,4</sup> Studies have shown that humans and other mammals are primarily infected by the CoVs of *Alphacoronavirus* and *Betacoronavirus* genera, whereas vertebrates are usually infected by *Gammacoronavirus* and *Deltacoronavirus*.<sup>5</sup>

To date, scientists have identified seven strains of CoVs that infect humans. Some strains such as HCoV-229E, HCoV-NL63,

Abbreviations: ACE2, angiotensin-converting enzyme 2; Ang, angiotensin; CoV, coronavirus; COVID-19, coronavirus disease 2019; cryo-EM, cryogenic electron microscopy; MERS-CoV, Middle East respiratory syndrome coronavirus; RAS, renin-angiotensin system; RBD, receptor-binding domain; rhACE2, recombinant human ACE2; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; TCGA, The Cancer Genome Atlas; TMPRSS2, transmembrane protease serine 2.

# <sup>2 of 12</sup> WILEY-

HCoV-OC43, and HCoV-HKU1 are regularly circulating in the human population and are responsible for respiratory infections.<sup>6,7</sup> People infected with these CoVs generally show mild symptoms of upper respiratory disease such as fever, sore throat, and coughing, and only on rare occasion, they may cause lower respiratory tract infections and severe pneumonia.<sup>8</sup> However, certain human CoV strains, such as SARS-CoV-2, SARS-CoV, and Middle East respiratory syndrome CoV (MERS-CoV), are associated with higher fatality rates. SARS-CoV and MERS-CoV have previously claimed over a thousand lives in total during the CoV outbreaks in 2002 to 2003 and 2012, respectively.<sup>9,10</sup> In addition to human CoVs, animal CoVs also could cause a significant loss to the animal husbandry. One of the most recent examples is the swine acute diarrhea syndrome CoV outbreak in 2017 to 2018.<sup>11</sup> All these incidences signify that the CoVs have continually posed a serious threat to the well-being of humans and animals.<sup>9,10</sup>

Genome sequencing and phylogenetic analysis of the newly emerged SARS-CoV-2 has placed it under the genus *Betacoronavirus*, making it a close relative to another two notorious CoVs: SARS-CoV and MERS-CoV.<sup>4</sup> The *Betacoronavirus* also contains a range of bat CoV strains, including several strains that show high % sequence similarity to the SARS-CoV-2, suggesting that SARS-CoV-2 is likely originated from the bats.<sup>12-14</sup> However, the intermediate reservoir of SARS-CoV-2 remains unclear.

The surface of all CoVs has a characteristic crown-like structure known as the spike protein (commonly referred to as S protein). The spike protein can bind to a specific cell membrane receptor and it is the key mediator for CoV entry into host cells.<sup>15</sup> Previously, angiotensin-converting enzyme 2 (ACE2) was identified as the target receptor for SARS-CoV.<sup>16-18</sup> Current studies have indicated that the novel SARS-CoV-2 also uses the ACE2 receptor.<sup>14,19</sup> Here, we review the involvement of ACE2 in human CoV infections by discussing the significances of ACE2 in relation to CoV evolution, cross-species transmissibility, and COVID-19 susceptibility. Lastly, we provide our perspectives on COVID-19 treatment and prevention. Also, since COVID-19 is an ongoing pandemic, some of the first-hand data discussed in this review are sourced from non-peer-reviewed preprints.

#### 2 | PROPERTIES AND BIOLOGICAL FUNCTIONS OF ACE2

ACE2, a homologue of ACE, was firstly described 20 years ago.<sup>20,21</sup> Both ACE2 and ACE are zinc metalloproteases that play crucial roles in the renin-angiotensin system (RAS), a system that regulates blood pressure, fluid, and electrolyte homeostasis.<sup>22,23</sup>

Human ACE2 is a protein with 805 aa encoded by the ACE2 gene (HGNC: 13557) while ACE is a larger protein consists of 1306 aa encoded by the ACE gene (HGNC: 2707). ACE2 and ACE share approximately 40% identity and 61% similarity in their aa sequences.<sup>21</sup> Despite the similarity, ACE and ACE2 do not share the same substrate specificity.<sup>24</sup> Also, ACE inhibitors that commonly used for treating high blood pressure or cardiovascular and kidney diseases, such as captopril, enalaprilat, and lisinopril, are ineffective against

ACE2.<sup>24</sup> In the RAS, ACE2 acts as a potent counter-regulator against ACE.<sup>25</sup> Physiologically, ACE converts inactive decapeptide angiotensin (Ang) I into vasoconstrictor Ang II and degrades vasodilator bradykinin, leading to increased blood pressure.<sup>20</sup> ACE2, on the other hand, decreases blood pressure by competing with ACE to hydrolyze Ang I into the nonapeptide Ang-(1–9), and at the same time degrades Ang II into Ang-(1–7) and promote the release of vasodilator bradykinin.<sup>20,26</sup>

ACE2 and ACE are mainly expressed in the cell membrane of vascular endothelial cells found in various organs. Generally, ACE is more widespread than ACE2 with highest levels of expression observed in, but not limited to, gastrointestinal tract, kidney, and lung.<sup>21,27</sup> For ACE2, gallbladder, gastrointestinal tract, heart, kidney, and testis are the primary organs of expression.<sup>27,28</sup> Both ACE2 and ACE can be secreted from the cell surface into the circulation or urine.<sup>20,29,30</sup> Aberrant expression of ACE or ACE2 is associated with many diseases, including hypertension, lung injury, and cardiovascular, renal, and liver diseases.<sup>31-33</sup> ACE2 is also known to be involved in humanand animal-CoV infections. The high-resolution cryogenic electron microscopy (cryo-EM) structure of full-length human ACE2 was recently revealed, and its interactions with SARS-CoV or SARS-CoV-2 were determined.<sup>34</sup>

#### 3 | ACE2 AND CORONAVIRUS PATHOGENESIS

The interactions between spike protein and host receptor are critical for CoV pathogenesis. The spike protein is a crown-shaped class I viral membrane fusion protein distributed throughout the surface of all CoVs.<sup>35</sup> It is made up of a short intracellular tail and a large ectodomain connected by a single-pass transmembrane anchor.<sup>36</sup> The ectodomain consists of two subunits: three S1 subunit heads resting above a trimeric S2 subunit stalk.<sup>37</sup> The S1 subunit is responsible for host receptor-binding while the S2 subunit is accountable for creating an entrance for the viral genomes to invade the host cells by fusing the viral and host membranes.<sup>35,38</sup> Structural studies on the S1 subunit have revealed two receptor-binding domains (RBDs) that can interact with a variety of receptors. Specifically, the N-terminal domain mainly binds sugar receptors and CEACAM1 in mouse hepatitis CoV<sup>39-42</sup> whereas the C-terminal domain appears to bind protein receptors (eg, APN, ACE2, and DDP4) more exclusively.<sup>38,42-46</sup> In order to bind a host-cell receptor, the RBD undergoes hinge-like conformational movements that either buried (lying state; receptorinaccessible state) or exposed (standing state; receptor-accessible state) its receptor-binding regions.<sup>47</sup> Some CoVs, such as lineage A Betacoronavirus, have a shorter spike-like envelope-associated hemagglutinin-esterase that acts as a receptor-destroying enzyme.48,49 Structures and functions of CoV spike proteins are reviewed extensively in Li.<sup>2</sup>

So far, three CoVs (HCoV-NL63, SARS-CoV, and SARS-CoV-2) have shown to utilize human ACE2 receptor. However, the RBDs of these three ACE2-utilizing human CoVs do not share identical sequences. Previously, the interactions between ACE2 and S1 domain

of SARS-CoV spike protein was firstly reported by Li et al,<sup>16</sup> and the structure of ACE2-spike protein complex had been determined.<sup>50</sup> Research indicated that the ACE2 protein could bind directly with the extended tyrosine-enriched loop on the RBD (residues 424-494) of SARS-CoV spike protein<sup>50,51</sup> and that the distantly-related ACE protein did not have the same function as ACE2.<sup>16</sup> Interestingly, although HCoV-NL63 also interacts with ACE2, its RBD shares no sequence and structural similarity with the SARS-CoV.<sup>52</sup> In fact, unlike SARS-CoV that binds the ACE2 through a continuous subdomain, HCoV-NL63 binds the receptor with three discontinuous beta-loops.<sup>52</sup> Research has indicated that both HCoV-NL63 and SARS-CoV bind overlapping regions of ACE2 and that mutations in the ACE2 protein generally resulted in the same binding properties of these two strains.<sup>17</sup> However, some mutations in the ACE2 protein only hinder the interactions of either HCoV-NL63 or SARS-CoV, indicating that both strains engage ACE2 differently.<sup>17,53</sup>

The engagement of SARS-CoV-2 with ACE2 is similar to SARS-CoV, and important features of the spike protein in SARS-CoV-2 and its most related CoVs are compared and summarized in Andersen et al.54 Virus infectivity study has indicated that the SARS-CoV-2 is able to utilize ACE2 of human, Chinese horseshoe bats, civet, and pig but was not able to use mouse ACE2.14 The crvo-EM structure of SARS-CoV-2 spike protein was recently determined.<sup>55</sup> Results indicated that the spike proteins of SARS-CoV and SARS-CoV-2 were highly homologous and structurally similar, with minor differences in the position of their RBDs in receptor-inaccessible states: RBD of SARS-CoV packs tightly against the N-terminal domain of the neighboring protomer whereas RBD of SARS-CoV-2 angles closer to the central cavity of the trimer.<sup>55</sup> Notably, the affinity between ACE2 and the spike protein of HCoV-NL63 is weaker compared to SARS-CoV and SARS-CoV-2, and although SARS-CoV and SARS-CoV-2 share similar spike protein and RBD sequences, the affinity between the spike protein of SARS-CoV-2 and ACE2 appears to be 10 to 20 folds higher than SARS-CoV and ACE2, suggesting that SARS-CoV-2 has a much higher human-to-human transmissibility compared to SARS-CoV and HCoV-NL63.<sup>6,52,53,55</sup> In addition, both SARS-CoV and SARS-CoV-2 use the same host transmembrane protease serine 2 (TMPRSS2) and possibly other proteases (eg, cathepsin B and cathepsin L) to cleave their spike protein and enhance viral entry.<sup>56,57</sup> Therefore, other host factors in addition to ACE2 are likely also contributed to human CoV pathogenesis and required further study.

#### 4 | ACE2 AND CORONAVIRUS EVOLUTION

Sequences in the spike proteins determine the tissue and species tropism of CoVs. Mutations in the RBD of spike protein not only could affect the virulence of CoVs, but also could alter viral host spectrum, allowing cross-species infection to happen.<sup>36,37,51</sup> RNA viruses can rapidly evolve and adapt to new environments due to their high mutation rates associated with the infidelity of RNA polymerase.<sup>58</sup> This feature of RNA viruses increases the probabilities of CoVs to invade a new host population. In fact, a substantial number of emerging pathogens that caused major epidemics in the past few decades are by RNA viruses.  $^{\rm 59}$ 

Recognizing the origin and host of CoVs is extremely critical for disease control and prevention. Understanding the underlying molecular mechanism of human receptor usage by different CoV strains is equally important. Bats are likely the ancestors of five out of seven human CoVs.<sup>3,15,60,61</sup> They are the natural reservoirs for CoVs and many other viruses such as Ebola, Hendra, Marburg, and Nipah.<sup>62</sup> Studies have suggested that the diverse viral inhabitants within and between bat species may perhaps promote co-evolution of these zoonotic viruses, further increasing the chances of cross-species transmission.<sup>62-64</sup> The relationships of Betacoronavirus RBDs between batand human-CoVs are reviewed in Cui et al<sup>3</sup> and Lu et al.<sup>65</sup> A review article from Fan et al has postulated that China is a hotspot for future bat-orientated CoV outbreaks due to multiple reasons, including the track record of bat CoV outbreaks in human and animals, high population density, great wildlife diversity, and coexistence of diverse viruses in bats.<sup>61</sup> However, CoV outbreaks could likewise happen anywhere in the world since mutations of CoVs in bats or other wild animals are occurred by chance.

Despite overwhelming data pointed toward bats as the progenitor of most human CoVs, the evidence of bat CoVs to cross-transmit directly to humans is still lacking.<sup>60</sup> Studies have suggested that all human CoVs have or require intermediate reservoirs before crosstransmitting from bats into humans.<sup>3</sup> For instance, the potential intermediate hosts of bat-orientated strains HCoV-229E, SARS-CoV, and MERS-CoV are identified as camelids.<sup>64,66</sup> palm civets.<sup>67</sup> and dromedary camels,<sup>68</sup> respectively. HCoV-NL63 is also originated from the bats, but the intermediate host remains unclear.<sup>64</sup> Native hosts of HCoV-OC43 and HCoV-HKU1 are likely rodents, and cattle might be the intermediate host for HCoV-OC43.8,52 As for SARS-CoV-2, the phylogenic analysis indicated that this new CoV strain is also of a bat origin (≈96% similarity),<sup>12-14</sup> and smuggled Malayan pangolins in China's border are currently the prime suspect of being its intermediate host.<sup>69</sup> The spike protein RBD of pangolin CoV strains showed ≈99% similarity to SARS-CoV-2, with only one aa difference. The binding capacity of pangolin RBD was tested, and it showed that the spike proteins of pangolin CoV and SARS-CoV-2 could potentially bind to both pangolin and human ACE2.<sup>69</sup> However, despite the high % similarity in the RBD region of pangolin CoV and SARS-CoV-2, they shared only a mere  $\approx$ 90% similarity of their overall genomes.<sup>69</sup> A few more similar studies also analyzed CoV strains isolated from pangolin in China, and although pangolin CoVs are considered phylogenetically closer to SARS-CoV-2, the % genome similarity was only 92.4% at best.<sup>70-72</sup> Since closely related CoVs should have higher genome similarity, like those observed between SARS-CoV and CoVs isolated from its potential intermediate host palm civet (up to 99.8% similarity), whether pangolin is truly the intermediate host for SARS-CoV-2 is still in question.<sup>73</sup>

All three human CoVs (HCoV-NL63, SARS-CoV, and SARS-CoV-2) that utilized ACE2 are originated from bats. Human CoV strains appeared to engage human ACE2 differently, mainly through targeting different positions of the ACE2 protein (reviewed in Li et  $a|^{17}$ ).

# 4 of 12 WILEY-

Generally, higher ACE2 affinity is correlated with higher infectibility of CoVs, and those strains with a higher ACE2 affinity may eventually be the dominant circulating strains in human. Few months into the COVID-19 outbreak, more than a hundred mutations were already detected in the SARS-CoV-2 genomes isolated from 103 COVID-19 patients.<sup>74</sup> These SARS-CoV-2 strains could be subdivided into two major types: the highly contagious and faster-growing L-type that found in  $\approx$ 70% of samples and the evolutionarily older and less aggressive S-type that found in  $\approx$ 30% of samples.<sup>74</sup> Similarly, SARS-CoV strains previously isolated from SARS patients showed vary ACE2 affinities: some strains had a moderate affinity for human ACE2 whereas some strains, especially those isolated during the late phase of the SARS-CoV outbreak, had a higher affinity.<sup>75,76</sup> SARS-like CoV strains isolated from palm civets during and after SARS-CoV outbreak showed high affinity for civet ACE2 but lower affinity for human ACE2, and consequently, higher infectivity in civet cells compared with human cells.<sup>73,77</sup> The spike protein RBDs of SARS-CoV and civet CoV only differ by two residues, and these differences are enough to cause SARS-CoV to bind human ACE2 more effectively than civet CoV.<sup>78</sup> Thus, the host range, tissue tropism, and receptor-binding property of CoVs can change dramatically simply due to a few mutations on the spike protein RBD. Previously, aa positions of human ACE2 that may play a role in host range and cross-species infection of CoVs were identified: K31, E35, D38, Y41, M82, and K353.<sup>10</sup>

Chinese horseshoe bats are considered the native host for SARS-CoV even though the overall genetic similarity between SARS-CoV and SARS-like civet CoVs is greater than that between SARS-CoV and SARS-like bat CoVs.<sup>79,80</sup> SARS-like bat CoVs share identical genome

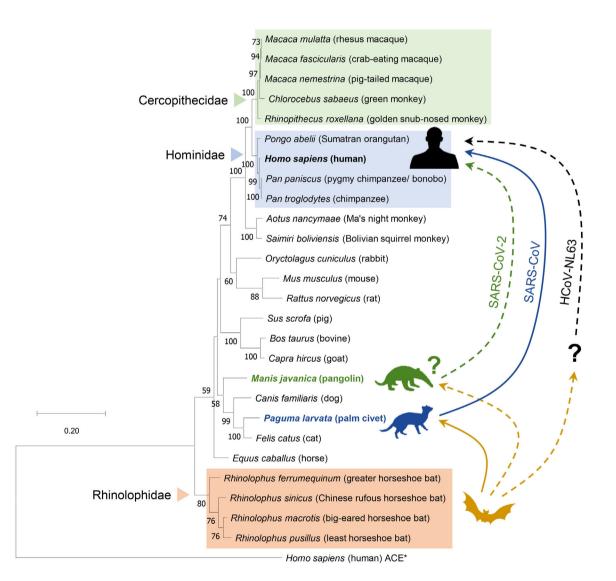


FIGURE 1 Phylogenetic tree of human and animal angiotensin-converting enzyme 2 (ACE2). Amino acid sequences were obtained from UniProt or National Center for Biotechnology Information and aligned by ClustalW using MEGA X (v10.1.7).<sup>88</sup> Maximum likelihood tree was constructed using Jones-Taylor-Thornton model with 100 bootstrap replications. Numbers at the branches represent bootstrap probability values ≥50%. The known and suspected hosts of human coronaviruses that utilized human ACE2 receptors (HCoV-NL63, SARS-CoV, and SARS-CoV-2) are shown. Solid arrows represent interspecies transmission with substantial evidence; broken arrows represent suspected interspecies transmission. \*Human ACE, a homologue of ACE2, was used as an outgroup

organizations and high sequence similarity with SARS-CoV, and these SARS-like CoVs can be found in diverse bat populations but not in other mammals.<sup>81,82</sup> Despite being phylogenetically related, the spike protein RBDs of SARS-like bat CoVs were not identical to SARS-CoV, and bat CoVs were generally not able to utilize human ACE2 or any ACE2 ortholog tested.<sup>83</sup> On the other hand, SARS-CoV was able to infect palm civet but was inefficient against bats.<sup>83,84</sup> However, in other studies, several bat CoV strains that had the ability to utilize human ACE2 were identified, suggesting that the direct spillover of novel CoVs from bat to human is certainly possible.<sup>12,85,86</sup>

The relationships between human CoVs and ACE2 of other different animal species have not been systematically assessed. Previously, SARS-CoV was not able to bind rat ACE2, but it could interact with mouse ACE2 with low affinity.<sup>10,84</sup> Comparing the ACE2 sequences of human and other mammalians may give us some clues on crossspecies receptor usage of SARS-CoV-2.87 Here, we performed a phylogenetic analysis of ACE2 from a range of mammals and found that human ACE2 are most closely related to other species in the same family Hominidae (chimpanzee and orangutan; Figure 1).<sup>88</sup> Furthermore, evolutionary inference suggests that species from the family Cercopithecidae (macaque and monkey) also have ACE2 similar to human ACE2 (Figure 1). Previously, human CoV strain OC43 was detected in fecal samples of macaque and chimpanzee, signifying the potential of CoV spillovers among human, macaque, chimpanzee, and monkey.<sup>89,90</sup> Unfortunately, research regarding CoVs in the abovementioned primates is scarce, and further studies are required to examine if one of these species could be the intermediate host for SARS-CoV-2. Furthermore, there is a rising concern regarding the transmission of COVID-19 in companion pets such as cats and dogs. Although so far there is no evidence that household pets can spread COVID-19, it appears that some animals, especially those in the family Felidae (eg, cat and Malayan tiger), may contract SARS-CoV-2 from humans.<sup>91,92</sup> At present, the roles of ACE2 in domestic animals in connection to their infection with SARS-CoV-2 plausibly transmitted from humans have remained unclear.

#### 5 | ACE2 EXPRESSION AND COVID-19 SUSCEPTIBILITY

The roles of ACE2 expression in SARS-CoV-2 pathogenesis and human COVID-19 susceptibility are largely unknown. Because SARS-CoV-2 uses ACE2 as the receptor, it is logical to assume that all human cells that express ACE2 are the potential targets for SARS-CoV-2. Therefore, the following questions regarding ACE2 expression in relation to COVID-19 susceptibility can be asked: (a) Which parts of the body express ACE2? (b) Which ACE2-expressing organs are prone to SARS-CoV-2 infection? (c) Is higher expression level of ACE2 equal to greater COVID-19 susceptibility?

ACE2 is expressed in a tissue- and species-specific pattern.<sup>93-95</sup> In human, ACE2 is expressed in virtually all organs, with gastrointestinal tract, heart, and kidney among the highest expressed organs.<sup>27,28,96</sup> Lung is the main target for SARS-CoV-2, but the overall expression of ACE2 in human lung vary significantly according to different studies-some studies indicated that lung expressed high levels of ACE2<sup>94</sup> while some suggested otherwise.<sup>27,97</sup> Using single-cell RNA sequencing, Zhao et al showed that the lung ACE2 explicitly expressed in a small population of type II alveolar cells, and these ACE2-expressing cells also highly expressed certain genes that involved in promoting viral reproduction and transmission.<sup>98</sup> Additionally, they found that age or smoking was not associated with ACE2-expressing lung cell number but revealed that Asian had higher lung ACE2 expression than white and African American, suggesting Asian might be more susceptible to COVID-19.<sup>98</sup> However, the above study only tested on eight samples (with only one Asian). In another study, 224 samples were collectively analyzed for ACE2 expression in healthy lungs, and results indicated that lung ACE2 expression was not significantly associated with age, gender, or race.<sup>99</sup> This study. however, did find that current and former smokers had higher ACE2 expression compare with nonsmokers. Additionally, ACE2 was specifically expressed in different lung cell types of smokers and nonsmokers, suggesting that not only smokers might be more susceptible to COVID-19, but the viral infection path could be different between smokers and nonsmokers.<sup>99</sup> A recent study also found increased ACE2 expression in lower airways of smokers and individuals with chronic obstructive pulmonary disease.<sup>100</sup> Further investigation is required to verify if higher lung ACE2 expression is associated with greater COVID-19 susceptibility.

Individuals with pre-existing disease may be more prone to COVID-19. In vitro study showed that human lung cells exposed to inflammatory cytokines, rhinovirus, H1N1 influenza, or CoV (SARS-CoV or MERS-CoV) resulted in increased expression of ACE2.<sup>101</sup> These data suggest that people with viral infection and/or high inflammatory cytokine levels could be more susceptible to COVID-19. Also, increased cytokines during virus infection may further accelerate COVID-19 infection.

Another two potential risk factors for COVID-19 are obesity and cancers.<sup>102</sup> Research has shown that human adipose tissues express high levels of ACE2.<sup>102</sup> In diabetic mice, the ACE2 expressions in the serum, liver, and pancreas were up-regulated.<sup>103</sup> By utilizing online databases (HCCDB, UALCAN, and GEPIA2), Jia et al showed that tumor tissues of specific types of cancers appeared to have higher levels of ACE2 compared with their adjacent noncancerous tissues.<sup>102</sup> Here, we analyzed ACE2 expression in 41 types of cancers obtained from The Cancer Genome Atlas (TCGA) database,<sup>104</sup> and found that for those that showed significant differences in their ACE2 expression between tumor and reference samples, almost all were up-regulated and only four samples (out of 482) from the papillary thyroid carcinoma showed down-regulated ACE2 (Figure 2A). In summary, it seems like tumor tissues tend to have up-regulated ACE2 expression, especially in neuroendocrine prostate cancer and ovarian serous cystadenocarcinoma. Our findings in Figure 2A may have clinical implications for COVID-19. Indeed, increased risk of COVID-19 and poorer prognosis was observed in cancer patients.<sup>105</sup> In addition, using similar methods as described previously,<sup>106</sup> we compared the ACE2 expression in lung cancer patients of various smoking statuses. 6 of 12 WILEY-

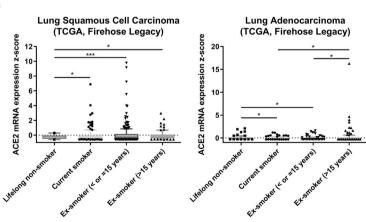
(A)

(~)		Down-Regulated	Up-Regulated	
	-20	0% 0'	% +2	.0%
	Samples (n)			
Acral Melanoma	33	0	3.03	1
Acute Myeloid Leukemia	173	0	2.31	
Adrenocortical Carcinoma	78	0	12.82	
Adult Soft Tissue Sarcomas	206	0	3.4	
Bladder Urothelial Carcinoma	407	0	2.21	
Breast Invasive Carcinoma	1082	0	0.74	
Cervical Squamous Cell Carcinoma	294	0	0.68	
Cholangiocarcinoma	36	0	2.78	
Colorectal Adenocarcinoma	592	0	10.3	
Diffuse Large B-Cell Lymphoma	48	0	8.33	
<b>Esophageal Adenocarcinoma</b>	181	0	6.63	
Glioblastoma Multiforme	155	0	6.45	
Head and Neck Squamous Cell Carcinoma	515	0	6.8	
Kidney Chromophobe	65	0	9.23	
Kidney Renal Clear Cell Carcinoma	510	0	7.65	
Liver Hepatocellular Carcinoma	366	0	6.01	
Lung Adenocarcinoma	510	0	2.16	
Lung Squamous Cell Carcinoma	484	0	4.96	
Mature B-cell malignancies	290	0	3.45	
Melanoma	21	0	4.76	
Mesothelioma	87	0	2.3	
Metastatic Melanoma	40	0	2.5	
Metastatic Prostate Adenocarcinoma	205	0	2.44	
Neuroendocrine Prostate Cancer	35	0	17.14	
Ovarian Serous Cystadenocarcinoma	300	0	16.33	
Pancreatic Adenocarcinoma	177	0	4.52	
Papillary Thyroid Carcinoma	482	0.829	8.3	
Pediatric Acute Lymphoid Leukemia	154	0	12.34	
Pediatric Neuroblastoma	139	0	5.04	
Pediatric Rhabdoid Tumor	43	0	11.68	
Pediatric Wilms' Tumor	125	0	13.6	
Pheochromocytoma and Paraganglioma Prostate Adenocarcinoma	178 493	0	1.69	
Prostate Adenocarcinoma Sarcoma	493 253	0 0	4.46 3.16	
Sarcoma Skin Cutaneous Melanoma	233 441	0	9.52	
Stomach Adenocarcinoma	441 412	0	5.34	
Testicular Germ Cell Cancer	412 150	0	6	
Testicular Germ Cell Cancer Thymoma	119	0	10.92	
Upper Tract Urothelial Carcinoma	32	0	6.25	
Uterine Corpus Endometrial Carcinoma	527	0	3.61	
Uveal Melanoma	80	0	8.75	
	-• [	2		1

Samples with altered ACE2 expression (%)

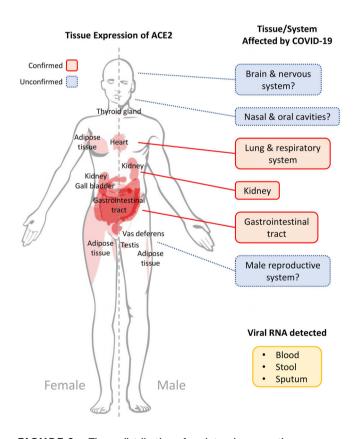
**FIGURE 2** Cancer tissues and the lungs of smokers generally contained higher levels of angiotensin-converting enzyme 2 (ACE2), and these tissues can be potentially more susceptible to COVID-19. A, ACE2 expression data of 41 types of cancers obtained from The Cancer Genome Atlas (TCGA) database<sup>104</sup>; mRNA expression z-score threshold was set at ±1.5. B, ACE2 expression in lung cancer patients with different smoking histories analyzed using similar methods as described previously<sup>106</sup>

(B)



We found that smokers, including ex-smokers, generally have upregulated ACE2 (Figure 2B). These results are to some extent in agreement with Cai's and Leung's studies<sup>99,100</sup> that smoking may increase lung ACE2 expression. SARS-CoV was previously shown not only able to affect the lung but also other organs, including brain, gastrointestinal tract, kidney, liver, and more.<sup>107,108</sup> There is no doubt that the lung is the main target for SARS-CoV-2. However, many COVID-19 patients also showed other symptoms in addition to respiratory symptoms, suggesting that SARS-CoV-2 could perhaps infect other organs (Figure 3). SARS-CoV-2 might infect other ACE2-expressing tissues via blood circulation as a recent report showed that SARS-CoV-2 RNA had been detected in the blood of some COVID-19 patients.<sup>109</sup> Intriguingly, it is recently indicated that higher circulating ACE2 is associated with milder symptoms of COVID-19, suggesting the protective role of secreted ACE2, probably by minimizing the direct contact between SARS-CoV-2 and ACE2-expressing tissues.<sup>110</sup> This may also explain why women and children with confirmed COVID-19 generally respond better to the disease because they appear to have higher levels of ACE2 in the blood.<sup>110</sup>

With a great proportion of COVID-19 patients developed renal abnormalities and acute kidney injury, the kidney seemed to be a major organ affected by the disease.<sup>97,111,112</sup> A consecutive cohort study of COVID-19 patients carried out by Cheng et al showed that older patients and/or those with kidney impairments had higher risks of in-hospital death.<sup>113</sup> The liver is another organ that may be prone to SARS-CoV-2 infection as studies have shown that ACE2 is enriched in the cholangiocytes, and liver injury is observed in some of the COVID-19 patients.<sup>114,115</sup> It is worth noting that although kidney and liver generally have higher ACE2 expressions than the lung, there is so



**FIGURE 3** Tissue distribution of angiotensin-converting enzyme 2 (ACE2) expression and potential COVID-19 susceptibility. Organ highlighted in red represents positive ACE2 expression. Human images and ACE2 expression data were obtained from The Human Protein Atlas (http://www.proteinatlas.org)<sup>27,96</sup>

far no evidence that the SARS-CoV-2 can directly infect the liver and kidney cells in the human body.

The gastrointestinal tract, especially the duodenum and small intestine, expressed high levels of ACE2.<sup>27,96</sup> Since about up to 10% of COVID-19 patients have shown gastrointestinal symptoms such as diarrhea, it is possible that SARS-CoV-2 can infect the gastrointestinal tract.<sup>116</sup> Clinically, the detection of viral RNA based on oral swab was used for confirmation of COVID-19 because the sputa of COVID-19 patients contained SARS-CoV-2 RNA during infection (before the onset of symptoms).<sup>117</sup> Interestingly, it was shown that in some cases, viral RNA could be detected in the sputum even after 2 weeks of clinical recovery.<sup>118</sup> In addition to sputum, SARS-CoV-2 RNA has been detected in the stools of a COVID-19 patient,<sup>119</sup> representing the possibility of fecal-oral transmission of SARS-CoV-2.<sup>120</sup> However, further tests are required to check if infectious SARS-CoV-2 is present in the stools, as well as the blood, sputa, and body fluids.

Reports regarding how COVID-19 could affect testis and brain have also emerged. Studies have suggested that COVID-19 may negatively impact male fertility<sup>112,121</sup> and may cause neurological symptoms and brain injury.<sup>122,123</sup> Furthermore, olfactory and taste disorders have recently been reported in some COVID-19 patients,<sup>124</sup> and whether such symptoms are consequences of neurological/brain damage or direct effects on the epithelial cells of nasal and oral mucosa by SARS-CoV-2 infection have yet to be answered. Latest studies have shown that ACE2 was highly expressed on the tongue and nasal epithelial cells, suggesting that the nasal and oral cavities could be susceptible to SARS-CoV-2 infection.<sup>125,126</sup>

# 6 | PERSPECTIVES: TREATMENT AND PREVENTION

Since the world has now become more interconnected than ever before, more frequent and faster-moving pandemics are anticipated. The ongoing COVID-19 pandemic has reminded us that the CoVs, with their rapidly evolving nature, continue to pose a serious global health threat. To combat the emerging COVID-19, several strategies have been used or proposed. However, most of the potential treatment options for COVID-19 discussed in this section have either not been assessed by randomized controlled clinical trials or are currently under clinical trials.

Hundreds of clinical trials targeting COVID-19 have been rapidly set up around the world amid COVID-19 outbreak. More than a few dozens of drugs are being tested, and some of them that attract the greatest attention include broad-spectrum antiviral remdesivir, anti-HIV protease inhibitors lopinavir/ritonavir, antimalaria chloroquine/ hydroxychloroquine, and traditional Chinese medicine.<sup>127,128</sup> Notably, on February 15, 2020, the Chinese health authority has approved Favilavir (formerly known as fapilavir) to be used in COVID-19 treatment, making it the first official anti-COVID-19 drug. Nevertheless, the efficiencies of all COVID-19 drugs have to be properly assessed by randomized double-blind placebo-controlled studies.<sup>129</sup> A healthy lifestyle is essential to keep the immune system active, and a balance host immune response is important to overcome virus infection.<sup>1</sup> Supplementations such as vitamin A and vitamin D could be used to strengthen the immune system and provide benefits in defending virus infection.<sup>130,131</sup> Specifically, regular intake of vitamin D is correlated with higher protection against respiratory tract infections.<sup>132</sup> Moreover, it is shown that zinc, an essential micronutrient, can inhibit viral RNA-dependent RNA polymerase of SARS-CoV and other viruses.<sup>133</sup> Selenium, another essential micronutrient involved in RAS and immune system,<sup>134</sup> has been shown to increase the immune responses of Se deficient patients and animal models upon CoV and other viral infections.<sup>135,136</sup>

In the century-long of war against viruses, vaccination is considered one of the most effective proactive measures to prevent the spread of viral diseases. Previously, multiple attempts have been made to create CoV vaccines, but these vaccines mainly only focused on SARS-CoV or MERS-CoV and have not yet undergone clinical trials.<sup>56,137</sup> However, in just less than 2 months after COVID-19 is declared a global pandemic, a few SARS-CoV-2 vaccines are already being trialed in humans as scientists in different countries raced to create the first effective COVID-19 vaccine. Spike protein of CoV is a major inducer of host immune responses and the main target of neutralizing antibodies during infection.<sup>37</sup> Recently, potential T- and B-cell epitopes of SARS-CoV-2 for vaccines and antibody-based neutralization have been identified using structural bioinformatic and machine learning techniques-the spike protein RBD region of 494-508 appear to be the best vaccine candidate.<sup>138</sup> DNA/RNA vaccines will likely be the most cost-effective form of vaccination against the fast mutating CoVs, if successfully developed. The fundamental idea of DNA- or RNA-based vaccine is to induce host primary immune response by injecting DNA/RNA that encodes a viral protein (eg. spike protein of CoV) into the body, and let the body cells produce the mimic protein for antibody stimulation and immunological memory.<sup>139,140</sup>

Based on the past success with HIV-1 treatment, gene therapybased strategy to combat COVID-19 could also be considered.<sup>141</sup> For instance, lung cells that targeted by SARS-CoV-2 could be geneedited to become COVID-19 resistant by knockdown or knockout of SARS-CoV-2 pathogenesis-related host gene(s) or permanent gene disruption of the CoV genome. Also, peptide mimetics or smallmolecule inhibitors have been developed to block viral entry or to suppress viral infection, and similar strategies can also be applied to COVID-19.142 Particularly, targeted immunotherapy using smallmolecule inhibitors to prevent cytokine storm syndrome has emerged as a potent treatment option for COVID-19 patients.<sup>143</sup> The ultimate objective of targeted immunotherapy is to optimize the cytokine productions and inflammatory responses upon viral infections.<sup>1</sup> Previously, ACE2 inhibitors have been tested in SARS-CoV and HCoV-NL63, but their antiviral activity against ACE2-utilizing CoVs has remained unknown.<sup>144,145</sup> Although it is possible to create an ACE2 inhibitor that targeting the ACE2 binding region of CoVs or create a recombinant human ACE2 (rhACE2) to trap SARS-CoV-2, the effects of these inhibitors or rhACE2 on the RAS should also be evaluated carefully. Studies showed that the use of serine protease inhibitors to block TMPRSS2 could increase survival in mice upon SARS-CoV infection and reduce the entry of SARS-CoV and SARS-CoV-2 from entering the human cells.<sup>57,146</sup> However, inhibiting host proteases will likely lead to adverse side effects, and therefore, this approach is considerably less attractive than targeting viral proteases. Recently, the crystal structure of SARS-CoV-2 main protease was determined, and this would facilitate the development of viral protease inhibitors specifically targeting SARS-CoV-2.<sup>147</sup> Lastly, reports have suggested that convalescent plasma collected from recovered COVID-19 patients could be used to treat patients in critical conditions, and this method should be further explored.<sup>148</sup>

To conclude, there are still many unknowns regarding COVID-19. Under the existing health emergency, critical knowledge of COVID-19 such as length of SARS-CoV-2 incubation period, route of transmission, viability outside a host, and intermediate host are urgently needed. Since there is currently no effective vaccine or treatment for COVID-19, several precautions should be taken to minimize the spread of this highly contagious virus. For individuals: avoid crowded places, keep distance with and wear fitted mask when sick people are around, maintain personal hygiene, wash hands regularly, avoid touching eyes, nose, and mouth, and stay isolated and seek medical assistance if show suspicious symptoms or happen to get in close contact with sick people; for relevant authorities: identify the source of infection and potential threats, provide sufficient diagnostic tools, offer accessible medical support, advertise proper public health knowledge, rapid update on epidemic information, and drastic measures such as temporary community lock-down may be necessary. Lastly, contact with wild animals should be regulated to prevent future SARS-like outbreaks.

#### ACKNOWLEDGEMENTS

This work was supported by the grants from the National Natural Science Foundation of China (Nos. 31771582 and 31271445), the Guangdong Natural Science Foundation of China (No. 2017A030313131), the "Thousand, Hundred, and Ten" project of the Department of Education of Guangdong Province of China, the Basic and Applied Research Major Projects of Guangdong Province of China (2017KZDXM035 and 2018KZDXM036), the "Yang Fan" Project of Guangdong Province of China (Andy T. Y. Lau-2016; Yan-Ming Xu-2015), and the "Young Innovative Talents" Project of Guangdong Province of China (2019KQNCX034). We would like to thank members of the Lau And Xu laboratory for critical reading of this manuscript.

#### CONFLICT OF INTEREST

The authors have no conflict of interest to disclose.

#### ORCID

Heng Wee Tan <sup>(1)</sup> https://orcid.org/0000-0001-5795-2449 Yan-Ming Xu <sup>(1)</sup> https://orcid.org/0000-0003-1124-0045 Andy T. Y. Lau <sup>(1)</sup> https://orcid.org/0000-0002-7146-7789

#### REFERENCES

1. Li G, Fan Y, Lai Y, et al. Coronavirus infections and immune responses. J Med Virol. 2020;92:424-432.

- Li F. Structure, function, and evolution of coronavirus spike proteins. Annu Rev Virol. 2016;3:237-261.
- Cui J, Li F, Shi ZL. Origin and evolution of pathogenic coronaviruses. Nat Rev Microbiol. 2019;17:181-192.
- Chen Y, Liu Q, Guo D. Emerging coronaviruses: genome structure, replication, and pathogenesis. J Med Virol. 2020;92:418-423.
- Woo PC, Lau SK, Lam CS, et al. Discovery of seven novel mammalian and avian coronaviruses in the genus *deltacoronavirus* supports bat coronaviruses as the gene source of *alphacoronavirus* and *betacoronavirus* and avian coronaviruses as the gene source of *gammacoronavirus* and *deltacoronavirus*. J Virol. 2012;86:3995-4008.
- 6. Abdul-Rasool S, Fielding BC. Understanding human coronavirus HCoV-NL63. *The Open Virol J.* 2010;4:76-84.
- 7. Fehr AR, Perlman S. Coronaviruses: an overview of their replication and pathogenesis. *Methods Mol Biol.* 2015;1282:1-23.
- Su S, Wong G, Shi W, et al. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. *Trends Microbiol.* 2016;24: 490-502.
- Chan JF, Lau SK, To KK, et al. Middle East respiratory syndrome coronavirus: another zoonotic betacoronavirus causing SARS-like disease. *Clin Microbiol Rev.* 2015;28:465-522.
- Sutton TC, Subbarao K. Development of animal models against emerging coronaviruses: from SARS to MERS coronavirus. *Virology*. 2015;479-480:247-258.
- 11. Zhou P, Fan H, Lan T, et al. Fatal swine acute diarrhoea syndrome caused by an HKU2-related coronavirus of bat origin. *Nature*. 2018; 556:255-258.
- Ge XY, Li JL, Yang XL, et al. Isolation and characterization of a bat SARS-like coronavirus that uses the ACE2 receptor. *Nature*. 2013; 503:535-538.
- 13. Wu F, Zhao S, Yu B, et al. A new coronavirus associated with human respiratory disease in China. *Nature*. 2020;579:265-269.
- Zhou P, Yang XL, Wang XG, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579: 270-273.
- Peck KM, Burch CL, Heise MT, Baric RS. Coronavirus host range expansion and Middle East respiratory syndrome coronavirus emergence: biochemical mechanisms and evolutionary perspectives. *Annu Rev Virol.* 2015;2:95-117.
- Li W, Moore MJ, Vasilieva N, et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature*. 2003; 426:450-454.
- Li W, Sui J, Huang IC, et al. The S proteins of human coronavirus NL63 and severe acute respiratory syndrome coronavirus bind overlapping regions of ACE2. Virology. 2007;367:367-374.
- He L, Ding Y, Zhang Q, et al. Expression of elevated levels of proinflammatory cytokines in SARS-CoV-infected ACE2<sup>+</sup> cells in SARS patients: relation to the acute lung injury and pathogenesis of SARS. *J Pathol.* 2006;210:288-297.
- Xu X, Chen P, Wang J, et al. Evolution of the novel coronavirus from the ongoing Wuhan outbreak and modeling of its spike protein for risk of human transmission. *Sci China Life Sci*. 2020;63:457-460.
- Donoghue M, Hsieh F, Baronas E, et al. A novel angiotensinconverting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9. *Circ Res.* 2000;87:1-9.
- 21. Tipnis SR, Hooper NM, Hyde R, Karran E, Christie G, Turner AJ. A human homolog of angiotensin-converting enzyme. *J Biol Chem.* 2000;275:33238-33243.
- Parvathy S, Oppong SY, Karran EH, et al. Angiotensin-converting enzyme secretase is inhibited by zinc metalloprotease inhibitors and requires its substrate to be inserted in a lipid bilayer. *Biochem J*. 1997;327:37-43.
- 23. Rivière G, Michaud A, Breton C, et al. Angiotensin-converting enzyme 2 (ACE2) and ACE activities display tissue-specific

sensitivity to undernutrition-programmed hypertension in the adult rat. *Hypertension*. 2005;46:1169-1174.

- Turner AJ, Tipnis SR, Guy JL, Rice GI, Hooper NM. ACEH/ACE2 is a novel mammalian metallocarboxypeptidase and a homologue of angiotensin-converting enzyme insensitive to ACE inhibitors. *Can J Physiol Pharmacol.* 2002;80:346-353.
- Wakahara S, Konoshita T, Mizuno S, et al. Synergistic expression of angiotensin-converting enzyme (ACE) and ACE2 in human renal tissue and confounding effects of hypertension on the ACE to ACE2 ratio. *Endocrinology*. 2007;148:2453-2457.
- Gorelik G, Carbini LA, Scicli AG. Angiotensin 1-7 induces bradykininmediated relaxation in porcine coronary artery. J Pharmacol Exp Ther. 1998;286:403-410.
- Pontén F, Jirström K, Uhlen M. The human protein atlas—a tool for pathology. J Pathol. 2008;216:387-393.
- Boehm M, Nabel EG. Angiotensin-converting enzyme 2–a new cardiac regulator. N Engl J Med. 2002;347:1795-1797.
- Lew RA, Warner FJ, Hanchapola I, et al. Angiotensin-converting enzyme 2 catalytic activity in human plasma is masked by an endogenous inhibitor. *Exp Physiol*. 2008;93:685-693.
- Lew RA, Warner FJ, Hanchapola I, et al. Characterization of angiotensin converting enzyme-2 (ACE2) in human urine. *Int J Pept Res Ther.* 2006;12:283-289.
- Kobori H, Nangaku M, Navar LG, Nishiyama A. The intrarenal reninangiotensin system: from physiology to the pathobiology of hypertension and kidney disease. *Pharmacol Rev.* 2007;59:251-287.
- Wong DW, Oudit GY, Reich H, et al. Loss of angiotensin-converting enzyme-2 (*Ace2*) accelerates diabetic kidney injury. *Am J Pathol*. 2007;171:438-451.
- Yang P, Gu H, Zhao Z, et al. Angiotensin-converting enzyme 2 (ACE2) mediates influenza H7N9 virus-induced acute lung injury. *Sci Rep.* 2014;4:7027.
- Yan R, Zhang Y, Li Y, et al. Structural basis for the recognition of the SARS-CoV-2 by full-length human ACE2. *Science*. 2020;367: 1444-1448.
- Beniac DR, Andonov A, Grudeski E, Booth TF. Architecture of the SARS coronavirus prefusion spike. *Nat Struct Mol Biol.* 2006;13: 751-752.
- Kirchdoerfer RN, Cottrell CA, Wang N, et al. Pre-fusion structure of a human coronavirus spike protein. *Nature*. 2016;531:118-121.
- Walls AC, Tortorici MA, Bosch BJ, et al. Cryo-electron microscopy structure of a coronavirus spike glycoprotein trimer. *Nature*. 2016; 531:114-117.
- Hofmann H, Simmons G, Rennekamp AJ, et al. Highly conserved regions within the spike proteins of human coronaviruses 229E and NL63 determine recognition of their respective cellular receptors. *J Virol.* 2006;80:8639-8652.
- Peng G, Sun D, Rajashankar KR, Qian Z, Holmes KV, Li F. Crystal structure of mouse coronavirus receptor-binding domain complexed with its murine receptor. *Proc Natl Acad Sci U S A*. 2011;108:10696-10701.
- 40. Peng G, Xu L, Lin YL, et al. Crystal structure of bovine coronavirus spike protein lectin domain. *J Biol Chem*. 2012;287:41931-41938.
- Promkuntod N, van Eijndhoven RE, de Vrieze G, et al. Mapping of the receptor-binding domain and amino acids critical for attachment in the spike protein of avian coronavirus infectious bronchitis virus. *Virology*. 2014;448:26-32.
- Liu C, Tang J, Ma Y, et al. Receptor usage and cell entry of porcine epidemic diarrhea coronavirus. J Virol. 2015;89:6121-6125.
- Yeager CL, Ashmun RA, Williams RK, et al. Human aminopeptidase N is a receptor for human coronavirus 229E. Nature. 1992;357:420-422.
- Wong SK, Li W, Moore MJ, Choe H, Farzan M. A 193-amino acid fragment of the SARS coronavirus S protein efficiently binds angiotensinconverting enzyme 2. *J Biol Chem.* 2004;279:3197-3201.

# 10 of 12 WILEY-

- 45. Du L, Zhao G, Kou Z, et al. Identification of a receptor-binding domain in the S protein of the novel human coronavirus Middle East respiratory syndrome coronavirus as an essential target for vaccine development. J Virol. 2013;87:11963-11963.
- Raj VS, Mou H, Smits SL, et al. Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC. *Nature*. 2013; 495:251-254.
- Yuan Y, Cao D, Zhang Y, et al. Cryo-EM structures of MERS-CoV and SARS-CoV spike glycoproteins reveal the dynamic receptor binding domains. *Nat Commun.* 2017;8:15092.
- Zeng Q, Langereis MA, van Vliet AL, et al. Structure of coronavirus hemagglutinin-esterase offers insight into corona and influenza virus evolution. *Proc Natl Acad Sci U S A*. 2008;105:9065-9069.
- 49. Woo PCY, Huang Y, Lau SKP, Yuen KY. Coronavirus genomics and bioinformatics analysis. *Viruses*. 2010;2:1804-1820.
- Li F, Li W, Farzan M, Harrison SC. Structure of SARS coronavirus spike receptor-binding domain complexed with receptor. *Science*. 2005;309:1864-1868.
- Li W, Wong SK, Li F, et al. Animal origins of the severe acute respiratory syndrome coronavirus: insight from ACE2-S-protein interactions. J Virol. 2006;80:4211-4219.
- Forni D, Cagliani R, Clerici M, Sironi M. Molecular evolution of human coronavirus genomes. *Trends Microbiol*. 2017;25:35-48.
- Hofmann H, Pyrc K, van der Hoek L, Geier M, Berkhout B, Pohlmann S. Human coronavirus NL63 employs the severe acute respiratory syndrome coronavirus receptor for cellular entry. *Proc Natl Acad Sci U S A*. 2005;102:7988-7993.
- 54. Andersen KG, Rambaut A, Lipkin WI, Holmes EC, Garry RF. The proximal origin of SARS-CoV-2. *Nat Med.* 2020;26:450-452.
- Wrapp D, Wang N, Corbett KS, et al. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science*. 2020;367: 1260-1263.
- de Wit E, van Doremalen N, Falzarano D, Munster VJ. SARS and MERS: recent insights into emerging coronaviruses. *Nat Rev Microbiol.* 2016;14:523-534.
- Hoffmann M, Kleine-Weber H, Schroeder S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell*. 2020;181:271-280.
- 58. Woolhouse ME, Gowtage-Sequeria S. Host range and emerging and reemerging pathogens. *Emerg Infect Dis.* 2005;11:1842-1847.
- Woolhouse ME, Haydon DT, Antia R. Emerging pathogens: the epidemiology and evolution of species jumps. *Trends Ecol Evol.* 2005; 20:238-244.
- 60. Banerjee A, Kulcsar K, Misra V, et al. Bats and coronaviruses. *Viruses*. 2019;11:1-15.
- Fan Y, Zhao K, Shi Z, et al. Bat coronaviruses in China. Viruses. 2019; 11:1-14.
- Plowright RK, Peel AJ, Streicker DG, et al. Transmission or withinhost dynamics driving pulses of zoonotic viruses in reservoir-host populations. *PLoS Negl Trop Dis*. 2016;10:e0004796.
- Anthony SJ, Johnson CK, Greig DJ, et al. Global patterns in coronavirus diversity. Virus Evol. 2017;3:vex012.
- 64. Tao Y, Shi M, Chommanard C, et al. Surveillance of bat coronaviruses in Kenya identifies relatives of human coronaviruses NL63 and 229E and their recombination history. J Virol. 2017;91: e01953-e01916.
- Lu G, Wang Q, Gao GF. Bat-to-human: spike features determining 'host jump' of coronaviruses SARS-CoV, MERS-CoV, and beyond. *Trends Microbiol.* 2015;23:468-478.
- Corman VM, Eckerle I, Memish ZA, et al. Link of a ubiquitous human coronavirus to dromedary camels. *Proc Natl Acad Sci U S A*. 2016; 113:9864-9869.
- Song HD, Tu CC, Zhang GW, et al. Cross-host evolution of severe acute respiratory syndrome coronavirus in palm civet and human. *Proc Natl Acad Sci U S A*. 2005;102:2430-2435.

- Müller MA, Corman VM, Jores J, et al. MERS coronavirus neutralizing antibodies in camels, Eastern Africa, 1983–1997. *Emerg Infect Dis*. 2014;20:2093-2095.
- Han G. Pangolins harbor SARS-CoV-2-related coronaviruses. Trends Microbiol. 2020. https://doi.org/10.1016/j.tim.2020.04.001.
- Liu P, Jiang J, Wan X, et al. Are pangolins the intermediate host of the 2019 novel coronavirus (2019-nCoV)? *bioRxiv*. 2020. https:// doi.org/10.1101/2020.02.18.954628.
- Zhang T, Wu Q, Zhang Z. Pangolin homology associated with 2019-nCoV. *bioRxiv*. 2020. https://doi.org/10.1101/2020.02.19. 950253.
- Lam TT, Shum MH, Zhu HC, et al. Identifying SARS-CoV-2 related coronaviruses in Malayan pangolins. *Nature*. 2020. https://doi.org/ 10.1038/s41586-020-2169-0.
- Guan Y, Zheng BJ, He YQ, et al. Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. *Science*. 2003;302:276-278.
- Tang X, Wu C, Li X, et al. On the origin and continuing evolution of SARS-CoV-2. National Science Review. 2020; nwaa036.
- Chinese SMEC. Molecular evolution of the SARS coronavirus during the course of the SARS epidemic in China. *Science*. 2004;303:1666-1669.
- Liang G, Chen Q, Xu J, et al. Laboratory diagnosis of four recent sporadic cases of community-acquired SARS, Guangdong Province. *China Emerg Infect Dis.* 2004;10:1774-1781.
- 77. Liu L, Fang Q, Deng F, et al. Natural mutations in the receptor binding domain of spike glycoprotein determine the reactivity of crossneutralization between palm civet coronavirus and severe acute respiratory syndrome coronavirus. J Virol. 2007;81:4694-4700.
- Wu K, Peng G, Wilken M, Geraghty RJ, Li F. Mechanisms of host receptor adaptation by severe acute respiratory syndrome coronavirus. J Biol Chem. 2012;287:8904-8911.
- Lau SK, Woo PC, Li KS, et al. Severe acute respiratory syndrome coronavirus-like virus in Chinese horseshoe bats. *Proc Natl Acad Sci* U S A. 2005;102:14040-14045.
- Li W, Shi Z, Yu M, et al. Bats are natural reservoirs of SARS-like coronaviruses. *Science*. 2005;310:676-679.
- Tang XC, Zhang JX, Zhang SY, et al. Prevalence and genetic diversity of coronaviruses in bats from China. J Virol. 2006;80:7481-7490.
- Drexler JF, Gloza-Rausch F, Glende J, et al. Genomic characterization of severe acute respiratory syndrome-related coronavirus in European bats and classification of coronaviruses based on partial RNA-dependent RNA polymerase gene sequences. J Vol. 2010;84: 11336-11349.
- Ren W, Qu X, Li W, et al. Difference in receptor usage between severe acute respiratory syndrome (SARS) coronavirus and SARSlike coronavirus of bat origin. J Virol. 2008;82:1899-1907.
- Li W, Zhang C, Sui J, et al. Receptor and viral determinants of SARScoronavirus adaptation to human ACE2. EMBO J. 2005;24:1634-1643.
- Menachery VD, Yount BL Jr, Debbink K, et al. A SARS-like cluster of circulating bat coronaviruses shows potential for human emergence. *Nat Med.* 2015;21:1508-1513.
- Zeng LP, Gao YT, Ge XY, et al. Bat severe acute respiratory syndrome-like coronavirus WIV1 encodes an extra accessory protein, ORFX, involved in modulation of the host immune response. *J Virol.* 2016;90:6573-6582.
- Li R, Qiao S, Zhang G. Analysis of angiotensin-converting enzyme 2 (ACE2) from different species sheds some light on cross-species receptor usage of a novel coronavirus 2019-nCoV. J Infect. 2020;80: 469-496.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Mol Biol Evol*. 2018;35:1547-1549.
- Eichhorn W, Czerny CP. Enteric coronaviruses in primates. J Vet Med. 2010;35:709-712.

- 90. Mostl K. Coronaviridae, pathogenetic and clinical aspects: an update. Comp Immunol Microbiol Infect Dis. 1990;13:169-180.
- Daly N. Tiger tests positive for coronavirus at Bronx Zoo, first known case in the world, in National Geographic. 2020, National Geographic Society. https://www.nationalgeographic.com/animals/2020/04/ tiger-coronavirus-covid19-positive-test-bronx-zoo/.
- Shi J, Wen Z, Zhong G, et al. Susceptibility of ferrets, cats, dogs, and other domesticated animals to SARS-coronavirus 2. *Science*. 2020. https://doi.org/10.1126/science.abb7015.
- Harmer D, Gilbert M, Borman R, Clark KL. Quantitative mRNA expression profling of ACE 2, a novel homologue of angiotensin converting enzyme. *FEBS Lett*. 2002;532:107-110.
- Hamming I, Timens W, Bulthuis MLC, Lely AT, Navis GJ, van Goor H. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. J Pathol. 2004;203:631-637.
- Gembardt F, Sterner-Kock A, Imboden H, et al. Organ-specific distribution of ACE2 mRNA and correlating peptidase activity in rodents. *Peptides*. 2005;26:1270-1277.
- Uhlén M, Fagerberg L, Hallstrom BM, et al. Tissue-based map of the human proteome. *Science*. 2015;347:1260419.
- Li Z, Wu M, Guo J, et al. Caution on kidney dysfunctions of 2019-nCoV patients. *medRxiv*. 2020. https://doi.org/10.1101/2020. 02.08.20021212.
- Zhao Y, Zhao Z, Wang Y, et al. Single-cell RNA expression profiling of ACE2, the putative receptor of Wuhan 2019-nCov. *bioRxiv*. 2020. https://doi.org/10.1101/2020.01.26.919985.
- Cai G. Bulk and single-cell transcriptomics identify tobacco-use disparity in lung gene expression of ACE2, the receptor of 2019-nCov. medRxiv. 2020. https://doi.org/10.1101/2020.02.05.20020107.
- Leung JM, Yang CX, Tam A, et al. ACE-2 expression in the small airway epithelia of smokers and COPD patients: implications for COVID-19. Eur Respir J. 2020;55:2000688.
- Wang P, Cheng Y. Increasing host cellular receptor—angiotensinconverting enzyme 2 (ACE2) expression by coronavirus may facilitate 2019-nCoV infection. *bioRxiv*. 2020. https://doi.org/10.1101/ 2020.02.24.963348.
- Jia X, Yin C, Lu S, et al. Two things about COVID-19 might need attention. *Preprints*. 2020. https://doi.org/10.20944/preprints202 002.0315.v1.
- Roca-Ho H, Riera M, Palau V, et al. Characterization of ACE and ACE2 expression within different organs of the NOD mouse. Int J Mol Sci. 2017;18:1-13.
- Gao J, Aksoy BA, Dogrusoz U, et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal*. 2013;6:pl1.
- 105. Liang W, Guan W, Chen R, et al. Cancer patients in SARS-CoV-2 infection: a nationwide analysis in China. *Lancet Oncol.* 2020;21: 335-337.
- 106. Tan HW, Liang ZL, Yao Y, et al. Lasting DNA damage and aberrant DNA repair gene expression profile are associated with post-chronic cadmium exposure in human bronchial epithelial cells. *Cells*. 2019; 8:842.
- 107. Ding Y, He L, Zhang Q, et al. Organ distribution of severe acute respiratory syndrome (SARS) associated coronavirus (SARS-CoV) in SARS patients: implications for pathogenesis and virus transmission pathways. J Pathol. 2004;203:622-630.
- Xia H, Lazartigues E. Angiotensin-converting enzyme 2 in the brain: properties and future directions. J Neurochem. 2008;107:1482-1494.
- Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020;395: 497-506.
- Ciaglia E, Vecchione C, Puca AA. COVID-19 infection and circulating ACE2 levels: protective role in women and children. *Front Pediatr*. 2020;8:1-3.

- 111. Naicker S, Yang C, Hwang S, et al. The novel coronavirus 2019 epidemic and kidneys. *Kidney Int*. 2020;97:824-828.
- Fan C, Li K, Ding Y, et al. ACE2 expression in kidney and testis may cause kidney and testis damage after 2019-nCoV infection. *medRxiv*. 2020. https://doi.org/10.1101/2020.02.12.20022418.
- 113. Cheng Y, Luo R, Wang K, et al. Kidney disease is associated with inhospital death of patients with COVID-19. *Kidney Int.* 2020;97: 829-838.
- 114. Chai X, Hu L, Zhang Y, et al. Specific ACE2 expression in cholangiocytes may cause liver damage after 2019-nCoV infection. *bioRxiv*. 2020. https://doi.org/10.1101/2020.02.03.931766.
- Zhang C, Shi L, Wang FS. Liver injury in COVID-19: management and challenges. Lancet Gastroenterol Hepatol. 2020. https://doi.org/ 10.1016/S2468-1253(20)30057-1.
- 116. Wang D, Hu B, Hu C, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. JAMA. 2020;323:1061-1069.
- Zhang W, Du RH, Li B, et al. Molecular and serological investigation of 2019-nCoV infected patients: implication of multiple shedding routes. *Emerg Microbes Infect*. 2020;9:386-389.
- 118. Lauer SA, Grantz KH, Bi Q, et al. The incubation period of coronavirus disease 2019 (COVID-19) from publicly reported confirmed cases: estimation and application. *Ann Intern Med.* 2020;172: 577-582.
- 119. Holshue ML, DeBolt C, Lindquist S, et al. First case of 2019 novel coronavirus in the United States. N Engl J Med. 2020;382:929-936.
- 120. Yeo C, Kaushal S, Yeo D. Enteric involvement of coronaviruses: is faecal-oral transmission of SARS-CoV-2 possible? *Lancet Gastroenterol Hepatol.* 2020;5:335-337.
- 121. Wang Z, Xu X. scRNA-seq profiling of human testes reveals the presence of the ACE2 receptor, a target for SARS-CoV-2 infection in spermatogonia, leydig and sertoli cells. *Cells*. 2020;9:920.
- 122. Li YC, Bai WZ, Hashikawa T. The neuroinvasive potential of SARS-CoV2 may be at least partially responsible for the respiratory failure of COVID-19 patients. *J Med Virol*. 2020;92:552-555.
- 123. Mao L, Wang M, Chen S, et al. Neurological manifestations of hospitalized patients with COVID-19 in Wuhan, China: a retrospective case series study. *medRxiv*. 2020. https://doi.org/10.1101/2020.02. 22.20026500.
- 124. Giacomelli A, Pezzati L, Conti F, et al. Self-reported olfactory and taste disorders in patients with severe acute respiratory coronavirus 2 infection: a cross-sectional study. *Clin Infect Dis.* 2020. https://doi. org/10.1093/cid/ciaa330.
- 125. Xu H, Zhong L, Deng J, et al. High expression of ACE2 receptor of 2019-nCoV on the epithelial cells of oral mucosa. Int J Oral Sci. 2020;12:8.
- 126. Sungnak W, Huang N, Becavin C, et al. SARS-CoV-2 entry factors are highly expressed in nasal epithelial cells together with innate immune genes. *Nat Med.* 2020;26:681–687.
- 127. Costanzo M, De Giglio MAR, Roviello GN. SARS-CoV-2: recent reports on antiviral therapies based on lopinavir/ritonavir, darunavir/umifenovir, hydroxychloroquine, remdesivir, favipiravir and other drugs for the treatment of the new coronavirus. *Curr Med Chem.* 2020. https://doi.org/10. 2174/0929867327666200416131117.
- Luo E, Zhang D, Luo H, et al. Treatment efficacy analysis of traditional Chinese medicine for novel coronavirus pneumonia (COVID-19): an empirical study from Wuhan, Hubei Province, China. *Chin Med.* 2020;15:34.
- 129. Ferner RE, Aronson JK. Remdesivir in covid-19. BMJ. 2020;369: m1610.
- Mora JR, Iwata M, von Andrian UH. Vitamin effects on the immune system: vitamins a and D take centre stage. *Nat Rev Immunol.* 2008; 8:685-698.
- 131. Gruber-Bzura BM. Vitamin D and influenza–prevention or therapy? Int J Mol Sci. 2018;19:1-25.

- Martineau AR, Jolliffe DA, Hooper RL, et al. Vitamin D supplementation to prevent acute respiratory tract infections: systematic review and meta-analysis of individual participant data. *BMJ*. 2017;356:i6583.
- 133. te Velthuis AJW, van den Worm SHE, Sims AC, Baric RS, Snijder EJ, van Hemert MJ. Zn<sup>2+</sup> inhibits coronavirus and arterivirus RNA polymerase activity *in vitro* and zinc ionophores block the replication of these viruses in cell culture. *PLoS Pathog*. 2010;6:e1001176.
- Tan HW, Mo HY, Lau ATY, Xu YM. Selenium species: current status and potentials in cancer prevention and therapy. *Int J Mol Sci.* 2018;20:75.
- Harthill M. Review: micronutrient selenium deficiency influences evolution of some viral infectious diseases. *Biol Trace Elem Res.* 2011;143:1325-1336.
- 136. Guillin OM, Vindry C, Ohlmann T, Chavatte L. Selenium, selenoproteins and viral infection. *Nutrients*. 2019;11:2101.
- Graham RL, Donaldson EF, Baric RS. A decade after SARS: strategies for controlling emerging coronaviruses. *Nat Rev Microbiol*. 2013;11:836-848.
- 138. Fast E, Chen B. Potential T-cell and B-cell epitopes of 2019-nCoV. *bioRxiv*. 2020. https://doi.org/10.1101/2020.02.19.955484.
- 139. Forde GM. Rapid-response vaccines-does DNA offer a solution? Nat Biotechnol. 2005;23:1059-1062.
- Petsch B, Schnee M, Vogel AB, et al. Protective efficacy of *in vitro* synthesized, specific mRNA vaccines against influenza a virus infection. *Nat Biotechnol*. 2012;30:1210-1216.
- Huyghe J, Magdalena S, Vandekerckhove L. Fight fire with fire: gene therapy strategies to cure HIV. Expert Rev Anti Infect Ther. 2017;15:747-758.
- Dogo-Isonagie C, Lee SL, Lohith K, et al. Design and synthesis of small molecule-sulfotyrosine mimetics that inhibit HIV-1 entry. *Bioorg Med Chem.* 2016;24:1718-1728.

- 143. Cao X. COVID-19: immunopathology and its implications for therapy. Nat Rev Immunol. 2020;20:269-270.
- Huentelman MJ, Zubcevic J, Hernandez Prada JA, et al. Structurebased discovery of a novel angiotensin-converting enzyme 2 inhibitor. *Hypertension*. 2004;44:903-906.
- 145. Towler P, Staker B, Prasad SG, et al. ACE2 X-ray structures reveal a large hinge-bending motion important for inhibitor binding and catalysis. *J Biol Chem.* 2004;279:17996-18007.
- 146. Zhou Y, Vedantham P, Lu K, *et al*. Protease inhibitors targeting coronavirus and filovirus entry 2015; 116: 76–84.
- 147. Zhang L, Lin D, Sun X, et al. Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved  $\alpha$ -ketoamide inhibitors. *Science*. 2020;368:409-412.
- 148. Winkler AM, Koepsell SA. The use of convalescent plasma to treat emerging infectious diseases: focus on Ebola virus disease. *Curr Opin Hematol.* 2015;22:521-526.

How to cite this article: Tan HW, Xu Y-M, Lau ATY. Angiotensin-converting enzyme 2: The old door for new severe acute respiratory syndrome coronavirus 2 infection. *Rev Med Virol*. 2020;30:e2122. <u>https://doi.org/10.1002/</u>

rmv.2122