

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26

Control of tongue movements by the Purkinje cells of the cerebellum

Paul Hage, Mohammad Amin Fakharian, Alden M. Shoup, Jay S. Pi, Ehsan Sedaghat-Nejad, Simon P. Orozco, In Kyu Jang, Vivian Looi, Hisham Y. Elseweifi, Nazanin Mohammadrezaei, Alexander N. Vasserman, Toren Arginteanu, and Reza Shadmehr

Laboratory for Computational Motor Control, Dept. of Biomedical Engineering, Johns Hopkins School of Medicine, Baltimore, Maryland 21205, USA

Correspondence: Reza Shadmehr, Johns Hopkins School of Medicine, 410 Traylor Building, 720 Rutland Ave., Baltimore, MD 21205. Email: shadmehr@jhu.edu

Acknowledgements: The work was supported by grants from the National Science Foundation (CNS-1714623), and the NIH (R01-EB028156, R37-NS128416).

Author contributions: PH and RS conceived experiments. PH, MAF, AMS, JSP, ESN, and SPO collected the data. PH, MAF, IKJ, VL, HYE, NM, ANV, TA, and RS analyzed the data. PH and HYE made the figures and performed statistical analysis. RS wrote the manuscript.

Competing interests: None.

Data availability: <https://osf.io/WDXU4/>

ORCID ID: Paul Hage 0000-0002-7006-9877, In Kyu Jang 0000-0002-1832-3743, Vivian Looi 0000-0002-9392-4178, Mohammad Amin Fakharian 0000-0002-7741-8689, Simon Orozco 0000-0001-6092-7228, Jay Pi 0000-0003-4036-2877, Ehsan Sedaghat-Nejad 0000-0001-6732-1296, Hisham Y. Elseweifi 0009-0005-8008-5829, Nazanin Mohammadrezaei 0009-0007-0549-9946, Alexander N. Vasserman 0009-0002-9794-1984, Toren Arginteanu 0000-0002-6029-7323, Reza Shadmehr 0000-0002-7686-2569.

27 **Abstract**

28 We use our tongue much like our hands: to interact with objects and transport them. For example, we
29 use our hands to sense properties of objects and transport them in the nearby space, and we use our
30 tongue to sense properties of food morsels and transport them through the oral cavity. But what does
31 the cerebellum contribute to control of tongue movements? Here, we trained head-fixed marmosets to
32 make skillful tongue movements to harvest food from small tubes that were placed at sharp angles to
33 their mouth. We identified the lingual regions of the cerebellar vermis and then measured the
34 contribution of each Purkinje cell (P-cell) to control of the tongue by relying on the brief but complete
35 suppression that they experienced following an input from the inferior olive. When a P-cell was
36 suppressed during protraction, the tongue's trajectory became hypermetric, and when the suppression
37 took place during retraction, the tongue's return to the mouth was slowed. Both effects were amplified
38 when two P-cells were simultaneously suppressed. Therefore, suppression of P-cells in the lingual
39 vermis disrupted the forces that would normally decelerate the tongue as it approached the target.
40 Notably, the population simple spike activity peaked near deceleration onset when the movement
41 required precision (aiming for a tube), but not when the movement was for the purpose of grooming.
42 Thus, the P-cells appeared to signal when to stop protrusion as the tongue approached its target.

43 Introduction

44 We use our tongue to shape the air and generate sounds in order to communicate, and we use our
45 tongue to evaluate food morsels and transport them through the oral cavity in order to eat. These
46 skillful acts involve coordination of over 100 muscles (1), producing movements that are fundamental to
47 our existence. Damage to the cerebellum disrupts these movements, resulting in abnormal muscle
48 activation patterns (2) that bear a resemblance to ataxia of the arm (3). However, life without a
49 cerebellum in humans (4), or inactivation of the deep cerebellar nuclei in mice (5), do not eliminate
50 tongue movements. Rather, the movements become inaccurate. For example, if the activities of
51 Purkinje cells (P-cell) are disrupted via silencing of molecular layer interneurons, the tongue's trajectory
52 becomes erratic and the subject is no longer able to efficiently harvest liquid rewards (6). Thus, the role
53 of the cerebellum in control of the tongue may be similar to its function during control of the limbs (7)
54 and the eyes (8, 9): stopping the movement on target. But how might the cerebellum achieve this?

55 In primates, stimulation of the fastigial nucleus moves the tongue predominantly in the ventral-
56 dorsal axis, while stimulation of the dentate nucleus moves it mainly in the medial-lateral axis (10).
57 Notably, tongue muscles are most readily activated via stimulation of the fastigial nucleus (as compared
58 to the other cerebellar nuclei) (11), suggesting that the P-cells in the vermis play a prominent role in
59 control of the tongue. Unfortunately, there are no reports of P-cell activity in the vermis during targeted
60 tongue movements in any species, but more is known regarding activity of P-cells in Crus I and Crus II (in
61 rodents). For example, as a licking bout is about to start, many P-cells in Crus I and Crus II increase their
62 simple spikes (SS), while a smaller number exhibit a decrease (12). Once the licking begins, the SS rates
63 as a population are phase-locked to the rhythm of the lick, with peaks occurring near lick onset (5).
64 Complex spikes (CSs) also exhibit their highest rates during protraction (12, 13). However, it has been
65 difficult to understand the relationship between the activities of P-cells and control of the tongue.

66 To answer this question, we sought an animal model that had a long tongue and could skillfully
67 direct it to small targets. Marmosets are an attractive choice because they have a 21mm tongue which
68 they use to burrow into small holes and retrieve insects and sap (14). Indeed, they have an extraordinary
69 ability to control their tongue, vocalizing in order to label other marmosets during 2-way communication
70 (15).

71 As we trained head-fixed marmosets to make saccades to visual targets and then rewarded
72 them with food (16), we noticed that they could naturally bend and twist their tongue in order to insert
73 it into small tubes, even when the tubes were placed at 90° with respect to their mouth (17). Because
74 their harvest was difficult, they chose to do many saccade trials, allowing the food to accumulate, then
75 stopped working and claimed their cache by scooping the food out of the tube (17).

76 To quantify how the cerebellum was contributing to the control of the tongue, we recorded
77 from tongue modulated P-cells in lobule VI and VII of the vermis. Then, we relied on the fact that the
78 inferior olive not only transmitted unexpected sensory events to the cerebellum (8, 18–20), it also acted
79 as a stochastic perturbation that completely suppressed the P-cells (21, 22), which then resulted in a
80 small movement (23), or a disruption of the ongoing movement (24–26). Using spike-triggered averaging
81 on the climbing fiber input, we found that the resulting SS suppression altered the deceleration phase as
82 the tongue approached the target, producing hypermetria.

83 This hypermetria was replicated when the P-cells experienced a long period of SS pause without
84 a preceding CS. That is, both a CS-induced SS suppression, and a long SS pause independently had the

85 same effect on behavior: producing downstream forces that extended the tongue. Because as a
86 population, the SS rates were greatest during the deceleration phase of protraction, the results
87 suggested that the P-cells signaled downstream structures to stop the movement as the tongue
88 approached its target. Indeed, this strong engagement of the P-cells was present when the tongue was
89 aiming for a small tube, but not when the movement's purpose was to groom the face.

90

91 **Results**

92 We trained marmosets (n=3) to perform visually guided saccades in exchange for food (Fig. 1A). The
93 subjects performed a sequence of task-relevant saccades, at the end of which we delivered an
94 increment of food (slurry mixture of apple sauce and monkey chow). This food was presented via either
95 the left or the right tube for 50-300 consecutive trials, and then switched tubes. Because the food
96 amounts were small (0.015-0.02 mL), and the tubes were located at $\pm 90^\circ$ with respect to the mouth, the
97 harvest was effortful (17), requiring skillful movements toward a target that was just large enough to
98 accommodate the tongue (4.4 mm diameter tube). As a result, the subjects chose to work for a few
99 consecutive saccade trials (n=6.1 \pm 0.02 successful trials per work period), allowing the food to
100 accumulate, then stopped making saccades to targets, fixated the tube and harvest via a bout of licking
101 (n=22.03 \pm 0.04 licks per harvest period, Fig. 1B).

102 We tracked the motion of the tongue in the horizontal plane using DeepLabCut (27). The tongue
103 movements were of two general types: in the task relevant licks the subjects aimed for the tube ([video 1](#)),
104 whereas in the task-irrelevant licks the subjects groomed their mouth (Fig. 1A) ([video 2](#), [video 3](#),
105 [video 4](#)). During the 2-3 hour recording sessions the subjects performed n=4401 \pm 11 task relevant licks
106 (i.e., aimed at a tube, mean \pm SEM), and n=1310 \pm 4 task irrelevant licks (Fig. 1E).

107 Typically, the subjects began their harvest by licking the food near the tip of the tube ([video 5](#)),
108 but then as the food cache declined, they inserted their tongue into the tube ([video 6](#), [video 7](#)),
109 scooping out their reward. Thus, we divided the task relevant licks into two subtypes, those that aimed
110 for the edge of the tube and harvested the food that was near the tip (Fig. 1A, labeled 2&4), and those
111 that penetrated the tube and harvested the food that was deeper (Fig. 1A, labeled 1&5). Lick protraction
112 velocity was largest for inner tube licks, which also had the largest amplitude and longest protraction
113 duration (Supplementary Fig. S1). Duration of the protraction phase of the inner tube licks was longer
114 than the duration of retraction. For example, in subject 132F, inner tube licks had a protraction duration
115 of 201.3 \pm 0.16 ms vs. retraction duration of 133.6 \pm 0.12 ms (Supplementary Fig. S1). This is consistent
116 with the idea that in contrast to retraction, protraction required aiming which tended to accompany
117 longer duration movements.

118

119 *Climbing fibers were most active near lick onset*

120 We combined MRI and CT image guided procedures (16) to place heptodes and silicon probes in lobules
121 VI and VII of the vermis. Over the course of 3.5 years, we recorded from n=284 P-cells (Figs. 1C & 1E)
122 while the subjects performed 840,787 licks. A neuron was identified as a definitive P-cell (n=230)
123 because of the presence of complex spikes (CS). In addition, we included data from putative P-cells
124 (n=54) for which we could not isolate the CSs but the neuron was located in the P-cell layer and
125 exhibited 0 ms synchronous simple spike (SS) interactions with other confirmed P-cells (8, 9, 26, 28)
126 (Supplementary Fig. S2A).

127 Among our P-cells, the SS modulations were usually present for both tongue movements and
128 eye movements (Supplementary Fig. S2C). However, as our aim was to record from the P-cells that were
129 tongue modulated, in our population the P-cells were more strongly modulated by tongue movements
130 (paired t-test, $t(156)=7.96$, $p=3.3E-13$). This preferential encoding of tongue vs. eye was greater for
131 neurons that were located in lobule VI (Supplementary Fig. S2B).

132 Fig. 1D illustrates the activities of two P-cells near bout onset, as well as during licking. As the
133 bout began, one P-cell increased its SS activity, earlier when the tongue targeted the ipsilateral tube
134 (with respect to the site of recording), while another P-cell decreased its SS activity, earlier for the
135 contralateral tube. As the licking continued, the SS rates in both P-cells were modulated in a rhythmic
136 pattern. We focused on three periods during each lick: protraction acceleration period (Fig. 1D, a-b),
137 protraction deceleration period (b-c), and retraction acceleration period (c-d).

138 As a population, the $n=230$ confirmed P-cells exhibited CS rates that increased near the onset of
139 protraction (Figs. 2A), peaking around the time the tongue touched the tube, and then decreased below
140 baseline around retraction onset. The increased rates during protraction were larger for ipsilateral licks
141 (within cell difference, protraction period, $\text{mean}\pm\text{SEM}$: 0.077 ± 0.024 , $t(229)=26.7$, $p=1.9E-72$). Thus, as a
142 population, for both target directions, the phase of movement for which the CS rates were maximum
143 was protraction (termed CS-on phase).

144

145 *P-cell suppression produced overshooting during protraction and slowed return during retraction*

146 The climbing fiber input suppressed SS production (Fig. 2B, left: all P-cells, right: single P-cell), lasting an
147 average of 14.8 ± 0.44 ms during licking (time to 85% recovery of SS rate, $\text{mean}\pm\text{SEM}$). However, the CS
148 events were rare: a CS occurred in only $5.45\pm 0.01\%$ of the licks during protraction acceleration period
149 ($\text{mean}\pm\text{SEM}$), $9.28\pm 0.01\%$ of the licks during protraction deceleration period, and $6.59\pm 0.009\%$ of the
150 licks during retraction acceleration period (Fig. 2C). We collected a large number of licks per neuron
151 (4401 ± 11 licks, Fig. 1E), then performed CS-triggered averaging to ask whether the resulting SS
152 suppression affected the motion of the tongue.

153 For each P-cell we considered triplets of consecutive licks $\{n - 1, n, n + 1\}$ in which all three
154 licks were of the same type, i.e., contacted the same part of the tube (edge or inner). We then selected
155 those triplets in which there was a CS at only a single period in lick n , but no CS during any period in the
156 two neighboring licks $n - 1$, and $n + 1$. For example, consider licks in which there was a CS in one P-cell
157 during the acceleration period of protrusion (Supplementary Fig. S3). This acceleration period was brief
158 (49.9 ± 0.019 ms), during which the SS activity was normally rising and nearly identical in licks $n - 1$ and
159 $n + 1$ but suppressed during lick n (Supplementary Fig. S3, 1st row). We measured the change in SS
160 activity by comparing lick n with licks $n - 1$ and $n + 1$ and plotted the results of each comparison in Fig.
161 S3B, green & blue solid lines (labeled suppressed, top row, right column). As a control, we performed a
162 bootstrapping procedure in which we generated a pseudo data set for each P-cell by randomly assigning
163 the CS label to a lick and comparing it with its two temporally adjacent neighbors.

164 To ask whether there were any effects of the brief SS suppression on the tongue, we measured
165 the distance of the tongue tip to the mouth and also its angle with respect to the midline, and then
166 compared the trajectories in lick n with the neighboring licks in which the P-cell was not suppressed. The
167 effects appeared consistent (Supplementary Fig. S3A, red colors): following SS suppression, there was
168 little or no change in tongue kinematics. Statistical testing, which relied on bootstrapping procedure to

169 compute 95% confidence intervals (CI), demonstrated that the changes were within the error bounds.
170 Thus, the suppression during the acceleration period of protraction had no significant effects on tongue
171 trajectory (Supplementary Fig. S3C, measured via distance to the mouth, and its angle, at peak
172 protraction speed and peak displacement).

173 As the protraction continued, the effects of SS suppression became evident. If the suppression
174 occurred during the deceleration period of protraction (duration: 103.6 ± 0.04 ms), the tongue exhibited
175 hypermetria (Fig. 3A, second row), producing increased displacement and increased angle of the
176 tongue's trajectory (Figs. 3B, displacement: 0.37 ± 0.002 mm, 95%CI = $[-0.17 \ 0.14]$ mm, angle: 3.35 ± 0.02
177 deg, 95%CI = $[-1.14 \ 0.98]$ deg). Notably, the effects were consistent regardless of whether lick n was
178 compared to the previous lick ($n - 1$), or the subsequent lick ($n + 1$) (Fig. 3B, second row, blue and
179 green traces). Furthermore, the effects were consistent across the P-cells (Fig. 3A, red colors, also
180 Supplementary Fig. S4, left panel), and larger for contralateral licks (within cell difference, ipsilateral
181 minus contralateral, at lick endpoint, displacement mean \pm SEM: -0.15 ± 0.04 mm, $t(229) = -3.97$, $p = 9.6E-05$,
182 angle: -1.41 ± 0.37 , $t(229) = -3.79$, $p = 9.69E-05$).

183 These results hinted that SS suppression prevented normal deceleration, producing hypermetria
184 and a bending of the tongue away from the midline. If this interpretation is valid, then a similar
185 suppression during the retraction period should produce forces that are again in the direction of
186 protraction, now resisting the tongue's return. That is, if SS suppression during protraction sped the
187 movement outward, then the same suppression during retraction should now slow the movement. In
188 both cases, the suppression should bend the tongue away from the midline.

189 As retraction began, the population CS activity (Fig. 2A) had fallen below baseline, i.e., opposite
190 of the activity during protraction. Yet, if the CS occurred during the retraction acceleration period
191 (duration: 78.3 ± 0.02 ms), the resulting suppression was again an outward displacement of the tongue
192 and bending (Fig. 3D). A comparison of lick n with $n - 1$ or $n + 1$ revealed a consistent effect: an
193 increased distance of the tongue to the mouth and an increased angle (Fig. 3F, displacement:
194 1.29 ± 0.002 mm, 95%CI = $[-0.32 \ 0.22]$ mm, angle: 4.70 ± 0.02 deg, 95%CI = $[-1.15 \ 0.85]$ deg). These effects
195 were present for ipsilateral and contralateral licks (Fig. 3E), larger for the contralateral licks (within cell
196 difference, ipsilateral minus contralateral, at lick endpoint, displacement: -0.37 ± 0.09 mm, $t(229) = -3.97$,
197 $p = 4.8E-05$, angle: -2.06 ± 0.50 , $t(229) = -4.1$, $p = 2.71E-05$), and consistent across the P-cells (Fig. 3D, red
198 colors). Thus, the CS-induced SS suppression during retraction slowed the return of the tongue and
199 producing bending away from the midline.

200 In summary, when the climbing fiber input briefly suppressed the P-cells during the deceleration
201 period of the tongue's protrusion, the tongue overextended and bent away from the midline. When this
202 suppression occurred during retraction, the tongue's return was slowed and again bent away from the
203 midline. These effects were present for targets on both the ipsilateral and contralateral sides, but
204 greater when the target was contralateral. Thus, it appeared that SS suppression disrupted the motor
205 commands that would normally stop the tongue during protraction and return it during retraction. That
206 is, the downstream effect of suppression of P-cells was to produce forces that extended the tongue and
207 produced lateral bending.

208

209 *Hypermetria increased when pairs of P-cells were simultaneously suppressed*

210 Our dataset included $n=298$ pairs of simultaneously recorded P-cells. This allowed us to test whether
211 near simultaneous suppression of two P-cells had a greater effect on tongue kinematics as compared to
212 when only one of the two P-cells was suppressed.

213 As before, we collected triplets of consecutive licks $\{n - 1, n, n + 1\}$ where all the licks were of
214 the same type (directed toward the same part of the tube), but only lick n had a CS. We divided the
215 triplets based on whether a CS was present in only one of the P-cells, or both P-cells, then computed
216 trajectory differences between licks n and $n - 1$, as well as licks n and $n + 1$. Finally, for each pair of P-
217 cells we averaged $n - (n - 1)$ and $n - (n + 1)$ to increase statistical power (as there were far fewer
218 licks in which both P-cells experienced a CS during the same period of the movement).

219 We found that if two P-cells were suppressed during the deceleration phase of protraction, then
220 there was significantly greater displacement of the tongue, and bending, as compared to when only one
221 of the P-cells was suppressed (Fig. 4A, angle: $t(248) = -3.7167$, $p = 2.5E-04$, displacement: $t(248) = -$
222 5.5471 , $p = 7.43E-08$). Similarly, when the suppression occurred during retraction, the return phase of
223 the movement experienced a greater slowing in the case of two P-cells as compared to a single P-cell
224 (Fig. 4B, angle: $t(192) = 6.34$, $p = 1.6E-09$, displacement: $t(192) = 6.50$, $p = 6.79E-10$). Thus, near
225 simultaneous suppression of two P-cells roughly doubled the kinematic effects.

226

227 *Control studies*

228 The fact that a CS was present during a given period in lick n may have been because earlier in the
229 tongue's trajectory there was an event (for example, an error), that affected that movement, increasing
230 the likelihood of a CS, and resulting in compensatory movements that followed the CS. To check for this,
231 for each period during which we observed a CS we considered the tongue's trajectory in the same lick
232 but during the period preceding the CS. For example, for the licks in which there was a CS in the
233 protraction deceleration period we focused on the acceleration period of the same movement. By
234 comparing the lick in which the CS had occurred with its two neighbors, we found that in the period
235 before the CS had occurred tongue kinematics remained within the 95% confidence intervals of chance:
236 distance to mouth and angle of the tongue at peak speed were not different than chance (Fig. 3C, period
237 before peak speed, displacement: -0.11 ± 0.001 mm, 95%CI = $[-0.12 \ 0.09]$ mm, angle: 0.19 ± 0.01 deg,
238 95%CI = $[-0.73 \ 0.53]$ deg).

239 Next, we checked the licks in which there was a CS in the retraction period. We found that
240 before the CS had occurred, at the onset of retraction the distance to the mouth and tongue angle were
241 not different than if the CS had not occurred (Fig. 3F, protraction period before peak displacement,
242 displacement: 0.01 ± 0.002 mm, 95%CI = $[-0.16 \ 0.10]$ mm, angle: 0.68 ± 0.02 deg, 95%CI = $[-1.14 \ 0.83]$
243 deg). Moreover, during the preceding protraction in the same lick, at peak tongue speed, the distance
244 and angle were again not different than chance (displacement: -0.66 ± 0.003 mm, 95%CI = $[-0.70 \ 0.74]$
245 mm, angle: -2.16 ± 0.02 deg, 95%CI = $[-3.35 \ 3.58]$ deg). Yet, if during the return phase a CS was present,
246 the movement was slowed (Fig. 3F, before peak-speed period of retraction, displacement: 1.29 ± 0.002
247 mm, 95%CI = $[-0.32 \ 0.22]$ mm, angle: 4.70 ± 0.02 deg, 95%CI = $[-1.16 \ 0.85]$ deg). That is, the tongue's
248 trajectory before the CS was not significantly different than the neighboring licks, but after the CS-
249 triggered SS suppression the trajectories diverged.

250 As a further control, we considered movements in which there was a CS event just before
251 protrusion onset. In these movements, the SS rates were suppressed, but the rates recovered around 10

252 ms after lick onset (Supplementary Fig. S5, 1st row). Thus, despite the presence of a CS just before the
253 lick had started, the SS rates during the entire protrusion and retraction periods were intact. The
254 trajectory of the tongue as measured via distance and angle remained within the 95% confidence
255 interval bounds of chance (Supplementary Fig. S5, 3rd and 4th rows).

256

257 *Effect of SS suppression remained consistent across P-cells regardless of SS modulation*

258 For nearly every P-cell in our dataset, during both protraction and retraction, the CS-triggered SS
259 suppression was followed by an extension of the tongue and a lateral bending (Figs. 3A & 3D). This
260 consistency was surprising in light of the diversity that was present in the SS patterns: before the onset
261 of the bout, some P-cells had increased their SS rates with respect to baseline, while others had
262 decreased (Fig. 1D). Did the effects of SS suppression differ in these two groups?

263 In our data set of $n=142$ of P-cells with SSs, most cells ($n=123$) increased their SS rates before
264 the onset of the bout, but a minority exhibited a decrease ($n=31$) (Figs. 5A & 5B). We separated the P-
265 cells into SS increasers and decreasers and found that despite the differences in their SS patterns, the CS
266 rates increased in both groups near bout onset (Fig. 5C, peak CS firing rate change from baseline, SS
267 increasers: 0.15 ± 0.04 Hz, SS decreasers: 0.26 ± 0.10 Hz). We then compared tongue trajectories in
268 triplets of consecutive licks $\{n - 1, n, n + 1\}$, finding that during protraction, following SS suppression in
269 the deceleration period, in both groups of P-cells the tongue was displaced away from the mouth,
270 exhibiting a greater distance and angle (Fig. 5D, SS increasers, displacement: 0.38 ± 0.005 mm, 95%CI = [-
271 0.02 0.02] mm, angle: 3.72 ± 0.05 deg, 95%CI = [-0.16 0.15] deg, SS decreasers: displacement: 0.45 ± 0.04
272 mm, 95%CI = [0.04 -0.06] mm, angle: 4.28 ± 0.54 deg, 95%CI = [-0.16 0.15] deg). Similarly, during
273 retraction, SS suppression in both groups of P-cells produced a slowing of the tongue's return, resulting
274 in a greater distance and angle (Fig. 5E, SS increasers, displacement: 1.25 ± 0.01 mm, 95%CI = [-0.04 0.03]
275 mm, angle: 4.45 ± 0.05 deg, 95%CI = [-0.16 0.17] deg, SS decreasers: displacement: 1.04 ± 0.04 mm, 95%CI
276 = [-0.04 0.03] mm, angle: 3.48 ± 0.17 deg, 95%CI = [-0.16 0.17] deg).

277 Thus, regardless of the patterns of SS activity during licking among the various P-cells, the
278 downstream effect of CS-triggered SS suppression was extension of the tongue coupled with lateral
279 bending.

280

281 *SS pause without a CS was sufficient to produce hypermetria*

282 We had interpreted the kinematic effects that followed the CS-induced SS suppression as being caused
283 by SS suppression, not due to the presence of a CS. To check the validity of this conjecture, we
284 quantified the kinematic effects of non-CS-induced long SS pauses on the tongue's trajectory. To identify
285 a long pause, for each P-cell we considered all licks towards the same direction in which no CS occurred
286 at any point in the movement. We then identified the longest inter-spike interval (ISI) for the SSs in each
287 phase of each lick (phase refers to protraction deceleration period, etc.). For each phase under study,
288 and each P-cell, we labeled 25% of the licks with the largest ISIs as a "long-pause" lick.

289 Next, we considered triplets of consecutive licks $\{n - 1, n, n + 1\}$ of the same type in the same
290 direction in which none of the licks had a CS during any phase of the movement. We selected the subset
291 of triplets in which lick n had a long pause in only one phase of the movement, but no long pauses in any
292 phase of the neighboring licks. We then compared tongue trajectories between the lick that had a long
293 SS pause with its two neighboring licks.

294 On average, the duration of a long SS pause was 31.25 ± 1.2 ms during protraction deceleration,
295 and 34.25 ± 1.08 ms (mean \pm SEM) during retraction acceleration. If this pause occurred during the
296 protraction acceleration period, it produced hypermetria and bending of the tongue away from the
297 midline (Fig. 6A), and if it occurred during the retraction period it produced a slowing of the return and
298 also a bending away from the midline (Fig. 6B). The trajectory changes were quite similar to what we
299 had observed following a CS-induced SS suppression.

300 Thus, regardless of whether an SS pause was due to the arrival of a CS or not, the downstream
301 effects were the same: production of forces that extended the tongue, bending it away from the
302 midline.

303

304 *The SS rates peaked at deceleration onset, but only if the movement was reward relevant*

305 Across the cells, the CS rates peaked at approximately the time of maximum protraction velocity (Figs.
306 7A, left column), exhibiting a greater rate for movements toward the ipsilateral side (ipsilateral:
307 0.21 ± 0.02 Hz, contralateral: 0.16 ± 0.02 Hz, within cell difference, average CS rate, ipsilateral minus
308 contralateral, $t(229)=28.1$, $p=2.5E-76$). In contrast, the CS rates declined below baseline before the onset
309 of retraction. Thus, the CS-on phase across the P-cells was protraction.

310 To analyze the SS activities as a population, we had to consider the fact that during a bout, as
311 the SS rates modulated about a mean, the mean was not stationary (Fig. 5A). Rather, the mean SS rates
312 rose or fell before bout onset, then drifted back toward the values before start of the bout, reaching
313 pre-bout levels at the bout ended. Thus, to quantify modulation of SS rates, for each P-cell we
314 considered a 2 sec moving window to compute the running average of its firing rate, then computed the
315 SS rates with respect to this mean. The 2 sec window was chosen because it was roughly an order of
316 magnitude larger than the duration of a typical lick.

317 Given that a CS-triggered suppression in SS rates induced downstream forces that pushed the
318 tongue outwards, then if this region of the cerebellum was interested in stopping the ongoing
319 movement, during deceleration of protraction the SS rates should increase, thus commanding forces
320 that would stop the motion of the tongue as it neared the target. Indeed, the SS rates peaked near the
321 onset of lick deceleration, and were larger for contralateral licks (Fig. 7A, right column, ipsilateral:
322 11.58 ± 1.75 Hz, contralateral: 12.91 ± 1.95 Hz, within cell difference: $t(156)=13.6$, $p=2.9E-28$).

323 To test if SS modulation varied with tongue kinematics, we considered two conditions: when
324 licks had the same amplitude but different peak velocities (Fig. 7B), and when they had the same peak
325 velocity but different amplitudes (Fig. 7C). To consider licks of the same amplitude but different velocity,
326 we quantified lick vigor, defined as the peak speed of the protraction with respect to the speed
327 expected for a lick of the same amplitude (17, 29) (Supplementary Fig. S6). High vigor licks exhibited
328 greater SS rates at peak protraction speed (Fig. 7B, within cell difference, high vigor minus low vigor,
329 2.92 ± 0.60 Hz, $t(156)=14.1$, $p=1.2E-29$). That is, licks that required greater deceleration forces
330 accompanied greater SS rates near the onset of deceleration.

331 Next, we considered licks that had the same peak velocity but different amplitudes (Fig. 7C).
332 These two licks began with very similar velocity patterns, reaching on average identical peak velocity,
333 but the SS rates achieved a greater peak rate for the licks that had a longer deceleration period.
334 Moreover, the licks with the longer period of deceleration had a longer period of SS rate increase (Fig.
335 7C, time to baseline crossing, high duration: 126.80 ± 7.68 ms, low duration: 86.02 ± 7.23 ms, paired t-test,

336 $t(156)=7.84$, $p=6.75E-13$). As a result, the SS firing rates at peak velocity tended to increase with the
337 lick's peak amplitude (Fig. 7E).

338 All of these results were for tongue movements in which the subject aimed for the small food
339 tubes. Would the same patterns hold for movements that did not require such precision? To consider
340 this question, we turned to the licks that were not directed toward a food tube, which were often
341 grooming licks. These licks tended to be slower, with a peak speed that was roughly half the speed of
342 the licks directed toward the food tubes (Fig. 7D, peak velocity of food tube licks: 290.5 ± 3.6 mm/s, other
343 licks: 153.8 ± 2.5 mm/s). Remarkably, during protraction of these licks the CS rates were an order of
344 magnitude smaller than when the licks were toward the food tubes (Fig. 7D, paired t-test, tube licks vs.
345 grooming, combined directions, $t(229)=-3.27$, $p=0.00126$). Similarly, the SS rate modulations were much
346 smaller (Fig. 7D, right, paired t-test, tube vs. other, combined directions, $t(156)=8.6$, $p=8.44E-15$). In
347 sharp contrast, during retraction of these task-irrelevant licks, both the CS and the SS rates were
348 modulated below baseline by amounts that were roughly comparable to the rates of the tube directed
349 licks. That is, the fundamental difference in the P-cell activity between the tube directed and other licks
350 was in the protraction phase, the phase in which control of the tongue required precision.

351 In summary, when the licks were directed toward the food tube, the CS and SS rates peaked
352 during protraction and were greater when the movement had greater speed. Because SS suppression
353 produced forces that extended the tongue, the fact that SS activity peaked near deceleration onset was
354 consistent with the view that the downstream effects were to decelerate the tongue as it neared its
355 target. Remarkably, both the CS and SS modulation patterns were largely absent when the tongue
356 movements were not aimed at the food tube.

357

358 *Error in the tongue's trajectory was reported to the cerebellum via complex spikes*

359 Inserting the tongue into the tube required precision because the opening was only slightly larger than
360 the tongue. As a result, in roughly 15% of the licks the food was inside the tube, but the tongue missed
361 the entrance ([video 8](#), [video 9](#), and [video 10](#), also Supplementary Fig. S7). We labeled these as
362 unsuccessful licks because the tongue did not bend enough and instead hit the tube's outer edge. Were
363 these errors reported to the cerebellum?

364 To visualize the CS patterns as a function of the spatial location of the tongue, we needed to
365 compute $\Pr(CS|x)$, i.e., the probability of producing a complex spike given that the tongue was at a
366 given location. To arrive at this variable, we began with computing $p(x|CS)$, i.e., the probability density
367 of the position of the tongue's tip x , given that a CS occurred at time t . To compute this function, we
368 used spike triggered averaging to compute the average position of the tongue during the 50ms period
369 before the CS. We found that this likelihood depended on whether the tongue successfully entered the
370 tube or not. For licks that were successful and entered the tube, the likelihood $p(x_s|CS)$ was bimodal,
371 exhibiting a peak near lick onset, then a second peak near the food (Fig. 8A, left). For licks that were
372 unsuccessful and did not enter the tube, the likelihood $p(x_u|CS)$ was also bimodal, but now the second
373 peak was around the edge where the tongue had collided with the tube (Fig. 8B, left).

374 We next computed the marginal probability density $p(x)$ for the successful and unsuccessful
375 licks (Fig. 8A and 8B, right). This function estimated the probability of the tongue being at a given
376 position during the various licks. We then computed the prior probabilities $\Pr(CS)$, and the ratio of the
377 probabilities $p(x|CS) \Pr(CS) / p(x)$, arriving at the posterior probability $\Pr(CS|x_s)$ and $\Pr(CS|x_u)$. Each

378 posterior estimated the probability of observing a CS as a function of the location of the tongue during
379 successful and unsuccessful licks. Finally, we computed the error-induced spatial pattern of complex
380 spikes by subtracting the posteriors (Fig. 8C). The results revealed a spatial gradient that increased with
381 the tongue's distance from the mouth, suggesting that once we accounted for the CS modulations
382 associated with normal licking, the CS events that remained in the unsuccessful licks tended to occur
383 after the tongue had touched the far end of the tube.

384 To view these error-specific effects in another way, we plotted the CS rates as a function of time
385 with respect to the tube-touch event. For the successful licks, tube touch occurred when the tongue
386 crossed the tube's edge and entered the tube. In this case, the CS rates following tube touch were
387 depressed (Fig. 8D). In contrast, for the unsuccessful licks the tube touch indicated an error, and the CS
388 rates following this event showed a sharp increase (100 ms period following tube touch, CS rate in
389 unsuccessful vs. successful licks, paired t-test, $t(459)=8.09$ $p=5.3 \times 10^{-15}$).

390 Thus, when the food was inside the tube and the tongue successfully entered it, tube touch did
391 not elicit complex spikes. However, when the food was inside the tube and the tongue touched it, but
392 failed to enter it, now the tube touch event produced complex spikes.

393

394 **Discussion**

395 To quantify how the P-cells in the vermis contributed to control of the tongue, we trained marmosets to
396 make skillful movements, extending and bending their tongue to harvest food from small tubes that
397 were placed at 90° with respect to their mouth. Using spike-triggered averaging on the climbing fiber
398 input to each P-cell, we found that if the resulting SS suppression occurred during protraction, there was
399 a disruption in the deceleration phase of the movement, resulting in hypermetria as the tongue
400 approached the target. A suppression that occurred during retraction retarded the tongue's return.
401 When two P-cells were simultaneously suppressed, the kinematic effects magnified. Thus, regardless of
402 whether a P-cell was suppressed during protraction or retraction, the downstream effects were
403 production of forces that pulled the tongue outwards. These results were present for both ipsilateral
404 and contralateral targets, but greater when the target was contralateral.

405 Because during unperturbed movements the SS rates in the population peaked as the
406 movement began decelerating, we infer that the downstream effects were production of forces in the
407 direction of retraction. This suggests that the contributions of the cerebellum to control of tongue
408 movements may be similar to that of the eyes (30, 31) and the limbs (7): steering the movement and
409 stopping it as it nears the target.

410

411 *P-cells were modulated only for reward relevant movements*

412 When the purpose of the movement was to groom the face, during the protraction phase the CS rates
413 remained at baseline while the SS modulations were absent. This observation replicated our
414 observations during saccades: when saccades are aimed toward a reward relevant target, in lobule VII
415 the CS and SS rates are strongly modulated and the P-cell population predicts when the movement
416 should be stopped (26). When similar saccades are made without a reward relevant target, the CS
417 modulations are missing (8) and the SS rates no longer predict deceleration onset (26). Thus, for both
418 saccades and licking, the cerebellum is engaged only when the movement is reward relevant.

419 One reason for engagement of the P-cells during reward relevant movements may be the
420 greater accuracy requirements of those movements. Target position (for saccades) strongly engages the

421 neurons in the superior colliculus, which appear to transmit that information to the cerebellum via
422 mossy fibers (26). When the saccade is reward irrelevant, the encoding of the target location is muted in
423 the colliculus (32–34), as well as the mossy fibers (26). This implies that when that movement is not
424 reward relevant, the cerebellum is poorly informed of the goal of the movement. As a result, it cannot
425 assist in predicting when the movement is nearing the target and should be stopped.

426 Because the colliculus contains a topographic map of tongue movements (35), we conjecture
427 that like saccades, tongue movements that are not reward relevant will produce muted activity in the
428 colliculus.

429

430 *Climbing fibers were most active near movement onset*

431 We were surprised that the CS rates peaked not after the tongue contacted the tube, but around the
432 onset of protraction. This observation reproduced findings of Welsh et al. (13) in rats, who recorded
433 from the lateral cerebellum and found that the CS rates peaked near the onset of tongue protrusion,
434 even when the tongue was deafferented. Indeed, in many types of movements, including walking (36),
435 reaching (37–41), and moving the wrist (42), the CS rates peak near movement onset. For example, in
436 the oculomotor vermis, near saccade onset the olivary input informs the P-cells regarding the direction
437 of the upcoming movement (8). How might the inferior olive be involved in transmitting movement
438 information to the cerebellum?

439 A key observation is that the inferior olive not only receives input from the superior colliculus
440 (43–45), but that subthreshold stimulation of a region of the colliculus leads to CS production without
441 producing a movement (46). Thus, it is possible that the increased CS rates around the onset of
442 protraction reflect activities of regions that initiated the tongue movement, i.e., the motor cortex and
443 the superior colliculus (47). This prediction remains to be tested with simultaneous recordings from the
444 colliculus, motor cortex, and the cerebellum.

445

446 *Climbing fibers reported lick errors*

447 Because the tubes were placed at sharp angles to the mouth, roughly 15% of the licks failed to retrieve
448 the food. In these unsuccessful licks, the tongue did not bend enough and instead collided with the far
449 edge of the tube. The climbing fibers reported this error robustly, exhibiting a strong increase in rates
450 following the touch of the tube. Remarkably, when the touch event was not in error, i.e., the tongue
451 entered the tube, the CS rates were suppressed. Thus, the CS rates signaled a touch that was in error,
452 not a touch that was expected.

453

454 *Using the olivary input to infer a P-cell's contribution to behavior*

455 Complex spikes are rare events, occurring at around once per second. They briefly and completely
456 suppress the SS rates, which induce downstream effects on the cerebellar nuclei (48), potentially
457 producing movements (23). However, the CS rates are modulated to encode the direction of sensory
458 prediction errors (8, 18, 49, 50). Thus, it was critical to test that the kinematic effects that we measured
459 following a CS-induced SS suppression were not a consequence of a feedback response to kinematic
460 deviations that occurred before the CS.

461 We did this by comparing triplets of temporally adjacent licks, finding that while the tongue
462 trajectory preceding the CS event remained within the chance error bounds, the trajectory that followed

463 were robustly different than chance. Notably, the downstream effects of the CS-induced suppression
464 were in the same direction, i.e., extension of the tongue, regardless of whether the CS events occurred
465 during protraction or retraction. This is notable because the CS rates were maximum during protraction,
466 and minimum during retraction, yet their downstream effects were the same: pull the tongue outwards.

467 The idea that the olivary input may affect ongoing movements was noted by Ebner and
468 colleagues during reaching movements (24), and subsequently observed during saccades (25). For
469 example, during saccades following the CS-induced SS suppression, the eyes are pulled in the CS-on
470 direction of the P-cell. This is consistent with the fact that optogenetic increase in the SS rates
471 suppresses the cerebellar nuclei (51), pulling the eyes approximately in direction CS+180 (52). While
472 relying on the stochasticity in the olivary input has the disadvantage of lacking a causal manipulation, it
473 has the advantage of relating the kinematic effects to CS-on properties of each P-cell, something that
474 would not be possible with large scale optogenetic stimulation.

475

476 *A long SS pause had the same effect on behavior as a CS-induced SS suppression*

477 A CS is followed by SS suppression, but pauses in SS production can also occur because of other reasons,
478 including inhibitory input from the molecular layer interneurons (53). Here we found that the effects of
479 CS-induced SS suppression on tongue kinematics were largely the same as SS pauses that were not due
480 to arrival of a CS. In both cases, the result was a force that pulled the tongue outwards and bent it away
481 from the midline. This implies that the downstream effects on kinematics were not due to arrival of an
482 input from the inferior olive, but rather the suppression or pausing of SS production in the parent P-cell.

483

484 *A cortico-cerebellar network for control of the tongue*

485 As the subject prepared to initiate a licking bout, the P-cells exhibited a ramping activity. Previous
486 reports have noted that during this period there is ramping activity in the tongue regions of the motor
487 cortex (54), as well as in the fastigial (54) and the dentate nuclei (55). Inhibiting the motor cortex in mice
488 prevents both the onset and the termination of the licking bout (56), suggesting that both are active
489 processes that are cortically mediated. Inhibiting the fastigial during the ramping period disrupts
490 planning of the movement and removes the direction selectivity that the motor cortical cells exhibit
491 (54), while inhibiting the dentate disrupts the ramping activity in the motor cortex (55). Similarly,
492 exciting the P-cells in the vermis abolishes the ramping activity in motor cortex during the delay period
493 of a decision-making task (57). This implies that as one prepares to initiate a movement, the rising
494 activity in the motor cortex is controlled via a loop through the cerebellum.

495 Our results here suggest that once the tongue movement begins, there is a specific role for the
496 cerebellum in producing forces that would stop the protraction, especially if that movement is reward
497 relevant. We speculate that the cerebellum is informed via the mossy fibers of two kinds of information:
498 the location of the target, and a copy of the ongoing motor commands (26). The function of this region
499 of the cerebellum may be to use the copy of the motor commands to predict when the tongue is about
500 to reach the target and aid in production of commands that would stop the outward movement.

501

502 *Medial and lateral parts of the cerebellum may contribute to different aspects of tongue movements*

503 In humans, the tongue region of the cerebellum extends from lobule VI in the vermis laterally to the
504 hemispheres (58–60). Dysarthria is principally associated with damage in the paravermal regions of the

505 cerebellum (61). In macaques, stimulation of the fastigial nucleus moves the tongue in the protraction-
506 retraction axis, while stimulation of the dentate nucleus moves it in the lateral-medial axis (10). In mice,
507 activation of the P-cells in the lateral regions of lobule VI and VII during protraction bends the tongue
508 toward the ipsilateral side (62). When we consider these results together with our observations here,
509 what emerges is the conjecture that the P-cells in the vermis are important for control of
510 protraction/retraction, but the P-cells in the paravermis and hemispheres have a different role, possibly
511 in controlling how the tongue bends.

512

513 *Toward a general model of how the cerebellum controls movements*

514 Like the SS rates, the CS rates peaked near protraction peak velocity, then fell below baseline before the
515 onset of retraction. Thus, for both ipsilateral and contralateral movements, the “CS-on action” across
516 the P-cells was protraction, while the “CS-off action” was retraction. Notably, the downstream effects of
517 SS suppression were to extend the tongue. As a result, there was a correspondence between the vector
518 that described the CS-on action, and the vector that described the effects of SS suppression. This fact is
519 notable because the same principle holds for P-cells in the oculomotor region of the cerebellum during
520 saccadic eye movements (25, 26): the olivary input to an oculomotor P-cell is most active when a
521 saccade is planned in direction CS-on, and the downstream effects of that P-cell’s SS suppression is to
522 pull the eyes also in direction CS-on. Thus, for both eye movements and tongue movements, the olivary
523 input provides a vector based coordinate system (26) with which one might estimate the downstream
524 contributions of a P-cell to control of that movement (63, 64).

525 The key theoretical idea is that the inferior olive organizes the cerebellum so that the P-cells are
526 placed in competition with each other: for every P-cell that has a particular CS-on, effecting movements
527 along a particular potent vector, there is another that prefers the opposite vector (26). Unfortunately,
528 here we could not apply this theory to organize the P-cells into antagonist populations because nearly all
529 the cells in our database had a CS response that peaked during protraction. However, our theory (26)
530 predicts that there should be P-cells whose climbing fiber inputs prefer retraction. In these P-cells the SS
531 suppression should pull the tongue inward. If these P-cells exist, then their SS pattern would be
532 antagonistic to the SS pattern of the P-cells we found here, resulting in a population response in which
533 P-cells would compete with each other, perhaps producing a sum of activity that is a burst-pause
534 pattern, inhibiting then disinhibiting the nucleus as the tongue approaches the target.

535 Lingual dysfunction accompanies a host of symptoms, including vocal muscle dystonia (65),
536 problems in swallowing (66), and dysarthria (2, 61, 67), all of which share a link to the cerebellum.
537 Rehabilitation or cures for these symptoms will require a much better understanding of how the
538 cerebellum contributes to control and learning of tongue movements. Marmosets are exceptionally
539 skilled in shaping and twisting their tongue, using it almost like a finger. This makes them an attractive
540 new model to study the neural control of a body part that is essential for our existence.

541

542 **Methods**

543 Data were collected from three marmosets, *Callithrix Jacchus*, 2 male and 1 female, 350-370 g, subjects
544 125D (Mirza), 59D (Ramon), and 132F (Charlie), during a 3.5-year period. The marmosets were born and
545 raised in a colony that Prof. Xiaoqin Wang has maintained at the Johns Hopkins School of Medicine since
546 1996. The procedures on the marmosets were approved by the Johns Hopkins University Animal Care
547 and Use Committee in compliance with the guidelines of the United States National Institutes of Health.

548

549 *Data acquisition*

550 Following recovery from head-post implantation surgery, the animals were trained to make saccades to
551 visual targets and rewarded with a mixture of applesauce and lab diet (16). Visual targets were
552 presented on an LCD screen. Binocular eye movements were tracked at 1000 Hz using EyeLink in subject
553 R and M, and 2000 Hz using VPIX in subject C. Tongue movements were tracked with a 522 frame/sec
554 Sony IMX287 FLIR camera, with frames captured at 100 Hz.

555 We performed MRI and CT imaging on each animal and used the imaging data to design an
556 alignment system that defined trajectories from the burr hole to various locations in the cerebellar
557 vermis (16), including points in lobule VI and VII. We used a piezoelectric, high precision microdrive (0.5
558 micron resolution) with an integrated absolute encoder (M3-LA-3.4-15 Linear smart stage, New Scale
559 Technologies) to advance the electrode.

560 We recorded from lobules VI and VII of the cerebellum (Fig. 1C) using quartz insulated 4 fiber
561 (tetrode) or 7 fiber (heptode) metal core (platinum/tungsten 95/05) electrodes (Thomas Recording), and
562 64 channel checkerboard or linear high density silicon probes (M1 and M2 probes, Cambridge
563 Neurotech). We connected each electrode to a 32 or 64 channel head stage amplifier and digitizer
564 (RHD2132 and RHD2164, Intan Technologies, USA), and then connected the head stage to a
565 communication system (RHD2000 Evaluation Board, Intan Technologies, USA). Data were sampled at 30
566 kHz and band-pass filtered (2.5 - 7.6 kHz).

567 The silicon probes arrived with a polymer coating on the contacts that degraded with each
568 insertion into the brain (68). This degradation increased the impedance of the electrodes and
569 dramatically reduced the ability of the probe to isolate neurons. We found it essential to rejuvenate the
570 silicon probes by stripping and then re-depositing the polymer coating after every 3-4 insertions into the
571 brain (68).

572

573 *Behavioral protocol*

574 Each trial began with fixation of a center target after which a primary target appeared at one of 8
575 randomly selected directions at a distance of 5-6.5 deg. As the subject made a saccade to this primary
576 target, that target was erased, and a secondary target was presented at a distance of 2-2.5 deg, also at
577 one of 8 randomly selected directions. The subject was rewarded if following the primary saccade, it
578 made a corrective saccade to the secondary target, landed within 1.5 deg radius of the target center,
579 and maintained fixation for at least 200 ms. The food was provided via two small tubes (4.4 mm
580 diameter), one to the left and the other to the right of the animal, positioned at 90° with respect to the
581 mouth. A successful trial produced a food increment in one of the tubes and would continue to do so for
582 50-300 consecutive trials, then switch to the other tube. Because the food increment was small, the
583 subjects naturally chose to work for a few consecutive trials, tracking the visual targets and allowing the
584 food to accumulate, then stopped tracking and harvested the food via a licking bout. The subjects did
585 not work while harvesting, and often fixated the tube. As a result, the behavior consisted of a work

586 period of targeted saccades, followed by a harvest period of targeted licking, repeated hundreds of
587 times per session.

588 We measured eye movements during all phases of the task, including the bouts of licking. The
589 monkeys tended to fixate the tube while licking. We analyzed tongue movements using DeepLabCut
590 (69). Our network was trained on 89 video recordings of each subject with 15-25 frames extracted and
591 labeled from each recording. The network was built on the ResNet-152 pre-trained model, and then
592 trained over 1.03×10^6 iterations with a batch size of 8, using a GeForce GTX 1080Ti graphics processing
593 unit. A Kalman filter was further applied to improve quality and smoothness of the tracking, and the
594 output was analyzed in MATLAB to quantify lick events and kinematics. We tracked the tongue tip and
595 the edge of the food in the tube, along with control locations (nose position and tube edges). We
596 tracked all licks, regardless of whether they were aimed toward a tube, or not. Food-tube licks were
597 further differentiated based on whether they aimed to enter the tube (inner-tube licks) or hit the outer
598 edge of the tube (outer-edge licks). If any of these licks successfully contacted the food, we labeled that
599 lick as a success (otherwise, an unsuccessful lick).

600

601 *Tracking the tongue*

602 The following videos provide examples of the various types of licks, along with the kinematic measures
603 that we used to track each movement: [videos 1-10](#). Licks were categorized based on heuristics that
604 considered the position of the tongue relative to the tube opening and the food. We tracked 4 regions
605 of the tongue consisting of the tip, the midpoint, and the left and right edges. The midpoint was
606 computed based on the distance between the tip marker and the opening of the mouth, while the left
607 and right edges were computed based on the mid-distance between the tip and midpoint, positioned at
608 max laterality. Furthermore, we tracked the left and right edges of the opening of each reward tube as
609 well as the densest edge of the food contained within.

610 Licks were labeled as reward seeking when the region of the tongue within the marker
611 overlapped with the edge of the tube coordinates. Alternatively, licks were labeled as grooming when
612 no overlap occurred. Reward seeking licks were further labeled into subcategories, consisting of inner-
613 tube and outer-tube licks. Inner-tube labels were assigned when the tip, left, and right tongue markers
614 remained within the bounds of the tube edge markers. Outer-tube labels were assigned when at least
615 one marker exited the tube boundaries, conditioned on the tip having remained within at least 5 mm of
616 the tube opening.

617 Additional labels were assigned to each reward seeking licks, categorizing them as either
618 successful or unsuccessful licks. In all cases, overlap with food dictated these labels. Thus, to call a given
619 lick an unsuccessful lick, the position of the food within the tube, relative to the tongue, was considered.
620 For example, consider a scenario in which the food is depleted, requiring an inner-tube lick to scoop out
621 the remaining bolus. If the lick entered the tube and thus touched the food, it was considered a success.
622 If it did not enter the tube and thus did not touch the food, it was considered an unsuccessful lick.

623

624 *Neurophysiological analysis*

625 We used OpenEphys (70) for electrophysiology data acquisition, and then used P-sort (28) to identify the
626 simple and complex spikes in the heptodes and tetrodes recordings. We used Kilosort and Phi (71) to
627 identify the spikes for the silicon probes. Simple and complex spike instantaneous firing rate were
628 calculated from peri-event time histograms with 1 ms bin size. We used a Savitzky–Golay filter (2nd
629 order, 31 datapoints) to smooth the traces for visualization purposes.

630 Many P-cells in lobules VI and VII of the vermis were modulated during licking as well as during
631 saccades. Our data here were selected from recordings that isolated P-cells with strong tongue related
632 activity. The strength of behavioral modulation for each P-cell during saccades and licks was quantified
633 using a z-score (Supplementary Fig. S2B). This z-score was calculated for each behavior via the range of
634 the P-cell's average stimulus-aligned response divided by the standard deviation of this range, as
635 computed across 2,000 permuted responses. Range was defined as the maximum change in firing rate
636 from pre-behavior to post-behavior for a given response. This approach relies on the notion that if a cell
637 is responsive to a given stimulus, it will exhibit both a strong response (high range) and a consistent
638 response (low standard deviation of range values). Consistent with earlier work (9), the threshold for
639 significant modulation during licking was set at a z-score of 3.

640 CS baseline firing rates were computed by dividing the total number of spikes by the duration of
641 the entire recording. SS baseline firing rates were computed using two different methods depending on
642 the analysis. For bout related responses, baseline was defined as the average firing rate in a 300 ms
643 window preceding bout onset by 700 ms, i.e. during the [-1000 to -700] ms period. However, to analyze
644 the activities during individual licks, because the rates were not stationary but gradually changing from
645 the first to the last lick in the bout, baseline SS rates were computed using the average firing within a
646 sliding window of 2 seconds, consisting of 5-6 licks.

647 To explore how the SS rates changed with the kinematic parameters of the lingual movements,
648 we visualized the firing rates as a function of tongue endpoint position. The firing rates of each P-cell
649 during maximal tongue velocity were computed on a trial-by-trial basis and associated with the spatial
650 coordinates corresponding to the endpoint of that trial's lick. Single trial spike data was smoothed with a
651 Savitzky-Golay filter. Spatial coordinates were standardized across animals such that all contralateral
652 licks appear to the left and all ipsilateral licks to the right. A 100x50 grid was mapped onto the full range
653 of tongue endpoint values, and the population firing rates at each point were estimated using a natural
654 neighbor interpolation, effectively weighing contributions of neighboring firing rate values based on
655 proximity. The interpolated surface was then smoothed with a 2-D Gaussian filter to produce a
656 continuous heatmap. To ensure population-level robustness of firing rate values, a cell coverage mask
657 was then applied over the heatmap, removing any grid points that did not have at least 75% of the
658 available PCs (118/157 SS cells).

659

660 *Computing the kinematic effects of CS-induced SS suppression*

661 For each P-cell we considered triplets of tube-directed licks $\{n - 1, n, n + 1\}$, where all three licks were
662 of the same type, i.e., contacted the same part of the tube (edge or inner). We then selected the subset
663 of triplets in which there was a CS at only a single period in lick n , but no CS during any period in the two
664 neighboring licks $n - 1$, and $n + 1$. We then compared tongue trajectories between the lick that had a
665 CS with the two neighboring licks, i.e., $n - (n - 1)$ and $n - (n + 1)$.

666

667 *Computing the kinematic effects of SS pauses*

668 To assess if the perturbation of tongue movements was a consequence unique to the presence of a CS,
669 or rather the suppression of SSs, we considered the effect of SS pauses on the tongue trajectory during
670 licking, i.e., long ISI events that were not preceded by a CS.

671 For each P-cell, we selected the subset of all licks of the same type towards the same direction
672 in which no CSs occurred at any point in the movement. Let us call these the NoCS licks. Working only
673 with the NoCS licks, we sought to identify licks in which during a phase of interest (e.g., protraction

674 deceleration), there was a long pause in the SS production. However, we had to ensure that if there was
675 a long pause in one phase of the lick, it did not also occur in other phases of the same lick. That is, like
676 the CS analysis, to be eligible for this analysis a long SS pause had to occur only once during the lick.

677 There were 4 phases for each lick (protraction acceleration and deceleration, retraction
678 acceleration and deceleration), i.e., $p = 1, \dots, 4$. For each lick n , during each phase p , we found the
679 duration of the longest ISI that originated in that phase (regardless of whether it extended into the next
680 phase) and labeled it as $t_n^{(p)}$. Next, for each phase, we found the distribution of $t_n^{(p)}$. Licks with zero SSs
681 during the given phase were excluded from this distribution.

682 For example, suppose we were interested in labeling the licks in which during phase 1 there was
683 a long pause. A lick with a long pause in phase 1 could not have also had a long pause in another phase
684 of that same lick. We found the distribution of $t_n^{(2)}$, the distribution of $t_n^{(3)}$, and the distribution of $t_n^{(4)}$,
685 and then for each phase selected the top quartile (25% longest ISIs). We removed the licks with a long
686 pause in phase 2-4 for consideration. From among the remaining licks, we formed the distribution of
687 $t_n^{(1)}$, found the top quartile, and labeled those as having a long pause during phase 1. We labeled the
688 remaining 75% of licks in this population as not having a long pause during this phase.

689 We selected the subset of triplets in which lick n had a long SS pause in only one phase of the
690 movement, but no SS pause occurred in any phase of the two neighboring licks. We then compared
691 tongue trajectories between the lick that had a pause with the two neighboring licks. Traces were
692 averaged within directions and then across directions for each cell.

693

694 *Computing the effects of trajectory error on climbing fiber activity*

695 Roughly 15% of the licks failed to enter the tube and did not touch the food (Supplementary Fig. S5). To
696 visualize the complex spike patterns as a function of the spatial location of the tongue, we began with
697 computing $p(x(t - 25)|CS)$, i.e., given that a CS occurred at time t , the likelihood of the tongue's tip
698 location x at time $t - 25$ ms. We did this by averaging the position of the tip of the tongue during the
699 50ms period before the CS event. We separated the licks into successful licks (tongue entered the tube
700 and touched the food) and unsuccessful licks (tongue touched the tube but neither entered it nor
701 touched the food). The result was the likelihood $p(x_s|CS)$ for the successful licks and $p(x_u|CS)$ for the
702 unsuccessful licks.

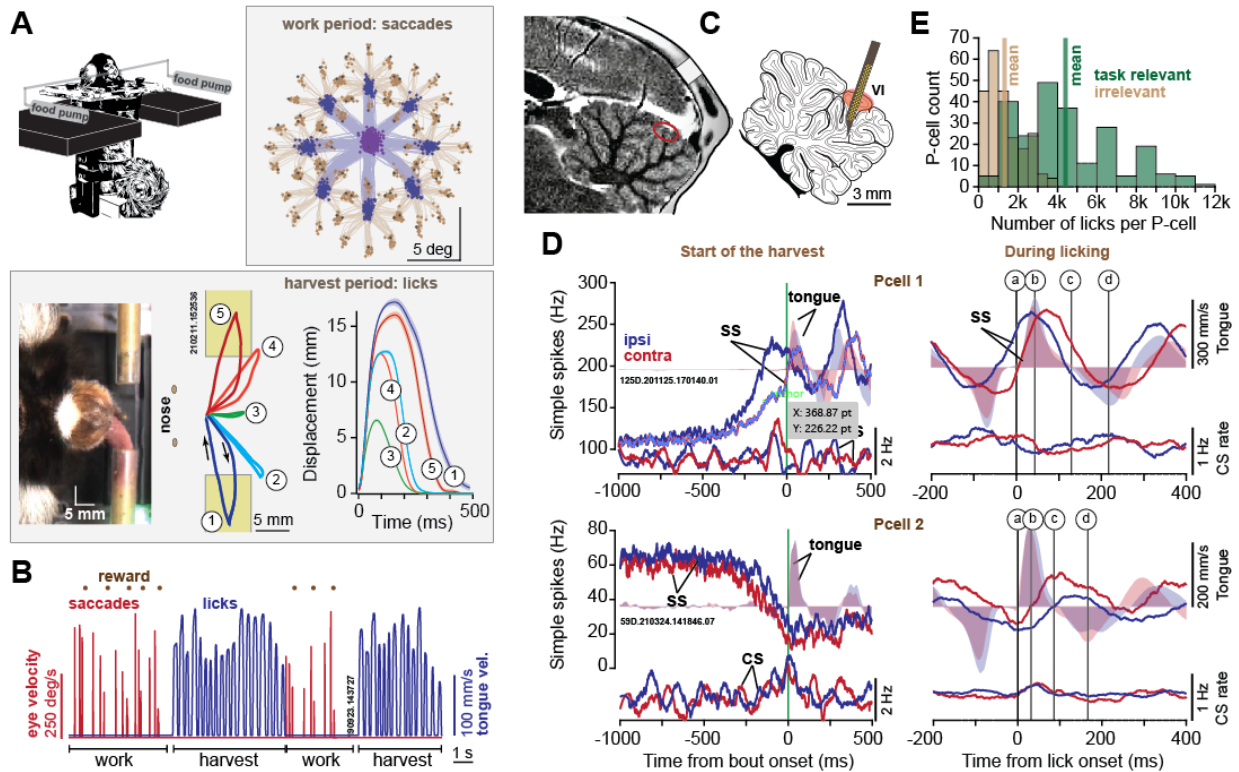
703 We next computed the marginal probability density $p(x)$ for each lick type, the prior $\Pr(CS)$
704 (from the average CS rate during a lick of that type, using a 50ms time bin), and then the ratio of the
705 probabilities $p(x|CS) \Pr(CS) / p(x)$. Finally, we computed the error-induced spatial pattern of complex
706 spikes by subtracting this ratio for the successful licks from the same ratio for the unsuccessful licks. To
707 reduce the noise associated with the far edges of each probability density function, for each ratio we
708 considered values that were in the 95% quantile of its distribution.

709

710 *Statistical analysis*

711 In order to compare the measured effect of SS suppression on tongue trajectory with what would be
712 expected to happen simply due to chance, we computed the bounds for the null hypothesis. To do so,
713 we used bootstrapping to compute 95% confidence intervals. We shuffled the assignment of CS tags
714 from the lick in which it had occurred to a randomly assigned lick of the same type. Using this pseudo-
715 data, we then selected triplets of consecutive tube-directed licks and computed trajectory differences
716 among neighboring licks, averaging $n - (n - 1)$ and $n - (n + 1)$. We computed this expected value for

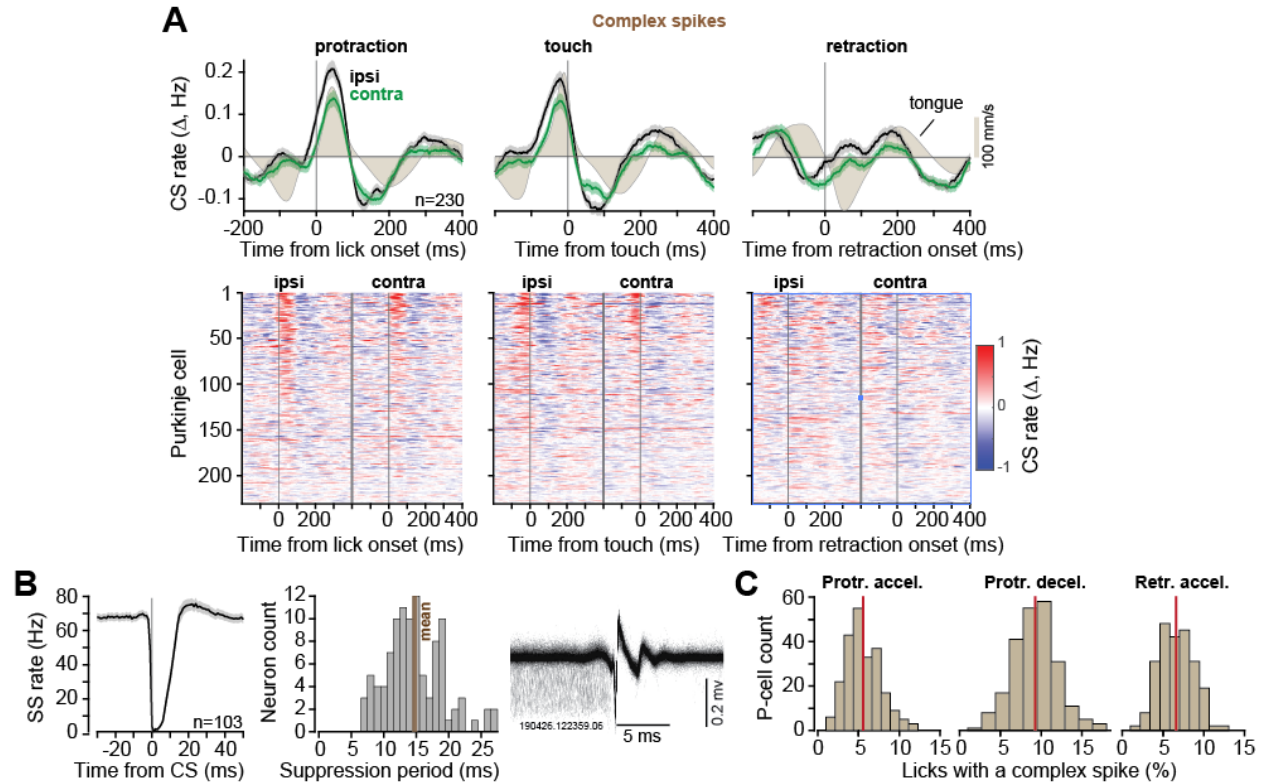
717 each cell, computed a mean across all the cells, and then repeated the shuffling 30 times to compute
718 95% confidence intervals.
719



720
721

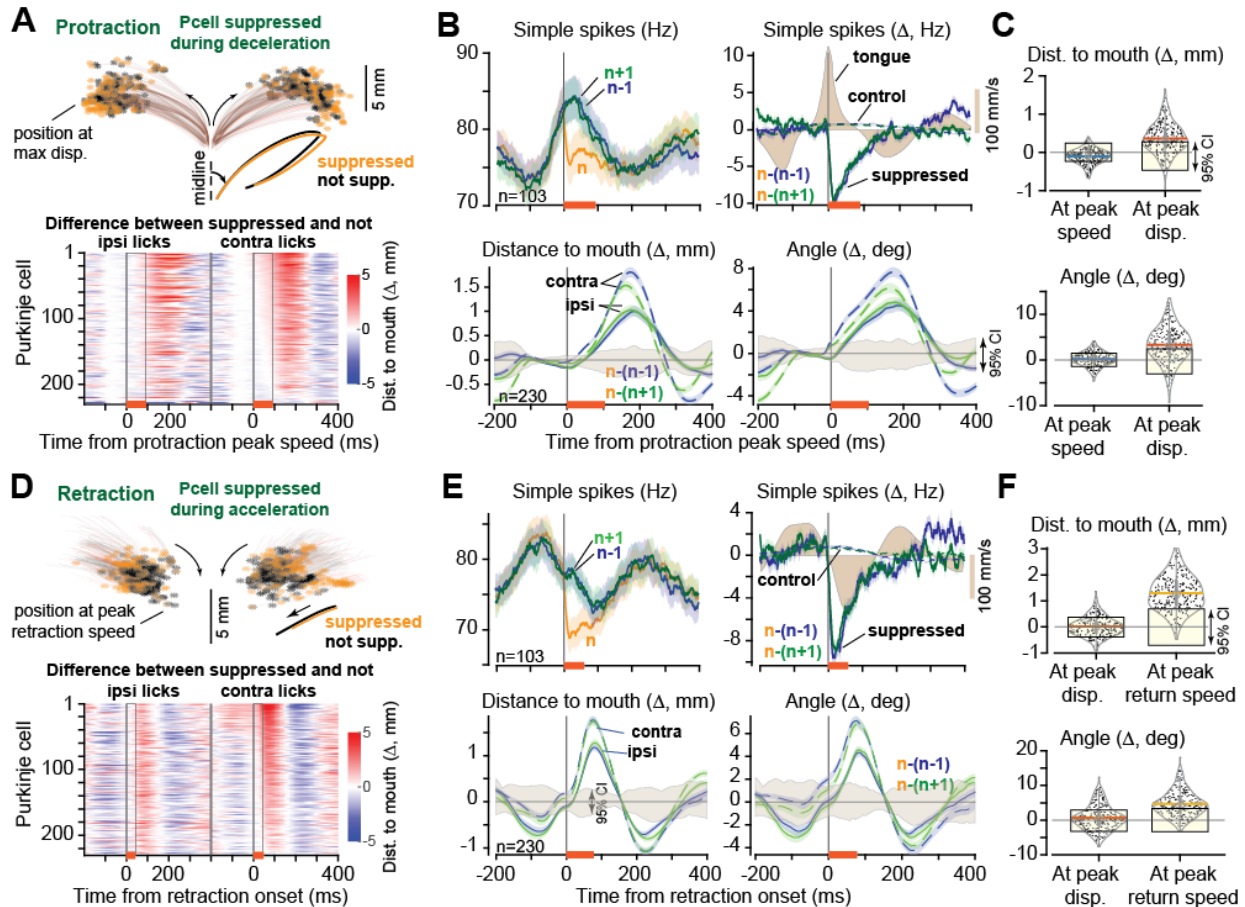
722 **Figure 1. Marmosets produced dexterous tongue movements during recordings from the cerebellar vermis. A.**
 723 Subjects made saccades to visual targets and received a small amount of food as reward via one of two tubes
 724 placed obliquely to the mouth. In the task relevant licks, they directed their tongue to the edge of the tube to
 725 harvest food near the tip (trajectories 2, 4), or inside the tube to harvest food that was deeper (trajectories 1, 5). In
 726 task irrelevant licks, they groomed their face (trajectory 3). **B.** Subjects chose to work for consecutive trials, making
 727 saccades and allowing the food to accumulate, then harvested their cache in bouts of licking. **C.** We employed
 728 silicon probes to record from lobule VI and VII of the vermis. **D.** Simple and complex spikes (SS, CS) of two Purkinje
 729 cells, aligned to bout onset and lick onset. A single lick was divided into acceleration period of protraction (a-b),
 730 deceleration period of protraction (b-c), and acceleration period of retraction (c-d). Filled color regions indicate
 731 tongue velocity. **E.** The number of task relevant (tube directed) and task irrelevant (grooming) licks recorded per
 732 neuron.

733
734



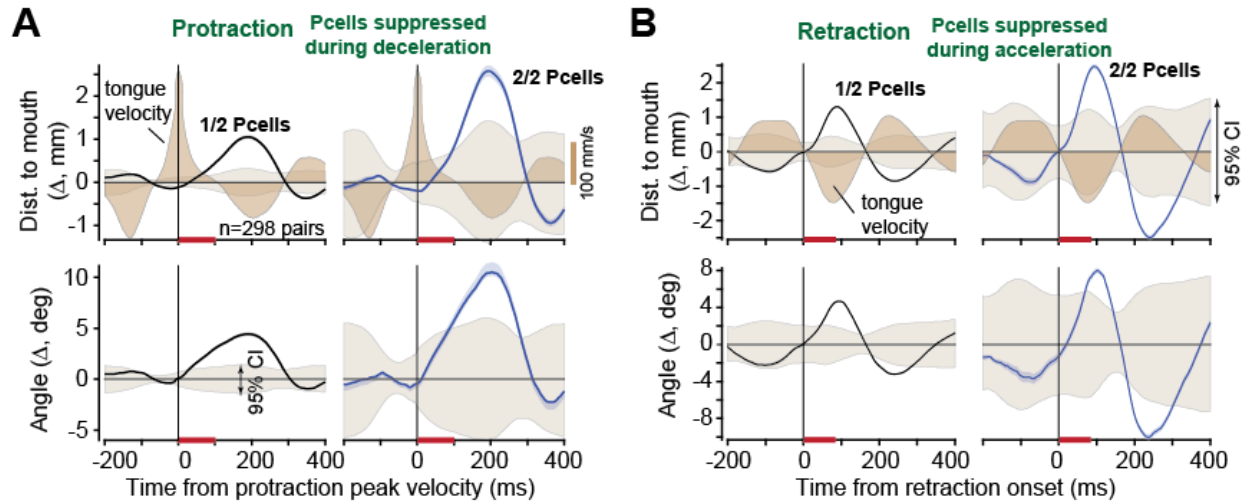
735
736
737
738
739
740
741
742
743

Figure 2. CS rates increased with protraction and decreased with retraction. **A.** CS rates across the population aligned to protraction onset, touch of the tube, and retraction onset. The second row shows CS activity across all P-cells, aligned to protraction, touch, and retraction, sorted based on CS rate for ipsilateral licks at lick onset. **B.** CS-induced SS suppression, averaged across the subset of P-cells for which both the CS and the SS were isolated (left). Examples of SS and CS waveforms for a single P-cell are shown at right. **C.** Percentage of licks with a CS during a specific period of time for each neuron. Vertical line indicates mean. Error bars are SEM.



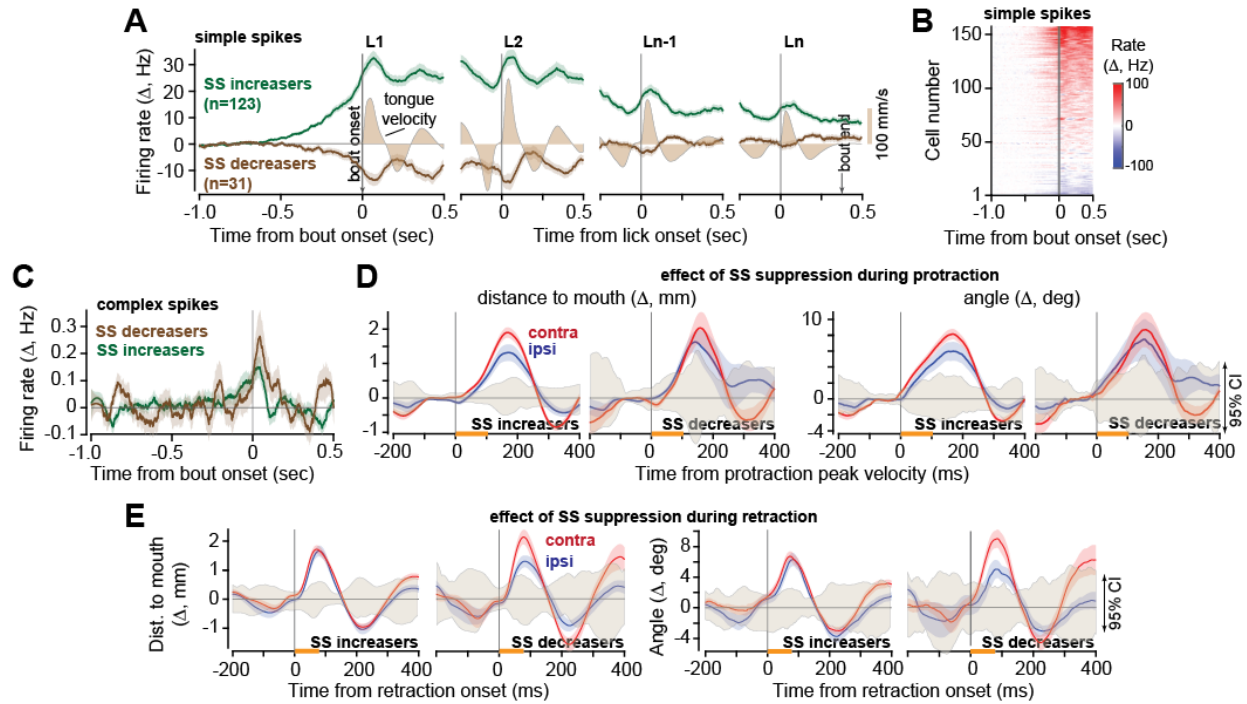
744
 745
 746
 747
 748
 749
 750
 751
 752
 753
 754
 755
 756
 757
 758
 759

Figure 3. CS-induced SS suppression produced hypermetria during protraction and slowing during retraction. A. Suppression took place during the deceleration period of protraction. Traces show average tongue trajectory during this period of protraction for each P-cell during SS suppressed and control licks. Ipsilateral licks are shown to the left and contralateral to the right. Heatmap quantifies change in endpoint trajectory between suppressed and control licks for each cell. Period of suppression is indicated by the orange bar at the bottom of the heatmap. **B.** Top row: SS rates for licks { , , }, where only lick experienced a CS. Filled color curves indicate tongue velocity. Second row: trajectory of the tongue in lick as compared to its two temporally neighboring licks. Trajectory is measured via distance from tip of the tongue to the mouth and angle of the tip with respect to midline. The filled region is 95% CI. **C.** Distance to mouth and angle in lick as compared to neighboring licks. Shaded region is 95% CI. **D-F.** Suppression during the acceleration period of retraction induced slowing. Same format as in parts A-C. In part F, tongue trajectory (displacement and angle) is similar before SS suppression (at peak displacement) but diverges after the suppression at peak return speed. Error bars are SEM.



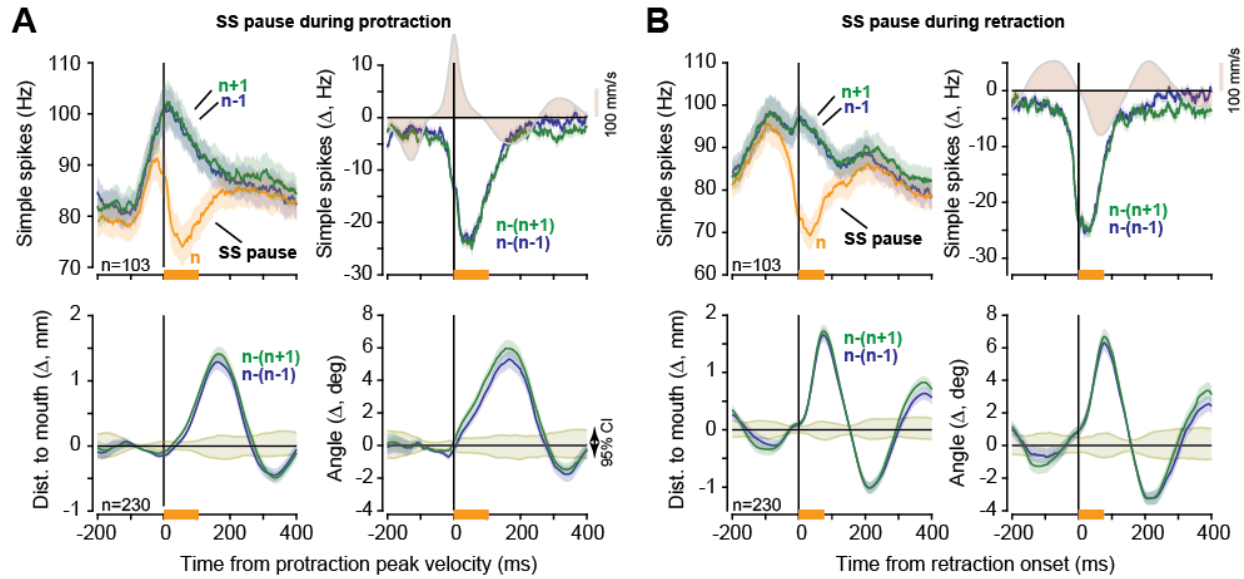
760
761
762
763
764
765
766
767
768
769

Figure 4. Suppression of multiple P-cells scaled the perturbation to the tongue. **A.** Kinematic effects of CS-induced P-cell suppression during the deceleration period of protraction (orange bar). Traces show change in tongue trajectory in suppressed licks vs. control licks, measured via distance from tip of the tongue to the mouth and angle of the tip with respect to midline. Gray shaded region is 95% CI. Brown filled region is tongue velocity. **B.** Kinematic effects of P-cell suppression during the acceleration period of retraction (orange bar). Same format as in part A. Error bars are SEM.



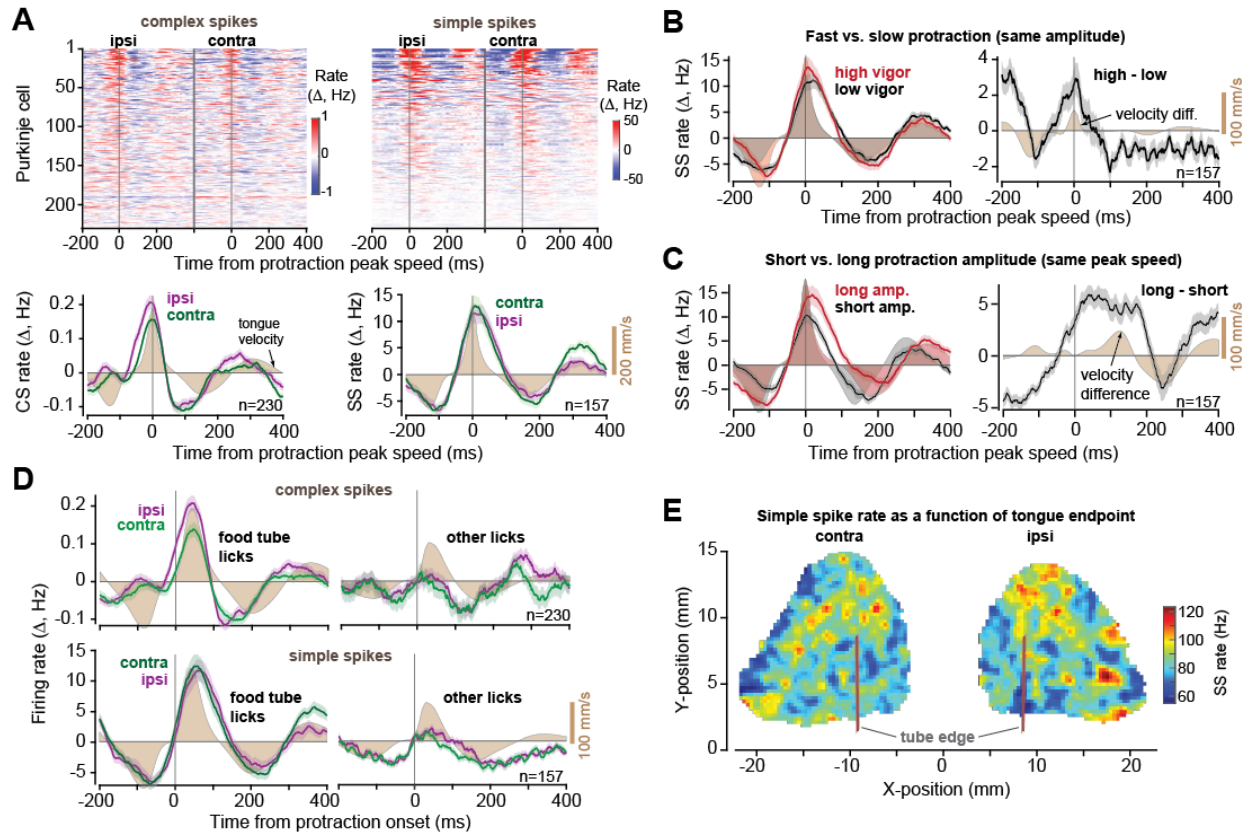
770
771
772
773
774
775
776
777
778

Figure 5. Effects of SS suppression remained consistent across P-cells regardless of SS modulation. **A.** Some of the P-cells exhibited an increase in SS rates before bout onset, while others exhibited a decrease. L1 is first lick in the bout, Ln is last lick. **B.** Change in SS rates for all P-cells (with respect to baseline), aligned to bout onset. **C.** Change in complex spike rate for the same P-cells. **D.** Kinematic effects of SS suppression during the deceleration period of protraction (orange bar). Shaded region is 95% CI. **E.** Kinematic effects of SS suppression during the acceleration period of retraction (orange bar). Error bars are SEM.



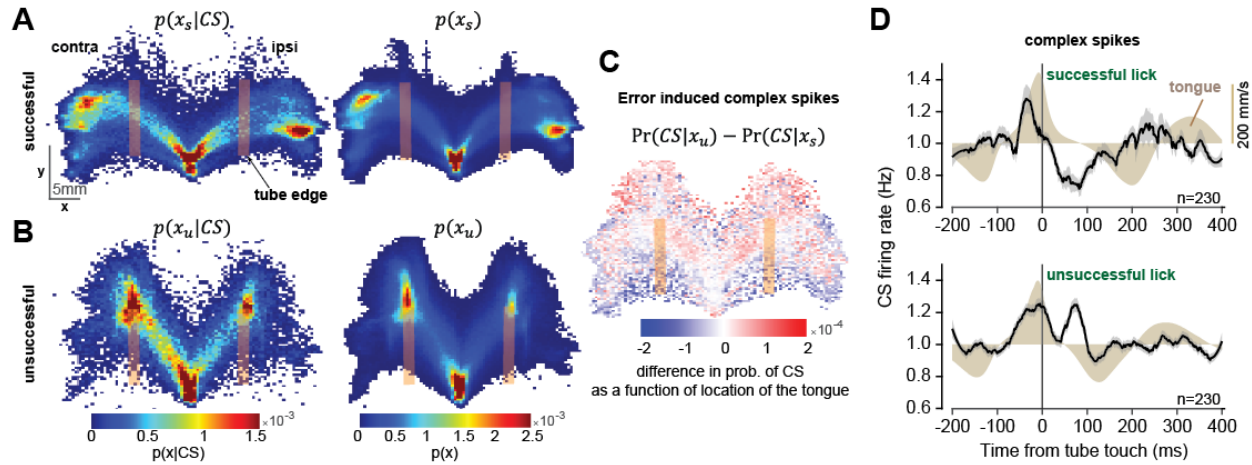
779
780

781 **Figure 6. SS pause without a CS was sufficient to produce hypermetria and bending of the tongue.** A. We
782 selected licks that did not have a CS but nevertheless experienced a long SS pause during the protraction
783 deceleration period. Top row: SS rates for lick n that experienced a pause and licks n-1 and n+1 that did not.
784 Bottom row: change in lick kinematics following the SS pause. B. Same as in part A, but for licks that experience a
785 long SS pause during the retraction acceleration period.



786
787
788
789
790
791
792
793
794
795
796
797

Figure 7. SS rates peaked at deceleration onset, but only if the lick was task relevant. **A.** CS and SS rates across the population aligned to protraction peak speed. **B.** Left figure shows SS rates for high and low vigor protractions (same amplitude). Right figure shows within cell difference in SS rates for high minus low vigor licks. Shaded regions are tongue speed for high and low vigor licks (left) and change in speed (right). **C.** Left figure shows SS rates for protractions that had long or short amplitudes but the same peak speed. Right figure shows within cell differences. Shaded regions are tongue speed for long and short licks (left) and change in speed (right). **D.** Modulation of CS and SS rates during task relevant and task irrelevant licks. Light shaded region is tongue speed for task relevant movements, while dark shaded region is tongue speed for task irrelevant movements. **E.** SS rates at peak velocity as a function of tongue position at maximum displacement. Error bars are SEM.



798

799 **Figure 8. Error in the tongue's trajectory induced complex spikes.** A. Successful licks: tongue entered the tube and
800 touched the food. Left subfigure shows the spatial likelihood at 25 ms before the CS event. Right subfigure shows
801 the probability of being at a spatial location. B. Unsuccessful licks: tongue did not enter the tube and collided with
802 the tube's edge. C. Spatial pattern of the error induced complex spikes (difference between unsuccessful and
803 successful licks in the posterior probability of CS as a function of the tongue's location). D. Complex spike rates
804 following tube touch for successful and unsuccessful licks.

805

806

807 **Video 1.** A sequence of 5 licks to the right tube. The top two plots show trajectory of the tip of the tongue and a
808 geometric representation of four markers on the tongue. The 2nd row shows the displacement, velocity, and angle
809 of the tongue as a function of time. The 3rd row shows the distance of the tip of the tongue to the food in the left
810 and the right tube. The first 3 licks are successful and enter the tube and contact the food. In the 4th and 5th licks
811 the tongue fails to enter the tube and do not contact the food. These licks are unsuccessful and are analyzed in Fig.
812 8.
813
814 **Video 2.** Example of a grooming lick. These licks aim to clean the regions around the mouth and are not aimed
815 toward the food tubes.
816
817 **Video 3.** Example of a grooming lick.
818
819 **Video 4.** Example of a bout of grooming licks.
820
821 **Video 5.** Example of an outer-tube lick. The food has accumulated beyond the edge of the tube and the subject
822 begins the bout by licking the food near the edge.
823
824 **Video 6.** Example of an inner-tube lick. The food is deep inside the tube and the subject enters the tube and
825 scoops the food out.
826
827 **Video 7.** Example of an inner-tube lick.
828
829 **Video 8.** Unsuccessful lick. The food is inside the tube, but the lick fails to enter it and instead goes under the tube.
830
831 **Video 9.** Unsuccessful lick. The food is inside the tube, but the lick fails to enter it and instead goes to the outer
832 edge.
833
834 **Video 10.** Unsuccessful lick. The food is inside the tube, but the lick fails to enter it and instead collides with the
835 edge.
836
837

- 838 1. W. J. M. Levelt, *Speaking: From Intention to Articulation* (MIT Press, 1993).
- 839 2. M. Gentil, EMG analysis of speech production of patients with Friedreich disease. *Clin. Linguist.*
840 *Phon.* **4**, 107–120 (1990).
- 841 3. T. Vilis, J. Hore, Central neural mechanisms contributing to cerebellar tremor produced by limb
842 perturbations. *J. Neurophysiol.* **43**, 279–291 (1980).
- 843 4. F. Yu, Q. J. Jiang, X. Y. Sun, R. W. Zhang, A new case of complete primary cerebellar agenesis:
844 clinical and imaging findings in a living patient. *Brain* **138**, e353 (2015).
- 845 5. J. L. Bryant, J. D. Boughter, S. Gong, M. S. LeDoux, D. H. Heck, Cerebellar cortical output encodes
846 temporal aspects of rhythmic licking movements and is necessary for normal licking frequency.
847 *Eur. J. Neurosci.* **32**, 41–52 (2010).
- 848 6. M. A. Gaffield, J. M. Christie, Movement Rate Is Encoded and Influenced by Widespread, Coherent
849 Activity of Cerebellar Molecular Layer Interneurons. *J. Neurosci.* **37**, 4751–4765 (2017).
- 850 7. M. I. Becker, A. L. Person, Cerebellar Control of Reach Kinematics for Endpoint Precision. *Neuron*
851 **103**, 335–348 (2019).
- 852 8. J. S. Pi, *et al.*, The olivary input to the cerebellum dissociates sensory events from movement plans.
853 *Proc. Natl. Acad. Sci.* **121**, e2318849121 (2024).
- 854 9. E. Sedaghat-Nejad, J. S. Pi, P. Hage, M. A. Fakharian, R. Shadmehr, Synchronous spiking of
855 cerebellar Purkinje cells during control of movements. *Proc. Natl. Acad. Sci.* **119**, e2118954119
856 (2022).
- 857 10. J. P. Bowman, L. D. Aldes, Organization of the cerebellar tongue representation in the monkey.
858 *Exp. Brain Res.* **39**, 249–259 (1980).
- 859 11. L. D. Aldes, J. P. Bowman, Representation of the tongue in the cerebellar nuclei of the monkey.
860 *Exp. Neurol.* **64**, 202–215 (1979).
- 861 12. M. A. Gaffield, B. A. Sauerbrei, J. M. Christie, Cerebellum encodes and influences the initiation,
862 performance, and termination of discontinuous movements in mice. *eLife* **11**, e71464 (2022).
- 863 13. J. P. Welsh, E. J. Lang, I. Sugihara, R. Llinas, Dynamic organization of motor control within the
864 olivocerebellar system. *Nature* **374**, 453–457 (1995).
- 865 14. K. Kubota, S. Hayama, Comparative anatomical and neurohistological observations on the tongues
866 of pigmy and common marmosets. *Anat. Rec.* **150**, 473–485 (1964).
- 867 15. G. Oren, *et al.*, Vocal labeling of others by nonhuman primates. *Science* **385**, 996–1003 (2024).
- 868 16. E. Sedaghat-Nejad, *et al.*, Behavioral training of marmosets and electrophysiological recording
869 from the cerebellum. *J. Neurophysiol.* **122**, 1502–1517 (2019).

- 870 17. P. Hage, *et al.*, Effort cost of harvest affects decisions and movement vigor of marmosets during
871 foraging. *eLife* **12**, RP87238 (2023).
- 872 18. D. J. Herzfeld, Y. Kojima, R. Soetedjo, R. Shadmehr, Encoding of error and learning to correct that
873 error by the Purkinje cells of the cerebellum. *Nat. Neurosci.* **21**, 736–743 (2018).
- 874 19. C. Ju, *et al.*, Neurons of the inferior olive respond to broad classes of sensory input while subject to
875 homeostatic control. *J. Physiol.* **597**, 2483–2514 (2019).
- 876 20. S. Tsutsumi, *et al.*, Purkinje Cell Activity Determines the Timing of Sensory-Evoked Motor Initiation.
877 *Cell Rep.* **33**, 108537 (2020).
- 878 21. Y. Sato, A. Miura, H. Fushiki, T. Kawasaki, Short-term modulation of cerebellar Purkinje cell activity
879 after spontaneous climbing fiber input. *J. Neurophysiol.* **68**, 2051–2062 (1992).
- 880 22. W. T. Thach, Somatosensory receptive fields of single units in cat cerebellar cortex. *J. Neurophysiol.*
881 **30**, 675–696 (1967).
- 882 23. V. Romano, *et al.*, Olivocerebellar control of movement symmetry. *Curr. Biol.* **32**, 654-670.e4
883 (2022).
- 884 24. M. L. Streng, L. S. Popa, T. J. Ebner, Climbing Fibers Control Purkinje Cell Representations of
885 Behavior. *J. Neurosci.* **37**, 1997–2009 (2017).
- 886 25. S. Z. Muller, *et al.*, Complex spikes perturb movements and reveal the sensorimotor map of
887 Purkinje cells. *Curr. Biol.* **33**, 4869-4879.e3 (2023).
- 888 26. M. A. Fakharian, A. M. Shoup, P. Hage, H. Y. Elseweifi, R. Shadmehr, A vector calculus for neural
889 computation in the cerebellum. [Preprint] (2024). Available at:
890 <https://www.biorxiv.org/content/10.1101/2024.11.14.623565v1> [Accessed 18 November 2024].
- 891 27. A. Mathis, *et al.*, DeepLabCut: markerless pose estimation of user-defined body parts with deep
892 learning. *Nat. Neurosci.* **21**, 1281–1289 (2018).
- 893 28. E. Sedaghat-Nejad, *et al.*, P-sort: an open-source software for cerebellar neurophysiology. *J.*
894 *Neurophysiol.* **126**, 1055–1075 (2021).
- 895 29. T. R. Reppert, *et al.*, Movement vigor as a trait-like attribute of individuality. *J. Neurophysiol.* **120**,
896 741–757 (2018).
- 897 30. Y. Kojima, F. R. Robinson, R. Soetedjo, Cerebellar fastigial nucleus influence on ipsilateral abducens
898 activity during saccades. *J. Neurophysiol.* **111**, 1553–1563 (2014).
- 899 31. C. Burrell, J. Quinet, L. Goffart, The caudal fastigial nucleus and the steering of saccades toward
900 a moving visual target. *J. Neurophysiol.* **120**, 421–438 (2018).
- 901 32. C. W. Mohler, R. H. Wurtz, Organization of monkey superior colliculus: intermediate layer cells
902 discharging before eye movements. *J. Neurophysiol.* **39**, 722–744 (1976).

- 903 33. D. M. Waitzman, T. P. Ma, L. M. Optican, R. H. Wurtz, Superior colliculus neurons mediate the
904 dynamic characteristics of saccades. *J. Neurophysiol.* **66**, 1716–1737 (1991).
- 905 34. M. E. Goldberg, C. J. Bruce, Primate frontal eye fields. III. Maintenance of a spatially accurate
906 saccade signal. *J. Neurophysiol.* **64**, 489–508 (1990).
- 907 35. B. S. Ito, Y. Gao, B. Kardon, J. H. Goldberg, A collicular map for touch-guided tongue control.
908 *Nature* 1–9 (2025). <https://doi.org/10.1038/s41586-024-08339-3>.
- 909 36. T. M. Hoogland, J. R. De Griijl, L. Witter, C. B. Canto, C. I. De Zeeuw, Role of Synchronous
910 Activation of Cerebellar Purkinje Cell Ensembles in Multi-joint Movement Control. *Curr. Biol.* **25**,
911 1157–1165 (2015).
- 912 37. S. Kitazawa, T. Kimura, P. B. Yin, Cerebellar complex spikes encode both destinations and errors in
913 arm movements. *Nature* **392**, 494 (1998).
- 914 38. A. L. Hewitt, L. S. Popa, T. J. Ebner, Changes in Purkinje cell simple spike encoding of reach
915 kinematics during adaption to a mechanical perturbation. *J. Neurosci.* **35**, 1106–1124 (2015).
- 916 39. W. Heffley, *et al.*, Coordinated cerebellar climbing fiber activity signals learned sensorimotor
917 predictions. *Nat. Neurosci.* **21**, 1431–1441 (2018).
- 918 40. M. J. Wagner, *et al.*, A neural circuit state change underlying skilled movements. *Cell* **184**, 3731-
919 3747.e21 (2021).
- 920 41. D. J. Calame, M. I. Becker, A. L. Person, Cerebellar associative learning underlies skilled reach
921 adaptation. *Nat. Neurosci.* 1–12 (2023). <https://doi.org/10.1038/s41593-023-01347-y>.
- 922 42. T. Ishikawa, *et al.*, Releasing dentate nucleus cells from Purkinje cell inhibition generates output
923 from the cerebrocerebellum. *PLoS. One.* **9**, e108774 (2014).
- 924 43. Y. Kojima, R. Soetedjo, Change in sensitivity to visual error in superior colliculus during saccade
925 adaptation. *Sci. Rep.* **7**, 9566 (2017).
- 926 44. Y. Kojima, R. Soetedjo, Elimination of the error signal in the superior colliculus impairs saccade
927 motor learning. *Proc. Natl. Acad. Sci. U.S.A* **115**, E8987–E8995 (2018).
- 928 45. P. J. May, S. Warren, Y. Kojima, The superior colliculus projection upon the macaque inferior olive.
929 *Brain Struct. Funct.* (2024). <https://doi.org/10.1007/s00429-023-02743-7>.
- 930 46. R. Soetedjo, Y. Kojima, A. F. Fuchs, How cerebellar motor learning keeps saccades accurate. *J.*
931 *Neurophysiol.* **121**, 2153–2162 (2019).
- 932 47. A. Thomas, *et al.*, Superior colliculus bidirectionally modulates choice activity in frontal cortex. *Nat.*
933 *Commun.* **14**, 7358 (2023).
- 934 48. T. Tang, T. A. Blenkinsop, E. J. Lang, Complex spike synchrony dependent modulation of rat deep
935 cerebellar nuclear activity. *eLife* **8**, e40101 (2019).

- 936 49. J. F. Medina, S. G. Lisberger, Links from complex spikes to local plasticity and motor learning in the
937 cerebellum of awake-behaving monkeys. *Nat. Neurosci.* **11**, 1185–1192 (2008).
- 938 50. Y. Yang, S. G. Lisberger, Role of Plasticity at Different Sites across the Time Course of Cerebellar
939 Motor Learning. *J. Neurosci.* **34**, 7077–7090 (2014).
- 940 51. J. Chaumont, *et al.*, Clusters of cerebellar Purkinje cells control their afferent climbing fiber
941 discharge. *Proc.Natl.Acad.Sci.U.S.A.* **110**, 16223–16228 (2013).
- 942 52. R. Soetedjo, G. D. Horwitz, Closed-Loop Optogenetic Perturbation of Macaque Oculomotor
943 Cerebellum: Evidence for an Internal Saccade Model. *J. Neurosci.* **44** (2024).
- 944 53. M. Jelitai, P. Puggioni, T. Ishikawa, A. Rinaldi, I. Duguid, Dendritic excitation–inhibition balance
945 shapes cerebellar output during motor behaviour. *Nat. Commun.* **7**, 13722 (2016).
- 946 54. Z. Gao, *et al.*, A cortico-cerebellar loop for motor planning. *Nature* **563**, 113–116 (2018).
- 947 55. F. P. Chabrol, A. Blot, T. D. Mrsic-Flogel, Cerebellar Contribution to Preparatory Activity in Motor
948 Neocortex. *Neuron* **103**, 506–519 (2019).
- 949 56. D. Xu, *et al.*, Cortical processing of flexible and context-dependent sensorimotor sequences.
950 *Nature* **603**, 464–469 (2022).
- 951 57. J. Zhu, H. Hasanbegović, L. D. Liu, Z. Gao, N. Li, Activity map of a cortico-cerebellar loop underlying
952 motor planning. *Nat. Neurosci.* 1–13 (2023). <https://doi.org/10.1038/s41593-023-01453-x>.
- 953 58. J. Diedrichsen, E. Zotow, Surface-Based Display of Volume-Averaged Cerebellar Imaging Data. *PLOS*
954 *ONE* **10**, e0133402 (2015).
- 955 59. K. Grabski, *et al.*, Functional MRI assessment of orofacial articulators: Neural correlates of lip, jaw,
956 larynx, and tongue movements. *Hum. Brain Mapp.* **33**, 2306–2321 (2012).
- 957 60. W. Grodd, E. Hülsmann, M. Lotze, D. Wildgruber, M. Erb, Sensorimotor mapping of the human
958 cerebellum: fMRI evidence of somatotopic organization. *Hum. Brain Mapp.* **13**, 55–73 (2001).
- 959 61. P. P. Urban, *et al.*, Cerebellar Speech Representation: Lesion Topography in Dysarthria as Derived
960 From Cerebellar Ischemia and Functional Magnetic Resonance Imaging. *Arch. Neurol.* **60**, 965–972
961 (2003).
- 962 62. L. Bina, *et al.*, Cerebellar control of targeted tongue movements. [Preprint] (2024). Available at:
963 <https://www.biorxiv.org/content/10.1101/2024.09.26.615128v1> [Accessed 6 November 2024].
- 964 63. T. J. H. Ruigrok, J. Voogd, Organization of projections from the inferior olive to the cerebellar nuclei
965 in the rat. *J. Comp. Neurol.* **426**, 209–228 (2000).
- 966 64. R. Apps, M. Garwicz, Anatomical and physiological foundations of cerebellar information
967 processing. *Nat.Rev.Neurosci.* **6**, 297–311 (2005).

- 968 65. A. Poologaindran, *et al.*, The effect of unilateral thalamic deep brain stimulation on the vocal
969 dysfunction in a patient with spasmodic dysphonia: interrogating cerebellar and pallidal neural
970 circuits. (2017). <https://doi.org/10.3171/2016.10.JNS161025>.
- 971 66. A. Sasegbon, S. Hamdy, The Role of the Cerebellum in Swallowing. *Dysphagia* **38**, 497–509 (2023).
- 972 67. R. Lechtenberg, S. Gilman, Speech disorders in cerebellar disease. *Ann. Neurol.* **3**, 285–290 (1978).
- 973 68. A. M. Shoup, *et al.*, Rejuvenating silicon probes for acute electrophysiology. [Preprint] (2024).
974 Available at: <https://www.biorxiv.org/content/10.1101/2024.02.20.581222v1> [Accessed 13 March
975 2024].
- 976 69. A. Mathis, *et al.*, DeepLabCut: markerless pose estimation of user-defined body parts with deep
977 learning. *Nat. Neurosci.* **21**, 1281–1289 (2018).
- 978 70. J. H. Siegle, *et al.*, Open Ephys: an open-source, plugin-based platform for multichannel
979 electrophysiology. *JNeural Eng* **14**, 045003 (2017).
- 980 71. M. Pachitariu, N. Steinmetz, S. Kadir, M. Carandini, H. K. D, Kilosort: realtime spike-sorting for
981 extracellular electrophysiology with hundreds of channels. *bioRxiv* 061481 (2016).
982 <https://doi.org/10.1101/061481>.

983

984