

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

ELSEVIER

Contents lists available at ScienceDirect

Microbes and Infection

journal homepage: www.elsevier.com/locate/micinf



From SARS-CoV to SARS-CoV-2: safety and broad-spectrum are important for coronavirus vaccine development



Cuiging Ma ^a, Shan Su ^b, Jiachao Wang ^a, Lin Wei ^a, Lanying Du ^c, Shibo Jiang ^{b, c, *}

- ^a Department of Immunology, Key Laboratory of Immune Mechanism and Intervention on Serious Disease in Hebei Province, Hebei Medical University, 050017, Shijiazhuang, China
- b Key Laboratory of Medical Molecular Virology (MOE/NHC/CAMS), School of Basic Medical Sciences, Shanghai Medical College, Fudan University, Shanghai,
- ^c Lindsley F. Kimball Research Institute, New York Blood Center, New York, NY, 10065, USA

ARTICLE INFO

Article history: Received 6 May 2020 Accepted 7 May 2020 Available online 11 May 2020

Keywords: 2019-nCoV SARS-CoV-2 HCoV-19 SARS-CoV Vaccine Cross-protection

ABSTRACT

The global pandemic of COVID-19 caused by SARS-CoV-2 (also known as 2019-nCoV and HCoV-19) has posed serious threats to public health and economic stability worldwide, thus calling for development of vaccines against SARS-CoV-2 and other emerging and reemerging coronaviruses. Since SARS-CoV-2 and SARS-CoV have high similarity of their genomic sequences and share the same cellular receptor (ACE2), it is essential to learn the lessons and experiences from the development of SARS-CoV vaccines for the development of SARS-CoV-2 vaccines. In this review, we summarized the current knowledge on the advantages and disadvantages of the SARS-CoV vaccine candidates and prospected the strategies for the development of safe, effective and broad-spectrum coronavirus vaccines for prevention of infection by currently circulating SARS-CoV-2 and other emerging and reemerging coronaviruses that may cause future epidemics or pandemics.

© 2020 Institut Pasteur. Published by Elsevier Masson SAS. All rights reserved.

In December 2019, the outbreak of an unexplained pneumonia similar to severe acute respiratory syndrome (SARS) in Wuhan, China was reported by the Health Commission of Hubei Province, China. This severe respiratory illness was identified by multiple diagnostic methods as an infection by a novel coronavirus [1–4], which was temporarily denoted as 2019-nCoV by World Health Organization [5], and renamed "severe acute respiratory syndrome coronavirus 2" (SARS-CoV-2) by the Coronavirus Study Group (CSG) of the International Committee on Taxonomy of Viruses (ICTV) [6]. The new coronavirus-associated pneumonia was designated as coronavirus disease 2019 (COVID-19) by WHO (https://www.who.int/emergencies/diseases/novelcoronavirus-2019 (accessed Feb 18, 2020)).

It has been originally reported that COVID-19 cases are associated with the exposure to a large seafood and animal market in Wuhan, suggesting an animal-to-human transmission. Later on, more epidemiologic data indicate a human-to-human transmission of

E-mail address: shibojiang@fudan.edu.cn (S. Jiang).

SARS-CoV-2 [7—11]. As of April 24, 2020, there are 84324 confirmed cases, 4642 death cases in China, and 2,626,334 confirmed cases, 181,938 cases of death in other countries (http://2019ncov.chinacdc.cn/2019-nCoV/global.html). Currently, the mortality rate in China is about 5.5%, however, about 6.9% globally. In China, there are 1303 existing confirmed cases, of which 983 are asymptomatic and vast majority are imported cases right now (http://2019ncov.chinacdc.cn/2019-nCoV/index.html). Therefore, the mortality rate in China should be closer to the real one. Of course, the mortality rate is closely related to the capacity of the local health care system. Hubei province was the most-affected area in China for the outbreak of COVID-19. At that time, local medical resources were extremely scarce; thus the mortality rate in Hubei reached 6.62% while that of the remaining regions of China is only 0.8%.

Different groups estimated that the basic reproductive number (R_0) for SARS-CoV-2 was approximately 2.68 [12], 2.2 [13], and even 3.8–6.47 [14–16]. The U.S. CDC has just adjusted R_0 to 5.7, meaning that each patient has spread infection to another 5.7 individuals on average. While the R_0 of SARS-CoV was estimated to be 2–5 [17], it seems that the infectivity of SARS-CoV-2 is stronger than that of SARS-CoV. SARS was the first new infectious disease identified in the twenty-first century. This acute, and often severe, respiratory illness originated in the Guangdong province of China in November 2002

^{*} Corresponding author. Key Laboratory of Medical Molecular Virology (MOE/NHC/CAMS), School of Basic Medical Sciences, Shanghai Medical College, Fudan University, Shanghai, China.

[18]. By the end of June 2003, there were 8450 cases and 810 deaths caused by SARS (www.cdc.gov/mmwr/mguide_sars.html). The overall fatality of SARS is about 10% in the general population, but approximately 50% in patients aged 65 years old and older [19]. Although the mortality rate of COVID-19 is not as high as that of SARS in 2002-2003, the number of confirmed cases has markedly surpassed that of SARS. In addition, the incubation period of COVID-19 ranged from 1 to 14 days, with an average of 10 days, and the patients in the incubation period or presymptomatic infected-individuals could potentially transmit the virus to uninfected people, which makes the infectivity of SARS-CoV-2 far exceeds that of SARS-CoV. SARS-CoV-2 is mainly transmitted through droplets and close contact, while aerosol is also a potential mode of transmission. Additionally, people of all ages are susceptible to the novel coronavirus, while older males with comorbidities are most vulnerable and more likely to develop severe and even fatal respiratory diseases [7,20]. The most thorough and strict control measures, including social distancing, limiting the flow of people and transportation, and canceling public activities, have taken effect in China. But the pandemic has spread to more than 200 countries and the best time window for controlling has been missed out. The SARS-CoV-2 seems highly possible to exist for a long time and the world is entering a period of normalized prevention and control. Currently, there is no specific drug for the virus. Although the drugs currently considered effective for COVID-19 are chloroquine and remdesivir, the latter one is a nucleotide analogue pro-drug that can inhibit RNA dependent RNA synthetase (RdRp) of virus [21], the existing data cannot fully prove the efficacy and safety of these drugs for treating SARS-CoV-2 infection. Therefore, it is urgent to develop an effective vaccine to prevent SARS-CoV-2 infection.

1. Biological characteristics of SARS-CoV-2

SARS-CoV-2 is an enveloped single-stranded positive-sense RNA virus. Generally, coronavirus falls into four genera- α , β , γ , δ on the basis of their phylogenetic relationships and genomic structures. α and β coronaviruses infect only mammals while the γ and δ coronaviruses mainly infect birds [22–25]. Among them, there are 7 species that can infect human beings, that is, common coronavirus including HCoV-229E and HCoV-NL63 that belong to α coronaviruses, HCoV-OC43 and CoV-HKU1 that belong to β coronaviruses, and three highly pathogenic viruses, SARS-CoV, Middle East respiratory syndrome coronavirus (MERS-CoV), and SARS-CoV-2 that all belong to β coronaviruses [17,26,27]. The novel coronavirus was initially isolated from three patients and the complete genome sequences were obtained. The three SARS-CoV-2 genomes clustered together within the sarbecovirus subgenus, which shows the typical β-coronavirus organization: a 5' untranslated region (UTR), replicase complex (orf1ab), S gene, E gene, M gene, N gene, 3' UTR, and several unidentified nonstructural open reading frames [1]. Chinese researchers have found that a gene sequence of coronavirus carried by Chinese rhinolophine is 96% identical with that of SARS-CoV-2, thus rhinolophine is considered as the natural host of SARS-CoV-2 [27,28]. Unfortunately, the intermediate host has not been determined yet, although it is speculated that pangolin may be the intermediate host. Based on the results of gene comparison, it was showed that SARS-CoV-2 was closely related (with 88% identity) to two bat-derived SARS-like coronaviruses, bat-SL-CoVZC45 and bat-SL-CoVZXC21, collected in 2018 in Zhoushan, eastern China, and SARS-CoV (about 79% identity), but was more distant from MERS-CoV (about 50% identity) [29]. Furthermore, according to the rich knowledge about SARS-CoV and the newly released sequence of SARS-CoV-2, the receptor-binding domain (RBD) of SARS-CoV-2, including its receptor-binding motif (RBM) that directly contacts angiotensin-converting enzyme 2 (ACE2), is similar to that of SARS-CoV, strongly suggesting that SARS-CoV-2 uses ACE2 as its receptor [30]. Currently, more structural and functional studies confirmed that ACE2 acted as the receptor for SARS-CoV-2 [31,32].

2. The strategies for previous development of SARS-CoV vaccines

The above analysis in gene level revealed that SARS-CoV-2 was very similar to SARS-CoV. Since the outbreak of SARS at the end of 2002, scientists have developed a variety of vaccines against SARS-CoV using their own vaccine platforms [33-37]. However, most vaccines are in preclinical studies, and only a few of them have been reported to under assessment in clinical trials (https://clinicaltrials. gov/ct2/show/nct03615911: https://clinicaltrials.gov/ct2/show/ nct0339578). Currently, no vaccine has been approved for the prevention of SARS. Since effective antiviral strategies to control SARS-CoV and SARS-CoV-2 infections are still lacking, vaccination is still regarded as the major approach for preventing potential reemergence of SARS-CoV, and more importantly, for preventing and controlling the current outbreak of SARS-CoV-2. Very recently, the Coalition for Epidemic Preparedness Innovations (CEPI) has reported that the global vaccine R&D effort in response to the COVID-19 pandemic is unprecedented in terms of scale and speed. As of 8 April 2020, the global COVID-19 vaccine R&D landscape includes 115 vaccine candidates, of which 5 advanced candidates including mRNA-1273 from Moderna, Ad5-nCoV from CanSino Biologicals, INO-4800 from Inovio, and LV-SMENP-DC and pathogenspecific aAPC from Shenzhen Geno-Immune Medical Institute have recently moved into clinical development, and the lead developers are mainly in the United States and China (Fig. 1) [38]. It sounds exciting, but we should always be cautious. Previously, a number of SARS vaccine candidates were suspended by the U.S. FDA because of the safety concern. Here, we summarized the advantages and disadvantages of the vaccine candidates against SARS-CoV to provide experience for the vaccine R&D of SARS-CoV-2 and avoid possible risks as far as possible.

2.1. Inactivated vaccines: advantages and disadvantages

It is known that most of the current influenza vaccines are inactivated vaccines, which plays a very important role in

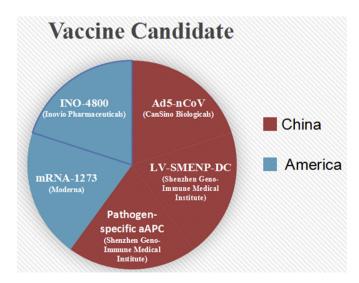


Fig. 1. Distribution of clinical-phage vaccine candidates for COVID-19 and their lead developers.

protecting people from influenza virus infection. Some approaches, such as formaldehyde, UV light, and β-propiolactone have been applied in the preparation of inactivated vaccines against SARS-CoV, which generally induced robust serumneutralizing antibody (Ab) response in different immunized animals [39-41]. For instance, He Y, et al. showed that high-titer antibodies elicited by the inactivated SARS-CoV in the immunized animals could recognize S protein, especially the receptorbinding domain (RBD) in the S1 subunit, and potently block SARS-CoV entry [42]. Some groups evaluated the safety of the inactivated vaccine for SARS-CoV and the results showed that all vaccines induced serum neutralizing antibody and significant reductions of SARS-CoV after viral challenge. However, challenge of mice given any of the inactivated vaccines led to occurrence of lung lesion based on th2-biased immunopathology with prominent eosinophil infiltration [43-45]. In addition, Fan Luo et al. intramuscularly immunized Chinese rhesus macaques with 2.5 µg betapropiolactone-inactivated whole SARS-CoV Z-1 vaccine, and boosted the immunization on day 7. Although this study revealed no exacerbation of infection in the immunized macaques following the challenge with SARS-CoV NS-1, the authors suggested that an antibody-dependent enhancement (ADE) assay should be performed in vitro to examine whether the immune sera induced by inactivated vaccine would mediate ADE in some cell lines such as K562, THP-1, and monkey's peripheral blood mononuclear cells [46]. Therefore, we should be cautious in using the inactivated SARS-CoV as a vaccine since it may cause harmful immune and/or inflammatory responses.

2.2. Live-attenuated vaccines: advantages and disadvantages

Historically and contemporarily, live-attenuated vaccines have been used successfully in controlling measles, mumps, rubella, polio, yellow fever, and chickenpox infections and outbreaks [47,48]. Some researchers constructed recombinant forms of the full-length SARS-CoV S protein or RBD of S protein that was encoded into the highly attenuated modified vaccinia virus Ankara (MVA) [49], recombinant adeno-associated virus (rAAV) [50], or an attenuated parainfluenza virus (BHPIV3), respectively [51,52]. BALB/c mice were inoculated intranasally or intramuscularly with MVA/S and neutralizing antibodies were induced. Moreover, MVA/ S administered by either route elicited protective immunity, as shown by reduced titers of SARS-CoV in the upper and lower respiratory tracts of the mice after viral challenge. Du L.et al. evaluated the immune protection of a rAAV encoding RBD vaccine in mouse model by intranasal inoculation, which induced strong mucosal immune responses and provided long-term protection against SARS-CoV infection [53]. Other groups reported that immunization of monkeys via the respiratory tract with BHPIV3/SARS-S induced the production of SARS-CoV-neutralizing serum antibodies. After the challenge with SARS-CoV, all monkeys inoculated with BHPIV3/ SARS-S did not shed any SARS-CoV while monkeys in control group shed SARS-CoV for 5-8 days [49,52]. Weingartl H et al. immunized ferrets with rMVA-S and the immunized ferrets developed a more rapid and vigorous neutralizing antibody response compared with control animals; however, after challenge with SARS-CoV, they also exhibited stronger inflammatory responses in liver tissue than control animals, suggesting that vaccination with rMVA-S is associated with enhanced hepatitis [54], which is not seen in the above infection model. It is suggested that we should not only use mice or monkeys as animal model, but also use ferrets as the infection model, because of its sensitivity to SARS-CoV. Thus, sometimes we do not observe the pathological damage caused by vaccine immunization, which may be that the animal model used is not representative enough.

Notably, live-attenuated vaccines are associated with the risk of reversion by either mutation- or recombination—driven processes, which can cause dangerous outbreaks in unvaccinated populations, including animals [48]. Several groups have tried to develop novel strategies in controlling fidelity and attenuating RNA virus pathogenesis. In the case of coronavirus, some mutants have been identified to prevent reversion to repaired virulence [55,56]. Rachel L. Graham et al. provided a proof that rewiring the transcription regulatory networks (TRNs) limited reversion in an live-attenuated coronavirus vaccine candidate which are effective against SARS-CoV [56]. Some SARS-CoV vaccine candidates were tested in clinical trials, showing that they could elicit antibody responses and are safe [57,58], although their long-term safety profiles have not been reported.

2.3. Vaccines based on full-length S protein: advantages and disadvantages

Envelope S protein is the most important surface protein of coronavirus (Fig. 2), which is related to the viral infectivity. The S protein mediates receptor binding and membrane fusion [59,60] and is crucial for determining host tropism and transmission capacity [61,62]. Generally, the S protein of coronavirus is functionally divided into the S1 subunit, responsible for receptor binding, and the S2 subunit, responsible for cell membrane fusion [63-66]. By aligning SARS-CoV-2 S protein sequence with those of SARS-CoV and several bat-SL-CoVs, we predicted that the cleavage site for generating S1 and S2 subunits is located at R694/S695 [67]. In addition, it has been reported that S protein has strong immunogenicity and can induce high titer neutralizing antibody [63]. RBD of SARS-CoV S1 is located in S318-510 and the key RBM is S425-494, of which R453 is critical for the complex formation [68]. Notably, N479 and T487 of the RBD are important for the high-affinity association of S protein with ACE2 [69-72]. Residues 442, 472 and 480 also contribute to receptor recognition and host range of SARS-CoV, although not as much as residues 479 and 487 [25,73,74]. Besides, a point mutation at R441 or D454 of the RBD of SARS-CoV disrupts the antigenic structure and binding activity of RBD to ACE2 [75,76]. By comparing the amino acid sequence of S protein between SARS-CoV-2 and SARS CoV, it was found that the similarities of primary sequence between SARS-CoV-2-S and SARS-CoV-S (isolated from human, civet or bat) are around 76%-78% for the whole protein, around 73%–76% for the RBD, and 50%–53% for the RBM [7,30].

In S1 subunit, SARS-CoV-2 and SARS-CoV shared around 50 conserved amino acids, and the three-dimensional structure of SARS-CoV-2 RBD was composed of a core and an external subdomain, which was more similar to that of SARS-CoV [29]. We aligned the sequence of S protein between SARS-CoV (NCBI Reference Sequence: NC_004718.3) and SARS-CoV-2 (NCBI Reference Sequence: NC_045512.2). As shown in Fig. 3A, the five key residues responsible for the binding of the SARS-CoV RBD (residues 442, 472, 479, 480 and 487; SARS-CoV numbering) to the ACE2 receptor were variable in the SARS-CoV-2 RBD (residues 455, 485, 493, 494 and 501; SARS-CoV-2 numbering). Although the five critical residues in SARS-CoV-2 RBM were different with that of SARS-CoV, residue Gln493 provide favorable interactions with human ACE2, consistent with SARS-CoV-2's capacity for human cell infection. Other critical residues in SARS-CoV-2 RBM (particularly Asn501) are compatible with, but not ideal for, binding human ACE2, suggesting that SARS-CoV-2 has acquired capacity for human-tohuman transmission. Phylogenetic analysis indicates that SARS-CoV-2 also potentially recognizes ACE2 from a diversity of animal species (except for mice and rats), implicating that these animal species may act as intermediate hosts or animal models for SARS-

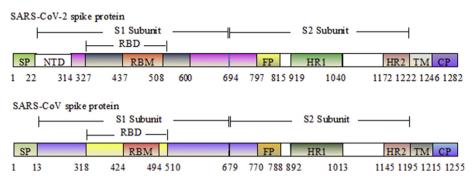


Fig. 2. Schematic of spike protein structure of SARS-CoV-2 and SARS-CoV. SP, signal peptide; NTD: N-terminal domain; RBD, receptor-binding domain; RBM, receptor-binding motif; FP, fusion peptide; HR, heptad repeat; TM, transmembrane domain; CP, cytoplasm domain.

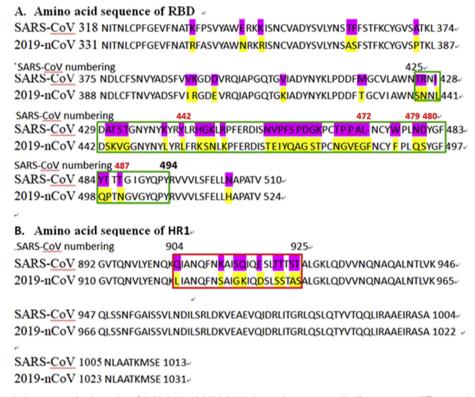


Fig. 3. Specific amino acid variations among the S proteins of SARS-CoV and SARS-CoV-2. Letters in magenta and yellow represent different amino acids in the corresponding sequences of SARS-CoV and SARS-CoV-2, respectively. A. Sequence alignment of SARS-CoV and SARS-CoV-2 RBDs. The red number indicates the core amino acids in RBD when it binds to receptor ACE2. The green frame is the amino acid sequence of RBM. B. Sequence alignment of SARS-CoV and SARS-CoV-2 HR1. The red frame is the location where the variable amino acid residues between SARS-CoV and SARS-CoV-2 HR1.

CoV-2 infections [30]. S2 subunit of coronavirus is divided into two domains, HR1 and HR2. We confirmed that SARS-CoV-2 HR1 and HR2 regions are able to interact with each other to form six helix bundle (6-HB), which brings the viral and cellular membranes in close proximity for fusion [64,77]. As shown in Fig. 3B, several amino acid residues in HR1 of SARS-CoV-2, which are concentrated from 922 to 943 were variable with SARS-CoV (residues 904–925), while HR2 is completely identical between SARS-CoV-2 and SARS-CoV (not shown), suggesting that SARS-CoV-2 S2, especially HR2 is a conserved target for pan-coronavirus vaccine. Together, these results suggest that S protein plays an important role in the infection and immunogenicity of coronavirus which promotes some research teams to prepare a vaccine based on full-length S protein of SARS-CoV-2.

A number of vaccine candidates based on the full-length S protein of SARS-CoV have been reported in succession and

generally could induce high level of immune response and potent protective immunity [35,78-80]. For example, the expression of full-length S protein and its trimer of urbani strain by recombinant baculovirus can induce adequate neutralizing antibody against human and palm civet SARS pseudoviruses that bears S proteins of homologous or heterologous SARS-CoV variants (such as Tor2, GD03T13, and SZ3 strains) in vaccinated mice [81]. These reports suggested that the full-length S protein is highly immunogenic and able to induce protection against SARS-CoV challenge. Besides, the neutralizing antibodies alone may be able to suppress virus proliferation. However, the risk in safety and final protective effect of the full-length S protein-based vaccine should not be underestimated. Some groups reported that although full-length S protein of SARS-CoV elicited a neutralizing antibody response, it also induced harmful immune responses including triggered infection of human B cells or other immune cells by SARS-CoV, and enhanced

SARS-CoV infection with anti-SARS-CoV-S immune-serum after challenge with homologous SARS-CoV [82-84]. Last year, Liu L. et al. identified that anti-S protein IgG, in productively infected lungs, caused severe acute lung injury by skewing inflammationresolving response in SARS-CoV macaque models. Subsequently, these antibodies were injected into healthy macaques that were then challenged with SARS-CoV. Interestingly, the macaques receiving the antibodies showed more severe lung injury than the control group, indicating that Antibody induced by SARS S protein vaccine candidate may cause antibody dependent enhancement (ADE) [85]. Thus, it is speculated that the S protein of SARS-CoV contains some epitopes, which may induce humoral or cellular immune responses that exacerbate the pathology in some coronavirus infections [86]. Protein Science, an American biotechnology company, once spent tens of millions to make a full-length S-protein vaccine of SARS-CoV. However, the vaccine development was suspended because of the safety problems in the immunized animals. Exactly, Liu G. et al. found that peptide S597-603 of S protein induced antibodies that enhanced infection both in vitro and in non-human primates [87]. Perhaps, S protein-based vaccines against SARS-CoV could be engineered to avoid ADE via elimination of the S597-603 epitope.

2.4. RBD-based subunit vaccine: advantages and disadvantages

Full length S protein has been truncated into RBD as a vaccine candidate. RBD of SARS-CoV-S contains major antigenic epitopes that can induce not only neutralizing antibodies but also CD8⁺ T cell-responses [43,63,88–91]. He Y.et al. identified the neutralizing epitopes on the RBD of SARS-CoV-S by a panel of 27 mAbs isolated from the immunized mice. Six structure-dependent epitopes and two adjacent linear epitopes were found, suggesting that the epitopes capable of inducing highly potent neutralizing Ab responses are mainly conformational epitopes [92]. In addition, the neutralizing epitopes in MERS-CoV RBD was confirmed to be mainly conformational as well [93,94]. The modifications in amino acid residues of MERS-CoV-S-RBD based on the structure design could improve its protection against MERS-CoV infection [94], enlightening that S-RBD or modified S-RBD of SARS-CoV-2 is promising for the development of vaccine against SARS-CoV-2.

We have compared the immunogenicity and immune protection of RBD vaccine with full-length S protein. The results suggested that the RBD, from either SARS-CoV or MERS-CoV, had induced Ab responses with higher titer, more potent neutralizing activity and provided longer-term protection in vivo against SARS-CoV infection compared with that induced by full-length S protein. More importantly, SARS-CoV RBD did not cause immune damage in the animal model while full-length S protein could do [51,95-97]. It is speculated that there are some super-antigens in the region beyond the RBD of the full-length S protein. In this way, after the full-length S protein enters the animal model, it will induce abundant nonspecific antibodies and immune cell activation, subsequently eliciting a large amount of cytokines, which will lead to severe immune damage, just like the pathogenic course of SARS-CoV. But why the antibody titer induced by full-length S protein is not as high as that induced by RBD. It is speculated that the antigen competition might result in a lower level of neutralizing antibody induced by fulllength S protein than that induced by RBD. Therefore, RBD-based subunit vaccine may be the ideal and safer alternative for the development of coronavirus vaccine.

Interestingly, some groups also found that the recombinant RBDs derived from the S protein of Tor2, GD03, and SZ3, the representative strains of human 2002–2003 and 2003–2004 SARS-CoV and palm civet SARS-CoV, respectively, induced high titers of cross-neutralizing Abs against pseudoviruses expressing S

proteins of Tor2, GD03, and SZ3 [98], suggesting that the major neutralizing epitopes of SARS-CoV are maintained in the process of cross species transmission, thus RBD-based vaccines could induce extensive protection against human and animal SARS-CoV variants. Probably, the RBD of S protein may serve as one of the most effective and safest vaccines for the prevention of SARS.

Since both SARS-CoV-2 and SARS-CoV recognize ACE2 on the cell surface as the receptor, can RBD-based SARS-CoV-specific vaccines cross protect individuals from SARS-CoV-2 infection? Unfortunately, Ying T. et al. showed that the most potent SARS-CoV-specific neutralizing antibodies (e.g., m396, CR3014) that target the ACE2 binding site of SARS-CoV failed to bind SARS-CoV-2 S protein, indicating that the different amino acids in RBD between SARS-CoV and SARS-CoV-2 has an important effect on the cross-reactivity of neutralizing antibodies [99].

2.5. Vaccines based on S2 subunit: advantages and disadvantages

SARS-CoV S2 subunit which is responsible for fusion between virus and target cell membranes is expected to be a vaccine candidate against SARS-CoV [66,100,101]. Several groups showed that S2 of SARS-CoV and/or MERS-CoV S protein could induce neutralizing antibodies although their neutralizing potency is generally lower than that of antibodies specific for RBD [102-104]. However, Guo et al. investigated the immune responses against the recombinant S2 fragment (residues 681-980) in BALB/c mice, and found that the S2 fragment could induce specific cellular immune response and a high level of total IgG but little neutralizing antibodies against infection by SARS-CoV [105]. Additionally, Zeng F et al. purified the total S1- and S2-specific IgG from mice immunized with recombinant S1 or S2 proteins, respectively, and found that anti-S1 and anti-S2 IgGs were able to abolish the binding between S protein and its cellular receptor(s), although anti-S1 IgG showed a significantly higher blocking efficiency [106]. As shown in Fig. 2B, the amino acid sequences of HR1 and HR2 are highly conserved and homologous between SARS-CoV and SARS-CoV-2. In fact, these domains are also highly conserved in other coronaviruses [104], thus the S2 subunit has potential to be used as a target for the development of pan-CoV vaccine against divergent virus strains. Altogether, these results suggested that the S2 domain of SARS-CoV S protein, as a vaccine candidate, may be able to prevent the infection of SARS-CoV.

SARS-CoV can also bind to host cells through alternative receptors, such as DC-SIGN (dendritic cell-specific intercellular adhesion molecule-3-grabbing non-integrin) and/or LSIGN (liver/ lymph node-SIGN) [107,108]. Seven asparagine-linked glycosylation sites in the S protein, including residues at positions 109, 118, 119, 158, 227, 589 and 699, are crucial for DC-SIGN-or L-SIGN-mediated virus entry [109]. Notably, a SARS-CoV-specific human monoclonal antibody, CR3022, could bind potently with SARS-CoV-2 RBD, however, the epitope of CR3022 does not overlap with the ACE2 binding site within SARS-CoV-2 RBD [99], suggesting that SARS-CoV-2 may also have other receptor binding sites except for ACE2, which should be noted in our vaccine design against SARS-CoV-2. Besides, optimization of sequences, components, or immunization routes, inclusion of appropriate adjuvants, or application of combinational immunization approaches are usually required for preparing an effective vaccine [110,111].

All these above results indicated that the modification of the SARS-CoV S protein, specifically, by retaining the conserved amino acids, especially the conserved sequence in the RBD region, and removing the possible epitopes that may cause immune damage may be a good strategy to prepare a broad-spectrum vaccine against SARS-CoV and SARS-CoV-related virus.

3. Implication for development of safe and broad-spectrum vaccines against SARS-CoV-2 and other human coronaviruses

Very recently, many research teams have launched the development of vaccines against SARS-CoV-2. The safety is a crucial issue that we need to be aware of when making vaccines. Meanwhile, there is another issue needs attention. The human and civet isolates of SARS-CoV nestle phylogenetically within the spectrum of SARS-CoV-like (SL-CoV) strains isolated from the bats, and the civets may have served as intermediate amplification hosts that enable SARS-CoV to cross species for animal-to-human transmission [70,112-115]. Additionally, RNA virus might well mutate in ways that would make previously effective vaccines useless. Thus, development of a safe and effective vaccine against both human and animal SARS-CoV strains is highly important for preventing future SARS outbreaks. On the other hand, given the high prevalence and wide distribution of coronaviruses, increasing human-animal interface activities, the large genetic diversity and frequent recombination of CoV genomes, novel coronaviruses are likely to emerge periodically in humans due to frequent crossspecies infections and occasional spillover events. A broadspectrum SARS vaccine that is effective against all the coronaviruses using ACE2 as their receptor can be applied immediately when we face the recurrent threat of coronavirus. Particularly, taken SARS-CoV-2 as an example, the virus might disappear before the vaccine is developed. Therefore, development of safe and broad-spectrum vaccines against SARS-CoV-2 and other SARS-CoVrelated virus is of great significance for protecting human from CoV infection. Based on the above analysis of the advantages and disadvantages of various vaccines against SARS-CoV, the modified S protein is expected to be a candidate for a cross protective vaccine against SARS-CoV and SARS-CoV-related virus. Notably, we have recently demonstrated that a vaccine based on RBD of S protein against SARS-CoV could effectively cross neutralize SARS-CoV-2. Moreover, several scientists have tried to design a vaccine against all β -coronavirus, especially these ACE2-associated ones.

In addition to antigens, adjuvants can enhance the immune response and reduce the amount of antigen required for each dose of vaccine, so that more doses of vaccine can be produced and provided to more people. The choice of adjuvants also affects the efficacy of respiratory virus vaccines and adjuvant has also been applied in the R&D of SARS-CoV-2 vaccine. Lung is a complex organ, which is rich in immune cells, such as macrophages. The pulmonary epithelial cells (AECs) also have the characteristics of some immune cells, such as secreting a large number of pulmonary surfactant A, D (PS-A, PS-D). The PS layer forms a strong barrier to prevent nanoparticles and hydrophilic molecules from accessing them, so as to protect the AECs from pathological damage [116]. Type I interferons (IFN-Is) are the chief immune mediators to protect the body from viral infections, and can be secreted by AECs as well as immune cells following viral infection. Therefore, the activation of stimulator of interferon genes (STING) in these two cell types can effectively enhance the production of IFN-I, and recapitulate the immune responses provoked by viral infection [117]. However, it is still a great challenge to delivery STNG agonists into the cytosol of AECs without destroying the integrity of PS layer. Recently, two research teams from China and the United States cooperated to ingeniously use PSbiomimetic liposomes (PS-GAMP) to encapsulated 2',3'-cyclic guanosine monophosphate-adenosine monophosphate (cGAMP), a natural and effective STING agonist [118,119]. Because of its resemblance to PS, PS-GAMP disguises "self" and escapes from immune surveillance following intranasal immunization. Together with SP-A and SP-D, PS-GAMP enters the alveolar macrophages (AMs), from which PS-GAMP fluxes into AECs through gap junctions between AMs and AECs. Subsequently, the STING pathway in AMs and AECs was activated without breaching PS and alveolar epithelial barriers. Through this mechanism, the adjuvant PS-GAMP combined with inactivated H1N1 vaccine generated broad-spectrum cross protection not only effectively against H1N1, but also against heterosubtypic H3N2, H5N1 and H7N9 virus infection within two days after a single immunization. This cross protection lasted for at least 6 months and induced the persistence of lung CD8⁺ TRM cells in vivo. It is also showed that PS-GAMP could enhance the recruitment and differentiation of CD11b⁺ dendritic cells (DC) and CD8⁺ T cells, as well as the humoral response [120]. Thus, PS-GAMP strategy is a promising "universal" mucosal adjuvant for developing broad-spectrum vaccines against betacoronavirus lineage B (SARS-CoV, SARS-CoV-2, SARSr-CoVs).

4. Conclusion and prospects

There was a significantly harmful immune response after immunizing animals with inactivated vaccine against SARS-CoV following challenge with the homotypic virus, although no obvious safety problem was observed in animal model after immunization. In addition, the inactivated vaccine against one serotype of dengue virus (DENV) can enhance individual infection with other serotypes [121]. These observations suggested that harmful response after immunization with inactivated vaccine against SARS-CoV-2 might also happen. Even if the inactivated SARS-CoV-2 vaccine does not elicit serious harmful immune response, it may enhance the infection of the mutated and/or another novel coronavirus by ADE. The subunit vaccine based on RBD can avoid ADE. However, RBM cannot serve as a universal vaccine candidate because of its large variation among different virus strains. The biggest advantage of mRNA and DNA vaccines or vector vaccines that is currently under clinical trials in the United States is that they do not need adjuvants. However, there are still potential safety risks of these mRNA or DNA vaccines. Therefore, the modified RBD combined with T cell epitopes and effective mucosal adjuvants will be the best choice for the preparation of a broad-spectrum and safe coronavirus vaccine. Of course, it should be noted that the choice of vaccine depends on the mortality rate of the disease. For infectious diseases with high mortality, such as EBOV, the effectiveness of vaccines is more important than safety. However, for low mortality disease (such as COVID-19), the safety of vaccines is more important than effectiveness. Additional safety concerns relate to effectiveness and safety against antigenic variants of SARS-CoV and for safety of vaccinated persons exposed to other coronaviruses.

In a word, a safe and effective vaccine for COVID-19 is urgently needed. However, regulators need to evaluate its safety through a series of virus strains and more than one animal model before allowing the COVID-19 vaccines to be used in humans. Simultaneously, regulators should also see strong preclinical evidence that vaccines can prevent infection and we should speed up the R&D progress on the premise of ensuring the safety and effectiveness of the vaccine [122]. In addition, the vaccine preparation and adjuvant selection for SARS-CoV-2 in the future should attach great importance to its broad-spectrum cross protection, so that if there is similar virus diseases emerged in humans or animals, the vaccine can be applied immediately to exert its protective effect.

Declaration of Competing Interest

The authors declare no conflict of interest.

Acknowledgments

This work was supported by grants from the National Natural Science Foundation of China (81971474), and Scientific and

Technological Research Projects of Colleges and Universities in Hebei Province (ZD2018001).

References

- [1] Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in China, 2019. N Engl J Med 2020;382:727–33.
- [2] Zhang N, Wang L, Deng X, Liang R, Su M, He C, et al. Recent advances in the detection of respiratory virus infection in humans. J Med Virol 2020;92: 408–17.
- [3] Jiang S, Shi Z-L. The first disease X is caused by a highly transmissible acute respiratory syndrome coronavirus. Virol Sin 2020. https://doi.org/10.1007/ s12250-020-00206-5 [online ahead of print].
- [4] Yu F, Du L, Ojcius DM, Pan C, Jiang S. Measures for diagnosing and treating infections by a novel coronavirus responsible for a pneumonia outbreak originating in Wuhan, China. Microb Infect 2020;22:74–9.
- [5] WHO. Coronavirus. Available at: https://www.who.int/health-topics/coronavirus. [Accessed 20 January 2020].
- [6] Coronaviridae Study Group of the International Committee on Taxonomy of Viruses. The species Severe acute respiratory syndromerelated coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2. Nat Microbiol 2020;5: 536–44.
- [7] Chan JF-W, Yuan S, Kok K-H, To KK-W, Chu H, Yang J, et al. A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating personto-person transmission: a study of a family cluster. Lancet 2020;395:514–23.
- [8] Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 2020;395: 497–506.
- [9] Messonnier NE. CDC advises travelers to avoid all nonessential travel to China: media statement. Available at: https://www.cdc.gov/media/releases/ 2020/s0128-travelers-avoid-china.html. [Accessed 28 January 2020].
- [10] Phan LT, Nguyen TV, Luong QC, Nguyen TV, Nguyen HT, Le HQ, et al. Importation and human-to-human transmission of a novel coronavirus in Vietnam. N Engl J Med 2020;382:872–4.
- [11] Jiang S, Xia S, Ying T, Lu L. A novel coronavirus (2019-nCoV) causing pneumonia-associated respiratory syndrome. Cell Mol Immunol 2020;17: 554
- [12] Wu JT, Leung K, Leung GM. Nowcasting and forecasting the potential domestic and international spread of the 2019-nCoV outbreak originating in Wuhan, China: a modelling study. Lancet 2020;395:689–97.
- [13] Li Q, Guan X, Wu P, Wang X, Zhou L, Tong Y, et al. Early transmission dynamics in Wuhan, China, of novel coronavirus—infected pneumonia. N Engl J Med 2020;382:1199–207.
- [14] Liu T, Hu J, Xiao J, He G, Kang M, Rong Z, et al. Time-varying transmission dynamics of Novel Coronavirus Pneumonia in China. bioRxiv 2020. https:// doi.org/10.1101/2020.01.25.919787 [online ahead of print].
- [15] Read JM, Bridgen JR, Cummings DA, Ho A, Jewell CP. Novel coronavirus 2019nCoV: early estimation of epidemiological parameters and epidemic predictions. medRxiv 2020. https://doi.org/10.1101/2020.01.23.20018549 [online ahead of print].
- [16] Shen M, Peng Z, Xiao Y, Zhang L. Modelling the epidemic trend of the 2019 novel coronavirus outbreak in China. bioRxiv 2020. https://doi.org/10.1101/ 2020.01.23.916726.
- [17] Bauch CT, Lloyd-Smith JO, Coffee MP, Galvani AP. Dynamically modeling SARS and other newly emerging respiratory illnesses: past, present, and future. Epidemiology 2005;16:791–801.
- [18] Zhong N, Zheng B, Li Y, Poon L, Xie Z, Chan K, et al. Epidemiology and cause of severe acute respiratory syndrome (SARS) in Guangdong, People's Republic of China, in February, 2003. Lancet 2003;362:1353–8.
- [19] Donnelly CA, Ghani AC, Leung GM, Hedley AJ, Fraser C, Riley S, et al. Epidemiological determinants of spread of causal agent of severe acute respiratory syndrome in Hong Kong. Lancet 2003;361:1761–6.
- [20] Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. Lancet 2020;395:507–13.
- [21] Holshue ML, DeBolt C, Lindquist S, Lofy KH, Wiesman J, Bruce H, et al. First case of 2019 novel coronavirus in the United States. N Engl J Med 2020;382: 929–36.
- [22] Woo PC, Lau SK, Lam CS, Lau CC, Tsang AK, Lau JH, et al. Discovery of seven novel Mammalian and avian coronaviruses in the genus deltacoronavirus supports bat coronaviruses as the gene source of alphacoronavirus and betacoronavirus and avian coronaviruses as the gene source of gammacoronavirus and deltacoronavirus. J Virol 2012;86:3995—4008.
- [23] Su S, Wong G, Shi W, Liu J, Lai AC, Zhou J, et al. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. Trends Microbiol 2016;24:490–502.
- [24] Forni D, Cagliani R, Clerici M, Sironi M. Molecular evolution of human coronavirus genomes. Trends Microbiol 2017;25:35–48.
- [25] Cui J, Li F, Shi Z-L. Origin and evolution of pathogenic coronaviruses. Nat Rev Microbiol 2019;17:181–92.
- [26] Benvenuto D, Giovannetti M, Ciccozzi A, Spoto S, Angeletti S, Ciccozzi M. The 2019-new coronavirus epidemic: evidence for virus evolution. J Med Virol 2020;92:455—9.

- [27] Zhou P, Yang X-L, Wang X-G, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 2020;579:270—3.
- [28] Zhou P, Yang X-L, Wang X-G, Hu B, Zhang L, Zhang W, et al. Discovery of a novel coronavirus associated with the recent pneumonia outbreak in humans and its potential bat origin. bioRxiv 2020. https://doi.org/10.1101/ 2020.01.22.914952.
- [29] Lu R, Zhao X, Li J, Niu P, Yang B, Wu H, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. Lancet 2020;395:565—74.
- [30] Wan Y, Shang J, Graham R, Baric RS, Li F. Receptor recognition by novel coronavirus from Wuhan: an analysis based on decade-long structural studies of SARS. J Virol 2020;94. e00127-20.
 [31] Wang Q, Zhang Y, Wu L, Niu S, Song C, Zhang Z, et al. Structural and func-
- [31] Wang Q, Zhang Y, Wu L, Niu S, Song C, Zhang Z, et al. Structural and functional basis of SARS-CoV-2 entry by using human ACE2. Cell 2020;181:1–11. https://doi.org/10.1016/j.cell.2020.03.045.
- [32] Monteil V, Kwon H, Prado P, Hagelkruys A, Wimmer RA, Stahl M, et al. Inhibition of SARS-CoV-2 infections in engineered human tissues using clinical-grade soluble human ACE2. Cell 2020;181:1–9. https://doi.org/10.1016/j.cell.2020.04.004.
- [33] Jiang S, He Y, Liu S. SARS vaccine development. Emerg Infect Dis 2005;11: 1016–20.
- [34] Zeng F, Chow KYC, Hon CC, Law KM, Yip CW, Chan KH, et al. Characterization of humoral responses in mice immunized with plasmid DNAs encoding SARS-CoV spike gene fragments. Biochem Biophys Res Commun 2004;315: 1134—9.
- [35] Du L, He Y, Zhou Y, Liu S, Zheng B, Jiang S. The spike protein of SARS-CoV a target for vaccine and therapeutic development. Nat Rev Microbiol 2009;7: 226–36.
- [36] Pimentel TAPF, Yan Z, Jeffers SA, Holmes KV, Hodges RS, Burkhard P. Peptide nanoparticles as novel immunogens: design and analysis of a prototypic Severe Acute Respiratory Syndrome Vaccine. Chem Biol Drug Des 2009;73: 53–61.
- [37] Du L, He Y, Jiang S, Zheng B. Development of subunit vaccines against severe acute respiratory syndrome. Drugs Today (Barc). 2008;44:63–73.
- [38] Thanh Le T, Andreadakis Z, Kumar A, Gomez Roman R, Tollefsen S, Saville M, et al. The COVID-19 vaccine development landscape. Nat Rev Drug Discov 2020:19:305–6
- [39] Xiong S, Wang Y-F, Zhang M-Y, Liu X-J, Zhang C-H, Liu S-S, et al. Immunogenicity of SARS inactivated vaccine in BALB/c mice. Immunol Lett 2004;95: 139–43.
- [40] Takasuka N, Fujii H, Takahashi Y, Kasai M, Morikawa S, Itamura S, et al. A subcutaneously injected UV-inactivated SARS coronavirus vaccine elicits systemic humoral immunity in mice. Int Immunol 2004;16:1423–30.
- [41] Tang L, Zhu Q, Qin E, Yu M, Ding Z, Shi H, et al. Inactivated SARS-CoV vaccine prepared from whole virus induces a high level of neutralizing antibodies in BALB/c mice. DNA Cell Biol 2004;23:391–4.
- [42] He Y, Li J, Du L, Yan X, Hu G, Zhou Y, et al. Identification and characterization of novel neutralizing epitopes in the receptor-binding domain of SARS-CoV spike protein: revealing the critical antigenic determinants in inactivated SARS-CoV vaccine. Vaccine 2006;24:5498–508.
- [43] He Y, Zhou Y, Siddiqui P, Jiang S. Inactivated SARS-CoV vaccine elicits high titers of spike protein-specific antibodies that block receptor binding and virus entry. Biochem Biophys Res Commun 2004;325:445–52.
- [44] Tseng C-T, Sbrana E, Iwata-Yoshikawa N, Newman PC, Garron T, Atmar RL, et al. Immunization with SARS coronavirus vaccines leads to pulmonary immunopathology on challenge with the SARS virus. PloS One 2012;7: e35421.
- [45] Wang D, Lu J. Glycan arrays lead to the discovery of autoimmunogenic activity of SARS-CoV. Physiol Genom 2004;18:245–8.
- [46] Luo F, Liao FL, Wang H, Tang HB, Hou W. Evaluation of antibody-dependent enhancement of SARS-CoV infection in rhesus macaques immunized with an inactivated SARS-CoV vaccine. Virol Sin 2018;33:201–4.
- [47] Lauring AS, Jones JO, Andino R. Rationalizing the development of live attenuated virus vaccines. Nat Biotechnol 2010;28:573–9.
- [48] Vignuzzi M, Wendt E, Andino R. Engineering attenuated virus vaccines by controlling replication fidelity. Nat Med 2008;14:154–61.
- [49] Bisht H, Roberts A, Vogel L, Bukreyev A, Collins PL, Murphy BR, et al. Severe acute respiratory syndrome coronavirus spike protein expressed by attenuated vaccinia virus protectively immunizes mice. Proc Natl Acad Sci Unit States Am 2004;101:6641–6.
- [50] Liu RY, Wu L-Z, Huang B-J, Huang J-L, Zhang Y-L, Ke M-L, et al. Adenoviral expression of a truncated S1 subunit of SARS-CoV spike protein results in specific humoral immune responses against SARS-CoV in rats. Virus Res 2005;112:24–31.
- [51] Du L, He Y, Wang Y, Zhang H, Ma S, Wong CK, et al. Recombinant adenoassociated virus expressing the receptor-binding domain of severe acute respiratory syndrome coronavirus S protein elicits neutralizing antibodies: implication for developing SARS vaccines. Virology 2006;353:6–16.
- [52] Bukreyev A, Lamirande EW, Buchholz UJ, Vogel LN, Elkins WR, St Claire M, et al. Mucosal immunisation of African green monkeys (Cercopithecus aethiops) with an attenuated parainfluenza virus expressing the SARS coronavirus spike protein for the prevention of SARS. Lancet 2004;363: 2122–7.

- [53] Du L, Zhao G, Lin Y, Sui H, Chan C, Ma S, et al. Intranasal vaccination of recombinant adeno-associated virus encoding receptor-binding domain of severe acute respiratory syndrome coronavirus (SARS-CoV) spike protein induces strong mucosal immune responses and provides long-term protection against SARS-CoV infection. J Immunol 2008;180:948-56.
- [54] Weingartl H, Czub M, Czub S, Neufeld J, Marszal P, Gren J, et al. Immunization with modified vaccinia virus Ankara-based recombinant vaccine against severe acute respiratory syndrome is associated with enhanced hepatitis in ferrets. J Virol 2004;78:12672–6.
- [55] Graham RL, Becker MM, Eckerle LD, Bolles M, Denison MR, Baric RS. A live, impaired-fidelity coronavirus vaccine protects in an aged, immunocompromised mouse model of lethal disease. Nat Med 2012;18:1820–6.
- [56] Graham RL, Deming DJ, Deming ME, Yount BL, Baric RS. Evaluation of a recombination-resistant coronavirus as a broadly applicable, rapidly implementable vaccine platform. Commun Biol 2018;1:1–10.
- [57] Lin JT, Zhang JS, Su N, Xu JG, Dongs X-P. Safety and immunogenicity from a Phase I trial of inactivated severe acute respiratory syndrome coronavirus vaccine. Antivir Ther 2007:12:1107—13.
- [58] Martin JE, Louder MK, Holman LSA, Gordon IJ, Enama ME, Larkin BD, et al. A SARS DNA vaccine induces neutralizing antibody and cellular immune responses in healthy adults in a Phase I clinical trial. Vaccine 2008;26: 6338–43
- [59] Li F. Structure, function, and evolution of coronavirus spike proteins. Annu Rev Virol 2016;3:237–61.
- [60] Xia S, Yan L, Xu W, Agrawal AS, Algaissi A, Tseng CK, et al. A pan-coronavirus fusion inhibitor targeting the HR1 domain of human coronavirus spike. Sci Adv 2019;5:eaav4580.
- [61] Lu G, Wang Q, Gao GF. Bat-to-human: spike features determining 'host jump'of coronaviruses SARS-CoV, MERS-CoV, and beyond. Trends Microbiol 2015;23:468-78.
- [62] Wang Q, Wong G, Lu G, Yan J, Gao GF. MERS-CoV spike protein: targets for vaccines and therapeutics. Antivir Res 2016;133:165–77.
- [63] He Y, Zhou Y, Liu S, Kou Z, Li W, Farzan M, et al. Receptor-binding domain of SARS-CoV spike protein induces highly potent neutralizing antibodies: implication for developing subunit vaccine. Biochem Biophys Res Commun 2004;324:773—81.
- [64] Xia S, Zhu Y, Liu M, Lan Q, Xu W, Wu Y, et al. Fusion mechanism of 2019nCoV and fusion inhibitors targeting HR1 domain in spike protein. Cell Mol Immunol 2020:1—3.
- [65] Taguchi F, Shimazaki YK. Functional analysis of an epitope in the S2 subunit of the murine coronavirus spike protein: involvement in fusion activity. J Gen Virol 2000;81:2867—71.
- [66] Deng Y, Liu J, Zheng Q, Yong W, Lu M. Structures and polymorphic interactions of two heptad-repeat regions of the SARS virus S2 protein. Structure 2006;14:889–99.
- [67] Jiang S, Du L, Shi Z. An emerging coronavirus causing pneumonia outbreak in Wuhan, China: calling for developing therapeutic and prophylactic strategies. Emerg Microb Infect 2020;9:275–7.
- [68] Zhang Y, Zheng N, Hao P, Cao Y, Zhong Y. A molecular docking model of SARS-CoV S1 protein in complex with its receptor, human ACE2. Comput Biol Chem 2005;29:254–7.
- [69] Li W, Zhang C, Sui J, Kuhn JH, Moore MJ, Luo S, et al. Receptor and viral determinants of SARS-coronavirus adaptation to human ACE2. EMBO J 2005;24:1634–43.
- [70] Song H-D, Tu C-C, Zhang G-W, Wang S-Y, Zheng K, Lei L-C, et al. Cross-host evolution of severe acute respiratory syndrome coronavirus in palm civet and human. Proc Natl Acad Sci Unit States Am 2005;102:2430–5.
- [71] Qu X-X, Hao P, Song X-J, Jiang S-M, Liu Y-X, Wang P-G, et al. Identification of two critical amino acid residues of the severe acute respiratory syndrome coronavirus spike protein for its variation in zoonotic tropism transition via a double substitution strategy. J Biol Chem 2005;280:29588–95.
- [72] Li F. Structural analysis of major species barriers between humans and palm civets for severe acute respiratory syndrome coronavirus infections. J Virol 2008;82:6984—91.
- [73] Li F. Receptor recognition and cross-species infections of SARS coronavirus. Antivir Res 2013;100:246–54.
- [74] Wu K, Peng G, Wilken M, Geraghty RJ, Li F. Mechanisms of host receptor adaptation by severe acute respiratory syndrome coronavirus. J Biol Chem 2012;287:8904–11.
- [75] Wong SK, Li W, Moore MJ, Choe H, Farzan M. A 193-amino acid fragment of the SARS coronavirus S protein efficiently binds angiotensin-converting enzyme 2. J Biol Chem 2004;279:3197—201.
- [76] He Y, Li J, Jiang S. A single amino acid substitution (R441A) in the receptorbinding domain of SARS coronavirus spike protein disrupts the antigenic structure and binding activity. Biochem Biophys Res Commun 2006;344: 106–13.
- [77] Xia S, Liu M, Wang C, Xu W, Lan Q, Feng S, et al. Inhibition of SARS-CoV-2 (previously 2019-nCoV) infection by a highly potent pan-coronavirus fusion inhibitor targeting its spike protein that harbors a high capacity to mediate membrane fusion. Cell Res 2020;30:343-55.
- [78] Kam YW, Kien F, Roberts A, Cheung YC, Lamirande EW, Vogel L, et al. Antibodies against trimeric S glycoprotein protect hamsters against SARS-CoV challenge despite their capacity to mediate FcγRII-dependent entry into B cells in vitro. Vaccine 2007;25:729–40.

- [79] Du L, Zhao G, Chan CC, Li L, He Y, Zhou Y, et al. A 219-mer CHO-expressing receptor-binding domain of SARS-CoV S protein induces potent immune responses and protective immunity. Viral Immunol 2010;23:211–9.
- [80] Du L, Zhao G, Chan CCS, Sun S, Chen M, Liu Z, et al. Recombinant receptorbinding domain of SARS-CoV spike protein expressed in mammalian, insect and E. coli cells elicits potent neutralizing antibody and protective immunity. Virology 2009;393:144–50.
- [81] He Y, Li J, Heck S, Lustigman S, Jiang S. Antigenic and immunogenic characterization of recombinant baculovirus-expressed severe acute respiratory syndrome coronavirus spike protein: implication for vaccine design. J Virol 2006;80:5757–67.
- [82] Jaume M, Yip MS, Cheung CY, Leung HL, Li PH, Kien F, et al. Anti-severe Acute Respiratory Syndrome Coronavirus spike antibodies trigger infection of human immune cells via a pH- and cysteine protease-Independent FcγR pathway. J Virol 2011;85:10582–97.
- [83] Wang SF, Tseng S-P, Yen C-H, Yang J-Y, Tsao C-H, Shen C-W, et al. Antibody-dependent SARS coronavirus infection is mediated by antibodies against spike proteins. Biochem Biophys Res Commun 2014;451:208–14.
- [84] Jaume M, Yip M, Kam Y, Cheung C, Kien F, Roberts A, et al. SARS CoV subunit vaccine: antibody-mediated neutralisation and enhancement. Hong Kong Med J 2012;18(Suppl 2):31–6.
- [85] Liu L, Wei Q, Lin Q, Fang J, Wang H, Kwok H, et al. Anti-spike IgG causes severe acute lung injury by skewing macrophage responses during acute SARS-CoV infection. JCI Insight 2019;4:e123158.
- [86] Wu GF, Dandekar AA, Pewe L, Perlman S. The role of CD4 and CD8 T cells in MHV-IHM-induced demyelination. Adv Exp Med Biol 2001:494:341–7.
- MHV-JHM-induced demyelination. Adv Exp Med Biol 2001;494:341–7.

 [87] Wang Q, Zhang L, Kuwahara K, Li L, Liu Z, Li T, et al. Immunodominant SARS coronavirus epitopes in humans elicited both enhancing and neutralizing effects on infection in non-human primates. ACS Infect Dis 2016;2:361–76.
- [88] Chen Z, Zhang L, Qin C, Ba L, Christopher EY, Zhang F, et al. Recombinant modified vaccinia virus Ankara expressing the spike glycoprotein of severe acute respiratory syndrome coronavirus induces protective neutralizing antibodies primarily targeting the receptor binding region. J Virol 2005;79: 2678–88
- [89] He Y, Zhu Q, Liu S, Zhou Y, Yang B, Li J, et al. Identification of a critical neutralization determinant of severe acute respiratory syndrome (SARS)associated coronavirus: importance for designing SARS vaccines. Virology 2005;334:74–82.
- [90] Zhi Y, Kobinger GP, Jordan H, Suchma K, Weiss SR, Shen H, et al. Identification of murine CD8 T cell epitopes in codon-optimized SARS-associated coronavirus spike protein. Virology 2005;335:34–45.
- [91] Du L, Zhao G, Li L, He Y, Zhou Y, Zheng B-J, et al. Antigenicity and immunogenicity of SARS-CoV S protein receptor-binding domain stably expressed in CHO cells. Biochem Biophys Res Commun 2009;384:486—90.
- [92] He Y, Lu H, Siddiqui P, Zhou Y, Jiang S. Receptor-binding domain of severe acute respiratory syndrome coronavirus spike protein contains multiple conformation-dependent epitopes that induce highly potent neutralizing antibodies. J Immunol 2005;174:4908–15.
- [93] Du L, Yang Y, Zhou Y, Lu L, Li F, Jiang S. MERS-CoV spike protein: a key target for antivirals. Expert Opin Ther Targets 2017;21:131—43.
- [94] Zhou Y, Yang Y, Huang J, Jiang S, Du L. Advances in MERS-CoV vaccines and therapeutics based on the receptor-binding domain. Viruses 2019;11:60.
- [95] Du L, Zhao G, He Y, Guo Y, Zheng B-J, Jiang S, et al. Receptor-binding domain of SARS-CoV spike protein induces long-term protective immunity in an animal model. Vaccine 2007;25:2832–8.
- [96] Du L, Zhao G, Lin Y, Chan C, He Y, Jiang S, et al. Priming with rAAV encoding RBD of SARS-CoV S protein and boosting with RBD-specific peptides for T cell epitopes elevated humoral and cellular immune responses against SARS-CoV infection. Vaccine 2008;26:1644–51.
- [97] Ma C, Wang L, Tao X, Zhang N, Yang Y, Tseng C-TK, et al. Searching for an ideal vaccine candidate among different MERS coronavirus receptor-binding fragments—the importance of immunofocusing in subunit vaccine design. Vaccine 2014;32:6170–6.
- [98] He Y, Li J, Li W, Lustigman S, Farzan M, Jiang S. Cross-neutralization of human and palm civet severe acute respiratory syndrome coronaviruses by antibodies targeting the receptor-binding domain of spike protein. J Immunol 2006;176:6085–92.
- [99] Tian X, Li C, Huang A, Xia S, Lu S, Shi Z, et al. Potent binding of 2019 novel coronavirus spike protein by a SARS coronavirus-specific human monoclonal antibody. Emerg Microb Infect 2020;9:382–5.
- [100] Basso LGM, Vicente EF, Crusca E, Cilli EM, Costa-Filho AJ. SARS-CoV fusion peptides induce membrane surface ordering and curvature. Sci Rep 2016;6: 37131.
- [101] Yan Z, Holmes KV, Hodges RS. Expression and characterization of recombinant S2 subunit of SARS-coronavirus S fusion protein. Adv Exp Med Biol 2009;611:153—4.
- [102] Wang L, Shi W, Joyce MG, Modjarrad K, Zhang Y, Leung K, et al. Evaluation of candidate vaccine approaches for MERS-CoV. Nat Commun 2015;6:1–11.
- [103] Chen Y, Lu S, Jia H, Deng Y, Zhou J, Huang B, et al. A novel neutralizing monoclonal antibody targeting the N-terminal domain of the MERS-CoV spike protein. Emerg Microb Infect 2017;6:e37.
- [104] Elshabrawy HA, Coughlin MM, Baker SC, Prabhakar BS. Human monoclonal antibodies against highly conserved HR1 and HR2 domains of the SARS-CoV spike protein are more broadly neutralizing. PloS One 2012;7:e50366.

- [105] Guo Y, Sun S, Wang K, Zhang S, Zhu W, Chen Z. Elicitation of immunity in mice after immunization with the S2 subunit of the severe acute respiratory syndrome coronavirus. DNA Cell Biol 2005;24:510-5.
- [106] Zeng F, Hon CC, Yip CW, Law KM, Yeung YS, Chan KH, et al. Quantitative comparison of the efficiency of antibodies against S1 and S2 subunit of SARS coronavirus spike protein in virus neutralization and blocking of receptor binding: implications for the functional roles of S2 subunit. FEBS Lett 2006;580:5612–20.
- [107] Jeffers SA, Tusell SM, Gillim-Ross L, Hemmila EM, Achenbach JE, Babcock GJ, et al. CD209L (L-SIGN) is a receptor for severe acute respiratory syndrome coronavirus. Proc Natl Acad Sci Unit States Am 2004;101:15748–53.
- [108] Yang Z-Y, Huang Y, Ganesh L, Leung K, Kong W-P, Schwartz O, et al. pH-dependent entry of severe acute respiratory syndrome coronavirus is mediated by the spike glycoprotein and enhanced by dendritic cell transfer through DC-SIGN. J Virol 2004;78:5642—50.
- [109] Han DP, Lohani M, Cho MW. Specific asparagine-linked glycosylation sites are critical for DC-SIGN-and L-SIGN-mediated severe acute respiratory syndrome coronavirus entry. J Virol 2007;81:12029—39.
- [110] Zhang N, Jiang S, Du L. Current advancements and potential strategies in the development of MERS-CoV vaccines. Expert Rev Vaccines 2014;13:761–74.
- [111] Zhang N, Channappanavar R, Ma C, Wang L, Tang J, Garron T, et al. Identification of an ideal adjuvant for receptor-binding domain-based subunit vaccines against Middle East respiratory syndrome coronavirus. Cell Mol Immunol 2016;13:180–90.
- [112] Cyranoski D, Abbott A. Virus detectives seek source of SARS in China's wild animals. Nature 2003:423:467.

- [113] Guan Y, Zheng B, He Y, Liu X, Zhuang Z, Cheung C, et al. Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. Science 2003;302:276–8.
- [114] Li W, Shi Z, Yu M, Ren W, Smith C, Epstein JH, et al. Bats are natural reservoirs of SARS-like coronaviruses. Science 2005;310:676–9.
- [115] Lau SK, Woo PC, Li KS, Huang Y, Tsoi H-W, Wong BH, et al. Severe acute respiratory syndrome coronavirus-like virus in Chinese horseshoe bats. Proc Natl Acad Sci Unit States Am 2005;102:14040—5.
- [116] Whitsett JA, Wert SE, Weaver TE. Alveolar surfactant homeostasis and the pathogenesis of pulmonary disease. Annu Rev Med 2010;61:105–19.
- [117] Ablasser A, Goldeck M, Cavlar T, Deimling T, Witte G, Röhl I, et al. cGAS produces a 2'-5'-linked cyclic dinucleotide second messenger that activates STING. Nature 2013;498:380—4.
- [118] Wu J, Sun L, Chen X, Du F, Shi H, Chen C, et al. Cyclic GMP-AMP is an endogenous second messenger in innate immune signaling by cytosolic DNA. Science 2013:339:826–30.
- [119] Li X-D, Wu J, Gao D, Wang H, Sun L, Chen ZJ. Pivotal roles of cGAS-cGAMP signaling in antiviral defense and immune adjuvant effects. Science 2013;341:1390—4.
- [120] Wang J, Li P, Yu Y, Fu Y, Jiang S, Lu L, et al. Pulmonary surfactant-biomimetic nanoparticles potentiate heterosubtypic influenza immunity. Science 2020:367. eaau0810.
- [121] Whitehead SS, Blaney JE, Durbin AP, Murphy BR. Prospects for a dengue virus vaccine. Nat Rev Microbiol 2007;5:518—28.
- [122] Jiang S. Don't rush to deploy COVID-19 vaccines and drugs without sufficient safety guarantees. Nature 2020;579:321.