

## Research Article

# Enhanced Antitumor Activity with Combining Effect of mTOR Inhibition and Microtubule Stabilization in Hepatocellular Carcinoma

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Mammalian target of rapamycin (mTOR) and the microtubules are shown to be potential targets for treating hepatocellular carcinoma (HCC). PI3K/Akt/mTOR activation is associated with resistance to microtubule inhibitors. Here, we evaluated the antitumor activity by cotargeting of the mTOR (using allosteric mTOR inhibitor everolimus) and the microtubules (using novel microtubule-stabilizing agent patupilone) in HCC models. *In vitro* studies showed that either targeting mTOR signaling with everolimus or targeting microtubules with patupilone was able to suppress HCC cell growth in a dose-dependent manner. Cotargeting of the mTOR (by everolimus) and the microtubules (by patupilone, at low nM) resulted in enhanced growth inhibition in HCC cells (achieving maximal growth inhibition of 60–87%), demonstrating potent antitumor activity of this combination. *In vivo* studies showed that everolimus treatment alone for two weeks was able to inhibit the growth of Hep3B xenografts. Strikingly, the everolimus/patupilone combination induced a more significant antitumor activity. Mechanistic study demonstrated that this enhanced antitumor effect was accompanied by marked cell apoptosis induction and antiangiogenic activity, which were more significant than single-agent treatments. Our findings demonstrated that the everolimus/patupilone combination, which had potent antitumor activity, was a potential therapeutic strategy for HCC.

## 1. Introduction

Hepatocellular carcinoma (HCC) is the third most common cause of cancer-related deaths worldwide [1, 2]. Surgical resection and liver transplantation are the two mainstays of curative treatment for HCC, but can only be applied to the early stage of HCC [3, 4]. The majority of patients with HCC are not amenable to, or eventually failed, locoregional therapies and have to be considered for systemic treatment. Although sorafenib (a multikinase inhibitor of VEGFR, PDGFR, and Raf) has been approved for the treatment of HCC as the first-line therapy for unresectable HCC, the outlook of patients with advanced disease remains dismal

[5, 6]. These reasons exemplify the need to design more effective therapeutic strategies.

Everolimus (RAD001, Afinitor), a rapamycin analogue, is an oral mammalian target of rapamycin (mTOR) inhibitor. mTOR is a key effector in the PI3K/Akt/mTOR pathway and it plays a critical role in regulating cell proliferation, survival, and angiogenesis [7]. Everolimus has been approved for the treatment of papillary renal carcinoma, pancreatic neuroendocrine tumor, some types of breast cancer, and subependymal giant cell astrocytoma associated with tuberous sclerosis [8–11]. In HCC, a phase I/II study of everolimus has been conducted in patients with advanced HCC and

antitumor activity was observed, with time to progression of 3.9 months and disease control rate of 44% [12].

However, to enhance the efficacy of everolimus [13, 14], evaluation for potential synergism with other classes of anticancer agents is warranted. Recent gene expression profiling studies suggested microtubules to be an important target for therapeutic intervention in HCC [15–17]. Furthermore, several studies demonstrated the involvement of mTOR pathway in resistance to microtubule-targeting chemotherapeutic agents [18, 19]. This led us to hypothesize that the cotargeting of mTOR and microtubules would be a potent therapeutic strategy for HCC. Indeed, in a previous study, we showed that combination of mTOR inhibitor temsirolimus and microtubule-targeting agent vinblastine had marked antitumor effect in HCC both *in vitro* and *in vivo* [20].

Patupilone, a macrocyclic polyketide, is a microtubule-stabilizing agent that belongs to the epothilone class. It binds to the  $\beta$ -tubulin subunit of microtubules [21, 22]. *In vitro* evidence indicates that patupilone is a more potent inducer of tubulin dimerization and is more effective in stabilizing preformed microtubules than taxanes [22, 23]. In HCC cell lines, patupilone is 4- to 130-fold more potent than taxanes [24]. Clinical studies of patupilone in solid tumor types including lung and ovarian cancers demonstrated high potency in its anticancer activity [25–27].

In the current study, we investigated the antitumor efficacy of everolimus in HCC, either alone or in combination with the novel microtubule-destabilizing agent, patupilone, in both *in vitro* and *in vivo* models of HCC.

## 2. Materials and Methods

**2.1. Reagents.** Everolimus (RAD001/Afinitor) and patupilone (Epothilone B, EPO906) were obtained from Novartis Pharma (Basel, Switzerland) and dissolved in DMSO at a stock concentration of 10 mM and stored at  $-20^{\circ}\text{C}$ . The following antibodies were used in the study: anti-mTOR, anti-pi-mTOR (ser2448), anti-Akt, anti-pi-Akt (ser473), anti-p70S6k, anti-pi-p70S6k (Thr389), anti-S6, anti-pi-S6 (ser240/244), anti-4E-BP1, anti-pi-4E-BP1 (ser65), anti-cleaved PARP (all from Cell Signaling Technology, Beverly, MA, USA), and anti-actin (Calbiochem, Nottingham, UK).

**2.2. Cell Culture.** Human hepatocellular carcinoma cell lines Hep3B, HepG2, PLC/PRF/5, and SNU398 were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA) and Huh7 was obtained from Japanese Collection of Research Bioresources (JCRB, Japan). Hep3B, HepG2, Huh7, and PLC/PRF/5 were cultured in Dulbecco's modified Eagle medium with Glutamax-1 (HyClone, Logan, UT, USA) supplemented with 10% fetal bovine serum, FBS (HyClone). SNU398 was cultured in complete RPMI-1640 medium (HyClone) containing 10% FBS (HyClone). All cells were cultured under a humidified atmosphere of 5%  $\text{CO}_2$  at  $37^{\circ}\text{C}$  as previously described [17].

**2.3. Cell Viability Assay.** Cells (8000–18000 cells per well) were treated with either vehicle (DMSO) or increasing concentrations of everolimus (ranging from 0.1 nM to 20  $\mu\text{M}$ ) or patupilone (ranging from 0.01 nM to 1  $\mu\text{M}$ ) for 48 and 72 hrs. For combination treatment, cells were treated with increasing concentrations of everolimus and low concentration of patupilone (0.5 nM). Cell viability was determined by MTT assay as previously described [28]. The percentage growth inhibition was calculated as  $(\text{OD}_{\text{vehicle}} - \text{OD}_{\text{drug}}) / \text{OD}_{\text{vehicle}} \times 100\%$ . The  $\text{IC}_{50}$  value was determined as the drug concentration at which half of the maximal growth inhibition was observed.

**2.4. Western Blotting.** Protein lysates were obtained as previously described [28]. Protein lysates (25–50  $\mu\text{g}$ ) were separated by SDS-PAGE (sodium dodecyl sulfate-polyacrylamide gel electrophoresis) and transferred to nitrocellulose membranes. After primary and secondary antibody incubations, the signal was detected by autoradiography using SuperSignal West Pico Chemiluminescent Substrate (Thermo Fisher Scientific, Rockford, IL, USA).

**2.5. HCC Xenograft Study.** Four-to-six-week-old male athymic nude mice (nu/nu) were used for the establishment of HCC xenografts. All experiments were conducted under license from the Department of Health and according to animal ethics approval from the University Animal Experimentation Ethics Committee, the Chinese University of Hong Kong. HCC cells ( $3 \times 10^6$  of Hep3B cells suspended in 200  $\mu\text{L}$  serum-free medium) were inoculated into the dorsal flanks of mice by subcutaneous injection. Mice were randomized into four groups. Treatments were started on day 20 after inoculation. The 4 treatment groups were (1) vehicle control, (2) everolimus alone (2.5 mg/kg, twice a week, orally), (3) patupilone alone (0.5 mg/kg, once a week, i.p.), and (4) a combination of everolimus and patupilone (two agents administered in an alternating schedule, not as mixed solution, with same doses and schedules as single agents alone). Tumor growth was monitored twice weekly and tumor volume was calculated using the formula of  $((\text{Length} \times \text{Width}^2) / 2)$  as previously published [29].

**2.6. Immunohistochemistry and Microvessel Density (MVD) Determination.** Immunohistochemistry was performed as previously described [30]. Tumor microvessels were stained with a rabbit anti-CD34 antibody (1 : 100 dilution, Santa Cruz Biotechnology). IHC score approach was applied to assess the intensity of staining for each xenograft specimen. The IHC score ranged from 1 to 4, 1 = -ve to weak, 2 = weak to moderate, 3 = moderate to strong, and 4 = strongest staining.

**2.7. Statistical Analysis.** All data were presented as mean  $\pm$  SEM. Student's *t*-test (with Welch's correction) was performed using GraphPad Prism 4.0 software (GraphPad Software, Inc., San Diego, CA, USA). Results were considered as statistically significant if  $P < 0.05$ .

TABLE 1: Average tumor volume and tumor weight during treatments in Hep3B xenograft model.

Treatments	Average tumor volume (mm <sup>3</sup> )		Average tumor weight (μg)
	Week 1	Week 2	End of experiment
Veh	122.66 ± 15.73	352.97 ± 40.47	434.70 ± 60.43
Eve	81.75 ± 9.32 (*P < 0.05)	218.56 ± 25.25 (*P < 0.05)	308.60 ± 44.10 (P = 0.11)
Pat	92.40 ± 12.03 (P = 0.14)	239.41 ± 33.81 (*P < 0.05)	346.10 ± 56.76 (P = 0.29)
Eve/Pat	65.14 ± 8.62 (**P < 0.01)	138.57 ± 16.57 (***P < 0.001)	228.10 ± 37.20 (**P < 0.01)

\*P < 0.05 versus vehicle control group; \*\*P < 0.01 versus vehicle control group; \*\*\*P < 0.001 versus vehicle control group.

### 3. Results

**3.1. Everolimus Inhibited HCC Cell Proliferation with Effective Inhibition of mTOR Signaling.** To examine the effects of everolimus on HCC cell proliferation, five HCC cell lines (SNU398, Hep3B, HepG2, PLC/PRF/5, and Huh7) were treated with everolimus at increasing concentrations (0.1 nM–20 μM). As early as 48 hrs upon treatment, everolimus was able to induce dose-dependent growth inhibition in all five cell lines tested, with a maximal achievable growth inhibition of ~90–95% at 20 μM concentration. Among these HCC cell lines tested, SNU398 was the most everolimus-sensitive (average IC<sub>50</sub> = 2.10 ± 0.25 μM), while HepG2 was the most resistant one (average IC<sub>50</sub> = 8.84 ± 0.70 μM). The remaining three cell lines, Hep3B, Huh7, and PLC/5, had intermediate sensitivities (Figures 1(a) and 1(b)).

Next, we examined the effects of everolimus on mTOR signaling in HCC cells. In HepG2, Hep3B, and SNU398 cells, everolimus (0.1 μM) was able to elicit marked inhibition of mTOR signaling at 48 hrs, sustaining up to 72 hrs (Figure 1(c)). This was indicated by significant inhibition of phospho-mTOR (ser2448), as well as effective inhibition of its downstream effectors, including phospho-p70S6k (Thr389), phospho-S6 (ser240/244), and phospho-4E-BP1 (ser65) (Figure 1(c)). Our results showed that everolimus can abrogate mTOR activation and its downstream targets in HCC cells. It is noted that different extent of upregulation of phospho-Akt (ser473) was observed in the three cell lines upon everolimus treatment (Supplementary Figure 1(a) available online at <http://dx.doi.org/10.1155/2013/103830>), implicating a possible feedback upregulation of p-Akt by everolimus.

**3.2. Patupilone Inhibited HCC Cell Proliferation.** In present study, we examined the effects of patupilone on HCC cell proliferation in five HCC cell lines (SNU398, Hep3B, HepG2, PLC/PRF/5, and Huh7). Cells were treated with patupilone at increasing concentrations (0.01–1 μM). Dose-dependent inhibition of cell proliferation was observed in all of these five cell lines after being treated with patupilone for 48 hrs. Among these HCC cell lines tested, HepG2 was the most everolimus-sensitive (average IC<sub>50</sub> = 0.41 ± 0.07 μM), while Huh7 was the most resistant one with IC<sub>50</sub> > 10 μM. The

remaining three cell lines, Hep3B, SNU398, and PLC/5, had intermediate sensitivities (Figures 2(a) and 2(b)).

**3.3. Enhanced Antitumor Activity of Everolimus/Patupilone Combination In Vitro.** Studies in cervical and ovarian cancers revealed that activation of the PI3K/Akt/mTOR pathway is associated with resistance to microtubule-targeting agents, implicating a potential benefit of combined targeting of both the microtubules and the PI3K/Akt/mTOR pathway [18, 31]. Previous study by our group has shown synergistic antitumor effect of temsirolimus (an mTOR inhibitor) and vinblastine (a microtubule targeting agent) [20]. Here we examined the *in vitro* antitumor activity of everolimus/patupilone combination in HepG2, Hep3B, and SNU398 cells. As shown in Figure 3(a), the Hep3B cell line was only moderately sensitive to high dose of everolimus treatment at 48 hrs (with a maximal growth inhibition of only 48.09% even at 5 μM concentration). Patupilone alone at low concentration (0.5 nM) only inhibited Hep3B proliferation by 20%. Strikingly, this low-dose patupilone with everolimus was able to enhance the growth inhibitory activity of everolimus (64.85 ± 1.46%, *n* = 3, *P* < 0.01) as early as 48 hrs. Similar findings were observed in the everolimus-sensitive SNU398 cells. A maximum growth inhibition of 86.93 ± 0.81% was observed in Huh7 cells with everolimus/patupilone combination (*n* = 3, *P* < 0.01) (Figure 3(a)). An enhanced growth inhibitory effect was also observed in the everolimus-resistant HepG2 cells, achieving 59.26 ± 1.07% maximal growth inhibition as early as 48 hrs (Figure 3(a)). Our findings in multiple HCC cell lines demonstrated marked therapeutic efficacy with such combination therapy.

**3.4. Everolimus/Patupilone Combination Elicited Potent Antitumor Activity In Vivo.** The striking *in vitro* anticancer activity of this everolimus/patupilone combination compelled us to examine if this combination would be effective *in vivo*. Using established xenograft models of Hep3B (with intermediate everolimus-sensitivity, Figures 1(a) and 1(b)), we found that one week of everolimus treatment alone was able to inhibit the growth of Hep3B tumors, when compared to vehicle alone (average tumor volume of 81.75 ± 9.32 mm<sup>3</sup> versus 122.66 ± 15.73 mm<sup>3</sup> in vehicle alone; *n* = 15, *P* < 0.05) (Figure 4(a) and Table 1). An additional week of everolimus treatment also elicited significant change in tumor volume

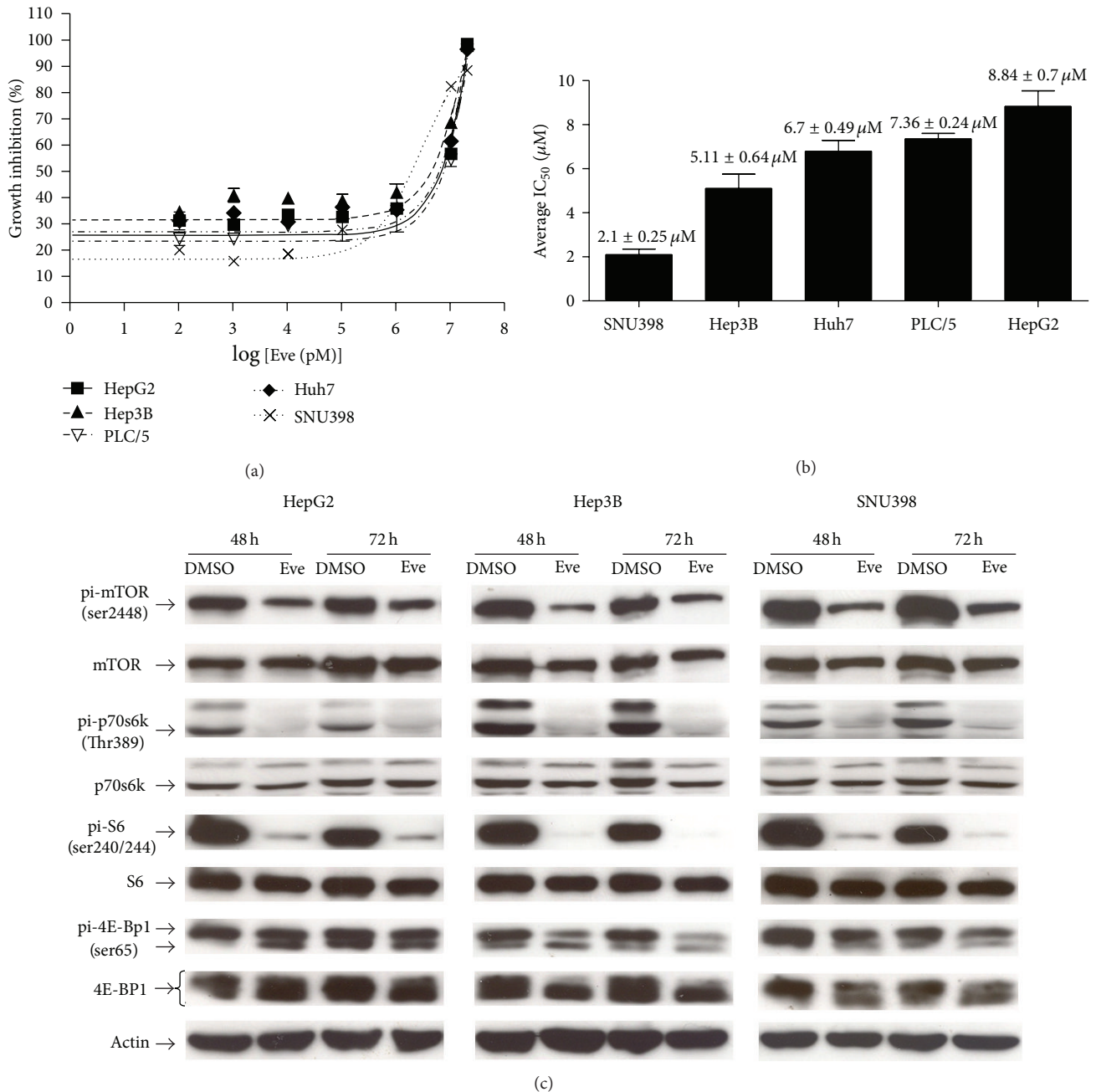


FIGURE 1: Everolimus inhibited proliferation and mTOR signaling in HCC cell lines. (a) Dose-dependent inhibition of HCC cell proliferation by everolimus. The effect of everolimus on cell viability was assessed by MTT assay. Dose-response curves of everolimus for all HCC cell lines were shown. Similar results were observed in 3 independent experiments. (b) Average IC<sub>50</sub> values of everolimus in HCC cell lines. Cumulative results from 3 independent experiments were shown as mean ± SEM. (c) Everolimus inhibited the mTOR pathway in HCC cells. HepG2, Hep3B, and SNU398 cells ( $3 \times 10^5$ ) were treated with 0.1 μM everolimus (hereafter labeled as Eve) or DMSO control for 48 hrs and 72 hrs. The expression levels of the mTOR pathway components, pi-mTOR (ser2448), mTOR, pi-p70S6K (Thr389), p70S6K, pi-S6 (ser240/244), S6, pi-4E-BP1 (ser65), and 4E-BP1, and actin were examined by western blotting. Similar results were observed in 3 independent experiments.

(average tumor volume of  $218.56 \pm 25.25 \text{ mm}^3$  versus  $352.97 \pm 40.47 \text{ mm}^3$  in the vehicle-treated group;  $n = 15$ ,  $P < 0.05$ ), consistent with the *in vitro* observation that these cells are moderately sensitive to everolimus (Figures 1(a) and 1(b)). Patupilone alone seemed to achieve a moderate degree of growth inhibition. However, as reported in an early study in

which higher dose of patupilone was administered intraperitoneally [32], higher concentration of patupilone was lethal to mice in the present study (data not shown), thus limiting dose escalation of patupilone in mice. Consistent with the marked *in vitro* growth inhibitory activity of everolimus/patupilone combination, we found that this combination was able to

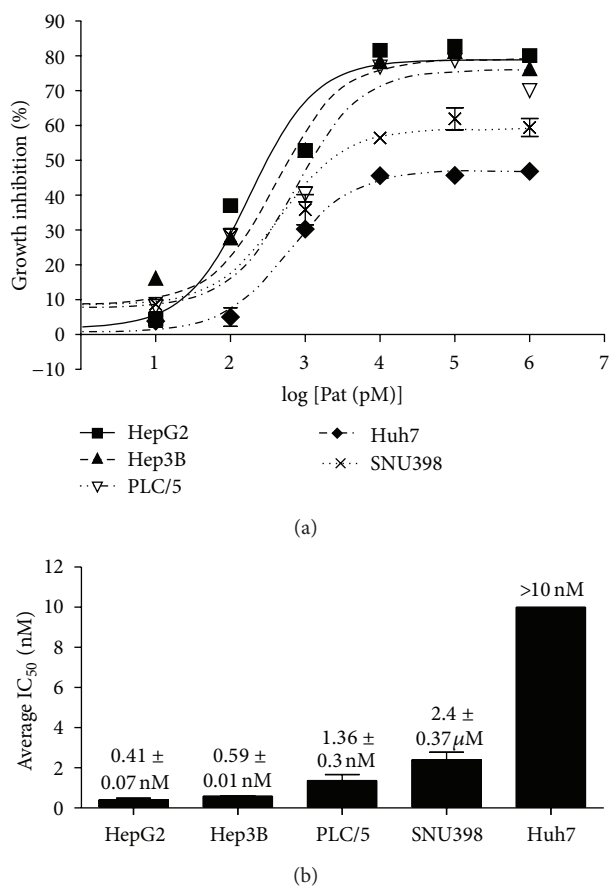


FIGURE 2: Patupilone inhibited proliferation in HCC cell lines. (a) Dose-dependent inhibition of HCC cell proliferation by patupilone. The effect of patupilone on cell viability was assessed by MTT assay. Dose-response curves of everolimus for all HCC cell lines were shown. Similar results were observed in 3 independent experiments. (b) Average IC<sub>50</sub> values of patupilone in HCC cell lines. Cumulative results from 3 independent experiments were shown as mean ± SEM.

inhibit Hep3B tumor growth significantly as early as 4 days after treatment (i.e., 24 days after tumor cell inoculation in Figure 4(a)). The most remarkable observation was that with only 2 weeks of treatment, the final tumor volume of the combination group was  $138.57 \pm 16.57 \text{ mm}^3$  versus  $352.97 \pm 40.47 \text{ mm}^3$  in the vehicle-treated group (~2.5-fold difference,  $n = 15$ ,  $P < 0.001$ ),  $218.56 \pm 25.25 \text{ mm}^3$  in the everolimus only group, and  $239.41 \pm 33.81 \text{ mm}^3$  in the patupilone only group (Figure 4(a) and Table 1). The final tumor weight of the combination group was  $228.10 \pm 37.20 \mu\text{g}$  versus  $434.70 \pm 60.43 \mu\text{g}$  in the vehicle-treated group ( $n = 15$ ,  $P < 0.01$ ),  $308.60 \pm 44.10 \mu\text{g}$  in the everolimus-only group, and  $346.10 \pm 56.76 \mu\text{g}$  in the patupilone-only group (Figure 4(b)). The treatment was tolerable by all groups with no deaths (data not shown).

**3.5. Everolimus/Patupilone Combination Did Not Further Suppress mTOR Signaling in HCC Models.** In order to examine the mechanism of such an enhanced antitumor

activity of this combination, we examined the effects of this everolimus/patupilone combination on mTOR signaling pathway in HCC cells. As shown in Figure 3(b), everolimus/patupilone combination did not result in further suppression of mTOR signaling when compared to everolimus treatment alone, while patupilone alone did not alter mTOR signaling in HepG2, Hep3B, and SNU398 cells (Figure 3(b)). These results indicate that the enhanced antiproliferative effect of the everolimus/patupilone combination is probably unrelated to further suppression of mTOR signaling in HCC cells. Note that the feedback activation of Akt still persisted with the everolimus/patupilone combination treatment in all the three cell lines (Supplementary Figure 1(b)), suggesting that the efficacy of this combination was probably not due to inhibition of this Akt feedback in HCC cells.

In fact, these *in vitro* findings were also confirmed in the respective *in vivo* models as well. As shown in Figure 4(c), pi-S6 and pi-mTOR levels were reduced in Hep3B tumors treated with either everolimus alone or with the combination, while patupilone did not suppress the two phosphoprotein levels.

### 3.6. Everolimus/Patupilone Combination-Induced Cell Apoptosis and Exerted Antiangiogenic Effect in HCC Models.

Next, we examined if the marked antitumor activity of the combination was due to possible induction of apoptosis in these HCC models, as the PI3K/Akt/mTOR signaling pathway is known to be crucial for cell survival. As shown in Figure 5, PARP cleavage was readily detected (by IHC) in Hep3B tumors treated with everolimus and patupilone alone and further increased in tumors treated with the combination (Figures 5(a) and 5(b)). These results implied that the observed antitumor effect was at least partly mediated by cell apoptosis induced in the combination treatment.

In addition to the observed cell apoptosis induction in HCC xenografts, we also found that this combination was able to induce a significant reduction in microvessel density (MVD) in Hep3B models as compared to vehicle control (Figure 5), suggesting potent antiangiogenic activity of this combination in HCC models. As shown in Figure 5(b), administration of everolimus or patupilone alone in Hep3B xenografts for 15 days was able to inhibit MVD by 44.4% and 33.3%, respectively, while the combination inhibited MVD by 52% ( $n = 10$ ,  $P < 0.001$  versus vehicle control group).

## 4. Discussion

In this study, we report the enhanced antitumor activity of cotargeting of mTOR (by everolimus) and the microtubules (by patupilone) in both *in vivo* and *in vitro* models of HCC, in which induction of cell apoptosis and inhibition of angiogenesis were detected. The observed additive to synergistic inhibitory effects of the everolimus/patupilone combination on HCC cell growth in multiple cell lines of HCC *in vitro* was further supported by the Hep3B xenograft model, where a potent antitumor and antiangiogenic effects were observed with only two cycles of this combination treatment. Our

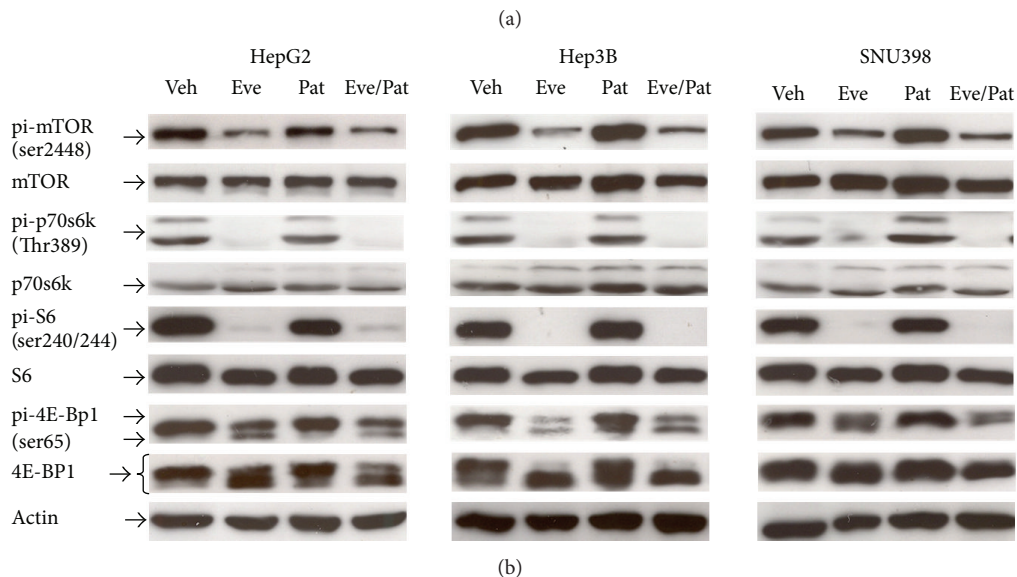
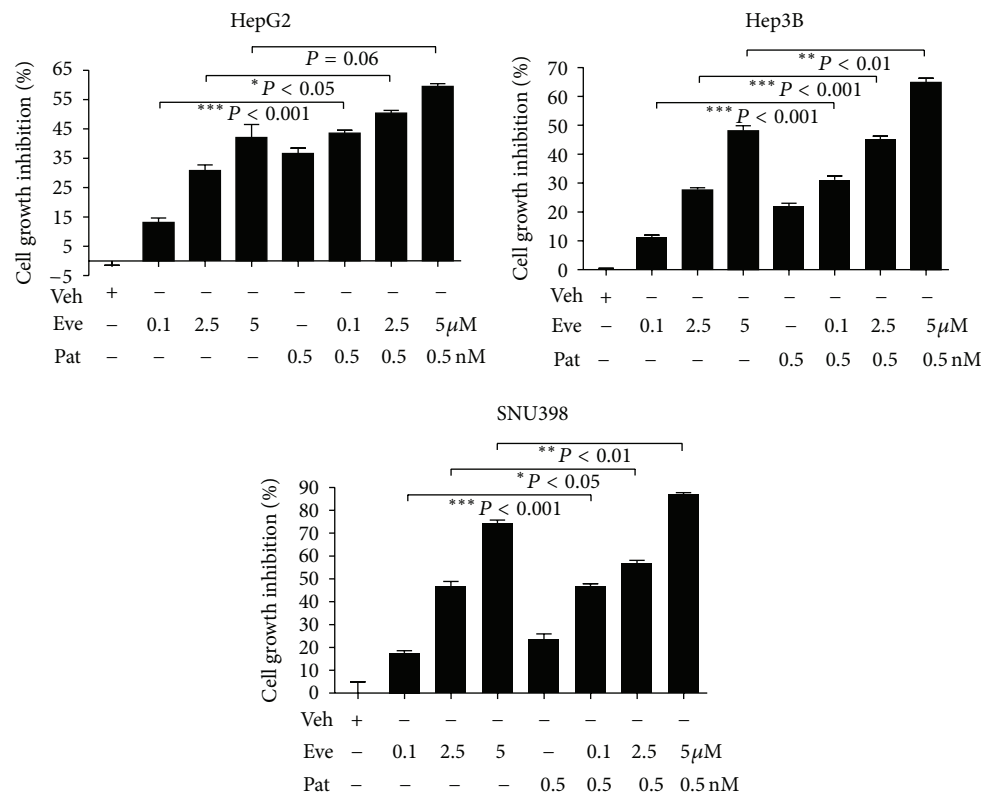


FIGURE 3: Enhanced antitumor activity of the everolimus/patupilone combination in HCC cell lines. (a) Effects of everolimus/patupilone in HCC cell lines. HepG2, Hep3B, and SNU398 cells ( $1 \times 10^4$ ) were treated with various concentrations of everolimus in combination with 0.5 nM patupilone (Pat) for 24 hrs. Cell viability was assessed by MTT assay. Cumulative results from 3 independent experiments were shown as mean  $\pm$  SEM (\* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  versus everolimus-treated group). (b) The mTOR signaling in HCC cells was not further suppressed by the everolimus/patupilone combination treatment. HepG2, Hep3B, and SNU398 cells ( $3 \times 10^5$ ) were treated with everolimus (0.1  $\mu$ M) and/or patupilone (Pat) (0.5 nM) for 24 hrs. The everolimus/patupilone combination is abbreviated as Eve/Pat hereafter. The expression levels of the mTOR pathway components, pi-mTOR (ser2448), mTOR, pi-p70S6K (Thr389), p70S6K, pi-S6 (ser240/244), S6, pi-4E-BP1 (ser65), and 4E-BP1, and actin were examined by western blotting. Similar results were observed in 3 independent experiments.

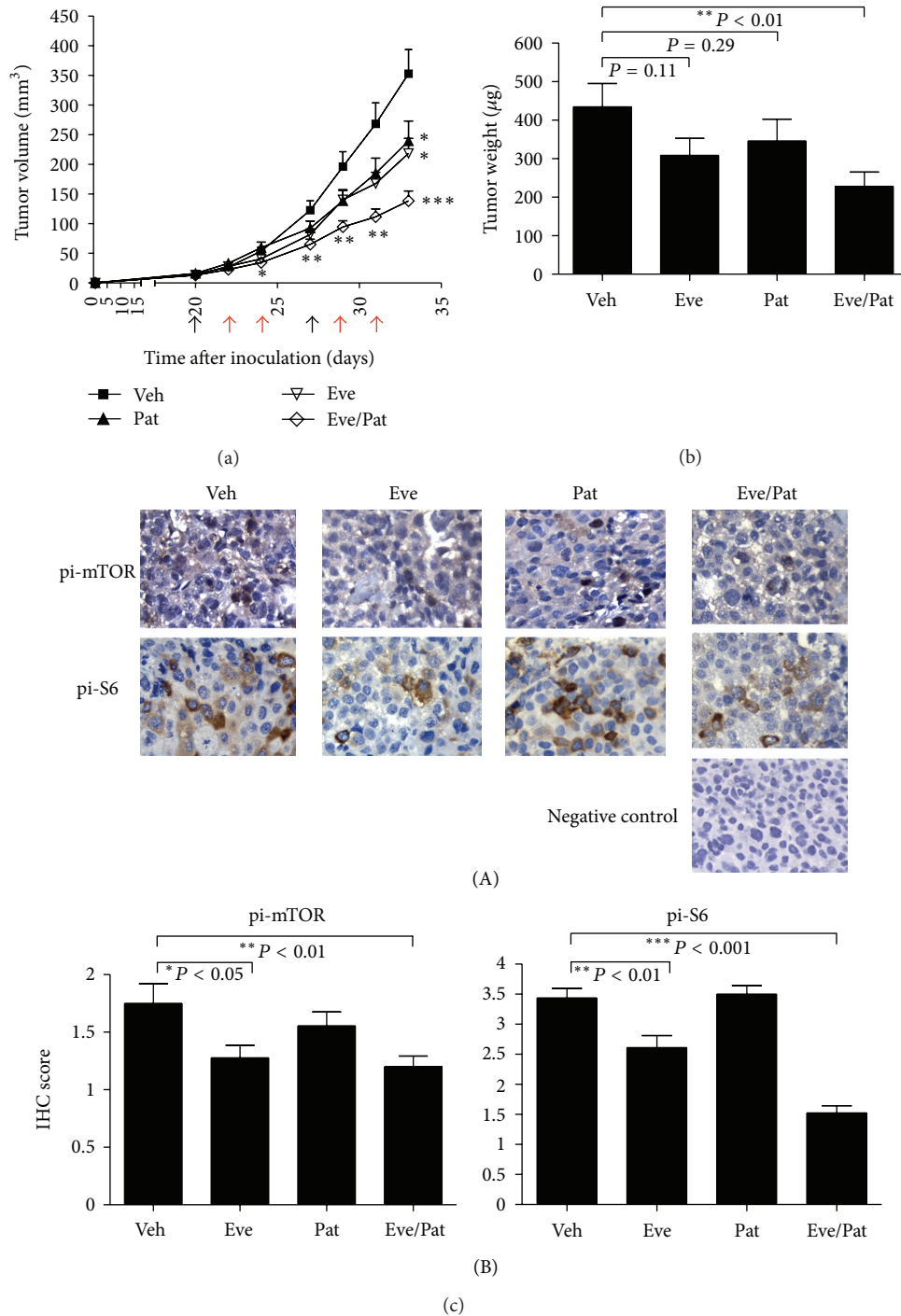


FIGURE 4: Potent antitumor effects of the everolimus/patupilone combination in *in vivo* models of HCC. Hep3B cells ( $3 \times 10^6$  cells) were inoculated into nude mice by subcutaneous injection. Drug treatments were started on day 20 after inoculation. Mice received administration of drugs for two weeks (black arrow: patupilone i.p. injection; red arrow: everolimus orally given). (a) Treatment of mice with everolimus, patupilone, or combination suppressed tumor growth in established xenografts of Hep3B. Tumor growth was monitored twice weekly. Arrows indicated time of drug administration. The *in vivo* antitumor activity of everolimus/patupilone combination was more significant than either agent alone ( $n = 10$  per group,  $*P < 0.05$ ,  $**P < 0.01$ ,  $***P < 0.001$  versus vehicle group). (b) Tumor weight of Hep3B xenografts in each group. The tumor weight of everolimus/patupilone combination group was significantly reduced ( $n = 10$  per group,  $**P < 0.01$  versus vehicle group). (c) The mTOR signaling in HCC cells was not further suppressed by the everolimus/patupilone combination treatment in Hep3B xenograft. Tumor xenografts were harvested, fixed, and stained for pi-mTOR and pi-S6 by immunohistochemistry. Representative images (400x magnification) were shown. Quantitation of pi-mTOR and pi-S6 staining using immunohistochemistry scoring was shown in (B) ( $n = 10$  per group,  $*P < 0.05$ ,  $**P < 0.01$ ,  $***P < 0.001$  versus vehicle group).

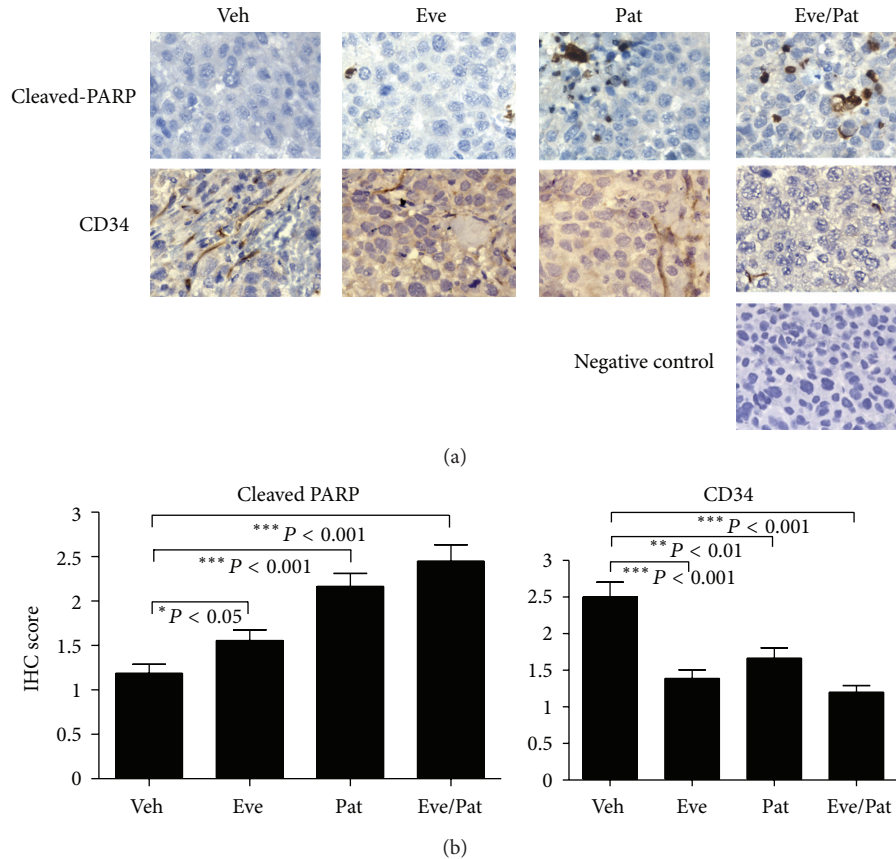


FIGURE 5: Everolimus/patupilone combination induced cell apoptosis and exerted antiangiogenic effect in HCC models. Tumor xenografts were harvested, fixed, and stained for cleaved PARP and CD34 by immunohistochemistry. Representative images (400x magnification) were shown. Quantitation of cleaved PARP and CD34 (microvessel density, MVD) staining was shown in (b) ( $n = 10$  per group,  $*P < 0.05$ ,  $**P < 0.01$ ,  $***P < 0.001$  versus vehicle group).

results indicate that the combination of everolimus with patupilone could be a highly effective regimen for HCC treatment, which warrants further clinical investigations in HCC patients.

We found that the HCC cell lines studied have demonstrated a similar sensitivity towards mTOR targeting by everolimus alone, with their  $IC_{50}$  ranging from 2.10 to 8.84  $\mu$ M. Previous studies in other cancers have indicated that mTOR targeting may elicit cytostatic effects rather than effective eradication of tumor cells [33, 34], suggesting that a combination of mTOR targeting with cytotoxic agents may be advantageous. Therefore, in search for a rational combination with everolimus, we decided to choose a combination with a microtubule-targeting agent, patupilone, based on the following evidence: (1) microtubule-targeting is believed to be a prominent druggable target in HCC [15–17], more importantly, (2) dual targeting of mTOR and microtubule by temsirolimus and vinblastine has recently shown sustained and potent antitumor effect in HCC models [20], and, lastly, (3) patupilone has been reported to be the most potent microtubule-targeting agent for HCC [35]. Indeed, we found that all the HCC cell lines that were tested were sensitive to patupilone, with the lowest  $IC_{50}$  being 0.41 nM.

Further, when everolimus was combined with very low dose of patupilone (0.5 nM), enhanced effect was observed in HCC cell lines with a maximal achievable growth inhibition of about 70–90%. More interestingly, we found that the superior antitumor activity of the addition of patupilone in HCC models was not contributed to further suppression of mTOR signaling pathway compared with everolimus alone, implicating mTOR-independent effects on growth inhibition with this combination.

When further investigating the mechanism involved, it was revealed that the combined treatment significantly induced cell apoptosis and suppressed angiogenesis, suggesting these two events to be the contributing mechanisms of the synergistic growth inhibition in HCC models. We found that PARP cleavage, which is a hallmark of cell apoptosis, was significantly increased in Hep3B xenograft tumors with the combined treatment versus vehicle control, although this effect seems to be mainly attributable to patupilone. This finding is consistent with the previous reports that mTOR targeting may only elicit cytostatic effects rather than cytotoxic effects [33, 34]. At the same time, microvessel density (MVD) was significantly reduced in tumors treated with the combination. In fact, the antiangiogenic effect by mTOR



inhibitor and microtubule-targeting agent combination has been reported. Marimpietri et al. recently demonstrated that combination of rapamycin and vinblastine enhanced the therapeutic effect on human neuroblastoma growth, apoptosis, and angiogenesis [36]. Moreover, rapamycin/vinblastine combination was found to exert antiangiogenic effects in an endothelial cell line EA.hy926 [37]. A previous study by our group has also shown that temsirolimus/vinblastine combination had marked antiangiogenic effect in HCC. In the current study, we further demonstrated the antiangiogenic effect with mTOR/microtubule targeting.

Everolimus is currently undergoing a phase III clinical trial in HCC. The earlier phase I/II study of everolimus has shown modest antitumor activity, with median progression-free survival of 3.8 months and overall survival of 8.4 months in patients with advanced HCC [38]. As a novel microtubule-targeting agent, patupilone has only shown modest antitumor effect as a single agent in a phase II study conducted in advanced HCC, with progression-free survival of 3 months and disease stabilization rate of 44% [39]. Based on the data from the current study, we were able to show for the first time that combination of a very low dose of patupilone with everolimus was able to result in a much stronger antitumor effect when compared to either of the single agents alone in HCC models.

## 5. Conclusions

In conclusion, our study demonstrated that the combination of everolimus with low dose of patupilone could be a highly effective regimen for the treatment of HCC. Clinical investigation into the role of such combination in HCC patients is warranted.

## Abbreviations

HCC: Hepatocellular carcinoma  
 mTOR: Mammalian target of rapamycin  
 4E-BP1: eIF4E binding protein  
 MVD: Microvessel density  
 MDR: Multidrug resistant gene.

## Conflict of Interests

All authors declare no financial/commercial conflict of interests regarding the study.

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## References

- [1] J. He, D. Gu, X. Wu et al., "Major causes of death among men and women in China," *New England Journal of Medicine*, vol. 353, no. 11, pp. 1124–1134, 2005.
- [2] D. M. Parkin, "Global cancer statistics in the year 2000," *Lancet Oncology*, vol. 2, no. 9, pp. 533–543, 2001.
- [3] M. Schwartz, "Liver transplantation for hepatocellular carcinoma," *Gastroenterology*, vol. 127, no. 5, supplement 1, pp. S268–S276, 2004.
- [4] R. T. P. Poon, S. T. Fan, C. M. Lo et al., "Improving survival results after resection of hepatocellular carcinoma: a prospective study of 377 patients over 10 years," *Annals of Surgery*, vol. 234, no. 1, pp. 63–70, 2001.
- [5] A. L. Cheng, Y. K. Kang, Z. Chen et al., "Efficacy and safety of sorafenib in patients in the Asia-Pacific region with advanced hepatocellular carcinoma: a phase III randomised, double-blind, placebo-controlled trial," *The Lancet Oncology*, vol. 10, no. 1, pp. 25–34, 2009.
- [6] J. M. Llovet, S. Ricci, V. Mazzaferro et al., "Sorafenib in advanced hepatocellular carcinoma," *New England Journal of Medicine*, vol. 359, no. 4, pp. 378–390, 2008.
- [7] M. A. Bjornsti and P. J. Houghton, "The TOR pathway: a target for cancer therapy," *Nature Reviews Cancer*, vol. 4, no. 5, pp. 335–348, 2004.
- [8] S. S. Agarwala and S. Case, "Everolimus (RAD001) in the treatment of advanced renal cell carcinoma: a review," *Oncologist*, vol. 15, no. 3, pp. 236–245, 2010.
- [9] K. L. Yim, "Everolimus and mTOR inhibition in pancreatic neuroendocrine tumors," *Cancer Management and Research*, vol. 4, pp. 207–214, 2012.
- [10] J. Baselga, M. Campone, M. Piccart et al., "Everolimus in postmenopausal hormone-receptor-positive advanced breast cancer," *New England Journal of Medicine*, vol. 366, no. 6, pp. 520–529, 2012.
- [11] M. P. Curran, "Everolimus: in patients with subependymal giant cell astrocytoma associated with tuberous sclerosis complex," *Pediatric Drugs*, vol. 14, no. 1, pp. 51–60, 2012.
- [12] L. S. Blaszkowsky, T. A. Abrams, R. A. Miksad et al., "Phase I/II study of everolimus in patients with advanced hepatocellular carcinoma (HCC)," *Journal of Clinical Oncology*, ASCO Annual Meeting, Article ID e14542, 2010.
- [13] S. Mabuchi, D. A. Altomare, D. C. Connolly et al., "RAD001 (Everolimus) delays tumor onset and progression in a transgenic mouse model of ovarian cancer," *Cancer Research*, vol. 67, no. 6, pp. 2408–2413, 2007.
- [14] C. Wu, M. Wangpaichitr, L. Feun et al., "Overcoming cisplatin resistance by mTOR inhibitor in lung cancer," *Molecular Cancer*, vol. 4, article 25, 2005.
- [15] M. A. Patil, M. S. Chua, K. H. Pan et al., "An integrated data analysis approach to characterize genes highly expressed in hepatocellular carcinoma," *Oncogene*, vol. 24, no. 23, pp. 3737–3747, 2005.
- [16] C. Y. Tung, C. H. Jen, M. T. Hsu, H. W. Wang, and C. H. Lin, "A novel regulatory event-based gene set analysis method for exploring global functional changes in heterogeneous genomic data sets," *BMC Genomics*, vol. 10, article 26, 2009.
- [17] Q. Zhou, A. K. K. Ching, W. K. C. Leung et al., "Novel therapeutic potential in targeting microtubules by nanoparticle albumin-bound paclitaxel in hepatocellular carcinoma," *International Journal of Oncology*, vol. 38, no. 3, pp. 721–731, 2011.
- [18] L. S. Fariied, A. Fariied, T. Kanuma et al., "Inhibition of the mammalian target of rapamycin (mTOR) by rapamycin increases chemosensitivity of CaSki cells to paclitaxel," *European Journal of Cancer*, vol. 42, no. 7, pp. 934–947, 2006.

- [19] D. J. VanderWeele, R. Zhou, and C. M. Rudin, "Akt up-regulation increases resistance to microtubule-directed chemotherapeutic agents through mammalian target of rapamycin," *Molecular Cancer Therapeutics*, vol. 3, no. 12, pp. 1605–1613, 2004.
- [20] Q. Zhou, V. W. Lui, C. P. Lau et al., "Sustained antitumor activity by co-targeting mTOR and the microtubule with temsirolimus/vinblastine combination in hepatocellular carcinoma," *Biochemical Pharmacology*, vol. 83, no. 9, pp. 1146–1158, 2012.
- [21] J. H. Nettles, H. Li, B. Cornett, J. M. Krahn, J. P. Snyder, and K. H. Downing, "The binding mode of epothilone A on  $\alpha,\beta$ -tubulin by electron crystallography," *Science*, vol. 305, no. 5685, pp. 866–869, 2004.
- [22] K. H. Altmann, G. Bold, G. Caravatti et al., "Epothilones and their analogs—potential new weapons in the fight against cancer," *Chimia*, vol. 54, no. 11, pp. 612–621, 2000.
- [23] R. J. Kowalski, P. Giannakakou, and E. Hamel, "Activities of the microtubule-stabilizing agents epothilones A and B with purified tubulin and in cells resistant to paclitaxel (Taxol)," *Journal of Biological Chemistry*, vol. 272, no. 4, pp. 2534–2541, 1997.
- [24] T. S. K. Mok, E. Choi, D. Yau et al., "Effects of patupilone (epothilone B; EPO906), a novel chemotherapeutic agent, in hepatocellular carcinoma: an in vitro study," *Oncology*, vol. 71, no. 3-4, pp. 292–296, 2007.
- [25] E. H. Rubin, J. Rothermel, F. Tesfaye et al., "Phase I dose-finding study of weekly single-agent patupilone in patients with advanced solid tumors," *Journal of Clinical Oncology*, vol. 23, no. 36, pp. 9120–9129, 2005.
- [26] K. Osterlind, J. M. Sanchez, P. Zatloukal et al., "Phase I/II dose escalation trial of patupilone every 3 weeks in patients with non-small cell lung cancer," *Journal of Clinical Oncology*, vol. 23, no. 16, p. 647S, 2005.
- [27] W. M. Smit, J. Sufliarsky, S. Spanik et al., "Phase I/II dose-escalation trial of patupilone every 3 weeks in patients with resistant/refractory ovarian cancer," *European Journal of Cancer*, vol. 3, no. 2, pp. 261–262, 2005.
- [28] V. W. Y. Lui, A. L. Boehm, P. Koppikar et al., "Antiproliferative mechanisms of a transcription factor decoy targeting signal transducer and activator of transcription (STAT) 3: the role of STAT1," *Molecular Pharmacology*, vol. 71, no. 5, pp. 1435–1443, 2007.
- [29] V. W. Y. Lui, D. M. Yau, E. Y. Wong et al., "Cucurbitacin I elicits anoikis sensitization, inhibits cellular invasion and in vivo tumor formation ability of nasopharyngeal carcinoma cells," *Carcinogenesis*, vol. 30, no. 12, pp. 2085–2094, 2009.
- [30] V. W. Y. Lui, D. M. S. Yau, C. S. F. Cheung et al., "FGF8b oncogene mediates proliferation and invasion of Epstein-Barr virus-associated nasopharyngeal carcinoma cells: implication for viral-mediated FGF8b upregulation," *Oncogene*, vol. 30, no. 13, pp. 1518–1530, 2011.
- [31] L. Hu, J. Hofmann, Y. Lu, G. B. Mills, and R. B. Jaffe, "Inhibition of phosphatidylinositol 3'-kinase increases efficacy of paclitaxel in in vitro and in vivo ovarian cancer models," *Cancer Research*, vol. 62, no. 4, pp. 1087–1092, 2002.
- [32] T. C. Chou, X. G. Zhang, A. Balog et al., "Desoxyepothilone B: an efficacious microtubule-targeted antitumor agent with a promising in vivo profile relative to epothilone B," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 95, no. 16, pp. 9642–9647, 1998.
- [33] K. H. Tam, Z. F. Yang, C. K. Lau, C. T. Lam, R. W. C. Pang, and R. T. P. Poon, "Inhibition of mTOR enhances chemosensitivity in hepatocellular carcinoma," *Cancer Letters*, vol. 273, no. 2, pp. 201–209, 2009.
- [34] L. Wang, W. Y. Shi, Z. Y. Wu et al., "Cytostatic and anti-angiogenic effects of temsirolimus in refractory mantle cell lymphoma," *Journal of Hematology & Oncology*, vol. 3, article 30, 2010.
- [35] T. S. K. Mok, E. Choi, D. Yau et al., "Effects of patupilone (epothilone B; EPO906), a novel chemotherapeutic agent, in hepatocellular carcinoma: an in vitro study," *Oncology*, vol. 71, no. 3-4, pp. 292–296, 2007.
- [36] D. Marimpietri, C. Brignole, B. Nico et al., "Combined therapeutic effects of vinblastine and rapamycin on human neuroblastoma growth, apoptosis, and angiogenesis," *Clinical Cancer Research*, vol. 13, no. 13, pp. 3977–3988, 2007.
- [37] N. Campostrini, D. Marimpietri, A. Totolo et al., "Proteomic analysis of anti-angiogenic effects by a combined treatment with vinblastine and rapamycin in an endothelial cell line," *Proteomics*, vol. 6, no. 15, pp. 4420–4431, 2006.
- [38] A. X. Zhu, T. A. Abrams, R. Miksad et al., "Phase 1/2 study of everolimus in advanced hepatocellular carcinoma," *Cancer*, vol. 117, no. 22, pp. 5094–5102, 2011.
- [39] A. P. Venook, R. Poon, Y. K. Kang et al., "Evaluation of patupilone as monotherapy in patients with advanced hepatocellular carcinoma (HCC)," *Journal of Clinical Oncology*, vol. 25, ASCO Annual Meeting Proceedings, no. 18S, Article ID 15055, 2007.