REVIEW

Alpha and Beta Synucleins: From Pathophysiology to Clinical Application as Biomarkers

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ABSTRACT: The synuclein family includes three neuronal proteins, named α -synuclein, β -synuclein, and γ -synuclein, that have peculiar structural features. α -synuclein is largely known for being a key protein in the pathophysiology of Parkinson's disease (PD) and other synucleinopathies, namely, dementia with Lewy bodies and multisystem atrophy. The role of β -synuclein and γ -synuclein is less well understood in terms of physiological functions and potential contribution to human diseases. α -synuclein has been investigated extensively in both cerebrospinal fluid (CSF) and blood as a potential biomarker for synucleinopathies. Recently, great attention has been also paid to β -synuclein, whose CSF and blood levels seem to reflect synaptic damage and neurodegeneration independent of the presence of synucleinopathy. In this review, we aim to provide an

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overview on the pathophysiological roles of the synucleins. Because γ -synuclein has been poorly investigated in the field of synucleinopathy and its pathophysiological roles are far from being clear, we focus on the interactions between α -synuclein and β -synuclein in PD. We also discuss the role of α -synuclein and β -synuclein as potential biomarkers to improve the diagnostic characterization of synucleinopathies, thus highlighting their potential application in clinical trials for disease-modifying therapies. © 2022 The Authors. *Movement Disorders* published by Wiley Periodicals LLC on behalf of International Parkinson and Movement Disorder Society

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The synucleins are a family of the following three conserved neuronal proteins: α -synuclein (140 amino acids), β -synuclein (134 amino acids), and γ -synuclein (127 amino acids).¹ α -Synuclein has been investigated extensively in neurodegenerative disorders that are referred to as synucleinopathies, including Parkinson's disease (PD), dementia with Lewy bodies (DLB), and multiple system atrophy (MSA).² These disorders share the main pathological feature of accumulation, misfolding, and aggregation of α -synuclein, which progressively leads to large intracellular aggregates, namely, Lewy bodies and glial cytoplasmic inclusions. Although α -synuclein is relatively well characterized, the relevance of the two other synucleins for physiological neuronal functioning, as well as their potential contribution to neurodegeneration in synucleinopathies, is still poorly understood. At a clinical level, several studies have focused on the possible application of α -synuclein as a candidate fluid biomarker for synucleinopathies. The interest is also growing for β -synuclein, which has been recently measured in cerebrospinal fluid (CSF) and blood

from patients affected by synucleinopathies and other neurodegenerative disorders. In this scenario, a major question arises: How can our understanding of synuclein pathophysiology impact the applicability of these proteins as body fluid biomarkers?

In this review, we first summarize the recent evidence of the pathophysiological roles of synucleins. Because the physiological roles of γ -synuclein, as well as its involvement in neurodegeneration, are far from being clear, we focus on α -synuclein and β -synuclein. Second, we discuss the role of α -synuclein and β -synuclein as potential biomarkers to improve the diagnostic characterization of patients affected by synucleinopathies. Their potential use in research settings, that is, clinical trials for novel disease-modifying treatments targeting α -synuclein accumulation and misfolding, is also discussed.

Distribution and Structural Features of Synucleins

Synucleins are mainly expressed in the central nervous system (CNS) and encoded by the *SNCA* (α -synuclein), *SNCB* (β -synuclein), and *SNCG* (γ -synuclein) genes.³ α -Synuclein is abundant in neurons, where it represents 1% of all cytosolic proteins,⁴ but other sites of expression are identified in glial and blood cells.⁵ In neurons, α -synuclein and β -synuclein colocalize physiologically in presynaptic terminals,⁶ but with aging, α -synuclein may redistribute to the neuronal soma with a relative decrease in synaptic levels.⁷ The production of α -synuclein is greatly induced during CNS development, when the protein promotes synaptogenesis and maturation of neuronal precursors.^{8,9} β -Synuclein is likewise largely expressed in the CNS and retina, whereas γ -synuclein expression mainly occurs in retinal and peripheral neurons.¹⁰⁻¹²

Synucleins are usually defined as intrinsically disordered proteins, and their structural peculiarities largely influence their pathophysiological activities.¹³ The amino acid sequence of synucleins is distinguished into three main regions: N-terminus, nonamyloid component (NAC) region, and C-terminus. The N-terminus is highly conserved among the three synucleins and is responsible for their lipid-binding properties.¹⁴ When bound to lipids, the N-terminal domain assumes an α -helical structure. The N-terminal helicity negatively correlates with aggregation potential. It progressively decreases from β -synuclein and γ -synuclein to α -synuclein and is even lower in PD-associated α -synuclein mutants.^{15,16} The central NAC region is remarkably aggregation prone in α -synuclein and constitutes the cross- β -sheet motifs within pathological aggregates.¹³ By contrast, the NAC region of β-synuclein has a central deletion of 11 amino acids that greatly decreases its tendency to aggregate.¹⁷ The C-terminal domain profoundly differs among synucleins and regulates their solubility depending on its length and charge.¹⁸ In particular, the high C-terminal flexibility of α-synuclein enhances its propensity to aggregate, whereas β-synuclein is substantially more rigid as a result of a polyproline secondary structure.^{16,19} The P123H β-synuclein mutation, which causes familial DLB clusters, seems to confer to β-synuclein an α-synucleinlike behavior by increasing the flexibility of its C-terminus.^{20,21} In comparison, γ-synuclein has a relatively shorter C-terminal domain with fewer acidic residues.²²

In the cytosolic environment, the physiological forms of α -synuclein are still not adequately defined because α -synuclein can be found alternatively in multimeric and monomeric species.^{13,23,24} However, these findings have not been widely replicated. Several cytosolic species of α -synuclein could be in dynamic equilibrium between free and membrane-bound states. A transient multimerization process would be therefore crucial for the functioning of α -synuclein and, if compromised, might lead to pathological aggregation.

Intriguingly, despite being highly conserved neuronal proteins, knock-out (KO) of a single synuclein does not induce severe alterations in overall neuronal wellness in animal models. α -Synuclein KO mice experienced only moderate synaptic dysfunction, especially in nigral dopaminergic neurons²⁵⁻²⁷ and, surprisingly, double α -synuclein/ β -synuclein²⁸ or α -synuclein/ γ -synuclein KO mice²⁹ show similar and not worse alterations. Only triple-synuclein KO models have a severe motor phenotype with large synaptic degeneration and early mortality, which could be rescued by overexpression of monomeric α -synuclein.³⁰

Physiological Roles of Synucleins

 α -Synuclein exerts several physiological functions in neurons, mainly as a key regulator of vesicular trafficking and neurotransmitter exocytosis (Fig. 1). Its most important function is to participate in vesicular dynamics by assisting the assembly of the soluble N-ethylmaleimidesensitive factor attachment proteins receptors (SNARE), which make up a pivotal macromolecular complex of synaptic terminals.³¹ Moreover, α-synuclein actively modulates both the shape and the dimension of presynaptic vesicles by directly binding their lipid surface.³² In dopaminergic neurons, α-synuclein acts as a central regulator of dopamine (DA) metabolism by inhibiting DA synthesis^{33,34} and inducing both storage in presynaptic vesicles³⁵ and extracellular reuptake of the neurotransmitter.³⁶ Inside presynaptic vesicles, DA colocalizes with antioxidants,³⁷ while free cytosolic DA can undergo oxidative reactions that release toxic DA-derived compounds. such as DA-quinone, aminochrome, and 5,6-indole-quinone.³⁸ Some of these products can even interact with β -synuclein and promote its aggregation.³⁹ Indeed, by regulating DA synthesis and storage, α -synuclein might contribute to protect dopaminergic neurons from oxidation-



FIG. 1. Main pathophysiological roles of α -synuclein (blue) and β -synuclein (purple) in neurons. (**A**) In the neuronal body, α -synuclein participates in mitochondrial functioning and motility, contributing to redirecting damaged mitochondria to mitophagy. When this pathway is impaired, misfolded α -synuclein may induce accumulation of dysmorphic organelles (ie, mitochondria) and formation of neuronal inclusions. The physiological impact of α -synuclein on nuclear functions is unclear, whereas pathological α -synuclein might be related to alterations in the transcription process. To what extent β -synuclein and misfolded β -synuclein affect neuronal organelles and nuclei is currently unknown. (**B**) α -Synuclein interacts with several cyto-skeletal components, among which the axonal microtubules are destabilized by misfolded and aggregated α -synuclein. It is unknown whether β -synuclein interacts with microtubules and other structural proteins. (**C**) The most relevant functions of α -synuclein are exerted in presynaptic terminals and mainly deal with regulation of vesicular trafficking and homeostasis of neurotransmitters in both dopaminergic neurons and other cell types. Physiologically, α -synuclein promotes storage of dopamine into presynaptic vesicles and its release through exocytosis. In synucleinopathies, the synthesis of dopamine might be promoted by dopamine dyshomeostasis, thus contributing to neuronal damage. L-DOPA, levodopa; SNARE, soluble N-ethylmaleimide-sensitive factor attachment proteins receptors.

induced toxicity. The regulatory activities of α -synuclein on synapse homeostasis might also be of great relevance in other neurotransmitter systems, which have been only partially examined, that is, other monoaminergic centers of the brain.

In addition to synapses, α -synuclein plays significant physiological roles in other cellular organelles. Mitochondria have a well-established role in PD pathology² and might be associated with Lewy body formation.⁴⁰ α -Synuclein participates in mitochondrial energetics by directly interacting with several mitochondrial proteins, such as the respiratory chain complexes⁴¹ and adenosine triphosphate (ATP) synthase.⁴² Thus, it is not surprising that both α -synuclein deficiency and α -synuclein over-expression affect ATP levels and predispose to energy crises.⁴² Intriguingly, α -synuclein is also involved in

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mitochondrial motility and mitophagy by promoting the expression of Miro proteins, which are anchoring proteins that connect mitochondria to microtubules and redirect mitochondria to mitophagy when damaged.⁴³ Indeed, accumulation of α -synuclein induces retention of Miro proteins on the mitochondrial surface and delays mitophagy of damaged mitochondria⁴³ (Fig. 1). α-Synuclein interacts with several cytoskeletal components, namely, actin, spectrins, tau, and microtubules. In physiological conditions, α -synuclein sequesters actin monomers and inhibits their polymerization.⁴⁴ Spectrins are peculiar cytoskeletal partners of α -synuclein. β -Spectrin, together with α -synuclein, physiologically regulates vesicular dynamics in presynaptic terminals.⁴⁵ In contrast, α -spectrin binds only to pathological α-synuclein species and promotes cytoskeleton instability.⁴⁶ In microtubules, the activity of α -synuclein is similar to that of dynamases, and by binding directly to free and assembled tubulin, α -synuclein influences its polymerazion rate.⁴⁷⁻⁴⁹ Morever, α -synuclein and tubulin compete for the same binding domain on tau protein,⁵⁰ a microtubule-associated protein related to the pathophysiology of Alzheimer's disease (AD) and other neurodegenerative disorders that are referred to as tauopathies: by promoting tau phosphorylation, α -synuclein may inhibit the stabilizing action exerted by tau on microtubules.51,52 Although nuclear α -synuclein species are of great research interest, the pathophysiological relationship between α -synuclein and the nucleus is still unclear, especially because there are methodological issues affecting the detection of α -synuclein in the nuclear environment.⁵³ Inside this α -synuclein is mainly organelle. found in its S129-phosphorylated form $(p-\alpha-synuclein)$,⁵⁴ which is a posttranslationally modified species frequently found in pathological aggregates.⁵⁵ α -Synuclein can bind directly to more than 300 different regions of DNA and may downregulate the transcription of a variety of genes, mainly involved in cell cycle control and DNA repair systems.^{54,56,57} α -Synuclein also interplays with epigenetic targets, such as minichromosome maintenance proteins and histones.56,58,59

Most studies of the roles of α -synuclein in neuronal physiology have focused on dopaminergic neurons, as they are frequently affected by α -synuclein misfolding pathologies. However, it should be further clarified whether and to what extent α -synuclein affects the general functioning of central neurons, such as dopaminergic cells, but also peripheral neuronal groups, for example, the myoenteric cholinergic neurons of the gut.^{60,61}

Interestingly, α -synuclein takes part in the physiology of nonneuronal cells, particularly blood cells, where it is found in high quantities in erythrocytes, leukocytes, and platelets. Erythrocytes are the main peripheral source of α -synuclein and account for 99% of α -synuclein in blood.⁵ In mature erythrocytes, α -synuclein exerts both structural functions, by tethering cytoskeletal proteins and the cytoplasmic membrane,⁶² and metabolic activities, by regulating iron homeostasis.^{63,64} In animal models, blood α -synuclein concentrations correlate with hematocrit and hemoglobin levels, and α -synuclein KO mice experience iron deficiency and moderate anemia.^{65,66} Furthermore, α -synuclein may play a significant role during hematopoiesis because erythroid precursors produce α -synuclein during maturation.⁶⁷ Platelets and leukocytes similarly rely on α -synuclein for their physiological functioning, mainly depending on α -synuclein-mediated assembly of SNARE complexes, which are also important in these cells.^{68,69}

 β -Synuclein and γ -synuclein are expressed in retinal neurons and have been associated with glaucoma-related neurodegeneration.^{70,71} To date, no convincing hypothesis has been proposed for the roles of β -synuclein in neurons. but it may act as a general synaptic chaperone.⁷² In adrenal chromaffin cells, it regulates vesicle dynamics and secretory rate with mechanisms similar to those occurring in neurons.⁷³ As opposed to α -synuclein, β -synuclein is not expressed in blood cells, thus peripheral β -synuclein might exclusively derive from brain neurons. Two main homeostatic functions are attributed to γ -synuclein in neurons and other cell types. First, y-synuclein regulates cytoskeletal remodeling during the cell-cycle,⁷⁴ which has been linked to development and progression of brain tumors and peripheral carcinomas.^{70,75} In addition, γ -synuclein exerts metabolic activities during the feed-fast cycle in hypothalamus and adipose tissue.⁷

Synucleins in Neurological Diseases

Misfolding and accumulation of α -synuclein leads to formation of neuronal inclusions in PD and DLB and of oligodendrocytic bodies in MSA. a-Synuclein aggregation is a multistep process that is thought to progress from accumulation of monomeric a-synuclein to nucleation of small soluble oligomers and to formation of large insoluble fibrils. Soluble oligomeric species of α -synuclein are reported to be more neurotoxic than large fibrils, which may instead have protective effects by sequestering free α -synuclein monomers.^{2,77} However, recent findings have pointed out that α -synuclein fibrils may also be a source of toxic soluble oligomers.⁷⁸ Degradation of α -synuclein species depends mainly on the autophagic lysosomal pathway (ALP) and, to a lesser extent, on the ubiquitin-proteasome system. ALP dysfunctions are key biological features of synucleinopathies and lead to multiorganelle toxicity as both a cause and a consequence of α -synuclein aggregation.⁷⁹ Accumulation of damaged and dysmorphic organelles that are improperly cleared in case of ALP dysfunction (eg, mitochondria) may contribute to the formation of Lewy bodies being stuck and tethered together by α -synuclein molecules.⁴⁰

Among the factors that modulate the dynamics of α -synuclein aggregation,³⁵ β -synuclein deserves to be mentioned. It has been shown that β -synuclein can act as an antiaggregation agent against α -synuclein and even more efficiently against amyloid-β peptides by directly binding monomers or early-stage oligomers and preventing them from further growth.⁸⁰⁻⁸² Nevertheless, the apparent protective action of β-synuclein on protein misfolding should not be misinterpreted because, in certain conditions, β-synuclein behaves rather as an amyloidogenic protein and contributes to neurodegeneration.^{39,83} Indeed, the antiaggregation properties of β -synuclein occur only when α -synuclein and β -synuclein are coexpressed at fixed ratios⁸⁴ while at acidic pH, a condition that can be found in lysosomal compartments of the cell, α -synuclein and β-synuclein aggregate at similar rates.⁸⁵ Rather than being a proper antiaggregation agent, β -synuclein could be a less aggregation-prone protein that appears beneficial only when compared with proteins with a higher amyloidogenic potential, such as α -synuclein and amyloid- β peptides. β-Synuclein could thus express its pathological potential only in particular conditions. Indeed, overexpression of α -synuclein or β -synuclein produces similar pathological features in preclinical models, mainly associated with ALP dysfunction^{72,79} and mitochondrial impairment.³⁹ However, α -synuclein pathology determines an overall more severe phenotype and uniquely impacts mitochondrial motility.^{43,84}

Although α -synuclein pathology may affect several cerebral regions and lead to widespread neurodegeneration,⁸⁶ dopaminergic neurons show some unique vulnerability factors. This vulnerability might be partially related to DA dyshomeostasis because DA and DA-derived products are able to promote aggregation of both α -synuclein and β -synuclein, especially in the presence of an excessive oxidative burden.^{39,84} Moreover, it is still not clear whether the presence of neuromelanin, a pigment uniquely expressed in nigral neurons among the dopaminergic centers of the brain, 37,38 has a role in α -synuclein pathology. Indeed, neuromelanin derives from DA metabolites and was found to boost the production of α -synuclein.⁸⁷ α -Synuclein, DA, and neuromelanin, together with β -synuclein, might thus contribute to the selective vulnerability of nigral dopaminergic neurons in synucleinopathies.

Another issue not yet fully clarified concerns the possible involvement of β -synuclein and γ -synuclein in synucleinopathies and other neurological disorders. β -Synuclein-positive inclusions have been observed in PD and DLB hippocampi⁸⁸ as well as in MSA cerebellum.⁸⁹ Interestingly, β -synuclein has also been associated with neuroinflammatory disorders because anti- β -synuclein immunity drives the gray matter damage in progressive forms of multiple sclerosis.⁹⁰ In addition to their typical neuropathological picture, synucleinopathies,⁷² AD,⁹¹ brain tumors,⁷⁵ and amyotrophic lateral sclerosis⁹² show γ -synuclein-positive inclusions. All of these findings suggest a possible relationship between synucleins and a wide range of neurological conditions.

α-Synuclein as Biomarker: State of the Art and Future Perspectives

The diagnosis of PD and other synucleinopathies still relies on clinical and imaging criteria.93,94 However, given the frequency of incorrect and delayed diagnosis even when these criteria are properly applied,⁹⁵ the diagnostic work-up of synucleinopathies is moving toward a molecular-based assessment, following the route traced by AD.96 As the most relevant protein in synucleinopathies, α -synuclein has been proposed to be a fluid and neuroimaging biomarker for synucleinopathies. Attempts to target α -synuclein pathology with neuroimaging biomarkers have so far met with limited success. The main problems in α -synuclein-targeting compounds include the lack of specificity and the difficulty of crossing the blood-brain barrier (BBB). On one hand, polyaromatic molecules able to cross the BBB and bind α -synuclein aggregates have shown cross-reactivity with amyloid- β plaques.⁹⁷ On the other hand, specific antibodies to α-synuclein aggregates do not adequately cross the BBB when administered intravenously.⁹⁸ Finally, peptides and antibodies labeled with positron emission tomography and single-photon emission computed tomography tracers are not suitable agents given the short half life of the isotopes of choice.98 Hence, brain imaging for synucleinopathies cannot directly picture α -synuclein pathology to date but can only demonstrate the structural and functional consequences.

Most of the contributions to the diagnostic characterization of patients with synucleinopathies derive from research on fluid biomarkers, especially in the CSF. α -Synuclein has been extensively investigated as a candidate fluid biomarker for synucleinopathies because it is a central protein in the pathophysiology of these disorders.⁹⁹ As assessed in several investigations and metaanalyses, CSF total α -synuclein (t- α -synuclein) concentrations have shown an overall slight decrease in patients with PD compared with healthy and neurological controls.100-105 Similar findings were found in DLB and MSA,^{106,107} with no differences among synucleinopathies except for one meta-analysis that showed lower values in MSA compared with PD.¹⁰¹ These findings indicate that CSF α -synuclein may be a possible biomarker of synucleinopathy (Table 1). However, lower α -synuclein concentrations were also reported in other conditions likely not related to α -synuclein misfolding, such as corticobasal syndrome and vascular parkinsonism, thus limiting the utility of CSF t- α -synuclein for discriminating PD from the atypical parkinsonisms.¹⁰³

Increased CSF t- α -synuclein levels were observed in AD and Creutzfeldt-Jakob disease (CJD),^{106,108-110} and,

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Candidate biomarker	Reference	Species	Method	Cohort	Main findings
CSF β-synuclein	Oeckl et al 2016 ¹⁰⁸	β-synuclein	MS	30 Controls, 19 PD, 13 PDD, 6 DLB, 18 AD, 6 CJD, 15 PSP, 7 CBS	 β-synuclein AD vs. controls β-synuclein in CJD vs. all other groups
	Oeckl et al 2020 ¹³⁴	β-synuclein	SW	110 Controls, 25 PD, 13 PDD/DLB, 145 AD, 25 CJD, 15 bvFTD, 30 ALS	↑ β-synuclein in AD vs. controls ↑ β-synuclein in CJD vs. all other groups
	Halbgebauer et al 2020 ¹³⁵	β-synuclein	ELISA	60 Controls, 46 LBD, 151 AD, 23 CJD, 29 bvFTD, 18 ALS	↑ β-synuclein in AD vs. controls ↑ β-synuclein in CJD vs. all other groups
Blood β-synuclein	Oeckl et al 2020 ¹³⁴	β-synuclein	SM	93 Controls, 25 PD, 13 PDD/DLB, 136 AD, 25 CJD, 10 bvFTD, 29 ALS	 β-synuclein in AD vs. controls β-synuclein in CJD vs. all other groups
CSF α-synuclein	Eusebi et al 2017 ¹⁰³	t-α-synuclein, o-α-synuclein, p-α-synuclein	ELISA, Luminex, TR-FRET, electro-chemiluminescence	Meta-analysis of 34 studies with a total of 1428 controls, 2070 PD, 396 DLB, 309 MSA, 259 PSP, 55 CBS, 22 VaP	\downarrow t- α -synuclein, \uparrow o- α -synuclein and \uparrow p- α -synuclein in PD vs. controls = t- α -synuclein in PD, DLB, MSA, PSP, CBS, and VaP
	Sako et al 2014 ¹⁰²	t-α-synuclein	ELISA, Luminex, TR-FRET	Meta-analysis of 9 studies with a total of 537 controls, 843 PD, 130 MSA, 98 PSP	<pre>L t-α-synuclein in PD vs. PSP and controls L t-α-synuclein in MSA vs. PSP and controls = t-α-synuclein in PD and MSA</pre>
	Zhou et al 2015 ¹⁰¹	t-α-synuclein o-α-synuclein	ELISA, Luminex, TR-FRET	Meta-analysis of 12 studies with a total of 783 Controls, 1131 PD, 192 DLB, 179 MSA, 92 PSP	
	Gao et al 2015 ¹⁰⁰	t-α-synuclein	ELISA, Luminex, MS, TR-FRET	Meta-analysis of 5 studies with a total of 399 controls, 412 PD, DLB,	↓ t-α-synuclein in PD vs. controls

(Continues)

Candidate biomarker	Reference	Species	Method	Cohort	Main findings
				148 AD, 31 FTD, 32 MSA	= t-α-synuclein in PD, DLB and MSA
Blood α-synuclein	Besong-Agbo et al 2013^{143}	t-α-synuclein	ELISA	46 Controls, 34 PD, 42 AD	= t- α -synuclein in PD, AD and controls
	Foulds et al 2013 ¹²⁰	t-α-synuclein p-α-synuclein	ELISA	91 Controls, 189 PD	 = t-α-synuclein in PD and controls ↑ p-α-synuclein in PD vs. controls
	Shi et al 2014 ¹⁴⁴	t-α-synuclein Exosomal α-synuclein	Luminex	215 Controls, 267 PD	 = t-α-synuclein in PD and controls ↑ Exosomal α-synuclein in PD vs. controls
	Ishii et al 2015 ¹⁴⁵	t-α-synuclein	ELISA	103 Controls, 53 PD	↓ t-α-synuclein in PD vs. controls
	Williams et al 2016 ¹¹⁹	o-α-synuclein	ELISA	5 controls, 9 PD, 6 AD	\uparrow o- $\alpha\text{-synuclein}$ in PD vs. AD and controls
	Ding et al 2017^{146}	t-α-synuclein	ELISA	23 Controls, 84 PD	f t-α-synuclein in PD vs. controls
	Lin et al 2017 ¹⁴⁷	t-α-synuclein	IMR	34 Controls, 80 PD	 t-α-synuclein in PD vs. controls t-α-synuclein in PDD vs. PD
	Vicente Miranda et al 2017 ¹²³	Y125-p-α-synuclein Y39-n-α-synuclein Glycated α-synuclein SUMOylated α-synuclein	Immunoblotting and densitometry	30 Controls, 58 PD	↑ Y125-p- α -synuclein, Y39-n- α -synuclein and glycated α -synuclein, and \downarrow SUMOylated α -synuclein in PD vs. controls
	Malec-Litwinowicz et al 2018 ¹⁴⁸	t-α-synuclein	ELISA	38 Controls, 58 PD	= t- α -synuclein in PD and controls
	Abd Elhadi et al 2019 ¹²¹	t-α-synuclein p-α-synuclein	ELISA	45 Controls, 32 PD, 14 PDD	 [†] t-α-synuclein in PD vs. PDD and controls [†] p-α-synuclein in PD vs. controls controls
	Chang et al 2020 ¹⁴⁹	$t-\alpha$ -synuclein	IMR	40 Controls, 48 PD	\uparrow t- α -synuclein in PD vs. controls
	Li et al 2021 ¹²²	Erythrocytic p-α-synuclein	ELISA	334 Controls, 333 PD	↑ p-α-synuclein in PD vs. controls

TABLE 1 Continued

	Reference	Species	Drug candidate	Cohort	Main findings
Clinical trials	Pagan et al 2016 ¹⁴⁰	CSF t-α-synuclein	Nilotinib	12 PDD/DLB	t-α-synuclein after 2 and 6 months with 150-mg nilotinib
	Pagan et al 2019 ¹⁴¹	CSF t-α-synuclein, CSF 0-α-synuclein	Nilotinib	75 PD	 = t-α-synuclein from 1 to 4 hours after drug administration ↓ o-α-synuclein 3 hours after administration of 400-mg nilotinib
	Shin et al 2019 ¹⁴²	CSF ο-α-synuclein, plasma ο-α-synuclein	KM-819	88 Healthy volunteers	= CSF and plasma o- α -synuclein 7 days after drug administration
	NCT03888222	Plasma t-α-synuclein, plasma o-α-synuclein	Bosutinib	30 DLB	Ongoing
	NCT03996460	CSF and plasma t-α-synuclein, CSF and plasma 0-α-synuclein	K0706	45 DLB	Ongoing
Abbreviations: AD, Alz Lewy bodies; ELISA, e disease; PDD, Parkinso	zheimer disease; ALS, amyotrophi nzyme-linked immunosorbent ass m's disease with dementia: PSP, 1	c lateral sclerosis; bvFTD, behavioral variant frontot say; IMR, immunomagnetic reduction; IBD, Lewy progresive supranuclear palsy; t-α-synuclein, total	temporal dementia; CBS, corti body disorders; MS, mass spec α-synuclein; p-α-synuclein, pl	:obasal syndrome; CJD, Creutzfeldt-J rometry; MSA, multiple system atroj oosphorylated α-synuclein at serine 1	akob disease; CSF, cerebrospinal fluid; DLB, dementia with hy; o-α-synuclein, oligometic α-synuclein; PD, Parkinson's 29: TR-FRET, time-resolved fluorescence energy transfer.

VaP, vascular parkinsonism; SUMO, small ubiquitin-like modifier; Y125-p-α-synuclein, phosphorylated α-synuclein at tyrosine 125; Y39-n-α-synuclein, nitrated α-synuclein at tyrosine 39.

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variably, even in progressive supranuclear palsy.^{101-103,111} These findings suggest a possible further role of α -synuclein as a marker of synaptic damage when its concentrations increase by being released from degenerating synapses. The two components of α -synuclein aggregation and synaptic derangement could variably influence α -synuclein measurements and partially justify the conflicting results reported. In this regard, longitudinal studies in PD have found increasing CSF t- α -synuclein concentrations during the disease course and described associations of t-α-synuclein with disease duration, total tau levels, and clinical motor scores.¹¹²⁻¹¹⁴ Higher t-α-synuclein values at baseline correlated with faster motor decline,¹¹³ whereas the predictive value of *a*-synuclein for cognitive decline in synucleinopathies has been inconsistent.99 A clear-cut interpretation is challenging because these studies have largely not been replicated¹¹⁵ and have shown some methodological concerns, such as the suboptimal CSF sample storage and the lack of standardized procedures. Moreover, the possible mechanisms underlying the longitudinal increase of t- α -synuclein in synucleinopathies, as well as its real meaning on a prognostic level, if it has any, are not vet clarified.

Together with t- α -synuclein concentrations, pathological species of α -synuclein, including S129-phosphorylated $(p-\alpha-synuclein),$ oligomeric $(o-\alpha-synuclein),$ and proaggregating forms, have been tested as potential diagnostic and prognostic biomarkers because they may more precisely identify the underlying synucleinopathy.¹¹⁶ CSF p- α -synuclein and o- α -synuclein levels were reported to be higher in PD than in controls,^{101,103} but their diagnostic accuracy, when considered singularly, is unsatisfactory for use in clinical practice.¹¹⁷ Better performances were instead obtained with combinations of α -synuclein species and other biomarkers, such as the o-a-synuclein/ t- α -synuclein ratio¹¹⁷ and the panel consisting of p- α -synuclein uclein, o-a-synuclein/t-a-synuclein ratio, and phosphorylated tau at threonine 181.118

Investigations of α -synuclein in peripheral blood have shown conflicting results, and the diagnostic utility of blood α -synuclein species for synucleinopathies should be further assessed before considering any routine clinical use. The main hampering issues concern data reproducibility and preanalytical confounders, such as the release of a-synuclein from blood cells.⁹⁹ Plasma and serum t- α -synuclein levels were reported to be either higher, lower, or not significantly altered in patients with PD compared with controls, whereas more concordant results were provided by assessing pathological forms of α -synuclein (Table 1). Increased values of p- α -synuclein and o- α -synuclein were found in the blood and erythrocytes of patients with PD compared with controls,¹¹⁹⁻¹²² but other posttranslationally modified species of α -synuclein have also been assessed.¹²³ Further studies on larger and longitudinally characterized cohorts will be of great help to verify the reliability and reproducibility of these preliminary findings.

As an extremely intriguing possibility, seed amplification assays (SAAs) are novel ultrasensitive protein amplification methods able to detect protein aggregates in biological samples.^{124,125} SAAs are already used with success for prion disease diagnostics and have now been implemented for identification of patients affected by synucleinopathy. In several works, SAAs showed excellent diagnostic performance in discriminating synucleinopathies from controls and from other neurodegenerative diseases since the prodromal disease stages.¹²⁶⁻¹²⁹ Recently, pathological aggregates from patients with PD and MSA could be distinguished in CSF by using a combination of biochemical and biophysical methods on the products of α -synuclein SAAs.¹³⁰ SAAs have also been applied successfully in other biological matrices, such as olfactory mucosa and skin biopsies, revealing high diagnostic accuracy in discriminating MSA from tauopathies and PD from controls, respectively.^{131,132} Further improvement in standardization and validation in different biological matrices is, however, required for the routine use of SAAs in clinical practice.^{124,133}

β -Synuclein as a Candidate Biomarker for Synaptic Degeneration

β-Synuclein has been assessed in a few studies as a body fluid biomarker for neurological disorders (Table 1). In 2016, the first quantitative data on CSF β -synuclein concentrations were provided by using an innovative mass spectrometry approach. No difference was found in patients with PD compared with controls. but PD with dementia (PDD) and DLB cases showed slightly higher β -synuclein values that became significantly increased when considering the β-synuclein/ α -synuclein ratio. This suggested that β -synuclein could be a potential biomarker of cognitive decline rather than of motor impairment in synucleinopathies.¹⁰⁸ Among neurodegenerative disorders other studied. CSF β -synuclein levels were found to be increased only in patients with CJD and AD.^{108,134} These results on CSF β-synuclein have been recently replicated with immunoassays, showing an excellent correlation with the antibody-free, mass spectrometry method.¹³⁵ Similarly to previous reports, only patients affected by AD and CJD, but not by synucleinopathies or other neurodegenerative disorders, showed increased CSF β -synuclein levels.¹³⁵

Overall, these findings indicate that CSF β -synuclein alone is not a reliable diagnostic biomarker for synucleinopathies but, rather, might be a candidate biomarker for synaptic degeneration given its localization in presynaptic terminals.¹³⁵ Moreover, the fact that β -synuclein is not expressed in blood cells makes it even more promising as a peripheral blood biomarker in comparison with α -synuclein and other synaptic proteins. For instance, neurogranin, which is one of the best characterized synaptic biomarkers, was repeatedly found to be elevated in the CSF of patients with AD, but similar changes were never observed in blood, probably because of the additional synthesis of this protein outside the CNS.¹³⁶ On the other hand, β -synuclein is only expressed in central neurons, and its blood levels more likely reflect synaptic degeneration. Accordingly, higher β -synuclein levels were found in the serum of patients with AD compared with healthy controls and other neurodegenerative diseases.¹³⁴ This finding has been confirmed in two validation cohorts.¹³⁴

 γ -Synuclein has been poorly investigated as a fluid biomarker for neurological disorders. CSF levels of γ -synuclein were measured in only a single study in patients affected by neurodegenerative diseases by quantitative mass spectrometry and showed increased levels in AD and CJD.¹⁰⁸ Interestingly, γ -synuclein is currently being tested as a potential fluid biomarker in brain and peripheral tumors.¹³⁷

Use of Synucleins as Biomarkers in Pharmacological Trials

In clinical trials for disease-modifying drugs, reliable biomarkers reflecting the underlying pathological process may help to enroll more homogeneous cohorts of patients and provide objective measures to better assess target engagement and outcome profile, thus improving the quality and the applicability of the results.¹³⁸ CSF α -synuclein has been used in clinical trials as an outcome measure, but the results are not yet available for all of them (Table 1).

Nilotinib is a protein kinase inhibitor being tested for PD and DLB to enhance intracellular degradation of α -synuclein via ALP.¹³⁹ In a clinical trial of efficacy, safety, and tolerability of nilotinib, 12 patients with PDD/DLB were randomized to receive either 150-mg or 300-mg nilotinib for 6 months. CSF was obtained at baseline and after 2 and 6 months of treatment. With respect to baseline levels, CSF t- α -synuclein was reduced at 2 and 6 months only in the 150-mg dosage group.¹⁴⁰ In a following trial of the pharmacokinetics and pharmacodynamics of nilotinib, 75 patients with PD were randomly assigned into five groups that received a single daily dose of placebo or 150-, 200-, 300-, or 400-mg nilotinib, with CSF collected 1 to 4 hours after drug administration. Nilotinib has been shown to enter the CNS in a dose-independent manner, and a single 200-mg dosage appeared optimal for impacting CSF biomarkers, including DA metabolites and α -synuclein. However, no changes were found in CSF t- α -synuclein, whereas $o-\alpha$ -synuclein levels were significantly reduced in the 400-mg group 3 hours after administration.¹⁴¹ In a more recent trial of KM-819, a Fas-associated factor 1 inhibitor, in healthy volunteers, CSF samples were collected at baseline and 7 days after the last drug

administration, and no changes were observed in CSF o- α -synuclein.¹⁴² A randomized, placebo-controlled study evaluating the safety and tolerability of bosutinib, another protein kinase inhibitor, in DLB started in 2019, and both plasma t- α -synuclein and o- α -synuclein were used as outcome measures (ClinicalTrials.gov identifier NCT03888222). Similarly, CSF and plasma levels of t- α -synuclein and o- α -synuclein are being used in an ongoing clinical trial of K0706, a tyrosine kinase inhibitor, in patients with DLB (ClinicalTrials.gov identifier NCT03996460) (Table 1).

The dual behavior of CSF t- α -synuclein as a marker of synucleinopathy and synaptic derangement poses relevant challenges for interpreting changes of its concentrations as an outcome measure and for verifying target engagement. To overcome these issues, CSF α -synuclein species could be combined with other synaptic biomarkers, such as β -synuclein, to better picture the synaptic dysfunction without being biased by α -synuclein pathology. Furthermore, given the promising results obtained in AD and prion disease, β -synuclein could be also used alone as a reliable outcome measure for clinical trials assessing novel drug candidates for preserving synaptic integrity.¹³⁵

So far, CSF t- α -synuclein has never been considered among the inclusion criteria for enrolling patients because of its low diagnostic performance in discriminating synucleinopathies from other neurodegenerative disorders. In this scenario, α -synuclein SAAs could be a reliable diagnostic test for the selection of patients thanks to their efficacy in identifying and partially differentiating synucleinopathies.¹³⁰ SAAs are primarily designed to produce a dichotomous result about the presence or absence of pathological α -synuclein, but because the seeding profiles depend on the amount of aggregated α -synuclein in biological samples, they might be suitable for monitoring the effects of drugs countering α -synuclein pathology.

The application of α -synuclein and β -synuclein as fluid biomarkers for clinical research is thus more than welcome, especially in association with more traditional markers including clinical scores and brain imaging. Their use in research would add valuable information with implications for patient selection and outcome monitoring in clinical trials. Nonetheless, several methodological issues, such as the standardization of quantitative procedures, should be addressed before these biomarkers can be used routinely in clinical practice.

Concluding Remarks and Future Perspectives

Most of the research on synucleins has so far mainly focused on α -synuclein pathophysiology in synucleinopathies. In recent years, we have begun to broaden our view to the whole synuclein family to define their physiological role in neurons, characterize their pathological involvement in neurological disorders, and most important, develop new diagnostic and therapeutic tools for such conditions.

At a physiological level, α -synuclein and β -synuclein seem to act as housekeeping proteins in presynaptic terminals that regulate synaptic homeostasis and transmission. However, they might have wider activities in neuronal and nonneuronal populations. At a pathological level, although α-synuclein is well known to aggregate, the modulatory properties attributed to β-synuclein are still not clear. On one hand, β -synuclein can mitigate α -synuclein-induced toxicity when coexpressed in certain conditions. On the other hand, β -synuclein also has an aggregation potential that can cause neurodegenerative features similar to those described for α -synuclein. It is possible that β -synuclein can interfere with the aggregation dynamics of α -synuclein but, at the same time, maintains the potential to aggregate under the appropriate conditions. In addition, β-synuclein-positive inclusions have been found in brain areas associated with cognition in patients with synucleinopathies and other neurodegenerative disorders, even without colocalizing with α -synuclein aggregates. Whether these features have an impact in vivo is still an open question. Although very recent and not numerous, investigations on CSF indicate that β-synuclein concentrations do not reflect an underlying synucleinopathy but, rather, may relate to ongoing synaptic degeneration. In this view, the association with β -synuclein could improve the diagnostic performance of α -synuclein given the dual behavior of the latter as a biomarker of synucleinopathy or of synaptic derangement. In addition, because β-synuclein is specifically expressed in neurons, both its CSF and blood concentrations might be reliably applied as a surrogate biomarker of synaptic damage in neurodegenerative disorders other than synucleinopathies, such as AD. The use of α -synuclein and β -synuclein as biomarkers in research settings and routine clinical practice requires further investigations to overcome methodological issues concerning their measurements, but the results so far are encouraging. Our knowledge about synuclein pathophysiology adds greater insight into the neurobiology of synucleinopathies and allows us to better understand the strengths and limitations of their potential application as fluid biomarkers.

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Data Availability Statement

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

References

- George JM. The synucleins. Genome Biol 2002;3(1):3002.1– 3002.6. https://doi.org/10.1186/gb-2001-3-1-reviews3002
- 2. Kalia LV, Lang AE. Parkinson's disease. Lancet 2015;386(9996): 896–912. https://doi.org/10.1016/S0140-6736(14)61393-3
- Goedert M, Jakes R, Spillantini MG. The synucleinopathies: twenty years on. J Parkinsons Dis 2017;7(S1):S51–S69. doi:https://doi. org/10.3233/JPD-179005
- Stefanis L. Alpha-synuclein in Parkinson's disease. Cold Spring Harb Perspect Med 2012;2(2):a009399. https://doi.org/10.1101/ cshperspect.a009399
- Barbour R, Kling K, Anderson JP, et al. Red blood cells are the major source of alpha-synuclein in blood. Neurodegener Dis 2008; 5(2):55–59. https://doi.org/10.1159/000112832
- Wilhelm BG, Mandad S, Truckenbrodt S, et al. Composition of isolated synaptic boutons reveals the amounts of vesicle trafficking proteins. Science 2014;344(6187):1023–1028. https://doi.org/10. 1126/science.1252884
- Chu Y, Kordower JH. Age-associated increases of alpha-synuclein in monkeys and humans are associated with nigrostriatal dopamine depletion: is this the target for Parkinson's disease? Neurobiol Dis 2007;25(1):134–149. https://doi.org/10.1016/j.nbd.2006.08.021
- 8. Murphy DD, Rueter SM, Trojanowski JQ, Lee VM. Synucleins are developmentally expressed, and alpha-synuclein regulates the size of the presynaptic vesicular pool in primary hippocampal neurons. J Neurosci 2000;20(9):3214–3220.
- Rideout HJ, Dietrich P, Savalle M, Dauer WT, Stefanis L. Regulation of alpha-synuclein by bFGF in cultured ventral midbrain dopaminergic neurons. J Neurochem 2003;84(4):803–813. https://doi. org/10.1046/j.1471-4159.2003.01574.x
- Beyer K, Munoz-Marmol AM, Sanz C, Marginet-Flinch R, Ferrer I, Ariza A. New brain-specific beta-synuclein isoforms show expression ratio changes in Lewy body diseases. Neurogenetics 2012;13(1):61–72. https://doi.org/10.1007/s10048-011-0311-8
- Ji H, Liu YE, Jia T, et al. Identification of a breast cancer-specific gene, BCSG1, by direct differential cDNA sequencing. Cancer Res 1997;57(4):759–764.
- Buchman VL, Adu J, Pinon LG, Ninkina NN, Davies AM. Persyn, a member of the synuclein family, influences neurofilament network integrity. Nat Neurosci 1998;1(2):101–103. https://doi.org/ 10.1038/349
- Villar-Piqué A, Lopes da Fonseca T, Outeiro TF. Structure, function and toxicity of alpha-synuclein: the Bermuda triangle in synucleinopathies. J Neurochem 2016;139:240–255. https://doi.org/ 10.1111/jnc.13249
- Landau M. Getting in charge of β-synuclein fibrillation. J Biol Chem 2017;292(39):16380–16381. https://doi.org/10.1074/jbc. H117.780528
- Vamvaca K, Volles MJ, Lansbury PT. The first N-terminal amino acids of α-synuclein are essential for α-helical structure formation in vitro and membrane binding in yeast. J Mol Biol 2009;389(2): 413–424. https://doi.org/10.1016/j.jmb.2009.03.021
- 16. Bertoncini CW, Rasia RM, Lamberto GR, et al. Structural characterization of the intrinsically unfolded protein β -synuclein, a natural negative regulator of α -synuclein aggregation. J Mol Biol 2007; 372(3):708–722. https://doi.org/10.1016/j.jmb.2007.07.009
- Williams JK, Yang X, Baum J. Interactions between the intrinsically disordered proteins β-synuclein and α-synuclein. Proteomics 2018; 18(21–22):e1800109. https://doi.org/10.1002/pmic.201800109
- Breydo L, Wu J, Uversky V. α-Synuclein misfolding and Parkinson's disease. Biochim Biophys Acta 2012;1822:261–285. https://doi.org/10.1016/j.bbadis.2011.10.002
- Uversky VN, Li J, Souillac P, et al. Biophysical properties of the synucleins and their propensities to fibrillate: inhibition of alpha-synuclein assembly by beta- and gamma-synucleins. J Biol Chem 2002;277(14): 11970–11978. https://doi.org/10.1074/jbc.M109541200
- Janowska MK, Baum J. The loss of inhibitory C-terminal conformations in disease associated P123H β-synuclein. Protein Sci 2016; 25(1):286–294. https://doi.org/10.1002/pro.2798

- Sharma K, Mehra S, Sawner AS, et al. Effect of disease-associated P123H and V70M mutations on β-synuclein fibrillation. ACS Chem Nerosci 2020;11(18):2836–2848. https://doi.org/10.1021/ acschemneuro.0c00405
- Ducas VC, Rhoades E. Quantifying interactions of β-synuclein and γ-synuclein with model membranes. J Mol Biol 2012;423(4):528– 539. https://doi.org/10.1016/j.jmb.2012.08.008
- Bartels T, Choi JG, Selkoe DJ. α-Synuclein occurs physiologically as a helically folded tetramer that resists aggregation. Nature 2011; 477(7362):107–110. https://doi.org/10.1038/nature10324
- Dettmer U, Newman AJ, Soldner F, et al. Parkinson-causing α-synuclein missense mutations shift native tetramers to monomers as a mechanism for disease initiation. Nat Commun 2015;6(1): 7314. https://doi.org/10.1038/ncomms8314
- Abeliovich A, Schmitz Y, Fariñas I, et al. Mice lacking alphasynuclein display functional deficits in the nigrostriatal dopamine system. Neuron 2000;25(1):239–252. https://doi.org/10.1016/ s0896-6273(00)80886-7
- Cabin DE, Shimazu K, Murphy D, et al. Synaptic vesicle depletion correlates with attenuated synaptic responses to prolonged repetitive stimulation in mice lacking alpha-synuclein. J Neurosci 2002; 22(20):8797–8807.
- Al-Wandi A, Ninkina N, Millership S, Williamson SJM, Jones PA, Buchman VL. Absence of alpha-synuclein affects dopamine metabolism and synaptic markers in the striatum of aging mice. Neurobiol Aging 2010;31(5):796–804. https://doi.org/10.1016/j. neurobiolaging.2008.11.001
- Chandra S, Fornai F, Kwon HB, et al. Double-knockout mice for α- and β-synucleins: effect on synaptic functions. Proc Natl Acad Sci U S A 2004;101(41):14966–14971. https://doi.org/10.1073/ pnas.0406283101
- Robertson DC, Schmidt O, Ninkina N, Jones PA, Sharkey J, Buchman VL. Developmental loss and resistance to MPTP toxicity of dopaminergic neurones in substantia nigra pars compacta of gamma-synuclein, alpha-synuclein and double alpha/gammasynuclein null mutant mice. J Neurochem 2004;89(5):1126–1136. https://doi.org/10.1111/j.1471-4159.2004.02378.x
- Greten-Harrison B, Polydoro M, Morimoto-Tomita M, et al. αβγ-Synuclein triple knockout mice reveal age-dependent neuronal dysfunction. Proc Natl Acad Sci U S A 2010;107(45):19573– 19578. https://doi.org/10.1073/pnas.1005005107
- 31. Burré J. The synaptic function of $\alpha\mbox{-synuclein}.$ J Parkinsons Dis 2015;5(4):699–713. https://doi.org/10.3233/JPD-150642
- Westphal CH, Chandra SS. Monomeric Synucleins generate membrane curvature. J Biol Chem 2013;288(3):1829–1840. https://doi. org/10.1074/jbc.M112.418871
- Perez RG, Waymire JC, Lin E, Liu JJ, Guo F, Zigmond MJ. A role for alpha-synuclein in the regulation of dopamine biosynthesis. J Neurosci 2002;22(8):3090–3099.
- Tehranian R, Montoya SE, Van Laar AD, Hastings TG, Perez RG. Alpha-synuclein inhibits aromatic amino acid decarboxylase activity in dopaminergic cells. J Neurochem 2006;99(4):1188–1196. https://doi.org/10.1111/j.1471-4159.2006.04146.x
- Ryskalin L, Busceti CL, Limanaqi F, Biagioni F, Gambardella S, Fornai F. A focus on the beneficial effects of alpha synuclein and a re-appraisal of synucleinopathies. 2018;19(6):598–611. Curr Protein Pept Sci, https://doi.org/10.2174/1389203718666171117110028
- Bellucci A, Navarria L, Falarti E, et al. Redistribution of DAT/αsynuclein complexes visualized by "in situ" proximity ligation assay in transgenic mice modelling early Parkinson's disease. PLoS One 2011;6(12):e27959. https://doi.org/10.1371/journal.pone.0027959
- Haining R, Achat-Mendes C. Neuromelanin, one of the most overlooked molecules in modern medicine, is not a spectator. Neural Regen Res 2017;12(3):372. https://doi.org/10.4103/1673-5374. 202928
- Muñoz P, Huenchuguala S, Paris I, Segura-Aguilar J. Dopamine oxidation and autophagy. Parkinsons Dis 2012;2012:920953. https://doi.org/10.1155/2012/920953
- Raina A, Leite K, Guerin S, et al. Dopamine promotes the neurodegenerative potential of β-synuclein. J Neurochem 2021;156(5):674– 691. https://doi.org/10.1111/jnc.15134

- Shahmoradian SH, Lewis AJ, Genoud C, et al. Lewy pathology in Parkinson's disease consists of crowded organelles and lipid membranes. Nat Neurosci 2019;22(7):1099–1109. https://doi.org/10. 1038/s41593-019-0423-2
- Elkon H, Don J, Melamed E, Ziv I, Shirvan A, Offen D. Mutant and wild-type alpha-synuclein interact with mitochondrial cytochrome C oxidase. J Mol Neurosci 2002;18(3):229–238. https:// doi.org/10.1385/JMN:18:3:229
- Ludtmann MHR, Angelova PR, Ninkina NN, Gandhi S, Buchman VL, Abramov AY. Monomeric alpha-Synuclein exerts a physiological role on brain ATP synthase. J Neurosci 2016;36(41): 10510–10521. https://doi.org/10.1523/JNEUROSCI.1659-16.2016
- Shaltouki A, Hsieh CH, Kim MJ, Wang X. Alpha-synuclein delays mitophagy and targeting Miro rescues neuron loss in Parkinson's models. Acta Neuropathol 2018;136(4):607–620. https://doi.org/ 10.1007/s00401-018-1873-4
- 44. Sousa VL, Bellani S, Giannandrea M, et al. α-Synuclein and its A30P mutant affect Actin cytoskeletal structure and dynamics. Mol Biol Cell 2009;20(16):3725–3739. https://doi.org/10.1091/mbc. E08-03-0302
- Lee HJ, Lee K, Im H. α-Synuclein modulates neurite outgrowth by interacting with SPTBN1. Biochem Biophys Res Commun 2012; 424(3):497–502. https://doi.org/10.1016/j.bbrc.2012.06.143
- 46. Ordonez DG, Lee MK, Feany MB. α-Synuclein induces mitochondrial dysfunction through Spectrin and the Actin cytoskeleton. Neuron 2018;97(1):108–124.e6. https://doi.org/10.1016/j.neuron. 2017.11.036
- Cartelli D, Aliverti A, Barbiroli A, et al. α-Synuclein is a novel microtubule dynamase. Sci Rep 2016;6(1):33289. https://doi.org/ 10.1038/srep33289
- Chen L, Jin J, Davis J, et al. Oligomeric α-synuclein inhibits tubulin polymerization. Biochem Biophys Res Commun 2007;356(3):548– 553. https://doi.org/10.1016/j.bbrc.2007.02.163
- Zhou RM, Huang YX, Li XL, et al. Molecular interaction of α-synuclein with tubulin influences on the polymerization of microtubule in vitro and structure of microtubule in cells. Mol Biol Rep 2010;37(7):3183–3192. https://doi.org/10.1007/s11033-009-9899-2
- Prots I, Veber V, Brey S, et al. α-Synuclein oligomers impair neuronal microtubule-kinesin interplay. J Biol Chem 2013;288(30): 21742–21754. https://doi.org/10.1074/jbc.M113.451815
- Duka T, Duka V, Joyce JN, Sidhu A. α-Synuclein contributes to GSK-3β-catalyzed tau phosphorylation in Parkinson's disease models. FASEB J 2009;23(9):2820–2830. https://doi.org/10.1096/ fj.08-120410
- Gąssowska M, Czapski GA, Pająk B, Cieślik M, Lenkiewicz AM, Adamczyk A. Extracellular α-synuclein leads to microtubule destabilization via GSK-3β-dependent tau phosphorylation in PC12 cells. PLoS One 2014;9(4):e94259. https://doi.org/10.1371/journal. pone.0094259
- Delic V, Chandra S, Abdelmotilib H, et al. Sensitivity and specificity of Phospho-Ser129 α-synuclein monoclonal antibodies. J Comp Neurol 2018;526(12):1978–1990. https://doi.org/10.1002/cne. 24468
- Pinho R, Paiva I, Jerčić KG, et al. Nuclear localization and phosphorylation modulate pathological effects of alpha-synuclein. Hum Mol Genet 2019;28(1):31–50. https://doi.org/10.1093/hmg/ddy326
- 55. Fujiwara H, Hasegawa M, Dohmae N, et al. Alpha-synuclein is phosphorylated in synucleinopathy lesions. Nat Cell Biol 2002; 4(2):160–164. https://doi.org/10.1038/ncb748
- Paiva I, Pinho R, Pavlou MA, et al. Sodium butyrate rescues dopaminergic cells from alpha-synuclein-induced transcriptional deregulation and DNA damage. Hum Mol Genet 2017;26(12):2231– 2246. https://doi.org/10.1093/hmg/ddx114
- 57. Vasudevaraju P, Guerrero E, Hegde ML, Collen TB, Britton GB, Rao KS. New evidence on α-synuclein and tau binding to conformation and sequence specific GC* rich DNA: relevance to neurological disorders. J Pharm Bioallied Sci 2012;4(2):112–117. https:// doi.org/10.4103/0975-7406.94811
- Kontopoulos E, Parvin JD, Feany MB. Alpha-synuclein acts in the nucleus to inhibit histone acetylation and promote neurotoxicity.

Hum Mol Genet 2006;15(20):3012-3023. https://doi.org/10.1093/hmg/ddl243

- Outeiro TF, Kontopoulos E, Altmann SM, et al. Sirtuin 2 inhibitors rescue alpha-synuclein-mediated toxicity in models of Parkinson's disease. Science 2007;317(5837):516–519. https://doi.org/10.1126/ science.1143780
- Garcia-Reitboeck P, Anichtchik O, Dalley JW, et al. Endogenous alpha-synuclein influences the number of dopaminergic neurons in mouse substantia nigra. Exp Neurol 2013;248:541–545. https:// doi.org/10.1016/j.expneurol.2013.07.015
- Swaminathan M, Fung C, Finkelstein DI, Bornstein JC, Foong JPP. α-Synuclein regulates development and function of cholinergic enteric neurons in the mouse colon. Neuroscience 2019;423:76–85. https://doi.org/10.1016/j.neuroscience.2019.10.029
- Witt SN. Molecular chaperones, α-synuclein, and neurodegeneration. Mol Neurobiol 2013;47(2):552–560. https://doi. org/10.1007/s12035-012-8325-2
- Belaidi AA, Bush AI. Iron neurochemistry in Alzheimer's disease and Parkinson's disease: targets for therapeutics. J Neurochem 2016;139((suppl 1)):179–197. https://doi.org/10.1111/jnc.13425
- Davies P, Moualla D, Brown DR. Alpha-synuclein is a cellular ferrireductase. PLoS One 2011;6(1):e15814. https://doi.org/10.1371/ journal.pone.0015814
- Tashkandi H, Shameli A, Harding CV, Maitta RW. Ultrastructural changes in peripheral blood leukocytes in α-synuclein knockout mice. Blood Cells Mol Dis 2018;73:33–37. https://doi.org/10.1016/ j.bcmd.2018.09.001
- Rodriguez-Araujo G, Nakagami H, Takami Y, et al. Low alphasynuclein levels in the blood are associated with insulin resistance. Sci Rep 2015;5:12081. https://doi.org/10.1038/srep12081
- Araki K, Sugawara K, Hayakawa EH, et al. The localization of α-synuclein in the process of differentiation of human erythroid cells. Int J Hematol 2018;108(2):130–138. https://doi.org/10.1007/ s12185-018-2457-8
- Hashimoto M, Yoshimoto M, Sisk A, et al. NACP, a synaptic protein involved in Alzheimer's disease, is differentially regulated during megakaryocyte differentiation. Biochem Biophys Res Commun 1997;237(3):611–616. https://doi.org/10.1006/bbrc.1997.6978
- Pei Y, Maitta RW. Alpha synuclein in hematopoiesis and immunity. Heliyon 2019;5(10):e02590. https://doi.org/10.1016/j.heliyon. 2019.e02590
- Liu Y, Tapia ML, Yeh J, He RC, Pomerleu D, Lee RK. Differential gamma-synuclein expression in acute and chronic retinal ganglion cell death in the retina and optic nerve. Mol Neurobiol 2020;57(2): 698–709. https://doi.org/10.1007/s12035-019-01735-1
- Liu H, Mercieca K, Anders F, Prokosch V. Hydrogen sulfide and β-synuclein are involved and interlinked in the aging glaucomatous retina. J Ophthalmol 2020;2020:8642135. https://doi.org/10.1155/ 2020/8642135
- Fujita M, Sekigawa A, Sekiyama K, Sugama S, Hashimoto M. Neurotoxic conversion of beta-synuclein: a novel approach to generate a transgenic mouse model of synucleinopathies? J Neurol 2009;256((suppl 3)):286–292. https://doi.org/10.1007/s00415-009-5246-8
- Abbineni PS, Bohannon KP, Bittner MA, Axelrod D, Holz RW. Identification of β-synuclein on secretory granules in chromaffin cells and the effects of α- and β-synuclein on post-fusion BDNF discharge and fusion pore expansion. Neurosci Lett 2019;699:134– 139. https://doi.org/10.1016/j.neulet.2019.01.056
- 74. Surgucheva I, McMahon B, Surguchov A. γ-Synuclein has a dynamic intracellular localization. Cell Motil 2006;63(8):447–458. https://doi.org/10.1002/cm.20135
- Fung KM, Rorke LB, Giasson B, Lee VMY, Trojanowski JQ. Expression of alpha-, beta-, and gamma-synuclein in glial tumors and medulloblastomas. Acta Neuropathol 2003;106(2):167–175. https://doi.org/10.1007/s00401-003-0718-x
- Millership S, Ninkina N, Rochford JJ, Buchman VL. γ-Synuclein is a novel player in the control of body lipid metabolism. Adipocyte 2013;2(4):276–280. https://doi.org/10.4161/adip.25162

- 77. Kanaan NM, Manfredsson FP. Loss of functional alpha-synuclein: a toxic event in Parkinson's disease? J Parkinsons Dis 2012;2(4): 249–267. https://doi.org/10.3233/JPD-012138
- 78. Cascella R, Chen SW, Bigi A, et al. The release of toxic oligomers from α -synuclein fibrils induces dysfunction in neuronal cells. Nat Commun 2021;12(1):1814. https://doi.org/10.1038/s41467-021-21937-3
- Bellomo G, Paciotti S, Gatticchi L, Parnetti L. The vicious cycle between α-synuclein aggregation and autophagic-lysosomal dysfunction. Mov Disord 2020;35(1):34–44. https://doi.org/10.1002/ mds.27895
- Tsigelny IF, Bar-On P, Sharikov Y, et al. Dynamics of alphasynuclein aggregation and inhibition of pore-like oligomer development by beta-synuclein. FEBS J 2007;274(7):1862–1877. https:// doi.org/10.1111/j.1742-4658.2007.05733.x
- Yang X, Williams JK, Yan R, Mouradian MM, Baum J. Increased dynamics of α-Synuclein fibrils by β-synuclein leads to reduced seeding and cytotoxicity. Sci Rep 2019;9(1):17579. https://doi.org/ 10.1038/s41598-019-54063-8
- Lee D, Paik SR, Choi KY. Beta-synuclein exhibits chaperone activity more efficiently than alpha-synuclein. FEBS Lett 2004;576(1–2): 256–260. https://doi.org/10.1016/j.febslet.2004.08.075
- Evans T, Kok WL, Cowan K, Hefford M, Anichtchik O. Accumulation of beta-synuclein in cortical neurons is associated with autophagy attenuation in the brains of dementia with Lewy body patients. Brain Res 2018;1681:1–13. https://doi.org/10.1016/j. brainres.2017.12.026
- Taschenberger G, Toloe J, Tereshchenko J, et al. β-Synuclein aggregates and induces neurodegeneration in dopaminergic neurons. Ann Neurol 2013;74(1):109–118. https://doi.org/10.1002/ana. 23905
- Ducas VC, Rhoades E. Investigation of intramolecular dynamics and conformations of α-, β- and γ-synuclein. PLoS One 2014;9(1): e86983. https://doi.org/10.1371/journal.pone.0086983
- Goedert M, Spillantini MG, Del Tredici K, Braak H. 100 years of Lewy pathology. Nat Rev Neurol 2013;9(1):13–24. https://doi.org/ 10.1038/nrneurol.2012.242
- Pan T, Zhu J, Hwu WJ, Jankovic J. The role of alpha-synuclein in melanin synthesis in melanoma and dopaminergic neuronal cells. PLoS One 2012;7(9):e45183. https://doi.org/10.1371/journal.pone. 0045183
- Galvin JE, Uryu K, Lee VM, Trojanowski JQ. Axon pathology in Parkinson's disease and Lewy body dementia hippocampus contains alpha-, beta-, and gamma-synuclein. Proc Natl Acad Sci U S A 1999;96(23):13450–13455. https://doi.org/10.1073/pnas.96.23. 13450
- Mori F, Nishie M, Yoshimoto M, Takahashi H, Wakabayashi K. Reciprocal accumulation of beta-synuclein in alpha-synuclein lesions in multiple system atrophy. Neuroreport 2003;14(14): 1783–1786. https://doi.org/10.1097/00001756-200310060-00005
- Lodygin D, Hermann M, Schweingruber N, et al. β-Synucleinreactive T cells induce autoimmune CNS grey matter degeneration. Nature 2019;566(7745):503–508. https://doi.org/10.1038/s41586-019-0964-2
- Surgucheva I, Newell KL, Burns J, Surguchov A. New α- and γ-synuclein immunopathological lesions in human brain. Acta Neuropathol Commun 2014;2:132. https://doi.org/10.1186/s40478-014-0132-8
- 92. Peters OM, Shelkovnikova T, Highley JR, et al. Gamma-synuclein pathology in amyotrophic lateral sclerosis. Ann Clin Transl Neurol 2015;2(1):29–37. https://doi.org/10.1002/acn3.143
- Postuma RB, Berg D, Stern M, et al. MDS clinical diagnostic criteria for Parkinson's disease: MDS-PD clinical diagnostic criteria. Mov Disord 2015;30(12):1591–1601. https://doi.org/10. 1002/mds.26424
- McKeith IG, Boeve BF, Dickson DW, et al. Diagnosis and management of dementia with Lewy bodies: fourth consensus report of the DLB consortium. Neurology 2017;89(1):88–100. https://doi.org/ 10.1212/WNL.00000000004058

- Tolosa E, Wenning G, Poewe W. The diagnosis of Parkinson's disease. Lancet Neurol 2006;5(1):75–86. https://doi.org/10.1016/ S1474-4422(05)70285-4
- Jack CR, Bennett DA, Blennow K, et al. NIA-AA research framework: toward a biological definition of Alzheimer's disease. Alzheimers Dement 2018;14(4):535–562. https://doi.org/10.1016/j. jalz.2018.02.018
- Fodero-Tavoletti MT, Mulligan RS, Okamura N, et al. In vitro characterisation of BF227 binding to alpha-synuclein/Lewy bodies. Eur J Pharmacol 2009;617(1–3):54–58. https://doi.org/10.1016/j. ejphar.2009.06.042
- Brooks DJ, Tambasco N. Imaging synucleinopathies. Mov Disord 2016;31(6):814–829. https://doi.org/10.1002/mds.26547
- 99. Parnetti L, Gaetani L, Eusebi P, et al. CSF and blood biomarkers for Parkinson's disease. Lancet Neurol 2019;18(6):573–586. https://doi.org/10.1016/S1474-4422(19)30024-9
- 100. Gao L, Tang H, Nie K, et al. Cerebrospinal fluid alpha-synuclein as a biomarker for Parkinson's disease diagnosis: a systematic review and meta-analysis. Int J Neurosci 2015;125(9):645–654. https://doi.org/10.3109/00207454.2014.961454
- 101. Zhou B, Wen M, Yu WF, Zhang CL, Jiao L. The diagnostic and differential diagnosis utility of cerebrospinal fluid α-synuclein levels in Parkinson's disease: a meta-analysis. Parkinsons Dis 2015;2015: 1–11. https://doi.org/10.1155/2015/567386
- 102. Sako W, Murakami N, Izumi Y, Kaji R. Reduced alpha-synuclein in cerebrospinal fluid in synucleinopathies: evidence from a metaanalysis: reduced alpha-synuclein in synucleinopathies. Mov Disord 2014;29(13):1599–1605. https://doi.org/10.1002/mds.26036
- 103. Eusebi P, Giannandrea D, Biscetti L, et al. Diagnostic utility of cerebrospinal fluid α -synuclein in Parkinson's disease: a systematic review and meta-analysis. Mov Disord 2017;32(10):1389–1400. https://doi.org/10.1002/mds.27110
- 104. Parnetti L, Chiasserini D, Bellomo G, et al. Cerebrospinal fluid tau/α-synuclein ratio in Parkinson's disease and degenerative dementias. Mov Disord 2011;26(8):1428–1435. https://doi.org/10. 1002/mds.23670
- 105. Simonsen AH, Kuiperij B, El-Agnaf OMA, et al. The utility of α -synuclein as biofluid marker in neurodegenerative diseases: a systematic review of the literature. Biomark Med 2016;10(1):19–34. https://doi.org/10.2217/BMM.14.105
- 106. Wennström M, Surova Y, Hall S, et al. Low CSF levels of both α -synuclein and the α -synuclein cleaving enzyme neurosin in patients with synucleinopathy. PLoS One 2013;8(1):e53250. https://doi.org/10.1371/journal.pone.0053250
- 107. Magdalinou NK, Paterson RW, Schott JM, et al. A panel of nine cerebrospinal fluid biomarkers may identify patients with atypical parkinsonian syndromes. J Neurol Neurosurg Psychiatry 2015; 86(11):1240–1247. https://doi.org/10.1136/jnnp-2014-309562
- 108. Oeckl P, Metzger F, Nagl M, et al. Alpha-, beta-, and gammasynuclein quantification in cerebrospinal fluid by multiple reaction monitoring reveals increased concentrations in Alzheimer's and Creutzfeldt-Jakob disease but no alteration in synucleinopathies. Mol Cell Proteomics 2016;15(10):3126–3138. https://doi.org/10. 1074/mcp.M116.059915
- 109. Hall S, Öhrfelt A, Constantinescu R, et al. Accuracy of a panel of 5 cerebrospinal fluid biomarkers in the differential diagnosis of patients with dementia and/or parkinsonian disorders. Arch Neurol 2012;69(11):1445–1452. https://doi.org/10.1001/archneurol.2012. 1654
- 110. Tateno F, Sakakibara R, Kawai T, Kishi M, Murano T. Alphasynuclein in the cerebrospinal fluid differentiates synucleinopathies (Parkinson disease, dementia with Lewy bodies, multiple system atrophy) from Alzheimer disease. Alzheimer Dis Assoc Disord 2012; 26(3):213–216. https://doi.org/10.1097/WAD.0b013e31823899cc
- 111. Bäckström DC, Eriksson Domellöf M, Linder J, et al. Cerebrospinal fluid patterns and the risk of future dementia in early, incident Parkinson disease. JAMA Neurol 2015;72(10):1175–1182. https:// doi.org/10.1001/jamaneurol.2015.1449
- 112. Majbour NK, Vaikath NN, Eusebi P, et al. Longitudinal changes in CSF alpha-synuclein species reflect Parkinson's disease

progression. Mov Disord 2016;31(10):1535–1542. https://doi.org/ 10.1002/mds.26754

- Hall S, Surova Y, Öhrfelt A, Zetterberg H, Lindqvist D, Hansson O. CSF biomarkers and clinical progression of Parkinson disease. Neurology 2015;84(1):57–63. https://doi.org/10.1212/ WNL.0000000000001098
- 114. Hall S, Surova Y, Öhrfelt A, et al. Longitudinal measurements of cerebrospinal fluid biomarkers in Parkinson's disease. Mov Disord 2016;31(6):898–905. https://doi.org/10.1002/mds.26578
- Mollenhauer B, Caspell-Garcia CJ, Coffey CS, et al. Longitudinal analyses of cerebrospinal fluid α-Synuclein in prodromal and early Parkinson's disease. Mov Disord 2019;34(9):1354–1364. https:// doi.org/10.1002/mds.27806
- Parnetti L, Cicognola C, Eusebi P, Chiasserini D. Value of cerebrospinal fluid α-synuclein species as biomarker in Parkinson's diagnosis and prognosis. Biomark Med 2016;10(1):35–49. https://doi.org/ 10.2217/bmm.15.107
- Parnetti L, Farotti L, Eusebi P, et al. Differential role of CSF alphasynuclein species, tau, and Aβ42 in Parkinson's disease. Front Aging Neurosci 2014;6:53. https://doi.org/10.3389/fnagi.2014. 00053
- 118. Majbour NK, Vaikath NN, van Dijk KD, et al. Oligomeric and phosphorylated alpha-synuclein as potential CSF biomarkers for Parkinson's disease. Mol Neurodegener 2016;11:7. https://doi.org/ 10.1186/s13024-016-0072-9
- Williams SM, Schulz P, Sierks MR. Oligomeric α-synuclein and β-amyloid variants as potential biomarkers for Parkinson's and Alzheimer's diseases. Eur J Neurosci 2016;43(1):3–16. https://doi. org/10.1111/ejn.13056
- 120. Foulds PG, Diggle P, Mitchell JD, et al. A longitudinal study on α -synuclein in blood plasma as a biomarker for Parkinson's disease. Sci Rep 2013;3:2540. https://doi.org/10.1038/srep02540
- 121. Abd Elhadi S, Grigoletto J, Poli M, Arosio P, Arkadir D, Sharon R. α-Synuclein in blood cells differentiates Parkinson's disease from healthy controls. Ann Clin Transl Neurol 2019;6(12):2426–2436. https://doi.org/10.1002/acn3.50944
- 122. Li XY, Li W, Li X, et al. Alterations of erythrocytic phosphorylated alpha-synuclein in different subtypes and stages of Parkinson's disease. Front Aging Neurosci 2021;13:623977. https://doi.org/10.3389/fnagi.2021.623977
- 123. Vicente Miranda H, Cássio R, Correia-Guedes L, et al. Posttranslational modifications of blood-derived alpha-synuclein as biochemical markers for Parkinson's disease. Sci Rep 2017;7(1):13713. https://doi.org/10.1038/s41598-017-14175-5
- 124. Paciotti S, Bellomo G, Gatticchi L, Parnetti L. Are we ready for detecting α-Synuclein prone to aggregation in patients? The case of "protein-Misfolding cyclic amplification" and "real-time quakinginduced conversion" as diagnostic tools. Front Neurol 2018;9:415. https://doi.org/10.3389/fneur.2018.00415
- 125. Russo MJ, Orru CD, Concha-Marambio L, et al. High diagnostic performance of independent alpha-synuclein seed amplification assays for detection of early Parkinson's disease. Acta Neuropathol Commun 2021;9(1):179. https://doi.org/10.1186/s40478-021-01282-8
- Fairfoul G, McGuire LI, Pal S, et al. Alpha-synuclein RT-QuIC in the CSF of patients with alpha-synucleinopathies. Ann Clin Transl Neurol 2016;3(10):812–818. https://doi.org/10.1002/acn3.338
- 127. Shahnawaz M, Tokuda T, Waragai M, et al. Development of a biochemical diagnosis of Parkinson disease by detection of α -synuclein misfolded aggregates in cerebrospinal fluid. JAMA Neurol 2017;74(2):163–172. https://doi.org/10.1001/jamaneurol. 2016.4547
- 128. Groveman BR, Orrù CD, Hughson AG, et al. Rapid and ultrasensitive quantitation of disease-associated α-synuclein seeds in brain and cerebrospinal fluid by αSyn RT-QuIC. Acta Neuropathol Commun 2018;6(1):7. https://doi.org/10.1186/s40478-018-0508-2
- 129. Kang UJ, Boehme AK, Fairfoul G, et al. Comparative study of cerebrospinal fluid α-synuclein seeding aggregation assays for diagnosis of Parkinson's disease: CSF α-synuclein seeding aggregation for PD diagnosis. Mov Disord 2019;34(4):536–544. https://doi.org/10. 1002/mds.27646

- Shahnawaz M, Mukherjee A, Pritzkow S, et al. Discriminating α-synuclein strains in Parkinson's disease and multiple system atrophy. Nature 2020;578(7794):273–277. https://doi.org/10.1038/ s41586-020-1984-7
- 131. De Luca CMG, Elia AE, Portaleone SM, et al. Efficient RT-QuIC seeding activity for α-synuclein in olfactory mucosa samples of patients with Parkinson's disease and multiple system atrophy. Transl Neurodegener 2019;8:8–24. https://doi.org/10.1186/ s40035-019-0164-x
- 132. Wang Z, Becker K, Donadio V, et al. Skin α-synuclein aggregation seeding activity as a novel biomarker for Parkinson disease. JAMA Neurol 2021;78(1):1–11. https://doi.org/10.1001/jamaneurol.2020. 3311
- 133. Bellomo G, Bologna S, Cerofolini L, et al. Dissecting the interactions between human serum albumin and α-synuclein: new insights on the factors influencing α-synuclein aggregation in biological fluids. J Phys Chem B 2019;123(20):4380–4386. https://doi.org/10. 1021/acs.jpcb.9b02381
- 134. Oeckl P, Halbgebauer S, Anderl-Straub S, et al. Targeted mass spectrometry suggests Beta-Synuclein as synaptic blood marker in Alzheimer's disease. J Proteome Res 2020;19(3):1310–1318. https://doi.org/10.1021/acs.jproteome.9b00824
- Halbgebauer S, Oeckl P, Steinacker P, et al. Beta-synuclein in cerebrospinal fluid as an early diagnostic marker of Alzheimer's disease. J Neurol Neurosurg Psychiatry 2020;92:349–356. https://doi. org/10.1136/jnnp-2020-324306
- 136. De Vos A, Jacobs D, Struyfs H, et al. C-terminal neurogranin is increased in cerebrospinal fluid but unchanged in plasma in Alzheimer's disease. Alzheimers Dement 2015;11(12):1461–1469. https://doi.org/10.1016/j.jalz.2015.05.012
- Winham CL, Le T, Jellison ER, Silver AC, Levesque AA, Koob AO. γ-Synuclein induces human cortical astrocyte proliferation and subsequent BDNF expression and release. Neuroscience 2019;410:41– 54. https://doi.org/10.1016/j.neuroscience.2019.04.057
- Gaetani L, Paolini Paoletti F, Bellomo G, et al. CSF and blood biomarkers in neuroinflammatory and neurodegenerative diseases: implications for treatment. Trends Pharmacol Sci 2020;41(12): 1023–1037. https://doi.org/10.1016/j.tips.2020.09.011
- 139. Karuppagounder SS, Brahmachari S, Lee Y, Dawson VL, Dawson TM, Ko HS. The c-Abl inhibitor, nilotinib, protects dopaminergic neurons in a preclinical animal model of Parkinson's disease. Sci Rep 2014;4:4874. https://doi.org/10.1038/srep04874
- 140. Pagan F, Hebron M, Valadez EH, et al. Nilotinib effects in Parkinson's disease and dementia with Lewy bodies. J Parkinsons Dis 2016;6(3):503–517. https://doi.org/10.3233/JPD-160867
- 141. Pagan FL, Hebron ML, Wilmarth B, et al. Pharmacokinetics and pharmacodynamics of a single dose nilotinib in individuals with Parkinson's disease. Pharmacol Res Perspect 2019;7(2):e00470. https://doi.org/10.1002/prp2.470
- 142. Shin W, Lim KS, Kim MK, et al. A first-in-human study to investigate the safety, tolerability, pharmacokinetics, and pharmacodynamics of KM-819 (FAS-associated factor 1 inhibitor), a drug for Parkinson's disease, in healthy volunteers. Drug Des Devel Ther 2019;13:1011–1022. https://doi.org/10.2147/DDDT.S198753
- 143. Besong-Agbo D, Wolf E, Jessen F, et al. Naturally occurring α-synuclein autoantibody levels are lower in patients with Parkinson disease. Neurology 2013;80(2):169–175. https://doi.org/ 10.1212/WNL.0b013e31827b90d1
- 144. Shi M, Liu C, Cook TJ, et al. Plasma exosomal α-synuclein is likely CNS-derived and increased in Parkinson's disease. Acta Neuropathol 2014;128(5):639–650. https://doi.org/10.1007/s00401-014-1314-y
- 145. Ishii R, Tokuda T, Tatebe H, et al. Decrease in plasma levels of α -synuclein is evident in patients with Parkinson's disease after elimination of heterophilic antibody interference. PLoS One 2015; 10(4):e0123162. https://doi.org/10.1371/journal.pone.0123162
- 146. Ding J, Zhang J, Wang X, et al. Relationship between the plasma levels of neurodegenerative proteins and motor subtypes of Parkinson's disease. J Neural Transm (Vienna) 2017;124(3):353– 360. https://doi.org/10.1007/s00702-016-1650-2

- 147. Lin CH, Yang SY, Horng HE, et al. Plasma α-synuclein predicts cognitive decline in Parkinson's disease. J Neurol Neurosurg Psychiatry 2017;88(10):818–824. https://doi.org/10.1136/jnnp-2016-314857
- 148. Malec-Litwinowicz M, Plewka A, Plewka D, et al. The relation between plasma α-synuclein level and clinical symptoms or signs of

Parkinson's disease. Neurol Neurochir Pol 2018;52(2):243–251. https://doi.org/10.1016/j.pjnns.2017.11.009

149. Chang CW, Yang SY, Yang CC, Chang CW, Wu YR. Plasma and serum alpha-synuclein as a biomarker of diagnosis in patients with Parkinson's disease. Front Neurol 2020;10:1388. https://doi.org/ 10.3389/fneur.2019.01388