



## CD28 Superagonist Shock and Blockage of Motogenic T Cell Cascade

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## INTRODUCTION

T cell motility is arrested by T cell receptor (TCR) recognition of cognate peptide-MHC (pMHC) complexes on antigen-presenting cells (1). Co-stimulation through CD28 is required for immune responses but its influence on T cell motility is less clear than that of the TCR. Ligation of CD28 and TCR collaborate to block a distinct protease-controlled step in the motogenic T cell cascade directed by the large transmembrane receptor low density lipoprotein receptor-related protein 1 (LRP1) and its high molecular weight ligand thrombospondin-1 (TSP-1) (2–9). Here I describe how blockage of this protease step is a likely explanation for the instantaneous arrest and cytokine storm elicited by the CD28 superagonist TGN1412 in a clinical trial in 2006 (10), which illustrates the power of the LRP1-targeted protease-dependent T cell control.

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Sundqvist K-G (2021) CD28 Superagonist Shock and Blockage of Motogenic T Cell Cascade. Front. Immunol. 12:670864. doi: 10.3389/fimmu.2021.670864 LRP1- AND TSP-1-DIRECTED MOTOGENIC CASCADE

LRP-1 and its coreceptor calreticulin trigger T cell motility and integrin-dependent adhesive contacts through a crosslinking cascade *via* the NH-terminal region of TSP-1 and interaction of the COOH-terminal region with CD47 (2–9). LRP1 and TSP-1 have a high turnover, their expression depends on sensing of components of the extracellular matrix (ECM) and other cells and they promote motility in response to ligation of integrin and chemokine receptors. Shedding of LRP1, which is attributed to a disintegrin and metalloprotease 10 (ADAM10), prevents accumulation of LRP1 and TSP-1 on the cells, and hence integrin-dependent adhesion and T cell activation, while maintaining polar cell shape and motility (9). Ligation of integrin and chemokine receptors antagonizes shedding through stimulation of transport of TSP-1 to the cell surface, whereupon TSP-1 targets shedding (5, 9). However, unlike a broad-spectrum MMP inhibitor, ligation of integrin or chemokine receptors by their natural ligands merely reduces but does not abrogate shedding.

## LIGATION OF TCR/CD3 AND CD28 TARGETS LRP1 AND TSP-1

The LRP1- and TSP-1-directed sensing and motility mechanism probably plays a crucial role for T cell search and surveillance to bring TCR and CD28 in contact with cognate pMHC complexes and B7. It is therefore interesting that these interactions target LRP1 and TSP-1 (2–9). Ligation of CD28 antagonizes shedding of LRP1 directly, whereas ligation of TCR/CD3 in collaboration with integrin

ligands increases TSP-1 on cells and TSP-1 inhibits shedding of LRP1 (4, 5, 9). This indicates that ligation of the TCR/CD3 complex and CD28 collaborate to prevent shedding of LRP1. Ligation of TCR/CD3 in a memory T cell clone enhanced cell surface expression of LRP1 suggesting a direct effect on shedding (4) but an impact on CD28 of the antigen-presenting cells used to maintain this clone cannot be excluded.

T cell activation by antigen upregulates TSP-1 synthesis and down-regulates LRP1 synthesis (8). The expression of LRP1 and TSP-1 in activated cells requires CD28 co-stimulation, since blocking co-stimulation by soluble CTLA-4 virtually abolishes expression of TSP-1 and LRP1 (8). The decreased LRP1 synthesis probably contributes to reduce motility, whereas the antigeninduced potentiation of TSP-1 expression, in collaboration with LRP1, supports integrin-dependent adhesive contacts with antigen-presenting cells.

Besides the initial effects of antigen stimulation on LRP1 and TSP1, antigen-induced IL-2 may stimulate TSP-1 synthesis contributing to subsequent effects on motility and perhaps also on other cells through secreted TSP-1 (6).

#### CD28 SUPERAGONISTS AND BLOCKAGE OF COUNTER-ADHESIVE STEP OF THE MOTOGENIC T CELL CASCADE

A single dose of the superagonistic CD28 antibody TGN1412 induced cytokine release syndrome in healthy human volunteers (10). This reflects capacity to crosslink CD28, binding to Fc-receptors, and activation of effector memory cells (11–14). Preculture at high cell density also increases the sensitivity of T cells to activation by antigen and TGN1412. This is thought to reflect tonic signaling (12), which is further discussed below.

Rodent experiments show that the primary effect of superagonistic CD28 antibodies is instantaneous arrest and trapping of T cells in lymphoid tissues associated with irregular cell shape, increased cell size and activation (15). This is accompanied by increased capacity to bind to integrin ligands and most likely reflects adhesion and cytoplasmic spreading, and probably caused the lymphopenia developed in the clinical trial (10). The TGN1412 trial and accompanying studies hence show that ligation of CD28 targets T cell motility and adhesion, like ligation of the TCR, which induces polarizing interactions with antigen-presenting cells (1, 16) contrasting with the apolar adhesion by CD28 superagonists (15). These similar effects probably require a mechanism targeted *via* both TCR and CD28, a criterion fulfilled by the motogenic cascade directed by LRP1 and TSP-1) (2–9).

A clue to the mechanism behind the disastrous TGN1412 trial is that ligation of CD28 antagonizes shedding of LRP1 (**Figure 1**) (2, 9). Ligation of CD28 using a conventional CD28 antibody has a moderate inhibitory effect on shedding of LRP1, whereas broad-spectrum MMP inhibition abrogates shedding. This induces adhesion and apolar spreading of T cells contacting integrin ligands, and a characteristic irregular periphery with pseudopodia (2, 9), which mimics the appearance of arrested cells in rodents treated with superagonistic CD28 antibodies (15). Like TGN1412, broad-spectrum MMP inhibition promotes excessive T cell production of cytokines and, in addition, of chemokines and growth factors (19). This reinforces the conclusion that the CD28-targeted protease of the motogenic LRP1-TSP-1 cascade controls T cell adhesion and activation.

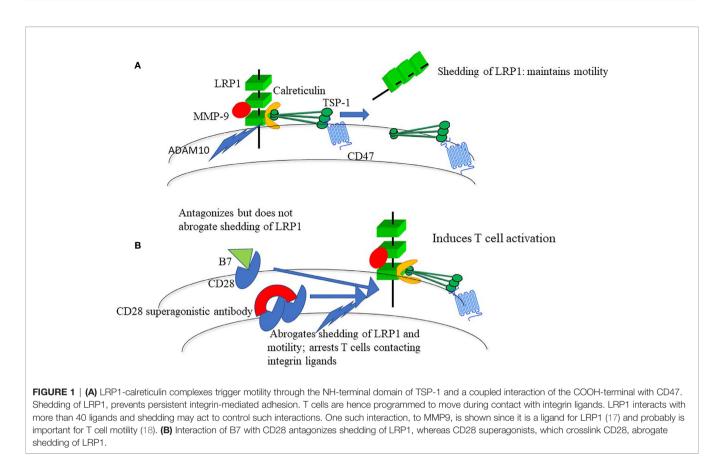
# THE MOTOGENIC LRP1-TSP-1 CASCADE AND TONIC SIGNALING

Constitutive low level signaling, often called tonic, is thought to drive T cell motility and responsiveness to antigen and TGN1412 through TCR self MHC interaction (12, 20–22). The conditions promoting tonic signaling and responsiveness to antigen and TGN1412 have several similarities to the motogenic cascade suggesting a shared mechanistic background. Maintenance of motility through the motogenic cascade hence also depends on contact with other cells but also on contact with ECM (2, 23–25). Additional evidence for a shared background of the tonic signaling responsible for the action of TGN1412 and the motogenic cascade is that absence of cell contact causes loss of sensitivity to TGN1412 and antigen in blood T cells (12), which resembles that T cell motility is low in blood and high in lymphoid tissue (26).

Motility dependent on sensing of ECM components and other cells (23–25) by the motogenic LRP1-TSP-1-directed cascade (2) implies a high degree of adaptivity to different environments in comparison with motility driven by self-recognition. In tissues, T cells frequently encounter ECM components, such as collagen and fibronectin, besides other cells, and it is therefore logical that contact both with cellular and non-cellular components supports T cell motility. In support of a basal motility mechanism dependent on environmental sensing, cell contact supports protein synthesis (27) and protein synthesis is required for motility of amebae and lymphocytes (24, 28–30).

# OKT3-INDUCED CYTOKINE RELEASE SYNDROME

OKT3 induces self-limited cytokine release and spontaneously reversible clinical symptoms (31), similar to but less pronounced than the TGN1412-induced life-threatening stereotypic symptoms in all individuals (10). These TCR/CD3-mediated effects, together with the fact that OKT3 induces T cell arrest (32), therefore resemble the effects of broad spectrum MMP inhibition on motility and cytokine release. This indicates that OKT3 targets the protease step of the LRP1- and TSP-1-directed motogenic cascade. OKT3 can activate T cells *in vitro* without preculture at high density, unlike TGN1412, which probably reflects independence of accessory cells and tonic signaling (12). This probably means that the cells do not have to be motile to bind OKT3, in contrast to TGN1412, which is presented bound to environmental Fc receptors.



## LRP1 AND TSP-1 CONTROL OF CELL SIGNALING

LRP1 and TSP-1 may regulate T cell functions, such as adhesion and activation, by connecting cell surface receptors and signaling pathways. LRP1 directs T cell motility through JAK tyrosine kinase and PI3K (3) and evidence from non-lymphoid cells indicate that LRP1 is a coreceptor for other receptors and an integrator of signaling pathways through its more than 40 binding sites for other molecules (33–36). One example is the influence of LRP1 on glucose metabolism (37), which is crucial for T cell activation and requires CD28 (38). Additional examples of integration of signaling pathways is that LRP1 is a receptor for TGF- $\beta$ , acts in concert with other TGF- $\beta$  receptor types (39), and together with TSP-1 (40) control TGF- $\beta$  action, which may be important since TGF- $\beta$  has a crucial role for T cell function with far-reaching implications for Th17 and Treg cells (41).

## CONCLUSIONS AND PERSPECTIVES

Encounter of the TCR and CD28 with cognate pMHC complexes and B7 on antigen-presenting cells hence seems to block a protease of a molecular cascade regulating T cell motility and integrin-dependent contacts. The powerful effects of broadspectrum MMP inhibition on T cell motility and activation and the similarities of these effects with the TGN1412- induced cytokine storm indicate that this protease control is important. This control seems to maintain cell surface expression of LRP1 and associated ligands at a low level, enough for triggering motogenic signals but insufficient for integrin-dependent adhesive contacts and activation. The low level of stable LRP1 expression on the cells due to shedding probably prevents LRP1, by virtue of its multiple binding sites for other molecules (33–36), to connect with them and signaling pathways critical for activation.

Unlike broad-spectrum MMP inhibition and superagonistic CD28 antibodies, which both may effectively inhibit the protease controlling cell surface expression of LRP1 to abrogate shedding, pMHC complexes and B7 seem to collaborate to inhibit this protease. The putative existence of this control with a driving effect on motility is evident in T cells with upregulated integrins in an environment presenting the corresponding integrin ligands (42). In addition, this control may be of vital importance to prevent triggering of adverse immune responses, which indeed is demonstrated by the cytokine release syndrome induced by superagonistic CD28 antibodies. So, the counteradhesive motogenic protease mechanism may function as an important checkpoint, decisive for the behavior and fate of T cells, coupled to the encounter of the TCR and CD28 with pMHC complexes and B7. This protease mechanism may further prevent the large LRP1 molecule from interference with TCR-pMHC interactions and generate force on TCR-pMHC bonds in T cell antigen recognition, which is a matter of debate (43).

The dependence of the motogenic molecular cascade on sensing of the environment implies a constitutive property of

crucial importance for the search for antigen and immune surveillance. The protease step endows this cascade with a fundamental control of triggering of immune responses. A basic regulation of T cell motility and adhesive contacts distinct from but responsive to the TCR and CD28 is logical from an evolutionary perspective. Adaptive immunity evolved in jawed vertebrates is hence likely to be intimately dependent on phylogenetically old proteins, including LRP1, TSP-1, and integrins, and functions dependent on them, such as motility and adhesion, evolved at earlier stages of evolution.

#### REFERENCES

- Henrickson SE, Mempel TR, Mazo IB, Artyomov MN, Zheng H, Peixoto A, et al. T Cell Sensing of Antigen Dose Governs Interactive Behavior With Dendritic Cells and Sets a Threshold for T Cell Activation. *Nat Immunol* (2008) 9:282–91. doi: 10.1038/ni1559
- Sundqvist KG. T Cell Motility-How is it Regulated? Front Immunol (2020) 11. doi: 10.3389/fimmu.2020.588642
- Li SS, Forslöw A, Sundqvist KG. Autocrine Regulation of T Cell Motility by Calreticulin-Thrombospondin-1 Interaction. J Immunol (2005) 174:654–61. doi: 10.4049/jimmunol.174.2.654
- Li SS, Liu Z, Uzunel M, Sundqvist KG. Endogenous Thrombospondin-1 is a Cell-Surface Ligand for Regulation of Integrin-Dependent T-lymphocyte Adhesion. *Blood* (2006) 108:3112–20. doi: 10.1182/blood-2006-04-016832
- Liu Z, Christensson M, Forslöw A, De Meester I, Sundqvist KG. A CD26controlled Cell Surface Cascade for Regulation of T Cell Motility and Chemokine Signals. J Immunol (2009) 183:3616–24. doi: 10.4049/ jimmunol.0804336
- Bergström SE, Bergdahl E, Sundqvist KG. A Cytokine-Controlled Mechanism for Integrated Regulation of T-lymphocyte Motility, Adhesion and Activation. *Immunol* (2013) 140:441–55. doi: 10.1111/imm.12154
- Talme T, Bergdahl E, Sundqvist KG. Regulation of T-lymphocyte Motility, Adhesion and De-Adhesion by a Cell Surface Mechanism Directed by Low Density Lipoprotein Receptor-Related Protein 1 and Endogenous Thrombospondin-1. *Immunol* (2014) 142:176–92. doi: 10.1111/imm.12229
- Bergström SE, Uzunel M, Talme T, Bergdahl E, Sundqvist KG. Antigen-Induced Regulation of T-cell Motility, Interaction With Antigen-Presenting Cells and Activation Through Endogenous Thrombospondin-1 and its Receptors. *Immunol* (2015) 144:687–703. doi: 10.1111/imm.12424
- Panezai J, Bergdahl E, Sundqvist KG. T-Cell Regulation Through a Basic Suppressive Mechanism Targeting Low-Density Lipoprotein Receptor-Related Protein 1. *Immunol* (2017) 152:308–27. doi: 10.1111/imm.12770
- Suntharalingam G, Perry MR, Ward S, Brett SJ, Castello-Cortes A, Brunner MD, et al. Cytokine Storm in a Phase1 Trial of the Anti–CD28 Monoclonal Antibody TGN1412. *NEngl JMed* (2006) 355:1018–28. doi: 10.1056/ NEJMoa063842
- Hunig T, Dennehy K. CD28 Superagonists: Mode of Action and Therapeutic Potential. *Immunol Lett* (2005) 100(1):21–8. doi: 10.1016/j.imlet.2005.06.012
- Römer PS, Berr S, Avota E, Na S-J, Battaglia M, ten Berge I, et al. Preculture At High Cell Density Increases Sensitivity of T-cell Responses, Revealing Cytokine Release by CD28 Superagonist TGN1412. *Blood* (2011) 118:6772– 82. doi: 10.1182/blood-2010-12-319780
- Bartholomaeus P, Semmler LY, Bukur T, Boisguerin V, Römer PS, Tabares P, et al. Cell Contact-Dependent Priming and Fc Interaction With CD32+ Immune Cells Contribute to the TGN1412-triggered Cytokine Response. *J Immunol* (2014) 192:2091–8. doi: 10.4049/jimmunol.1302461
- Eastwood D, Findlay L, Poole S, Bird C, Wadhwa M, Moore M, et al. Monoclonal Antibody TGN1412 Trial Failure Explained by Species Differences in CD28 Expression on CD4+ Effector Memory T–Cells. Br J Pharmacol (2010) 161:512–26. doi: 10.1111/j.1476-5381.2010.00922.x
- Muller N, Van den Brandt J, Odoardi F, Tischner D, Herath J, Flugel A, et al. A CD28 Superagonistic Antibody Elicits 2 Functionally Distinct Waves of T Cell Activation in Rats. J Clin Invest (2008) 118:1405–16. doi: 10.1172/JCI32698

#### **AUTHOR CONTRIBUTIONS**

The author confirms being the sole contributor of this work and has approved it for publication.

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- Ueda H, Morphew MK, McIntosh JR, Davis MM. CD4+T-Cell Synapses Involve Multiple Distinct Stages. *Proc Natl Acad Sci (USA)* (2011) 108:17099– 104. doi: 10.1073/pnas.1113703108
- Hahn-Dantona E, Ruiz JF, Bornstein P, Strickland DK. The Low Density Lipoprotein Receptor-Related Protein Modulates Levels of Matrix Metalloproteinase 9 (MMP-9) by Mediating its Cellular Catabolism. J Biol Chem (2001) 276:15498–503. doi: 10.1074/jbc.M100121200
- Ivanoff A, Ivanoff J, Hultenby K, Sundqvist KG. Infiltrative Capacity of T Leukemia Cell Lines: A Distinct Functional Property Coupled to Expression of Matrix Metalloproteinase-9 (MMP-9) and Tissue Inhibitor of Metalloproteinases-1 (TIMP-1). *Clin Exp Metast* (1999) 17:695–711. doi: 10.1023/A:1006749304315
- Eaton VL, Lerret NM, Velasquez-Lopera MM, John R, Caicedo RM, DeCresce RP, et al. Enhanced Allograft Survival and Modulation of T-cell Alloreactivity Induced by Inhibition of MMP/ADAM Enzymatic Activity. *Am J Transplant* (2008) 8:507–16. doi: 10.1111/j.1600-6143.2007.02097.x
- Kim H-E, Cho J-H. T Cell's Sense of Self: A Role of Self-Recognition in Shaping Functional Competence of Naïve T Cells. *Imm Netw* (2017) 17:201– 13. doi: 10.4110/in.2017.17.4.201
- Fischer UB, Jacovetty EL, Mendelros RB, Goudy BD, Zell T, Swanson J-B, et al. MHC Class II Deprivation Impairs CD4 T Cell Motility and Responsiveness to Antigen-Bearing Dendritic Cells in Vivo. *Proc Natl Acad Sci* (2007) 104:7181–6. doi: 10.1073/pnas.0608299104
- Stefanova I, Dorfmann JR, Germain RN. Self-Recognition Promotes the Foreign Antigen Sensitivity of Naïve T Lymphocytes. *Nature* (2002) 420:429–34. doi: 10.1038/nature01146
- Sundqvist K-G, Otteskog P. Anchorage and Lymphocyte Function: Collagen and the Maintenance of Motile Shape in T Cells. *Immunology* (1986) 58:365–9.
- Arencibia I, Pedari L, Sundqvist KG. Induction of Motility and Alteration of Surface Membrane Polypeptides in Lymphocytes by Contact With Autologous and Allogeneic Fibroblasts. *Exp Cell Res* (1987) 172:124–33. doi: 10.1016/0014-4827(87)90099-1
- Sundqvist KG, Otteskog P. A Labile Antigen Complex At the Lymphocyte Surface: The Effect of the Substratum on its Expression. J Immunol (1987) 1383:825–32.
- Sundqvist KG, Otteskog P, Mellstedt H. Compartment Dependence of Tlymphocyte Motility. Scand J Immunol (1986) 23:647–54. doi: 10.1111/j.1365-3083.1986.tb02000.x
- Ben-Ze'ev A, Farmer SR, Penman S. Protein Synthesis Requires Cell-Surface Contact While Nuclear Events Respond to Cell Shape in Anchorage-Dependent Fibroblasts. *Cell* (1980) 21:365–72. doi: 10.1016/0092-8674(80)90473-0
- Wilkinson PC. The Locomotor Capacity of Human Lymphocytes and its Enhancement by Cell Growth. *Immunol* (1986) 57:281–9.
- Wiebke JL, Quinlan WM, Graham L, Doerschuk CM. Effect of Protein Synthesis Inhibition by Cycloheximide on Lymphocyte Circulation Lab Invest. *Laboratory Investigation* (1994) 70:392–8.
- Clotworthy M, Traynor D. On the Effects of Cycloheximide on Cell Motility and Polarisation in Dictyostelium Discoideum. BMC Cell Biol (2006) 7:5. doi: 10.1186/1471-2121-7-5
- Chatenoud L, Ferran C, Legendre C, Thouard I, Merite S, Reuter A, et al. In Vivo Cell Activation Following OKT3 Administration. Systemic Cytokine Release and Modulation by Corticosteroids. *Transplantation* (1990) 49:697– 702. doi: 10.1097/00007890-199004000-00009

- Wiemer AJ, Subramanya Hegde S, Jenny E, Gumperz JE, Huttenlocher A. A Live Imaging Cell Motility Screen Identifies Prostaglandin E2 as a T Cell Stop Signal Antagonist. J Immunol (2011) 187:3663–70. doi: 10.4049/ jimmunol.1100103
- Lillis AP, Van Duyn LB, Murphy-Ullrich JE, Strickland DK. LDL Receptor-Related Protein 1: Unique Tissue-Specific Functions Revealed by Selective Gene Knockout Studies. *Physiol Rev* (2008) 88:887–918. doi: 10.1152/ physrev.00033.2007
- Boucher P, Herz J. Signaling Through LRP1: Protection From Atherosclerosis and Beyond. *Biochem Pharmacol* (2011) 1(81):1–5. doi: 10.1016/ j.bcp.2010.09.018
- El Asmar Z, Terrand J, Jenty M, Host L, Mlih M. Controlled by LRP1 (Receptor-Related Protein 1) Cytoplasmic and Extracellular Domains Limit Cellular Cholesterol Accumulation. J Biol Chem (2016) 291:5116–27. doi: 10.1074/jbc.M116.714485
- 36. Boucher P, Li W-P, Matz RL, Takayama Y, Auwerx J, Anderson RGW, et al. LRP1 Functions as an Atheroprotective Integrator of TGFbeta and PDFG Signals in the Vascular Wall: Implications for Marfan Syndrome. *PloS One* (2007) 16:e448. doi: 10.1371/journal.pone.0000448
- Liu CC, Hu J, Tsai CW, Yue M, Melrose HJ, Kanekiyo T, et al. Neuronal LRP1 Regulates Glucose Metabolism and Insulin Signaling in the Brain. *J Neurosci* (2015) 35:5851–9. doi: 10.1523/JNEUROSCI.5180-14.2015
- Jacobs SR, Herman CE, Maciver NJ, Wofford JA, Wieman HL, Hammen JJ, et al. Glucose Uptake is Limiting in T Cell Activation and Requires Cd28-Mediated Akt-Dependent and Independent Pathways. J Immunol (2008) 180:4476–86. doi: 10.4049/jimmunol.180.7.4476

- 39. Huang SS, Ling T-Y, Tseng W-F, Huang Y-H, Tang F-M, Leal SM, et al. Cellular Growth Inhibition by IGFBP-3 and TGF- $\beta_1$ requires LRP-1. FASEB (2003) 17:2068–89. doi: 10.1096/fj.03-0256com
- Murphy-Ullrich JE, Suto MJ. Thrombospondin-1 Regulation of Latent TGF-β Activation: A Therapeutic Target for Fibrotic Disease. *Matrix Biol* (2018) 68-69:28–43.
- Oh SA, Li MO. Tgf-β: Guardian of T Cell Function. J Immunol (2013) 191:3973–9. doi: 10.4049/jimmunol.1301843
- Sundqvist KG, Hauzenberger D, Hultenby K, Bergström SE. T Lymphocyte Infiltration of Two- and Three-Dimensional Collagen Substrata by an Adhesive Mechanism. *Exp Cell Res* (1993) 206:100–10. doi: 10.1006/excr.1993.1125
- Limozin L, Bridge M, Bongrand P, Dushek O, van der Merwe MA, Robert P. Tcr-pMHC Kinetics Under Force in a Cell-Free System Show No Intrinsic Catch Bond, But a Minimal Encounter Duration Before Binding. PNAS (2019) 116:16943–8. doi: 10.1073/pnas.1902141116

**Conflict of Interest:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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