MAJOR HISTOCOMPATIBILITY COMPLEX-RESTRICTED ANTIGEN RECEPTOR ON T CELLS VIII. Role of the LFA-1 Molecule

BY WILLIAM T. GOLDE, JOHN W. KAPPLER, JULIA GREENSTEIN,* BERNARD MALISSEN,[‡] LEROY HOOD,[‡] and PHILIPPA MARRACK

From the Department of Medicine, National Jewish Hospital and Research Centers, and the Departments of Microbiology and Immunology; Biophysics, Biochemistry and Genetics; and Medicine, University of Colorado Health Sciences Center, Denver, Colorado 80206; the *Department of Pediatric Oncology and Pathology, Dana Farber Cancer Center, Boston, Massachusetts 02115; and the [‡]Division of Biology, California Institute of Technology,

Pasadena, California 91125

A subset of T cell surface antigens have been shown to play a role in the response of T cells to target cells. These molecules do not seem to be part of the antigen (Ag)-specific, major histocompatibility complex (MHC)-restricted receptor on T cells since they do not vary between T cell clones, yet antibodies against these molecules very frequently inhibit responses of T cells to Ag/MHC (1-4). For example, the lymphocyte function-associated antigen-1 (LFA-1) seems to be active in a broad range of T cell functions with no relation to the specificity of the response (5, 6).

In murine systems, all reports so far of the effects of monoclonal antibody directed against LFA-1 on T cell function have analyzed the interaction of T cells with cells of leukocytic lineage. In this paper we present data which show that anti-LFA-1 does not inhibit the responses of T cell hybridomas to Ag presented by nonleukocytic fibroblast cells derived from the L cell line, nor to polyvalent anti-T cell receptor antibodies, although the responses of these same T cell hybridomas to Ag presented by leukocytic cells are sensitive to the antibody. These results imply that LFA-1 does not play an essential role in T cell activation, and that the molecule may interact with ligands on leukocytic target cells which are absent from L cells.

Materials and Methods

Culture Medium. Cells were cultured in medium prepared by the recipe of Mishell and Dutton (7), with the exception that the medium was supplemented before use with 10% nutrient cocktail and contained 10% fetal bovine serum, 5×10^{-5} M 2-mercaptoethanol, and 50 µg/ml gentamicin (Shering Corp., Kenilworth, NJ). Transfected fibroblast lines were cultured in the same medium with hypoxanthine, aminopterin, and thymidine (HAT) added to maintain selection for cells expressing the transfected genes.

This work was supported by U. S. Public Health Service grants A1-17134 and A1-20317 and American Cancer Society grant IM-49. It was done during the tenure by J. W. Kappler of faculty research award 218 from the American Cancer Society. J. Greenstein was supported by U. S. Public Health Service postdoctoral fellowship CA 07324.

J. EXP. MED. © The Rockefeller University Press · 0022-1007/85/03/0635/06 \$1.00 635 Volume 161 March 1985 635-640

TABLE I							
Monoclonal Antibodies	Used	in	These	Studies			

Antibody	Specificity	Source	Reference	
121/7.7	LFA-1	Culture SN	8	
GK-1.5	L3T4	Culture SN	3	
KI12-98.15	Self-D ^d receptor on T cell hybrid 3DT-52.5	Culture SN	9	
КЈ1-26.1	Receptor for OVA/I-A ^d on T cell hybridoma, DO- 11.10	Culture SN	9	

Reagents. Keyhole limpet hemocyanin (KLH) and ovalbumin (OVA), obtained from Sigma Chemical Co., St. Louis, MO, were used as Ag in these experiments. Alkaline phosphatase-conjugated rabbit anti-rat immunoglobulin and p-nitrophenyl phosphate were also purchased from Sigma Chemical Co.

Monoclonal Antibodies. Monoclonal antibodies were prepared as cell-free culture supernatants (SN) of hybridoma c ures grown to a cell concentration of 10⁶ cells/ml. A list of the monoclonal antibodies used in these experiments is given in Table I. Monoclonal antibody culture supernatants were used neat.

T Cell Hybridomas. T cell hybridomas for these experiments were prepared by fusing Ag/MHC-reactive T cell blasts to BW5147 by standard procedures (10). The properties of these T cell hybridomas have been described elsewhere (10).

Transfected Fibroblast Lines. Two different transfected fibroblast lines were used in these studies. Both are products of transfections of L cells, C3H-derived fibroblast tumor cells, made thymidine kinase negative (TK⁻) to render them HAT sensitive. Transfections were done with plasmid constructs containing the genes of interest plus the thymidine kinase gene. The first cell line, CA14.11.14, was transfected with the genes for I-A^k (11). The second cell line, a generous gift of Dr. John Seidman (Harvard University, Cambridge, MA) was transfected with the genes for D^d (12).

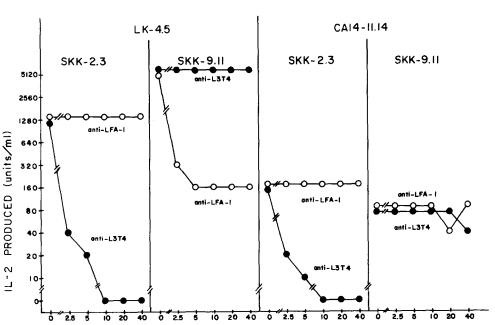
Interleukin 2 (IL-2) Assay for T Cell Hybridoma Activity. T cell hybridomas' recognition of Ag/MHC or stimulation by polyvalent anti-receptor antibody was assayed by the secretion of IL-2 by these hybridomas (10). None of these hybridomas made detectable IL-2 in the absence of the appropriate Ag/MHC or other stimulus. To induce IL-2 production, 10⁴ hybridoma T cells were incubated for 24 h with the appropriate Ag and 10^5 antigen-presenting cells (APC), or with anti-receptor antibodies coupled to Sepharose beads in 250-µl cultures. Ag concentrations were always 500-1,000 µg/ml.

Results are expressed as units per milliliter of IL-2 secreted by the T cell hybridomas under different conditions. 10 U/ml IL-2 was the minimum detectable using this assay. A20.2J, a BALB/c B cell lymphoma, and P815, a DBA/2 mastocytoma, were used to stimulate a D^d-specific T cell hybridoma; LK-4.5, a B cell hybridoma, was used to present Ag to I-A^k-restricted T cell hybridomas.

ELISA for Cell Surface Antigens. We assayed for the expression of cell surface antigens by the standard enzyme-linked immunosorbent assay (ELISA) (13). This assay was slightly modified to use a vacuum plate system from Millipore Corp. (Bedford, MA).

Results and Discussion

It has been reported previously that the L cell transfectant CA14-11.14 expresses the transfected gene product I-A^k in sufficient concentration to serve as a presenting cell to I-A^k-restricted, Ag-specific T cell hybridomas. This is illustrated in Fig. 1, which shows that two I-A^k-restricted T cell hybridomas, SKK-2.3 and SKK-9.11, respond to KLH plus the transfected L cell, although somewhat less well than to KLH plus an equal number of cells of the B cell hybridoma, LK-4.5. The blocking effects of anti-L3T4 were equivalent when using either presenting line together with the anti-L3T4-sensitive T cell hybridoma, SKK-2.3. A different result was observed using anti-LFA-1 as the blocking antibody. The anti-LFA-1-sensitive hybrid SKK-9.11 was effectively blocked



UI MONOCLONAL ANTIBODY SN/CULTURE

FIGURE 1. The response of a T cell hybridoma to KLH/I-A^k is unaffected by the blocking effects of anti-LFA-1 antibody when an I-A-transfected fibroblast is used as the presenting cell. The T cell hybridomas SKK-2.3 and SKK-9.11 were incubated with KLH plus the B cell hybridoma LK-4.5 or the I-A^k-transfected fibroblast line, CA14-11.14. Different concentrations of anti-L3T4 (\bigcirc) or anti-LFA-1 (\bigcirc) antibodies were added to these cultures. The effects of these antibodies on the responses of the T cell hybridomas were measured 24 h later by monitoring the concentrations of IL-2 secreted.

when LK-4.5 was the presenting cell line, but such blocking effects were lost when the CA14-11.14-transfected fibroblast was the presenting cell (Fig. 1).

The effects of anti-L3T4 on the responses of the T cell hybridomas, regardless of the presenting cell used, is consistent with the hypothesis that the target of L3T4 interaction is a nonpolymorphic determinant on the Ia molecule (14). The fact that anti-LFA-1 had no effect on the response of SKK-9.11 when L cells were used as presenting cells suggests that LFA-1 does not play a role in this interaction, discussed more fully below.

A similar result was seen when the D^d-specific T hybridoma, 3DT-52.5, was used (Fig. 2). This hybridoma responded comparably to the BALB/c B cell lymphoma A20.2J, the DBA/2 mastocytoma P815, and the D^d-transfected L cell line T4.8.3. KJ12-98.5, an antibody specific for the clonotypic receptor on 3DT-52.5, blocked the response of the T cell hybridoma to all of these cell lines comparably. However, the blocking effects of anti-LFA-1 were seen only when the two leukocytic cell lines, A20.2J and P815, were used as targets. No such blocking was observed in the response of 3DT-52.5 to T4.8.3.

The blocking effects of anti-LFA-1 do not seem to depend on expression of this glycoprotein by the APC themselves since, as shown in Table II, neither A20-2J nor the L cells and their transfectants, bore LFA-1. Thus anti-LFA-1 antibodies were able to inhibit Ag presentation by A20-2J to some T cell

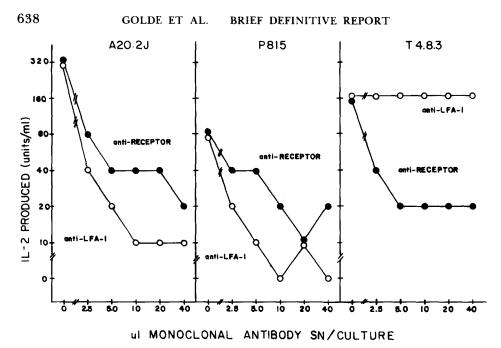


FIGURE 2. The response of a class I-specific T cell hybridoma is insensitive to the blocking effects of anti-LFA-1 monoclonal antibody when a transfected fibroblast is used as the stimulator. The T cell hybridoma, 3DT-52.5, was incubated with the D^d-presenting cell lines, A20-2J, P815, or T4.8.3 in the presence of various concentrations of anti-3DT-52.5 receptor antibody (\bullet) or anti-LFA-1 (O). The effects of these antibodies on responses were measured 24 h later by monitoring concentrations of 1L-2 secreted.

Cell line	ELISA OD × 10 ⁸ * after assay with 121/7.7 anti-LFA-1			
3DT-52.5	661 ± 88			
SKK-2.3	705 ± 45			
SKK-9.11	392 ± 37			
A20.2J	28 ± 37			
P815	287 ± 42			
LK-4.5	-3 ± 11			
T4.8.3	-68 ± 29			
L(TK ⁻)	37 ± 67			

TABLE II Expression of Cell Surface Antigens by These Cell Lines as Measured by ELISA

* $OD_{405} \pm SE \times 10^{-3}$.

hybridomas, even though this cell line was LFA-1⁻. This result implies that LFA-1 inhibition acts on the LFA-1-bearing T cell hybridomas in these cultures, and argues against LFA-1 functioning in a like-like interaction.

It is difficult to account for the variations in the effects of anti-LFA-1 that depend on the APC in culture, given the conclusion that the primary targets of these antibodies are T cell hybridomas rather than APC. One explanation could be that the LFA-1 molecule is involved in the binding of some T cells to their targets, as an "accessory molecule" that interacts with an unknown ligand (not a

 TABLE III

 Anti-LFA-1 Does Not Inhibit the Response of T Cell Hybridomas to Polyvalent Anti-receptor

Antibodies

T. 11 hat 14 m		IL-2 secreted in the presence of anti-LFA-1 SN at:					
T cell hybridoma	KJ1-26-Sepharose	0*	2.5	5.0	10	20	40
		U/ml					
DO-11.10	OVA/A20-2]	320	40	20	10	10	<10
DO-11.10	K11-26-Sepharose beads	320	320	320	320	320	160
3DT-52.5	A20-2J	640	320	160	80	40	40
3DT-52.5	KJ1-26-Sepharose beads	160	160	160	160	160	160

* Microliters of anti-LFA-1 per 250-µl culture of SN from I21/7.7.

class I or class II molecule) on the target cells. If this explanation is correct, we would have to propose that LFA-1 is not involved in the interaction of T cells with L cells, perhaps because L cells do not bear the ligand for LFA-1.

If LFA-1 is simply acting as an accessory molecule in the binding of T cells and their targets, then the molecule should not play a role in the interactions of T cells with noncellular targets. To test this possibility, we challenged two T cell hybridomas, DO-11.10 and 3DT-52.5, either with stimulating B cell targets, bearing OVA/I-A^d or D^d, respectively, or with stimulatory anti-receptor antibodies coupled to Sepharose beads. As shown in Table III, anti-LFA-1 inhibited very well the responses of these T cells to Ag/MHC presented by the B cell lymphoma, A20-2J. The antibody had no effect on the responses of these T cells to polyvalent anti-receptor antibody, however.

From these experiments we concluded, as before, that LFA-1 did not play an essential role in T cell activation for IL-2 production, but rather was involved in the interaction of T cells with certain target cells, which did not include L cells and their derivatives.

Our future experiments will test whether LFA-1 functions when normal mouse fibroblasts are used as APC for T cell hybridomas, instead of L cells. This is the case in man, since Collins et al. (15) have shown that human dermal fibroblasts, induced to express class II molecules by gamma interferon (IFN- γ), can be killed by class II-specific cytotoxic T cells, and that this killing is blocked by anti-LFA-1 antibodies. With this in mind, we have been unable to show that Ag presentation by L cell lines becomes sensitive to anti-LFA-1 antibodies if the cells are preincubated with IFN- γ (data not shown); it remains to be determined if this is a property of L cells only or if it also extends to normal mouse fibroblasts.

Summary

We show that the LFA-1 molecule on T cells does not play a role in the stimulation of T cell hybridomas by certain targets, namely antigen presented by L cell derivatives or polyvalent anti-receptor antibody. These results suggest that LFA-1 may act by binding to ligands that are not present on all cells. We hope this result will help us and others to establish the true role of LFA-1 in T cell responses.

We would like to thank Dr. Seidman for the kind gift of the D^d-transfected L cells, and Drs. Dialynas, Fitch, and Trowbridge for the gift of monoclonal antibodies. We also thank Ella Kushnir, James Leibson, and Janice White for their excellent technical assistance and Edna Squillante and Donna J. Thompson for their superb secretarial help.

Received for publication 10 December 1984.

References

- Meuer, S. C., S. F. Schlossman, and E. L. Reinherz. 1982. Clonal analysis of human cytotoxic T lymphocytes: T4⁺ and T8⁺ effector T cells recognize products of different major histocompatibility complex regions. *Proc. Natl. Acad. Sci. USA*. 79:4395.
- MacDonald, H. R., A. L. Glasebrook, C. Bron, A. Kelso, and J.-C. Cerottini. 1982. Clonal heterogeneity in the functional requirement for Lyt-2,3 molecules on cytotoxic T lymphocytes (CTL): possible implications for the affinity of CTL receptors. *Immunol. Rev.* 68:69.
- Dialynas, D. P., Z. S. Quan, K. A. Wall, A. Pierres, J. Quintans, M. R. Loken, M. Pierres, and F. Fitch. 1983. Characterization of the murine T cell surface molecule, designated L3T4, identified by monoclonal antibody GK-1.5: similarity of L3T4 to the human Leu 3/T4 molecule and the possible involvement of L3T4 in class II MHC antigen reactivity. J. Immunol. 131:2445.
- Meuer, S. C., K. A. Fitzgerald, R. E. Hussey, J. C. Hodgdon, S. F. Schlossman, and E. L. Reinherz. 1983. Clonotypic structures involved in antigen-specific human T cell function. Relationship to the T3 molecular complex. J. Exp. Med. 157:705.
- Davingnon, D., E. Martz, T. Reynolds, K. Kurzinger, and T. A. Springer. 1981. Lymphocyte function-associated antigen (LFA-1): a surface antigen distinct from Lyt-2,3 that participates in T lymphocyte-mediated killing. *Immunology*. 78:4535.
- 6. Pierres, M., C. Goridis, and P. Goldstein. 1982. Inhibition of murine T cell-mediated cytolysis by a rat monoclonal Ab immunoprecipitating two lymphoid cell surface polypeptides of 94,000 and 180,000 molecular weight. *Eur. J. Immunol.* 13:424.
- 7. Mishell, R., and R. W. Dutton. 1967. Immunization of dissociated spleen cell cultures from normal mice. J. Exp. Med. 126:423.
- 8. Trowbridge, I. S., and M. B. Ovary. 1981. Molecular complexity of leukocyte surface glycoproteins related to the macrophage differentiation antigen MAC-1. J. Exp. Med. 154:1517.
- 9. Kappler, J. W., R. Kubo, K. Haskins, J. White, and P. Marrack. 1983. The mouse Tcell receptor: comparison of MHC-restricted receptors on two T cell hybridomas. *Cell*. 34:727.
- Kappler, J. W., B. Skidmore, J. White, and P. C. Marrack. 1981. Antigen-inducible, H-2-restricted, interleukin 2-producing T cell hybridomas. Lack of independent antigen and H-2 recognition. J. Exp. Med. 153:1198.
- 11. Malissen, B., M. Peele-Price, J. M. Goverman, M. McMillan, J. White, J. Kappler, P. Marrack, A. Pierres, M. Pierres, and L. Hood. 1984. Gene transfer of H-2 class II genes: antigen presentation by mouse fibroblast and hamster B cell lines. *Cell.* 36:39.
- Margulies, D. H., G. A. Evans, K. Ozato, R. D. Camerini, K. Tamaka, E. Appella, and J. G. Seidman. 1983. Expression of H-2D^d and H-2L^d mouse major histocompatibility antigen genes in L cells after DNA-mediated gene transfer. J. Immunol. 130:463.
- 13. Marrack, P., R. Endres, R. Shimonkevitz, A. Zlotnik, D. Dialynas, F. Fitch, and J. Kappler. 1983. The major histocompatibility complex-restricted antigen receptor on T cells. II. Role of the L3T4 Product. J. Exp. Med. 158:1077.
- 14. Greenstein, J. L., P. Marrack, J. Kappler, and S. J. Burakoff. 1984. The role of L3T4 in recognition of Ia by acytotoxic H-2^d-specific T cell hybridoma. *J. Exp. Med.* 159:1213.
- 15. Collins, T., A. M. Krensky, C. Clayberger, W. Fiers, M. A. Gimbrone, S. J. Burakoff, and J. S. Pober. 1984. Human cytolytic T lymphocyte interactions with vascular endothelium and fibroblasts: role of effector and target cell molecules. *J. Immunol.* 133:1878.