

## Review

# Interaction between lipid metabolism and macrophage polarization in atherosclerosis

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## SUMMARY

Atherosclerosis (AS) is a chronic inflammatory condition associated with lipid deposition. The interaction between abnormal lipid metabolism and the inflammatory response has been identified as the underlying cause of AS. Lipid metabolism disorders are considered the basis of atherosclerotic lesion formation and macrophages are involved in the entire process of AS formation. Macrophages have a high degree of plasticity, and the change of their polarization direction can determine the progress or regression of AS. The disturbances in bioactive lipid metabolism affect the polarization of different phenotypes of macrophages, thus, affecting lipid metabolism and the expression of key signal factors. Therefore, understanding the interaction between lipid metabolism and macrophages as well as their key targets is important for preventing and treating AS and developing new drugs. Recent studies have shown that traditional Chinese medicines play a positive role in the prevention and treatment of AS, providing a basis for clinical individualized treatment.

## INTRODUCTION

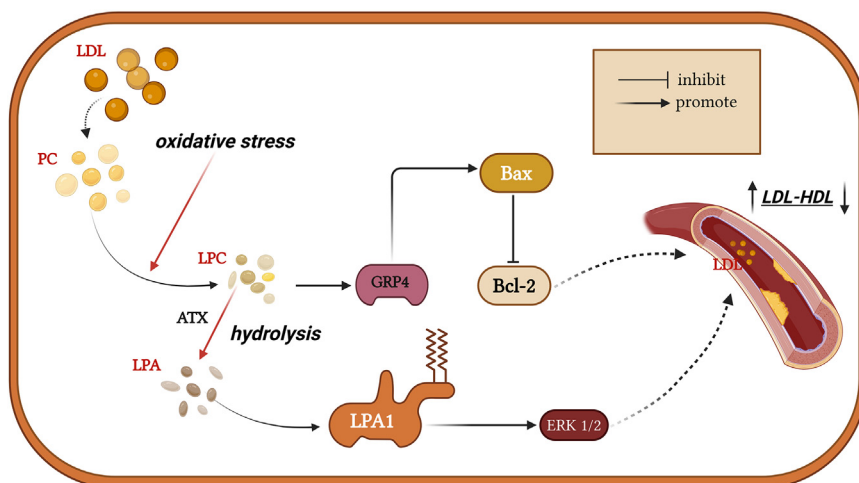
Atherosclerosis (AS) is a disease in which lipid plaque-like deposits (atheromatous plaques or atherosclerotic plaques) form in the walls of medium or large arteries, resulting in reduced blood flow or blockage of blood outflow, and is a chronic inflammatory response lesion.<sup>1–3</sup> Impaired lipid metabolism is believed to be the root cause of AS. When lipid metabolism is compromised, lipids accumulate in the plasma and then accumulate in large amounts on the inner walls of the blood vessels, forming plaques. In contrast to unstable atherosclerotic plaques, stable plaques develop at a slower rate and are less likely to result in vascular embolisms. Although statins can reduce lipid levels, they do not reduce AS-associated morbidity because the “residual risk” caused by long-term inflammation has not been eliminated, suggesting that treatment of AS cannot simply regulate lipids but should be combined with anti-inflammatory effects, so there are strategies to prevent and control AS by the interaction of lipid metabolism and inflammation.<sup>4</sup>

In complex lesions such as AS, macrophages participate in the initiation, growth, rupture, and final healing stages of arterial plaque formation. Macrophages in plaques are highly heterogeneous and plastic, enabling them to alter their phenotype and function in response to changes in the plaque microenvironment.<sup>5–7</sup> The two main phenotypes associated with AS are M1

and M2. M1 macrophages are mainly involved in immune and inflammatory responses, increasing the expression of inflammatory factors and exacerbating AS progression, whereas M2 macrophages are mainly involved in the repair and regeneration processes and produce anti-inflammatory factors to delay AS progression.<sup>8–11</sup> The M1/M2 phenotypes of macrophages are highly correlated with the inflammatory response; thus, the inter-transformation of M1 and M2 is a potential target for clinically resolving AS. Although previous studies demonstrated the influence of lipids and macrophages on AS, the relationship between lipid metabolism and macrophages, as well as the mechanism whereby their interactions affect AS, is very complex, and the relevant comprehensive elaboration still lacks. Therefore, this study describes how the interaction between lipid metabolism and macrophages influences the occurrence, development, and outcome of AS to provide direction for in-depth studies.

According to traditional Chinese medicine, the core etiology and pathogenesis of AS is “phlegm-blood stasis mutual damage/co-existence,” stimulation of blood vessels to induce toxicity, and toxic damage to the veins and channels. In a sense, the phlegm described in traditional Chinese medicine is similar to the accumulation of abnormal lipids caused by disordered lipid metabolism in Western medicine, which continuously develops into stasis deposited in the blood vessels to form plaques; this process is similar to the pathogenesis of lipid dysregulation





**Figure 1. Low-density lipoprotein (LDL)-associated lipids promote atherosclerotic lesions**

① Main constituent of LDL, phosphatidylcholine (PC), is oxidized and converted to lysophosphatidylcholine (LPC), which regulates the expression of Bax and Bcl-2 through activation of GPR4, inducing apoptosis of vascular endothelial cells, and promoting atherosclerosis; ② LPC is hydrolyzed to lysophosphatidic acid (LPA) by autotaxin (ATX). LPA activates the extracellular signal-regulated kinase 1/2 (ERK1/2) pathway by stimulating lysophosphatidic acid receptor 1 (LPA1), promoting the proliferation and migration of vascular smooth muscle cells, and contributing to atherosclerosis.

leading to inflammatory reactions in modern medicine.<sup>12–14</sup> In the process of preventing and controlling the occurrence and development of AS, we should combine the theory of formulae under the guidance of traditional Chinese medicine's diagnosis and treatment with the symptomatic treatment of pathogenesis and fully utilize the multi-target and multi-pathway characteristics of Chinese medicine.<sup>15</sup> Clinical studies have proven that the combination of activating blood stasis and clearing heat and detoxification preparations cannot only reduce blood lipid levels but also control inflammation and induce changes in the direction of macrophage polarization, which can effectively control the development of AS.<sup>16</sup> The discriminative thinking of Chinese medicine, in the modern medical environment that advocates individualized treatment, is able to combine the holistic concepts of Chinese medicine to give play to its characteristics.

## LIPID METABOLISM AND AS

### Lipids that promote the development of AS

Previous studies showed that the serum total cholesterol level is closely correlated with AS and can be used as a basis for risk stratification for a long time. Excessive accumulation and blocked efflux of cholesterol are important factors that promote AS; most lipids that contribute to AS are related to cholesterol.<sup>17–19</sup> When lipid metabolism is disrupted, the body consumes excessive amounts of lipids. The liver synthesizes high levels of very low-density lipoproteins (VLDL), which are then converted into low-density lipoproteins (LDL). LDL is a cholesterol-rich lipoprotein that carries cholesterol into cells of the peripheral tissue. An increase in the LDL concentration in the blood leads to the accumulation of excess cholesterol in the blood vessels, indicating a state of inflammation characterized by high LDL and low high-density lipoprotein (HDL) levels in the body. This condition gradually leads to the formation of atherosclerotic plaques.<sup>20–22</sup> The prevalent lipids that influence AS include LDL and its oxidation-related derivatives, as well as ceramide, among others.

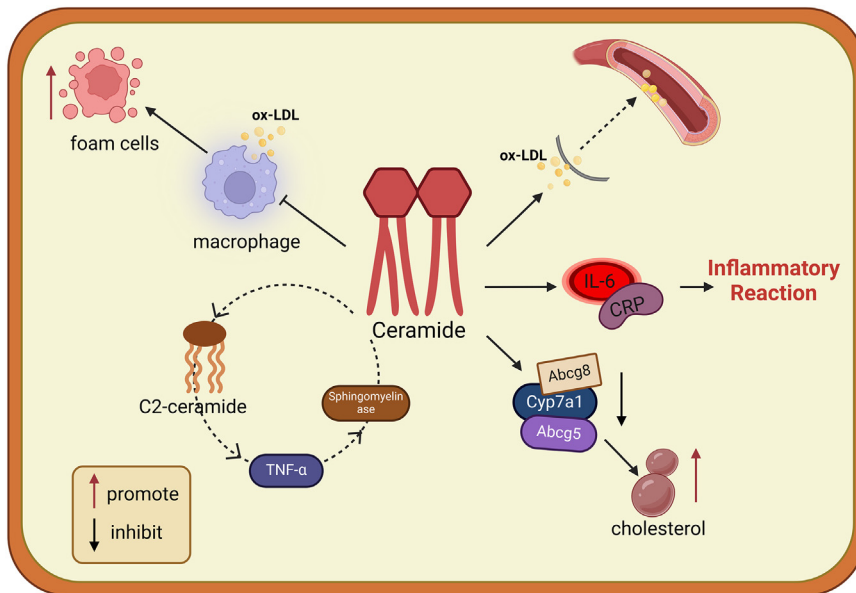
### LDL and its oxidation-associated lipids

Oxidation of LDL is a complex process. Oxidative stress in the subendothelium induces oxidative modification of LDL, which in-

creases the affinity of LDL for macrophage scavenger receptors, such as class A (SR-A1 and SR-A2) and class B (CD36) receptors, allowing the accumulation of oxidized LDL (ox-LDL) in macrophages and triggering a pro-inflammatory macrophage response. This process is thought to be a key step in AS lesion formation and progression.<sup>23–25</sup> Ox-LDL is more readily taken up by macrophages via scavenger receptors, generating foam cells, in the process largely mediated by the action of oxidized phospholipids (ox-PL). This finding was validated in a transgenic mouse assay which demonstrated that ox-PL promotes inflammation and AS.<sup>26–29</sup> Ox-PL can also interact with specific receptors on vascular cells (CD36, TLR2, and TLR4) to upregulate the expression of pro-inflammatory factors, such as interleukin (IL)-1 $\beta$ , IL-6, and IL-8, thereby triggering AS pathology.<sup>30–32</sup> Furthermore, ox-LDL acts as an important proinflammatory factor that elicits an inflammatory response through the lysophosphatidylcholine (LPC)-autotaxin-lysophosphatidic acid (LPA) pathway.<sup>33</sup> In addition, other lipids involved in LDL oxidation all affect AS, under oxidative stress, phosphatidylcholine (PC) in LDL molecules is converted to LPC, which activates G protein-coupled receptor 4 (GPR4) to regulate the expression of Bax and Bcl-2. Bcl-2 is an oncogene that inhibits apoptosis, and Bax is the most important apoptotic gene in the body. Bax can form heterodimers with Bcl-2 to inhibit Bcl-2 and induce apoptosis of vascular endothelial cells. In addition, LPC can increase vascular endothelial permeability and stimulate the expression of pro-inflammatory molecules, such as adhesion molecules and cytokines, demonstrating pro-atherogenic effects.<sup>33,34</sup> Zhaowei Cai et al. analyzed the correlation between common lipids and atherosclerotic parameters. They found that LPC was positively correlated with serum cholesterol levels, intima-media areas, and plaque areas.<sup>35</sup> Autotaxin hydrolyzes LPC to LPA, which activates the G-protein-coupled lysophosphatidic acid receptor 1 (LPA1) on the membrane of the vascular smooth muscle. This activation triggers the extracellular signal-regulated kinase 1/2 (ERK1/2) pathway, promoting smooth vascular muscle cell proliferation and migration. Ultimately, AS is induced (Figure 1).<sup>33</sup>

### Ceramides

It is worthwhile for us to explore. In addition to common lipids such as cholesterol and LDL, which promote AS, research



**Figure 2. Pathway whereby ceramides promote atherosclerosis (AS) development**

① Tumor necrosis factor (TNF)- $\alpha$  stimulates ceramide formation by activating neutral and acidic sphingomyelinase, and C2-ceramide, in turn, stimulates the expression of TNF- $\alpha$  by venous endothelial cells. TNF- $\alpha$  and ceramide may form a vicious cycle; ② ceramides exacerbate AS lesions by increasing cholesterol levels by downregulating the expression of the cholesterol elimination genes Cyp7a1, Abcg5, and Abcg8; ③ promoting foam cell formation by inhibiting macrophage digestion of oxidized low-density lipoprotein (ox-LDL), exacerbating AS; ④ by promoting *trans*-intimal transport of ox-LDL, ox-LDL is retained in the vascular endothelium, exacerbating AS; ⑤ by promoting the production of interleukin (IL)-6 and C-reactive protein (CRP), it exerts a pro-inflammatory effect and aggravates AS.

indicates that ceramide, a key component of sphingolipids, is regarded as a potential risk factor for cardiovascular disease and is involved in various physiological processes linked to AS.<sup>35–40</sup> The concentration of ceramide was notably increased in arterial plaques and showed a positive correlation with total cholesterol, triglyceride, and ox-LDL levels. This finding reveals a correlation between ceramide and AS development.<sup>37,41,42</sup> Ceramide exacerbates AS dyslipidemia by downregulating genes involved in cholesterol elimination, such as cholesterol 7 $\alpha$ -hydroxylase (Cyp7a1), ATP-binding cassette subfamily G member 5 (Abcg5), and ATP-binding cassette subfamily G member 8 (Abcg8).<sup>43</sup> It retains ox-LDL in the vascular endothelium by facilitating its translocation across the endothelium or promotes foam cell formation by impairing macrophage-digestion of aggregated LDL.<sup>44,45</sup> Furthermore, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) stimulates ceramide formation by activating neutral and acidic sphingomyelinases. C2-ceramide stimulates human umbilical vein endothelial cells to express TNF- $\alpha$ . Ceramide and TNF- $\alpha$  form a vicious circle to enhance inflammation or promote production of inflammatory factors IL-6 and C-reactive protein, which in turn affects AS (Figure 2).<sup>37,46–48</sup>

### Lipids that protect against the development of AS HDL

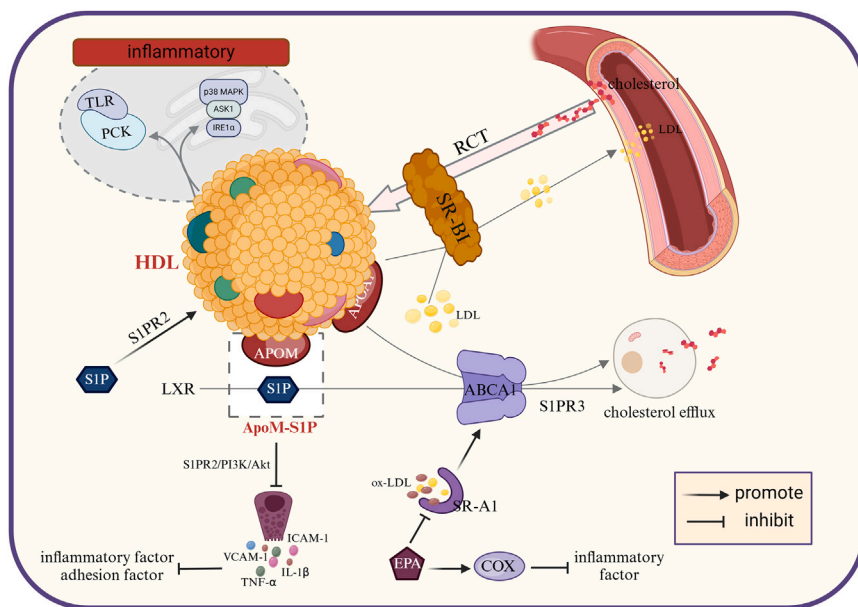
Among the lipids that cause AS, LDL, as an important subject of study, is commonly referred to as the “bad lipid,” whereas HDL, as the “good lipid,” is on the opposite side. However, recent studies have demonstrated that viewing HDL simply as a “good lipid” is overly simplistic.

HDL has a complex composition, including proteins, lipids, and their regulatory factors and indicating that HDL affects AS through multiple pathways. HDL is considered a “lipid scavenger” because of its ability to take up excess cholesterol from the vascular wall and achieve reverse cholesterol transport. The final step of reverse cholesterol transport is selective lipid

uptake of HDL-C, which is mediated by the type I class B scavenger receptor (SR-BI), which exerts a protective effect

against AS.<sup>49–52</sup> Apolipoprotein A1 (ApoA1), the most abundant apolipoprotein in HDL, helps maintain the structure of HDL and removes excess cellular cholesterol using ATP-binding cassette transporter 1 (ABCA1).<sup>53</sup> Additionally, ApoA1 competes with LDL to bind to SR-BI, thereby limiting SR-BI-mediated LDL deposition in the subarterial lumen and delaying AS.<sup>54</sup> HDL participates in the reverse transportation of ox-LDL, reduces damage caused by ox-LDL, inhibits the proliferation of vascular smooth muscle cells induced by growth factors, and prevents AS.<sup>55</sup>

Nevertheless, despite the well-established anti-atherosclerotic properties of HDL, the additional mechanisms and impacts of its intricate constituents have not been extensively explored. In recent years, several studies have begun to revisit the structure of HDL and found that HDL is a rather complex family that undergoes constant remodeling processes in circulation. HDL has anti-inflammatory, antioxidant, anti-aggregation, and anti-clotting functions in addition to its role in reversing cholesterol.<sup>56–59</sup> Its complexity is also gradually revealing that HDL has not only anti-AS properties but sometimes the opposite. Early studies focused on the anti-inflammatory effects of HDL,<sup>60–65</sup> but in recent years, investigations shows that HDL and ApoA1 also have pro-inflammatory functions. HDL exerts pro-inflammatory effects by enhancing protein kinase C activation in response to Toll-like receptor (TLR) ligands. It also activates inositol-requiring enzyme 1 $\alpha$  (IRE1 $\alpha$ )/apoptosis signal-regulating kinase-1 (ASK1)/p38 MAPK signaling due to excessive cellular cholesterol depletion, leading to pro-inflammatory endoplasmic reticulum stress responses to exert pro-inflammatory effects<sup>63,66,67</sup> (Figure 3). These findings may reveal the singularity and limitations in our search for therapeutic targets for AS. Simultaneously, they may offer new ideas for exploring targets that are more appropriate and effective for preventing and treating AS.



**Figure 3. Role of high-density lipoprotein (HDL) and related lipids in atherosclerosis**

① HDL ingests excess cholesterol from the vessel wall, enabling reverse cholesterol transport (RCT); ② APOA1, the most abundant apolipoprotein on HDL, removes excess cellular cholesterol via ATP-binding cassette transporter ABCA1; ③ APOA1 competes with LDL for SR-B1 binding and limits SR-B1-mediated low-density lipoprotein (LDL) deposition in the subarterial lumen; ④ HDL responds to Toll-like receptor (TLR) ligands to enhance PCK activation exerting pro-inflammatory effects; ⑤ activation of the IRE1 $\alpha$ /ASK1/p38 MAPK signaling pathway leads to a pro-inflammatory endoplasmic reticulum stress response, resulting in pro-inflammatory effects; ⑥ ApoM-S1P inhibits the secretion of IL-1 $\beta$ , TNF- $\alpha$ , ICAM-1, and VCAM-1 from human umbilical vein endothelial cells (HUVECs) through the S1PR2/PI3K/Akt signaling pathway and downregulates the expression of inflammatory and adhesion factors; ⑦ S1P loading enhances the anti-inflammatory effect of HDL via S1PR2; ⑧ S1P regulates cholesterol outflow in a positive feedback manner dependent on S1PR3 via ABCA1 outputs; ⑨ EPA has many functions, including inhibiting the

expression of inflammatory factors through cyclooxygenase (COX), decreasing the gene expression of the scavenger receptor SR-A1, reducing the phagocytosis of ox-LDL, and further increasing ABCA1 expression, thereby promoting cholesterol and phospholipid output.

### Sphingosine-1-phosphate

Sphingosine-1-phosphate (S1P), a lipid closely related to HDL, is produced by two isotypes of sphingosine kinase, namely SphK-1 and SphK-2. S1P functions as a signaling molecule in cell membrane signal transduction pathways. Both S1P and ceramide are called “bioactive sphingolipids,” but their functions in the development of atherosclerosis are antagonistic.<sup>68,69</sup> S1P binds to apolipoprotein M (apoM), one of the essential components of HDL. ApoM-S1P protects endothelial cells by inhibiting ox-LDL-induced secretion of IL-1 $\beta$ , TNF- $\alpha$ , ICAM-1, and VCAM-1 from human umbilical vein endothelial cells (HUVECs) through the S1PR2/PI3K/Akt signaling pathway. This inhibition significantly downregulates the expression of inflammatory and adhesion factors.<sup>70,71</sup> Therapeutic S1P loading via S1PR2 enhances the anti-inflammatory effects of HDL. S1P is also previously identified as an unidentified intermediate in the liver X receptors (LXR)-stimulated ABCA1-mediated cholesterol efflux and can delay AS progression by being exported from macrophages via ABCA1 and regulating cholesterol efflux in a positive feedback manner dependent on S1PR3<sup>72–74</sup> (Figure 3). Furthermore, research has demonstrated that S1P possesses various roles pertinent to macrophage physiology and is capable of inhibiting the activation of inflammatory macrophages through multiple mechanisms. For example, S1P selectively weakens Toll-like receptor 2 signal transduction through negative crosstalk of S1PR1/2 and Toll-like receptor 2 signal transduction pathways in macrophages, thereby preventing inflammatory macrophage activation.<sup>75</sup> Mitigate inflammation through the suppression of nuclear factor kappa B (NF- $\kappa$ B) activity in macrophages.<sup>76</sup> The activation of S1P elicited a Th2 response, which resulted in a substantial decrease in macrophage populations due to the knockout of SphK1.<sup>77</sup> In light of the aforementioned experimental findings, it would be reasonable to investigate

the relationship between S1P and diseases associated with M2 anti-inflammatory macrophages, such as AS.

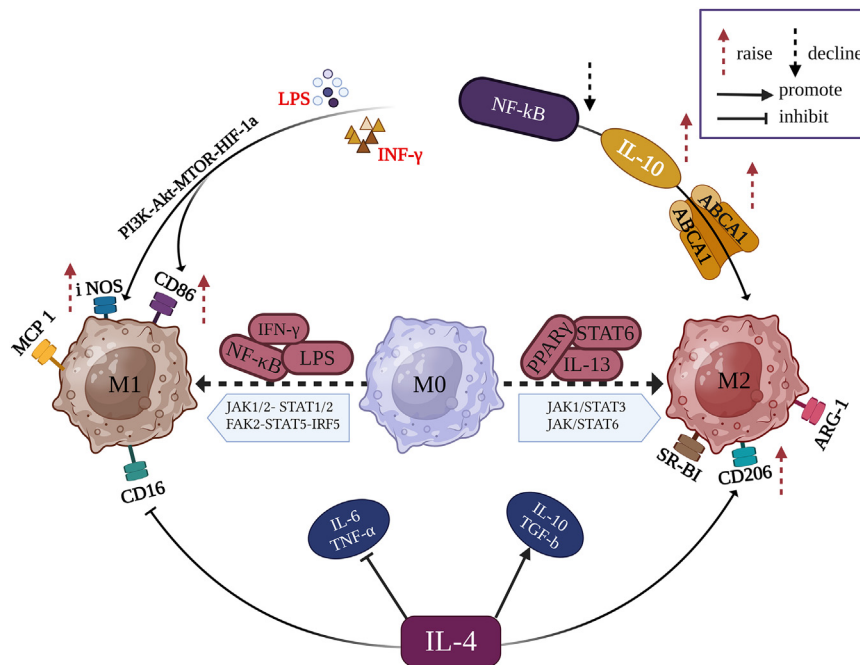
### Other lipids

In addition to the two lipids mentioned previously, it is more noteworthy that polyunsaturated fatty acids act together from multiple species and pathways.<sup>78</sup> Omega-3 polyunsaturated fatty acid (PUFA), an important group of polyunsaturated fatty acids, delays AS progression by reducing the expression of inflammatory molecules, decreasing fat synthesis, and promoting lipolysis.<sup>79</sup> For instance, EPA inhibits the expression of inflammatory factors by producing anti-inflammatory products via cyclooxygenase (COX) and reduces the formation of AS by decreasing the gene expression of scavenger receptors (SR-A1 and CD36), lowering the phagocytosis of ox-LDL, and further increasing the expression of ABCA1 to enhance the efflux of cholesterol and phospholipids<sup>79–81</sup> (Figure 3). In addition, lipids, such as nitro-oleic acid (NO<sub>2</sub>-OA), oxysterols, and conjugated linoleic acid (CLA) exert anti-AS effects by reducing lipid accumulation, increasing cholesterol efflux, and decreasing pro-inflammatory factor production. These effects have been demonstrated in numerous experiments.<sup>82–85</sup> In a word, lipid metabolism disorder, as the basis of AS, makes lipid play a dual role in regulating AS. However, it cannot be ignored that in the case of complex lesions such as AS, macrophages participate in the whole process and also have multiple regulatory effects on the development of AS, especially the different phenotypes under the high plasticity of macrophages.

### MACROPHAGE POLARIZATION

#### Macrophage polarization under classical pathways

Macrophages can be transformed into M1 macrophages through the JAK1/2- signal transduction and transcriptional



**Figure 4. Summary of the polarization environment of macrophages**

phenotype.<sup>94</sup> In addition, the glycolytic pathway is a metabolic signaling pathway, and the lactate it produces is a key mediator of macrophage polarization. An increase in lactate levels activates hypoxia-inducible factor (HIF)-1 $\alpha$  and HIF-2 $\alpha$ , promoting M1 macrophages and M2 macrophages, respectively.<sup>95</sup> Lactate produced by endothelial cell glycolysis can also reverse M2 macrophage polarization through monocarboxylate transporter-1-dependent signal transduction.<sup>96</sup>

### Macrophage polarization in AS

In the case of complex AS lesions, the plasticity of macrophages significantly affect AS progression.<sup>97,98</sup> M1-type polarization, also known as pro-inflammatory polarization, releases pro-inflammatory factors TNF- $\alpha$  and IL-1 $\beta$ . It exhibits a low cholesterol scavenging capacity, destabilizes plaques, and promotes the development of AS. In contrast, M2 polarization has an anti-inflammatory effect; these cells secrete cytokines such as those found in inflammatory zone 1 (Fizz1), arginase 1 (Arg-1), and IL-10. M2 polarization also leads to low foam cell formation. These cells exhibit high phagocytic activity, an increased ability to clear apoptotic cells, and can produce anti-inflammatory factors that inhibit the development of AS.<sup>10,99</sup> Furthermore, IL-1 $\beta$ , MCP1, and iNOS can be used as M1-type cell markers, while ARG1, IL-10, IL-4, and CD206 can be used as M2-type cell markers. The levels of M1 and M2 markers reflect the transformation of M1-and M2-type macrophages, respectively. Altering the environment in which the two macrophages are located is a viable strategy for achieving from M1 to M2 polarization. This transformation from M1 to M2 in AS has gradually become a research goal that we are pursuing. Notably, macrophage plasticity is not purely M1 and M2 polarization; with advances in technology, more phenotypes of macrophages have been identified and recognized. The M2 phenotype alone can be categorized as M2a, M2b, M2c, and M2d; normally, M2 macrophages exhibit anti-inflammatory effects, yet M2b macrophages deviate from this anti-inflammatory mode as they maintain high levels of inflammatory cytokine production.<sup>100</sup> In addition to the M1 and M2 phenotypes, the M(Hb), Mhem, Mox, and M4 phenotypes have also been detected, and it is precisely because of the diversity of macrophage phenotypes that the complete reflection of the *in vivo* situation by the M1/M2 phenotypes has become more complex and difficult,<sup>6</sup> but most current studies still classify macrophages into the M1 and M2 classes to evaluate their properties and functions; therefore working on finding a complete pathway from M1 to M2 is also more conducive to realizing a true reflection of the *in vivo* situation by M1/M2.<sup>101</sup>

activator (STAT)1/2 and FAK2-STAT5-IRF5 pathways in the presence of lipopolysaccharide (LPS), IFN- $\gamma$ , and NF- $\kappa$ B. In the presence of IL-4, PPAR $\gamma$ , STAT3, STAT6, interferon regulatory factor 4 (IRF4), and a hypoxic environment, macrophages transform into M2 macrophages through the JAK1/STAT3 signaling pathway and JAK/STAT6 signaling pathway.<sup>86</sup> Macrophages treated with LPS/IFN- $\gamma$  showed elevated expression of iNOS and CD86. LPS induced macrophage polarization toward the M1 phenotype through the PI3K-Akt-MTOR-HIF-1 $\alpha$  signaling pathway, which was further enhanced by IFN- $\gamma$ . In IL-4-treated cells and mice, the expression of JAK1 and STAT6 was significantly elevated, promoting macrophage polarization toward the M2 subtype by activating Wnt- $\beta$ -catenin signaling through enhanced nuclear translocation of  $\beta$ -catenin<sup>87–90</sup> (Figure 4).

### Macrophage polarization under non-classical pathway

In addition to the classical pathway, macrophage phenotypes also undergo specific shifts in the context of autophagy and glycolysis. Impaired autophagy has been shown to facilitate the polarization of macrophages toward the M1 phenotype.<sup>91</sup> Conversely, enhancing autophagic flux in macrophages through the action of ubiquitin-specific protease 19 (USP 19) or through the administration of small molecule compounds that stimulate autophagy can drive macrophage polarization toward the M2 phenotype. In the regulation of autophagy, mTOR and NF- $\kappa$ B are involved. Inhibiting the mTOR pathway and stimulating macrophages to polarize toward M1. The induction of NF- $\kappa$ B p65 cytoplasmic ubiquitination leads to its degradation by p62-mediated autophagy. At the same time, inhibition of NF- $\kappa$ B activity saved by autophagy drives macrophages to shift to the M2 phenotype.<sup>92,93</sup> CCL2 and IL-6, which are effective factors for macrophages to induce autophagy, can also trigger the M2

NF- $\kappa$ B plays a crucial role in the development of AS. Inhibiting the NF- $\kappa$ B regulatory pathway reduces aortic stiffness, mitigates inflammation, enhances IL-10 secretion, upregulates ABCA1 expression to facilitate cholesterol efflux from foam cells, and triggers the transformation of macrophages into the M2 type.<sup>8</sup> For instance, rutin and hirudin He-D inhibit macrophage migration by targeting IK $\alpha$  and IK $\gamma$  in the NF- $\kappa$ B signaling pathway; n-Butyric acid (NBA) mediated by GPR43- $\beta$ -receptorstatin-2 and lipoxin A4 (LXA4) inhibits IRF5 activity and promotes interferon regulatory factor 4 (IRF4) activation via FPR2-mediated inhibition. These processes downregulate p-NF- $\kappa$ B p65 activity, reducing M1 marker expression and increasing the mRNA levels of M2 markers, facilitating the transformation of macrophages from the M1 phenotype to the M2 phenotype, thereby improving the inflammatory response of macrophages. Consequently, NF- $\kappa$ B plays a role in AS.<sup>102–105</sup> With the current technological advancements, M1 to M2 conversion is no longer limited to drug research. New technologies and therapies have emerged that show potential for use in treating AS, including sonodynamic therapy (SDT), a treatment developed from photodynamic therapy.<sup>106–108</sup> Previous studies,<sup>109</sup> showed that SDT narrowed and stabilized advanced AS plaques by modulating M1 to M2 macrophage polarization. It enhanced anti-AS effects and had an earlier onset of action than statins, achieving M1 to M2 conversion at a technical level. In conclusion, the discovery of more target pathways for M1 to M2 conversion in AS is expected to reverse AS progression and prevent AS-related acute events.

## LIPID METABOLISM AND MACROPHAGE POLARIZATION

In the complex lesions of AS, lipid-lowering through lipid metabolism alone or the anti-inflammatory actions of macrophages is effective treatment that has been extensively studied. Previously, we described inflammatory lipids such as ox-LDL and ox-PL promote the development of AS. In contrast, anti-inflammatory lipids, such as HDL, S1P, and EPA delay AS development. Additionally, distinct macrophage phenotypes are known to have differential impacts on the course of AS progression, which further complicates the understanding of this multifactorial disease. However, whether lipid metabolism and macrophages interact with each other to affect AS has not been systematically reported. Therefore, herein, we focused on their interactions and related mechanisms to explore whether lipid metabolism and macrophage interactions can serve as potentially effective targets for the clinical treatment of AS, and provide more information for new drug research and development in the treatment of AS.

### Interaction between lipid metabolism and macrophage polarization

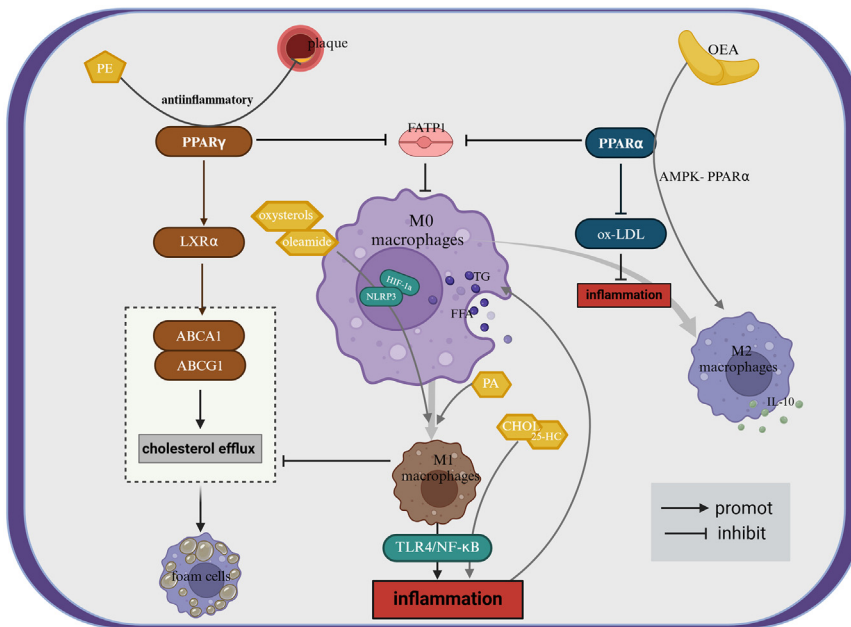
In AS lesions, macrophages are exposed to accumulated lipids and their derivatives. Lipid metabolism plays a crucial and indispensable role in modulating the function of macrophages, with particular emphasis on fatty acids. Metabolic discrepancies in fatty acid biosynthesis (FA) and fatty acid oxidation (FAO) induce distinct phenotypes of macrophages.<sup>110</sup> Research indicates that microRNA-33 (miR-33) facilitates the development of the inflam-

matory M1-like macrophage phenotype, which is linked to metabolic disorders. Conversely, the suppression of miR-33 has been shown to metabolically reprogram macrophages toward the M2 phenotype by enhancing FAO. It is posited that miR-33 plays a regulatory role in the polarization of macrophages between the M1 and M2 phenotypes by modulating the equilibrium between cellular glycolysis and FAO.<sup>111</sup> At the same time, as phagocytes, macrophages take up different forms of lipids, such as LDL, VLDL, and oxidized lipoproteins, from phagocytosed dying cells and the microenvironment through phagocytosis, macropinocytosis, and scavenger receptor-mediated pathways. The ingested lipids are processed by acid lipases in lysosomes, resulting in the production of free fatty acids and cholesterol. And the generation of M2 macrophages is partly dependent on fatty acid uptake and oxidation, and one of the sources of fatty acids is through lysosomal lipolysis uptake mediated by lysosomal acid lipases.<sup>112,113</sup> In macrophages, sterol regulatory element-binding proteins (SREBPs) and LXRs are expressed at high levels, exerting influence on the transcriptional regulation of lipid metabolism and modulating the release of cytokines. LPS treatment augments the activity of macrophage SREBP-1a via NF- $\kappa$ B, and macrophages with SREBP-1a deficiency are unable to generate inflammatory factors subsequent to LPS stimulation, thereby indicating that lipid metabolism in M1 macrophages is correlated with inflammasome activation.<sup>114</sup> In contrast, M2 macrophages are distinguished by the activation of LXR, which regulates cholesterol homeostasis and lipid synthesis. Overexpression or activation of LXR $\alpha$  is capable of suppressing the M1 response and inflammation through the inhibition of NF- $\kappa$ B activity.<sup>115</sup>

In general, these findings elucidate the interaction between lipid metabolism and macrophage polarization, thereby providing significant support for the research on metabolic diseases, particularly AS. Subsequently, we will discuss the relationship between specific macrophage phenotypes and lipid metabolism, as well as their influence on AS.

### Lipid metabolism and macrophage polarization in AS Lipid metabolism and M1 macrophages in AS

In the preceding text, we have discussed the impact of ox-LDL on the occurrence of AS through the accumulation of foam cells. Herein, we will first conduct a further exploration of the relationship between ox-LDL and macrophage polarization and investigate its influence on AS. Ox-LDL is capable of promoting the signal transduction of the PI3K/Akt/mTOR pathway and blocking the autophagic process of macrophages, resulting in an increase in p62 expression.<sup>116</sup> The accumulated p62 activates NF- $\kappa$ B through the phosphorylation of inhibitor of  $\kappa$ B kinase (IKK), leading to the nuclear translocation of NF- $\kappa$ B, thereby facilitating the transcription of inflammatory factors and inducing M1 polarization. Additionally, ox-LDL binds to the receptor CD36, causing the uptake of long-chain fatty acids (LCFAs). In macrophages stimulated by ox-LDL, the expression of fatty acid-binding protein 4 (FABP 4) and acyl-CoA synthetase 1 (ASCL 1) is upregulated,<sup>117,118</sup> and after processing the LCFA, it is transported to the mitochondrial matrix through carnitine palmitoyl transferase CPTI and CPTII.<sup>119,120</sup> Consequently, LCFA accumulates in mitochondria, leading to the downregulation of FAO and mitochondrial oxidative phosphorylation (OXPHOS), thereby promoting



**Figure 5. Mechanisms associated with “lipid-inflammatory” interactions in atherosclerosis (AS)**

① Palmitic acid (PA) converts macrophages to the M1 type, which exacerbate the inflammatory response by upregulating the TLR4/NF- $\kappa$ B signaling pathway; ② excess free cholesterol and 25-hydroxycholesterol (25-HC) promote NF- $\kappa$ B-mediated inflammatory responses through the TLR4 pathway; ③ oxysterols and oleamide polarize macrophages toward the M1 type by activating HIF-1 $\alpha$  or NLRP3 inflammatory vesicles in macrophages; ④ activation of PPAR $\alpha$  and PPAR $\gamma$  attenuates free fatty acid (FFA) and TG accumulation in macrophages by inhibiting FATP1; ⑤ activation of PPAR $\alpha$  and PPAR $\gamma$  attenuates FFA and TG accumulation in macrophages by inhibiting FATP1; ⑥ macrophages accelerate the conversion to foam cells by reducing the expression of ABCA1 and ABCG1 expression; ⑦ phosphatidylethanolamine (PE) improves AS plaque stability through PPAR $\gamma$ -mediated anti-inflammatory properties; ⑧ oleoyl ethanol amide (OEA) activates PPAR $\alpha$ , which, inhibits ox-LDL production and reverses the inflammatory response; it promotes M2 macrophage polarization via the AMPK-PPAR $\alpha$  pathway.

macrophage M1 polarization and exacerbating AS lesions. Furthermore, within macrophages, the TLR/NF- $\kappa$ B pathway is a significant player associated with inflammation, and more lipids exert the macrophage polarization effect under the classical pathway through this signaling. The saturated fatty acid palmitic acid (PA), which converts macrophages to the M1 type, induces M1 macrophages, and exacerbates the inflammatory response by upregulating the TLR4/NF- $\kappa$ B signaling pathway. This, in turn, promotes lipid synthesis in cells. Excess free cholesterol and 25-hydroxycholesterol (25-HC) increase susceptibility to apoptosis by activating the p38 mitogen-activated protein kinase (p38MAPK) signaling pathway or promoting NF- $\kappa$ B-mediated expression of pro-inflammatory genes via the Toll-like receptor 4 (TLR4) pathway. These steps enhance inflammatory responses in lipid-hyperlipidemic macrophages, thereby exacerbating AS plaque instability<sup>121,122</sup>; the same is true for oxidized lipoproteins and cholesterol linoleic acid (Figure 5).<sup>123–125</sup> It is worth noting that in the relationship between lipid metabolism and macrophage polarization, some lipids directly affect M1 polarization through inflammatory factors. For example, oxysterol and oleamide promote the production of the pro-inflammatory factor IL-1 $\beta$  by activating HIF-1 $\alpha$  or NLRP3 inflammasome in macrophages, creating an environment conducive to the polarization of M1 macrophages.<sup>126,127</sup> Specifically, ox-PL upregulates the expression of activating transcription factor 4 (ATF4) directly through the PERK/eIF2 $\alpha$  axis, thereby promoting M1-type macrophage.<sup>128</sup> In contrast, M1-type macrophages, as pro-inflammatory macrophages, release several pro-inflammatory factors, such as TNF- $\alpha$ , IL-1 $\beta$ , and IL-6, which influence lipid efflux and, thus, AS development. M1-type macrophages downregulate cholesterol efflux, develop pathological processes such as vascular endothelial cell damage, platelet aggregation, and vascular smooth muscle cell pro-

liferation. Therefore, decreasing the expression of scavenger receptors and cholesterol efflux transporter proteins, thereby decreasing the ability to remove lipids from plaques, promoting the accumulation of lipids within the lesion, and accelerating the transformation of macrophages into foam cells; the released TNF- $\alpha$  also triggers the adipocyte inflammatory response, exacerbating the release of inflammatory free fatty acids (FAA) from adipocytes, while secreting MCP-1, leading to the further infiltration of monocytes from the bloodstream.<sup>129–132</sup> In summary, lipids and M1 macrophages can interact through multiple pathways, exacerbating the formation of AS plaques (Table 1).

#### Lipid metabolism and M2 macrophages in AS

The relationship between M2 polarization and lipids in macrophages is somewhat simpler compared to that of M1 polarization. Most physiologically active lipids inhibit the production of inflammatory factors and promote the secretion of M2 macrophage markers, placing macrophages in an environment that favors M2 polarization, which in turn promotes macrophage polarization toward M2-type macrophages. Phosphatidyl serine liposome (PSL) stimulation reduces the expression of pro-inflammatory cytokines IL-1 $\alpha$ , IL-1 $\beta$ , IL-12, IL-6, and TNF- $\alpha$ ; enhances the secretion of IL-10 and TGF- $\beta$  and the expression of the M2 macrophage biomarker CD206; and provides an environment conducive to the differentiation of macrophages toward M2, thus phosphatidylserine (PS) significantly enhances the M2-like polarization of macrophages via the PSRS-STAT3-JMJD3 pathway.<sup>133</sup> Phosphatidyl ethanolamine (PE) decreases the expression of inflammatory factors NOS, CCL2, ICAM-1, and VCAM-1; increases the aortic expression of SR-BI, a phenotypic marker of M2 macrophages; and promotes the transformation of macrophages into an anti-inflammatory M2 type. The same is true of the polyunsaturated fatty acid docosahexaenoic acid (DHA).<sup>134</sup> Moreover, a certain degree of M2 polarization is

**Table 1. The interaction between lipid metabolism and macrophage polarization**

Polarization pathway	Mode of action	Lipids	Lipid impact macrophage signaling pathways	Direction of polarization	Macrophages influence lipid metabolic pathways
Classical pathway	Indirect	Saturated fatty acid palmitic acid	/	M1	TLR4/NF- $\kappa$ B
		Cholesterol/25-hydroxycholesterol	TLR4/p38MAPK/NF- $\kappa$ B	M1	Increased susceptibility to apoptosis
		Oxidized lipoproteins	TLR4/NF- $\kappa$ B	M1	Activate the inflammation factor
		Cholesterol linoleic acid	TLR4/NF- $\kappa$ B	M1	Activate the inflammation factor
		Phosphatidylserine	PSRS-STAT3-JMJD	M2	Raised cholesterol efflux gene
	Direct	Oxysterol	Activate HIF-1a/NLRP3 and promote IL-1b production	M1	Decrease the expression of scavenger receptor and cholesterol efflux transporter
		Oleamide	Activate HIF-1a/NLRP3 and promote IL-1b production	M1	Decrease the expression of scavenger receptor and cholesterol efflux transporter
		ox-PL	Upregulation of ATF4 through PERK/eIF2 $\alpha$ axis	M1	Decrease the expression of scavenger receptor and cholesterol efflux transporter
		Phosphatidyl serine liposome	Reduce proinflammatory cytokines, enhance CD206 expression	M2	Raised cholesterol efflux gene
		Phosphatidyl ethanolamine	Reduce inflammation factor expression, increase SR - B expression	M2	Raised cholesterol efflux gene
Non-classical pathway	Autophagy	ox-LDL	Block macrophage autophagy, increase P62 expression, and activate NF- $\kappa$ B via IKK	M1	Inflammatory cytokines produced
		ox-LDL	Downregulation of FAO and OXPHOS	M1	Lipid accumulation
	Metabolic reprogramming	Fatty acid	Biosynthesis	M1	Activate the inflammasome
		Fatty acid	Uptake and oxidation	M2	Pathways mediated by phagocytosis, macropinocytosis, and scavenger receptors

reliant on the uptake and oxidation of fatty acids, and it promotes wound-healing processes via IL-4 and IL-13 signal transduction (Table 1). Polarized M2 macrophages enhance the production of anti-inflammatory substances, have anti-inflammatory, repair, and regenerative functions, affect the accumulation and efflux of lipids, clear lipid deposits and cell debris in blood vessels, promote the repair of vascular endothelial cells and the remodeling of blood vessel walls, downregulate the expression of SR and the uptake of cholesterol genes, and upregulate the efflux genes of cholesterol through the secretion of IL-10. These processes reduce the uptake of lipids, increase the exclusion of lipids, reduce the accumulation of lipids in cells, and then delay the development of AS.<sup>135,136</sup> However, not all lipids are involved in M2 polarization impede AS development, as a “bioactive sphingolipid”, S1P induces polarization of M2-type macrophages through IL-4 secretion and signal transduction, thereby upregulating cholesterol efferent genes and downregulating cholesterol uptake genes, reducing cholesterol accumulation, and inhibiting apoptosis signals, namely the activation of cas-

pase 3 and 8, thereby enhancing the stability of AS plaques. However, by inhibiting S1P lyase, a long-term increase in endogenous S1P levels promotes AS,<sup>135,137</sup> contradicting the results of previous studies and may lead to an exciting new area of research. In conclusion, this review elaborates on the impact of the interaction between lipid metabolism and macrophage polarization on AS, which may offer new insights for the development of individualized clinical treatment regimens involving combined lipid-inflammation medications. Furthermore, in-depth exploration of the interaction mechanism between the two will also contribute to the discovery of new drugs and new therapeutic targets for AS treatment.

#### **“Lipid-inflammatory” interaction mechanism between lipid metabolism and macrophage polarization**

In the foregoing content, we delved into the intricate relationship between lipid metabolism and macrophage polarization within the context of AS. Our exploration revealed that the reciprocal influence between these two elements predominantly converges on the “lipid-inflammatory” interplay. Beyond the well-established

TLR/NF- $\kappa$ B canonical pathway, the peroxisome proliferator-activated receptor (PPAR) emerges as a pivotal target that demands meticulous consideration. Previous studies showed that activation of PPAR $\alpha$  and PPAR $\gamma$  attenuates accumulation of total FFAs and triglycerides in macrophages by inhibiting fatty acid transporter 1 (FATP1).<sup>138</sup> PPAR $\gamma$  participates in the regulation of adipocyte differentiation and lipid metabolism, regulates the expression of cholesterol efflux genes including ABCA1 and ABCG1 by activating LXR $\alpha$ , inhibits cholesterol outflow by blocking the PPAR $\gamma$ -LXR $\alpha$ -ABCA1/ABCG1 pathway, thus regulating lipid accumulation in AS plaques.<sup>139–142</sup> Furthermore, activation of PPAR $\gamma$  effectively suppresses macrophage inflammation and likely facilitates the conversion of pro-inflammatory M1 macrophages into anti-inflammatory M2 macrophages. In contrast, inhibition of PPAR $\gamma$  is associated with a decreased M1/M2 ratio of macrophages. This evidence suggests that PPAR $\gamma$  co-regulates AS through lipid-lowering and anti-inflammatory mechanisms.<sup>143,144</sup> For instance, phosphatidylethanolamine (PE), a multifunctional phospholipid rich in cells, improves the stability of AS plaques through PPAR $\gamma$ -mediated anti-inflammatory properties; oleoyl ethanol amide (OEA), an endogenous PPAR $\alpha$  ligand, activates PPAR  $\alpha$  and then inhibits the production of ox-LDL in mouse macrophages, reversing the ox-LDL-induced pathological effects in AS animals, including AS plaque formation and inflammation. Additionally, it promotes M2 macrophage polarization and inhibits M1 macrophage polarization through the AMPK-PPAR  $\alpha$  pathway (Figure 5).<sup>134,145–147</sup> Overall, these lipids affect the direction of macrophage polarization through the anti-inflammatory and lipid-regulating effects of PPAR, thus affecting AS. PPAR may become a very important research target to identify the interaction between lipid metabolism and macrophage polarization and its effects on AS.

### THE STRATEGY OF TRADITIONAL CHINESE MEDICINE TO PREVENT AND CONTROL AS BY REGULATING BLOOD LIPID AND ANTI-INFLAMMATION

For AS, the most widely used lipid-lowering drugs in the clinic in Western countries include niacin, phenoxy aromatic acids, and statins.<sup>148</sup> Statins, the most widely studied drugs, can slow AS by reducing lipid levels<sup>149–151</sup> but are associated with side effects such as AS muscle pain, liver injury, and increased diabetes risk.<sup>152</sup> Therefore, clinical research on alternative AS drugs has increased.

Traditional Chinese medicines, characterized by minimal toxic and side effects, have the unique advantage of multi-target and multi-component synergy in disease management. They align closely with the treatment strategies for AS. Chinese medicine posits that herbs that promote blood circulation, alleviate blood stasis, and eliminate heat and toxins can greatly contribute to reducing inflammation, combating fat accumulation, providing antioxidant effects, and preventing thrombosis. In addition, these herbs are believed to play crucial roles in delaying AS progression.<sup>153</sup> Based on the specific categorization of effects, salidroside has been shown to significantly decrease levels of cholesterol, fatty acids, and other biosynthetic products, thereby stabilizing AS by downregulating the gene expression of sterol regulatory element binding proteins (Srebf1 and Srebf2). Ethyl

acetate extract of *Patrinia villosa* (PVJEE) has an anti-AS effect by downregulating the LPC of glycerol phospholipid metabolic pathway and inhibiting the apoptosis of vascular endothelial cells. Gypenosides promote autophagy and affect LncTUG1/miR-26a to improve lipid deposition and prevent and treat atherosclerosis.<sup>148,154,155</sup> Curcumin upregulates the expression of PPAR $\gamma$  in M1 macrophages, which in turn promotes the expression of CD36 and ABCA1. By supporting cholesterol homeostasis, curcumin exerts an anti-atherosclerotic effect.<sup>97</sup> Secondly, in terms of anti-inflammation, ginsenoside Rb1 demonstrates its effects by inducing the production of anti-inflammatory cytokines such as IL-4 and IL-13, as well as by suppressing the synthesis of pro-inflammatory factors. Additionally, it facilitates the polarization of M2 macrophages through various signaling pathways, including JAK-STAT, PPARS, and AMPK, thereby contributing to the attenuation of AS.<sup>156–158</sup> Similarly, laminaran enhance the expression of genes associated with anti-inflammatory factors while inhibiting the expression of genes linked to inflammation. Furthermore, they promote autophagic activity in macrophages and encourage the polarization of these cells toward the M2 phenotype by modulating relevant signaling pathways, thus playing a role in the prevention and progression of AS.<sup>159</sup> Tanshinone II A has been shown to effectively inhibit the M1 polarization of macrophages induced by ox-LDL, with the underlying mechanism potentially involving the suppression of miR-375, which activates KLF4.<sup>160</sup> Furthermore, convallaria toxin (CNT), a naturally occurring cardioside, has been found to reduce the secretion of inflammatory cytokines, activate the PPAR $\gamma$  signaling pathway, and promote the M2 phenotype of macrophages, thereby contributing to the regulation of AS.<sup>161</sup> In addition to the previously discussed active monomer components, it is worth mentioning the role of traditional Chinese medicine compounds. Huang-Lian-Jie-Du decoction, recognized as a representative prescription for heat-clearing and detoxification in Chinese medicine, has been shown to reduce the expression of pro-inflammatory factors associated with M1 macrophage polarization. Simultaneously, it enhances the expression of CD163 and Arg1 markers linked to M2 polarization, thereby influencing the progression of AS by modulating macrophage polarization.<sup>162</sup> The Shenlian formula, an empirically derived clinical formulation for the treatment of AS, is supported by multiple lines of evidence demonstrating its efficacy in alleviating AS. This is achieved through mechanisms such as promoting LAL-LXR $\alpha$ -mediated cholesterol ester degradation and cholesterol efflux, modulating inflammatory responses, and improving blood flow and lipid profiles.<sup>163–166</sup> In conclusion, the results of these experiments indicate that both the active monomer components of traditional Chinese medicine and its composite compounds play a role in the prevention and management of AS through various mechanisms related to lipid metabolism and macrophage polarization (Table 2).

### CONCLUSION AND OUTLOOK

AS pathogenesis is complex and abnormal lipid metabolism and inflammatory reactions occur throughout its occurrence and development. Many lipid components in the body are important for the maintenance of physiological functions. When the

**Table 2. Monomer composition and compound prescription of traditional Chinese medicine regulating the development of AS**

Type of action	lipids	Signaling pathways	Role purpose	Affect the AS
Reduce lipids	Salidroside	Downregulated Srebf1 and Srebf2 expression	Decrease of lipid synthesis	Stable AS plaque
	PVJEE	Downregulate the metabolic pathway of glycerol phospholipid	Inhibition of vascular endothelial cell apoptosis	Anti-as effect
	Gypenosides	Promote autophagy and affect Inctug1/mir-26a	Improve lipid deposition	Mitigate the occurrence of AS
	Curcumin	Raised ppar $\gamma$ expression, and promote the CD36/ABCA1 expression	Supports cholesterol homeostasis	Stable AS plaque
	Shenlian formula	LAL-lxr $\alpha$	Cholesterol efflux	Mitigate the occurrence of AS
Anti-inflammation	Ginsenoside Rb1	JAK-STAT/PPARS/AMPK	Promote the M2 macrophage polarization	Anti-AS effect
	Laminaran	Inhibit the expression of inflammation-related cytokines	Promote the M2 macrophage polarization	Anti-AS effect
	Tanshinone II A	Inhibition of mir-375 activated KLF4	Prevent polarization of M1 macrophages	Anti-AS effect
	Convallaria toxin (CNT)	Activate the ppar $\gamma$ signaling pathway	Promote the M2 macrophage polarization	Anti-AS effect
	Huang-Lian-Jie-Du decoction	Decreased the expression of proinflammatory factor, increased the expression of CD 163 and Arg 1	Promote M1 polarization to M2	Anti-AS effect

composition, quantity, and proportion of lipids change, their metabolic balance in the body is disrupted; the number and proportion of oleamide, ceramide, lysophosphatidylcholine, and other pro-inflammatory lipids increase; and macrophages are stimulated to M1 type transformation via a series of signaling pathways. However, although lipid-lowering drugs used to treat AS can reduce blood lipid levels, they often fail to have the expected effect on AS development and incidence of cardiovascular and cerebrovascular diseases.<sup>167</sup> This leads the researchers to further study and find that improving the inflammatory response can also regulate lipid levels in AS plaques.<sup>168,169</sup> Lipids such as phosphatidylethanolamine, S1P, phosphatidylserine, and  $\omega$ -3 PUFA in plaques can promote the polarization of M2 macrophages and play an anti-inflammatory role. The proposal of “lipid-inflammation” interactions provides clues for the diagnosis and treatment idea for clinical treatment of AS. This review aims to expand the scope of drug targets for the clinical management of AS by elucidating the interrelationships, conditions, and pathways of reciprocal alterations between lipid metabolism and macrophage-mediated inflammatory responses. It suggests that it will become a trend to develop personalized treatment regimens for combined drug use, tailored to patients’ lipid metabolism and macrophage polarization. Through syndrome differentiation and treatment and prescription compatibility, traditional Chinese medicine plays a role in regulating the occurrence and development of AS. However, further treatment based on syndrome differentiation and scientific compatibility will play a more important role in preventing and treating AS and reducing the incidence of cardiovascular and cerebrovascular diseases caused by lipid metabolism and macrophage polar-

ization. At the same time, it can also prove that the prescription of traditional Chinese medicine is scientific and effective.

## LIMITATIONS OF THE STUDY

With the development of technology, more and more phenotypes of macrophages are being identified, and only M2 phenotypes can be divided into M2a, M2b, M2c, and M2d. M2 macrophages usually show anti-inflammatory effects, but M2b macrophages deviate from this anti-inflammatory model because they maintain high levels of inflammatory cytokines. In this article we only study the general M2 phenotype, its more detailed classifications and functions have not been systematically reported, which will be included in our next research plan.

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## AUTHOR CONTRIBUTIONS

X.W. and Z.X. contributed equally in this work; conceptualization, Y.D. and S.X.; writing—original draft preparation, X.W. and Z.X.; writing—review and editing, X.W., Z.X., J.Z., Y.C., and Q.L.; supervision, Q.Y.; project administration, X.C. and B.L. All authors have read and agreed to the published version of the manuscript.

## DECLARATION OF INTERESTS

The authors declare no competing interests.

# REFERENCES

1. Wolf, D., and Ley, K. (2019). Immunity and Inflammation in Atherosclerosis. *Circ. Res.* 124, 315–327. <https://doi.org/10.1161/CIRCRESAHA.118.313591>.
2. Rasheed, A., and Rayner, K.J. (2021). Macrophage Responses to Environmental Stimuli During Homeostasis and Disease. *Endocr. Rev.* 42, 407–435. <https://doi.org/10.1210/edrv/bnab004>.
3. Zhu, Y., Xian, X., Wang, Z., Bi, Y., Chen, Q., Han, X., Tang, D., and Chen, R. (2018). Research Progress on the Relationship between Atherosclerosis and Inflammation. *Biomolecules* 8, 80. <https://doi.org/10.3390/biom8030080>.
4. Pradhan, A.D., Aday, A.W., Rose, L.M., and Ridker, P.M. (2018). Residual Inflammatory Risk on Treatment With PCSK9 Inhibition and Statin Therapy. *Circulation* 138, 141–149. <https://doi.org/10.1161/CIRCULATIONAHA.118.034645>.
5. Zhao, P., Zhou, W., Zhang, Y., Li, J., Zhao, Y., Pan, L., Shen, Z., Chen, W., and Hui, J. (2020). Aminoxyacetic acid attenuates post-infarct cardiac dysfunction by balancing macrophage polarization through modulating macrophage metabolism in mice. *J. Cell Mol. Med.* 24, 2593–2609. <https://doi.org/10.1111/jcmm.14972>.
6. Hou, P., Fang, J., Liu, Z., Shi, Y., Agostini, M., Bernassola, F., Bove, P., Candi, E., Rovella, V., Sica, G., et al. (2023). Macrophage polarization and metabolism in atherosclerosis. *Cell Death Dis.* 14, 691. <https://doi.org/10.1038/s41419-023-06206-z>.
7. Huang, J., Ding, X., Dong, Y., and Zhu, H. (2024). Growth Differentiation Factor-15 Orchestrates Inflammation-Related Diseases via Macrophage Polarization. *Discov. Med.* 36, 248–255. <https://doi.org/10.24976/Discover.Med.202436181.23>.
8. Gong, L., Suo, Y., Wang, X., and Guo, M. (2016). Research Progress on TCM Prevention and Treatment of Atherosclerosis Based on the Differentiation of Macrophage Subtypes. *J. Basic Chin. Med.* 22, 723–726. <https://doi.org/10.19945/j.cnki.issn.1006-3250.2016.05.054>.
9. Zhou, Q., Sun, H., Yu, D., and Liu, S. (2020). Mechanisms of M1/M2 macrophage polarization in different diseases. *Chin. Pharmacol. Bull.* 36, 1502–1506.
10. Leitinger, N., and Schulman, I.G. (2013). Phenotypic polarization of macrophages in atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* 33, 1120–1126. <https://doi.org/10.1161/ATVBAHA.112.300173>.
11. Muñoz-García, J., Cochonneau, D., Télétchéa, S., Moranton, E., Lanoe, D., Brion, R., Lézot, F., Heymann, M.-F., and Heymann, D. (2021). The twin cytokines interleukin-34 and CSF-1: masterful conductors of macrophage homeostasis. *Theranostics* 11, 1568–1593. <https://doi.org/10.7150/thno.50683>.
12. HU, X., CHI, L., ZHANG, W., DUAN, N., and FENG, Z. (2014). Developments in Inflammation Mechanism of Atherosclerosis and Pleiotropic Effects of Atorvastatin. *Adv. Cardiovasc. Dis.* 35, 261–265.
13. SHI, L., and ZHAO, C. (2021). Research Progress in Treating Atherosclerosis with Traditional Chinese Medicine. *Mod. Diagn. Treat.* 32, 1528–1530.
14. WANG, D., FENG, R., SHI, S., WEI, N., and HU, Y. (2021). Research progress on anti-atherosclerosis mechanism of phlegm and stasis treatment. *China Med. Her.* 18, 53–55. <https://doi.org/10.20047/j.issn1673-7210.2021.30.013>.
15. PEI, Y., YANG, G., CHEN, Z., SHAO, Y., CHENG, Z., SONG, N., WEN, Z., DUAN, J., and ZHANG, Z. (2020). Establishment of New Theoretical System for Chinese Medical Principle of Treatment from Phlegm and Blood Stasis, Strengthening Spleen Being the Key in Treatment of Atherosclerosis. *Chin. Arch. Tradit. Chin. Med.* 38, 32–34. <https://doi.org/10.13193/j.issn.1673-7717.2020.08.007>.
16. Wang, S. (2018). Overview of Components of Simiao Yong'an Decoction and Pharmacological Mechanisms in the Treatment of Atherosclerosis. *Tradit. Chin. Med.* 7, 7–12. <https://doi.org/10.12677/TCM.2018.71002>.
17. Tarasov, K., Ekroos, K., Suoniemi, M., Kauhanen, D., Sylvänne, T., Hurme, R., Gouni-Berthold, I., Berthold, H.K., Kleber, M.E., Laaksonen, R., and März, W. (2014). Molecular lipids identify cardiovascular risk and are efficiently lowered by simvastatin and PCSK9 deficiency. *J. Clin. Endocrinol. Metab.* 99, E45–E52. <https://doi.org/10.1210/jc.2013-2559>.
18. Wang, D., Yang, Y., Lei, Y., Tzvetkov, N.T., Liu, X., Yeung, A.W.K., Xu, S., and Atanasov, A.G. (2019). Targeting Foam Cell Formation in Atherosclerosis: Therapeutic Potential of Natural Products. *Pharmacol. Rev.* 71, 596–670. <https://doi.org/10.1124/pr.118.017178>.
19. Gao, L.-N., Zhou, X., Lu, Y.-R., Li, K., Gao, S., Yu, C.-Q., and Cui, Y.-L. (2018). Dan-Lou Prescription Inhibits Foam Cell Formation Induced by ox-LDL via the TLR4/NF-κB and PPARγ Signaling Pathways. *Front. Physiol.* 9, 590. <https://doi.org/10.3389/fphys.2018.00590>.
20. Rahmati-Ahmadabad, S., Broom, D.R., Ghanbari-Niaki, A., and Shirvani, H. (2019). Effects of exercise on reverse cholesterol transport: A systematized narrative review of animal studies. *Life Sci.* 224, 139–148. <https://doi.org/10.1016/j.lfs.2019.03.058>.
21. Plat, J., Baumgartner, S., Vanmierlo, T., Lütjohann, D., Calkins, K.L., Burin, D.G., Guthrie, G., Thijs, C., Te Velde, A.A., Vreugdenhil, A.C.E., et al. (2019). Plant-based sterols and stanols in health & disease: “Consequences of human development in a plant-based environment?”. *Prog. Lipid Res.* 74, 87–102. <https://doi.org/10.1016/j.plipres.2019.02.003>.
22. Ding, X., Zhang, W., Li, S., and Yang, H. (2019). The role of cholesterol metabolism in cancer. *Am. J. Cancer Res.* 9, 219–227.
23. Weber, C., and Noels, H. (2011). Atherosclerosis: current pathogenesis and therapeutic options. *Nat. Med.* 17, 1410–1422. <https://doi.org/10.1038/nm.2538>.
24. Soppert, J., Lehrke, M., Marx, N., Jankowski, J., and Noels, H. (2020). Lipoproteins and lipids in cardiovascular disease: from mechanistic insights to therapeutic targeting. *Adv. Drug Deliv. Rev.* 159, 4–33. <https://doi.org/10.1016/j.addr.2020.07.019>.
25. Moore, K.J., and Freeman, M.W. (2006). Scavenger receptors in atherosclerosis: beyond lipid uptake. *Arterioscler. Thromb. Vasc. Biol.* 26, 1702–1711. <https://doi.org/10.1161/01.ATV.0000229218.97976.43>.
26. Devaraj, S., and Jialal, I. (1996). Oxidized low-density lipoprotein and atherosclerosis. *Int. J. Clin. Lab. Res.* 26, 178–184.
27. Que, X., Hung, M.-Y., Yeang, C., Gonen, A., Prohaska, T.A., Sun, X., Diehl, C., Määttä, A., Gaddis, D.E., Bowden, K., et al. (2018). Oxidized phospholipids are proinflammatory and proatherogenic in hypercholesterolaemic mice. *Nature* 558, 301–306. <https://doi.org/10.1038/s41586-018-0198-8>.
28. van der Valk, F.M., Bekkering, S., Kroon, J., Yeang, C., Van den Bossche, J., van Buul, J.D., Ravandi, A., Nederveen, A.J., Verberne, H.J., Scipione, C., et al. (2016). Oxidized Phospholipids on Lipoprotein(a) Elicit Arterial Wall Inflammation and an Inflammatory Monocyte Response in Humans. *Circulation* 134, 611–624. <https://doi.org/10.1161/CIRCULATIONAHA.116.020838>.
29. Ambrogini, E., Que, X., Wang, S., Yamaguchi, F., Weinstein, R.S., Tsimikas, S., Manolagas, S.C., Witztum, J.L., and Jilka, R.L. (2018). Oxidation-specific epitopes restrain bone formation. *Nat. Commun.* 9, 2193. <https://doi.org/10.1038/s41467-018-04047-5>.
30. Witztum, J.L., and Lichtman, A.H. (2014). The Influence of Innate and Adaptive Immune Responses on Atherosclerosis. *Annu. Rev. Pathol.* 9, 73–102. <https://doi.org/10.1146/annurev-pathol-020712-163936>.
31. Tsimikas, S., and Witztum, J.L. (2024). Oxidized phospholipids in cardiovascular disease. *Nat. Rev. Cardiol.* 21, 170–191. <https://doi.org/10.1038/s41569-023-00937-4>.
32. Sun, X., Seidman, J.S., Zhao, P., Troutman, T.D., Spann, N.J., Que, X., Zhou, F., Liao, Z., Pasillas, M., Yang, X., et al. (2020). Neutralization of Oxidized Phospholipids Ameliorates Non-alcoholic Steatohepatitis. *Cell Metab.* 31, 189–206.e8. <https://doi.org/10.1016/j.cmet.2019.10.014>.

33. Bao, L., Qi, J., Wang, Y.W., Xi, Q., Tserennadmid, T., Zhao, P.F., Qi, J., and Damirin, A. (2018). The atherogenic actions of LPC on vascular smooth muscle cells and its LPA receptor mediated mechanism. *Biochem. Biophys. Res. Commun.* 503, 1911–1918. <https://doi.org/10.1016/j.bbrc.2018.07.135>.
34. Leskova, G.F., Kaplun, A.P., Bezrukov, D.A., and Lvovsky, A.I. (2020). Effect of Phosphatidylcholine Nanosomes on Phospholipid Composition of the Plasma Membranes in Liver Cells and Blood Serum in Experimental Atherosclerosis. *Bull. Exp. Biol. Med.* 170, 181–184. <https://doi.org/10.1007/s10517-020-05028-9>.
35. Cai, Z., Deng, L., Fan, Y., Ren, Y., Ling, Y., Tu, J., Cai, Y., Xu, X., and Chen, M. (2023). Dysregulation of Ceramide Metabolism Is Linked to Iron Deposition and Activation of Related Pathways in the Aorta of Atherosclerotic Miniature Pigs. *Antioxidants* 13, 4. <https://doi.org/10.3390/antiox13010004>.
36. Cogolludo, A., Villamor, E., Perez-Vizcaino, F., and Moreno, L. (2019). Ceramide and Regulation of Vascular Tone. *Int. J. Mol. Sci.* 20, 411. <https://doi.org/10.3390/ijms20020411>.
37. Shu, H., Peng, Y., Hang, W., Li, N., Zhou, N., and Wang, D.W. (2022). Emerging Roles of Ceramide in Cardiovascular Diseases. *Aging Dis.* 13, 232–245. <https://doi.org/10.14336/AD.2021.0710>.
38. Uchida, Y., Uchida, Y., Kobayashi, T., Shirai, S., Hiruta, N., Shimoyama, E., and Tabata, T. (2017). Detection of Ceramide, a Risk Factor for Coronary Artery Disease, in Human Coronary Plaques by Fluorescent Angioscopy. *Circ. J.* 81, 1886–1893. <https://doi.org/10.1253/circj.CJ-17-0363>.
39. Vandanmagsar, B., Youm, Y.-H., Ravussin, A., Galgani, J.E., Stadler, K., Mynatt, R.L., Ravussin, E., Stephens, J.M., and Dixit, V.D. (2011). The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat. Med.* 17, 179–188. <https://doi.org/10.1038/nm.2279>.
40. York, A.G., Skadow, M.H., Oh, J., Qu, R., Zhou, Q.D., Hsieh, W.-Y., Mow, W.K., Brewer, J.R., Kaffé, E., Williams, K.J., et al. (2024). IL-10 constrains sphingolipid metabolism to limit inflammation. *Nature* 627, 628–635.
41. Ichi, I., Nakahara, K., Miyashita, Y., Hidaka, A., Kutsukake, S., Inoue, K., Maruyama, T., Miwa, Y., Harada-Shiba, M., Tsumishima, M., et al. (2006). Association of ceramides in human plasma with risk factors of atherosclerosis. *Lipids* 41, 859–863. <https://doi.org/10.1007/s11745-006-5041-6>.
42. Edsfieldt, A., Dunér, P., Ståhlman, M., Mollet, I.G., Ascietto, G., Grufman, H., Nitulescu, M., Persson, A.F., Fisher, R.M., Melander, O., et al. (2016). Sphingolipids Contribute to Human Atherosclerotic Plaque Inflammation. *Arterioscler. Thromb. Vasc. Biol.* 36, 1132–1140. <https://doi.org/10.1161/ATVBAHA.116.305675>.
43. Zhang, X., Zhang, Y., Wang, P., Zhang, S.-Y., Dong, Y., Zeng, G., Yan, Y., Sun, L., Wu, Q., Liu, H., et al. (2019). Adipocyte Hypoxia-Inducible Factor 2 $\alpha$  Suppresses Atherosclerosis by Promoting Adipose Ceramide Catabolism. *Cell Metab.* 30, 937–951.e5. <https://doi.org/10.1016/j.cmet.2019.09.016>.
44. Li, W., Yang, X., Xing, S., Bian, F., Yao, W., Bai, X., Zheng, T., Wu, G., and Jin, S. (2014). Endogenous ceramide contributes to the transcytosis of oxLDL across endothelial cells and promotes its subendothelial retention in vascular wall. *Oxid. Med. Cell. Longev.* 2014, 823071. <https://doi.org/10.1155/2014/823071>.
45. Singh, R.K., Haka, A.S., Brumfield, A., Grosheva, I., Bhardwaj, P., Chin, H.F., Xiong, Y., Hla, T., and Maxfield, F.R. (2017). Ceramide activation of RhoA/Rho kinase impairs actin polymerization during aggregated LDL catabolism. *J. Lipid Res.* 58, 1977–1987. <https://doi.org/10.1194/jlr.M076398>.
46. Marcos-Ramiro, B., García-Weber, D., and Millán, J. (2014). TNF-induced endothelial barrier disruption: beyond actin and Rho. *Thromb. Haemost.* 112, 1088–1102. <https://doi.org/10.1160/TH14-04-0299>.
47. Sawada, M., Kiyono, T., Nakashima, S., Shinoda, J., Naganawa, T., Hara, S., Iwama, T., and Sakai, N. (2004). Molecular mechanisms of TNF- $\alpha$ -induced ceramide formation in human glioma cells: P53-mediated oxidant stress-dependent and -independent pathways. *Cell Death Differ.* 11, 997–1008. <https://doi.org/10.1038/sj.cdd.4401438>.
48. Hirokawa, M., Kitabayashi, A., Kuroki, J., and Miura, A.B. (2000). Induction of tissue factor production but not the upregulation of adhesion molecule expression by ceramide in human vascular endothelial cells. *Tohoku J. Exp. Med.* 191, 167–176. <https://doi.org/10.1620/tjem.191.167>.
49. Acton, S., Rigotti, A., Landschulz, K.T., Xu, S., Hobbs, H.H., and Krieger, M. (1996). Identification of Scavenger Receptor SR-BI as a High Density Lipoprotein Receptor. *Science* 271, 518–520. <https://doi.org/10.1126/science.271.5248.518>.
50. Ren, K., Zhu, X., Zheng, Z., Mo, Z.-C., Peng, X.-S., Zeng, Y.-Z., Ou, H.-X., Zhang, Q.-H., Qi, H.-Z., Zhao, G.-J., and Yi, G.H. (2018). MicroRNA-24 aggravates atherosclerosis by inhibiting selective lipid uptake from HDL cholesterol via the post-transcriptional repression of scavenger receptor class B type I. *Atherosclerosis* 270, 57–67. <https://doi.org/10.1016/j.atherosclerosis.2018.01.045>.
51. Feig, J.E., Shamir, R., and Fisher, E.A. (2008). Atheroprotective effects of HDL: beyond reverse cholesterol transport. *Curr. Drug Targets* 9, 196–203. <https://doi.org/10.2174/138945008783755557>.
52. Rader, D.J. (2006). Molecular regulation of HDL metabolism and function: implications for novel therapies. *J. Clin. Investig.* 116, 3090–3100. <https://doi.org/10.1172/JCI30163>.
53. von Eckardstein, A., Nordestgaard, B.G., Remaley, A.T., and Catapano, A.L. (2023). High-density lipoprotein revisited: biological functions and clinical relevance. *Eur. Heart J.* 44, 1394–1407. <https://doi.org/10.1093/eurheartj/ehac605>.
54. Fung, K.Y.Y., Ho, T.W.W., Xu, Z., Neculai, D., Beauchemin, C.A.A., Lee, W.L., and Fair, G.D. (2024). Apolipoprotein A1 and high-density lipoprotein limit low-density lipoprotein transcytosis by binding SR-B1. *J. Lipid Res.* 65, 100530. <https://doi.org/10.1016/j.jlr.2024.100530>.
55. Tian, D. (2003). Research progress of high density lipoprotein and its receptors. *J. Int. Pharm. Res.* 1, 1–5. <https://doi.org/10.13220/j.cnki.jipr.2003.01.001>.
56. Pirillo, A., Catapano, A.L., and Norata, G.D. (2019). Biological Consequences of Dysfunctional HDL. *Curr. Med. Chem.* 26, 1644–1664. <https://doi.org/10.2174/0929867325666180530110543>.
57. Kosmas, C.E., Sourlas, A., Guzman, E., and Kostara, C.E. (2022). Environmental Factors Modifying HDL Functionality. *Curr. Med. Chem.* 29, 1687–1701. <https://doi.org/10.2174/0929867328666210714155422>.
58. Darabi, M., and Kontush, A. (2022). High-density lipoproteins (HDL): Novel function and therapeutic applications. *Biochim. Biophys. Acta. Mol. Cell Biol. Lipids* 1867, 159058. <https://doi.org/10.1016/j.bbalip.2021.159058>.
59. Jia, C., Anderson, J.L.C., Gruppen, E.G., Lei, Y., Bakker, S.J.L., Dullaart, R.P.F., and Tietge, U.J.F. (2021). High-Density Lipoprotein Anti-Inflammatory Capacity and Incident Cardiovascular Events. *Circulation* 143, 1935–1945. <https://doi.org/10.1161/CIRCULATIONAHA.120.050808>.
60. Thacker, S.G., Zarzour, A., Chen, Y., Alciçek, M.S., Freeman, L.A., Sviridov, D.O., Demosky, S.J., and Remaley, A.T. (2016). High-density lipoprotein reduces inflammation from cholesterol crystals by inhibiting inflammasome activation. *Immunology* 149, 306–319. <https://doi.org/10.1111/imm.12638>.
61. Murphy, A.J., Woollard, K.J., Hoang, A., Mukhamedova, N., Stirzaker, R.A., McCormick, S.P.A., Remaley, A.T., Sviridov, D., and Chin-Dusting, J. (2008). High-density lipoprotein reduces the human monocyte inflammatory response. *Arterioscler. Thromb. Vasc. Biol.* 28, 2071–2077. <https://doi.org/10.1161/ATVBAHA.108.168690>.
62. De Nardo, D., Labzin, L.I., Kono, H., Seki, R., Schmidt, S.V., Beyer, M., Xu, D., Zimmer, S., Lahmann, C., Schildberg, F.A., et al. (2014). High-density lipoprotein mediates anti-inflammatory reprogramming of

- macrophages via the transcriptional regulator ATF3. *Nat. Immunol.* 15, 152–160. <https://doi.org/10.1038/ni.2784>.
63. Fotakis, P., Kothari, V., Thomas, D.G., Westerterp, M., Molusky, M.M., Altin, E., Abramowicz, S., Wang, N., He, Y., Heinecke, J.W., et al. (2019). Anti-Inflammatory Effects of HDL (High-Density Lipoprotein) in Macrophages Predominate Over Proinflammatory Effects in Atherosclerotic Plaques. *Arterioscler. Thromb. Vasc. Biol.* 39, e253–e272. <https://doi.org/10.1161/ATVBAHA.119.313253>.
64. Yvan-Charvet, L., Kling, J., Pagler, T., Li, H., Hubbard, B., Fisher, T., Sparrow, C.P., Taggart, A.K., and Tall, A.R. (2010). Cholesterol efflux potential and antiinflammatory properties of high-density lipoprotein after treatment with niacin or anacetrapib. *Arterioscler. Thromb. Vasc. Biol.* 30, 1430–1438. <https://doi.org/10.1161/ATVBAHA.110.207142>.
65. Suzuki, M., Pritchard, D.K., Becker, L., Hoofnagle, A.N., Tanimura, N., Bammler, T.K., Beyer, R.P., Bumgarner, R., Vaisar, T., de Beer, M.C., et al. (2010). High-density lipoprotein suppresses the type I interferon response, a family of potent antiviral immunoregulators, in macrophages challenged with lipopolysaccharide. *Circulation* 122, 1919–1927. <https://doi.org/10.1161/CIRCULATIONAHA.110.961193>.
66. Smoak, K.A., Aloor, J.J., Madenspacher, J., Merrick, B.A., Collins, J.B., Zhu, X., Cavigiolio, G., Oda, M.N., Parks, J.S., and Fessler, M.B. (2010). Myeloid differentiation primary response protein 88 couples reverse cholesterol transport to inflammation. *Cell Metab.* 11, 493–502. <https://doi.org/10.1016/j.cmet.2010.04.006>.
67. van der Vorst, E.P.C., Theodorou, K., Wu, Y., Hoeksema, M.A., Goossens, P., Bursill, C.A., Aliyev, T., Huitema, L.F.A., Tas, S.W., Wolfs, I.M.J., et al. (2017). High-Density Lipoproteins Exert Pro-inflammatory Effects on Macrophages via Passive Cholesterol Depletion and PKC-NF- $\kappa$ B/STAT1-IRF1 Signaling. *Cell Metab.* 25, 197–207. <https://doi.org/10.1016/j.cmet.2016.10.013>.
68. Manzo, O.L., Nour, J., Sasset, L., Marino, A., Rubinelli, L., Palikhe, S., Smimmo, M., Hu, Y., Bucci, M.R., Borczuk, A., et al. (2024). Rewiring Endothelial Sphingolipid Metabolism to Favor S1P Over Ceramide Protects From Coronary Atherosclerosis. *Circ. Res.* 134, 990–1005. <https://doi.org/10.1161/CIRCRESAHA.123.323826>.
69. Piccoli, M., Cirillo, F., Ghiroldi, A., Rota, P., Coviello, S., Tarantino, A., La Rocca, P., Lavata, I., Creo, P., Signorelli, P., et al. (2023). Sphingolipids and Atherosclerosis: The Dual Role of Ceramide and Sphingosine-1-Phosphate. *Antioxidants* 12, 143. <https://doi.org/10.3390/antiox12010143>.
70. Zheng, Z., Zeng, Y., Zhu, X., Tan, Y., Li, Y., Li, Q., and Yi, G. (2019). ApoM-S1P Modulates Ox-LDL-Induced Inflammation Through the PI3K/Akt Signaling Pathway in HUVECs. *Inflammation* 42, 606–617. <https://doi.org/10.1007/s10753-018-0918-0>.
71. Blaho, V.A., Galvani, S., Engelbrecht, E., Liu, C., Swendeman, S.L., Kono, M., Proia, R.L., Steinman, L., Han, M.H., and Hla, T. (2015). HDL-bound sphingosine-1-phosphate restrains lymphopoiesis and neuroinflammation. *Nature* 523, 342–346. <https://doi.org/10.1038/nature14462>.
72. Keul, P., Polzin, A., Kaiser, K., Gräler, M., Dannenberg, L., Daum, G., Heusch, G., and Levkau, B. (2019). Potent anti-inflammatory properties of HDL in vascular smooth muscle cells mediated by HDL-S1P and their impairment in coronary artery disease due to lower HDL-S1P: a new aspect of HDL dysfunction and its therapy. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* 33, 1482–1495. <https://doi.org/10.1096/fj.201801245R>.
73. Bot, M., Van Veldhoven, P.P., de Jager, S.C.A., Johnson, J., Nijstad, N., Van Santbrink, P.J., Westra, M.M., Van Der Hoeven, G., Gijbels, M.J., Müller-Tidow, C., et al. (2013). Hematopoietic sphingosine 1-phosphate lyase deficiency decreases atherosclerotic lesion development in LDL-receptor deficient mice. *PLoS One* 8, e63360. <https://doi.org/10.1371/journal.pone.0063360>.
74. Vaidya, M., Jentsch, J.A., Peters, S., Keul, P., Weske, S., Gräler, M.H., Mladenov, E., Iliakis, G., Heusch, G., and Levkau, B. (2019). Regulation of ABCA1-mediated cholesterol efflux by sphingosine-1-phosphate signaling in macrophages. *J. Lipid Res.* 60, 506–515. <https://doi.org/10.1194/jlr.M088443>.
75. Hammad, S.M., Crellin, H.G., Wu, B.X., Melton, J., Anelli, V., and Obeid, L.M. (2008). Dual and distinct roles for sphingosine kinase 1 and sphingosine 1 phosphate in the response to inflammatory stimuli in RAW macrophages. *Prostag. Other Lipid Mediat.* 85, 107–114. <https://doi.org/10.1016/j.prostaglandins.2007.11.002>.
76. Dueñas, A.I., Aceves, M., Fernández-Pisonero, I., Gómez, C., Orduña, A., Crespo, M.S., and García-Rodríguez, C. (2008). Selective attenuation of Toll-like receptor 2 signalling may explain the atheroprotective effect of sphingosine 1-phosphate. *Cardiovasc. Res.* 79, 537–544. <https://doi.org/10.1093/cvr/cvn087>.
77. Lai, W.-Q., Goh, H.H., Bao, Z., Wong, W.S.F., Melendez, A.J., and Leung, B.P. (2008). The Role of Sphingosine Kinase in a Murine Model of Allergic Asthma. *J. Immunol.* 180, 4323–4329. <https://doi.org/10.4049/jimmunol.180.6.4323>.
78. Ménégaut, L., Jalil, A., Thomas, C., and Masson, D. (2019). Macrophage fatty acid metabolism and atherosclerosis: The rise of PUFAs. *Atherosclerosis* 291, 52–61. <https://doi.org/10.1016/j.atherosclerosis.2019.10.002>.
79. Cao, Y., Wang, W.-Q., Chen, C., Qin, Y.-T., and Guo, X.-M. (2018). Structure and metabolism of  $\omega$ -3 polyunsaturated fatty acid and its relationship with atherosclerosis. *Chin. J. Arterioscler.* 26, 633–643.
80. Zhao, Y., Yao, L., Zhang, X., and Zhu, Y. (2020). Atherosclerosis protection mechanism of omega-3 polyunsaturated fatty acid metabolites. *Chin. J. Arterioscler.* 28, 461–467.
81. Song, Y., Zhang, L.-J., Li, H., Gu, Y., Li, F.-F., Jiang, L.-N., Liu, F., Ye, J., and Li, Q. (2013). Polyunsaturated fatty acid relatively decreases cholesterol content in THP-1 macrophage-derived foam cell: partly correlates with expression profile of CIDE and PAT members. *Lipids Health Dis.* 12, 111. <https://doi.org/10.1186/1476-511X-12-111>.
82. Vazquez, M.M., Gutierrez, M.V., Salvatore, S.R., Puiatti, M., Dato, V.A., Chiabrando, G.A., Freeman, B.A., Schopfer, F.J., and Bonacci, G. (2020). Nitro-oleic acid, a ligand of CD36, reduces cholesterol accumulation by modulating oxidized-LDL uptake and cholesterol efflux in RAW264.7 macrophages. *Redox Biol.* 36, 101591. <https://doi.org/10.1016/j.redox.2020.101591>.
83. Adamson, S., and Leitinger, N. (2011). Phenotypic modulation of macrophages in response to plaque lipids. *Curr. Opin. Lipidol.* 22, 335–342. <https://doi.org/10.1097/MOL.0b013e32834a97e4>.
84. Leonarduzzi, G., Gargiulo, S., Gamba, P., Perrelli, M.G., Castellano, I., Sapino, A., Sottero, B., and Poli, G. (2010). Molecular signaling operated by a diet-compatible mixture of oxysterols in up-regulating CD36 receptor in CD68 positive cells. *Mol. Nutr. Food Res.* 54, S31–S41. <https://doi.org/10.1002/mnfr.200900493>.
85. McClelland, S., Cox, C., O'Connor, R., de Gaetano, M., McCarthy, C., Cryan, L., Fitzgerald, D., and Belton, O. (2010). Conjugated linoleic acid suppresses the migratory and inflammatory phenotype of the monocyte/macrophage cell. *Atherosclerosis* 211, 96–102. <https://doi.org/10.1016/j.atherosclerosis.2010.02.003>.
86. Guan, X., and Liu, P. (2019). Advances in relationship between macrophage polarization and atherosclerosis and intervention effects of traditional Chinese medicine. *Shanghai J. Tradit. Chin. Med.* 53, 105–108. <https://doi.org/10.16305/j.1007-1334.2019.03.024>.
87. Zhang, T., Xiang, C., Xia, Y., Ma, L., and Zhang, B. (2021). Non-targeted lipomics analysis of M1 macrophages. *Chin. J. Pathophysiol.* 37, 711–717.
88. Gong, M., Zhuo, X., and Ma, A. (2017). STAT6 Upregulation Promotes M2 Macrophage Polarization to Suppress Atherosclerosis. *Med. Sci. Monit. Basic Res.* 23, 240–249. <https://doi.org/10.12659/msmbr.904014>.
89. He, Y., Gao, Y., Zhang, Q., Zhou, G., Cao, F., and Yao, S. (2020). IL-4 Switches Microglia/macrophage M1/M2 Polarization and Alleviates Neurological Damage by Modulating the JAK1/STAT6 Pathway Following ICH. *Neuroscience* 437, 161–171. <https://doi.org/10.1016/j.neuroscience.2020.03.008>.

90. Yang, S., Yuan, H.-Q., Hao, Y.-M., Ren, Z., Qu, S.-L., Liu, L.-S., Wei, D.-H., Tang, Z.-H., Zhang, J.-F., and Jiang, Z.-S. (2020). Macrophage polarization in atherosclerosis. *Clin. Chim. Acta* 507, 142–146. <https://doi.org/10.1016/j.cca.2019.10.034>.
91. Liu, T., Wang, L., Liang, P., Wang, X., Liu, Y., Cai, J., She, Y., Wang, D., Wang, Z., Guo, Z., et al. (2021). USP19 suppresses inflammation and promotes M2-like macrophage polarization by manipulating NLRP3 function via autophagy. *Cell. Mol. Immunol.* 18, 2431–2442. <https://doi.org/10.1038/s41423-020-00567-7>.
92. Chang, C.-P., Su, Y.-C., Lee, P.-H., and Lei, H.-Y. (2013). Targeting NFKB by autophagy to polarize hepatoma-associated macrophage differentiation. *Autophagy* 9, 619–621. <https://doi.org/10.4161/auto.23546>.
93. Chang, C.-P., Su, Y.-C., Hu, C.-W., and Lei, H.-Y. (2013). TLR2-dependent selective autophagy regulates NF- $\kappa$ B lysosomal degradation in hepatoma-derived M2 macrophage differentiation. *Cell Death Differ.* 20, 515–523. <https://doi.org/10.1038/cdd.2012.146>.
94. Roca, H., Varsos, Z.S., Sud, S., Craig, M.J., Ying, C., and Pienta, K.J. (2009). CCL2 and Interleukin-6 Promote Survival of Human CD11b+ Peripheral Blood Mononuclear Cells and Induce M2-type Macrophage Polarization. *J. Biol. Chem.* 284, 34342–34354. <https://doi.org/10.1074/jbc.M109.042671>.
95. Sanjurjo, L., Castelblanco, E., Julve, J., Villalmanzo, N., Téllez, É., Ramirez-Morros, A., Alonso, N., Mauricio, D., and Sarrias, M.-R. (2023). Contribution of Elevated Glucose and Oxidized LDL to Macrophage Inflammation: A Role for PRAS40/Akt-Dependent Shedding of Soluble CD14. *Antioxidants* 12, 1083. <https://doi.org/10.3390/antiox12051083>.
96. Zhang, J., Muri, J., Fitzgerald, G., Gorski, T., Gianni-Barrera, R., Maschlein, E., D'Hulst, G., Gilardoni, P., Turiel, G., Fan, Z., et al. (2020). Endothelial Lactate Controls Muscle Regeneration from Ischemia by Inducing M2-like Macrophage Polarization. *Cell Metab.* 37, 1136–1153.e7. <https://doi.org/10.1016/j.cmet.2020.05.004>.
97. Momtazi-Borojeni, A.A., Abdollahi, E., Nikfar, B., Chaichian, S., and Ekhlas-Hundrieser, M. (2019). Curcumin as a potential modulator of M1 and M2 macrophages: new insights in atherosclerosis therapy. *Heart Fail. Rev.* 24, 399–409. <https://doi.org/10.1007/s10741-018-09764-z>.
98. Lee, S.G., Oh, J., Bong, S.K., Kim, J.S., Park, S., Kim, S., Park, S., Lee, S.H., and Jang, Y. (2018). Macrophage polarization and acceleration of atherosclerotic plaques in a swine model. *PLoS One* 13, e0193005. <https://doi.org/10.1371/journal.pone.0193005>.
99. Liu, Z., Liu, Y., Song, G., Zhang, C., Chen, D., Guo, F., and Li, Y. (2020). Research progress of foam cell in atherogenesis. *Shaanxi Med. J.* 49, 1363–1366+1369.
100. Mosser, D.M. (2003). The many faces of macrophage activation. *J. Leukoc. Biol.* 73, 209–212. <https://doi.org/10.1189/jlb.0602325>.
101. Xiao, Y., Huang, X., Xia, Y., Ding, M., Li, A., Yang, B., and She, Q. (2024). Role of dysregulated macrophage subpopulation ratios and functional changes in the development of coronary atherosclerosis. *J. Gene Med.* 26, e3626. <https://doi.org/10.1002/jgm.3626>.
102. Yuan, J., Lin, F., Chen, L., Chen, W., Pan, X., Bai, Y., Cai, Y., and Lu, H. (2022). Lipoxin A4 regulates M1/M2 macrophage polarization via FPR2-IRF pathway. *Inflammopharmacology* 30, 487–498. <https://doi.org/10.1007/s10787-022-00942-y>.
103. Ma, H., Yang, L., Liu, Y., Yan, R., Wang, R., Zhang, P., Bai, Z., Liu, Y., Ren, Y., Li, Y., et al. (2023). Butyrate suppresses atherosclerotic inflammation by regulating macrophages and polarization via GPR43/HDAC-miRNAs axis in ApoE<sup>-/-</sup> mice. *PLoS One* 18, e0282685. <https://doi.org/10.1371/journal.pone.0282685>.
104. Wang, K., Cao, Q., Yang, Q., Wei, Q., Zhao, J., Wang, Y., Hou, J., and Song, S. (2022). Study on the regulatory effect of leech peptide HE-D on macrophages in atherosclerosis by transcriptome sequencing. *J. Ethnopharmacol.* 294, 115380. <https://doi.org/10.1016/j.jep.2022.115380>.
105. Li, B., Ji, Y., Yi, C., Wang, X., Liu, C., Wang, C., Lu, X., Xu, X., and Wang, X. (2022). Rutin Inhibits Ox-LDL-Mediated Macrophage Inflammation and Foam Cell Formation by Inducing Autophagy and Modulating PI3K/ATK Signaling. *Molecules* 27, 4201. <https://doi.org/10.3390/molecules27134201>.
106. Yao, J., Gao, W., Wang, Y., Wang, L., Diabakte, K., Li, J., Yang, J., Jiang, Y., Liu, Y., Guo, S., et al. (2020). Sonodynamic Therapy Suppresses Neovascularization in Atherosclerotic Plaques via Macrophage Apoptosis-Induced Endothelial Cell Apoptosis. *JACC Basic Transl. Sci.* 5, 53–65. <https://doi.org/10.1016/j.jacbts.2019.10.007>.
107. Jiang, Y., Fan, J., Li, Y., Wu, G., Wang, Y., Yang, J., Wang, M., Cao, Z., Li, Q., Wang, H., et al. (2021). Rapid reduction in plaque inflammation by sonodynamic therapy in patients with symptomatic femoropopliteal peripheral artery disease: A randomized controlled trial. *Int. J. Cardiol.* 325, 132–139. <https://doi.org/10.1016/j.ijcard.2020.09.035>.
108. Cao, Z., Yuan, G., Zeng, L., Bai, L., Liu, X., Wu, M., Sun, R., Chen, Z., Jiang, Y., Gao, Q., et al. (2022). Macrophage-Targeted Sonodynamic/Photothermal Synergistic Therapy for Preventing Atherosclerotic Plaque Progression Using CuS/TiO<sub>2</sub> Heterostructured Nanosheets. *ACS Nano* 16, 10608–10622. <https://doi.org/10.1021/acsnano.2c02177>.
109. Chen, Y., Wang, H., Pan, J., Guo, Y., Hu, Y., Huang, X., Zhou, Y., Deng, Q., and Zhou, Q. (2024). Macrophage-targeted ultrasound nanobubbles for highly efficient sonodynamic therapy of atherosclerotic plaques by modulating M1-to-M2 polarization. *Atherosclerosis* 389, 117423. <https://doi.org/10.1016/j.atherosclerosis.2023.117423>.
110. Viola, A., Munari, F., Sánchez-Rodríguez, R., Scolaro, T., and Castegna, A. (2019). The Metabolic Signature of Macrophage Responses. *Front. Immunol.* 10, 1462. <https://doi.org/10.3389/fimmu.2019.01462>.
111. Ouimet, M., Ediriweera, H.N., Gundra, U.M., Sheedy, F.J., Ramkhalawon, B., Hutchison, S.B., Rinehold, K., van Solingen, C., Fullerton, M.D., Cecchini, K., et al. (2015). MicroRNA-33-dependent regulation of macrophage metabolism directs immune cell polarization in atherosclerosis. *J. Clin. Investig.* 125, 4334–4348. <https://doi.org/10.1172/JCI81676>.
112. Tabas, I., and Bornfeldt, K.E. (2016). Macrophage Phenotype and Function in Different Stages of Atherosclerosis. *Circ. Res.* 118, 653–667. <https://doi.org/10.1161/CIRCRESAHA.115.306256>.
113. Huang, S.C.-C., Everts, B., Ivanova, Y., O'Sullivan, D., Nascimento, M., Smith, A.M., Beatty, W., Love-Gregory, L., Lam, W.Y., O'Neill, C.M., et al. (2014). Cell-intrinsic lysosomal lipolysis is essential for alternative activation of macrophages. *Nat. Immunol.* 15, 846–855. <https://doi.org/10.1038/ni.2956>.
114. Franchi, L., Eigenbrod, T., Muñoz-Planillo, R., and Núñez, G. (2009). The inflammasome: a caspase-1-activation platform that regulates immune responses and disease pathogenesis. *Nat. Immunol.* 10, 241–247. <https://doi.org/10.1038/ni.1703>.
115. Zelcer, N., and Tontonoz, P. (2006). Liver X receptors as integrators of metabolic and inflammatory signaling. *J. Clin. Investig.* 116, 607–614. <https://doi.org/10.1172/JCI27883>.
116. Zhang, X., Qin, Y., Wan, X., Liu, H., Lv, C., Ruan, W., He, L., Lu, L., and Guo, X. (2021). Rosuvastatin exerts anti-atherosclerotic effects by improving macrophage-related foam cell formation and polarization conversion via mediating autophagic activities. *J. Transl. Med.* 19, 62. <https://doi.org/10.1186/s12967-021-02727-3>.
117. Makowski, L., Boord, J.B., Maeda, K., Babaev, V.R., Uysal, K.T., Morgan, M.A., Parker, R.A., Suttles, J., Fazio, S., Hotamisligil, G.S., and Linton, M.F. (2001). Lack of macrophage fatty-acid-binding protein aP2 protects mice deficient in apolipoprotein E against atherosclerosis. *Nat. Med.* 7, 699–705. <https://doi.org/10.1038/89076>.
118. Kanter, J.E., Kramer, F., Barnhart, S., Averill, M.M., Vivekanandan-Giri, A., Vickery, T., Li, L.O., Becker, L., Yuan, W., Chait, A., et al. (2012). Diabetes promotes an inflammatory macrophage phenotype and atherosclerosis through acyl-CoA synthetase 1. *Proc. Natl. Acad. Sci. USA* 109, E715–E724. <https://doi.org/10.1073/pnas.1111600109>.

119. Thorp, E.B. (2019). Mitochondrial Indigestion After Lipid Scavenging. *Circ. Res.* 125, 1103–1105. <https://doi.org/10.1161/CIRCRESAHA.119.316200>.
120. He, Y., and Liu, T. (2023). Oxidized low-density lipoprotein regulates macrophage polarization in atherosclerosis. *Int. Immunopharmacol.* 120, 110338. <https://doi.org/10.1016/j.intimp.2023.110338>.
121. Sun, Y., Ishibashi, M., Seimon, T., Lee, M., Sharma, S.M., Fitzgerald, K.A., Samokhin, A.O., Wang, Y., Sayers, S., Aikawa, M., et al. (2009). Free cholesterol accumulation in macrophage membranes activates Toll-like receptors and p38 mitogen-activated protein kinase and induces cathepsin K. *Circ. Res.* 104, 455–465. <https://doi.org/10.1161/CIRCRESAHA.108.182568>.
122. Canfrán-Duque, A., Rotllán, N., Zhang, X., Andrés-Blasco, I., Thompson, B.M., Sun, J., Price, N.L., Fernández-Fuertes, M., Fowler, J.W., Gómez-Coronado, D., et al. (2023). Macrophage-Derived 25-Hydroxycholesterol Promotes Vascular Inflammation, Atherogenesis, and Lesion Remodeling. *Circulation* 147, 388–408. <https://doi.org/10.1161/CIRCULATION.122.059062>.
123. Wu, H.-M., Ni, X.-X., Xu, Q.-Y., Wang, Q., Li, X.-Y., and Hua, J. (2020). Regulation of lipid-induced macrophage polarization through modulating peroxisome proliferator-activated receptor- $\gamma$  activity affects hepatic lipid metabolism via a Toll-like receptor 4/NF- $\kappa$ B signaling pathway. *J. Gastroenterol. Hepatol.* 35, 1998–2008. <https://doi.org/10.1111/jgh.15025>.
124. Bae, Y.S., Lee, J.H., Choi, S.H., Kim, S., Almazan, F., Witztum, J.L., and Miller, Y.I. (2009). Macrophages generate reactive oxygen species in response to minimally oxidized low-density lipoprotein: toll-like receptor 4- and spleen tyrosine kinase-dependent activation of NADPH oxidase 2. *Circ. Res.* 104, 210–218. <https://doi.org/10.1161/CIRCRESAHA.108.181040>.
125. Jinnouchi, H., Guo, L., Sakamoto, A., Torii, S., Sato, Y., Cornelissen, A., Kuntz, S., Paek, K.H., Fernandez, R., Fuller, D., et al. (2020). Diversity of macrophage phenotypes and responses in atherosclerosis. *Cell. Mol. Life Sci.* 77, 1919–1932. <https://doi.org/10.1007/s00018-019-03371-3>.
126. Wisitpongpan, P., Potup, P., and Usuwanthim, K. (2022). Oleamide-Mediated Polarization of M1 Macrophages and IL-1 $\beta$  Production by Regulating NLRP3-Inflammasome Activation in Primary Human Monocyte-Derived Macrophages. *Front. Immunol.* 13, 856296. <https://doi.org/10.3389/fimmu.2022.856296>.
127. Thomas, C., Leleu, D., and Masson, D. (2022). Cholesterol and HIF-1 $\alpha$ : Dangerous Liaisons in Atherosclerosis. *Front. Immunol.* 13, 868958. <https://doi.org/10.3389/fimmu.2022.868958>.
128. Zhu, X., Yang, L., Han, X., Huang, C., Huang, G., Wei, T., Shu, L., and Xu, J. (2023). Oxidized phospholipids facilitate calcific aortic valve disease by elevating ATF4 through the PERK/eIF2 $\alpha$  axis. *Aging* 15, 6834–6847. <https://doi.org/10.18632/aging.204875>.
129. Zhang, Z., Zhai, L., Lu, J., Sun, S., Wang, D., Zhao, D., Sun, L., Zhao, W., Li, X., and Chen, Y. (2020). Shen-Hong-Tong-Luo Formula Attenuates Macrophage Inflammation and Lipid Accumulation through the Activation of the PPAR- $\gamma$ /LXR- $\alpha$ /ABCA1 Pathway. *Oxid. Med. Cell. Longev.* 2020, 3426925. <https://doi.org/10.1155/2020/3426925>.
130. Hashizume, M., and Mihara, M. (2012). Atherogenic effects of TNF- $\alpha$  and IL-6 via up-regulation of scavenger receptors. *Cytokine* 58, 424–430. <https://doi.org/10.1016/j.cyto.2012.02.010>.
131. Zhang, N., Lei, J., Lei, H., Ruan, X., Liu, Q., Chen, Y., and Huang, W. (2015). MicroRNA-101 overexpression by IL-6 and TNF- $\alpha$  inhibits cholesterol efflux by suppressing ATP-binding cassette transporter A1 expression. *Exp. Cell Res.* 336, 33–42. <https://doi.org/10.1016/j.yexcr.2015.05.023>.
132. Poledne, R., and Králová Lesná, I. (2021). Adipose tissue macrophages and atherogenesis - a synergy with cholesterolaemia. *Physiol. Res.* 70, S535–S549. <https://doi.org/10.33549/physiolres.934745>.
133. Liang, X., Luo, M., Shao, B., Yang, J.-Y., Tong, A., Wang, R.-B., Liu, Y.-T., Jun, R., Liu, T., Yi, T., et al. (2022). Phosphatidylserine released from apoptotic cells in tumor induces M2-like macrophage polarization through the PSR-STAT3-JMJD3 axis. *Cancer Commun.* 42, 205–222. <https://doi.org/10.1002/cac2.12272>.
134. Rinne, P., Guillaumat-Prats, R., Rami, M., Bindila, L., Ring, L., Lyytikäinen, L.-P., Raitoharju, E., Oksala, N., Lehtimäki, T., Weber, C., et al. (2018). Palmitoylethanolamide Promotes a Proresolving Macrophage Phenotype and Attenuates Atherosclerotic Plaque Formation. *Arterioscler. Thromb. Vasc. Biol.* 38, 2562–2575. <https://doi.org/10.1161/ATVBAHA.118.311185>.
135. Park, S.-J., Lee, K.-P., Kang, S., Lee, J., Sato, K., Chung, H.Y., Okajima, F., and Im, D.-S. (2014). Sphingosine 1-phosphate induced anti-atherogenic and atheroprotective M2 macrophage polarization through IL-4. *Cell. Signal.* 26, 2249–2258. <https://doi.org/10.1016/j.cellsig.2014.07.009>.
136. Yang, H., Chen, S., Tang, Y., and Dai, Y. (2011). Interleukin-10 down-regulates oxLDL induced expression of scavenger receptor A and Bak-1 in macrophages derived from THP-1 cells. *Arch. Biochem. Biophys.* 512, 30–37. <https://doi.org/10.1016/j.abb.2011.05.017>.
137. Keul, P., Peters, S., von Wnuck Lipinski, K., Schröder, N.H., Nowak, M.K., Duse, D.A., Polzin, A., Weske, S., Gräler, M.H., and Levkau, B. (2022). Sphingosine-1-Phosphate (S1P) Lyase Inhibition Aggravates Atherosclerosis and Induces Plaque Rupture in ApoE-/-Mice. *Int. J. Mol. Sci.* 23, 9606. <https://doi.org/10.3390/ijms23179606>.
138. Ye, G., Gao, H., Wang, Z., Lin, Y., Liao, X., Zhang, H., Chi, Y., Zhu, H., and Dong, S. (2019). PPAR $\alpha$  and PPAR $\gamma$  activation attenuates total free fatty acid and triglyceride accumulation in macrophages via the inhibition of Fatp1 expression. *Cell Death Dis.* 10, 39. <https://doi.org/10.1038/s41419-018-1135-3>.
139. Zheng, S., Huang, H., Li, Y., Wang, Y., Zheng, Y., Liang, J., Zhang, S., Liu, M., and Fang, Z. (2021). Yin-xing-tong-mai decoction attenuates atherosclerosis via activating PPAR $\gamma$ -LXR $\alpha$ -ABCA1/ABCG1 pathway. *Pharmacol. Res.* 169, 105639. <https://doi.org/10.1016/j.phrs.2021.105639>.
140. Chistiakov, D.A., Melnichenko, A.A., Myasoedova, V.A., Grechko, A.V., and Orekhov, A.N. (2017). Mechanisms of foam cell formation in atherosclerosis. *J. Mol. Med.* 95, 1153–1165. <https://doi.org/10.1007/s00109-017-1575-8>.
141. Cai, Y., Wang, Z., Li, L., He, L., Wu, X., Zhang, M., and Zhu, P. (2022). Neuropeptide Y regulates cholesterol uptake and efflux in macrophages and promotes foam cell formation. *J. Cell Mol. Med.* 26, 5391–5402. <https://doi.org/10.1111/jcmm.17561>.
142. Remmerie, A., and Scott, C.L. (2018). Macrophages and lipid metabolism. *Cell. Immunol.* 330, 27–42. <https://doi.org/10.1016/j.cellimm.2018.01.020>.
143. Shou, X., Wang, Y., Jiang, Q., Chen, J., and Liu, Q. (2023). miR-126 promotes M1 to M2 macrophage phenotype switching via VEGFA and KLF4. *PeerJ* 11, e15180. <https://doi.org/10.7717/peerj.15180>.
144. Song, F., Li, J.-Z., Wu, Y., Wu, W.-Y., Wang, Y., and Li, G. (2021). Ubiquitinated ligation protein NEDD4L participates in MiR-30a-5p attenuated atherosclerosis by regulating macrophage polarization and lipid metabolism. *Mol. Ther. Nucleic Acids* 26, 1303–1317. <https://doi.org/10.1016/j.omtn.2021.10.030>.
145. Hao, T., Fang, W., Xu, D., Chen, Q., Liu, Q., Cui, K., Cao, X., Li, Y., Mai, K., and Ai, Q. (2023). Phosphatidylethanolamine alleviates OX-LDL-induced macrophage inflammation by upregulating autophagy and inhibiting NLRP1 inflammasome activation. *Free Radic. Biol. Med.* 208, 402–417. <https://doi.org/10.1016/j.freeradbiomed.2023.08.031>.
146. Fan, A., Wu, X., Wu, H., Li, L., Huang, R., Zhu, Y., Qiu, Y., Fu, J., Ren, J., and Zhu, C. (2014). Atheroprotective effect of oleoylethanolamide (OEA) targeting oxidized LDL. *PLoS One* 9, e85337. <https://doi.org/10.1371/journal.pone.0085337>.
147. Chen, Z., Zhuo, R., Zhao, Y., Yang, L., Zhou, Y., Cheng, X., Peng, L., Jin, X., and Wang, Y. (2020). Oleoylethanolamide stabilizes atherosclerotic plaque through regulating macrophage polarization via AMPK-PPAR $\alpha$

- pathway. *Biochem. Biophys. Res. Commun.* 524. <https://doi.org/10.1016/j.bbrc.2020.01.103>.
148. Song, T., Wang, P., Li, C., Jia, L., Liang, Q., Cao, Y., Dong, P., Shi, H., and Jiang, M. (2021). Salidroside simultaneously reduces *de novo* lipogenesis and cholesterol biosynthesis to attenuate atherosclerosis in mice. *Biomed. Pharmacother.* 134, 111137. <https://doi.org/10.1016/j.biopha.2020.111137>.
149. Morofuji, Y., Nakagawa, S., Ujifuku, K., Fujimoto, T., Otsuka, K., Niwa, M., and Tsutsumi, K. (2022). Beyond Lipid-Lowering: Effects of Statins on Cardiovascular and Cerebrovascular Diseases and Cancer. *Pharmaceuticals* 15, 151. <https://doi.org/10.3390/ph15020151>.
150. Yu, P., Xiong, T., Tenedero, C.B., Lebeau, P., Ni, R., MacDonald, M.E., Gross, P.L., Austin, R.C., and Trigatti, B.L. (2018). Rosuvastatin Reduces Aortic Sinus and Coronary Artery Atherosclerosis in SR-B1 (Scavenger Receptor Class B Type 1)/ApoE (Apolipoprotein E) Double Knockout Mice Independently of Plasma Cholesterol Lowering. *Arterioscler. Thromb. Vasc. Biol.* 38, 26–39. <https://doi.org/10.1161/ATVBAHA.117.305140>.
151. Wan, J., Yang, J., Lei, W., Xiao, Z., Zhou, P., Zheng, S., and Zhu, P. (2023). Anti-Oxidative, Anti-Apoptotic, and M2 Polarized DSPC Liposome Nanoparticles for Selective Treatment of Atherosclerosis. *Int. J. Nanomed.* 18, 579–594. <https://doi.org/10.2147/IJN.S384675>.
152. Gulshan, K. (2023). Crosstalk Between Cholesterol, ABC Transporters, and PIP2 in Inflammation and Atherosclerosis. *Adv. Exp. Med. Biol.* 1422, 353–377. [https://doi.org/10.1007/978-3-031-21547-6\\_13](https://doi.org/10.1007/978-3-031-21547-6_13).
153. Liang, Q., Zhang, L., and Lv, J. (2024). Research progress on chemical constituents and pharmacological effects of Chuanxiong Rhizoma. *J. Xinxiang Med. Coll.* 41, 275–285.
154. Su, D., Liao, L., Zeng, Q., Liao, Z., Liu, Y., Jin, C., Zhu, G., Chen, C., Yang, M., Ai, Z., and Song, Y. (2022). Study on the new anti-atherosclerosis activity of different Herba patriniae through down-regulating lysophosphatidylcholine of the glycerophospholipid metabolism pathway. *Phytomedicine* 94, 153833. <https://doi.org/10.1016/j.phymed.2021.153833>.
155. Song, N., and Cao, H. (2021). Gypenoside regulates long non-coding RNA TUG1/miR-26a by interfering with mitochondrial apoptosis on hepatic lipid deposition of ApoE<sup>-/-</sup> AS mice. *Nat. Prod. Res. Dev.* 33, 1178. <https://doi.org/10.16333/j.1001-6880.2021.7.013>.
156. Jang, S.-I., Jeong, S.-I., Kim, K.-J., Kim, H.-J., Yu, H.-H., Park, R., Kim, H.-M., and You, Y.-O. (2003). Tanshinone IIA from *Salvia miltiorrhiza* inhibits inducible nitric oxide synthase expression and production of TNF- $\alpha$ , IL-1 $\beta$  and IL-6 in activated RAW 264.7 cells. *Planta Med.* 69, 1057–1059. <https://doi.org/10.1055/s-2003-45157>.
157. Zhang, X., Liu, M.-H., Qiao, L., Zhang, X.-Y., Liu, X.-L., Dong, M., Dai, H.-Y., Ni, M., Luan, X.-R., Guan, J., and Lu, H.-X. (2018). Ginsenoside Rb1 enhances atherosclerotic plaque stability by skewing macrophages to the M2 phenotype. *J. Cell Mol. Med.* 22, 409–416. <https://doi.org/10.1111/jcmm.13329>.
158. Hu, P. (2019). Research Progress on Chemical Constituents and Pharmacological Effects of *Salvia miltiorrhiza*. *Adv. Clin. Med.* 09, 127–132. <https://doi.org/10.12677/ACM.2019.92021>.
159. Li, X.-Y., Wang, Y.-J., Chen, S., Pan, L.-H., Li, Q.-M., Luo, J.-P., and Zha, X.-Q. (2022). Laminaria japonica Polysaccharide Suppresses Atherosclerosis via Regulating Autophagy-Mediated Macrophage Polarization. *J. Agric. Food Chem.* 70, 3633–3643. <https://doi.org/10.1021/acs.jafc.1c07483>.
160. Chen, W., Li, X., Guo, S., Song, N., Wang, J., Jia, L., and Zhu, A. (2019). Tanshinone IIA harmonizes the crosstalk of autophagy and polarization in macrophages via miR-375/KLF4 pathway to attenuate atherosclerosis. *Int. Immunopharmacol.* 70, 486–497. <https://doi.org/10.1016/j.intimp.2019.02.054>.
161. Zhang, Y., Shi, X., Han, J., Peng, W., Fang, Z., Zhou, Y., Xu, X., Lin, J., Xiao, F., Zhao, L., and Lin, Y. (2021). Convallatoxin Promotes M2 Macrophage Polarization to Attenuate Atherosclerosis Through PPAR $\gamma$ -Integrin  $\alpha$ v $\beta$ 5 Signaling Pathway. *Drug Des. Devel. Ther.* 15, 803–812. <https://doi.org/10.2147/DDDT.S288728>.
162. Cai, Y., Wen, J., Ma, S., Mai, Z., Zhan, Q., Wang, Y., Zhang, Y., Chen, H., Li, H., Wu, W., et al. (2021). Huang-Lian-Jie-Du Decoction Attenuates Atherosclerosis and Increases Plaque Stability in High-Fat Diet-Induced ApoE<sup>-/-</sup> Mice by Inhibiting M1 Macrophage Polarization and Promoting M2 Macrophage Polarization. *Front. Physiol.* 12, 666449. <https://doi.org/10.3389/fphys.2021.666449>.
163. Nie, C.-X., Du, X.-K., Yang, L.-N., Li, M.-J., Liu, L., Chen, Y., Yang, Q., Weng, X.-G., Cai, W.-Y., Dong, Y., et al. (2023). Shenlian extract protected ox-LDL-loaded macrophages against ER stress by promoting LAL-LXR $\alpha$  mediated cholesterol flux. *J. Ethnopharmacol.* 317, 116721. <https://doi.org/10.1016/j.jep.2023.116721>.
164. Li, Y., Guo, Y., Chen, Y., Wang, Y., You, Y., Yang, Q., Weng, X., Li, Q., Zhu, X., Zhou, B., et al. (2015). Establishment of an interleukin-1 $\beta$ -induced inflammation-activated endothelial cell-smooth muscle cell-mononuclear cell co-culture model and evaluation of the anti-inflammatory effects of tanshinone IIA on atherosclerosis. *Mol. Med. Rep.* 12, 1665–1676. <https://doi.org/10.3892/mmr.2015.3668>.
165. Xing-Xing, C., Ri-Jin, H., Xin-Ge, W., Cai-Ying, Y., Qing, Y., Ying, C., Qi, L., Xiao-Xin, Z., Lihong, Y., Long, C., and Yu, D. (2024). Mechanistic exploration of the shenlian formula in the suppression of atherosclerosis progression via network pharmacology and *in vivo* experimental validation. *J. Ethnopharmacol.* 333, 118347. <https://doi.org/10.1016/j.jep.2024.118347>.
166. Guo, Y., Liu, X.C., Wang, Y.J., Li, Q., Yang, Q., Weng, X.G., Chen, Y., Cai, W.Y., Kan, X.X., Chen, X., et al. (2016). Effects of Shenlian extract on experimental atherosclerosis in ApoE-deficient mice based on ultrasound biomicroscopy. *BMC Compl. Alternative Med.* 16, 469. <https://doi.org/10.1186/s12906-016-1449-6>.
167. Cai, D., Liu, H., Liu, D., Hao, H., and He, C. (2020). Advances in the Research on Lipid Metabolism and Lipid-lowering Drugs. *Prog. Pharm. Sci.* 44, 379–386.
168. Li, J. (2022). Modern conception of the relationship between dyslipidemia and atherosclerosis. *Chin. Circ. J.* 37, 212–214.
169. Tuñón, J., Badimón, L., Bochaton-Piallat, M.-L., Cariou, B., Daemen, M.J., Egido, J., Evans, P.C., Hoefer, I.E., Ketelhuth, D.F.J., Lutgens, E., et al. (2019). Identifying the anti-inflammatory response to lipid lowering therapy: a position paper from the working group on atherosclerosis and vascular biology of the European Society of Cardiology. *Cardiovasc. Res.* 115, 10–19. <https://doi.org/10.1093/cvr/cvy293>.