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Respiratory uncoupling by increased H⁺ or K⁺ flux is beneficial for heart mitochondrial turnover of reactive oxygen species but not for permeability transition

Saori Morota^{1,2}, Sarah Piel¹ and Magnus J Hansson^{1,3*}

Abstract

Background: Ischemic preconditioning has been proposed to involve changes in mitochondrial H⁺ and K⁺ fluxes, in particular through activation of uncoupling proteins and ATP-sensitive K⁺ channels (MitoK_{ATP}). The objectives of the present study were to explore how increased H⁺ and K⁺ fluxes influence heart mitochondrial physiology with regard to production and scavenging of reactive oxygen species (ROS), volume changes and resistance to calcium-induced mitochondrial permeability transition (mPT).

Results: Isolated rat heart mitochondria were exposed to a wide concentration range of the protonophore CCCP or the potassium ionophore valinomycin to induce increased H⁺ and K⁺ conductance, respectively. Simultaneous monitoring of mitochondrial respiration and calcium retention capacity (CRC) demonstrated that the relative increase in respiration caused by valinomycin or CCCP correlated with a decrease in CRC, and that no level of respiratory uncoupling was associated with enhanced resistance to mPT. Mitochondria suspended in hyperosmolar buffer demonstrated a dose-dependent reduction in CRC with increasing osmolarity. However, mitochondria in hypoosmolar buffer to increase matrix volume did not display increased CRC. ROS generation was reduced by both K⁺- and H⁺-mediated respiratory uncoupling. The ability of heart mitochondria to detoxify H₂O₂ was substantially greater than the production rate. The H₂O₂ detoxification was dependent on respiratory substrates and was dramatically decreased following calcium-induced mPT, but was unaffected by uncoupling via increased K⁺ and H⁺ conductance.

Conclusion: It is concluded that respiratory uncoupling is not directly beneficial to rat heart mitochondrial resistance to calcium overload irrespective of whether H⁺ or K⁺ conductance is increased. The negative effects of respiratory uncoupling thus probably outweigh the reduction in ROS generation and a potential positive effect by increased matrix volume, resulting in a net sensitization of heart mitochondria to mPT activation.

Keywords: Ischemic preconditioning, Mitochondrial permeability transition, Potassium channels, Respiratory uncoupling, Reactive oxygen species

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Background

Hearts exposed to brief periods of ischemia exhibit considerable protection from subsequent ischemia reperfusion-injury, so-called ischemic preconditioning [1,2]. Activation of mitochondrial permeability transition (mPT) has been demonstrated to play a critical role in several animal models of disease including ischemia-reperfusion injury, and inhibition of the mPT regulator cyclophilin D is cardioprotective in animal models and in man [3-6]. The mechanisms involved in preconditioning are not fully established but inhibition of mPT is generally regarded as an important end effect [7,8].

Changes in mitochondrial H^+ or K^+ conductance may critically influence mitochondrial function. Several different pathways involving altered H^+ or K^+ entry into the mitochondrial matrix have been proposed to modify resistance to ischemia-reperfusion injury. Mitochondrial K^+ channel activation, in particular the putative ATP-sensitive K^+ channel (MitoK_{ATP}), has been extensively investigated as a mediator of ischemic preconditioning [9]. Uncoupling proteins - *i.e.* mitochondrial transporter proteins structurally related to UCP1, which uncouples mitochondrial respiration in brown adipose tissue - have also been explored as mediators of ischemic preconditioning [10]. Activation of K^+ channels and H^+ translocators will both uncouple mitochondrial oxidative phosphorylation to some extent. So-called mild uncoupling [11] has been suggested to reduce the sensitivity of mitochondria to undergo calcium-induced mPT both by reducing the $\Delta\Psi_m$ -dependent calcium uptake and by attenuating reactive oxygen species (ROS) production of mitochondria [12,13]. Low concentrations of protonophores can induce a ROS-dependent cardioprotection in ischemia-reperfusion injury [14]. Similarly, activation of mitochondrial K^+ channels has been proposed to reduce the generation of ROS [15,16]. However, results are not consistent as the opposite relation also has been found with an increase in mitochondrial ROS production following K^+ channel activation [17]. A signaling pathway linking MitoK_{ATP} and mPT has also been proposed where an increased generation of H_2O_2 activates an mPT-associated PKC ϵ [18,19]. The increased K^+ conductance following mitochondrial K^+ channel activation may also increase matrix volume [20]. We have previously demonstrated that enhanced matrix volume in brain mitochondria increased their resistance to calcium-induced mPT [21].

The objective of the present study was to explore how increased H^+ and K^+ conductance influence a number of heart mitochondrial physiological parameters relevant for ischemia-reperfusion injury, including respiration, calcium retention capacity (CRC), matrix volume, ROS production and ROS scavenging. Further, we wanted to explore whether increased matrix volume in heart mitochondria exert a beneficial effect on CRC.

In summary, we demonstrate that decreased matrix volume reduces mitochondrial resistance to calcium-induced mPT whereas increased volume did not alter it. H_2O_2 generation of mitochondria was reduced by both K^+ - and H^+ -mediated respiratory uncoupling whereas scavenging of H_2O_2 , which was substantially greater than the production rate, was unaffected by uncoupling unless the inner mitochondrial membrane integrity was disrupted. Increased H^+ or K^+ flux dose-dependently decreased mitochondrial calcium retention capacity and no level of uncoupling demonstrated any positive effect on heart mitochondrial resistance to calcium-induced mPT.

Results

Matrix volume

Activation of mitochondrial K^+ channels with influx of K^+ along its electrochemical gradient may lead to different extents of matrix volume increase depending on the balance of K^+ entry versus activity of the K^+/H^+ exchanger. Here we explored the relation between heart mitochondrial matrix volume and the mitochondrial resistance to calcium-induced mPT by infusing calcium to mitochondria suspended in media with different osmolarities. The extent of mitochondrial calcium accumulation before activation of permeability transition, the calcium retention capacity (CRC), is plotted against the concentration of KCl and the resulting approximate medium osmolarity in Figure 1C. Changes in mitochondrial light absorbance are depicted in Figure 1A, and respiratory function was evaluated in parallel experiments (Figure 1B). CRC was dose-dependently decreased in relation to medium osmolarity in hyperosmolar buffer, compared to the standard buffer containing 125 mM KCl (300 mOsm/l). Mitochondria with reduced matrix volume were thus more sensitive to calcium-induced mPT. The reduced matrix volume tended to increase respiration with a significant increase in state 4 respiration at 275 mM KCl but no significant effect on state 3 respiration. In contrast, increasing mitochondrial matrix volume by reducing the KCl concentration and osmolarity of the media from 125 mM to 75 mM had no apparent effect on CRC but state 3 respiration and the corresponding respiratory control ratio (RCR) were reduced. Exchanging KCl for LiCl did not alter the osmolarity effect; there was no difference in CRC between mitochondria suspended in 50 mM LiCl plus 125 mM KCl buffer and mitochondria suspended in 175 mM KCl buffer. Increasing osmolarity 200 mOsm/l using sucrose instead of KCl likewise decreased CRC (data not shown). In summary, matrix volume correlated to mitochondrial resistance to calcium-induced mPT as reduced volume dose-dependently reduced CRC. However, increased

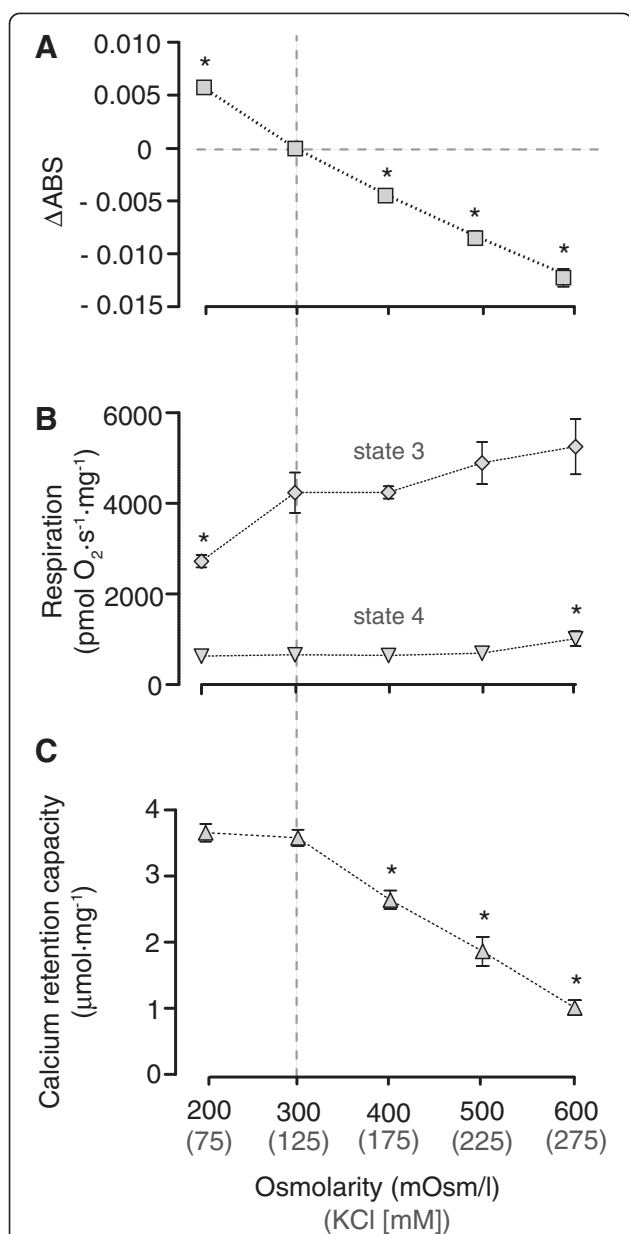


Figure 1 Matrix volume effects on heart mitochondrial respiration and calcium retention capacity (CRC). Heart mitochondrial matrix volume was modulated by changing the osmolarity of the buffer using different concentrations of KCl. The KCl concentrations and the respective approximate medium osmolarities are depicted on the x-axis. The standard incubation medium, 125 mM KCl, is marked by vertical dashed line (A) Changes in matrix volume were monitored by assaying changes in light absorbance at 520 nm (Δ ABS). (B) Respiration rates at state 3 (during ADP phosphorylation) and state 4 (after ADP was converted to ATP) are demonstrated. (C) Calculations of mitochondrial CRC at the different buffer osmolarities. Values are means \pm S.E.M. * indicates $p < 0.05$ compared to control (125 mM KCl).

matrix volume failed to increase heart mitochondrial resistance to mPT and negatively affected respiratory function.

Uncoupling and resistance to mPT

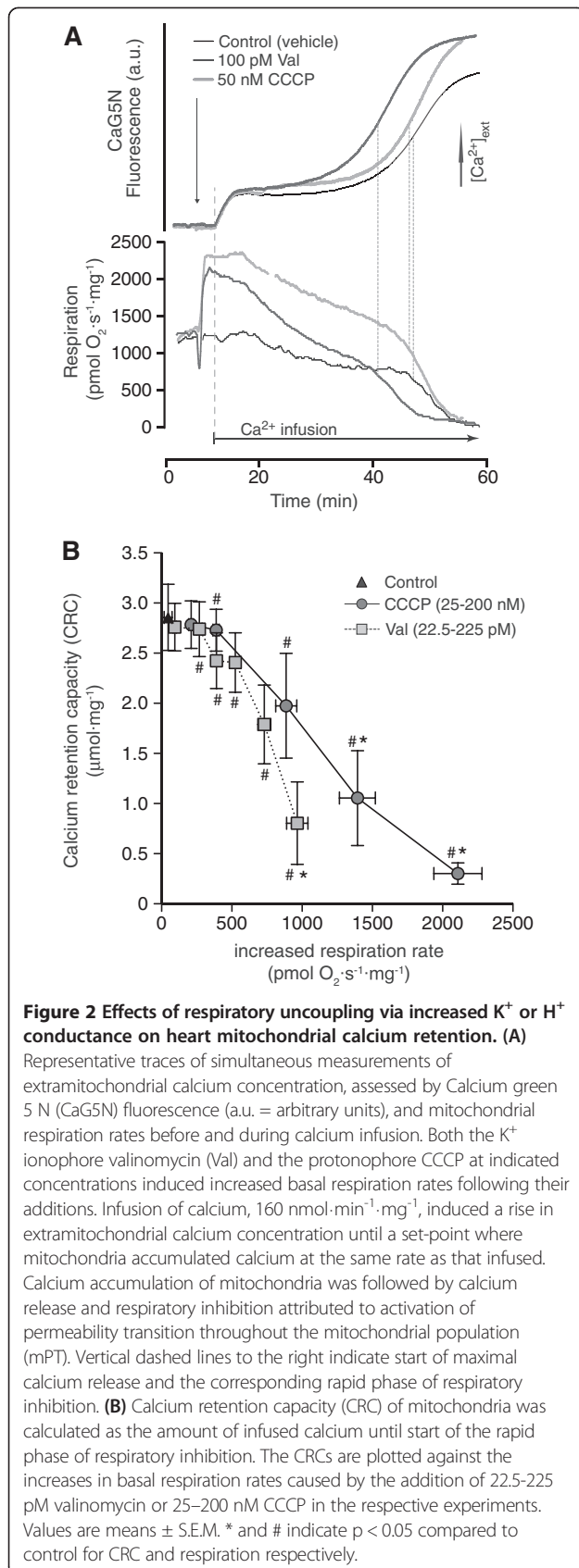
Increased H^+ or K^+ conductance will lead to an augmented uncoupling of mitochondrial respiration since the proton motive force is utilized for ion exchange rather than ATP synthesis. In order to evaluate if uncoupling can reduce the sensitivity of heart mitochondria to undergo calcium-induced mPT we performed a detailed simultaneous analysis of mitochondrial CRC and respiration (Figure 2). Mitochondria were exposed to wide concentration ranges of the K^+ carrier valinomycin, (22.5 to 225 μ M) and the protonophore CCCP (25 to 200 nM). The uncoupling effects of valinomycin or CCCP were calculated by measuring the added rates of respiration following their addition. Representative traces of a control and two of the lower concentrations of the respective compounds are demonstrated in Figure 2A. When mPT is activated throughout the mitochondrial population, respiration is decreased at least partly due to a loss of NADH/NAD⁺ from the mitochondrial matrix [21], and the decrease in respiration occurs simultaneously to the rapid release of calcium from mitochondria following the period of calcium uptake (Figure 2A). Plotting mitochondrial CRC against the increased respiration rate demonstrated a dose-dependent reduction in CRC for both the K^+ carrier and the protonophore (Figure 2B). Linear regression analysis of the individual CCCP and valinomycin data points demonstrated a significant correlation between increases in basal respiratory rates and decreased CRC. The r^2 values were 0.76 ($p < 0.0001$) and 0.45 ($p = 0.0001$) for CCCP and valinomycin, respectively. At higher concentrations of the two compounds, valinomycin caused a more pronounced reduction of CRC compared to CCCP at comparable degrees of uncoupling. In summary, no level of respiratory uncoupling demonstrated any positive effect on heart mitochondrial resistance to calcium-induced mPT.

Effects of diazoxide and cyclosporin A

The putative activator of MitoK_{ATP}, diazoxide, was evaluated for effects on respiration and CRC (in separate experiments) at 30, 100 and 200 μ M. State 4 respiration was slightly but significantly elevated at 100 and 200 μ M diazoxide (862.2 ± 44 and 937.1 ± 33 , respectively, compared to 659.2 ± 30 pmol O₂·s⁻¹·mg⁻¹ for control). The corresponding respiratory control ratios (state3/state4) were reduced but none of the tested concentrations of diazoxide had any significant effect on CRC (data not shown). The cyclophilin D inhibitor CsA (1 μ M), in contrast, significantly increased CRC from 3.5 ± 0.4 to 4.7 ± 0.2 μ mol·mg⁻¹.

ROS generation

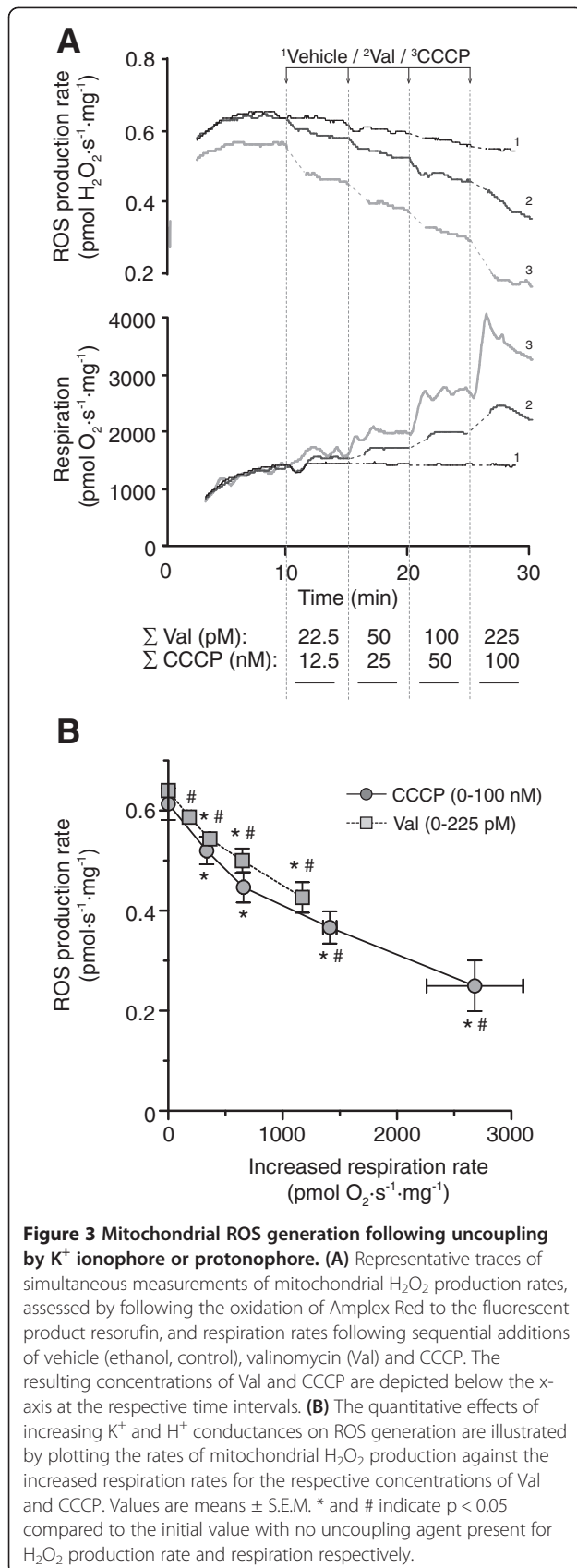
To evaluate whether there is a qualitative or quantitative difference between increased H^+ and K^+ conductance on ROS generation, heart mitochondria were exposed to



repetitive additions of valinomycin (22.5 to 225 pM) and CCCP (25 to 100 nM). The H₂O₂ production rates were correlated to simultaneous measurements of increased respiration rates. The K⁺ carrier and the protonophore dose-dependently reduced mitochondrial ROS generation corresponding to extent of respiratory uncoupling (Figure 3). Linear regression analysis of the individual CCCP and valinomycin data points demonstrated a significant correlation between increases in basal respiratory rates and decreased H₂O₂ production. The r² values were 0.65 (p = 0.0003) and 0.85 (p < 0.0001) for CCCP and valinomycin, respectively. There was no significant difference between the slopes for CCCP or valinomycin. ROS generation was also evaluated in mitochondria exposed to a continuous calcium infusion in presence of 50 pM valinomycin or 100 nM CCCP which both reduced H₂O₂ detection from 4.3 ± 0.2 to 3.77 ± 0.1 and 3.3 ± 0.1 pmol·s⁻¹·mg⁻¹, respectively. In summary, there was no qualitative or quantitative difference on the effect on heart mitochondrial H₂O₂ generation between increased H⁺ and K⁺ conductance.

ROS scavenging

A reduced capacity of mitochondria to detoxify ROS could potentially be more important for organ failure than gross mitochondrial ROS generation. The steady-state production of ROS is generally assessed by probes with higher affinity for H₂O₂ than the endogenous mitochondrial scavenging pathways, and the balance between mitochondrial generation and detoxification of H₂O₂ is therefore not reflected by such an assay [22,23]. Further, there is no accumulation of H₂O₂ in suspensions of isolated intact mitochondria in contrast to mitochondria following mPT [22-24]. Therefore, to calculate the rate of H₂O₂ removal, the heart mitochondrial suspensions were exposed to exogenous H₂O₂ and the ROS detection system was added after specific time points to determine residual H₂O₂ concentrations (Figure 4). Following a large bolus load of calcium to induce extensive mPT (Figure 4B) or when the experiments were performed without respiratory substrates (data not shown) the H₂O₂ removal was dramatically diminished, and the exogenously added H₂O₂ remained in the mitochondrial suspension to a large extent. However, valinomycin and CCCP had no effect on H₂O₂ removal (Figure 4C) when tested at concentrations that displayed prominent effects on respiration and CRC. When compared in the same set of experiments under standard conditions, the mitochondrial H₂O₂ scavenging rate was more than 10-fold greater than the detected production rate of H₂O₂ (Figure 4D). Thus, the ROS scavenging of heart mitochondria was dependent on active substrate oxidation and required an intact inner membrane. Whereas uncoupling by increased H⁺ and K⁺ conductance reduced ROS production, it did not affect ROS scavenging.

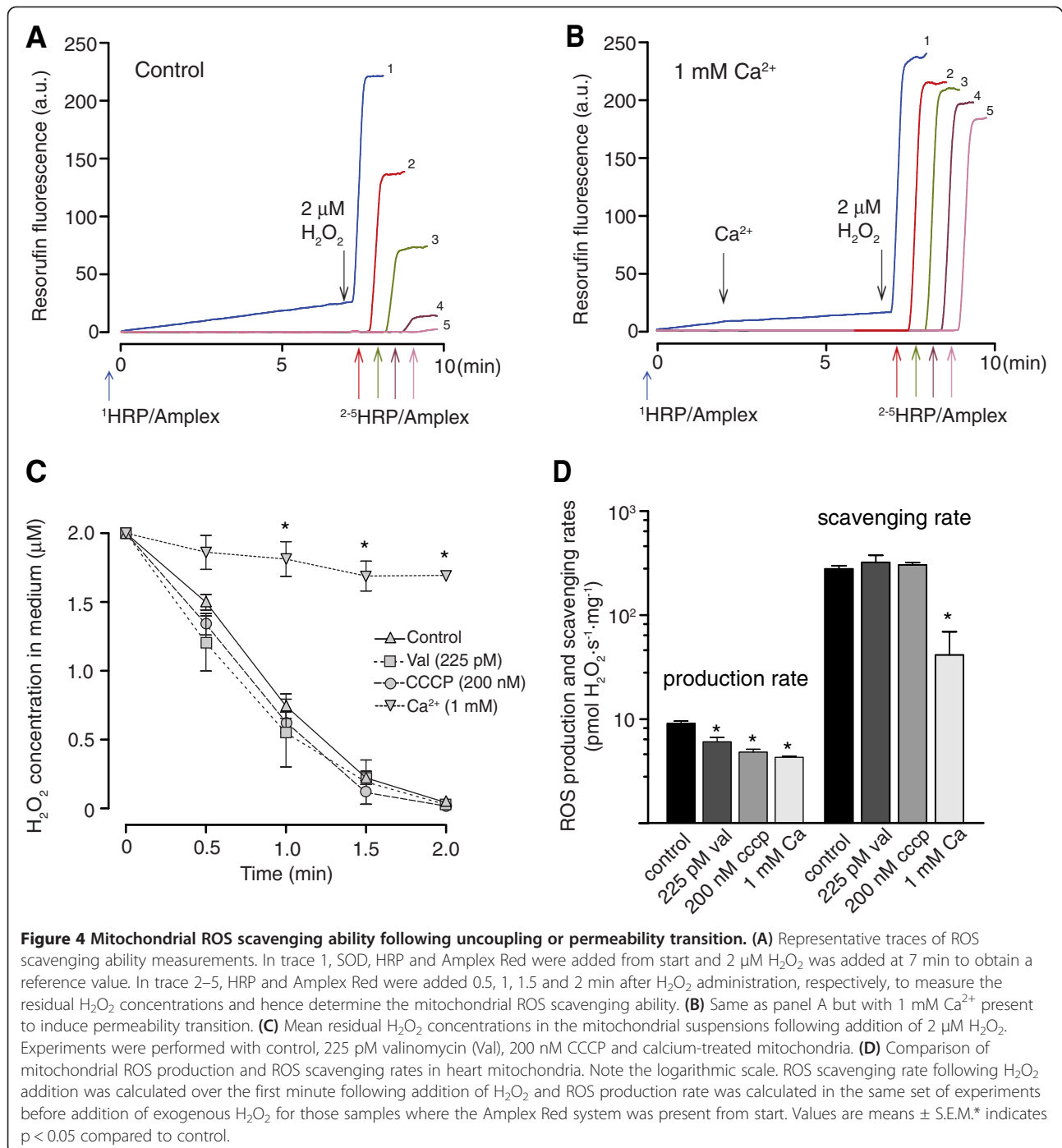


Discussion

Several hypotheses on the mechanisms of ischemic preconditioning involve activation of channels or transporter proteins affecting the ion flux, most notably for K⁺ and H⁺, across the inner mitochondrial membrane. Increasing mitochondrial K⁺ or H⁺ conductance will have several physiological consequences. The aim of the present work was to explore how some of these alterations relate to commonly proposed mechanisms for ischemia-reperfusion injury in heart mitochondria.

An extensive uncoupling will cause a dramatic effect on normal mitochondrial function, but so-called mild uncoupling has been suggested to reduce the sensitivity of mitochondria to undergo calcium-induced mPT by limiting the calcium uptake and ROS production of mitochondria [12,25]. The CRC assay is a sensitive and quantitative method for evaluating mitochondrial susceptibility to mPT induction [26], and by simultaneous monitoring of respiration the specificity is increased [21]. Here, we demonstrate that heart mitochondrial CRC is dose-dependently reduced by uncoupling irrespective of whether H⁺ or K⁺ conductance is increased, and that the reduced CRC is caused by mPT (as demonstrated by the respiratory decrease). There was no tendency of any beneficial effect at lower concentrations of the K⁺ carrier valinomycin or the protonophore CCCP. At the highest tested concentration of CCCP, 200 nM, the respiratory increase was less than a state 2 to state 3 transition but the ability to retain Ca²⁺ was virtually abolished and mitochondria underwent permeability transition at a very low calcium load. Ca²⁺ is the principal trigger of mPT and a reduction in matrix Ca²⁺ would arguably reduce mPT activation. However, it is the free Ca²⁺ ions that trigger mPT and most calcium in mitochondria is retained as inactive calcium-phosphate complexes [26]. The formation of calcium-phosphates is highly dependent on an alkaline matrix environment. A small decrease in ΔpH, as occurs with uncoupling following H⁺ entry, will increase the concentration of free Ca²⁺ even if the electrophoretic driving force for Ca²⁺ uptake is reduced. Uncoupling may also directly increase the propensity of the mPT pore to open due to a more oxidized redox state and a reduced mitochondrial membrane potential (ΔΨ_m) [27,28].

Increasing K⁺ flux into the mitochondria may affect several factors important for calcium handling and mPT regulation, including ΔΨ_m, ΔpH, ROS generation and matrix volume, which are somewhat different to increasing H⁺ flux. For example, increasing K⁺ or H⁺ conductance may be expected to have opposite effects on ΔpH due to activation of the K⁺/H⁺ exchanger by the former. Whether an alteration in any of these parameters is beneficial or not to the resistance of mitochondria to the dramatic consequences of calcium overload depend both



on their direct and potential indirect effects on the mPT pore components, as pointed out above for the effect of ΔpH on free Ca^{2+} . In a previous study, we found that low increases in K^+ but not H^+ conductance was beneficial to brain mitochondrial resistance to calcium-induced permeability transition, and that the beneficial effect probably was mediated by an increase in matrix volume [21]. In the present study, there was no qualitative difference between

increasing H^+ or K^+ conductance on CRC. The negative effects of uncoupling thus predominate over other potential direct beneficial effects of increasing K^+ conductance in rat heart mitochondria.

Changes in mitochondrial matrix volume influence respiratory function [29–31] and we have previously demonstrated that increased matrix volume in brain mitochondria either caused by an elevated influx of K^+ or by suspending

mitochondria in hypoosmolar KCl medium increases CRC [21]. In the present study we explored respiratory changes and CRC in heart mitochondria suspended in hypo- or hyperosmolar KCl media. Increasing medium osmolarity by KCl (or LiCl) dose-dependently decreased CRC indicating a positive correlation between matrix volume and CRC. A possible confounder to increasing osmolarity with KCl or LiCl is the simultaneous change in ionic strength which may affect *e.g.* cytochrome *c* charge interactions [32]. However, increasing medium osmolarity with sucrose likewise decreased CRC. In contrast to the effects of increasing medium osmolarity, decreasing the osmolarity further than the standard conditions did not cause the expected increase in CRC. A plausible explanation may be the reduced respiratory capacity of heart mitochondria in the hypoosmolar medium. An increase in matrix volume has been shown to cause release of cytochrome *c* thus reducing respiratory capacity in isolated liver mitochondria [33]. The same authors also found a reduced calcium uptake in mitochondria following exposure to a hypoosmotic shock. In a limited set of experiments in heart mitochondria (data not shown) we found that there was a tendency for exogenously added cytochrome *c* to increase respiration more in hypoosmolar media compared to hyperosmolar conditions, but these findings were not explored further. It is thus plausible that any beneficial effect on heart mitochondrial CRC by an enlarged matrix volume following increased K^+ flux is counteracted by a respiratory inhibition and a direct negative effect of uncoupling. Even though the majority of studied characteristics described herein were similar to previous findings in brain mitochondria, there seems to be a distinct organ specific difference in relation to the response to increased K^+ conductance and matrix volume.

ROS production of mitochondria has since long been recognized to increase with elevated proton-motive force [34,35]. This is particularly evident when isolated mitochondria are supplemented with solely succinate as respiratory substrate to support so-called reverse electron transport through respiratory complex I [36]. Ischemic preconditioning reduces oxidative damage in mitochondria, and a reduction of oxidative stress may mediate a reduced propensity for mPT activation [37]. Previous studies have demonstrated both increased and decreased ROS generation following augmented K^+ flux in mitochondria [15-19]. In the present study, both increased K^+ and H^+ flux dose-dependently reduced the production of H_2O_2 in heart mitochondria using complex I substrates. There was no apparent qualitative or quantitative difference between increased K^+ and H^+ flux; rather, the level of ROS reduction was in proportion to the extent of uncoupling. However, the reduced ROS generation did not translate into an increased resistance to mPT induction.

The capacity of mitochondria to detoxify ROS is less explored but likely equally or more important than mitochondrial ROS production in oxidative stress [23] and there is no clear evidence whether mitochondria function as net source or sink of ROS [38]. Heart mitochondria are equipped with effective antioxidant systems for scavenging of ROS and dysfunction of these are involved in cardiovascular disease [39]. Malfunction of mitochondrial thioredoxin reductase causes dilated cardiomyopathy in man [40] and aggravates ischemia-reperfusion injury in mice [41], while overexpression of glutaredoxin-2 reduces ischemia-reperfusion injury in mice [42]. The mitochondrial antioxidative systems are dependent on intact respiratory function. Reduced glutathione and NADPH provide antioxidant defences against H_2O_2 mediated damage through the action of *e.g.* the glutathione peroxidase family, peroxiredoxins and glutaredoxins. The reduction of GSH and thioredoxin reductase is dependent on NADPH generation from *e.g.* the proton motive force-driven transhydrogenase [43]. Although uncoupling of mitochondrial respiration reduces ROS production, any interference with ROS scavenging may result in an overall negative effect on mitochondrial ROS handling. Therefore, we investigated the heart mitochondrial capacity of H_2O_2 removal under conditions of uncoupling and calcium overload. The mitochondrial H_2O_2 scavenging rate was more than 10-fold greater than the detected production rate of H_2O_2 under standard conditions. Whereas uncoupling decreased H_2O_2 production it did not affect H_2O_2 scavenging. In contrast, calcium overload leading to mPT dramatically reduced H_2O_2 scavenging in line with previous reports [23,24]. Any effects of calcium unrelated to mPT activation were not explored in the present study.

We did not detect any specific effect of the MitoK_{ATP} opener diazoxide. The compound uncoupled respiration at high concentrations, which also has been demonstrated previously [20]. The present experiments were not primarily designed to explore the putative MitoK_{ATP}. Such experiments are usually performed within the first few minutes upon incubation in experimental medium and the present experiments examining general effects of increased H^+ and K^+ conductance required a longer time frame. It has been noted that the MitoK_{ATP} is sensitive to rapid inactivation [44] and there is also a current debate over its existence [9].

Conclusions

The present findings suggest that there are no direct beneficial effects of increased K^+ or H^+ flux on heart mitochondrial resistance to mPT. Even though there was a decrease in the generation of ROS and a positive correlation between matrix volume and CRC in hyperosmotic media, the negative effects related to respiratory uncoupling predominated with a net sensitization of heart

mitochondria to mPT activation. Importantly though, uncoupling did not hamper mitochondrial ROS scavenging capacity. It is concluded that respiratory uncoupling is not beneficial to rat heart mitochondrial resistance to calcium overload regardless of whether H⁺ or K⁺ conductance is increased. The present study is limited by its specific examination of mitochondria in their isolated state in contrast to their integrated function *in vivo*. Nevertheless, the results imply that the direct physiological effects of increased H⁺ or K⁺ flux on ROS generation, matrix volume and calcium handling in heart mitochondria are not in themselves sufficient to mediate ischemic preconditioning in the heart.

Methods

Isolation of heart mitochondria

Adult male Wistar rats (Harlan Scandinavia ApS, Allerød, Denmark, 350–450, 500–600 g) were allowed access to water and food *ad libitum* prior to use. All animal procedures were approved by the Malmö/Lund Ethical Committee for Animal Research (M44-07, M99-10). Following a brief exposure to halothane in order to minimize stress, animals were decapitated [45]. Hearts were rapidly transferred into ice-cold solution and mitochondria were isolated using trypsinization, homogenization and differential centrifugation at 4°C as previously described [46]. Protein content was measured using Bradford analysis.

Calcium retention capacity (CRC), mitochondrial respiration and matrix volume changes

A luminescence spectrometer LS-50B (Perkin-Elmer, Emeryville, CA) with a temperature controlled cuvette holder or an Oxygraph-2 k with a Titration-Injection micro Pump TIP-2 k (Oroboros instruments, Innsbruck Austria) was used to measure mitochondrial CRC. Experiments were performed at 37°C. Mitochondria, 75 µg/ml, were suspended in buffer containing 125 mM KCl, 20 mM Trizma base, 2 mM Pi (K₂PO₄), 1 mM MgCl₂, 1 µM EGTA, 200 µM ATP, 10 µM BSA and 5 mM of the NADH-linked respiratory substrates malate and glutamate, pH 7.1. For experiments assessing matrix volume effects, the osmolarity of the buffer was altered by using different KCl concentrations, 75–275 mM, corresponding to approximately 200–600 mOsm/l. In experiments evaluating modulation of the putative MitoK_{ATP}, 30, 100 or 200 µM diazoxide was added before addition of mitochondria. For evaluation of cyclophilin D modulation of mPT, 1 µM CsA was used. The mitochondrial suspensions were infused with 160 nmol CaCl₂·min⁻¹·mg⁻¹ in presence of 1 µg/ml oligomycin and 50 µM ADP. In luminescence spectrometer experiments, mitochondrial Ca²⁺ uptake/release was monitored by the excitation ratio of the extramitochondrial calcium-sensitive fluorescent probe Fura 6 F (250 nM, Ex. 340/380 nm, Em. 509 nm). In

experiments monitoring oxygen consumption, mitochondrial Ca²⁺ uptake and release was monitored by respiration changes during Ca²⁺ infusion [21]. In one set of experiments, respiration and extramitochondrial calcium were monitored simultaneously by following the fluorescence of Calcium Green 5 N (100 nM) in the oxygraph using a O2k-Fluorescence LED2-Module (Oroboros instruments, Innsbruck Austria).

CRC was calculated as the amount of infused calcium from the start of mitochondrial calcium uptake until start of maximal calcium release or start of the rapid phase of respiratory inhibition. The effects of changing medium osmolarities on respiratory states were investigated under conditions described above except that no adenine nucleotides were present from start. State 3 respiration was induced by adding 250 µM ADP and state 4 respiration was measured after all ADP had been converted to ATP. Matrix volume changes were evaluated by measuring light absorbance at 520 nm in a Hitachi U-2800 Spectrophotometer (Tokyo, Japan).

Mitochondrial ROS production and scavenging

Mitochondrial H₂O₂ generation was detected by the oxidation of 1 µM Amplex Red to the fluorescent product resorufin in presence of HRP and SOD (0.5 and 20 U/ml, respectively) [24]. Experiments were performed under the same conditions as described above. Simultaneous measurement of respiration and H₂O₂ generation was performed in the oxygraph [47]. The oxidation of Amplex Red was calibrated with repetitive additions of 0.1 µM H₂O₂, and the calibrated H₂O₂ concentrations and production rates were monitored. Mitochondrial ROS scavenging experiments were performed in the luminescence spectrometer. In contrast to following the continuous Amplex Red oxidation, HRP and Amplex Red were added at specific time points (0.5 – 2.0 min) following addition of 2 µM H₂O₂ to the mitochondrial suspensions in order to detect the remaining H₂O₂ concentration and to establish the rate of mitochondrial H₂O₂ detoxification.

Statistical analyses

All average results are presented as mean ± S.E.M. and were evaluated using one-way ANOVA or repeated measures ANOVA for comparisons within experimental runs followed by Dunnett's multiple comparison post hoc test using GraphPad Prism v5.0 software. All experiments were replicated at least 3 times in separate mitochondrial preparations. Differences were considered significant where *P* < 0.05.

Competing interests

MJH has equity interest and receives consultancy fees and SP is a part-time employee of NeuroVive Pharmaceutical AB, which is active in the field of mitochondrial medicine including development of cyclophilin D inhibitors.

Authors' contributions

SM and SP performed experiments. SM and MJH planned and designed the experiments, analysed the results and wrote the manuscript. All authors read and approved the final manuscript.

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References

- Murry CE, Jennings RB, Reimer KA: **Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium.** *Circulation* 1986, **74**:1124–1136.
- Yellon DM, Downey JM: **Preconditioning the myocardium: from cellular physiology to clinical cardiology.** *Physiol Rev* 2003, **83**:1113–1151.
- Griffiths EJ, Halestrap AP: **Protection by Cyclosporin A of ischemia/reperfusion-induced damage in isolated rat hearts.** *J Mol Cell Cardiol* 1993, **25**:1461–1469.
- Argaud L, Gateau-Roesch O, Muntean D, Chalabreysse L, Loufouat J, Robert D, Ovize M: **Specific inhibition of the mitochondrial permeability transition prevents lethal reperfusion injury.** *J Mol Cell Cardiol* 2005, **38**:367–374.
- Shanmuganathan S, Hausenloy DJ, Duchon MR, Yellon DM: **Mitochondrial permeability transition pore as a target for cardioprotection in the human heart.** *Am J Physiol Heart Circ Physiol* 2005, **289**:H237–H242.
- Piot C, Croisille P, Staat P, Thibault H, Rioufol G, Mewton N, Elbelghiti R, Cung TT, Bonnefoy E, Angoulvant D, et al: **Effect of cyclosporine on reperfusion injury in acute myocardial infarction.** *N Engl J Med* 2008, **359**:473–481.
- Lim SY, Davidson SM, Hausenloy DJ, Yellon DM: **Preconditioning and postconditioning: the essential role of the mitochondrial permeability transition pore.** *Cardiovasc Res* 2007, **75**:530–535.
- Halestrap AP, Clarke SJ, Khalilulin I: **The role of mitochondria in protection of the heart by preconditioning.** *Biochim Biophys Acta* 2007, **1767**:1007–1031.
- Garlid KD, Halestrap AP: **The mitochondrial K(ATP) channel-Fact or fiction?** *J Mol Cell Cardiol* 2012, **52**:578–583.
- McLeod CJ, Aziz A, Hoyt RF Jr, McCoy JP Jr, Sack MN: **Uncoupling proteins 2 and 3 function in concert to augment tolerance to cardiac ischemia.** *J Biol Chem* 2005, **280**:33470–33476.
- Skulachev VP: **Role of uncoupled and non-coupled oxidations in maintenance of safely low levels of oxygen and its one-electron reductants.** *Q Rev Biophys* 1996, **29**:169–202.
- Baines CP: **The cardiac mitochondrion: nexus of stress.** *Annu Rev Physiol* 2010, **72**:61–80.
- Sedlic F, Sepac A, Pravdic D, Camara AK, Bienengraeber M, Brzezinska AK, Wakatsuki T, Bosnjak ZJ: **Mitochondrial depolarization underlies delay in permeability transition by preconditioning with isoflurane: roles of ROS and Ca²⁺.** *Am J Physiol Cell Physiol* 2010, **299**:C506–515.
- Brennan JP, Southworth R, Medina RA, Davidson SM, Duchon MR, Shattock MJ: **Mitochondrial uncoupling, with low concentration FCCP, induces ROS-dependent cardioprotection independent of KATP channel activation.** *Cardiovasc Res* 2006, **72**:313–321.
- Ozcan C, Bienengraeber M, Dzeja PP, Terzic A: **Potassium channel openers protect cardiac mitochondria by attenuating oxidant stress at reoxygenation.** *Am J Physiol Heart Circ Physiol* 2002, **282**:H531–539.
- Vanden Hoek T, Becker LB, Shao ZH, Li CQ, Schumacker PT: **Preconditioning in cardiomyocytes protects by attenuating oxidant stress at reperfusion.** *Circ Res* 2000, **86**:541–548.
- Andrukhiv A, Costa AD, West IC, Garlid KD: **Opening mitoKATP increases superoxide generation from complex I of the electron transport chain.** *Am J Physiol Heart Circ Physiol* 2006, **291**:H2067–2074.
- Costa AD, Quinlan CL, Andrukhiv A, West IC, Jaburek M, Garlid KD: **The direct physiological effects of mitoK(ATP) opening on heart mitochondria.** *Am J Physiol Heart Circ Physiol* 2006, **290**:H406–415.
- Costa AD, Jakob R, Costa CL, Andrukhiv K, West IC, Garlid KD: **The mechanism by which the mitochondrial ATP-sensitive K⁺ channel opening and H₂O₂ inhibit the mitochondrial permeability transition.** *J Biol Chem* 2006, **281**:20801–20808.
- Kowaltowski AJ, Seetharaman S, Pauczek P, Garlid KD: **Bioenergetic consequences of opening the ATP-sensitive K(+) channel of heart mitochondria.** *Am J Physiol Heart Circ Physiol* 2001, **280**:H649–657.
- Hansson MJ, Morota S, Teilmum M, Mattiasson G, Uchino H, Elmer E: **Increased potassium conductance of brain mitochondria induces resistance to permeability transition by enhancing matrix volume.** *J Biol Chem* 2010, **285**:741–750.
- Nohl H, Kozlov AV, Gille L, Staniek K: **Cell respiration and formation of reactive oxygen species: facts and artefacts.** *Biochem Soc Trans* 2003, **31**:1308–1311.
- Andreyev AY, Kushnareva YE, Starkov AA: **Mitochondrial metabolism of reactive oxygen species.** *Biochemistry (Mosc)* 2005, **70**:200–214.
- Hansson MJ, Mansson R, Morota S, Uchino H, Kallur T, Sumi T, Ishii N, Shimazu M, Keep MF, Jegorov A, Elmer E: **Calcium-induced generation of reactive oxygen species in brain mitochondria is mediated by permeability transition.** *Free Radic Biol Med* 2008, **45**:284–294.
- Holmuhamedov EL, Wang L, Terzic A: **ATP-sensitive K⁺ channel openers prevent Ca²⁺ overload in rat cardiac mitochondria.** *J Physiol* 1999, **519**(Pt 2):347–360.
- Chalmers S, Nicholls DG: **The relationship between free and total calcium concentrations in the matrix of liver and brain mitochondria.** *J Biol Chem* 2003, **278**:19062–19070.
- Hunter DR, Haworth RA: **The Ca²⁺-induced membrane transition in mitochondria. I. The protective mechanisms.** *Arch Biochem Biophys* 1979, **195**:453–459.
- Bernardi P: **Modulation of the mitochondrial cyclosporin A-sensitive permeability transition pore by the proton electrochemical gradient. Evidence that the pore can be opened by membrane depolarization.** *J Biol Chem* 1992, **267**:8834–8839.
- Halestrap AP: **The regulation of the matrix volume of mammalian mitochondria in vivo and in vitro and its role in the control of mitochondrial metabolism.** *Biochim Biophys Acta* 1989, **973**:355–382.
- Devin A, Espie P, Guerin B, Rigoulet M: **Energetics of swelling in isolated hepatocytes: a comprehensive study.** *Mol Cell Biochem* 1998, **184**:107–121.
- Nicholls DG, Lindberg O: **Inhibited respiration and ATPase activity of rat liver mitochondria under conditions of matrix condensation.** *FEBS Lett* 1972, **25**:61–64.
- Kossekov G, Atanasov B, Bolli R, Azzi A: **Ionic-strength-dependence of the oxidation of native and pyridoxal 5'-phosphate-modified cytochromes c by cytochrome c oxidase.** *Biochem J* 1989, **262**:591–596.
- Gogvadze V, Robertson JD, Enoksson M, Zhivotovskiy B, Orrenius S: **Mitochondrial cytochrome c release may occur by volume-dependent mechanisms not involving permeability transition.** *Biochem J* 2004, **378**:213–217.
- Boveris A, Chance B: **The mitochondrial generation of hydrogen peroxide. General properties and effect of hyperbaric oxygen.** *Biochem J* 1973, **134**:707–716.
- Korshunov SS, Skulachev VP, Starkov AA: **High protonic potential actuates a mechanism of production of reactive oxygen species in mitochondria.** *FEBS Lett* 1997, **416**:15–18.
- Toime LJ, Brand MD: **Uncoupling protein-3 lowers reactive oxygen species production in isolated mitochondria.** *Free Radic Biol Med* 2010, **49**:606–611.
- Clarke SJ, Khalilulin I, Das M, Parker JE, Heesom KJ, Halestrap AP: **Inhibition of mitochondrial permeability transition pore opening by ischemic preconditioning is probably mediated by reduction of oxidative stress rather than mitochondrial protein phosphorylation.** *Circ Res* 2008, **102**:1082–1090.
- Brown GC, Borutaite V: **There is no evidence that mitochondria are the main source of reactive oxygen species in mammalian cells.** *Mitochondrion* 2012, **12**:1–4.
- Berndt C, Lillig CH, Holmgren A: **Thiol-based mechanisms of the thioredoxin and glutaredoxin systems: implications for diseases in the cardiovascular system.** *Am J Physiol Heart Circ Physiol* 2007, **292**:H1227–1236.

40. Sibbing D, Pfeufer A, Perisic T, Mannes AM, Fritz-Wolf K, Unwin S, Sinner MF, Gieger C, Gloeckner CJ, Wichmann HE, *et al*: **Mutations in the mitochondrial thioredoxin reductase gene TXNRD2 cause dilated cardiomyopathy.** *Eur Heart J* 2011, **32**:1121–1133.
41. Horstkotte J, Perisic T, Schneider M, Lange P, Schroeder M, Kiermayer C, Hinkel R, Ziegler T, Mandal PK, David R, *et al*: **Mitochondrial thioredoxin reductase is essential for early postischemic myocardial protection.** *Circulation* 2011, **124**:2892–2902.
42. Nagy N, Malik G, Tosaki A, Ho YS, Maulik N, Das DK: **Overexpression of glutaredoxin-2 reduces myocardial cell death by preventing both apoptosis and necrosis.** *J Mol Cell Cardiol* 2008, **44**:252–260.
43. Nicholls DG, Budd SL: **Mitochondria and neuronal survival.** *Physiol Rev* 2000, **80**:315–360.
44. Wojtovich AP, Brookes PS: **The endogenous mitochondrial complex II inhibitor malonate regulates mitochondrial ATP-sensitive potassium channels: implications for ischemic preconditioning.** *Biochim Biophys Acta* 2008, **1777**:882–889.
45. Morota S, Hansson MJ, Ishii N, Kudo Y, Elmer E, Uchino H: **Spinal cord mitochondria display lower calcium retention capacity compared with brain mitochondria without inherent differences in sensitivity to cyclophilin D inhibition.** *J Neurochem* 2007, **103**:2066–2076.
46. Morota S, Manolopoulos T, Eyjolfsson A, Kimblad PO, Wierup P, Metzsch C, Blomquist S, Hansson MJ: **Functional and pharmacological characteristics of permeability transition in isolated human heart mitochondria.** *PLoS One* 2013, **8**:e67747.
47. Hickey AJ, Renshaw GM, Speers-Roesch B, Richards JG, Wang Y, Farrell AP, Brauner CJ: **A radical approach to beating hypoxia: depressed free radical release from heart fibres of the hypoxia-tolerant epaulette shark (*Hemiscyllium ocellatum*).** *J Comp Physiol B* 2012, **182**:91–100.

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