



# Article In Utero Exposure to Caffeine and Acetaminophen, the Gut Microbiome, and Neurodevelopmental Outcomes: A Prospective Birth Cohort Study

Hannah E. Laue <sup>1,\*,†</sup>, Yike Shen <sup>2,†</sup>, Tessa R. Bloomquist <sup>2</sup>, Haotian Wu <sup>2</sup>, Kasey J. M. Brennan <sup>2</sup>, Raphael Cassoulet <sup>3</sup>, Erin Wilkie <sup>4</sup>, Virginie Gillet <sup>4</sup>, Anne-Sandrine Desautels <sup>4</sup>, Nadia Abdelouahab <sup>4</sup>, Jean Philippe Bellenger <sup>3</sup>, Heather H. Burris <sup>5,6</sup>, Brent A. Coull <sup>7</sup>, Marc G. Weisskopf <sup>8</sup>, Wei Zhang <sup>9</sup>, Larissa Takser <sup>4,10</sup> and Andrea A. Baccarelli <sup>2</sup>

- <sup>1</sup> Department of Epidemiology, Geisel School of Medicine, Dartmouth College, Hanover, NH 03755, USA
- <sup>2</sup> Department of Environmental Health Sciences, Columbia University Mailman School of Public Health, New York, NY 10032, USA; ys3419@cumc.columbia.edu (Y.S.); tb2715@cumc.columbia.edu (T.R.B.); hw2694@cumc.columbia.edu (H.W.); kaseyjmbrennan@gmail.com (K.J.M.B.); andrea.baccarelli@columbia.edu (A.A.B.)
- <sup>3</sup> Département de Chimie, Université de Sherbrooke, Sherbrooke, QC J1K 2R1, Canada; raphael.cassoulet@hotmail.fr (R.C.); jean-philippe.bellenger@usherbrooke.ca (J.P.B.)
- <sup>4</sup> Département de Pédiatrie, Université de Sherbrooke, Sherbrooke, QC J1K 2R1, Canada; erin.wilkie@usherbrooke.ca (E.W.); virginie.gillet@usherbrooke.ca (V.G.); infoasdpsy@gmail.com (A.-S.D.); nadia.abdelouahab@usherbrooke.ca (N.A.); larissa.takser@usherbrooke.ca (L.T.)
- <sup>5</sup> Department of Pediatrics, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA; burrish@chop.edu
- <sup>6</sup> Division of Neonatology, Department of Pediatrics, Children's Hospital of Philadelphia, Philadelphia, PA 19104, USA
- <sup>7</sup> Department of Biostatistics, Harvard T.H. Chan School of Public Health, Boston, MA 02115, USA; bcoull@hsph.harvard.edu
- Department of Environmental Health, Harvard T.H. Chan School of Public Health, Boston, MA 02115, USA; mweissko@hsph.harvard.edu
- Department of Preventive Medicine, Northwestern University Feinberg School of Medicine, Chicago, IL 60611, USA; wei.zhang1@northwestern.edu
- <sup>10</sup> Département de Psychiatrie, Faculté de Médicine et Sciences de la Santé, Université de Sherbrooke, Sherbrooke, QC J1K 2R1, Canada
- \* Correspondence: hannah.e.laue@dartmouth.edu
- † These authors contributed equally to this work.

Abstract: Pregnant individuals are exposed to acetaminophen and caffeine, but it is unknown how these exposures interact with the developing gut microbiome. We aimed to determine whether acetaminophen and/or caffeine relate to the childhood gut microbiome and whether features of the gut microbiome alter the relationship between acetaminophen/caffeine and neurodevelopment. Forty-nine and 85 participants provided meconium and stool samples at 6-7, respectively, for exposure and microbiome assessment. Fecal acetaminophen and caffeine concentrations were quantified, and fecal DNA underwent metagenomic sequencing. Caregivers and study staff assessed the participants' motor and cognitive development using standardized scales. Prenatal exposures had stronger associations with the childhood microbiome than concurrent exposures. Prenatal acetaminophen exposure was associated with a trend of lower gut bacterial diversity in childhood [ $\beta = -0.17$  Shannon Index, 95% CI: (-0.31, -0.04)] and was marginally associated with differences in the relative abundances of features of the gut microbiome at the phylum (Firmicutes, Actinobacteria) and gene pathway levels. Among the participants with a higher relative abundance of Proteobacteria, prenatal exposure to acetaminophen and caffeine was associated with lower scores on WISC-IV subscales. Acetaminophen during bacterial colonization of the naïve gut is associated with lasting alterations in childhood microbiome composition. Future studies may inform our understanding of downstream health effects.



Citation: Laue, H.E.; Shen, Y.; Bloomquist, T.R.; Wu, H.; Brennan, K.J.M.; Cassoulet, R.; Wilkie, E.; Gillet, V.; Desautels, A.-S.; Abdelouahab, N.; et al. In Utero Exposure to Caffeine and Acetaminophen, the Gut Microbiome, and Neurodevelopmental Outcomes: A Prospective Birth Cohort Study. *Int. J. Environ. Res. Public Health* **2022**, *19*, 9357. https://doi.org/10.3390/ ijerph19159357

Academic Editor: Halina B. Röllin

Received: 27 June 2022 Accepted: 28 July 2022 Published: 30 July 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Keywords: acetaminophen; caffeine; microbiome; neurodevelopment; children's health

### 1. Introduction

Acetaminophen, the only analgesic and antipyretic recommended during pregnancy, is used by at least two-thirds of pregnant people and is also one of the most frequently used medications in childhood [1–3]. Recent evidence suggests potential adverse neurodevelopment resulting from exposure, including impaired cognition and increased risk of attention deficit-hyperactivity disorder [4,5], although findings are inconsistent [6,7]. Similarly, physicians recommend that caffeine intake in pregnancy be limited to less than 200 mg per day [8]. This recommendation is partially due to evidence from animal models suggesting that high-dose prenatal exposure causes decreased brain weight, neural tube and central nervous system defects, and adverse behavioral outcomes [9–11], but epidemiologic human studies are inconclusive [12–14]. Studies of childhood caffeine consumption are limited, but it is estimated to average 0.3–1 mg/kg/day in the United States [15].

Factors that confer resilience or susceptibility to these exposures may explain discrepancies in prior findings but have been understudied. One such factor is the gut microbiome, the myriad microorganisms that reside in the gastrointestinal tract [16]. Some bacteria contribute to the metabolism of potentially neurotoxic compounds, including acetaminophen [17], while others may alleviate harmful effects by producing beneficial compounds. In contrast, pathogens may exacerbate adverse associations. By examining microbial features that alter the relationships of caffeine and acetaminophen with neurodevelopment, we may begin to understand the inconsistencies of prior studies. Further, we may identify bacterial species or byproducts that can be used therapeutically to reduce adverse effects when exposure is unavoidable due to limited antipyretic options in pregnancy.

Bacteria colonize the naïve infant gut after birth, and their ability to establish communities is affected by the environment they experience, including chemical exposures [18–20]. Acetaminophen and caffeine cross the placenta [21–23], undergo limited metabolism by the fetus [24,25], and thus accumulate in meconium, making it an ideal matrix to assess exposure experienced by early colonizers of the gut microbiome. Although the infant gut microbiome is highly variable, early exposures can have lasting impacts that are detectable into childhood [26]. Additionally, with the ingestion of acetaminophen and caffeine continuing into childhood, stool remains an informative exposure matrix for microbiome studies. Few studies have examined whether either acetaminophen or caffeine at any age is associated with changes in the microbiome, and none have examined prenatal exposure [27–30].

We aimed to understand the role of the childhood gut microbiome in the associations of two of the most common medications/exposures in pregnancy, acetaminophen, and caffeine, with neurodevelopment in the GESTation and Environment (GESTE) cohort.

## 2. Materials and Methods

## 2.1. Study Participants

Patients were recruited to the GESTE cohort in early pregnancy or at the time of birth between 2007 and 2009 at the University of Sherbrooke, Quebec, Canada (cohort description Supplemental Methods). When the children were aged 6–7 years old, they underwent a battery of neurocognitive tests. In addition, a convenient sample of children provided stool samples for microbiome analysis. This analysis included children with stool samples at age 6–7 that underwent metagenomic sequencing, completed the neurocognitive tests, and had exposures assessed in the meconium (prenatal, n = 49) and/or stool at age 6–7 (cross-sectional, n = 85). Written informed consent was obtained from parents. All of the study protocols were approved by the Institutional Review Boards of the University of Sherbrooke, Harvard T.H. Chan School of Public Health, and Columbia University.

## 2.2. Exposure Assessment

Meconium was collected from infants who passed it after birth at the hospital and stored at -80 °C, and the stools were collected by caregivers at the participants' homes when they were 6–7 years old. Caffeine and acetaminophen were extracted from the feces and quantified as previously described [6,31]. Caffeine exposures were log<sub>2</sub> transformed for all analyses to reduce the influence of outlier concentrations. Approximately half of the meconium and 90% of the childhood stool had non-detectable exposure to acetaminophen; thus, we dichotomized exposure as exposed (detected) and non-exposed (below the limit of quantification, LOQ). The details on the LOQ for both exposures are available in the Supplemental Methods.

#### 2.3. Microbiome Assessment

Total DNA was extracted from childhood stool using established protocols [32]. Metagenomic sequencing was conducted at New York University's Langone Genome Technology Center. After preprocessing and quality control as previously described [32], bacterial species and their pathways were annotated with MetaPhlAn 3 and HUMAnN 3 (MetaCyc) [33], respectively, and normalized to relative abundance tables (i.e., divided by the total number of reads for each sample) [34]. Bacterial alpha diversity was quantified with the Shannon [35] and Pielou [36] Indices, calculated from species relative abundance tables with the *phyloseq* package [37]. Species beta diversity was measured with UniFrac (weighted and unweighted) [38], Bray-Curtis [39], and Jaccard [40] distances.

## 2.4. Neurodevelopmental Assessment

When the participants were 6–7 years old, caregivers completed the Questionnaire sur le Trouble de L'Acquisition de la Coordination (QTAC), the French equivalent of the Developmental Coordination Disorder Questionnaire. Through this, they compared their child's motor development to their peers [41]. In addition, a trained member of the study staff administered five subtests from the Wechsler Intelligence Scale for Children, 4th edition (WISC-IV) to the children, including Block Design, Coding, Digit Span, Information, and Vocabulary [42]. To assess an underlying component of intelligence not captured by individual subtests, we created a summary score of the five completed subtests (sum of five subtests; WISCsum).

#### 2.5. Statistical Methods

The covariates were selected based on their potential to confound the association between the exposures and the microbiome based on prior knowledge. In final analyses, we adjusted for sex, ever breastfeeding, mode of birth (vaginal or caesarean), and family income (Supplemental Methods). In sensitivity analyses, we adjusted for maternal IQ (Supplemental Methods) and for both exposure windows concurrently. The alpha diversity indices were linearly regressed against caffeine or acetaminophen, adjusting for covariates. A Benjamini–Hochberg false discovery rate (FDR) of q < 0.1 was considered statistically significant [43]. Differences in beta diversity related to caffeine and acetaminophen were tested with the *adonis* function, a permutational analysis of variance [44]. Microbiome Multivariable Association with Linear Models (MaAsLin2) was used to determine whether acetaminophen or caffeine showed a trend of association (q < 0.1) with bacterial phyla, species, or pathways with a prevalence above 10% [45].

We have previously reported no association between acetaminophen exposure and QTAC/WISC-IV scores [6], but we hypothesized that features of the microbiome might modify the relationships between prenatal acetaminophen or caffeine and neurodevelopment. To limit multiple testing penalties, we screened the most abundant (top 10%) bacterial species (n = 34), pathways (n = 36), and all phyla (n = 9) for interactions with caffeine and acetaminophen using linear regression. Model fit was assessed using likelihood ratio tests. All of the statistical analyses were conducted in the R statistical environment (v4.0.2; R

Core Team, Vienna, Austria). Code is available at https://github.com/YikeShen (accessed on 27 June 2022).

#### 3. Results

The population characteristics among the participants included in the analysis were similar to the full cohort examined at age 6–7 years (Table 1). The GESTE cohort is relatively homogeneous in demographics (primarily upper-middle-class French-Canadians). The mean maternal age at recruitment was 29 years, and more than half of the participants were nulliparous. Breastfed children (87.8%) were overrepresented in the analytical population. The average family income was higher at follow-up than at delivery, but our analytical cohorts were similar to the full cohort. The percentage of the participants with detectable levels of acetaminophen in their meconium was similar in the overall and analytical populations (Table 1). All of the participants had detectable levels of caffeine in the meconium, with a median concentration of 390 ng/g meconium in the pilot population compared to 399 ng/g meconium in all children with available meconium who were followed to an age of 6 years (Table 1). The median caffeine concentration was significantly lower in childhood (24 ng/g stool, 100% detection), and fewer (10%) participants had detectable levels of acetaminophen (Table 1).

**Table 1.** Characteristics of the GESTE meconium analytic population (N = 49), cross-sectional analytic population (n = 85), population with meconium (N = 197), and overall population (N = 365) at 6–7-year-old follow-up [expressed as mean  $\pm$  SD or n (%), unless specified otherwise].

	Full Cohort (n = 365)	Population with Meconium (n = 197)	Meconium Analytic Population (n = 49)	Cross-Sectional Analytic Population (n = 85)
amily Characteristics				
Aaternal Age at Recruitment	$29.1\pm4.44$	$29.4\pm4.45$	$29.1\pm3.85$	$28.9\pm4.09$
years)	$27.1 \pm 1.11$	$2).1 \pm 1.10$	20.1 ± 0.00	20.7 ± 4.07
Maternal Pre-pregnancy Body				
Mass Index (kg/m²)				
Available	$25.3\pm5.95$	$25.4 \pm 5.85$	$25.5\pm 6.30$	$25.2\pm6.01$
Missing	68 (18.6)	13 (6.6)	12 (24.5)	16 (18.8)
Family Income (CAD <sup>a</sup> )				
Available	$70,800 \pm 41,200$	$71,\!600 \pm 47,\!300$	$69,600 \pm 34,600$	93,900 $\pm$ 47,100 <sup>b</sup>
Missing	20 (5.5)	14 (7.1)	4 (8.2)	6 (7.1) <sup>b</sup>
arity				
Nulliparous	205 (56.2)	113 (57.4)	26 (53.1)	46 (54.1)
Parous	158 (43.3)	83 (42.1)	23 (46.9)	39 (45.9)
Missing	2 (0.5)	1 (0.5)	0(0)	0(0)
Birth Characteristics	~ /	~ /		
Gestational Age (weeks)	$39.1 \pm 1.43$	$39.1 \pm 1.44$	$39.4 \pm 1.09$	$39.4 \pm 1.18$
Birth Mode				
Vaginal	299 (81.9)	162 (82.2)	40 (81.6)	68 (80)
Caesarean Section	66 (18.1)	35 (17.8)	9 (18.4)	17 (20)
Child Sex				
Male	199 (54.5)	107 (54.3)	25 (51.0)	44 (51.8)
Female	166 (45.5)	90 (45.7)	24 (49.0)	41 (48.2)
Child Birthweight (g)	$3400 \pm 486$	$3370 \pm 484$	$3440 \pm 419$	$3460 \pm 445$
Breast Feeding Status				
Ever breastfed	284 (77.8)	159 (80.7)	43 (87.8)	68 (80.0)
Never breastfed	65 (17.8)	30 (15.2)	4 (8.2)	14 (16.5)
Missing	16 (4.4)	8 (4.1)	2 (4.1)	3 (3.5)
Neurological Outcomes	10 (111)	0 (111)	= ()	0 (0.0)
VISC-IV <sup>c</sup> : Block Design	$9.54 \pm 2.92$	$9.43 \pm 2.88$	$10.4\pm2.99$	$10.3\pm2.98$
VISC-IV: Coding	$10.5 \pm 2.94$	$10.5 \pm 2.92$	$10.1 \pm 2.55$ $10.8 \pm 2.56$	$10.5 \pm 2.67$ $11.1 \pm 2.67$
VISC-IV: Digit Span	$9.40 \pm 2.58$	$9.39 \pm 2.61$	$10.0 \pm 2.26$	$9.81 \pm 2.18$
VISC-IV: Information	$9.55 \pm 2.29$	$9.37 \pm 2.28$	$9.45 \pm 2.17$	$9.81 \pm 2.23$
VISC-IV: Vocabulary	$10.4 \pm 2.70$	$10.1 \pm 2.76$	$10.3 \pm 3.29$	$10.6 \pm 2.97$
VISC-IV Summary Score	$49.4 \pm 8.36$	$48.9 \pm 8.17$	$51.0 \pm 7.70$	$51.7 \pm 7.86$
DTAC <sup>d</sup>	$49.4 \pm 0.00$ $61.0 \pm 8.91$	$40.9 \pm 0.17$ $61.6 \pm 8.32$	$62.2 \pm 8.20$	$62.1 \pm 8.11$

Table 1.	Cont.
----------	-------

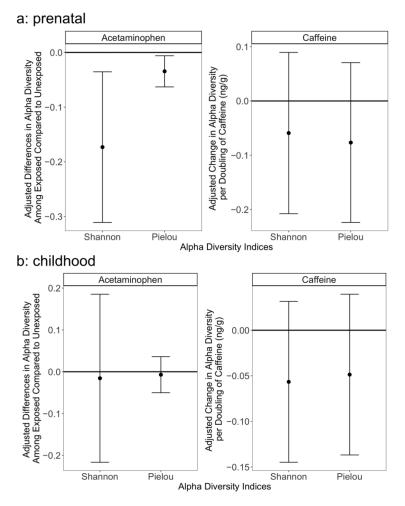
	Full Cohort (n = 365)	Population with Meconium (n = 197)	Meconium Analytic Population (n = 49)	Cross-Sectional Analytic Population (n = 85)
Maternal Intelligence Quotient				
>95 percentile			30 (61.2)	
$\leq 95$ percentile			19 (38.8)	
Exposure of Interest				
Caffeine (median [25th percentile, 75th percentile]; ng/g meconium) Acetaminophen	-	399 [2.82, 5170]	390 [15.3, 3110]	24.7 [0.184, 231] <sup>e</sup>
Detected	-	100 (50.8)	20 (40.8)	7 (8.2) <sup>b</sup>
Not Detected	-	97 (49.2)	29 (59.2)	78 (91.8) <sup>b</sup>

<sup>a</sup> Canadian dollars. <sup>b</sup> Family income at year 6–7 follow-up. <sup>c</sup> Wechsler Intelligence Scale for Children, 4th edition.

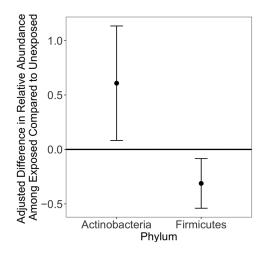
<sup>d</sup> Questionnaire sur le Trouble de L'Acquisition de la Coordination. <sup>e</sup> Exposure concentrations in year 6–7 children stool.

Prenatal acetaminophen exposure was associated with lower alpha diversity using multiple indices [e.g.,  $\beta = 0.17$  lower Shannon Index comparing exposed to unexposed (95% CI: -0.21, -0.09), q = 0.044; Figure 1, Table S1]. At the phylum level, prenatal acetaminophen exposure showed a trend of association with lower relative abundance of Firmicutes [ $\beta = -0.09$  comparing exposed to unexposed (95% CI: -0.16, -0.03), q = 0.062] and higher relative abundance of Actinobacteria [Figure 2, Table S2,  $\beta = 0.183$ , (95% CI: 0.03, 0.34), q = 0.085], but there were no associations at the species level (Table S3). In sensitivity analyses co-adjusting for cross-sectional exposure, the associations were of a similar magnitude (Table S2). The microbiomes of children exposed to prenatal acetaminophen had higher relative abundance of genes in the succinate fermentation to butanoate pathway [Figure 3, Table S4, MetaCyc Pathway ID: PWY-5677;  $\beta = 0.33$ , (95%) CI: 0.17, 0.49), q = 0.072]. In contrast, prenatal caffeine exposure was not significantly associated with alpha diversity or the relative abundance of bacterial species, phyla, or functional pathways (Figures 1 and 3, Tables S1, S2, S5 and S6). Neither prenatal exposure was associated with differences in beta diversity (Table S7). In general, cross-sectional exposure was not associated with differences in the microbiome, even when restricting to subjects with meconium exposures, although caffeine exposure in stool at age 6-7 years was associated with an increased relative abundance of three pathways (Table S6): methionine synthesis [MetaCyc Pathway ID: PWY-5345;  $\beta = 0.24$  (0.12, 0.35), q = 0.018], assimilatory sulfate reduction [MetaCyc Pathway ID: SO4ASSIM-PWY;  $\beta = 0.24$  (0.12, 0.36), q = 0.018], and sulfate assimilation and cysteine synthesis [MetaCyc Pathway ID: SULFATE-CYS-PWY;  $\beta = 0.24 \ (0.12, 0.35), q = 0.018].$ 

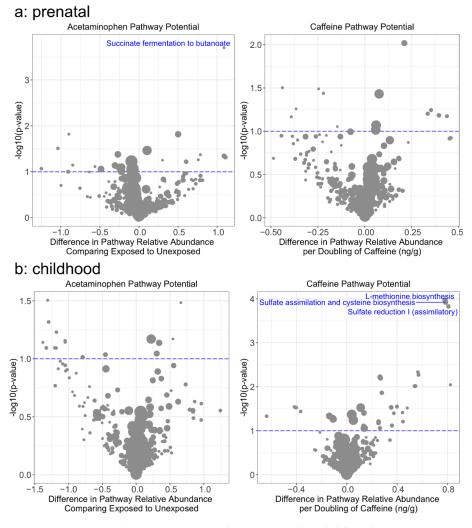
When testing whether the most abundant phyla, species, or pathways modified the association between exposure to acetaminophen/caffeine and neurodevelopmental scores, we found that the relative abundance of Proteobacteria modified the neurodevelopmental effects of both acetaminophen and caffeine (Figure 4). The participants with detectable concentrations of acetaminophen had slightly higher scores on WISC-IV subtests, as previously reported, [6] but this benefit was weakened among those with higher levels of Proteobacteria (Table S8). In contrast, higher concentrations of caffeine related to lower scores on several WISC-IV subtests, and these effects were exacerbated among children with higher levels of Proteobacteria (Table S9). No species (Tables S10 and S11) or functional pathway (Tables S12 and S13) modified the exposure–outcome relationship. Adjusting for maternal IQ did not meaningfully change the interpretation of results (Tables S8–S13).



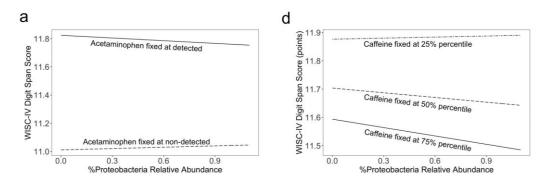
**Figure 1.** Association between acetaminophen/caffeine concentrations in (**a**) meconium (n = 49) and (**b**) 6–7-year-old stool (n = 85) and alpha diversity. Diversity was measured with Shannon Diversity Index and Pielou Evenness Index. Whiskers represent 95% confidence intervals. Models are adjusted for whether the child was ever breastfed, sex, mode of birth, and family income.



**Figure 2.** Association between prenatal exposure to acetaminophen and phyla (n = 49). Whiskers represent 95% confidence intervals. Models adjust for whether the child was ever breastfed, sex, mode of birth, and family income.

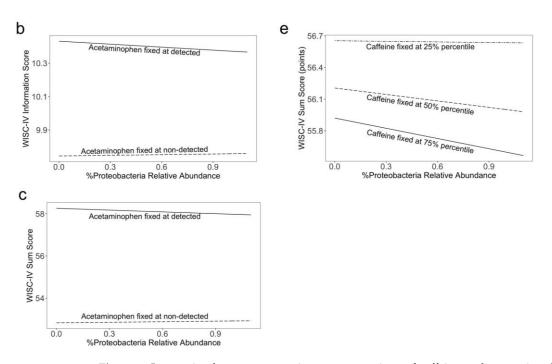


**Figure 3.** Association between (**a**) prenatal (n = 49) and (**b**) childhood exposure (n = 85) to caffeine or acetaminophen and relative abundance of functional pathways. Effect estimates for caffeine are expressed per doubling of exposure. Effect estimates for acetaminophen are comparing exposed to unexposed. Models are adjusted for whether the child was ever breastfed, sex, mode of birth, and socioeconomic status. Point size is proportional to average relative abundance of the represented pathway. Pathways with an FDR *q*-value < 0.1 are labeled. Full names: L-methionine biosynthesis = Superpathway of L-methionine biosynthesis (by sulfhydrylation); Sulfate assimilation and cysteine biosynthesis.



7 of 14

Figure 4. Cont.



**Figure 4.** Interaction between meconium concentrations of caffeine and acetaminophen with Proteobacteria (n = 49). Models are adjusted for whether the child was ever breastfed, sex, mode of birth, and socioeconomic status. In (a-c), solid lines show the association when acetaminophen is detected, and dashed lines show the association when acetaminophen is not detected. In (d-e), solid lines show the association when acetaminophen is not detected. In (d-e), solid lines show the association when caffeine is at the 75th percentile, dashed lines show when caffeine is at the 50th percentile, and dash-dot lines show when caffeine is at the 25th percentile. Subfigures (a,d) show the association between Proteobacteria and WISC-IV Digit Span scores; subfigures (c,e) show the association between Proteobacteria and WISC-IV Information scores; subfigures (c,e) show the association between Proteobacteria and WISC-IV summary scores.

### 4. Discussion

In this preliminary study of the potential role of the microbiome as a modifier of the associations between prenatal exposure to non-prescription consumer products and neurodevelopment, we found suggestive evidence that certain childhood gut bacteria may modify the association between caffeine/acetaminophen and cognitive development. Additionally, prenatal acetaminophen exposure was associated with alterations in the gut microbiome in childhood. These findings, if confirmed in larger and more diverse cohorts, may explain differences in the prior literature and suggest therapeutic interventions in cases where exposure is unavoidable.

Prenatal acetaminophen showed a trend of association with lower microbial diversity in childhood, accompanied by decreased relative abundance of Firmicutes and increased relative abundance of Actinobacteria. Higher taxonomic diversity is generally thought to indicate a healthier system in adults, as it contains greater functional diversity to complement the host's response to exposures [46]. Infants have low taxonomic diversity that increases with the introduction of solid foods [47]. Similarly, vaginally delivered infants have relatively low levels of Firmicutes and high levels of Actinobacteria compared to later samples [26]. It is possible that prenatal exposure to acetaminophen delays the maturation of the gut microbiome to a more adult-like composition. This hypothesis should be explored with more regular exposure assessment and microbiome sampling. In addition, the reason that no species level associations were detected could be due to the relatively small sample size. Future investigations with a larger sample size could help elucidate differences at the species level. Prenatal acetaminophen was associated with an increased relative abundance of genes in the pathway of succinate fermentation to butanoate. Metatranscriptomic or metabolomic analyses could inform whether there are functional differences as a result of differences in the relative abundance of this pathway and whether there are downstream health consequences.

In contrast, we did not observe any associations between prenatal caffeine exposure and gut microbial features. While no epidemiologic studies of changes in the gut microbiome resulting from caffeine exposure have been reported, rats who consumed caffeinated coffee with their chow experienced an increase in Enterobacteriaceae and *Clostridium leptum* [27]. These results may be due to other chemical compounds found in coffee (e.g., antioxidants), but another study found that rats treated with high, repeated doses of caffeine (not reflective of the fetal experience) had increased *Lactobacillus* compared to untreated rats [28]. It is possible that our study was underpowered to detect these changes or that caffeine exposure in the naïve gut is not relevant to later microbiome composition. Although we did not observe differences in species or phylum relative abundance related to caffeine exposure, several gene pathways had increased relative abundance with increasing exposure. For example, concurrent exposure was associated with an increased relative abundance of methionine synthesis genes. This may be in response to increased methyl group availability, as caffeine metabolism requires demethylation. In vitro models may inform the underlying mechanism of this and other associations.

The associations of both acetaminophen and caffeine with cognitive domains were modified by the relative abundance of Proteobacteria in the childhood gut. Proteobacteria includes many notorious pathogens, including *Escherichia, Shigella*, and *Salmonella*, all in the Gammaproteobacteria. An elevated relative abundance of the phylum is associated with worse cognitive performance or impairment in clinical [48,49], population-based [50,51], and animal studies [52], in addition to being considered a marker of gut dysbiosis [53]. Our findings suggest that in addition to the direct negative relationship between Proteobacteria relative abundance and cognitive scores observed in prior studies, it may modify the associations of caffeine and acetaminophen with cognition in an adverse manner. In the case of acetaminophen, Proteobacteria weakened the beneficial impact of acetaminophen on short-term memory, general knowledge, and our measure of overall cognition. In contrast, in the case of caffeine, Proteobacteria exacerbated the adverse impact of caffeine on memory and cognition. This may suggest that Proteobacteria acts through similar pathways (e.g., immune response or oxidative stress) as caffeine and acetaminophen in their associations with cognition [53–55].

There is substantial interest in the microbiome as a mediator of health effects. We did not uncover bacterial features at 6–7 years that indicate they may mediate the caffeine/acetaminophen-cognition association (i.e., no feature associated with an exposure that is also associated with cognition in our cohort). However, our study is underpowered to detect small effects and does not discount the possibility that the microbiome mediates these or other associations between chemicals and health outcomes. Importantly, our findings highlight the need to consider whether features of the microbiome, especially Proteobacteria, can alter host response to exogenous exposures, including pharmaceuticals. Because Proteobacteria are overrepresented in the microbiomes of patients with inflammatory bowel diseases [56] and cognitive decline [50,51], compared to healthy controls, this could be particularly important when designing precision medications for these outcomes.

The associations of caffeine and acetaminophen with the microbiome may differ when considering prenatal and mid-childhood exposure due to windows of microbial susceptibility to these exposures or due to differences in the exposure matrix. Fecal concentrations of exposure represent the environment of the gut bacteria more directly than other biomarkers, thus are an intriguing option for microbiome studies. However, meconium begins to accumulate in the second trimester and captures cumulative exposure over the last six months of pregnancy [57], while stool accumulates over a much shorter period (~24 h), capturing more acute exposures [58]. This is captured by the differences in caffeine concentrations between meconium and stool in our study. In addition to differences arising from acute vs. chronic exposures, meconium captures exposure in the naïve gut, which may have a larger impact on long-term microbiome composition by altering the suitability of the gut environment for early colonizers of the intestine. Future animal studies with repeated exposure or epidemiological studies with more frequent exposure assessment could inform windows of susceptibility of the pediatric gut microbiome.

Our study should be considered within the context of its limitations. Due to the small sample size, we could adjust for a limited number of confounders. We prioritized confounders of the association between exposure and the microbiome and selected those that likely have the strongest confounding effects. Ideally, we would control for factors that confound the association between caffeine exposure and neurodevelopment and factors that confound the association between the microbiome and neurodevelopment. Our exposure assessment assay only quantified concentrations of the parent compounds (acetaminophen/caffeine) and not their metabolites. Both caffeine and acetaminophen are known to cross the placenta, whereas the metabolites are more rapidly excreted, and the fetus has limited capacity to metabolize either compound [23,59,60]. Therefore, we captured the majority of fetal exposure from the second trimester (when meconium begins accumulating) to birth. An assessment of the early-life microbiome in relation to prenatal acetaminophen/caffeine exposure may reveal transient disturbances in the microbiome that relate to neurodevelopment during a sensitive window. Our cross-sectional exposure data reduce the possibility that our findings result from correlations between prenatal and childhood exposures. There is potential for reverse causation due to our fecal samples and cognitive measures being collected at the same age.

This novel study also has many strengths. This is one of the first studies to examine the effects of chemicals on the developing microbiome and the first to examine prenatal pharmaceutical exposure. As such, it is particularly interesting and innovative that we measured exposure in the meconium, a matrix that represents the naïve gut environment that bacteria colonize in early life. Further, we used a quantitative measure of exposure to acetaminophen and caffeine rather than relying on maternal reports of intake during pregnancy. Self-report may be under-reported due to the perception of caffeine consumption during pregnancy as a risk behavior or subject to recall bias in cases where the exposure data are collected retrospectively. The longitudinal design and temporality of associations remove the possibility of reverse causation, a common concern of microbiome epidemiology studies. Our use of metagenomic sequencing to profile the childhood microbiome allowed us to examine many aspects of the microbiome, including functional potential, which may result from exposure or modify the exposure–outcome relationship.

#### 5. Conclusions

In conclusion, we identified an association between prenatal exposure to acetaminophen and gut microbiome composition in mid-childhood. Larger studies are needed to confirm these results and examine the mechanisms by which in utero exposures affect long-term gut microbiome development. Specifically, studies should collect perinatal maternal and infant microbiome samples to determine whether exposures are associated with the maternal taxa, which are then passed to the infant or if exposures act on the microbiome by affecting what taxa are able to colonize the naïve gut. In addition, we found evidence that certain bacteria may modify the relationships between exogenous exposures and health outcomes, a possible explanation for discrepancies in prior studies on these exposures. The potential for the microbiome to confer resilience or susceptibility to toxic exposures should be explored more fully, including varying exposures, windows of exposure, and outcomes. Overall, this study positions the microbiome as a critical, modifiable feature for future environmental health studies.

**Supplementary Materials:** The following are available online at https://www.mdpi.com/article/ 10.3390/ijerph19159357/s1, Supplemental Methods, Table S1. Associations of acetaminophen and caffeine with childhood bacterial diversity, Table S2. Associations of acetaminophen and caffeine exposure with bacterial phylum relative abundance, Table S3. Difference in bacterial species relative abundance comparing those exposed to acetaminophen to unexposed, Table S4. Difference in MetaCyc pathway relative abundance comparing acetaminophen exposed to unexposed, Table S5. Change in bacterial species relative abundance per doubling caffeine (ng/g), Table S6. Change in MetaCyc pathway relative abundance per doubling caffeine (ng/g), Table S7. Associations of acetaminophen and caffeine with beta diversity, Table S8. Acetaminophen-phylum interactions in association with neurodevelopmental outcomes, Table S9. Caffeine-phylum interactions in association with neurodevelopmental outcomes, Table S10. Caffeine-species interactions in association with neurodevelopment, Table S11. Acetaminophen-species interactions in associations with neurodevelopment, Table S12. Caffeine-pathway interactions in association with neurodevelopment, Table S13. Acetaminophen-pathway interactions in association with neurodevelopment, Table S13. Acetaminophen-pathway interactions in association with neurodevelopment, Table S13. Acetaminophen-pathway interactions in association with neurodevelopment. References [61–64] are cited in the supplementary materials

Author Contributions: Conceptualization, H.E.L., H.H.B., B.A.C., M.G.W. and A.A.B.; Data Curation, H.E.L., Y.S., T.R.B., K.J.M.B., R.C., E.W., V.G., A.-S.D., N.A., J.P.B., L.T. and A.A.B.; Formal Analysis, H.E.L., Y.S. and H.W.; Funding Acquisition, H.E.L., K.J.M.B., J.P.B., W.Z., L.T. and A.A.B.; Investigation, H.E.L., Y.S., T.R.B., R.C., J.P.B.; Methodology, H.E.L., Y.S., H.W. and W.Z..; Project Administration, T.R.B., K.J.M.B. and V.G.; Supervision, A.A.B.; Visualization, H.E.L., Y.S. and H.W.; Writing—Original Draft Preparation, H.E.L. and Y.S.; Writing—Review & Editing, H.E.L., Y.S., T.R.B., H.W., K.J.M.B., R.C., E.W., V.G., A.-S.D., N.A., J.P.B., B.A.C., M.G.W., W.Z., L.T. and A.A.B. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was supported by the National Institutes of Environmental Health (grant numbers R01ES027845, R21ES024841, P30ES009089, P30ES000002, and K23ES022242), the National Institutes of Health Office of the Director (UH3OD023275), the Canadian Institutes of Health Research (MOP-84551), and Canadian Research Chair grant CRC-950-230570 from the Natural Sciences and Engineering Research Council of Canada.

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Institutional Review Boards of the University of Sherbrooke (05-057-S1, 2008-103; 17 December 2015), Harvard T.H. Chan School of Public Health (IRB15-0810; 10 March 2015), and Columbia University (AAAR5408; renewed 14 April 2022).

**Informed Consent Statement:** Informed consent was obtained from caregivers of all subjects involved in the study.

**Data Availability Statement:** Epidemiologic data are not available due to their sensitive and potentially identifiable nature. Requests to work with the GESTation and Environment cohort should be addressed to Larissa Takser.

**Conflicts of Interest:** The authors declare no conflict of interest. The sponsors had no role in the design, execution, interpretation, or writing of the study.

## References

- McKenna, L.; McIntyre, M. What over-the-counter preparations are pregnant women taking? A literature review. J. Adv. Nurs. 2006, 56, 636–645. [CrossRef]
- Werler, M.M.; Mitchell, A.A.; Hernandez-Diaz, S.; Honein, M.A. Use of over-the-counter medications during pregnancy. Am. J. Obstet. Gynecol. 2005, 193, 771–777. [CrossRef]
- Vernacchio, L.; Kelly, J.P.; Kaufman, D.W.; Mitchell, A.A. Medication Use Among Children <12 Years of Age in the United States: Results From the Slone Survey. *Pediatrics* 2009, 124, 446–454. [CrossRef]
- Baker, B.H.; Lugo-Candelas, C.; Wu, H.; Laue, H.E.; Boivin, A.; Gillet, V.; Aw, N.; Rahman, T.; Lepage, J.-F.; Whittingstall, K. Association of prenatal acetaminophen exposure measured in meconium with risk of attention-deficit/hyperactivity disorder mediated by frontoparietal network brain connectivity. *JAMA Pediatrics* 2020, 174, 1073–1081. [CrossRef]
- 5. Bauer, A.Z.; Kriebel, D.; Herbert, M.R.; Bornehag, C.-G.; Swan, S.H. Prenatal paracetamol exposure and child neurodevelopment: A review. *Horm. Behav.* **2018**, *101*, 125–147. [CrossRef]
- Laue, H.E.; Cassoulet, R.; Abdelouahab, N.; Serme-Gbedo, Y.K.; Desautels, A.-S.; Brennan, K.J.; Bellenger, J.-P.; Burris, H.H.; Coull, B.A.; Weisskopf, M.G.; et al. Association between meconium acetaminophen and childhood neurocognitive development in GESTE, a Canadian cohort study. *Toxicol. Sci.* 2019, *167*, 138–144. [CrossRef]
- Bertoldi, A.D.; Rifas-Shiman, S.L.; Boing, A.C.; Silva Dal Pizzol, T.; Miranda, V.I.A.; Silveira, M.P.T.; Freitas Silveira, M.; Domingues, M.R.; Santos, I.S.; Bassani, D.G.; et al. Associations of acetaminophen use during pregnancy and the first year of life with neurodevelopment in early childhood. *Paediatr. Perinat. Epidemiol.* 2020, 34, 267–277. [CrossRef] [PubMed]
- 8. World Health Organization. *WHO Recommendations on Antenatal Care for a Positive Pregnancy Experience;* WHO Press: Geneva, Switzerland, 2016.

- 9. Nehlig, A.; Debry, G. Potential teratogenic and neurodevelopmental consequences of coffee and caffeine exposure: A review on human and animal data. *Neurotoxicology Teratol.* **1994**, *16*, 531–543. [CrossRef]
- Ma, Z.-l.; Qin, Y.; Wang, G.; Li, X.-d.; He, R.-r.; Chuai, M.; Kurihara, H.; Yang, X. Exploring the Caffeine-Induced Teratogenicity on Neurodevelopment Using Early Chick Embryo. *PLoS ONE* 2012, 7, e34278. [CrossRef]
- 11. Sobotka, T.J. Neurobehavioral Effects of Prenatal Caffeine. Ann. N. Y. Acad. Sci. 1989, 562, 327–339. [CrossRef]
- 12. Del-Ponte, B.; Santos, I.S.; Tovo-Rodrigues, L.; Anselmi, L.; Munhoz, T.N.; Matijasevich, A. Caffeine consumption during pregnancy and ADHD at the age of 11 years: A birth cohort study. *BMJ Open* **2016**, *6*, e012749. [CrossRef] [PubMed]
- 13. Loomans, E.M.; Hofland, L.; van der Stelt, O.; van der Wal, M.F.; Koot, H.M.; Van den Bergh, B.R.; Vrijkotte, T.G. Caffeine intake during pregnancy and risk of problem behavior in 5- to 6-year-old children. *Pediatrics* **2012**, *130*, e305–e313. [CrossRef] [PubMed]
- Morgan, S.; Koren, G.; Bozzo, P. Is caffeine consumption safe during pregnancy? *Can. Fam. Physician* 2013, *59*, 361–362. [PubMed]
   Castellanos, F.X.; Rapoport, J.L. Effects of caffeine on development and behavior in infancy and childhood: A review of the
- published literature. *Food Chem. Toxicol.* 2002, 40, 1235–1242. [CrossRef]
  16. Turnbaugh, P.J.; Ley, R.E.; Hamady, M.; Fraser-Liggett, C.M.; Knight, R.; Gordon, J.I. The Human Microbiome Project. *Nature* 2007, 449, 804–810. [CrossRef] [PubMed]
- 17. Malfatti, M.A.; Kuhn, E.A.; Murugesh, D.K.; Mendez, M.E.; Hum, N.; Thissen, J.B.; Jaing, C.J.; Loots, G.G. Manipulation of the Gut Microbiome Alters Acetaminophen Biodisposition in Mice. *Sci. Rep.* **2020**, *10*, 4571. [CrossRef] [PubMed]
- Laue, H.E.; Brennan, K.J.M.; Gillet, V.; Abdelouahab, N.; Coull, B.A.; Weisskopf, M.G.; Burris, H.H.; Zhang, W.; Takser, L.; Baccarelli, A.A. Associations of prenatal exposure to polybrominated diphenyl ethers and polychlorinated biphenyls with long-term gut microbiome structure: A pilot study. *Environ. Epidemiol.* 2019, *3*, e039. [CrossRef] [PubMed]
- 19. Laue, H.E.; Moroishi, Y.; Jackson, B.P.; Palys, T.J.; Madan, J.C.; Karagas, M.R. Nutrient-toxic element mixtures and the early postnatal gut microbiome in a United States longitudinal birth cohort. *Environ. Int.* **2020**, *138*, 105613. [CrossRef]
- 20. Chiu, K.; Warner, G.; Nowak, R.A.; Flaws, J.A.; Mei, W. The Impact of Environmental Chemicals on the Gut Microbiome. *Toxicol. Sci.* **2020**, *176*, 253–284. [CrossRef]
- 21. Goldstein, A.; Warren, R. Passage of caffeine into human gonadal and fetal tissue. *Biochem. Pharmacol.* **1962**, *11*, 166–168. [CrossRef]
- 22. Levy, G.; Garrettson, L.K.; Soda, D.M. Letter: Evidence of placental transfer of acetaminophen. *Pediatrics* **1975**, *55*, 895. [CrossRef] [PubMed]
- 23. Nitsche, J.F.; Patil, A.S.; Langman, L.J.; Penn, H.J.; Derleth, D.; Watson, W.J.; Brost, B.C. Transplacental Passage of Acetaminophen in Term Pregnancy. *Am. J. Perinatol.* **2017**, *34*, 541–543. [CrossRef] [PubMed]
- 24. Mian, P.; Allegaert, K.; Conings, S.; Annaert, P.; Tibboel, D.; Pfister, M.; van Calsteren, K.; van den Anker, J.N.; Dallmann, A. Integration of Placental Transfer in a Fetal-Maternal Physiologically Based Pharmacokinetic Model to Characterize Acetaminophen Exposure and Metabolic Clearance in the Fetus. *Clin. Pharmacokinet.* **2020**, *59*, 911–925. [CrossRef] [PubMed]
- Grosso, L.M.; Bracken, M.B. Caffeine Metabolism, Genetics, and Perinatal Outcomes: A Review of Exposure Assessment Considerations during Pregnancy. Ann. Epidemiol. 2005, 15, 460–466. [CrossRef]
- Yassour, M.; Vatanen, T.; Siljander, H.; Hämäläinen, A.-M.; Härkönen, T.; Ryhänen, S.J.; Franzosa, E.A.; Vlamakis, H.; Huttenhower, C.; Gevers, D.; et al. Natural history of the infant gut microbiome and impact of antibiotic treatment on bacterial strain diversity and stability. *Sci. Transl. Med.* 2016, *8*, 343ra381. [CrossRef] [PubMed]
- Cowan, T.E.; Palmnäs, M.S.A.; Yang, J.; Bomhof, M.R.; Ardell, K.L.; Reimer, R.A.; Vogel, H.J.; Shearer, J. Chronic coffee consumption in the diet-induced obese rat: Impact on gut microbiota and serum metabolomics. *J. Nutr. Biochem.* 2014, 25, 489–495. [CrossRef] [PubMed]
- Scorza, C.; Piccini, C.; Martinez Busi, M.; Abin Carriquiry, J.A.; Zunino, P. Alterations in the Gut Microbiota of Rats Chronically Exposed to Volatilized Cocaine and Its Active Adulterants Caffeine and Phenacetin. *Neurotox. Res.* 2018. [CrossRef]
- Clayton, T.A.; Baker, D.; Lindon, J.C.; Everett, J.R.; Nicholson, J.K. Pharmacometabonomic identification of a significant hostmicrobiome metabolic interaction affecting human drug metabolism. *Proc. Natl. Acad. Sci. USA* 2009, 106, 14728–14733. [CrossRef]
- 30. Maier, L.; Pruteanu, M.; Kuhn, M.; Zeller, G.; Telzerow, A.; Anderson, E.E.; Brochado, A.R.; Fernandez, K.C.; Dose, H.; Mori, H.; et al. Extensive impact of non-antibiotic drugs on human gut bacteria. *Nature* **2018**, *555*, 623. [CrossRef]
- Haroune, L.; Cassoulet, R.; Lafontaine, M.P.; Belisle, M.; Garant, D.; Pelletier, F.; Cabana, H.; Bellenger, J.P. Liquid chromatographytandem mass spectrometry determination for multiclass pesticides from insect samples by microwave-assisted solvent extraction followed by a salt-out effect and micro-dispersion purification. *Anal. Chim. Acta* 2015, *891*, 160–170. [CrossRef]
- Shen, Y.; Laue, H.E.; Shrubsole, M.J.; Wu, H.; Bloomquist, T.R.; Larouche, A.; Zhao, K.; Gao, F.; Boivin, A.; Prada, D.; et al. Associations of Childhood and Perinatal Blood Metals with Children's Gut Microbiomes in a Canadian Gestation Cohort. *Environ. Health Perspect.* 2022, 130, 017007. [CrossRef] [PubMed]
- Caspi, R.; Altman, T.; Billington, R.; Dreher, K.; Foerster, H.; Fulcher, C.A.; Holland, T.A.; Keseler, I.M.; Kothari, A.; Kubo, A.; et al. The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of Pathway/Genome Databases. *Nucleic Acids Res.* 2013, 42, D459–D471. [CrossRef] [PubMed]
- Franzosa, E.A.; McIver, L.J.; Rahnavard, G.; Thompson, L.R.; Schirmer, M.; Weingart, G.; Lipson, K.S.; Knight, R.; Caporaso, J.G.; Segata, N. Species-level functional profiling of metagenomes and metatranscriptomes. *Nat. Methods* 2018, 15, 962–968. [CrossRef] [PubMed]

- 35. Shannon, C.E. A mathematical theory of communication. Bell Syst. Tech. J. 1948, 27, 379–423. [CrossRef]
- Pielou, E.C. The measurement of diversity in different types of biological collections. *J. Theor. Biol.* 1966, 13, 131–144. [CrossRef]
   McMurdie, P.J.; Holmes, S. phyloseq: An R package for reproducible interactive analysis and graphics of microbiome census data.
- PLoS ONE 2013, 8, e61217. [CrossRef] [PubMed]
   Lozupone, C.: Lladser, M.E.: Knights, D.: Stombaugh, I.: Knight, R. UniFrac: An effective distance metric for microbial community
- Lozupone, C.; Lladser, M.E.; Knights, D.; Stombaugh, J.; Knight, R. UniFrac: An effective distance metric for microbial community comparison. *ISME J.* 2011, 5, 169–172. [CrossRef] [PubMed]
- Bray, J.R.; Curtis, J.T. An Ordination of the Upland Forest Communities of Southern Wisconsin. Ecol. Monogr. 1957, 27, 325–349. [CrossRef]
- 40. Jaccard, P. The Distribution of the Flora in the Alpine Zone. New Phytol. 1912, 11, 37–50. [CrossRef]
- Martini, R.; St-Pierre, M.F.; Wilson, B.N. French Canadian cross-cultural adaptation of the Developmental Coordination Disorder Questionnaire '07: DCDQ-FC. *Can. J. Occup. Ther.* 2011, 78, 318–327. [CrossRef]
- 42. Wechsler, D. Wechsler Intelligence Scale for Children-WISC-IV.; Psychological Corporation: San Antonio, TX, USA, 2003.
- Benjamini, Y.; Hochberg, Y. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. J. R. Stat. Soc. Ser. B 1995, 57, 289–300. [CrossRef]
- 44. Oksanen, J.; Kindt, R.; Legendre, P.; O'Hara, B.; Stevens, M.H.H.; Oksanen, M.J.; Suggests, M. The vegan package. *Community Ecol. Package* **2007**, *10*, 719.
- Mallick, H.; Rahnavard, A.; McIver, L.J.; Ma, S.; Zhang, Y.; Nguyen, L.H.; Tickle, T.L.; Weingart, G.; Ren, B.; Schwager, E.H.; et al. Multivariable association discovery in population-scale meta-omics studies. *PLOS Comput. Biol.* 2021, 17, e1009442. [CrossRef]
- Lozupone, C.A.; Stombaugh, J.I.; Gordon, J.I.; Jansson, J.K.; Knight, R. Diversity, stability and resilience of the human gut microbiota. *Nature* 2012, 489, 220–230. [CrossRef] [PubMed]
- Vatanen, T.; Franzosa, E.A.; Schwager, R.; Tripathi, S.; Arthur, T.D.; Vehik, K.; Lernmark, Å.; Hagopian, W.A.; Rewers, M.J.; She, J.X.; et al. The human gut microbiome in early-onset type 1 diabetes from the TEDDY study. *Nature* 2018, 562, 589–594. [CrossRef] [PubMed]
- Ling, Y.; Gong, T.; Zhang, J.; Gu, Q.; Gao, X.; Weng, X.; Liu, J.; Sun, J. Gut Microbiome Signatures Are Biomarkers for Cognitive Impairment in Patients With Ischemic Stroke. *Front. Aging Neurosci.* 2020, 12, 511562. [CrossRef]
- 49. Liu, P.; Wu, L.; Peng, G.; Han, Y.; Tang, R.; Ge, J.; Zhang, L.; Jia, L.; Yue, S.; Zhou, K.; et al. Altered microbiomes distinguish Alzheimer's disease from amnestic mild cognitive impairment and health in a Chinese cohort. *Brain Behav. Immun.* **2019**, *80*, 633–643. [CrossRef] [PubMed]
- Manderino, L.; Carroll, I.; Azcarate-Peril, M.A.; Rochette, A.; Heinberg, L.; Peat, C.; Steffen, K.; Mitchell, J.; Gunstad, J. Preliminary Evidence for an Association Between the Composition of the Gut Microbiome and Cognitive Function in Neurologically Healthy Older Adults. J. Int. Neuropsychol. Soc. 2017, 23, 700–705. [CrossRef]
- 51. Tamana, S.K.; Tun, H.M.; Konya, T.; Chari, R.S.; Field, C.J.; Guttman, D.S.; Becker, A.B.; Moraes, T.J.; Turvey, S.E.; Subbarao, P.; et al. Bacteroides-dominant gut microbiome of late infancy is associated with enhanced neurodevelopment. *Gut Microbes* 2021, 13, 1930875. [CrossRef]
- Shi, H.; Ge, X.; Ma, X.; Zheng, M.; Cui, X.; Pan, W.; Zheng, P.; Yang, X.; Zhang, P.; Hu, M.; et al. A fiber-deprived diet causes cognitive impairment and hippocampal microglia-mediated synaptic loss through the gut microbiota and metabolites. *Microbiome* 2021, *9*, 223. [CrossRef]
- 53. Shin, N.R.; Whon, T.W.; Bae, J.W. Proteobacteria: Microbial signature of dysbiosis in gut microbiota. *Trends Biotechnol.* **2015**, *33*, 496–503. [CrossRef] [PubMed]
- Jetten, M.J.A.; Gaj, S.; Ruiz-Aracama, A.; de Kok, T.M.; van Delft, J.H.M.; Lommen, A.; van Someren, E.P.; Jennen, D.G.J.; Claessen, S.M.; Peijnenburg, A.A.C.M.; et al. Omics analysis of low dose acetaminophen intake demonstrates novel response pathways in humans. *Toxicol. Appl. Pharmacol.* 2012, 259, 320–328. [CrossRef] [PubMed]
- Al Reef, T.; Ghanem, E. Caffeine: Well-known as psychotropic substance, but little as immunomodulator. *Immunobiology* 2018, 223, 818–825. [CrossRef]
- Mukhopadhya, I.; Hansen, R.; El-Omar, E.M.; Hold, G.L. IBD—what role do Proteobacteria play? *Nat. Rev. Gastroenterol. Hepatol.* 2012, 9, 219–230. [CrossRef] [PubMed]
- 57. Park, B.Y.; Lee, B.K. Use of meconium in perinatal epidemiology: Potential benefits and pitfalls. *Ann. Epidemiol.* **2014**, *24*, 878–881. [CrossRef] [PubMed]
- 58. Heaton, K.W.; Radvan, J.; Cripps, H.; Mountford, R.A.; Braddon, F.E.; Hughes, A.O. Defecation frequency and timing, and stool form in the general population: A prospective study. *Gut* **1992**, *33*, 818–824. [CrossRef] [PubMed]
- Nehlig, A. Interindividual Differences in Caffeine Metabolism and Factors Driving Caffeine Consumption. *Pharmacol. Rev.* 2018, 70, 384. [CrossRef] [PubMed]
- 60. Hines, R.N. The ontogeny of drug metabolism enzymes and implications for adverse drug events. *Pharmacol. Ther.* **2008**, *118*, 250–267. [CrossRef] [PubMed]
- Solazzo, G.; Wu, H.; Laue, H.E.; Brennan, K.; Knox, J.M.; Gillet, V.; Bovin, A.; Abdelouahab, N.; Posner, J.; Raffanello, E.; et al. The association between prenatal concentrations of polybrominated diphenyl ether and child cognitive and psychomotor function. *Environ Epidemiol.* 2021, 5, e156. [CrossRef]

- 62. Sussman, T.J.; Baker, B.H.; Wakhloo, A.J.; Gillet, V.; Abdelouahab, N.; Whittingstall, K.; Lepage, J.-F.; St-Cyr, L.; Boivin, A.; Gagnon, A.; et al. The relationship between persistent organic pollutants and Attention Deficit Hyperactivity Disorder phenotypes: Evidence from task-based neural activity in an observational study of a community sample of Canadian mother-child dyads. *Environ Res.* **2022**, *206*, 112593. [CrossRef]
- 63. Abdelouahab, N.; Langlois, M.F.; Lavoie, L.; Corbin, F.; Pasquier, J.C.; Takser, L. Maternal and cord-blood thyroid hormone levels and exposure to polybrominated diphenyl ethers and polychlorinated biphenyls during early pregnancy. *Am. J. Epidemiol.* **2013**, *178*, 701–713. [CrossRef]
- 64. Raven, J.; Baker, B.H.; Wakhloo, A.J.; Gillet, V.; Abdelouahab, N.; Whittingstall, K.; Lepage, J.-F.; St-Cyr, L.; Boivin, A.; Gagnon, A.; et al. Raven Progressive Matrices. In *Handbook of nonverbal assessment*; Kluwer Academic/Plenum Publishers: New York, NY, USA, 2003; pp. 223–237.