**REVIEW ARTICLE** 





# Polymer-based nano-therapies to combat COVID-19 related respiratory injury: progress, prospects, and challenges

#### Md Mohosin Rana 🝺

Biomedical Engineering Graduate Program, University of Calgary, Calgary, AB, Canada

#### ABSTRACT

The recent coronavirus disease-2019 (COVID-19) outbreak has increased at an alarming rate, representing a substantial cause of mortality worldwide. Respiratory injuries are major COVID-19 related complications, leading to poor lung circulation, tissue scarring, and airway obstruction. Despite an in-depth investigation of respiratory injury's molecular pathogenesis, effective treatments have yet to be developed. Moreover, early detection of viral infection is required to halt the disease-related long-term complications, including respiratory injuries. The currently employed detection technique (quantitative real-time polymerase chain reaction or qRT-PCR) failed to meet this need at some point because it is costly, time-consuming, and requires higher expertise and technical skills. Polymer-based nanobiosensing techniques can be employed to overcome these limitations. Polymeric nanomaterials have the potential for clinical applications due to their versatile features like low cytotoxicity, biodegradability, bioavailability, biocompatibility, and specific delivery at the targeted site of action. In recent years, innovative polymeric nanomedicine approaches have been developed to deliver therapeutic agents and support tissue growth for the inflamed organs, including the lung. This review highlights the most recent advances of polymer-based nanomedicine approaches in infectious disease diagnosis and treatments. This paper also focuses on the potential of novel nanomedicine techniques that may prove to be therapeutically efficient in fighting against COVID-19 related respiratory injuries.

#### **ARTICLE HISTORY**

Received 25 January 2021 Accepted 17 March 2021

#### **KEYWORDS**

Polymer; nanocarrier; tissue engineering; hydrogel; ARDS; COVID-19; respiratory injury; scaffold

#### **GRAPHICAL ABSTRACT**



CONTACT Md Mohosin Rana 🐼 mdmohosin.rana@ucalgary.ca 💽 Biomedical Engineering Graduate Program, Schulich School of Engineering, University of Calgary, 2500 University Drive, North West, Calgary, T2N 1N4, AB, Canada.

© 2021 Informa UK Limited, trading as Taylor & Francis Group

Schematic illustration of potential polymer-based nanomedicine strategies for the diagnosis and treatment of COVID-19 related respiratory injury.

Abbreviations: SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2: COVID-19: Coronavirus disease 2019: WHO: World health organization; ALI: Acute lung injury; ARDS: Acute respiratory distress syndrome; DNA: Deoxyribonucleic acid; RNA: Ribonucleic acid; ACE2: Angiotensin-converting enzyme 2; AECII: Alveolar epithelial type II cells; TMPRSS2: Transmembrane serine protease 2; VILI: Ventilator-induced lung injury; VV-ECMO: venovenous-Extracorporeal membrane oxygenation; IL: Interleukin; NF- $\kappa$ B: Nuclear factor kappa-light-chain-enhancer of activated B cells; PMMA: Poly(methyl methacrylate); PAA: Poly(amino acid); QCM: quartz crystal microbalance; COPD: Chronic obstructive pulmonary disease; POEGMA: Poly(Oligo(ethylene glycol) monomethyl ether methacrylate); DOTA: 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetrakisacetic acid); GMA: Glvcidvl methacrvlate: DPA: 2-(Diisopropylamino)ethyl methacrylate; TPE-4SH: Tetrakis[4-(2-mercaptoethoxy)phenyl]ethylene; pHLIP: pH-low insertion peptide; PVAS: Poly(vinyl alcohol) sulfate; AMPS: 2-acrylamido-2-methylpropane sodium sulfonate; PPCM: Polyphenylene carboxymethylene; PLGA: Poly(lactic-co-glycolic acid); PNIPAm: Poly(N-isopropylacrylamide); PLA: Polylactic acid; PICA: Poly(isobutyl cyanoacrylate); pHEMA: Poly(2-hydroxyethyl methacrylate); DDS: Drug delivery system; PCL: Poly(*e*-caprolactone); PEG: Poly(ethylene glycol); VEGFR: Vascular endothelial growth factor receptor; SAHA: Suberoylanilide hydroxamic acid; DOX: Doxorubicin; HCC: Hepatocellular carcinoma; COPD: Chronic obstructive pulmonary disease; PAA: Poly(acrylic acid); AZT-TP: Azidothymidine-triphosphate; CDDP: Cis-platinum or cis-diamminedichloroplatinum(II); HIV: Human immunodeficiency virus; PAMAM: Polyamidoamine; HSV: Herpes simplex virus; PVP: Polyvinylpyrrolidone; VZV: Varicella zoster virus; PVL-co-PAVL: Poly(valerolactone)-co-poly(allyl-δ-valerolactone); PVA: Poly(vinyl alcohol); PLL: Poly-L-lysine; PPI: Polypropylene polybenzyl isocyanate; gp120: Envelope glycoprotein GP120; CD4: Cluster of differentiation 4; PCD: Polyanionic carbosilane dendrimer; MPN-HANP: Metal-phenolic networkcoated hyaluronic acid nanoparticles; PDEAAm: Poly(N,N-diethylacrylamide); PEO: Poly(ethylene oxide); PPO: Poly(phenylene oxide); PVCL: Poly(N-vinyl caprolactam); PDLLA: Poly(D, L-lactic acid); HAG: Hyaluronic acid hydrogel; ESCs: Embryonic stem cells; iPSCs: Induced pluripotent stem cells; MSC: Messenchymal stem cells; siRNA: Small interfering RNA; APCs: Antigen-presenting cells; MHC: Major histocompatibility complex; VANs: Vaccine adjuvant nanoparticles; CSPG: Chondroitin sulphate proteoglycan; ASCs: Adipose-derived stem cells;

# Background

The outbreak of the novel  $\beta$ -coronavirus (severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2); family: *Coronaviridae*) responsible for Corona Virus Infectious Disease-2019 or COVID-19 is considered the worst crisis since World War II.<sup>[1]</sup> This pandemic's impact is frightening as the human race faces a critical situation with mandatory lockdowns with a long-lasting dent in the world economy. Critically

ill patients can develop acute lung injury (ALI) and acute respiratory distress syndrome (ARDS) (about 30-40%), which is associated with high mortality.<sup>[2]</sup> ARDS is a catastrophic disease condition characterized by noncardiogenic pulmonary edema, decrease pulmonary compliance and acute onset of hypoxic respiratory failure.<sup>[3,4]</sup> All of these complications can subsequently trigger a cascade of other severe injuries, including multiple organ failure. Unfortunately, to date, contemporary therapeutic strategies to treat ALI/ARDS have not been rewarding.<sup>[5,6]</sup> Due to the complex pathogenesis and nature of the infection, some therapeutic targets for the blockade of specific cytokines and chemokines have failed to show an optimistic outcome.<sup>[7-9]</sup> Currently, only protective lung ventilation strategies are the accepted gold standard for ARDS treatment.<sup>[10]</sup> However, targeted delivery of anti-viral drugs, proteins, peptides, and silencing RNAs is some potential therapies for ARDS treatments.<sup>[11]</sup> Despite these potential candidates' prospects, their delivery to the lung is a significant challenge for potential use in preventing viral infection and treating the respiratory injury.<sup>[12,13]</sup> A major hurdle in lung tissue engineering is developing lung-appropriate scaffold materials for soft tissue regeneration.<sup>[14]</sup>

Nano-therapies have become an attractive approach to overcome these limitations and the targeted delivery of potential therapeutic candidates to the lung.<sup>[15]</sup> Nanocarriers are mainly designed to increase the biodistribution of therapeutic agents to target organs, which results in improved efficacy with minimizing drug toxicity.<sup>[16]</sup> Nanomaterials can also be designed to support the production of bioengineered lung tissue in lung damage repair.<sup>[17]</sup> Polymer chemistry offers the capability to develop a wide range of nanocarriers with broad classes of functional groups to provide unique possibilities to bypass the conventional limitations of viral infection prevention and respiratory injury treatments. Among different drug delivery systems proposed for pulmonary or respiratory applications, biodegradable polymeric nanocarriers' use represents more potentiality.<sup>[18]</sup> Moreover, biocompatible polymeric hydrogel materials are considered one of the most suitable options to use as scaffolds because of its capability to provide lung-appropriate three-dimensional (3D) architecture with mechanical properties required to help the breathing and gas exchange processes.<sup>[19]</sup>

This review article will first focus on lung pathophysiology during the development of ARDS in COVID-19 infected patients. In the next section, a short synopsis of polymer-based nanobiosensing approaches for SARS-CoV-2 virus detection will be given. Later on, conventional treatments of respiratory injury and their shortfalls will be explored. In the later part of this review, advances in polymer-based nanotherapies will be emphasized to control respiratory complications and the treatment of ARDS in COVID-19 infected patients.

#### ARDS in COVID-19 patients: Pathological changes of lungs

The primary target organ of SARS-CoV-2 is the respiratory tract, particularly the upper airways and lungs.<sup>[20]</sup> The virus initially reaches alveoli, and the spike protein of the virus binds to angiotensin-converting enzyme 2 (ACE2) and enters alveolar epithelial type-II (AECII) cells *via* transmembrane protease serine 2 (TMPRSS2) catalysis.<sup>[21]</sup> These cells act as a reservoir of the virus. Pulmonary dendritic cells and

macrophages sense the presence of viral antigens, thereby initiate an innate immune response to discharge immense amounts of proinflammatory cytokines, including Tumor Necrosis Factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 (IL-1) and IL-6, and interferon (IFN- $\gamma$ ), resulting in a 'cytokine storm'.<sup>[22]</sup> Elevated levels of secreted cytokines induce the disruption of the alveolar-capillary membrane. Moreover, these cytokines also induce endothelial contraction, resulting in vasodilation and increased vascular permeability.<sup>[23]</sup> The disrupted alveolar-capillary membrane integrity allows the plasma leakage into the alveoli and the lungs interstitial spaces. Osmotic pressure gradient loss leads to a leaky barrier, and enhanced sensitivity to hydrostatic forces is considered key in diffuse edema formation.<sup>[24]</sup> The formation of protein-rich edema (also known as exudate) in the alveolar spaces and interstitium leads to alveolar flooding; such events make it difficult to breathe and triggers hypoxemia, one of the common symptoms of COVID-19 related respiratory injury.<sup>[23,25]</sup> The flooded interstitium provokes alveoli contraction. Moreover, alveoli collapsing is also induced by the decreased surfactant level due to the damaged AECII cells. Pulmonary macrophages also release more  $TNF-\alpha$  and interleukins that move towards Polymorpho-nuclear Neutrophils (PMNs) via chemotactic phenomena.<sup>[26]</sup> Interleukins and TNF- $\alpha$  trigger PMNs of the inflamed tissue to invade the alveoli and release reactive oxygen species (ROS), neutrophil extracellular traps (NETs), and proteases that damage different cells existing in the lung inflammatory microenvironment, including endothelial cells and alveolar epithelial cells.<sup>[23]</sup> These disease outcomes expedite inflammatory clusters containing fibrinoid materials and multinucleated giant cells, carbon dioxide diffusion and gas exchange disorders, and vascular congestion.<sup>[27,28]</sup> All these pathological changes lead to intractable hypoxemia and a consolidation process that enhances and worsens the alveolar collapsing. In normal physiological conditions, the inhaled oxygen reaches the alveoli to oxygenate the blood which then returns to the heart and then to the body's different cells.<sup>[23]</sup> Hence, the alveolarcapillary membrane in a healthy human is very thin to help exchange of gases. In ALI/ ARDS patients, the inflammatory process is widespread in both alveoli and interstitium, making the lungs stiff, and it becomes more difficult to inflate due to fluid and inflammation; thus, pulmonary failure occurs in infected patients.

# Polymer-based nanomedicine strategies for SARS-CoV-2 detection

Generally, methods for detecting viral infections rely on the detection of genetic materials or unique markers of the pathogen itself. In the case of COVID-19, the currently employed detection technique is quantitative real-time polymerase chain reaction (qRT-PCR), where the detection mainly relies on the presence of RNA of the SARS-CoV-2 virus.<sup>[29]</sup> Besides, some combined approaches including RT-PCR, chest X-ray, CT-scans, identification of some biomarkers and their level (e.g. procalcitonin: low level, IL-6 and 10: high concentrations, C-reactive protein: elevated level, and lymphocyte counts: low level) in blood have also been practiced for the diagnosis of COVID-19 infected patients.<sup>[30]</sup> These techniques are labor-intensive, time-consuming, and cannot be available in resource-limited settings. Moreover, some false positive/negative responses are also reported in many cases.<sup>[31]</sup> Contrarily, nanomaterial-based sensing strategies are suitable for viral detection with better sensitivity and

selectivity, authenticity, scalability, specificity, and minimal false positive/negative responses.<sup>[32]</sup> Among different nano- and biosensing approaches, molecularly imprinted polymers (MIPs) provide potential applicability and physicochemical robustness for detecting viral pathogens.<sup>[33]</sup> Fabrication of MIPs is done by molecular imprinting of novel functional polymers with pre-designed molecular target selectivity. MIP-based sensors with unique selectivity and sensitivity can be employed for the detection of the SARS-CoV-2 virus. Previously, the detection of the Influenza virus, HIV, Zika virus, Ebola virus, and Dengue virus have been performed successfully by selecting virus-specific biomarkers as the recognition element.<sup>[34-38]</sup> For example. Wangchareansak et al. developed a MIP sensing tool in combination with the quartz crystal microbalance (QCM) method for proof-of-concept of Influenza A virus subtypes (H5N1, H5N3, H1N1, H1N3, and H6N1) screening.<sup>[37]</sup> They used acrylamide, methacrylic acid, methylmethacrylate, and N-vinylpyrrolidone as the polymer system for imprinting. Influenza A virus surface antigens are composed of two glycoproteins, i.e. hemagglutinin (HA) and neuraminidase (NA), that play a vital role in the subtype classification.<sup>[33]</sup> In that study, MIP was made for each Influenza virus subtype whereby each MIP possessed a better recognition property towards its original viral template. Findings of that study suggest that both the H and N domains play crucial roles in the molecular recognition of MIP. This report has opened a new option to screen Influenza A virus subtypes in unknown samples with detection limits of up to 105 particles/mL. Similarly, Tai et al. fabricated MIP-based film in the presence of a pentadecapeptide (15-mer peptide: linear epitope of the non-structural (NS1) protein of Dengue virus) onto a QCM chip.<sup>[38]</sup> Such epitope-mediated imprinting resulted in an enhanced polymer affinity toward the virus protein. They enhanced the binding effect using a monoclonal antibody to form a sandwich with the MIP-NS1 protein complex on the chip. These studies indicated that the detection of SARS-CoV-2 could be done by selecting a polymer belonging to the acrylic group (e.g. acrylamide, acrylic acid, methyl acrylate, ethyl acrylate, and methyl methacrylate) and applying the CoVspecific biomarker as the recognition element.<sup>[39]</sup> Only polymer-based biosensing approaches have been summarized here with their potential application to detect the SARS-CoV-2 virus. A more detailed discussion of other nanomaterial-based biosensors for CoV detection can be found elsewhere.<sup>[40]</sup>

#### Current therapeutic options to treat ARDS: Limitations and challenges

Despite the improved molecular understanding of the infection, there is still no specific treatment for ARDS. Some common therapeutic strategies include protective mechanical ventilation, prone-positioning ventilation, fluid-conservative strategy, and other supportive care.<sup>[41]</sup> These strategies have some limitations like the development of ventilation-induced lung injury (VILI), exacerbation of lung injury, and stimulating inflammatory reactions.<sup>[42]</sup> Veno-venous extracorporeal membrane oxygenation or VV-ECMO is an effective life-saving intervention to treat ARDS. However, the routine application of ECMO as salvage therapy in severe ARDS patients is a matter of debate.<sup>[43,44]</sup> Broad-spectrum antiviral therapy could be an option to treat COVID-19 related ARDS patients. However, a low success rate due to ongoing inflammatory response, the emergence of rapid mutation of SARS-CoV-2 strains, and antibiotic administration's timing make such pharmacologic treatments completely ineffective.<sup>[45]</sup>

Short-term use of a neuromuscular blockade in the early stage of moderate to severe ARDS improved survival rates by reducing epithelial and endothelial injury markers and systemic inflammations.<sup>[46]</sup> However, such pharmacological therapies have failed to show long-term benefit.<sup>[46]</sup> Moreover, a wide range of conventional anti-inflammatory drugs have been proposed to treat ARDS because these drugs work by regulating inflammatory signalling targets (Losartan: Angiotensin II receptor blocker, Tocilizumab, Siltuximab: Interleukin-6 or IL-6 inhibitor, Baricitinib: Janus kinase or JAK-1/2 inhibitor, Anakinra: IL-1 inhibitor, Polyphenolic compounds: Kinase inhibitors, and so on) within the inflammatory systems in the body.<sup>[8,47-49]</sup> Despite the potentiality of these proposed treatments, developing these therapeutics is lagging behind the need for them due to extensive research and clinical trials to prove their efficacy and safety.<sup>[50]</sup> Recently, Russel et al. investigated the efficacy of corticosteroid treatment for COVID-19 related lung injury.<sup>[51]</sup> This study suggested that dexamethasone is not effective enough to treat lung injury. Hence, to circumvent lifethreatening complications due to ARDS or other infection-related respiratory injuries, alternative therapeutic strategies are required on urgent basis to eradicate respiratory injuries and induce damaged lung tissue repair. In this regard, polymer-based nanotherapies have drawn the attention recently to overcome all the shortfalls of conventional treatments.

#### Polymer-based nano-therapies for respiratory injury treatment

#### Polymeric nanocarriers and drug delivery systems (DDS)

Polymers are soft materials that have been proposed and used in the preparation of nanomedicine. Natural and synthetic polymers with both hydrophilicity and hydrophobicity are used for such purpose. Proteins like albumin, gelatin, lectin, and polysaccharides such as cellulose, dextran, chitosan, and alginates are natural hydrophilic polymers that have been used as nanomedicine.<sup>[52]</sup> Polymethacrylate (PMMA), poly(lactic-co-glycolic acid) (PLGA), polystyrene, poly(N-isopropylacrylamide) (PNIPAm), polylactic acid (PLA), poly(isobutyl cyanoacrylate) (PICA), poly(hexyl cyanoacrylate) are some synthetic polymers with hydrophobic property for nanomedicine formulation.<sup>[53,54]</sup> Various polymeric nanocarriers using these materials have been designed for the controlled release of drugs.<sup>[55,56]</sup> To reduce the nonspecific interaction with healthy cells and serum proteins and avoid uptake by phagocytosis, surface modification with several functional groups onto polymeric nanocarriers has been designed over the past decades.<sup>[57]</sup> On the other hand, some polymeric materials are sensitive to environmental stimuli like temperature (e.g. PNIPAm) and pH (e.g. pHEMA).<sup>[58-60]</sup> Thereby, nanomedicine strategy from these stimuli-responsive materials can help prevent the drug or biomolecules degradation before reaching the target site of action which subsequently helps decrease the toxic effects of nonspecific sites and increase the bioavailability of the delivered therapeutic components.<sup>[58]</sup> The availability of stimuli-responsive polymer materials and flexibility of nanocarrier fabrication techniques with the application of target ligands on the nanocarrier surface

enables the design of biomolecule loaded nanocarriers to boost antiviral effects. There are nearly 90 antiviral drug candidates have been approved for the treatment of emerging viruses.<sup>[61]</sup> The administration of these drugs is often accompanied by side effects due to their accumulation in the body's off-target site. Some drugs require high concentrations in the body to become effective against the virus, causing toxic effects to host cells with other side effects. For example, ribavirin is associated with hemolytic anemia.<sup>[62]</sup> Most of the approved antiviral drugs are poorly water-soluble as well, which hinders their successful use.<sup>[52]</sup> Hence, the polymeric nanomedicine strategies can be a potential solution for delivering a broad range of active moieties like antiviral biologics and nucleic acids to the target site for the COVID-19 related respiratory injury treatments.

#### **Polymeric nanoparticles**

Polymeric nanoparticles are the primary type of nanocarriers that can change the pharmacokinetic parameters of the encapsulated drug compound, and controlled drug release helps to reduce required drug concentration for biological activity. Polymeric nanoparticles can be fabricated by a broad range of fabrication techniques like solvent evaporation, ionic gelation, spray-drying, living- or free-radical polymerization, nanoprecipitation, and polymer dispersion technique.<sup>[52]</sup> Nanoparticles as nanocarriers have some advantages over other drug delivery systems (DDS) like low toxicity, site-specific delivery and degradation, better cellular uptake, controlled release of incorporated drug molecules. They can be used as theranostics in antiviral therapy.<sup>[63]</sup> Nanoparticles with 100–500 nm in size, which can incorporate the drug molecules inside its core, are known as nanocapsules. In nanocapsule system, the targeted drug is infused in the inner core, surrounded by the polymeric shell. High drug loading, controlled release profile, and target specific delivery are vital features of nanocapsules.<sup>[64]</sup> For example, nanocapsule consisting of poly(isobutyl cyanoacrylate) core and polyethyleneimine shell has been designed to deliver azidothymidinetriphosphate (AZT-TP) into the cytoplasm directly.<sup>[65]</sup> Contrarily, nanoparticles with 10-200 nm in size are known as nanospheres and the drug molecules can be adsorbed onto its surface or embedded in the matrix of the particles.<sup>[66]</sup> This type of nanocarrier can resist the drug molecules from unwanted degradation, and rapid drug clearance is observed because of its smaller size. In a study, chitosan nanospheres were developed with an average size of 200 nm for HSV-1 and HSV-2 treatment.<sup>[67]</sup> This polymeric nanosphere loaded with acyclovir showed better permeation and higher potency against the viral treatment compared to free acyclovir itself. This kind of smaller nanocarrier also offers site-specific drug delivery and controlled drug release profile.

Nanoparticles developed by polymeric materials such as PLGA, PLA, poly(ethylene glycol)-poly(*ɛ*-caprolactone) (PEG-PCL), PLA-PEG, and some others have been studied exclusively as nanocarriers for the systematic delivery of drug molecules and biomolecules for the antiviral and other disease treatment (Figure 1).<sup>[68–72]</sup> Previously, Li et al. developed a unique cocktail therapeutic strategy containing biodegradable polymeric nanoparticles for antiviral treatment.<sup>[73]</sup> They developed this PEG-PLA-based cocktail nanoparticles to encapsulate HIV-1 entry inhibitor and conjugate with reverse transcriptase inhibitor, resulting in strong virucidal effects against HIV-1.



**Figure 1.** Different types of polymeric nanocarrier systems. (A) Different nanoparticles used to deliver antioxidants in chronic obstructive pulmonary disease (COPD) treatment. Reprinted from Xu et al.<sup>[68]</sup> Owing to some advantages of nanocarrier systems like small size, high stability, targeted deposition, sustained release, biodegradation, and reduced dosing frequency, they can be used to integrate and deliver hydrophobic and hydrophilic drugs in COPD treatment. Some of the promising novel DDS for COPD treatment include polymeric nanoparticles, micelles, dendrimers, microspheres and microparticles, nanoemulsion, lipid nanoparticles, and liposomes. Copyright 2020 BioMed Central. (B) PEG-PAA block copolymer endowed micelles with on-demand functionalities and specific targetability. Reprinted from Nishiyama et al.<sup>[69]</sup> Polymeric micelles can be engineered through block copolymers' self-assembly with a controllable size range of 10–100 nm. These micelles have a core-shell structure where biocompatible PEG shell surrounds the drug-loaded core. Copyright 2016 John Wiley & Sons Australia.

Polymeric nanoparticles can be administered in a systemic route (e.g. dermal, oral, intravenous, and so on) or directly into the lung *via* inhalation or intranasal route.<sup>[74]</sup> Nanoparticles containing antiviral drugs or small interfering RNA (siRNA) could be very effective if deliver through nasal epithelia and lungs in order to attack viruses that infect the respiratory tract, like Influenza viruses, Respiratory syncytial virus, and Rhinoviruses. Earlier, Jamali et al. developed a siRNA-chitosan nanoparticulate therapy that effectively target viral nucleoprotein to reduce virus infections.<sup>[75]</sup> They reported that the intranasal administration of this nanoparticles enhanced therapeutic effect on mice attacked with a lethal dose of Influenza virus, revealing *in vivo* antiviral activity of such nanoparticles. It is important to note that majority of the studies performed so far to assess the efficiency of polymer nanoparticles as DDS and recommendation are mostly based on preclinical data performed on lab animals, and these are not ready yet to administer in humans.<sup>[76]</sup>

Nevertheless, these investigations provide some promising hope to develop efficient polymeric nanoparticle-based nano-therapies to deliver antiviral drug molecules and immunomodulate cytokine storms in COVID-19 infected patients with respiratory complications.

#### **Polymeric micelles**

Polymeric micelles are amphiphilic block copolymers consisting of a hydrophobic core incorporate water-insoluble drugs and a hydrophilic shell that acts as a barrier to protect the drug. This type of nanocarrier structure allows higher drug loading and

minimizes the premature drug release in the off-target site.<sup>[77]</sup> Polymeric micelles can be used as a targeted drug delivery vehicle by surface modification with specific ligands. In a recent study, *ɛ*-caprolactone has been used as a hydrophobic core to encapsulate silibinin and further grafted with methoxy PEG (mPEG) to form amphiphilic block copolymeric micelles.<sup>[78]</sup> Such micelles are bioengineered auto-assembly copolymers formed in a liquid medium. Due to the micelle's hydrophilic outer laver, the whole micelle remains stable and biocompatible with tissues and blood. A recent study reported the development of polymeric micelles of isoniazid and rifampicin using the di-block polymer, PEG and PLA.<sup>[79]</sup> This formulated drug-loaded polymeric miceller delivery was reported to enhance the efficacy by reducing minimum inhibitory concentration (MIC) against Mycobacterium tuberculosis up to 8-fold. In a different study, polymeric micelles of isoniazid and rifampicin using ethylene oxidepropylene oxide tri-block copolymers, Pluronic® was developed.<sup>[80]</sup> A multifunctional PLA-b-PEG copolymer modified methyl-b-neuraminic acid (mNA) has been prepared as drug delivery micelles to treat Influenza virus infection.<sup>[81]</sup> It has been found that amantadine loaded in these micelles inhibit hemagglutination by binding to the hemagglutinin of Influenza viruses and efficiently alleviating viral infection.

The limited intracellular intake of antiviral drugs due to limited aqueous solubility is one of the major drawbacks to the successful treatment of respiratory illness in COVID-19 infected patients. Hence, due to the amphiphilic auto-assembly nature of polymeric micelles, these nanocarrier systems can be served as vehicles for delivering insoluble hydrophobic antiviral and anti-inflammatory therapeutics for the COVID-19 related ARDS treatments.

#### **Polymeric conjugates**

Conjugation of polymer with the targeted drug can be obtained by covalent bonding between a polymer and the therapeutic drug molecules. Polymeric conjugates are another potential delivery vehicle candidate for the treatment of COVID-19 because of several advantages like lower dosages of the required drugs that cause fewer side effects, decreasing the likelihood of drug resistance, and useful for the delivery of multiple drugs with different physicochemical properties.<sup>[82]</sup> A wide variety of negatively charged polymers and glycosaminoglycans like heparin, chondroitin sulphate, keratan sulphate, dermatan sulphate, and heparan sulphate have the potentiality to bind to the HIV envelope and resist the entry of viral particles inside the host cells.<sup>[83]</sup> Conjugation of azidothymidine (AZT) and ribavirin with synthetic polymers like methacrylates have been investigated broadly.<sup>[84]</sup> Results from these studies have shown greater antiviral potency with decreased toxicity. Conjugation of AZT with some natural polymers such as chitosan and dextrin via succinic ester linkage has shown longer plasma half-life and high drug loading capacity. In another study, conjugation of antiviral drug compound stavudine with chitosan by phosphoramide linkage has shown beneficial results in viral infection treatments.<sup>[84]</sup> Despite the benefits of polymer-drug conjugates for a wide range of treatments, optimizing the drug conjugation rate with polymers is still an uphill task. Some optimization challenges include controlling the interaction between two or more drugs and the release profile of individual drug from the conjugated cargo system.<sup>[85]</sup> Hence, it is recommended that high-throughput screening profiles are needed to understand the biological interactions and find out if there is any synergism present or not (Table 1).

#### Other polymeric carriers to treat respiratory injury

Dendrimers are radially symmetric, highly branched, monodisperse, and homogeneous, nanoparticles with greater ability to attach multiple functional groups on their surface. Like polymeric micelles, dendrimers are made up of a central core, an inner shell composed of repeating units of building blocks, and an outer shell with many functional groups attached. Such configuration of dendrimers enables them to encapsulate non-water soluble, hydrophobic therapeutic agents in their core and specific surface functional groups to allow them to interact with the target biological site to deliver encapsulated agents. Such nanocarriers can be used as theranostics due to their outstanding ability to uptake by cells, longer circulation times, and improved stability and solubility in targeted drug delivery. Some of the commercially available dendrimers are poly(propyleneimine) (PPI), polyamidoamine (PAMAM), and poly-Llysine (PLL).<sup>[112]</sup> Dendrimers' strong ability to interact with the viral cell surface and enhance antiviral activities can be used to treat viral infection in the host, such as HIV and Influenza virus infections.<sup>[113]</sup> PLL-based dendrimers with anionic naphthalene disulphonate surface have been designed to block the entry of HIV viruses by binding to the gp120 protein (viral envelope protein), thereby preventing the formation of CD4-gp120 complex.<sup>[114]</sup> Poly(phosphor-hydrazone) is a biodegradable dendrimer with end phosphoric acid functionalities, which have been proposed for anti-HIV activity.<sup>[115]</sup> Polyanionic carbosilane dendrimers (PCDs) have been designed, and combination therapy of PCDs with tenofovir and maraviroc has shown enhanced efficacy against HIV and minimizes the emergence of multidrug-resistant HIV mutants.<sup>[116]</sup> It has also been reported that some nanocarriers' surface properties have shown promising result in binding ACE2 receptor.<sup>[117]</sup> The cationic PAMAM nanoparticles have the property to bind to the ACE2 receptor, blocking angiotensin's cleavage, causing ARDS.<sup>[118]</sup>

A variety of sulfated polymers, including sulfated derivatives of PVA, polystyrene, poly(vinylsulfonic acid), poly(anethole sulfonate), and poly(2-acrylamido-2-methyl-1-propanesulfonic acid), have been reported earlier to inhibit HIV replication.<sup>[119]</sup> Sulfated polymers like poly(vinyl alcohol) sulfate (PVAS) have also proved their efficacy to inhibit HSV, Cytomegalovirus, Respiratory syncytial virus, Vesicular stomatitis virus, and Retroviruses.<sup>[120]</sup> Previously, Danial et al. investigated combining the antiviral lamivudine with a terpolymer synthesized from sulfonated side chains (2-acrylamido-2-methylpropane sodium sulfonate (AMPS)).<sup>[121]</sup> They found that at higher concentrations, the homopolymer poly(AMPS) combined with lamivudine exhibited nearly full inhibition against HIV infection. Polyphenylene carboxymethylene (PPCM) is a broad-spectrum antiviral polymer that binds to the viral envelope glycoproteins V3 loop and interferes with the interaction between gp120 and CD4<sup>+</sup>T cells.<sup>[122]</sup>

Table 1. Polymeric I	nanocarrier systems in a	anti-microbial drug delivery a	pplications.		
Nanocarrier system	Carrier material	Drug target	Diseases	Major findings	Ref.
Nanoparticle	PLGA + transferrin	Nevirapine	NIH :	Increased uptake in brain microvascular endothelial cells	[86]
	PLGA	Lamivudine	Herpes	High targeting ability	[87]
	PLGA	Combination therapy	HIV	Efficient drug entrapment (>/9%) Indibition of HIV infortion	88
		(юршаун ⊤ шолаун ⊤ еfavirenz)		Ffficient untake in nonimmune cells	
				High nuclear and membrane drug levels in infected cells	
	Chitosan	Lamivudine	AIN	Brain targeting	[89]
	Alginate	Pyrazinamide, ethambutol,	Tuberculosis	Higher drug payload,	[06]
		isoniazid, Rifampicin		Enhanced therapeutic efficacy,	
				Improved pharmacokinetic profile	
	PLGA	Rifampicin, Pyrazinamide	Tuberculosis	Higher drug payload	[11]
				Higher efficacy	
	PLGA	Efavirenz, Nevirapine	HIV	Increased permeability,	[92]
				Enhanced blood brain barrier (BBB) interacting ability	
	PLGA	Elvitegravir	AIN	Improved intracellular uptake	[63]
	CAB	Nevirapine	HIV	Enhanced efficacy	[94]
Nanosphere	PLA	Arjunglucoside	Leishmaniasis	Reduced toxicity	[95]
	PCL	Amphotericin B	Candidiasis	Reduced accumulation into the kidney	[96]
				Decreased toxicity	
	PEG-PLA	Acyclovir	Ocular HSV	Sustained release, no eye inflammation	[22]
	PEG-PECA	Acyclovir	Ocular HSV	Significant drug level increase in aqueous humor	[98]
Nanocapsule	PEG-PLA	Halofantrine	Malaria	Prolonged circulation time	[66]
	PCL	Indomethacin	Arthritis (anti-	Increased corneal penetration	[100]
			inflammatory)		
Micelle	Stearic acid	Lamivudine	Hepatitis B	Higher cellular uptake in infected hepatoblastoma cells,	[101]
	+ chitosan			High drug loading	
	PEG	Pyrazinamide	Tuberculosis	Significant therapeutic efficacy than original drug	[102]
	+ PAA				
	PCL	Acyclovir	HSV	Efficient drug uptake and delivery,	[103]
	+			Nontoxic in nature	
	PEG				
	PAMAM	Acyclovir	HSV, Shingles	Enhanced mucoadhesion	[104]
Dendrimer	PAMAM	Sulfamethoxazole	Influenza	Increased antimicrobial activity, Suctained drug release	[105]
	PEGylated lysine	Chloroguine phosphate	Malaria	Justice and stability,	[106]
				Enhanced drug circulation time	
					ontinued)

$\overline{\mathbf{D}}$	
ā.	
≝	
_	
_	
. <del></del>	
-	
-	
-	
0	
()	
-	
•	
÷.	
<del>.</del> -	
le 1.	
ole 1.	
ble 1.	

Table 1. Continued.					
Nanocarrier system	Carrier material	Drug target	Diseases	Major findings	Ref.
	PAMAM	Nadifloxacin, Prulifloxacin	Anti-bacterial	Improved water solubility	[107]
Nano-emulsion	PVP, methyl cellulose	Darunavir, Nelfinavir, Atazanavir	HIV, HSV, VZV, Shingles	Improved therapeutic efficacy	[108]
Nano-sponge	PVA, ethyl cellulose, PVL-co-PAVL	Acyclovir	HSV 2	Increased stability and solubility, formulation flexibility	[109]
Nano-dispersion	PVP, PEG, PVA	Efavirenz	HIV	Extended availability of drug, Increased solubility	[110]
Nanocrystal	PVA, PVP, cellulose derivative	Nevirapine	HIV	Improved bioavailability, Facilitates phagocytosis and targets the spleen	[111]



**Figure 2.** Stimuli-responsive polymer-based nanocarrier systems. (A) Thermo-responsive PNIPAmcellulose nanocrystals (CNC) hydrogels for wound dressing application. The thermal stability of this hybrid hydrogel decreased while the rheological property increased with increasing CNC content. This smart hydrogel showed good drug-loading ability at room temperature and sustained drugrelease at 37 °C. Reprinted from Zubik et al.<sup>[124]</sup> Copyright 2017 MDPI. (B) Core cross-linked micelles (CCL) fabricated with fluorescence and magnetic resonance (MR) dual imaging modalities from tetrakis[4-(2-mercaptoethoxy)phenyl]ethylene (TPE-4SH) fluorophores, and DOTA(Gd)-POEGMA— P(DPA-co-GMA) and benzaldehyde-POEGMA-b-P(DPA-co-GMA) deblock copolymers *via* co-assembly and click chemistry. Further, pH-responsive CCL micelles were fabricated with pH-low insertion peptide (pHLIP) through Schiff base linkage formation. Under a neutral pH state, pHLIP of the micelles formed coil state while turned to  $\alpha$ -helical conformation under acidic pH conditions. Such transition helps to enhance cellular internationalization and allows for imaging in live cells. Reprinted from Tian et al.<sup>[125]</sup> Copyright 2016 MDPI.

Polymer hydrogels are 3D crosslinked networks of hydrophilic polymer chains that can swell and hold a bulk amount of water while maintaining the polymers' structure. The crosslinking structure provides the physical integrity of the hydrogels and required mechanical strength as well. Hydrogels can be synthesized using both natural and synthetic biodegradable polymers. The hydrogels' high-water content can possess similarity (e.g. the higher degree of flexibility, biocompatibility) to that of normal tissue.<sup>[123]</sup> These advanced properties of hydrogels make them potential candidates for nanomedicine applications. Stimuli-responsive hydrogels are another potential nano-carrier system for the specific delivery of therapeutic agents (Figure 2).<sup>[124,126–128]</sup>

One example of such hydrogel-based delivery systems is thermoresponsive injectable hydrogels. Hydrogels of this group can show phase-transition behavior below and above the physiological temperature.<sup>[129]</sup> Thermoresponsive hydrogels including PNIPAm, poly(N,N-diethylacrylamide) (PDEAAm), poly(ethylene oxide)/poly(propylene oxide) (PEO/PPO), poly(N-isopropylmethacrylamide) (PNIPAm), poly(N-vinyl caprolactam) (PVCL), PEG-based biodegradable polyester copolymers have been designed and developed as drug delivery systems.<sup>[129,130]</sup>

Polymer-based cellular nanosponges are another novel nanomedicine strategy to combat COVID-19 related infections. Zhang et al. recently prepared two cellular nanosponges (i.e. Epithelial-nanosponge (NS) and Macrophage-nanosponge or M $\Phi$ -NS) by coating cell membranes of human lung epithelial cells and macrophages onto polymeric nanoparticle cores made from biodegradable PLGA.<sup>[131]</sup> These nanosponges carry the same identified and unidentified protein receptors required by

SARS-CoV-2 for cellular entry. The results obtained after incubation of these nanosponges indicate that both have the comparable ability to inhibit the viral infectivity of SARS-CoV-2. Moreover, M $\Phi$ -NS can neutralize the viral activity early to reduce the viral load in the host and the later stage of the infection. It is well established that macrophages play a significant role in the pathogenesis of respiratory virus infection.<sup>[131]</sup> Hence, M $\Phi$ -NS may play significant roles in treating inflammatory viral infections such as SARS-CoV-2 and related complications.

It is important to note that advances in polymer chemistry along with pathophysiological changes in human due to COVID-19 infection can enable us to develop smart biodegradable polymeric delivery systems with the great potential for controlled delivery of immunomodulatory therapeutic agents to treat respiratory injuries in critically ill patients.

# Polymer-based nanomedicine strategies for COVID-19 vaccine delivery

From the history of vaccine development, it is well established that vaccination is one of the most effective strategies to prevent and control the spread of infectious diseases, where naturally developed immunity induces protective long-term immune memory in patients.<sup>[132]</sup> In general, vaccines introduce specific viral antigens on the cell surface of antigen-presenting cells (APCs), particularly dendritic cells, embodied in the major histocompatibility complex (MHC) I and II.<sup>[133]</sup> Such an event triggers the adaptive immune system by recognizing these antigens as invaders and induces antibodies production or T cells to eliminate these unwanted invaders. Consequently, memory B cells in the body develop virus-specific antibodies on its cell surface, which triggers a fast immune response to clear the similar viral infection in the future. There are three different generations of vaccine formulations currently used to trigger immune responses against infection, including live attenuated (whole inactivated pathogen) vaccines or first-generation vaccines, recombinant subunit vaccines (second-generation), and RNA/DNA vaccines or third-generation vaccines.<sup>[134,135]</sup> Since the outbreak of COVID-19, several different vaccine candidates have been developed and reached clinical phases due to a high urgency to halt the pandemic.<sup>[136]</sup>

In the novel vaccine development for COVID-19, some studies have indicated that the viral S protein or receptor-binding domain (RBD) and N-terminal domain of S protein can be an excellent target for vaccine preparation in order to enhance the immunological response.<sup>[137]</sup> Different mRNA, DNA, and non-replicating adenovirus vector-based vaccines are under clinical trial to check their efficacy in COVID-19 treatment. The University of Oxford, in collaboration with AstraZeneca, developed a vaccine (AZD1222; formerly known as ChAdOX1) composed of a non-replicating adenovirus vector and able to replicate the S protein of SARS-CoV-2.<sup>[138]</sup> Some recently developed mRNA vaccine candidates are Moderna's mRNA-1273 (NCT04405076), Arcturus Therapeutics' LUNAR-COV19, BioNTech and Pfizer's BNT162a1, b1, b2, and c2, Globe Biotech's BANCOVID, and an CVnCoV developed by CureVac.<sup>[139–143]</sup> These mRNA vaccine candidates target the S protein (or a specific region of S protein) of the SARS-CoV-2 cell surface. On the other hand, vaccine

candidates developed by Inovio Pharmaceuticals (INO-4800), Genexine's GX-19, and Zydus Cadila's ZyCoV-D are some DNA vaccines targeting viral S protein.<sup>[144,145]</sup> Epivax is a cocktail vaccine made up of antigens (i.e. non-structural proteins and nucleoproteins) other than S protein to provide partial protection against the virus.<sup>[146]</sup> Gamaleva Research institute developed Gam-COVID-Vac, and CanSino Biologics developed Ad5-nCoV to fight against SARS-CoV-2.<sup>[147]</sup> Johnson & Johnson also developed a vaccine candidate (Ad26.COV2.S), a recombinant, replicationincompetent adenovirus serotype 26 (Ad26) vector encoding a stabilized full-length SARS-CoV-2S protein.<sup>[148]</sup> Previously, this Ad26 vector was approved by the European Medicines Agency for the Respiratory syncytial virus, Zika virus, and Ebola virus.<sup>[148,149]</sup> Vaccine made of Ad26 vector is considered safe and highly immunogenic.<sup>[149]</sup> A couple of vaccine candidates developed by Sinopharm in collaboration with the Beijing Institute of Biological Products are currently in phase III clinical trial<sup>[146]</sup>. Some other protein-based vaccines, including COVAX-19 by Vaxine PTY Ltd. and NVX-CoV2373 by Novavax, are under clinical trials to evaluate their efficacy against COVID-19.<sup>[150]</sup> So far, vaccine candidates developed by Pfizer-BioNTech, Moderna, Oxford-AstraZeneca, Johnson & Johnson, CanSino, Sinopharm, Gamaleya, and Sinovac have been approved by health regulatory agencies throughout the world for early and emergency use.<sup>[151]</sup>

In COVID-19 vaccine research, some significant challenges are finding practical approaches to stimulate both the T cell and B cell immunity against the virus and developing precise next-generation vaccine for patients with compromised immunity.<sup>[152]</sup> Besides, poor immunogenicity and premature degradation of the antigens in harsh in vivo conditions and failing to reach the target sites of DNA and RNA vaccines are some other limitations that result in the weak immune response.<sup>[153]</sup> Therefore, it is crucial to develop smart strategies to deliver the COVID-19 vaccine more protectively, providing enduring protection with enhanced patients' immunity. Nanocarrier-based vaccine delivery program can be an option to deliver vaccines to induce complimenting immunomodulatory effects. Polymeric nanocarriers can be used to deliver antigens without premature degradation and potential side-effects, which allow directed targeting of the vaccine towards APCs.<sup>[117]</sup> Additionally, with the growing interest in RNA and DNA vaccines to fight against coronavirus, combining them with nanoscale cargo devices will be an effective approach to overcome all the limitations mentioned above. It has been already reported that the nanocarrierbased strategy can be an effective approach to deliver small interfering RNA (siRNA) for the treatment of malignancies, infections, and autoimmune diseases.<sup>[154]</sup>

The vaccine delivery using nanocarrier systems can be done either by encapsulating the antigen or DNA/RNA within the nanocarrier or by attaching antigens on the nanocarrier surface.<sup>[155]</sup> The antigen encapsulated nanocarrier-based vaccine delivery strategy shows the potential efficacy that prolongs the antigen exposure towards the immune cells.<sup>[117]</sup> Antigens with amphoteric nature are suitable candidates to be adsorbed on polymeric nanocarrier surfaces like chitosan and dextran sulphate-based nanoparticles.<sup>[156]</sup> In such cases, nanocarriers are predesigned with stimuli-responsive properties like pH, temperature, and ionic strength to release antigens from the carrier surface inside patients' bodies. PLGA nanoparticles are suitable candidates for

encapsulating the antigens within the nanocarrier to provide extended and controlled biological release.<sup>[157]</sup> Previously, these PLGA nanoparticles have shown efficacy preclinically in carrying antigen vaccines like HBsAg, Malaria antigens, Bacillus anthracis spores to generate extended cellular and humoral immune response.<sup>[158,159]</sup> For example, Galloway et al. developed a PLGA nanoparticle-based Particle Replication in Non-wetting Templates (PRINT<sup>®</sup>) technology to deliver influenza vaccine antigens.<sup>[160]</sup> This vaccine delivery approach requires low amount of vaccine antigens to induce immune responses. Naked mRNAs are prone to degradation by extracellular RNases; therefore, it is essential to deliver them in a protective way to prolong their efficacy.<sup>[161]</sup> Some of the polymeric nanocarriers are under investigation to use for the delivery of mRNA-based vaccines. These mRNA nanocarriers include PEG-lipid functionalized dendrimers (200 nm), polyethyleneimine (PEI) nanoparticles (100 - 300 nm), chitosan (cationic) nanoparticles (300 - 600 nm), and protamine (cationic peptide) nanoliposomes (~100 nm).<sup>[156]</sup> Like mRNAs, naked DNAs also experience premature degradation by nucleases and require protective delivery to boost immune response.<sup>[156]</sup> Polymeric nanocarriers with DNA encapsulation helps to prevent premature inactivation of DNA and provide controlled release.<sup>[156]</sup> Composite PLGA nanocarriers with cationic glycol-chitosan or PEIs are under investigation to improve DNA loading efficiency, systemic protection, and controlled release.<sup>[157]</sup> Surface electroporation of DNA coated-PLGA nanoparticles has been developed recently to improve DNA/RNA delivery across the cells and nuclear membrane.<sup>[162]</sup> This electroporation strategy has shown efficient delivery of DNA to elicit T cell and B cell response in pigs.<sup>[162]</sup> This kind of portable electroporation approach is, therefore, become an attractive option in COVID-19 vaccine research. Vaccine adjuvant nanoparticles (VANs) are designed to tackle the shortfalls of conventional molecular adjuvant delivery and improve the efficacy and safety of the generated immune response.<sup>[163]</sup> Vaccines and vaccine adjuvants which induce T helper type 1 or Th1biased immune responses are highly preferrable to fight against COVID-19 or related viral infections.<sup>[164]</sup> It has been reported that five protein subunit vaccine candidates using a combination of antigen and adjuvant are in the pipeline of preclinical COVID-19 vaccine candidates.<sup>[165]</sup> Recently, a SARS-CoV-2 recombinant full-length S protein nanoparticle vaccine combined with the saponin-based Matrix-M<sup>TM</sup> adjuvant has been developed by Novavax.<sup>[150]</sup> This vaccine candidate is currently in phase I/II clinical trial (NCT04368988), and it has been demonstrated that the adjuvant triggers the entry of APCs into the injection site and induces the antigen presentation in local lymph nodes, resulting in enhanced immunological response.<sup>[166]</sup> Some in vivo studies of PLGA and calcium phosphate nanoparticles co-encapsulating both adjuvants and antigens have shown improved efficacy by inducing antigen uptake, APC activation, and higher antibody titers.<sup>[167]</sup> VANs like PLGA are also used to codeliver immunoregulatory drugs or self-antigens as adjuvants to trigger antigenspecific peripheral tolerance of autoreactive T cells and obstruct any possible autoimmune response as well.<sup>[156]</sup>

Emulsion-based adjuvants like Freund's adjuvant and montanide ISA51 are easy to develop at comparatively very low cost among different adjuvants available for vaccination. These adjuvants can be made as water-in-oil (W/O) emulsion with dispersed antigenic media and continuous oily phases.<sup>[168]</sup> These adjuvants based on emulsion have the advantages of improving the vaccine's nontoxicity and securing long-term protective immunity. Nevertheless, these adjuvants' preparation time, difficulties during injection by syringe, and localized toxicities at the injection sites are some common problems with these emulsions.<sup>[169]</sup> Synthetic low molecular weight block polymer has been proposed as a better replacement to reduce the toxicity of surfactants used during the manufacturing of such adjuvants. TiterMax is an ideal example of this kind of modified adjuvant for vaccination. TiterMax is a squalene-based W/O emulsion containing polyoxyethylene-polyoxypropylene-polyoxyethylene (POE-POP-POE) polymer.<sup>[170]</sup> To overcome the mild toxicity and poor degradability of TiterMax, recently, Huang et al. have reported multiphase emulsion based on the hydrophilic polymeric emulsifiers PEG-b-PLA, PEG-b-PCL, and PEG-b-PLACL in the antigen phase of oily ISA51-adjuvant-based vaccines.<sup>[171]</sup> This hydrophilic polymerstabilized ISA51 emulsion increase fluidity and conceptually diminish local reactions. The excellent biodegradability and biocompatibility of these modified adjuvants make them promising candidates for SARS-CoV-2 vaccine delivery applications. Huan et al. recently studied the effect of CoVaccine HT<sup>TM</sup> (W/O emulsion type adjuvant) against the SARS-CoV-2 spike S1 protein in mice.<sup>[172]</sup> The CoVaccine HT<sup>TM</sup> has already proven its efficacy in Malaria, Zika, Ebola, and other antiviral vaccine formulations.<sup>[173]</sup> This adjuvant is composed of negatively charged sucrose fatty acid sulfate ester and plant-derived squalene.<sup>[174]</sup> In that study, they compared the potency and efficacy of CoVaccine HT<sup>TM</sup> against two gold standard adjuvants (i.e. alum and Th2 adjuvant). They found that the CoVaccine HT<sup>TM</sup> induced cell-mediated immune responses, antigen-specific antibody titers and virus-neutralizing antibody titers significantly compared to alum adjuvant. High potency, efficacy, biodegradability, and biocompatibility are key features in these modified adjuvants that make them attractive for SARS-CoV-2 vaccine delivery applications.<sup>[164]</sup>

#### Polymers in lung tissue engineering applications

Tissue engineering is a process to reconstruct a tissue for clinical use or repair damaged ones by seeding stem cells (e.g. ESCs, iPSCs, and MSCs) on a biological scaffold with extracellular matrix (ECM) proteins. The seeded cells are expected to proliferate and differentiate into the proper cell populations, followed by reconstructing targeted organs or tissue for clinical applications. In ARDS injury, one way to alleviate lung injury is to regenerate physiologically functional lung tissues to replace the damaged tissue.<sup>[175]</sup> In tissue engineering approaches, a scaffold is used as a 3D substrate that maintains the specific biological environment for tissue regeneration, keeps the mechanical structure of the regenerated tissue or organ, and provokes a minimal toxicity level, low or no inflammation.<sup>[176]</sup> Scaffolds can be synthesized using polymeric materials either from natural or from synthetic sources (Figure 3).<sup>[177–179]</sup> The first 3D scaffold has been made of Gelfoam (vacuolar sponge based on gelatin) and used to culture rat fetal lung cells.<sup>[180]</sup> In that study, it has been reported that fetal lung cells survived both in and around the collagen matrix and proliferated into new epithelial and endothelial cells. In another study, chondroitin sulphate proteoglycans (CSPGs)



**Figure 3.** Polymer-based scaffolds in tissue engineering applications. (A) PVA/Collagen composite nanofibrous electrospun scaffold for application in tissue-engineered cornea. Reprinted from Wu et al.<sup>[177]</sup> Copyright 2018 MDPI. (B) Peptide/GO/ $\beta$ -TCP/PLGA scaffold from cryogenic 3D printing for critical-sized bone defect repair. Reprinted from Zhang et al.<sup>[178]</sup> Copyright 2019 MDPI. (C) Fabrication of an injectable, porous hyaluronic acid-based hydrogel by in-situ and bubble-forming hydrogel entrapment process. Reprinted from Wang et al.<sup>[179]</sup> Copyright 2020 MDPI.

scaffold has been developed for pulmonary tissue regeneration, and further studies indicated its potentiality to construct biomimetic matrices for inducing lung epithelial morphogenesis.<sup>[181]</sup>

On the other hand, synthetic materials are another potential candidate for use as scaffolds in tissue engineering applications, although these materials' low biocompatibility hinders their broad applications. Synthetic materials are combined with natural biomaterials to overcome this limitation and enhance scaffold properties. Elasticity and biodegradability are two key features for materials to be used as a scaffold for lung tissue engineering. Synthetic materials containing biodegradable polymers like PLGA are vigorously explored to develop porous scaffolds for tissue engineering.<sup>[182]</sup> Due to the lack of biocompatibility, synthetic polymers often fail to drive the differentiation of cultured cells.<sup>[183]</sup> Even sometimes, synthetic polymers coated with natural ECM proteins failed to guide seeded cells into required destiny despite the cells' initial attachment onto the scaffold. Therefore, it is necessary to employ some surface modification approaches (e.g. coating with instructive peptide domains) to enhance scaffolds' biocompatibility.<sup>[184]</sup> Besides, elastomeric polymers with more organotypic mechanical properties essential for cell growth and differentiation can also be useful for better cyclic respiration strain.<sup>[185]</sup> A widely used biodegradable polymer is poly(glycolic acid) (PGA), which has been used previously as a patch grafted to the rat's incised lung.<sup>[186]</sup> It is reported that the adipose-derived stem cells (ASCs) seeded onto PGA graft succeeded to regenerate alveolar and vascular tissues.<sup>[187]</sup> Another popular biodegradable hydrophobic polymer is poly(D,L-lactic acid) (PDLLA) that is suggested to be used as a scaffold for lung tissue due to its elasticity feature that resembles the lung environment.<sup>[188]</sup> Few polymeric hydrogels have been designed to meet certain requirements (i.e. mechanical strength, elasticity, stiffness, and controlled degradation kinetics) needed for lung tissue regeneration. A novel porous hydrogel scaffold has been developed recently from a blend of hyaluronic acid hydrogels (HAG) gel.<sup>[189]</sup> This hydrogel scaffold provided a lower inflammatory response, high elasticity with rapid hydration ability due to the interconnected porous network. This novel HAG gel fulfilled the characteristics compatible with lung engineering. Polymer-based porous matrices are considered ideal scaffolds because of their appropriate 3D structure, biocompatibility and biodegradability. Hence, they operate as appropriate substrate to induce stem cells' differentiation by regeneration of physiologically functional lung tissues.

# Scope of polymer-based nano-therapies to combat respiratory injury: Progress, prospects, and challenges

For the detection of respiratory viruses, various polymeric nanobiosensors, including MIP-based sensors, have been developed in recent years.<sup>[40]</sup> Modification and surface functionalization of MIPs are unique nanobiosensing strategies for faster and more specific detection of viral infection. In recent years, these low-cost, affordable, and highly selective detection systems have drawn the research community and biomedical industry's attention to replacing costly, time-consuming, and labor-intensive traditional detection techniques. It is well established that patients with severe COVID-19 infection experience long-term respiratory complications of the infection.<sup>[190]</sup> Therefore, detection of early-stage infection is vital to mitigate long-term complications. The low-level detection of a specific SARS-CoV-2 biomarker can be an option for early evaluation, management, and infection treatment. Nanobiosensors with multi-functionalities could also have the potential for immediate detection of the SARS-CoV-2 virus. Despite their potential application in virus detection, there are still some significant limitations like reliability, reproducibility, and diagnosis performance and accuracy. Therefore, more elaborative analyses and research should be conducted in the near future to solve these drawbacks.

It is well established that COVID-19 infected patients often experience hyperinflammation correlated with acute respiratory injury, e.g. ARDS. Such respiratory injury might cause severe, long-lasting damage to the lungs, resulting in a substantial reduction of the patient's life quality. Therefore, it is crucial to develop a unique treatment strategy to treat the consequences of COVID-19 infection, including attenuation of the inflammatory response leading to respiratory injury. In respiratory injuries, microvascular leakiness is the outcome of inflammatory vasoactive factors that induce permeability.<sup>[191]</sup> Hence, drugs can be administered systemically and will localize to the lungs by passive targeting.<sup>[192]</sup> A controlled drug delivery system is an innovative, passively targeted therapeutic strategy that maximizes efficacy and increases inflammation resolution, resulting in reducing collateral damage to healthy organs in patients. Polymeric nanoparticles as nanocarriers have already proved their ability in anti-viral treatment.<sup>[193]</sup> Such nanocarrier systems have shown improved and efficient drug delivery against HIV, Influenza virus, HSV, Respiratory syncytial virus, Zika virus, and Monkeypox virus.<sup>[194]</sup> Therefore, the nanoencapsulation of anti-viral drug candidates for COVID-19 may be an attractive and safer treatment strategy.<sup>[195]</sup> Theranostic nanoparticles as nanocarriers have already been investigated against various viral infections like SARS or MERS.<sup>[196]</sup> Theranostic nanocarriers can improve the drug delivery, ensure selective delivery of siRNA, block viral entry inside the cells, and trigger host cells' immune systems.<sup>[196]</sup> Therefore, biocompatible theranostic nanocarriers can be another promising strategy to deliver therapeutic agents *via* the intranasal route to combat against CoV-related respiratory injuries.

Despite the potentiality of polymeric nanocarrier systems, some bottlenecks must be addressed to facilitate its broader implementation. Since most recent studies have used *in vitro* approaches to evaluate the biocompatibility, it is crucial to ensure the safe use of nanomaterials inside the biological systems. Developing polymeric nanocarrier to deliver drugs, vaccines, genes, and other biologics in a controlled manner that can precisely cure the respiratory injury and other COVID-19 related complications is the ultimate goal of nanotechnology experts. The development of such smart polymeric nanocarrier with high efficacy and target specific functionality in the human body is very challenging to achieve. Additionally, the nanocarrier-based delivery system's efficacy is also related to the size, shape, and the surface charge of the nanoparticles.<sup>[16]</sup> It has been reported that spherical nanoparticles compared to rodshaped particles are more prone to phagocytosis by macrophages and APCs (e.g. dendritic cells).<sup>[197]</sup> Positively charged nanomaterials are taken up more easily by the epithelial cell membranes due to its anionic nature. Due to the multifaceted interactions between nanomaterials and biological systems, nanomaterials' fate and behavior can be changed under physiological conditions. A high dose of these agents may cause severe side effects inside human body due to the off-targeting feature can be worse than SARS-CoV-2 infection. However, it is challenging to foresee the response of the nanomaterials under harsh biological conditions particularly in SARS-CoV-2 virus infection. Once the nanocarriers reach the blood circulation inside the body, they can interact with proteins to form protein corona.<sup>[198]</sup> Other complications may appear when nanoparticles enter blood circulation due to the complex matrix containing ions, small molecules, proteins, and cells in the circulation.<sup>[199]</sup> Thus, the characterization of protein corona is a vital step to be investigated during polymeric nanomedicine development to treat COVID-19 related complications including respiratory injury. Moreover, reliable in vivo models are required to explore the toxicokinetic behavior of the nanoparticulate carriers in the body. In the case of aerosol therapy of nanoparticles, although the incidence of adverse events is minimized, however, some observational studies have indicated that there is potential to cause local or systemic toxicity in the form of coughing, airway irritation, bronchospasms, and in some cases pulmonary injury.<sup>[191]</sup> Therefore, a more detailed investigation to assess the safety profile of polymeric and other organic/inorganic nanoparticles that are considered for delivery via aerosol therapy is necessary. Off-targeting limitation of the

nanocarrier can be overcome by introducing stimuli-responsive nanocarrier that can deliver therapeutic agents to the infected respiratory system in a controlled and target specific manner to combat CoV-associated symptoms. Polymeric hydrogels (e.g. PEG) are widely used for the controlled release of biomolecules. Control of gelation is crucial in delivering therapeutic agents because appropriate dosing and release kinetics rely on the understanding of fundamental gelation kinetics of hydrogel-based nanocarrier systems.<sup>[200]</sup> Thermoresponsive injectable hydrogels (e.g. PNIPAm) is a type of hydrogel-based delivery system developed in recent years as a drug delivery system and cell encapsulation system.<sup>[129]</sup> These hydrogels are free-flowing solutions below physiological temperature, and after in vivo injection, they convert into non-flowing gels at body temperature. A key feature of living systems is the ability to sense and react to external environmental stimuli. Therefore, cells and tissues' physiological responses with advances in polymer chemistry can enable the development of stimuli-responsive biohybrid hydrogels that can functionally interconnect with the living systems. These hybrid hydrogel-based delivery systems can be a promising nanomedicine strategy for drug or other biomolecule delivery applications to treat CoV-related respiratory injury.

The addition of cells to the scaffold to produce tissue construct under the controlled addition of specific growth factors to support tissue growth in vitro is the classical tissue engineering approach. Interestingly, lung tissue reconstructs using this classic approach are not suitable because the reconstructed tissues lack appropriate vascularization and the intricate organizational patterns found in normal lung tissue.<sup>[201]</sup> Bioprinting offers the advantages of placing various cell types in layer-bylayer constructs into a soft scaffold using a computer-controlled design template to resolve this issue.<sup>[201]</sup> Concurrent printing of hydrogel scaffold containing biomolecules will allow for precise placement of cells and proteins within 3D structures of complex tissues like lung. This hydrogel-based organ printing approach will also provide the advantage for spatial control of the scaffold structure, the type and arrangement of cells, the thickness of the tissue, and the formation of capillaries and vessels to make physiologically functional lung tissue. Despite these technological advantages in polymer-based lung tissue engineering, precise control of hydrogel properties like porosity remains a major hurdle in scaffold design.<sup>[202]</sup> The porosity, pore architecture, and interconnectivity between pores are some significant features that play a vital role in cell survival, proliferation, and migration during tissue regeneration.<sup>[202]</sup> Control of these features can influence the scaffold's cell movement and movement of oxygen and nutrients and regulate cell attachment to the scaffold. Therefore, it is now crucial to develop hybrid stimuli-responsive hydrogel scaffolds with ECM components similar to natural tissue to provide cells and developing lung tissue with the environment required to function like natural tissue in a native *in vivo* environment.

#### **Concluding remarks**

Amid the COVID-19 pandemic, primary diagnostic tools for SARS-CoV-2 detection are mainly based on RT-PCR-based assays. Although PCR-based tools are broadly applied in recent times, such tools are only limited to detect viral nucleic acids.

Moreover, the testing capacity, cost, detection time, and availability are some issues of this technique. Contrarily, polymeric nanobiosensing devices are more versatile and can be used for antigen, antibody, and nucleic acid detection. These novel sensing tools can also provide rapid, reliable, broadly accessible, and low-cost diagnosis in this pandemic. A wide range of treatment strategies using polymer-based nanotechnologies has been developed and commercialized so far for different viral infections like HIV and HSV-1 and 2. Advancements in these nano-therapeutics developments can help to invent novel treatments and vaccines to tackle COVID-19 related complications to the next level. Although polymer-based nano-therapies offer a broad range of antiviral therapeutics opportunities, it is still in the infantile stage. Some of the major barriers in nanomedicine development are long-term toxicity, fabrication and characterization complexities, and large-scale production difficulties. Moreover, tissue engineering approaches in respiratory injury treatment is still limited and far from clinical use. Therefore, future perspective should focus on addressing and solving current drawbacks of polymeric nano-therapies to develop a revolutionary solution for the treatment of COVID-19 and other viral infection related respiratory injuries.

# **Disclosure statement**

No conflict of interest is reported.

#### ORCID

Md Mohosin Rana (D http://orcid.org/0000-0003-3299-6538

# References

- [1] Pal M, Berhanu G, Desalegn C, et al. Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2): an update. Cureus. 2020;12(3).
- [2] Flynn Makic MB. Prone position of patients with COVID-19 and acute respiratory distress syndrome. J Perianesth Nurs. 2020;35(4):437–438.
- [3] Piantadosi CA, Schwartz DA. The acute respiratory distress syndrome. Ann Intern Med. 2004;141(6):460-470.
- [4] Han J, Li Y, Li Y. Strategies to enhance mesenchymal stem cell-based therapies for acute respiratory distress syndrome. Stem Cells Int. 2019;2019:5432134.
- [5] Li L, Huang Q, Wang DC, et al. Acute lung injury in patients with COVID-19 infection. Clin Transl Med. 2020;10(1):20–27.
- [6] Patel VJ, Biswas Roy S, Mehta HJ, et al. Alternative and natural therapies for acute lung injury and acute respiratory distress syndrome. Biomed Res Int. 2018;2018: 2476824.
- [7] Puneet P, Moochhala S, Bhatia M. Chemokines in acute respiratory distress syndrome. Am J Physiol Lung Cell Mol Physiol. 2005;288(1):L3–L15.
- [8] Rana MM. Cytokine storm in COVID-19: potential therapeutics for immunomodulation. J Res Clin Med. 2020;8(1):38–38.
- [9] Hojyo S, Uchida M, Tanaka K, et al. How COVID-19 induces cytokine storm with high mortality. Inflamm Regen. 2020;40(1):37.
- [10] Beitler JR, Schoenfeld DA, Taylor Thompson B. Preventing ARDS: progress, promise, and pitfalls. Chest. 2014;146(4):1102–1113.

- [11] Osman N, Kaneko K, Carini V, et al. Saleem, I. Carriers for the targeted delivery of aerosolized macromolecules for pulmonary pathologies. Expert Opin Drug Deliv. 2018;15(8):821-834.
- [12] Patil JS, Sarasija S. Pulmonary drug delivery strategies: a concise, systematic review. Lung India. 2012;29(1):44–49.
- [13] Omri A. Pulmonary drug and vaccine delivery: therapeutic significance and major challenges. Expert Opin Drug Deliv. 2015;12(6):853-855.
- [14] Song HHG, Rumma RT, Ozaki CK, et al. Vascular tissue engineering: progress, challenges, and clinical promise. Cell Stem Cell. 2018;22(3):340–354.
- [15] Van Rijt SH, Bein T, Meiners S. Medical nanoparticles for next generation drug delivery to the lungs. Eur Respir J. 2014;44(3):765–774.
- [16] Din FU, Aman W, Ullah I, et al. Effective use of nanocarriers as drug delivery systems for the treatment of selected tumors. IJN. 2017;12:7291–7309.
- [17] Tsuchiya T, Doi R, Obata T, et al. Lung microvascular niche, repair, and engineering. Front Bioeng Biotechnol. 2020;8:105.
- [18] Lim YH, Tiemann KM, Hunstad DA, et al. Polymeric nanoparticles in development for treatment of pulmonary infectious diseases. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2016;8(6):842–871.
- [19] De Santis MM, Bölükbas DA, Lindstedt S, et al. How to build a lung: latest advances and emerging themes in lung bioengineering. Eur Respir J. 2018;52(1):1–19.
- [20] Hou YJ, Okuda K, Edwards CE, et al. SARS-CoV-2 reverse genetics reveals a variable infection gradient in the respiratory tract. Cell. 2020;182(2):429–446.e14.
- [21] Zhang H, Penninger JM, Li Y, et al. Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target. Intensive Care Med. 2020;46(4):586–590.
- [22] Kim JS, Lee JY, Yang JW, et al. Immunopathogenesis and treatment of cytokine storm in COVID-19. Theranostics. 2021;11(1):316-329.
- Badraoui R, Alrashedi MM, El-May MV, et al. Acute respiratory distress syndrome: a life threatening associated complication of SARS-CoV-2 infection inducing COVID-19. J Biomol Struct Dyn. 2020;1–10.
- [24] Sartori C, Matthay MA. Alveolar epithelial fluid transport in acute lung injury: new insights. Eur Respir J. 2002;20(5):1299–1313.
- [25] Matuschak GM, Lechner AJ. Acute lung injury and the acute respiratory distress syndrome: pathophysiology and treatment. Mo Med. 2010;107(4):252–258.
- [26] Zemans RL, Matthay MA. What drives neutrophils to the alveoli in ARDS? Thorax. 2017;72(1):1–3.
- [27] Gallelli L, Zhang L, Wang T, et al. Severe acute lung injury related to COVID-19 infection: a review and the possible role for escin. J Clin Pharmacol. 2020;60(7): 815–825.
- [28] Matthay MA, Zimmerman GA. Acute lung injury and the acute respiratory distress syndrome: four decades of inquiry into pathogenesis and rational management. Am J Respir Cell Mol Biol. 2005;33(4):319–327.
- [29] Udugama B, Kadhiresan P, Kozlowski HN, et al. Diagnosing COVID-19: the disease and tools for detection. ACS Nano. 2020;14(4):3822-3835.
- [30] Bhalla N, Pan Y, Yang Z, et al. Opportunities and challenges for biosensors and nanoscale analytical tools for pandemics: COVID-19. ACS Nano. 2020;14(7):7783–7807.
- [31] Surkova E, Nikolayevskyy V, Drobniewski F. False-positive COVID-19 results: hidden problems and costs. Lancet Respir Med. 2020;8(12):1167–1168.(20)30453-7.
- [32] Pokhrel P, Hu C, Mao H. Detecting the coronavirus (CoVID-19). ACS Sens. 2020; 5(8):2283-2297.
- [33] Malik AA, Nantasenamat C, Piacham T. Molecularly imprinted polymer for human viral pathogen detection. Mater Sci Eng C Mater Biol Appl. 2017;77:1341–1348.
- [34] Goud KY, Reddy KK, Khorshed A, et al. Electrochemical diagnostics of infectious viral diseases: trends and challenges. Biosens Bioelectron. 2021;180:113112.

- [35] Navakul K, Warakulwit C, Yenchitsomanus P. t, et al. A novel method for dengue virus detection and antibody screening using a graphene-polymer based electrochemical biosensor. Nanomedicine. 2017;13(2):549–557.
- [36] Lu CH, Zhang Y, Tang SF, et al. Sensing HIV related protein using epitope imprinted hydrophilic polymer coated quartz crystal microbalance. Biosens Bioelectron. 2012; 31(1):439-444.
- [37] Wangchareansak T, Thitithanyanont A, Chuakheaw D, et al. A novel approach to identify molecular binding to the influenza virus H5N1: screening using molecularly imprinted polymers (MIPs). Medchemcomm. 2014;5(5):617–621.
- [38] Tai DF, Lin CY, Wu TZ, et al. Recognition of dengue virus protein using epitopemediated molecularly imprinted film. Anal Chem. 2005;77(16):5140–5143.
- [39] Nandy Chatterjee T, Bandyopadhyay R. A molecularly imprinted polymer-based technology for rapid testing of COVID-19. Trans Indian Natl Acad Eng. 2020;5(2): 225–228.
- [40] Iravani S. Nano- and biosensors for the detection of SARS-CoV-2: challenges and opportunities. Mater Adv. 2020;1(9):3092–3103.
- [41] Murthy S, Gomersall CD, Fowler RA. Care for critically ill patients with COVID-19. JAMA. 2020;323(15):1499–1500.
- [42] Papazian L, Aubron C, Brochard L, et al. Formal guidelines: management of acute respiratory distress syndrome. Ann Intensive Care. 2019;9(1):1–18.
- [43] Aokage T, Palmér K, Ichiba S, et al. Extracorporeal membrane oxygenation for acute respiratory distress syndrome. J Intensive Care. 2015;3(1):17.
- [44] Wu MY, Huang CC, Wu TI, et al. Venovenous extracorporeal membrane oxygenation for acute respiratory distress syndrome in adults prognostic factors for outcomes. Medicine (United States). 2016;95(8):e2870.
- [45] Wu R, Wang L, Kuo HCD, et al. An update on current therapeutic drugs treating COVID-19. Curr Pharmacol Reports. 2020;6:56–70.
- [46] Ho ATN, Patolia S, Guervilly C. Neuromuscular blockade in acute respiratory distress syndrome: a systematic review and meta-analysis of randomized controlled trials. J Intensive Care. 2020;8:1–11. https://doi.org/10.1186/s40560-020-0431-z.
- [47] Russell B, Moss C, George G, et al. Associations between immune-suppressive and stimulating drugs and novel COVID-19 - a systematic review of current evidence. Ecancermedicalscience. 2020;14(1022): 1–43.
- [48] Curran CS, Rivera DR, Kopp JB. COVID-19 usurps host regulatory networks. Front Pharmacol. 2020;11:1278.
- [49] Bonam SR, Kaveri SV, Sakuntabhai A, et al. Adjunct immunotherapies for the management of severely ill COVID-19 patients. Cell Rep Med. 2020;1(2):100016.
- [50] Sivasankarapillai VS, Pillai AM, Rahdar A, et al. On facing the SARS-CoV-2 (COVID-19) with combination of nanomaterials and medicine: possible strategies and first challenges. Nanomaterials. 2020;10(5):852.
- [51] Russell CD, Millar JE, Baillie JK. Clinical evidence does not support corticosteroid treatment for 2019-NCoV lung injury. Lancet. 2020;395(10223):473-475.
- [52] Chakravarty M, Vora A. Nanotechnology-based antiviral therapeutics. Drug Deliv Transl Res. 2020;1-40.
- [53] Patel A, Patel M, Yang X, et al. Recent advances in protein and peptide drug delivery: a special emphasis on polymeric nanoparticles. Protein Pept Lett. 2014;21(11): 1102–1120.
- [54] Casalini T, Rossi F, Castrovinci A, et al. A perspective on polylactic acid-based polymers use for nanoparticles synthesis and applications. Front Bioeng Biotechnol. 2019; 7:259.
- [55] Daglar B, Ozgur E, Corman ME, et al. Polymeric nanocarriers for expected nanomedicine: current challenges and future prospects. RSC Adv. 2014;4(89):48639–48659.
- [56] Avramović N, Mandić B, Savić-Radojević A, et al. Polymeric nanocarriers of drug delivery systems in cancer therapy. Pharmaceutics. 2020;12(4):298.

- [57] Zhao J, Stenzel MH. Entry of nanoparticles into cells: the importance of nanoparticle properties. Polym Chem. 2018;9(3):259–272.
- [58] Vijayakameswara Rao N, Ko H, Lee J, et al. Recent progress and advances in stimuliresponsive polymers for cancer therapy. Front Bioeng Biotechnol. 2018;6:110.
- [59] Brighenti R, Li Y, Vernerey FJ. Smart polymers for advanced applications: a mechanical perspective review. Front Mater. 2020;7:196.
- [60] Mohd Amin MCI, Ahmad N, Halib N, et al. Synthesis and characterization of thermoand PH-responsive bacterial cellulose/acrylic acid hydrogels for drug delivery. Carbohydr Polym. 2012;88(2):465–473.
- [61] De Clercq E, Li G. Approved antiviral drugs over the past 50 years. Clin Microbiol Rev. 2016;29(3):695-747.
- [62] Soota K, Maliakkal B. Ribavirin induced hemolysis: a novel mechanism of action against chronic hepatitis C virus infection. World J Gastroenterol. 2014;20(43): 16184–16190.
- [63] Li C, Wang J, Wang Y, et al. Recent progress in drug delivery. Acta Pharm Sin B. 2019;9(6):1145–1162.
- [64] Haggag Y, Elshikh M, El-Tanani M, et al. Nanoencapsulation of sophorolipids in PEGylated poly(lactide-co-glycolide) as a novel approach to target colon carcinoma in the murine model. Drug Deliv Transl Res. 2020;10(5):1353–1366.
- [65] Hillaireau H, Le Doan T, Besnard M, et al. Encapsulation of antiviral nucleotide analogues azidothymidine-triphosphate and cidofovir in poly(iso-butylcyanoacrylate) nanocapsules. Int J Pharm. 2006;324(1):37–42.
- [66] Deng S, Gigliobianco MR, Censi R, et al. Polymeric nanocapsules as nanotechnological alternative for drug delivery system: current status, challenges and opportunities. Nanomaterials. 2020;10(5):847.
- [67] Donalisio M, Leone F, Civra A, et al. Acyclovir-loaded chitosan nanospheres from nano-emulsion templating for the topical treatment of herpesviruses infections. Pharmaceutics. 2018;10(2):46.
- [68] Xu Y, Liu H, Song L. Novel drug delivery systems targeting oxidative stress in chronic obstructive pulmonary disease: a review. J Nanobiotechnology. 2020;18(1):145.
- [69] Nishiyama N, Matsumura Y, Kataoka K. Development of polymeric micelles for targeting intractable cancers. Cancer Sci. 2016;107(7):867–874.
- [70] Suk JS, Xu Q, Kim N, et al. PEGylation as a strategy for improving nanoparticle-based drug and gene delivery. Adv Drug Deliv Rev. 2016;99:28–51.
- [71] Wu J, Yuan J, Ye B, et al. Dual-responsive core crosslinking glycopolymer-drug conjugates nanoparticles for precise hepatocarcinoma therapy. Front Pharmacol. 2018;9(Jul): 663.
- [72] Kim K-T, Lee J-Y, Kim D-D, et al. Recent progress in the development of poly(lacticco-glycolic acid)-based nanostructures for cancer imaging and therapy. Pharmaceutics. 2019;11(6):280.
- [73] Li W, Yu F, Wang Q, et al. Co-delivery of HIV-1 entry inhibitor and nonnucleoside reverse transcriptase inhibitor shuttled by nanoparticles: cocktail therapeutic strategy for antiviral therapy. AIDS. 2016;30(6):827–837.
- [74] Lu X, Zhu T, Chen C, et al. Right or left: the role of nanoparticles in pulmonary diseases. Int J Mol Sci. 2014;15(10):17577–17600.
- [75] Jamali A, Mottaghitalab F, Abdoli A, et al. Inhibiting influenza virus replication and inducing protection against lethal influenza virus challenge through chitosan nanoparticles loaded by SiRNA. Drug Deliv Transl Res. 2018;8(1):12–20.
- [76] Kudgus RA, Walden CA, McGovern RM, et al. Tuning pharmacokinetics and biodistribution of a targeted drug delivery system through incorporation of a passive targeting component. Sci Rep. 2014;4(1):1–9.
- [77] Jhaveri AM, Torchilin VP. Multifunctional polymeric micelles for delivery of drugs and SiRNA. Front Pharmacol. 2014;5:77.

- [78] Rad AH, Asiaee F, Jafari S, et al. Poly(ethylene glycol)-poly(ε-caprolactone)-based micelles for solubilization and tumor-targeted delivery of silibinin. Bioimpacts. 2020; 10(2):87–95.
- [79] Rani S, Gothwal A, Khan I, et al. Smartly engineered PEGylated Di-block nanopolymeric micelles: duo delivery of isoniazid and rifampicin against mycobacterium tuberculosis. AAPS PharmSciTech. 2018;19(7):3237–3248.
- [80] Sheth U, Tiwari S, Bahadur A. Preparation and characterization of anti-tubercular drugs encapsulated in polymer micelles. J Drug Deliv Sci Technol. 2018;48:422–428.
- [81] Ahn YS, Baik HJ, Lee BR, et al. Preparation of multifunctional polymeric micelles for antiviral treatment. Macromol Res. 2010;18(8):747–752.
- [82] Alven S, Nqoro X, Buyana B, et al. Polymer-drug conjugate, a potential therapeutic to combat breast and lung cancer. Pharmaceutics. 2020;12(5):406.
- [83] Aquino RS, Park PW. Glycosaminoglycans and infection. Front Biosci (Landmark Ed). 2016;21(6):1260–1277.
- [84] Smith AAA, Kryger MBL, Wohl BM, et al. Macromolecular (pro)drugs in antiviral research. Polym Chem. 2014;5(22):6407–6425.
- [85] Van Dongen MA, Dougherty CA, Banaszak Holl MM. Multivalent polymers for drug delivery and imaging: the challenges of conjugation. Biomacromolecules. 2014;15(9): 3215–3234.
- [86] Kuo YC, Lin PI, Wang CC. Targeting nevirapine delivery across human brain microvascular endothelial cells using transferrin-grafted poly(lactide-co-glycolide) nanoparticles. Nanomedicine (Lond). 2011;6(6):1011–1026.
- [87] Dhoke DM, Basaiyye SS, Khedekar PB. Development and characterization of L-HSA conjugated PLGA nanoparticle for hepatocyte targeted delivery of antiviral drug. J Drug Deliv Sci Technol. 2018;47:77–94.
- [88] Destache CJ, Belgum T, Christensen K, et al. Combination antiretroviral drugs in PLGA nanoparticle for HIV-1. BMC Infect Dis. 2009;9(198): 1–8.
- [89] Ramana LN, Sharma S, Sethuraman S, et al. Evaluation of chitosan nanoformulations as potent anti-HIV therapeutic systems. Biochim Biophys Acta. 2014;1840(1):476–484.
- [90] Ahmad Z, Pandey R, Sharma S, et al. Alginate nanoparticles as antituberculosis drug carriers: formulation development, pharmacokinetics and therapeutic potential. Indian J Chest Dis Allied Sci. 2006;48(3):171–176.
- [91] Kalombo L, Lemmer Y, Semete-Makokotlela B, et al. Spray-dried, nanoencapsulated, multi-drug anti-tuberculosis therapy aimed at once weekly administration for the duration of treatment. Nanomaterials. 2019;9(8):1167.
- [92] Grande F, Ioele G, Occhiuzzi MA, et al. Reverse transcriptase inhibitors nanosystems designed for drug stability and controlled delivery. Pharmaceutics. 2019;11(5):197.
- [93] Gong Y, Chowdhury P, Midde NM, et al. Novel elvitegravir nanoformulation approach to suppress the viral load in HIV-infected macrophages. Biochem Biophys Rep. 2017;12:214–219.
- [94] Varshosaz J, Taymouri S, Jafari E, et al. Formulation and characterization of cellulose acetate butyrate nanoparticles loaded with nevirapine for HIV treatment. J Drug Deliv Sci Technol. 2018;48:9–20.
- [95] Tyagi R, Lala S, Verma AK, et al. Targeted delivery of arjunglucoside I using surface hydrophilic and hydrophobic nanocarriers to combat experimental leishmaniasis. J Drug Target. 2005;13(3):161–171.
- [96] Zhou L, Zhang P, Chen Z, et al. Preparation, characterization, and evaluation of amphotericin B-loaded MPEG-PCL-g-PEI micelles for local treatment of oral Candida Albicans. Int J Nanomedicine. 2017;12:4269–4283.
- [97] Giannavola C, Bucolo C, Maltese A, et al. Influence of preparation conditions on acyclovir-loaded poly-d,l-lactic acid nanospheres and effect of PEG coating on ocular drug bioavailability. Pharm Res. 2003;20(4):584–590. [12739765]

- [98] Fresta M, Fontana G, Bucolo C, et al. Ocular tolerability and in vivo bioavailability of poly(ethylene glycol) (PEG)-coated polyethyl-2-cyanoacrylate nanosphere-encapsulated acyclovir. J Pharm Sci. 2001;90(3):288–297.
- [99] Mosqueira VCF, Loiseau PM, Bories C, et al. Efficacy and pharmacokinetics of intravenous nanocapsule formulations of halofantrine in plasmodium berghei-infected mice. Antimicrob Agents Chemother. 2004;48(4):1222–1228.
- [100] Calvo P, Vila-Jato JL, Alonso M. J. Comparative in vitro evaluation of several colloidal systems, nanoparticles, nanocapsules, and nanoemulsions, as ocular drug carriers. J Pharm Sci. 1996;85(5):530–536.
- [101] Li Q, Du YZ, Yuan H, et al. Synthesis of lamivudine stearate and antiviral activity of stearic acid-g-chitosan oligosaccharide polymeric micelles delivery system. Eur J Pharm Sci. 2010;41(3-4):498-507.
- [102] Silva M, Ricelli NL, Seoud OE, et al. Potential tuberculostatic agent: micelle-forming pyrazinamide prodrug. Arch Pharm (Weinheim). 2006;339(6):283–290.
- [103] Sawdon AJ, Peng CA. Polymeric micelles for acyclovir drug delivery. Colloids Surf B Biointerfaces. 2014;122:738–745.
- [104] Yandrapu SK, Kanujia P, Chalasani KB, et al. Development and optimization of thiolated dendrimer as a viable mucoadhesive excipient for the controlled drug delivery: an acyclovir model formulation. Nanomedicine. 2013;9(4):514–522.
- [105] Ma M, Cheng Y, Xu Z, et al. Evaluation of polyamidoamine (PAMAM) dendrimers as drug carriers of anti-bacterial drugs using sulfamethoxazole (SMZ) as a model drug. Eur J Med Chem. 2007;42(1):93–98.
- [106] Bhadra D, Bhadra S, Jain NK. PEGylated peptide dendrimeric carriers for the delivery of antimalarial drug chloroquine phosphate. Pharm Res. 2006;23(3):623–633.
- [107] Cheng Y, Qu H, Ma M, et al. Polyamidoamine (PAMAM) dendrimers as biocompatible carriers of quinolone antimicrobials: an in vitro study. Eur J Med Chem. 2007; 42(7):1032–1038.
- [108] Cojocaru FD, Botezat D, Gardikiotis I, et al. Nanomaterials designed for antiviral drug delivery transport across biological barriers. Pharmaceutics. 2020;12(2):171.
- [109] Srinivas P, Reddy A. Formulation and evaluation of isoniazid loaded nanosponges for topical delivery. PNT. 2015;3(1):68-76.
- [110] Mcdonald TO, Giardiello M, Martin P, et al. Antiretroviral solid drug nanoparticles with enhanced oral bioavailability: production, characterization, and in vitro-in vivo correlation. Adv Healthc Mater. 2014;3(3):400–411.
- [111] Sun B, Yeo Y. Nanocrystals for the parenteral delivery of poorly water-soluble drugs. Curr Opin Solid State Mater Sci. 2012;16(6):295-301.
- [112] Janaszewska A, Lazniewska J, Trzepiński P, et al. Cytotoxicity of dendrimers. Biomolecules. 2019;9(8):1-23.
- [113] Mhlwatika Z, Aderibigbe BA. Application of dendrimers for the treatment of infectious diseases. Molecules. 2018;23(9):2205.
- [114] Sepúlveda-Crespo D, Ceña-Díez R, Jiménez JL, et al. Mechanistic studies of viral entry: an overview of dendrimer-based microbicides as entry inhibitors against both HIV and HSV-2 overlapped infections. Med Res Rev. 2017;37(1):149–179.
- [115] Caminade AM, Turrin CO, Majoral JP. Biological properties of phosphorus dendrimers. New J Chem. 2010;34(8):1512–1524.
- [116] Briz V, Sepúlveda-Crespo D, Diniz AR, et al. Development of water-soluble polyanionic carbosilane dendrimers as novel and highly potent topical anti-HIV-2 microbicides. Nanoscale. 2015;7(35):14669–14683.
- [117] Heinrich MA, Martina B, Prakash J. Nanomedicine strategies to target coronavirus. Nano Today. 2020;35:100961.
- [118] Sun Y, Guo F, Zou Z, et al. Cationic nanoparticles directly bind angiotensin-converting enzyme 2 and induce acute lung injury in mice. Part Fibre Toxicol. 2015;12(1): 1–13.

- [119] Tan GT, Wickramasinghe A, Verma S, et al. Sulfonic acid polymers are potent inhibitors of HIV-1 induced cytopathogenicity and the reverse transcriptases of both HIV-1 and HIV-2. BBA - Mol. Basis Dis. 1993;1181(2):183–188.
- [120] Schols D, De Clercq E, Balzarini J, et al. Sulphated polymers are potent and selective inhibitors of various enveloped viruses, including herpes simplex virus, cytomegalovirus, vesicular stomatitis virus, respiratory syncytial virus, and toga-, arena- and retroviruses. Antivir Chem Chemother. 1990;1(4):233–240.
- [121] Danial M, Andersen AHF, Zuwala K, et al. Triple activity of lamivudine releasing sulfonated polymers against HIV-1. Mol Pharm. 2016;13(7):2397–2410.
- [122] Bianculli RH, Mase JD, Schulz MD. Antiviral polymers: past approaches and future possibilities. Macromolecules. 2020;53(21):9158–9186.
- [123] Caló E, Khutoryanskiy VV. Biomedical applications of hydrogels: a review of patents and commercial products. Eur Polym J. 2015;65:252–267.
- [124] Zubik K, Singhsa P, Wang Y, et al. Thermo-responsive poly(n-isopropylacrylamide)cellulose nanocrystals hybrid hydrogels for wound dressing. Polymers (Basel). 2017; 9(12):119.
- [125] Tian S, Liu G, Wang X, et al. PH-responsive tumor-targetable theranostic nanovectors based on core crosslinked (CCL) micelles with fluorescence and magnetic resonance (MR) dual imaging modalities and drug delivery performance. Polymers (Basel). 2016; 8(6):226.
- [126] Ghaeini-Hesaroeiye S, Razmi Bagtash H, Boddohi S, et al. Thermoresponsive nanogels based on different polymeric moieties for biomedical applications. Gels. 2020;6(3):20.
- [127] Chen J, Li G, Liu Q, et al. A photocleavable amphiphilic prodrug self-assembled nanoparticles with effective anticancer activity in vitro. Nanomaterials. 2019;9(6):860.
- [128] Shin JM, Choi GH, Song SH, et al. Metal-phenolic network-coated hyaluronic acid nanoparticles for PH-responsive drug delivery. Pharmaceutics. 2019;11(12):636.
- [129] Gandhi A, Paul A, Sen SO, et al. Studies on thermoresponsive polymers: phase behaviour, drug delivery and biomedical applications. Asian J Pharm Sci. 2015;10(2):99–107.
- [130] Ramos J, Imaz A, Forcada J. Temperature-sensitive nanogels: poly(n-vinylcaprolactam) versus poly(n-isopropylacrylamide). Polym Chem. 2012;3(4):852–856.
- [131] Zhang Q, Honko A, Zhou J, et al. Cellular nanosponges inhibit SARS-CoV-2 infectivity. Nano Lett. 2020;20(7):5570–5574.
- [132] Pulendran B, Ahmed R. Immunological mechanisms of vaccination. Nat Immunol. 2011;12(6):509–517.
- [133] Liu MA. The immunologist's grail: vaccines that generate cellular immunity. Proc Natl Acad Sci U S A. 1997;94(20):10496–10498.
- [134] Tahamtan A, Charostad J, Hoseini Shokouh SJ, et al. An overview of history, evolution, and manufacturing of various generations of vaccines. J Arch Mil Med. 2017; 5(e12315):1-7.
- [135] Finco O, Rappuoli R. Designing vaccines for the twenty-first century society. Front Immunol. 2014;5(Jan):12.
- [136] Dong Y, Dai T, Wei Y, et al. A systematic review of SARS-CoV-2 vaccine candidates. Sig Transduct Target Ther. 2020;5:237.
- [137] Yang J, Wang W, Chen Z, et al. A vaccine targeting the RBD of the S protein of SARS-CoV-2 induces protective immunity. Nature. 2020;586(7830):572–577.
- [138] Folegatti PM, Ewer KJ, Aley PK, et al. Safety and immunogenicity of the ChAdOx1 NCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, singleblind, randomised controlled trial. Lancet. 2020;396(10249):467–478.
- [139] Walsh EE, Frenck RW, Falsey AR, et al. Safety and immunogenicity of two RNAbased covid-19 vaccine candidates. N Engl J Med. 2020;383:2439–2450.
- [140] de Alwis R, Gan ES, Chen S, et al. A single dose of self-transcribing and replicating RNA based SARS-CoV-2 vaccine produces protective adaptive immunity in mice. Mol Ther. 2021 (In Press). doi: https://doi.org/10.1016/j.ymthe.2021.04.001.

- [141] Chandra Baray J, Maksudur Rahman Khan M, Mahmud A, et al. BANCOVID, the first D614G variant MRNA-based vaccine candidate against SARS-CoV-2 elicits neutralizing antibody and balanced cellular immune response. bioRxiv. 2020.
- [142] Jackson LA, Anderson EJ, Rouphael NG, et al. An MRNA vaccine against SARS-CoV-2—preliminary report. N Engl J Med. 2020;383:1920–1931.
- [143] Rauch S, Roth N, Schwendt K, et al. mRNA based SARS-CoV-2 vaccine candidate CVnCoV induces high levels of virus neutralizing antibodies and mediates protection in rodents. bioRxiv. 2020. doi: https://doi.org/10.1101/2020.10.23.351775
- [144] Shao L, Wu WS. Gene-delivery systems for IPS cell generation. Expert Opin Biol Ther. 2010;10(2):231-242.
- [145] Ullah I, Subbarao RB, Rho GJ. Human mesenchymal stem cells current trends and future prospective. Biosci Rep. 2015;35(2):191.
- [146] Mahalingam S, Ng WH, Liu X. Development of vaccines for SARS-CoV-2. F1000Res. 2020;9:991.
- [147] Logunov DY, Dolzhikova IV, Zubkova OV, et al. Safety and immunogenicity of an RAd26 and RAd5 vector-based heterologous prime-boost COVID-19 vaccine in two formulations: two open, non-randomised phase 1/2 studies from Russia. Lancet. 2020; 396(10255):887–897.
- [148] Sadoff J, Le Gars M, Shukarev G, Heerwegh, et al. Interim results of a phase 1–2a trial of Ad26.COV2.S covid-19 vaccine. N Engl J Med. 2021.
- [149] Anywaine Z, Whitworth H, Kaleebu P, et al. Safety and immunogenicity of a 2-dose heterologous vaccination regimen with Ad26.ZEBOV and MVA-BN-filo ebola vaccines: 12-month data from a phase 1 randomized clinical trial in Uganda and Tanzania. J Infect Dis. 2019;220(1):46–56.
- [150] Guebre-Xabier M, Patel N, Tian J-H, et al. NVX-CoV2373 vaccine protects cynomolgus macaque upper and lower airways against SARS-CoV-2 challenge. Vaccine. 2020; 38(50):7892–7896.
- [151] Zimmer C, Corum J, Wee S-L. Covid-19 vaccine tracker updates: the latest. The New York Times [accessed Mar 9, 2021]. https://www.nytimes.com/interactive/2020/science/ coronavirus-vaccine-tracker.html.
- [152] Pandey SC, Pande V, Sati D, et al. Vaccination strategies to combat novel corona virus SARS-CoV-2. Life Sci. 2020;256:117956.
- [153] Brisse M, Vrba SM, Kirk N, et al. Emerging concepts and technologies in vaccine development. Front Immunol. 2020;11:583077.
- [154] Li J, Xue S, Mao ZW. Nanoparticle delivery systems for SiRNA-based therapeutics. J Mater Chem B. 2016;4(41):6620-6639.
- [155] Kalam MA, Khan AA, Alshamsan A. Non-invasive administration of biodegradable nano-carrier vaccines. Am J Transl Res. 2017;9(1):15–35.
- [156] Chauhan G, Madou MJ, Kalra S, et al. Nanotechnology for COVID-19: therapeutics and vaccine research. ACS Nano. 2020;14(7):7760–7782.
- [157] Ghitman J, Biru EI, Stan R, et al. Review of hybrid PLGA nanoparticles: future of smart drug delivery and theranostics medicine. Mater Des. 2020;193:108805.
- [158] Catoira MC, Fusaro L, Di Francesco D, et al. Overview of natural hydrogels for regenerative medicine applications. J Mater Sci Mater Med. 2019;30(10):1–10.
- [159] Afewerki S, Sheikhi A, Kannan S, et al. Gelatin-polysaccharide composite scaffolds for 3D cell culture and tissue engineering: towards natural therapeutics. Bioeng Transl Med. 2019;4(1):96–115.
- [160] Galloway AL, Murphy A, DeSimone JM, et al. Development of a nanoparticle-based influenza vaccine using the PRINT technology. Nanomedicine. 2013;9(4):523–531.
- [161] Pardi N, Hogan MJ, Porter FW, et al. MRNA vaccines a new era in vaccinology. Nat Rev Drug Discov. 2018;17(4):261–279.
- [162] Lambricht L, Lopes A, Kos S, et al. Clinical potential of electroporation for gene therapy and DNA vaccine delivery. Expert Opin Drug Deliv. 2016;13(2):295–310.

- [163] Zhu M, Wang R, Nie G. Applications of nanomaterials as vaccine adjuvants. Hum Vaccin Immunother. 2014;10(9):2761–2774.
- [164] Bonam SR, Kotla NG, Bohara RA, et al. Potential immuno-nanomedicine strategies to fight COVID-19 like pulmonary infections. Nano Today. 2021;36:101051.
- [165] Kaur SP, Gupta V. COVID-19 vaccine: a comprehensive status report. Virus Res. 2020;288:198114.
- [166] Keech C, Albert G, Cho I, et al. Phase 1–2 trial of a SARS-CoV-2 recombinant spike protein nanoparticle vaccine. N Engl J Med. 2020;383:2320–2332.
- [167] Irvine DJ, Hanson MC, Rakhra K, et al. Synthetic nanoparticles for vaccines and immunotherapy. Chem Rev. 2015;115(19):11109–11146.
- [168] Aucouturier J, Dupuis L, Deville S, et al. Montanide ISA 720 and 51: a new generation of water in oil emulsions as adjuvants for human vaccines. Expert Rev Vaccines. 2002; 1(1):111–118.
- [169] Chang JCC, Diveley JP, Savary JR, et al. Adjuvant activity of incomplete freund's adjuvant. Adv Drug Deliv Rev. 1998;32(3):173–186.
- [170] Newman MJ, Balusubramanian M, Todd CW. Development of adjuvant-active nonionic block copolymers. Adv Drug Deliv Rev. 1998;32(3):199–223.
- [171] Huang MH, Huang CY, Lien SP, et al. Development of multi-phase emulsions based on bioresorbable polymers and oily adjuvant. Pharm Res. 2009;26(8):1856–1862.
- [172] Haun BK, Lai C-Y, Williams CA, et al. CoVaccine HT<sup>TM</sup> adjuvant potentiates robust immune responses to recombinant SARS-CoV-2 spike S1 immunization. Front Immunol. 2020;11:599587.
- [173] Kusi KA, Remarque EJ, Riasat V, et al. Safety and immunogenicity of multi-antigen AMA1-based vaccines formulated with CoVaccine HT<sup>TM</sup> and Montanide ISA 51 in rhesus macaques. Malar J. 2011;10(1):182.
- [174] Lakhan N, Stevens NE, Diener KR, et al. CoVaccine HT<sup>TM</sup> adjuvant is superior to Freund's adjuvants in eliciting antibodies against the endogenous alarmin HMGB1. J Immunol Methods. 2016;439:37–43.
- [175] Ye Q, Wang B, Mao J. The pathogenesis and treatment of the 'cytokine storm' in COVID-19. J Infect. 2020;80(6):607–613.
- [176] Nikolova MP, Chavali MS. Recent advances in biomaterials for 3D scaffolds: a review. Bioact Mater. 2019;4:271–292.
- [177] Wu Z, Kong B, Liu R, et al. Engineering of corneal tissue through an aligned PVA/ collagen composite nanofibrous electrospun scaffold. Nanomaterials. 2018;8(2):124.
- [178] Zhang Y, Wang C, Fu L, et al. Fabrication and application of novel porous scaffold in situ-loaded graphene oxide and osteogenic peptide by cryogenic 3D printing for repairing critical-sized bone defect. Molecules. 2019;24(9):1–20.
- [179] Wang L, Dong S, Liu Y, et al. Fabrication of injectable, porous hyaluronic acid hydrogel based on an in-situ bubble-forming hydrogel entrapment process. Polymers (Basel). 2020;12(5):1-15.
- [180] Andrade CF, Wong AP, Waddell TK, et al. Cell-based tissue engineering for lung regeneration. Am J Physiol Lung Cell Mol Physiol. 2007;292(2):L510–L518.
- [181] Shannon JM, McCormick-Shannon K, Burhans MS, et al. Chondroitin sulfate proteoglycans are required for lung growth and morphogenesis in vitro. Am J Physiol Cell Mol Physiol. 2003;285(6):L1323–L1336.
- [182] Guo B, Ma PX. Synthetic biodegradable functional polymers for tissue engineering: a brief review. Sci China Chem. 2014;57(4):490–500.
- [183] Mohammadi Nasr S, Rabiee N, Hajebi S, et al. Biodegradable nanopolymers in cardiac tissue engineering: from concept towards nanomedicine. Int J Nanomed. 2020;15: 4205–4224.
- [184] Zhu J, Marchant RE. Design properties of hydrogel tissue-engineering scaffolds. Expert Rev Med Devices. 2011;8(5):607–626.
- [185] Tebyanian H, Karami A, Nourani MR, et al. Lung tissue engineering: an update. J Cell Physiol. 2019;234(11):19256–19270.

- [186] Shigemura N, Okumura M, Mizuno S, et al. Lung tissue engineering technique with adipose stromal cells improves surgical outcome for pulmonary emphysema. Am J Respir Crit Care Med. 2006;174(11):1199–1205.
- [187] Deng M, Gu Y, Liu Z, et al. Endothelial differentiation of human adipose-derived stem cells on polyglycolic acid/polylactic acid mesh. Stem Cells Int. 2015;2015:1–11.
- [188] Pappalardo D, Mathisen T, Finne-Wistrand A. Biocompatibility of resorbable polymers: a historical perspective and framework for the future. Biomacromolecules. 2019; 20(4):1465–1477.
- [189] Yang R, Tan L, Cen L, et al. An injectable scaffold based on crosslinked hyaluronic acid gel for tissue regeneration. RSC Adv. 2016;6(20):16838-16850.
- [190] George PM, Barratt SL, Condliffe R, et al. Respiratory follow-up of patients with COVID-19 pneumonia. Thorax. 2020;75(11):1009–1016.
- [191] Sadikot RT, Kolanjiyil AV, Kleinstreuer C, et al. Nanomedicine for treatment of acute lung injury and acute respiratory distress syndrome. Biomed Hub. 2017;2(2):1–12.
- [192] Sadikot RT. Peptide nanomedicines for treatment of acute lung injury. In Methods in enzymology. Vol. 508. Academic Press Inc.; 2012, 508:315–324.
- [193] Patra JK, Das G, Fraceto LF, et al. Nano based drug delivery systems: recent developments and future prospects. J Nanobiotechnology. 2018;16(1):71.
- [194] Weiss C, Carriere M, Fusco L, et al. Toward nanotechnology-enabled approaches against the COVID-19 pandemic. ACS Nano. 2020;14(6):6383–6406.
- [195] Santos I, de A, Grosche VR, Bergamini FRG, et al. Antivirals against coronaviruses: candidate drugs for SARS-CoV-2 treatment? Front Microbiol. 2020;11:1818.
- [196] Itani R, Tobaiqy M, Faraj A. Al. Optimizing use of theranostic nanoparticles as a lifesaving strategy for treating COVID-19 patients. Theranostics. 2020;10(13):5932–5942.
- [197] Liu Y, Hardie J, Zhang X, et al. Effects of engineered nanoparticles on the innate immune system. Semin Immunol. 2017;34:25–32.
- [198] Richtering W, Alberg I, Zentel R. Nanoparticles in the biological context: surface morphology and protein corona formation. Small. 2020;16(39):2002162.
- [199] Wolfram J, Zhu M, Yang Y, et al. Safety of nanoparticles in medicine. CDT. 2015; 16(14):1671–1681.
- [200] Li J, Mooney DJ. Designing hydrogels for controlled drug delivery. Nat. Rev. Mater. 2016;1:16071.
- [201] Ashammakhi N, Ahadian S, Xu C, et al. Bioinks and bioprinting technologies to make heterogeneous and biomimetic tissue constructs. Mater Today Bio. 2019;1:100008.
- [202] Annabi N, Nichol JW, Zhong X, et al. Controlling the porosity and microarchitecture of hydrogels for tissue engineering. Tissue Eng Part B Rev. 2010;16(4):371–383. [InsertedFromOnline[pubmedMismatch]]