

# LXRs, SHP, and FXR in Prostate Cancer: Enemies or *Ménage à Quatre* With AR?

Marica Cariello<sup>1\*</sup>, Simon Ducheix<sup>2\*</sup>, Salwan Maqdas<sup>3,4,5</sup>,  
Silvère Baron<sup>3,4</sup>, Antonio Moschetta<sup>1,2,6\*\*</sup>,  
and Jean-Marc A. Lobaccaro<sup>1,2,3,4\*\*</sup>

Nuclear Receptor Signaling  
Volume 15: 1–10  
© The Author(s) 2018  
Article reuse guidelines:  
sagepub.com/journals-permissions  
DOI: 10.1177/1550762918801070  
journals.sagepub.com/home/nrs



## Abstract

Androgens and androgen receptor (AR, NR3C4) clearly play a crucial role in prostate cancer progression. Besides, the link between metabolic disorders and the risk of developing a prostate cancer has been emerging these last years. Interestingly, “lipid” nuclear receptors such as LXR $\alpha$ /NR1H3 and LXR $\beta$ /NR1H2 (as well as FXR $\alpha$ /NR1H4 and SHP/NR0B2) have been described to decrease the lipid metabolism, while AR increases it. Moreover, these former orphan nuclear receptors can regulate androgen levels and modulate AR activity. Thus, it is not surprising to find such receptors involved in the physiology of prostate. This review is focused on the roles of liver X receptors (LXRs), farnesoid X receptor (FXR), and small heterodimeric partner (SHP) in prostate physiology and their capabilities to interfere with the androgen-regulated pathways by modulating the levels of active androgen within the prostate. By the use of prostate cancer cell lines, mice deficient for these nuclear receptors and human tissue libraries, several authors have pointed out the putative possibility to pharmacologically target these receptors. These data open a new field of research for the development of new drugs that could overcome the castration resistance in prostate cancer, a usual phenomenon in patients.

## Keywords

prostate cancer, LXR, FXR, SHP, lipid metabolism

Received: June 20, 2017; Accepted: January 3, 2018

## Abbreviations

ABCA1, ATP-binding cassette A1; ADT, androgen deprivation therapy; AKT/PKB, protein kinase B; AMACR,  $\alpha$ -methylacyl CoA racemase; AP-1, activator protein-1; AR, androgen receptor; CDKN1A/p21CIP1, cyclin-dependent kinase inhibitor 1; CDKN1B/p27KIP1, cyclin-dependent kinase inhibitor 1B; CRPC, castration-resistant prostate cancer; DHT, dihydrotestosterone; ERK1/2, extracellular signal-regulated kinases; FXR, farnesoid X receptor; LXR, liver X receptor; MAPK, mitogen-activated protein kinase; MTOR/mTOR, mammalian target of rapamycin; NF $\kappa$ B1/p105, nuclear factor kappa-light-chain-enhancer of activated B cells; OATP, organic-anion-transporting polypeptide; PCa, prostate cancer; PI3K, phosphatidylinositol-4,5-bisphosphate 3-kinase; PSA, prostate-specific antigen; PTEN, Phosphatase and TENsin homolog; SRC/Src, proto-oncogene tyrosine-protein kinase; SHP, small heterodimeric partner; SLCO1B3/OATP1B3, solute carrier organic anion transporter family member 1B3; SLiMs, selective liver X receptor modulators; SOCS3, suppressor of cytokine signaling 3; SREBF/SREBP, sterol regulatory element-binding protein; UGT2, UDP glucuronosyltransferase 2.

## Introduction

### *Prostate Cancer at a Glance*

Even though the Greek physician Herophilos (335-280 BC) was the first to describe the prostate in man according to Galen of Pergamon (129-216 AC), the first surgical case of prostate cancer (PCa) was documented by George Langstaff

<sup>1</sup>“Aldo Moro” University of Bari, Italy

<sup>2</sup>Istituto Nazionale Biostrutture e Biosistemi, Roma, Italy

<sup>3</sup>Université Clermont Auvergne, Clermont-Ferrand, France

<sup>4</sup>Centre de Recherche en Nutrition Humaine d’Auvergne, Clermont-Ferrand, France

<sup>5</sup>CHU Clermont-Ferrand, France

<sup>6</sup>IRCCS Istituto Oncologico “Giovanni Paolo II,” Bari, Italy

\*M.C. and S.D. are equal first authors

\*\*A.M. and J.-M.A.L. are equal last authors

### Corresponding Author:

Jean-Marc A. Lobaccaro, Université Clermont Auvergne, GRéD, Faculté de Médecine, 28 Place Henri-Dunant, 63001 Clermont-Ferrand Cedex, France.

Email: j-marc.lobaccaro@uca.fr



in 1817 and histologically defined in 1853.<sup>1</sup> As expected, and despite the urban legend, PCa is not a modern phenomenon. Indeed, Ghabili et al<sup>1</sup> nicely reported that radiographic analyses of skeletons and of mummies pointed out that men have been affected by this tumor since stone and bronze ages. However, the reported prevalence was lower compared to the one of modern populations. Interestingly, PCa was also probably present in the Americas centuries prior to European colonization.<sup>1</sup>

Today, PCa, along with colorectal and breast cancer, displays a higher rate of prevalence in the developed countries, with about 6-fold difference in comparison with low-incidence countries. This increment has been linked to different risk factors and diagnostic practices.<sup>2</sup> Indeed, the incidence of PCa is constantly increasing due in part to new diagnostic methods,<sup>3</sup> to the increasing impact of prostate-specific antigen (PSA) testing, to the perceptions of PCa fear,<sup>4</sup> and also to the increase in life expectancy. For example, in high-income countries with a low and gradual increase in the rate of PSA testing, such as Japan or the United Kingdom, the prevalence of PCa continues to slightly increase.<sup>5</sup> However, the role of PSA testing in the reduction of PCa-related mortality rates at the population level is rather controverted.<sup>6,7</sup>

In contrary to some other cancers, PCa has a relatively slow evolution and about 85% of diagnosed PCa are in patients older than 65 years.<sup>8</sup> It is currently admitted that more men die with PCa rather than from it. Indeed, a study performed by Sakr et al<sup>9</sup> pointed out that 50% of the men of 50 years old have a latent PCa on autopsy analyses and that the initiating events leading to a clinically relevant PCa likely occur decades before. Nonetheless, the development and the etiology of the disease are still poorly understood, and various factors such as genetic/ethnic origin, diet, lifestyle, and environmental factors have been suggested to play a role.<sup>10</sup>

As already stated, great differences in the incidence of PCa have been observed depending on the ethnical origin or the country of the patients.<sup>11</sup> A Caucasian American has 30% less risk to develop a PCa compared with an African American,<sup>12,13</sup> but at the same time, Asians develop twice less PCa than Americans.<sup>14</sup> Yet the genetic background cannot explain everything because the first generation of immigrants from Asia living in the United States has a more important risk of PCa than those living in Asia.<sup>15</sup> Among various factors putatively identified, higher lipid intake in the United States has been pointed out.<sup>16</sup> A comparable observation had been done years before by Shimizu et al<sup>17</sup> in the Japanese population that moved to America.

### *PCa, Also a Matter of Metabolic Disorder*

As enlightened by previous epidemiological studies describing the link between the high lipid intake in Western countries and the risk of developing a PCa, the high prevalence of obesity and metabolic syndrome is associated with worse oncological outcomes in men with PCa<sup>18</sup>; the tumor is more

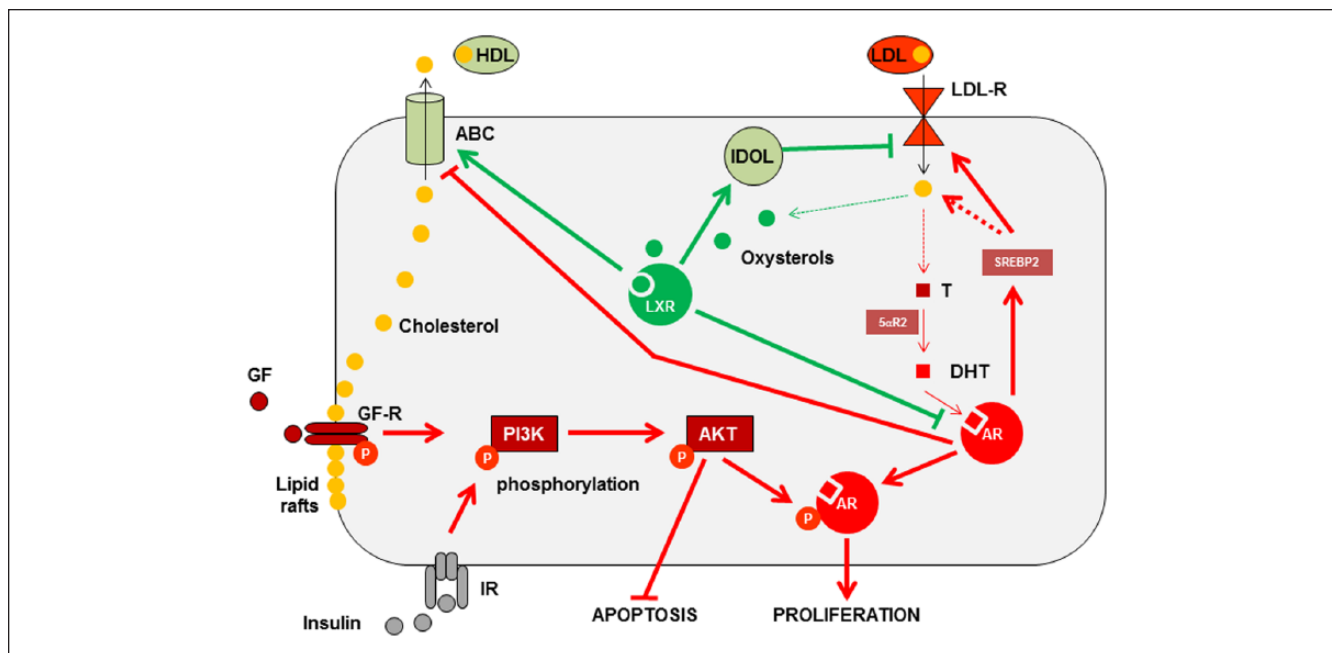
aggressive and the biochemical recurrence rate is higher.<sup>19</sup> In direction with this link, metformin, an antidiabetic drug that leads to significant improvement of metabolic syndrome parameters, has been shown to induce apoptosis in PCa cells.<sup>20,21</sup> However, the use of metformin for the treatment of patients with PCa is still a matter of debate.<sup>22</sup>

A high lipogenesis has also been associated to PCa, as it supplies the tumor with key membrane components such as phospholipids and cholesterol. Indeed, cancer cells being characterized by a higher rate of multiplication need to abundantly build membrane for that.<sup>23</sup> Indeed, Swinnen's group has been among the first to propose pharmacological inhibition of lipogenesis to induce apoptosis in cancer cell lines<sup>24</sup> and to reduce tumor growth in xenograft models, eg, by targeting squalene synthase by zaragozic acid<sup>25</sup> or by blocking acetyl-CoA carboxylases<sup>26</sup> using soraphen A.

Last but not least, cholesterol imbalance has been pointed out in PCa. Cholesterol accumulation in tumors is not a recent observation. White demonstrated in 1909 an "accumulation of crystals of lipid nature in tumors" and suggested that "cholesterol might be associated in some way with the regulation of cell proliferation."<sup>27</sup> Such cholesterol accumulation was also observed later on in skin cancer.<sup>28</sup> Then Swyer showed for the first time an increase of cholesterol content by 2-fold in a zone of the prostate affected by a hypertrophy compared with the surrounding healthy tissues.<sup>29</sup> Two mechanisms are put forward to explain the intracellular cholesterol accumulation: a higher circulating cholesterol uptake, and an increase in the accumulation of the mevalonate pathway enzymes.<sup>30,31</sup>

Yue et al identified aberrant accumulation of esterified cholesterol in lipid droplets of high-grade PCa and metastases using imaging data.<sup>32</sup> The authors showed that such cholesteryl ester accumulation was a consequence of loss of the tumor suppressor Phosphatase and TENSin homolog (PTEN), one of the most common genetic events in PCa, and thus subsequent activation of the phosphatidylinositol-4,5-bisphosphate 3-kinase (PI3K)/protein kinase B (PKB/AKT) pathway in PCa cells. In response to the loss of the PTEN, they identified the activation of the mammalian target of rapamycin (MTOR/mTOR) pathway and downstream activation of sterol regulatory element-binding protein (SREBF/SREBP), and the upregulation of LDL receptor (LDL-R<sup>32</sup>). However, blockade of mTOR by analogs of rapamycin has been shown to be inefficient in castration-resistant prostate cancer (CRPC) so far,<sup>33</sup> but more promising in radioresistant PCa.<sup>34</sup>

Hence, despite a higher accumulation of cholesterol within the tumor has been well demonstrated in PCa, no clear link has been however made between circulating cholesterol levels and high Gleason score, positive nodal status, and positive surgical margins.<sup>35</sup> Nevertheless, it has been tempted to test various compounds decreasing the levels of cholesterol in androgen-dependent or -independent cancer cell cultures or in animal models.<sup>36-41</sup> Altogether, sufficient data are lacking to support the use of statins for the primary



**Figure 1.** Interconnection between LXRs and AR in prostate cell.

*Note.* AR and DHT increase the proliferation. When growth factors bind their membrane receptors, they activate phosphorylation cascades, stimulate PI3K/AKT, and increase AR activity through its phosphorylation. Oxysterol-activated LXR induces ubiquitin ligase IDOL accumulation followed by degradation of LDL-R. ABC proteins increase both export of cholesterol and destructuration of lipid rafts, which in turn will decrease both AKT phosphorylation and inhibition of the apoptotic pathway. Green lines represent favorable effects on PCa management; red lines represent negative effects on PCa management. LXRs = liver X receptors; AR = androgen receptor; DHT = dihydrotestosterone; PI3K = phosphatidylinositol-4,5-bisphosphate 3-kinase; AKT = protein kinase B; LDL-R = LDL receptor; ABC = ATP-binding cassette; PCa = prostate cancer; T = testosterone; GF = growth factors; GF-R = growth factor receptor; P = phosphorylation; IR = insulin receptor; HDL = high density lipoprotein.

prevention of PCa. Meanwhile, statins have been associated with improved PCa-specific survival, particularly in men undergoing radiotherapy, suggesting usefulness of statins in secondary and tertiary prevention.<sup>42</sup> Yet more epidemiological and mechanistic studies are needed to eventually use statins in PCa.<sup>43</sup>

### Androgens and Androgen Receptor Control PCa Progression

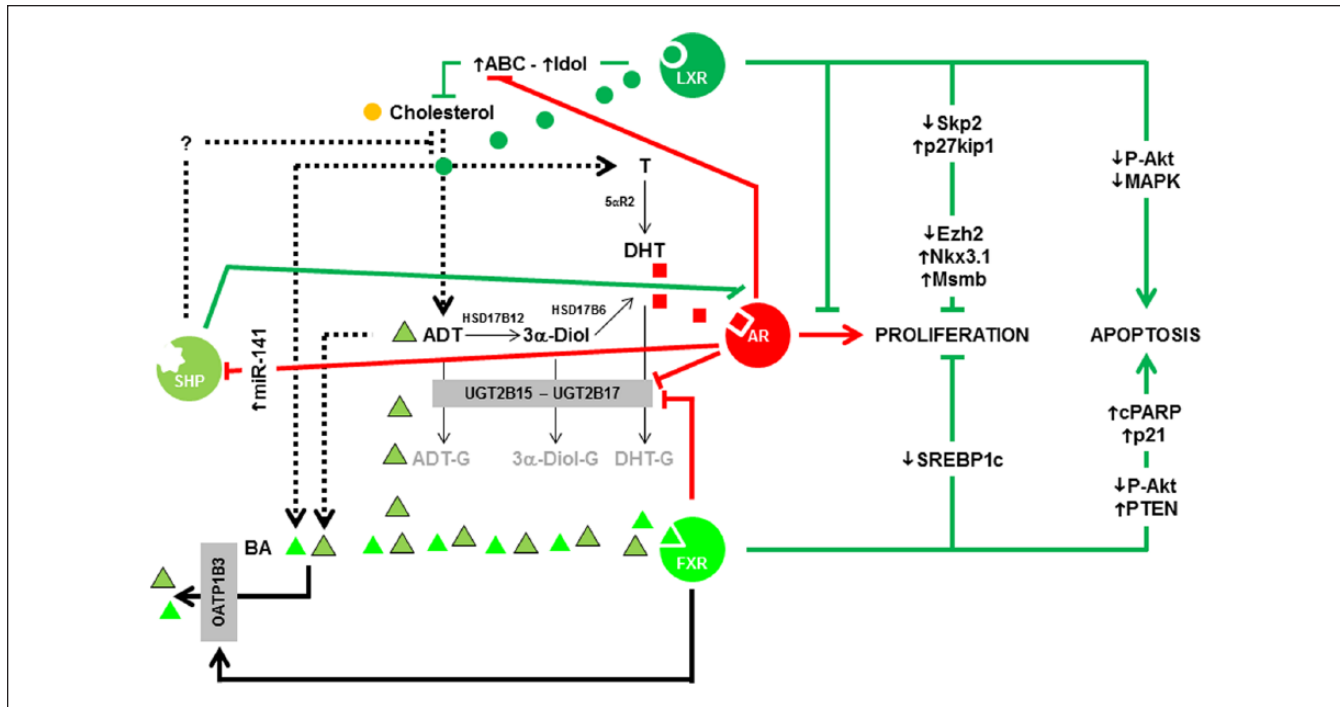
Androgens and androgen receptor (AR, NR3C4) play a crucial role in PCa. Indeed, since Huggins and Hodges' *princeps* article in 1941,<sup>44</sup> it has been admitted that PCa is driven by androgen levels and androgen activity through AR transcriptional regulation.

Among the various androgens, dihydrotestosterone (DHT) is the most active on AR to induce cell proliferation. DHT is synthesized from testosterone by 5 $\alpha$ -reductase 2 (SRD5A2) (Figure 1), which can be targeted in PCa by dutasteride, a 5 $\alpha$ -reductase inhibitor, despite some controversies.<sup>45</sup> DHT could also be synthesized from androsterone and 5 $\alpha$ -androstane-3 $\alpha$ ,17 $\beta$ -diol even in small amounts. On the opposite, androsterone, 5 $\alpha$ -androstane-3 $\alpha$ ,17 $\beta$ -diol, and DHT can be transformed in nonactive glucuronides by UDP glucuronosyltransferase 2 (UGT2) B15 and B17.<sup>46</sup> Note that

AR downregulates the expression of *UGT2B15/17* (Figure 2), thus decreasing the inactivation of androgens.<sup>47</sup>

Hence, PCa is initially an androgen-dependent disease; unfortunately, about 30% of patients will relapse after primary therapy.<sup>48</sup> Androgen deprivation therapy (ADT), by surgical or biochemical castration, is the main treatment for relapsed patients and provides temporary relief to tumor burden.<sup>49</sup> This step is performed by downregulating androgen production by using steroid synthesis inhibitors, as well as antiandrogens, that act at the level of AR.<sup>49</sup> However, most of the PCa will dedifferentiate to CRPC and inevitably will develop a more aggressive and metastatic cancer. Despite approved treatment options for this PCa stage (eg, taxane compounds; abiraterone that inhibits 17 $\alpha$ -hydroxylase involved in androgen synthesis; enzalutamide that blocks AR; sipuleucel-T, a therapeutic vaccine), drug resistance will eventually develop in few months.

The exact mechanism of transition from castration sensitive PCa to castration-resistant disease is still not fully understood, but AR definitely plays a key role as described by Bevan's group<sup>50</sup>; several mechanisms could be cited, among them an increased number of the AR encoding gene copies making them more sensitive to lower levels of androgens, development of cellular clones harboring mutations within the ligand-binding domain of AR and hence potentially activated



**Figure 2.** Summary of the interconnections among AR, LXRs, FXR, and SHP in prostate cell.

Note. AR and DHT have a central role in the proliferation of the epithelial cells. LXRs, FXR, and SHP have positive impacts in PCa by blocking AR transcriptional activity, decreasing proliferation and/or increasing apoptosis. FXR could also play a negative role by decreasing glucuronidation of androgens through the transcriptional regulation of UGT2B15/17 enzymes. Green lines represent favorable effects on PCa management; red lines represent negative effects on PCa management. See the article for more details about the various links. AR = androgen receptor; LXRs = liver X receptors; FXR = farnesoid X receptor; SHP = small heterodimeric partner; DHT = dihydrotestosterone; PCa = prostate cancer; MAPK = mitogen-activated protein kinase; ADT = androgen deprivation therapy; UGT2 = UDP glucuronosyltransferase 2; SREBP = sterol regulatory element-binding protein; PTEN = Phosphatase and TENsin homolog; ABC = ATP-binding cassette; BA = bile acid; cPARP = cleaved Poly (ADP-ribose) polymerase.

by steroids that usually do not bind AR, and modifications of AR coactivators or corepressors. Hence, despite an extremely low level of circulating androgens, AR remains active and continues to drive PCa progression.

Beside the classical ligand-regulation of the transcriptional activity, AR could also rapidly interact with the nonreceptor tyrosine kinase SRC/Src increasing cell proliferation through activation of the mitogen-activated protein kinase (MAPK)/extracellular signal-regulated kinases (ERK1/2) cascade,<sup>51</sup> or with PI3K/AKT signaling pathway (Figure 1), and controlling cell survival.<sup>52</sup> Based on that, innovative therapies will be necessary to counteract this so-called non-genomic signaling of AR during the establishment of metastatic CRPC.<sup>53</sup>

Finally, “androgens meet lipids” (Figure 1) as AR activation increases fatty acid synthesis<sup>23,54-56</sup> and SREBP2,<sup>30</sup> a key-player in de novo synthesis of cholesterol and its cellular uptake,<sup>57</sup> and decreases ATP-binding cassette A1 (ABCA1), a cholesterol export pump.<sup>58</sup> This last point is crucial because cholesterol is an obligatory precursor for testosterone and DHT synthesis, the only two active androgens on AR<sup>59</sup>; more importantly, tumor cells also have the ability to abnormally synthesize DHT from cholesterol<sup>60</sup> or from adrenal androgens.<sup>61,62</sup>

Together with AR, other nuclear receptors have been involved in PCa (for a review, see Leach et al<sup>50</sup>); among them the popular “*Ménage-à-trois*” LXR-FXR-SHP (liver X receptor–farnesoid X receptor–small heterodimeric partner) has been described to be the major player in the regulation of both cholesterol and bile acid homeostasis.<sup>63</sup> Altogether, targeting this new “*Ménage-à-quatre*” appears to be of importance to take care of the prostate. Interestingly, the role of these 3 nuclear receptors has been emerging these last years in ex vivo or in vivo experiments, especially with the generation of transgenic animals knock-out for LXRs. Noteworthy, The Cancer Genome Atlas pointed out that these nuclear receptors could present alterations of copy numbers of their respective encoding genes in PCa.<sup>50</sup>

### LXR $\alpha$ and LXR $\beta$ Are Involved in Prostate Physiology

LXR $\alpha$ /NR1H3 and LXR $\beta$ /NR1H2 (as well as FXR $\alpha$ /NR1H4 and SHP/NR0B2) are members of the nuclear receptor superfamily. They are composed of several functional domains, among them a central DNA-binding domain and a C-terminal ligand-binding domain.<sup>64</sup> At the end of the 1990s, Janowski et al demonstrated that LXRs are the bona fide



receptors for oxysterols,<sup>65,66</sup> oxidized derivatives of cholesterol. Hence, it was suggested that LXRs could regulate cholesterol homeostasis in the cell and was demonstrated thanks to the analysis of *Lxr*-deficient mice.<sup>67</sup> Since this seminal article, others groups have associated LXR roles to numerous physiological functions.<sup>64,68</sup>

In the prostate, Liao's group was the first to evoke a putative positive role of LXRs in PCa.<sup>58</sup> The authors showed that *ABCA1*, a bona fide LXR-target gene which increases cholesterol export, was downregulated by androgens (Figure 2) in LNCaP cells.<sup>58</sup> The same group identified that activation of LXRs by the synthetic agonist T0901317 decreased the percentage of S-phase LNCaP cells in a dose-dependent manner and increased the expression of cyclin-dependent kinase inhibitor CDKN1B/p27KIP1<sup>58</sup>, by decreasing the S-phase kinase associated protein 2 (SKP2) involved in the degradation of cell cycle inhibitors.<sup>69</sup> At last, Chuu et al demonstrated that LXRs and some of their target genes were decreased during the progression of androgen-dependent tumor to androgen-independent relapsed tumors in a xenograft model.<sup>70</sup> These data thus made a clear link between LXRs and the proliferative capacities of PCa cells (Figure 2). Likewise, we identified that *Lxr $\alpha$ ; $\beta$* -deficient mice fed a high cholesterol diet presented prostatic intraepithelial neoplasia characterized by an accumulation of the oncogene and histone methyl transferase Enhancer of Zeste Homolog 2 (*EZH2*) which results in the downregulation of the tumor suppressors microseminoprotein beta (*MSMB*) and NK3 homeobox 1 (*NKX3.1*).<sup>71</sup> It is noteworthy that overexpression of *EZH2* has been described in patients with an aggressive PCa.<sup>72</sup> *EZH2* controls prostate cell proliferation through the epigenetic silencing of *NKX3.1*<sup>73</sup> and *MSMB*.<sup>74</sup> In wild-type mice fed a high cholesterol diet, LXRs induce the transcription of Inducible Degradator of the LDL receptor MYLIP/IDOL (Figures 1 and 2), a ubiquitin ligase that targets LDLR, and of ABC transporters, altogether maintaining a controlled level of cholesterol and a low amount of *EZH2*.<sup>71</sup> So far, it has not been possible to dissociate the exact role of each LXR isoform as they both compensate each other.

In addition to the role of LXRs in the control of cellular cholesterol content and prostate cell proliferation, we showed that the activation of LXRs by various natural or synthetic ligands increases the level of apoptosis in LNCaP cells.<sup>75</sup> This phenomenon is linked to the presence of smaller and thinner lipid rafts after LXR stimulation and the downregulation of AKT phosphorylation in these lipid rafts. After having derived new models of epithelial cells from the dorsal prostate (MPECs) of wild-type or *Lxr*-deficient mice, Dufour et al enlightened that LXRs modulate AKT and MAPK phosphorylation accumulation, making them potential mediators of LXRs in cell cycle control<sup>76</sup> (Figure 2). Altogether, these results confirm that LXRs are becoming exquisite pharmacological targets for PCa, unless specific modulators, we called SLiMs (selective LXR modulators<sup>77,78</sup>), could be developed.

On the other side, LXRs also control AR activity. Indeed, *Lxr*-deficient mice also develop benign prostatic hyperplasia (BPH).<sup>79</sup> In man, BPH is clearly due to an excessive activity of AR and the production of DHT.<sup>80</sup> Using transgenic animals, we were able to demonstrate that LXR $\alpha$  acts as a key modulator of the cross talk between the stromal and epithelial compartments, which is essential for the integration of androgen signaling in the prostate and its effect on the epithelium.<sup>81</sup> Interestingly, Tsui et al<sup>82</sup> pointed out that LXR expression was higher in androgen-sensitive LNCaP cells than in other PCa cell lines. Moreover, T0901317-activated LXRs decrease AR accumulation and PSA production in LNCaP. Overall, AR and LXRs are definitively interconnected (Figure 1). These data open numerous opportunities to develop new therapeutic concepts especially in CRPC situations where AR activity could become independent of androgen levels. If LXRs negatively modulate AR accumulation, a SLiM could increase PCa cell apoptosis, as well as decrease AR activity, hence, bypassing the castration-resistant stage.

One of the most challenging issues in PCa is to slow down the metastatic potential of the tumor.<sup>83</sup> Fu et al described an interesting effect of GW3965, another LXR synthetic ligand, on LNCaP cells. Activation of LXR increases the suppressor of cytokine signaling 3 (SOCS3) accumulation, followed by a decrease of phosphorylated AKT, activator protein-1 (AP-1), and nuclear factor kappa-light-chain-enhancer of activated B cells (NF $\kappa$ B1/p105).<sup>84</sup> GW3965 also inhibited PCa invasion in xenografted mice, suggesting that LXRs could be targetable to prevent metastases.

However, these potential effects of LXRs in PCa treatment should be modulated. Indeed, it should be kept in mind that, in prostate, tumor cells are surrounded by immune cells such as dendritic cells that initiate immune responses, including antitumor activity after their CC chemokine receptor-7 (CCR7)-dependent migration to lymphoid organs. Activation of LXR $\alpha$  inhibits CCR7 expression dampening the antitumor immune responses.<sup>85</sup> Likewise, drugs inhibiting cholesterol synthesis (and thus LXR ligands), such as zaragozic acid, increase the efficacy of the treatments in xenografted mice with tumors.<sup>86</sup> Once again, these contradictory results enlighten the necessity to develop tissue- and cell-specific LXR ligands to decipher the exact roles of each LXR in each cell type. It should also be noticed that no variation of LXR $\alpha$  and/or LXR $\beta$  expression has been linked to cancer grades. This would imply to stratify patients' cohorts, which has not been performed so far.

## Putative Role of FXR in PCa

FXR/NR1H4 is the nuclear receptor for bile acids.<sup>87</sup> However, the inactive androgen androsterone has also been described as a potent activator of FXR.<sup>88</sup> This point is important because androsterone is present in prostate. Conversely, to LXR and the historical story of cholesterol accumulation

in prostate tumors, the link between FXR, bile acids, and prostate physiology is less evident.

The first “historical” link came with Wang and Schaffner<sup>89</sup> who observed that BIO 87-20 hamsters, developing spontaneous cystic prostate hypertrophy, had a lower prostate size and weight with much less distended prostatic acini when were treated with colestipol, a bile-acid-sequestering anion-exchange resin. More recently, it has been shown that bile acid content is increased in patients with a PCa treated with an ADT<sup>90</sup>; in parallel, ADT has been associated with an increased risk of diverse biliary diseases.<sup>91</sup> Alpha-methylacyl CoA racemase (AMACR) is overexpressed in PCa<sup>92</sup> and is a better diagnostic marker than PSA. AMACR plays a key role in the  $\beta$  oxidation of branched chain fatty acids and the bile acid intermediates dihydroxycholestanic acid and trihydroxycholestanic acid. Furthermore, AMACR is highly expressed in androgen-sensitive PCa cell lines and is required for the proliferation of these cells.<sup>93</sup>

Even though lithocholic acid selectively induces apoptosis in androgen-sensitive and -insensitive prostate cell lines,<sup>94,95</sup> few studies have focused on the molecular effects of FXR activation in androgen-sensitive or androgen-insensitive prostate cell lines. Indeed, FXR inhibits cell proliferation by decreasing lipid metabolism via targeting SREBP1c (Figure 2).<sup>96</sup> Likewise, chenodeoxycholic acid (CDCA, a natural FXR ligand) and GW4065 (a synthetic FXR agonist) decrease LNCaP cell proliferation by the induction of *PTEN* accumulation, which in turn decreases the phosphorylation of AKT and the survival pathway.<sup>97</sup> Interestingly, new derivatives of ursodeoxycholic acid and CDCA induce the apoptosis of human prostate androgen-insensitive carcinoma PC-3 cells by increasing cyclin-dependent kinase inhibitor 1 CDKN1A/p21CIP1 and the cleaved form of poly [ADP-ribose] polymerase 1 PARP1.<sup>98</sup> However, it has not been proved yet that FXR mediates the effect of these molecules. Finally, the most important point is that FXR accumulation was found to be significantly lower, at both mRNA and protein levels, in human PCa tissues compared with the pair-matched adjacent normal tissues.<sup>97</sup> Again, no correlation was made with grades/stages of the tumors by the authors. Based on these results, one can suggest that FXR ligands could have some benefit in the treatment of PCa.

Unfortunately, other data showed a negative role of bile acids and FXR in the development of PCa. First of all, androgen metabolite androsterone, which is also an activator of FXR, reduces the glucuronidation of androgens catalyzed by UGT2B15/B17 in an FXR-dependent manner in LNCaP cells.<sup>99</sup> Such an action would increase the levels of androsterone, 5 $\alpha$ -androstane-3 $\alpha$ ,17 $\beta$ -diol, and finally DHT, and thus activate AR and induce cell proliferation. On the contrary, the increase of androsterone could also neutralize the AR-induced proliferation by blocking the proliferation via FXR (Figure 2). This dual paradoxical effect of FXR is also seen in the regulation of the solute carrier organic anion transporter family member 1B3 SLC01B3/OATP1B3, which is an export

pump for steroids and bile acids<sup>100</sup> and seems to be upregulated by FXR in the prostate,<sup>101</sup> but not in the liver.<sup>102</sup> If this regulation is efficient in the prostate, FXR would deplete the cells in steroids (including bile acids) by increasing OATP1B3; bile acid depletion will act as a safety valve and will reduce FXR activity. More investigations are thus needed using adequate in vivo models to understand how FXR targeting could be interesting in PCa management.

## SHP, a Noncanonical Nuclear Receptor With Significant Potential in PCa

Short heterodimer partner (SHP/*NR0B2*) is an atypical orphan nuclear receptor: It lacks the classical DNA- and possesses a ligand-binding domain. Despite its strong repressive activity on other nuclear receptors such as AR, no known ligand for SHP has been identified so far.<sup>103</sup>

Initially described as downregulating bile acid synthesis in liver by decreasing 7 $\alpha$ -cholesterol hydroxylase CYP7A1 in an LXR-dependent manner,<sup>104</sup> SHP became a member of the Triad with LXR and FXR when it appeared that FXR was the nuclear receptor for bile acids<sup>105</sup> and SHP was one of its bona fide target genes.

Indeed, it was the discovery of synthetic SHP ligands that gave the opportunity to link SHP to PCa.<sup>106</sup> Some of these ligands had a strong inhibitory effect on proliferation and inducing effect on apoptosis in the PCa androgen-insensitive DU-145 cells (50% efficacy less than 1 $\mu$ M).

In spite of SHP role in the regulation of androgen synthesis directly in the testis<sup>107</sup> or indirectly *via* gonadotropin hormones,<sup>108</sup> such role has never been described so far in prostate, but cannot be excluded. Besides, SHP seems to play an antitumor role in many types of cancers<sup>109</sup>; unfortunately, SHP mRNA and/or protein accumulation has never been studied in prostate tumors. The centerpiece pointing out SHP as interesting in PCa comes from the fact that AR negatively regulates the amount of SHP (Figure 2).<sup>110</sup> Indeed, miR-141 which targets SHP is induced by AR in LNCaP cells. One could thus hypothesize that increasing concentration of active androgens would block SHP, which antagonizes AR activity and neutralizes the proliferative role of androgens.

## Conclusion and Perspectives

PCa incidence is drastically increasing in Westernized countries. Today, the main challenge is to have good diagnostic and prognostic markers that could help in the management of the patients. As androgens have been playing a central role in the progression of the tumors, most of these markers were previously obtained focusing on the screening of AR target genes. The involvement of other transcription factors, members or not of the nuclear receptor superfamily, has made possible to identify new signaling pathways regulating progression of the tumor until metastasis. LXR, FXR, and SHP have been associated for many years to the regulation of

metabolism. It is finally not surprising to find them as putative pharmacological targets to treat PCa, especially knowing that they can regulate androgen levels and AR activity (Figure 1). Altogether, developing new specific molecules regulating these nuclear receptors will give the opportunity to offer different therapeutic arsenals. This is probably the most challenging issue, especially in CRPC, which is the fate of almost all PCa.

### Author Contributions

M.C., S.D., S.M., S.B., A.M., and J.-M.A.L. collected the information, wrote the manuscript, and drew the figures; all authors read and approved the manuscript.

### Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

### Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: Part of this work was financed by grants from Université Blaise Pascal, Région Auvergne Rhône Alpes, Fond Européen de Développement Régional (FEDER), AAP Plan Cancer Environnement 2016.

### References

- Ghabili K, Tosoian JJ, Schaeffer EM, et al. The history of prostate cancer from antiquity: review of paleopathological studies. *Urology*. 2016;97:8-12.
- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin*. 2011;61:69-90.
- McDonald ML, Parsons JK. The case for tailored prostate cancer screening: an NCCN perspective. *J Natl Compr Canc Netw*. 2015;13:1576-1583.
- Cobran EK, Hall JN, Aiken WD. African-American and Caribbean-born men's perceptions of prostate cancer fear and facilitators for screening behavior: a pilot study. *J Cancer Educ*. 2018;33(3):640-648.
- Baade PD, Youlden DR, Krnjacki LJ. International epidemiology of prostate cancer: geographical distribution and secular trends. *Mol Nutr Food Res*. 2009;53:171-184.
- Andriole GL, Crawford ED, Grubb RL, et al. Mortality results from a randomized prostate-cancer screening trial. *N Engl J Med*. 2009;360:1310-1319.
- Schröder FH, Hugosson J, Roobol MJ, et al. Screening and prostate-cancer mortality in a randomized European study. *N Engl J Med*. 2009;360:1320-1328.
- Grönberg H. Prostate cancer epidemiology. *Lancet*. 2003;361:859-864.
- Sakr WA, Haas GP, Cassin BF, Pontes JE, Crissman JD. The frequency of carcinoma and intraepithelial neoplasia of the prostate in young male patients. *J Urol*. 1993;150:379-385.
- Perdana NR, Mochtar CA, Umbas R, Hamid ARA. The risk factors of prostate cancer and its prevention: a literature review. *Acta Med Indones*. 2016;48:228-238.
- Rebbeck TR. Prostate cancer genetics: variation by race, ethnicity, and geography. *Semin Radiat Oncol*. 2017;27:3-10.
- DeSantis CE, Siegel RL, Sauer AG, et al. Cancer statistics for African Americans, 2016: progress and opportunities in reducing racial disparities. *CA Cancer J Clin*. 2016;66:290-308.
- Pietro GD, Chornokur G, Kumar NB, Davis C, Park JY. Racial differences in the diagnosis and treatment of prostate cancer. *Int Neurourol J*. 2016;20:S112-S119.
- Tran HN, Li Y, Udaltsova N, Armstrong MA, Friedman GD, Klatsky AL. Risk of cancer in Asian Americans: a Kaiser Permanente cohort study. *Cancer Causes Control*. 2016;27:1197-1207.
- Cook LS, Goldoft M, Schwartz SM, Weiss NS. Incidence of adenocarcinoma of the prostate in Asian immigrants to the United States and their descendants. *J Urol*. 1999;161:152-155.
- Watanabe M, Nakayama T, Shiraishi T, Stemmermann GN, Yatani R. Comparative studies of prostate cancer in Japan versus the United States. A review. *Urol Oncol*. 2000;5:274-283.
- Shimizu H, Ross RK, Bernstein L, Yatani R, Henderson BE, Mack TM. Cancers of the prostate and breast among Japanese and white immigrants in Los Angeles County. *Br J Cancer*. 1991;63:963-966.
- Taylor RA, Lo J, Ascuí N, Watt MJ. Linking obesogenic dysregulation to prostate cancer progression. *Endocr Connect*. 2015;4:R68-R80.
- Gacci M, Russo GI, De Nunzio C, et al. Meta-analysis of metabolic syndrome and prostate cancer. *Prostate Cancer Prostatic Dis*. 2017;20(2):146-155.
- Ben Sahara I, Laurent K, Giuliano S, et al. Targeting cancer cell metabolism: the combination of metformin and 2-deoxyglucose induces p53-dependent apoptosis in prostate cancer cells. *Cancer Res*. 2010;70:2465-2475.
- Loubière C, Goiran T, Laurent K, Djabari Z, Tanti J-F, Bost F. Metformin-induced energy deficiency leads to the inhibition of lipogenesis in prostate cancer cells. *Oncotarget*. 2015;6:15652-15661.
- Hankinson SJ, Fam M, Patel NN. A review for clinicians: prostate cancer and the antineoplastic properties of metformin. *Urol Oncol*. 2017;35:21-29.
- Swinnen JV, Heemers H, van de Sande T, et al. Androgens, lipogenesis and prostate cancer. *J Steroid Biochem Mol Biol*. 2004;92:273-279.
- Kuemmerle NB, Rysman E, Lombardo PS, et al. Lipoprotein lipase links dietary fat to solid tumor cell proliferation. *Mol Cancer Ther*. 2011;10:427-436.
- Brusselmans K, Timmermans L, Van de Sande T, et al. Squalene synthase, a determinant of Raft-associated cholesterol and modulator of cancer cell proliferation. *J Biol Chem*. 2007;282:18777-18785.
- Beckers A, Organe S, Timmermans L, et al. Chemical inhibition of acetyl-CoA carboxylase induces growth arrest and cytotoxicity selectively in cancer cells. *Cancer Res*. 2007;67:8180-8187.
- White C. The occurrence of crystals in tumours. *J Pathol Bacteriol*. 1909;13:3-10.
- Vargas C. Cholesterol in cutaneous cancer. *Urol Cutan Rev*. 1932;310.



29. Swyer G. The cholesterol content of normal and enlarged prostates. *Cancer Res.* 1942;2:372-375.
30. Chen Y, Hughes-Fulford M. Human prostate cancer cells lack feedback regulation of low-density lipoprotein receptor and its regulator, SREBP2. *Int J Cancer.* 2001;91:41-45.
31. Thysell E, Surowiec I, Hörnberg E, et al. Metabolomic characterization of human prostate cancer bone metastases reveals increased levels of cholesterol. *PLoS ONE.* 2010;5:e14175.
32. Yue S, Li J, Lee S-Y, et al. Cholesteryl ester accumulation induced by PTEN loss and PI3K/AKT activation underlies human prostate cancer aggressiveness. *Cell Metab.* 2014;19:393-406.
33. Statz CM, Patterson SE, Mockus SM. mTOR inhibitors in castration-resistant prostate cancer: a systematic review. *Target Oncol.* 2017;12:47-59.
34. Chang L, Graham PH, Ni J, et al. Targeting PI3K/Akt/mTOR signaling pathway in the treatment of prostate cancer radioresistance. *Crit Rev Oncol Hematol.* 2015;96:507-517.
35. Wettstein MS, Saba K, Umbehr MH, et al. Prognostic role of preoperative serum lipid levels in patients undergoing radical prostatectomy for clinically localized prostate cancer. *Prostate.* 2017;77:549-556.
36. Gordon JA, Midha A, Szeitz A, et al. Oral simvastatin administration delays castration-resistant progression and reduces intratumoral steroidogenesis of LNCaP prostate cancer xenografts. *Prostate Cancer Prostatic Dis.* 2016;19:21-27.
37. Hoque A, Chen H, Xu X-C. Statin induces apoptosis and cell growth arrest in prostate cancer cells. *Cancer Epidemiol Biomarkers Prev.* 2008;17:88-94.
38. Sekine Y, Furuya Y, Nishii M, Koike H, Matsui H, Suzuki K. Simvastatin inhibits the proliferation of human prostate cancer PC-3 cells via down-regulation of the insulin-like growth factor 1 receptor. *Biochem Biophys Res Commun.* 2008;372:356-361.
39. Sivaprasad U, Abbas T, Dutta A. Differential efficacy of 3-hydroxy-3-methylglutaryl CoA reductase inhibitors on the cell cycle of prostate cancer cells. *Mol Cancer Ther.* 2006;5:2310-2316.
40. Zheng X, Cui X-X, Avila GE, et al. Atorvastatin and celecoxib inhibit prostate PC-3 tumors in immunodeficient mice. *Clin Cancer Res.* 2007;13:5480-5487.
41. Zhuang L, Kim J, Adam RM, Solomon KR, Freeman MR. Cholesterol targeting alters lipid raft composition and cell survival in prostate cancer cells and xenografts. *J Clin Invest.* 2005;115:959-968.
42. Alfaqih MA, Allott EH, Hamilton RJ, Freeman MR, Freedland SJ. The current evidence on statin use and prostate cancer prevention: are we there yet? *Nat Rev Urol.* 2017;14:107-119.
43. Murai T. Cholesterol lowering: role in cancer prevention and treatment. *Biol Chem.* 2015;396:1-11.
44. Huggins C, Hodges CV. Studies on prostatic cancer. I. The effect of castration, of estrogen and of androgen injection on serum phosphatases in metastatic carcinoma of the prostate. *Cancer Res.* 1941;1:293-297.
45. Lacy JM, Kyprianou N. A tale of two trials: the impact of 5 $\alpha$ -reductase inhibition on prostate cancer (Review). *Oncol Lett.* 2014;8:1391-1396.
46. Gauthier-Landry L, Bélanger A, Barbier O. Multiple roles for UDP-glucuronosyltransferase (UGT)2B15 and UGT2B17 enzymes in androgen metabolism and prostate cancer evolution. *J Steroid Biochem Mol Biol.* 2015 Jan;145:187-92.
47. Bao B-Y, Chuang B-F, Wang Q, et al. Androgen receptor mediates the expression of UDP-glucuronosyltransferase 2 B15 and B17 genes. *Prostate.* 2008;68:839-848.
48. Cornford P, Bellmunt J, Bolla M, et al. EAU-ESTRO-SIOG guidelines on prostate cancer. Part II: treatment of relapsing, metastatic, and castration-resistant prostate cancer. *Eur Urol.* 2017;71:630-642.
49. Lowrance WT, Roth BJ, Kirkby E, Murad MH, Cookson MS. Castration-resistant prostate cancer: AUA guideline amendment 2015. *J Urol.* 2016;195:1444-1452.
50. Leach DA, Powell SM, Bevan CL. Women in cancer thematic review: new roles for nuclear receptors in prostate cancer. *Endocr Relat Cancer.* 2016;23:T85-T108.
51. Migliaccio A, Castoria G, Di Domenico M, et al. Steroid-induced androgen receptor-oestradiol receptor beta-Src complex triggers prostate cancer cell proliferation. *EMBO J.* 2000;19:5406-5417.
52. Baron S, Manin M, Beaudoin C, et al. Androgen receptor mediates non-genomic activation of phosphatidylinositol 3-OH kinase in androgen-sensitive epithelial cells. *J Biol Chem.* 2004;279:14579-14586.
53. Leung JK, Sadar MD. Non-genomic actions of the androgen receptor in prostate cancer. *Front Endocrinol.* 2017;8:2.
54. Heemers H, Maes B, Fougelle F, Heyns W, Verhoeven G, Swinnen JV. Androgens stimulate lipogenic gene expression in prostate cancer cells by activation of the sterol regulatory element-binding protein cleavage activating protein/sterol regulatory element-binding protein pathway. *Mol Endocrinol.* 2001;15:1817-1828.
55. Heemers H, Verrijdt G, Organe S, et al. Identification of an androgen response element in intron 8 of the sterol regulatory element-binding protein cleavage-activating protein gene allowing direct regulation by the androgen receptor. *J Biol Chem.* 2004;279:30880-30887.
56. Swinnen JV, Verhoeven G. Androgens and the control of lipid metabolism in human prostate cancer cells. *J Steroid Biochem Mol Biol.* 1998;65:191-198.
57. Brown MS, Goldstein JL. Sterol regulatory element binding proteins (SREBPs): controllers of lipid synthesis and cellular uptake. *Nutr Rev.* 1998;56:S1-S3; discussion S54-S75.
58. Fukuchi J, Hiipakka RA, Kokontis JM, et al. Androgenic suppression of ATP-binding cassette transporter A1 expression in LNCaP human prostate cancer cells. *Cancer Res.* 2004;64:7682-7685.
59. Alioui A, Celhay O, Baron S, Lobaccaro J-MA. Lipids and prostate cancer adenocarcinoma. *Clin Lipidol.* 2014;9:643-655.
60. Dillard PR, Lin M-F, Khan SA. Androgen-independent prostate cancer cells acquire the complete steroidogenic potential of synthesizing testosterone from cholesterol. *Mol Cell Endocrinol.* 2008;295:115-120.
61. Luu-The V, Bélanger A, Labrie F. Androgen biosynthetic pathways in the human prostate. *Best Pract Res Clin Endocrinol Metab.* 2008;22:207-221.



62. Mohler JL, Titus MA, Bai S, et al. Activation of the androgen receptor by intratumoral bioconversion of androstane-diol to dihydrotestosterone in prostate cancer. *Cancer Res.* 2011;71:1486-1496.
63. Kalaany NY, Mangelsdorf DJ. LXRs and FXR: the Yin and Yang of cholesterol and fat metabolism. *Annu Rev Physiol.* 2006;68:159-191.
64. Maqdasy S, Trousson A, Tauveron I, Volle DH, Baron S, Lobaccaro J-MA. Once and for all, LXR $\alpha$  and LXR $\beta$  are gatekeepers of the endocrine system. *Mol Aspects Med.* 2016;49:31-46.
65. Janowski BA, Willy PJ, Devi TR, Falck JR, Mangelsdorf DJ. An oxysterol signalling pathway mediated by the nuclear receptor LXR alpha. *Nature.* 1996;383:728-731.
66. Janowski BA, Grogan MJ, Jones SA, et al. Structural requirements of ligands for the oxysterol liver X receptors LXRalpha and LXRbeta. *Proc Natl Acad Sci U S A.* 1999;96:266-271.
67. Peet DJ, Turley SD, Ma W, et al. Cholesterol and bile acid metabolism are impaired in mice lacking the nuclear oxysterol receptor LXR alpha. *Cell.* 1998;93:693-704.
68. Hong C, Tontonoz P. Liver X receptors in lipid metabolism: opportunities for drug discovery. *Nat Rev Drug Discov.* 2014;13:433-444.
69. Chuu C-P, Kokontis JM, Hiipakka RA, Liao S. Modulation of liver X receptor signaling as novel therapy for prostate cancer. *J Biomed Sci.* 2007;14:543-553.
70. Chuu C, Hiipakka RA, Kokontis JM, Fukuchi J, Chen R-Y, Liao S. Inhibition of tumor growth and progression of LNCaP prostate cancer cells in athymic mice by androgen and liver X receptor agonist. *Cancer Res.* 2006;66:6482-6486.
71. Pommier AJC, Dufour J, Alves G, et al. Liver x receptors protect from development of prostatic intra-epithelial neoplasia in mice. *PLoS Genet.* 2013;9:e1003483.
72. Varambally S, Dhanasekaran SM, Zhou M, et al. The polycomb group protein EZH2 is involved in progression of prostate cancer. *Nature.* 2002;419:624-629.
73. Kunderfranco P, Mello-Grand M, Cangemi R, et al. ETS transcription factors control transcription of EZH2 and epigenetic silencing of the tumor suppressor gene Nkx3.1 in prostate cancer. *PLoS ONE.* 2010;5:e10547.
74. Beke L, Nuytten M, Van Eynde A, Beullens M, Bollen M. The gene encoding the prostatic tumor suppressor PSP94 is a target for repression by the Polycomb group protein EZH2. *Oncogene.* 2007;26:4590-4595.
75. Pommier AJC, Alves G, Viennois E, et al. Liver X Receptor activation downregulates AKT survival signaling in lipid rafts and induces apoptosis of prostate cancer cells. *Oncogene.* 2010;29:2712-2723.
76. Dufour J, Pommier A, Alves G, et al. Lack of liver x receptors leads to cell proliferation in a model of mouse dorsal prostate epithelial cell. *PLoS ONE.* 2013;8:e58876.
77. Viennois E, Pommier AJC, Mouzat K, et al. Targeting liver X receptors in human health: deadlock or promising trail? *Expert Opin Ther Targets.* 2011;15:219-232.
78. Viennois E, Mouzat K, Dufour J, Morel L, Lobaccaro J-M, Baron S. Selective liver X receptor modulators (SLiMs): what use in human health? *Mol Cell Endocrinol.* 2012;351:129-141.
79. Kim H-J, Andersson LC, Bouton D, Warner M, Gustafsson J-A. Stromal growth and epithelial cell proliferation in ventral prostates of liver X receptor knockout mice. *Proc Natl Acad Sci U S A.* 2009;106:558-563.
80. Füllhase C, Schneider MP. 5-alpha-reductase inhibitors and combination therapy. *Urol Clin North Am.* 2016;43:325-336.
81. Viennois E, Esposito T, Dufour J, et al. Lxr $\alpha$  regulates the androgen response in prostate epithelium. *Endocrinology.* 2012;153:3211-3223.
82. Tsui K-H, Chung L-C, Feng T-H, et al. Divergent effect of liver X receptor agonists on prostate-specific antigen expression is dependent on androgen receptor in prostate carcinoma cells. *Prostate.* 2015;75:603-615.
83. Clement JM, Sweeney CJ. Evolving treatment of oligometastatic hormone-sensitive prostate cancer. *J Oncol Pract.* 2017;13:9-18.
84. Fu W, Yao J, Huang Y, et al. LXR agonist regulates the carcinogenesis of PCa via the SOCS3 pathway. *Cell Physiol Biochem.* 2014;33:195-204.
85. Villablanca EJ, Raccosta L, Zhou D, et al. Tumor-mediated liver X receptor-alpha activation inhibits CC chemokine receptor-7 expression on dendritic cells and dampens antitumor responses. *Nat Med.* 2010;16:98-105.
86. Lanterna C, Musumeci A, Raccosta L, et al. The administration of drugs inhibiting cholesterol/oxysterol synthesis is safe and increases the efficacy of immunotherapeutic regimens in tumor-bearing mice. *Cancer Immunol Immunother.* 2016;65:1303-1315.
87. Gadaleta RM, Cariello M, Sabbà C, Moschetta A. Tissue-specific actions of FXR in metabolism and cancer. *Biochim Biophys Acta.* 2015;1851:30-39.
88. Wang S, Lai K, Moy FJ, Bhat A, Hartman HB, Evans MJ. The nuclear hormone receptor farnesoid X receptor (FXR) is activated by androsterone. *Endocrinology.* 2006;147:4025-4033.
89. Wang GM, Schaffner CP. Effect of candicidin and colestipol on the testes and prostate glands of BIO 87.20 hamsters. *Invest Urol.* 1976;14:66-71.
90. Saylor PJ, Karoly ED, Smith MR. Prospective study of changes in the metabolomic profiles of men during their first three months of androgen deprivation therapy for prostate cancer. *Clin Cancer Res.* 2012;18:3677-3685.
91. Saylor PJ, Smith MR, O'Malley AJ, Keating NL. Androgen-deprivation therapy and risk for biliary disease in men with prostate cancer. *Eur Urol.* 2014;65:642-649.
92. Evans AJ.  $\alpha$ -methylacyl CoA racemase (P504S): overview and potential uses in diagnostic pathology as applied to prostate needle biopsies. *J Clin Pathol.* 2003;56:892-897.
93. Zha S, Ferdinandusse S, Denis S, et al.  $\alpha$ -methylacyl-CoA racemase as an androgen-independent growth modifier in prostate cancer. *Cancer Res.* 2003;63:7365-7376.
94. Gafar AA, Draz HM, Goldberg AA, et al. Lithocholic acid induces endoplasmic reticulum stress, autophagy and mitochondrial dysfunction in human prostate cancer cells. *PeerJ.* 2016;4:e2445.
95. Goldberg AA, Titorenko VI, Beach A, Sanderson JT. Bile acids induce apoptosis selectively in androgen-dependent and -independent prostate cancer cells. *PeerJ.* 2013;1:e122.

96. Liu N, Zhao J, Wang J, Teng H, Fu Y, Yuan H. Farnesoid X receptor ligand CDCA suppresses human prostate cancer cells growth by inhibiting lipid metabolism via targeting sterol response element binding protein 1. *Am J Transl Res.* 2016;8:5118-5124.
97. Liu J, Tong S-J, Wang X, Qu L-X. Farnesoid X receptor inhibits LNCaP cell proliferation via the upregulation of PTEN. *Exp Ther Med.* 2014;8:1209-1212.
98. Choi YH, Im EO, Suh H, Jin Y, Yoo YH, Kim ND. Apoptosis and modulation of cell cycle control by synthetic derivatives of ursodeoxycholic acid and chenodeoxycholic acid in human prostate cancer cells. *Cancer Lett.* 2003;199:157-167.
99. Kaeding J, Bouchaert E, Bélanger J, et al. Activators of the farnesoid X receptor negatively regulate androgen glucuronidation in human prostate cancer LNCAP cells. *Biochem J.* 2008;410:245-253.
100. Yang M, Xie W, Mostaghel E, et al. SLCO2B1 and SLCO1B3 may determine time to progression for patients receiving androgen deprivation therapy for prostate cancer. *J Clin Oncol.* 2011;29:2565-2573.
101. Svoboda M, Riha J, Wlcek K, Jaeger W, Thalhammer T. Organic anion transporting polypeptides (OATPs): regulation of expression and function. *Curr Drug Metab.* 2011;12:139-153.
102. Jung D, Elferink MGL, Stellaard F, Groothuis GMM. Analysis of bile acid-induced regulation of FXR target genes in human liver slices. *Liver Int.* 2007;27:137-144.
103. Zhang Y, Hagedorn CH, Wang L. Role of nuclear receptor SHP in metabolism and cancer. *Biochim Biophys Acta.* 2011;1812:893-908.
104. Lu TT, Makishima M, Repa JJ, et al. Molecular basis for feedback regulation of bile acid synthesis by nuclear receptors. *Mol Cell.* 2000;6:507-515.
105. Makishima M, Okamoto AY, Repa JJ, et al. Identification of a nuclear receptor for bile acids. *Science.* 1999;284:1362-1365.
106. Dawson MI, Xia Z, Liu G, et al. An adamantyl-substituted retinoid-derived molecule that inhibits cancer cell growth and angiogenesis by inducing apoptosis and binds to small heterodimer partner nuclear receptor: effects of modifying its carboxylate group on apoptosis, proliferation, and protein-tyrosine phosphatase activity. *J Med Chem.* 2007;50:2622-2639.
107. Volle DH, Duggavathi R, Magnier BC, et al. The small heterodimer partner is a gonadal gatekeeper of sexual maturation in male mice. *Genes Dev.* 2007;21:303-315.
108. Vega A, Martinot E, Baptissart M, et al. Identification of the link between the hypothalamo-pituitary axis and the testicular orphan nuclear receptor NR0B2 in adult male mice. *Endocrinology.* 2015;156:660-669.
109. Zou A, Lehn S, Magee N, Zhang Y. New insights into orphan nuclear receptor SHP in liver cancer. *Nucl Receptor Res.* 2015;2:101162.
110. Xiao J, Gong A-Y, Eischeid AN, et al. miR-141 modulates androgen receptor transcriptional activity in human prostate cancer cells through targeting the small heterodimer partner protein. *Prostate.* 2012;72:1514-1522.