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Copy Number Variation Analysis of IL22 and LCE3C in Different Subtypes of Psoriasis in a Chinese Han Population

Authors' Stu Data Statistic Data Inte Ianuscript F Literat Funds	Contribution: udy Design A a Collection B cal Analysis C erpretation D Preparation E ture Search F 6 Collection G	ABCDEF 1-4 BDF 1,4 CD 1-3 BC 1-3 CF 1-4 ACDEFG 1-4	Caihong Zhu (D) Wenmin Fei Wenjun Wang Lili Tang Jinping Gao Fusheng Zhou	 Department of Dermatology, The First Affiliated Hospital of Anhui Medical University, Hefei, Anhui, PR China Institute of Dermatology, The First Affiliated Hospital of Anhui Medical Univers Hefei, Anhui, PR China The Key Laboratory of Dermatology (Anhui Medical University), Ministry of Education, Hefei, Anhui, PR China Collaborative Innovation Center for Complex and Severe Dermatosis, Anhui Medical University, Hefei, Anhui, PR China 			
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Background: Material/Methods:		ackground:	Psoriasis is a chronic, immune-mediated and hyperp mental components. Copy number variations (CNV) of be predisposed to psoriasis vulgaris (PsV) in several e er CNVs of <i>IL22</i> and <i>LCE3C</i> are associated with differe dermic psoriasis, EP; and generalized pustular psorias	roliferative skin disease with both genetic and environ- f <i>IL22</i> and <i>LCE3C-LCE3B</i> deletion have been confirmed to ethnic groups. However, it remains to be clarified wheth- ent subtypes of psoriasis (psoriatic arthritis, PsA; erythro- sis, GPP).			
		/Methods:	We enrolled 897 Han Chinese individuals, including 478 patients and 419 healthy controls, and detected CNVs of <i>IL22</i> and <i>LCE3C</i> using the comparative CT method by real-time PCR, and Pearson's χ^2 test was used to evaluated the copy number difference among subtypes.				
Results:			CNVs of <i>IL22</i> were significantly higher in PsV than in healthy controls ($P<0.001$). CNV of <i>LCE3C</i> in PsV, PsA, and GPP groups were significantly lower compared to healthy controls. When linked with clinical parameters, mild psoriasis carried less <i>IL22</i> copy numbers than that in severe psoriasis ($P=0.043$). Neither <i>IL22</i> or <i>LCE3C</i> CNVs were associated with age of onset.				
Conclusions:		onclusions:	CNVs of <i>LCE3C</i> and <i>IL22</i> might differentially contribute to subtypes of psoriasis. These findings suggest complex and diverse genetic variations in and among different clinical subtypes of psoriasis.				
	I	Keywords:	Body Surface Area • DNA Copy Number Variations	• Psoriasis			
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Background

Psoriasis is an autoimmune-mediated disease that primarily affects the skin, nails, and joints [1,2]. The disease presents mainly in 4 presentations, and classifications have been proposed based on clinical course. PsV, characterized by erythematous and scaly plaques, is usually found on the elbows, knees, scalp, and back and affects 85-90% of patients with psoriasis [3-6]. PsA is a type of psoriasis that is associated with erosive polyarthritis and usually has a negative serologic test result for rheumatoid factor; it shares some common genetic background with psoriasis [6,7]. EP presents as widespread erythema with thick scales (exfoliation of the skin, which is quite different from the thick, adherent, and white scales of PsV) [5,6,8]. GPP is characterized by infiltration of neutrophil granulocytes in the epidermis to such an extent that clinically visible sterile pustules develop [5,6,9,10]. Both EP and GPP are severe subtypes of psoriasis that affect nearly the entire body surface. Approximately 30% of psoriasis patients develop PsA, EP, or GPP. Even though these clinical phenotypes share some common manifestations, their molecular mechanisms differ and remain to be classified further.

It was widely accepted that aberrant secretion of cytokines and skin barrier dysfunction are the 2 key features in psoriasis pathogenesis. Some cytokines, including IL22, IL17A, IL23, and beta-defensin 2 [11], and skin structural proteins such as S1001A and LCE [12] were identified as critical risk factors in disease development. Thanks to the development of modern genetic technologies, great progress has been made in understanding the genetic landscape and pathogenesis of psoriasis. Linkage analysis and genome-wide association studies (GWASs), as well as next-generation sequencing studies, have further illuminated the genetic signatures of psoriasis. Notably, GWASs have identified more than 100 single-nucleotide polymorphisms (SNPs) and susceptibility genes. However, these markers explain only a small portion of the information in the hereditary susceptibility of psoriasis [13], indicating that other types of genetic loci beyond SNPs, such as CNVs, would add to the disease susceptible lists.

CNVs, defined as deletions or duplications of DNA segments longer than 1 kb, cover approximately 12% of the human genome and encompass more nucleotides than do SNPs [14]. CNVs can affect human phenotypes by directly affecting gene expression levels through duplication or deletion of a specific DNA sequence, or by indirectly altering gene expression, such as affecting the transcriptional regulatory elements of corresponding genes [15-18]. Several studies revealed that CNVs are important genetic factors that predispose to psoriasis [13,19-24]. CNVs of *IL22* have been identified to be associated with PsV in the Estonian and other populations [24,25]. *LCE3C-LCE3B* deletion has been widely reported to be associated with PsV in several ethnic groups as well [21-23,25-28]. CNVs of the betadefensin gene cluster and *FCGR3B* are reported to be associated with psoriasis in German and Chinese populations [19,20]. Unfortunately, all studies thus far have focused on psoriasis vulgaris. Whether CNVs of *LCE3C* and *IL22* are associated with different subtypes of psoriasis (PsA, GPP, and EP) remains to be determined. In this study, we focused on the relationship between CNVs of *IL22*, *LCE3C* and PsV, PsA, EP, and GPP in a Chinese Han population.

Material and Methods

Subjects

We enrolled 897 Chinese Han individuals, including 478 patients (74 PsA, 69 EP, 52 GPP, and 283 PsV) and 419 healthy controls. None of the patients in the study had received systemic or physical therapy within the 2 weeks before a blood sample was drawn. All subjects were recruited from the First Affiliated Hospital of Anhui Medical University, China. Psoriasis patients with joint symptoms were confirmed with a diagnosis of PsA according to the Classification for Psoriatic Arthritis criteria [29]. The remaining patients with no joint symptoms were divided into PsV, EP, and GPP groups according to their clinical manifestations which state in the introduction section. All patients were diagnosed according to their individual clinical manifestation and/or histopathological performance by 2 experienced dermatologists (Table 1). Psoriasis patients were grouped as early- and late-onset psoriasis. Early-onset psoriasis is defined as onset before 40 years of age, while the rest are defined as late-onset psoriasis. Psoriasis severity is categorized as mild, moderate, and severe, which is guided by body surface area (BSA). BSA is preferential measurement in the US and other countries in both routine clinical practice and clinical trials [30,31]. BSA was assessed by a trained physician. The severity of psoriasis was classified as follows: mild: BSA \leq 3%, moderate: 3% < BSA \leq 10%, and severe: BSA >10%. Healthy controls were confirmed to have no personal or family history of autoimmune diseases such as psoriasis, rheumatoid arthritis, and systemic lupus erythematosus. This study was approved by the Ethics Committee of the First Affiliated Hospital of Anhui Medical University. Each participant signed the informed consent form.

Sample Preparation

Genomic DNA was extracted from venous blood using the Qiagen Genomic Flexi Gene DNA Kit51206 (Qiagen, Germany). DNA templates used for CNV analysis were prepared as follows: DNA quality was assessed by gel electrophoresis to confirm no DNA degradation had occurred and was then diluted to 20 ng/ μ l and an OD260/280 ratio range of 1.8-2.0 for utilization.

	Severity	No.	Age		Gender		
Phenotypes ¹			Average (range) ²	P ³	Male	Female	P ⁴
Healthy controls		419	35.7 (4-84)	/	268 (63.9%)	151 (36.0%)	/
All patients		478	39.6 (6-90)	<0.001	280 (58.6%)	198 (41.4%)	0.11
PsV		283	36.9 (7-78)	0.16	161 (56.8%)	122 (43.1%)	0.07
	Mild	45	40.6 (13-76)	/	29 (64.44%)	16 (35.56%)	/
	Moderate	203	36.2 (7-78)	0.07(<i>P</i> ⁵)	114 (56.16%)	89 (43.84%)	<0.001(P ⁶)
	Severe	35	36.2 (13-58)	0.184(<i>P</i> ⁵)	18 (51.43%)	17 (48.57%)	0.001(<i>P</i> ⁶)
PsA		74	45.1 (14-90)	<0.001	46 (62.2%)	28 (37.8%)	0.86
GPP		52	31.4 (6-71)	0.09	24 (46.2%)	28 (53.8%)	0.02
EP		69	51.0 (11-85)	<0.001	49 (83.1%)	20 (33.8%)	0.32

Table 1. Demographic and clinical data of study participants.

¹ Clinical phenotypes of controls and psoriasis patients. PsV – psoriasis vulgaris; PsA – psoriatic arthritis; GPP – generalized pustular psoriasis; EP – erythrodermic psoriasis. ² Age (in years) values are given as the mean and range (minimum–maximum). ³ Mann-Whitney test (each subgroup of patients versus controls). ⁴ Pearson's chi-square test (each subgroup of patients versus controls). ⁵ Mann-Whitney test (each subgroup of patients versus Mild group). ⁶ Pearson's chi-square test (each subgroup of patients in the Mild group).

Quantification of Copy Numbers

Copy number was determined using the $2^{-\Delta CT}$ method using TaqMan copy number targets and RNase P reference assays (assay ID: 4403328). TaqMan Copy Number Assays Hs00146600_cn (for *IL22*) and Hs02550639_cn (for *LCE3C*) (Applied Biosystems, Foster City, CA, USA) were used to detect CNVs of *IL22* and *LCE3C* genes using real-time quantitative polymerase chain reaction (RT-qPCR) in an ABI PRISM 7900HT Fast Real-time PCR instrument. Cycling conditions were held at 95°C for 10 minutes, and 45 cycles of 2 steps (95°C for 15 s and 60°C for 1 min). Data were analyzed using the SDS2.2 software package (Applied Biosystems, Foster City, CA, USA).

Statistical Analysis

Statistical analysis was performed using the R 3.4.0 program for Windows (<u>https://www.rproject.org</u>). Categorical variables were compared using Pearson's χ^2 test. Logistic regression was used to obtain the odds ratio (OR) and 95% confidence interval (CI) for copy numbers. Continuous variables were compared using the nonparametric Mann-Whitney and Kruskal-Wallis tests. Linear regression analysis was used to investigate the relationship between BSA and different CNV group. Statistical significance was assumed for *P* values <0.05.

Results

Clinical Data of All Participants

Patients were classified as PsV (n=283), PsA (n=74), EP (n=69), and GPP (n=52). The mean age of psoriasis patients and healthy controls was 39.6 ± 16.1 and 35.7 ± 16.6 years, respectively. The characteristics of subjects in the study are shown in **Table 1**. There was no significant difference in the distribution of male and female subjects between case and control cohorts (*P*>0.01).

CNV of IL22 in Psoriasis Subtypes

Copy numbers of the *IL22* gene ranged from 2 to 7 (mean \pm SD, 2.31 \pm 0.6) in controls and 2 to 5 (mean \pm SD, 2.36 \pm 0.59) in patients. Mean copy numbers of *IL22* were significantly higher in the PsV group compared to the control group (*P*=0.001, *t* test) (**Figure 1A**). No strong association was observed in comparisons between PsA, GPP, or EP and controls (**Figure 1A**). Allele frequency of *IL22* copy numbers significantly differed in cases and controls, with 35% patients carried 3 or more copies, while only 26.3% controls obtained more than 3 copies. (*P*=0.021, Pearson's χ^2 test; OR=0.036, 95% CI=0.18-0.82) (**Figure 2A**). Furthermore, those who had more than 2 copies of *IL22* were more likely to develop PsV (*P*=0.028, **Figure 2B**), suggesting IL22 copy numbers increased risk of affecting psoriasis.



Figure 1. Frequency distributions of IL22 and LCE3C CNVs. (A) Associations of *IL22* gene copy number with PsV. (B) *LCE3C* CNVs are associated with PsV, PsA and GPP. ** P<0.001, * P<0.05. HC – healthy control; PsV – psoriasis vulgaris; PsA – psoriatic arthritis; EP – erythrodermic psoriasis; GPP – generalized pustular psoriasis. (The figure was created by Microsoft Excel 2019 software, Microsoft, USA).



Figure 2. Distribution of IL22 in PsV, PsA and HC. (A) *IL22* copy numbers were significantly different between PsV and HC.
(B) More than 2 copies of *IL22* increased the risk of developing PsV. (C) PsA has lower *IL22* copy number compared to PsV.
(D) The PsV group had wider *IL22* copy number spectrums than the PsA group. HC – healthy control; PsV – psoriasis vulgaris; PsA – psoriatic arthritis. (The figure was created by Microsoft Excel 2019 software, Microsoft, USA).

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Figure 3. Distribution of LCE3C in PsV, GPP, and HC. (A) LCE3C copy numbers were significantly different between PsV and HC.
(B) More than 2 copies of LCE3C decrease the risk of developing PsV. (C) LCE3C copy numbers were significantly different between GPP and HC. (D) More than 2 copies of LCE3C decreased the risk of developing GPP. HC – healthy control; PsV – psoriasis vulgaris; GPP – generalized pustular psoriasis. (The figure was created by Microsoft Excel 2019 software, Microsoft, USA).

We then looked at copy number alterations among psoriasis subtypes. Mean copy numbers of *IL22* were higher in PsV than that in PsA groups (*P*=0.0025), but not in other subtypes (**Figure 1A**). Interestingly, PsV group presented wider copy number spectrums than PsA group, with 2-6 copies in PsV and 2-3 copies in PsA (**Figure 2D**). A distinct distribution was observed between the PsV and PsA groups (*P*=7.75E-10, OR=0.089, 95% CI=0.26-2.83, **Figure 2C**). These findings revealed *IL22* copy numbers might differentially associated with psoriasis subtypes.

CNV of LCE3C in Psoriasis Subtypes

The mean *LCE3C* copy number in the PsV group was significantly lower when compared to the control group (P<0.001) (**Figure 1B**). The copy number distribution difference was observed between PsV and control groups (P<0.001, OR=1.4, 95% CI=0.91-2.18, **Figure 3A**). Furthermore, those who had 2 copies of *LCE3C* were more likely to develop PsV (P=0.0018,

Figure 3B). The mean *LCE3C* copy number in PsA and GPP groups was lower compared to the control group (*P*=0.0046 and *P*=1.56E-04, respectively) (**Figure 1B**). The same distribution trend was observed when comparing the GPP and control groups (*P*=0.078, OR=0.13, 95% CI=0.06-0.31) (**Figure 3C**). Those who had 2 copies of *LCE3C* were more probably to develop GPP (*P*<0.05, **Figure 3D**). The copy numbers of *LCE3C* tended to be associated with EP, although the comparison did not reach significance (*P*=0.076).

CNV of IL22 Linked with Psoriasis Severity

Given the critical roles of *IL22* in psoriasis development, we evaluated the relationship between CNV of *IL22* and clinical parameters of psoriasis. We found patients with different *IL22* copy numbers obtained varied severity scores (mild, moderate, and severe) (**Figure 4A**). We also found the mean BSA scores were dramatically higher in patients with 2 copy numbers than that in 3 or more carriers, and the CNV of *IL22* was inversely

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Figure 4. Associations of IL22 and LCE3C copy numbers among different severities of psoriasis and healthy controls. (A) The distribution of *IL22* CNV was different and inversely correlated with the severity of psoriasis. (B) The distribution of *LCE3C* CNV was different and inversely correlated with the severity of psoriasis. (C) *IL22* copy numbers in mild psoriasis are higher than in severe psoriasis and in healthy controls. (D) *LCE3C* copy numbers in mild, moderate, and severe psoriasis were lower than in healthy controls. Mild – mild psoriasis; Moderate – moderate psoriasis; Severe – severe psoriasis; HC – healthy control; Blue Bar – the correlation between different CNV group and the BSA score. * *P*<0.05. (The figure was created by Microsoft Excel 2019 software, Microsoft, USA).

correlated with the severity of psoriasis (*P*=0.0348, R²=0.9316, **Figure 4A**). CNV of *IL22* in the mild group were significantly higher than in either the severe group or healthy controls (Mild: 2.53±0.694, Severe: 2.33±0.581, Control: 2.31±0.598, *P*_{Mild vs Severe}=0.043, *P*_{Mild vs Control}=0.019, respectively, **Figure 4C**). We also evaluated the relationship between CNV in the *LCE3C* gene and the clinical parameters of psoriasis. Similar to *IL22* copy numbers, BSA scores varied in patients with different *LCE3C* copy numbers (**Figure 4B**). But the correlation between CNV of *LCE3C* and the trends of BSA score were not found (**Figure 4B**). However, we noticed that CNV of *LCE3C* in mild, moderate and severe psoriasis being lower compared to healthy controls (*P*_{mild}=0.007, *P*_{moderate}=0.002, *P*_{severe}<0.001, respectively, **Figure 4D**).

For genetic background would be associated with early onset of psoriasis [32]. We assessed whether CNVs of *IL22* and *LEC3C* were risk factors for early onset, by comparing the copy number allele frequencies in early and late-onset 4 subtypes of psoriasis. No significant relationships were observed, indicating CNVs of *IL22* and *LEC3C* are not the key predisposing factors for early onset of psoriasis.

Discussion

Psoriasis is a common skin disorder with strong environmental and genetic components. Genome variation studies have revealed at least 100 disease-associated risk loci or genes, including CNVs in *IL22* and *LCE3C* [21-28]. Our group previously identified *LCE3C* copy numbers were strong risk factors in PsV predisposition. Several gene copy numbers have been associated with PsV in European, African, and other ethnic populations. For the phenotype difference among PsV, PsA, EP and GPP, we raised the question of whether copy number alteration of these genes might be involved in etiology of disease development. We selected *IL22* and *LCE3C* as candidates, mainly for 2 reasons. Firstly, CNVs of these 2 genes have been confirmed to be risk alleles in both Chinese and other populations, but have not been evaluated in psoriasis subtypes. Secondly, psoriasis was tightly associated with immune

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dysfunction and epidermal hyperproliferation. *IL22* and *LCE3C* are good candidates because of their fundamental roles in modulating immune microenvironment and barrier function of human skins respectively. This study will help better understanding the common and different aspect of psoriasis subtypes, from the perspective of *IL22* and *LCE3C* copy number alterations.

IL22 is a cytokine member of the *IL10* family. *IL22* targets external epithelial barriers and maintains immune homeostasis at barrier surfaces through the induction of regulatory genes [33]. As an important component of the cytokine network, *IL22* has long been implicated in driving psoriatic skin inflammation [24,34-36]. The concentration of *IL22* in serum was significantly higher in psoriasis patients than in healthy controls [37]. In psoriatic skin lesions, increased *IL22* pathway signaling (*IL20* subfamily members including *IL10*, *IL19*, *IL20*, *IL22*, *IL24*, and *IL26*) induces characteristic histological features, such as skin infiltration [36]. Furthermore, serum levels of *IL22* were positively correlated with the severity scores of GPP [38].

In this study, CNVs of IL22 were found to be significantly associated with PsV but not with other subtypes. A protective effect of lower IL22 copy number was observed, indicating that a higher copy number leads to increased risk of developing psoriasis vulgaris, while a lower copy number results in reduced disease risk [24], which was consistent with the Estonian study. A previous study has demonstrated the mRNA expression of IL22 in the samples with >2 copies was significantly higher than that in those with 2 copies [39]. Thus, we speculate that CNV of IL22 may directly affect the expression of IL22 and thus participate in the pathogenesis and clinical manifestations of psoriasis. Interestingly, IL22 copy number was inversely correlated with disease severity. This phenomenon might be due to our relatively small sample size in high copy number groups or the different distribution of sex ratio in patients with varying severity of psoriasis, but we raised the possibility that IL22 might be just a trigger factor that ignited the inflammatory circuit. We speculated that expression level of IL22 can be modulated by several mechanisms such as promoter methylation, miRNA, and/ or intergraded with some undetermined factors. IL22 CNVs is a genome variation that stably induces gene expression and lasts for a relatively long time. The disease severity was also determined by some other cytokines like IL17, IL12, and IL23. This hypothesis was confirmed by the fact that biological therapy targeted to IL22 was less effective than to IL17 or IL23 [40,41].

We also identified differential distribution of *IL22* CNV between PsA and PsV subtypes. *IL22* is expressed at a high level in the synovial fluid of PsA patients and can regulate synoviocyte proliferation [35]. The inverse correlation between *IL22* CNV and PsA in our study suggested *IL22* alone was insufficient to drive synovial inflammation, but needs additional cytokines, such as *IL17A*. There also some other possibilities such as inadequate sample size or bias in sample selection in the PsA group. These findings suggest that CNV of *IL22* has diverse roles in different subtypes and severities of psoriasis. It is reasonable to speculate that different subtypes and severities of psoriasis stem, at least in part, from differences in their *IL22* CNV.

The LCE3C gene is located in the LCE gene cluster, encoding stratum corneum proteins, within the epidermal differentiation complex on chromosome 1g21.3. Stratum corneum proteins are indispensable in the differentiation and disruption of the terminal epidermis [22]. LCE3C mRNA expression is detected in psoriasis lesions but not in normal skin [22]. Mutations in LCE3B/LCE3C confer a nearly 40% increased risk of development of plaque psoriasis [42]. Reduced CNVs in the LCE gene cluster were associated with psoriasis in Chinese, European, and American populations. Consistent with previous studies, we found that lower copy numbers of LCE3C were associated with PsV [23,43]. Our study also demonstrated associations between LCE3C CNV and PsA/GPP. In each group, except for EP, subjects with fewer LCE3C copies were more prone to developing psoriasis. This suggests that PsV, GPP, and PsA share a partially similar disease pathogenesis. We also found the CNV of LCE3C in mild, moderate, and severe psoriasis were lower than in healthy controls, suggesting LCE3C CNV may play a consistent role in patients with different severities of disease.

Conclusions

In conclusion, we observed that CNV of *IL22* contributes to the occurrence of PsV and might play different roles during the development of PsV and PsA. Reduced CNV of *IL22* increases the risk of developing severe psoriasis. CNV in *LCE3C* contributes to the risk of PsV, PsA, and GPP. Both CNV of *IL22* and *LCE3C* affect the severity of psoriasis. Our study reveals the complexity in associations of different CNVs with different subtypes of psoriasis. We demonstrate that CNVs have a profound effect on human genomic diversity and may cause disease or confer risk to complex disease traits. This information will aid in identifying new and suitable targets for diagnosis and therapy in different subtypes and different severities of psoriasis according to different CNV of *IL22* and *LCE3C* may help to define populations with high susceptibility to different subtypes of psoriasis.

Further research is needed to determine whether CNVs of *IL22* and *LCE3C* are predictors of psoriasis and the severity or clinical classification of psoriasis. *IL22* and *LCE3C* are located far apart, and the relations between the 2 genes are poorly understood. Therefore, the correlation between the 2 genes needs further analysis before combined analysis, which will be the focus of our future research.

Declaration of Figures' Authenticity

All figures submitted have been created by the authors who confirm that the images are original with no duplication and have not been previously published in whole or in part.

References:

- 1. Harden JL, Krueger JG, Bowcock AM. The immunogenetics of psoriasis: A comprehensive review. J Autoimmun. 2015;64:66-73
- Nikamo P, Cheuk S, Lysell J, et al. Genetic variants of the IL22 promoter associate to onset of psoriasis before puberty and increased IL-22 production in T cells. J Invest Dermatol. 2014;134:1535-41
- 3. Boehncke WH, Schon MP. Psoriasis. Lancet. 2015;386:983-94
- Palfreeman AC, McNamee KE, McCann FE. New developments in the management of psoriasis and psoriatic arthritis: A focus on apremilast. Drug Des Devel Ther. 2013;7:201-10
- 5. Griffiths CE, Barker JN. Pathogenesis and clinical features of psoriasis. Lancet. 2007;370:263-71
- Griffiths C, Barker J, Bleiker TO, et al. Rook's textbook of dermatology, 4 Volume Set: Wiley; 2016
- 7. Moll JM, Wright V. Psoriatic arthritis. Semin Arthritis Rheum. 1973;3:55-78
- Rosenbach M, Hsu S, Korman NJ, et al. Treatment of erythrodermic psoriasis: From the medical board of the National Psoriasis Foundation. J Am Acad Dermatol. 2010;62:655-62
- 9. Fujita H, Terui T, Hayama K, et al. Japanese guidelines for the management and treatment of generalized pustular psoriasis: The new pathogenesis and treatment of GPP. J Dermatol. 2018;45:1235-70
- Navarini AA, Burden AD, Capon F, et al. European consensus statement on phenotypes of pustular psoriasis. J Eur Acad Dermatol Venereol. 2017;31:1792-99
- 11. Hwang ST, Nijsten T, Elder JT. Recent highlights in psoriasis research. J Invest Dermatol. 2017;137:550-56
- Zhang XJ, Huang W, Yang S, et al. Psoriasis genome-wide association study identifies susceptibility variants within LCE gene cluster at 1q21. Nat Genet. 2009;41:205-10
- Yin X, Wineinger NE, Cheng H, et al. Common variants explain a large fraction of the variability in the liability to psoriasis in a Han Chinese population. BMC Genomics. 2014;15:87
- 14. Redon R, Ishikawa S, Fitch KR, et al. Global variation in copy number in the human genome. Nature. 2006;444:444-54
- 15. Kleinjan DA, van Heyningen V. Long-range control of gene expression: Emerging mechanisms and disruption in disease. Am J Hum Genet. 2005;76:8-32
- McCarroll SA, Hadnott TN, Perry GH, et al. Common deletion polymorphisms in the human genome. Nat Genet. 2006;38:86-92
- Rodriguez-Revenga L, Mila M, Rosenberg C, et al. Structural variation in the human genome: The impact of copy number variants on clinical diagnosis. Genet Med. 2007;9:600-6
- Stranger BE, Forrest MS, Dunning M, et al. Relative impact of nucleotide and copy number variation on gene expression phenotypes. Science. 2007;315:848-53
- Wu Y, Zhang Z, Tao L, et al. A high copy number of FCGR3B is associated with psoriasis vulgaris in Han Chinese. Dermatology. 2014;229:70-75
- Hollox EJ, Huffmeier U, Zeeuwen PL, et al. Psoriasis is associated with increased beta-defensin genomic copy number. Nat Genet. 2008;40:23-25
- Bassaganyas L, Riveira-Munoz E, Garcia-Aragones M, et al. Worldwide population distribution of the common LCE3C-LCE3B deletion associated with psoriasis and other autoimmune disorders. BMC Genomics. 2013;14:261
- 22. de Cid R, Riveira-Munoz E, Zeeuwen PL, et al. Deletion of the late cornified envelope LCE3B and LCE3C genes as a susceptibility factor for psoriasis. Nat Genet. 2009;41:211-15

- 23. Li M, Wu Y, Chen G, et al. Deletion of the late cornified envelope genes LCE3C and LCE3B is associated with psoriasis in a Chinese population. J Invest Dermatol. 2011;131:1639-43
- 24. Prans E, Kingo K, Traks T, et al. Copy number variations in IL22 gene are associated with Psoriasis vulgaris. Hum Immunol. 2013;74:792-95
- Zhou F, Shen C, Hsu YH, et al. DNA methylation-based subclassification of psoriasis in the Chinese Han population. Front Med. 2018;12:717-25
- Bashir S, Hassan I, Majid S, et al. Feasibility of establishing deletion of the late cornified envelope genes LCE3B and LCE3C as a susceptibility factor for psoriasis. Adv Biomed Res. 2016;5:109
- Coin LJ, Cao D, Ren J, et al. An exome sequencing pipeline for identifying and genotyping common CNVs associated with disease with application to psoriasis. Bioinformatics 2012;28:i370-i74
- Huffmeier U, Bergboer JG, Becker T, et al. Replication of LCE3C-LCE3B CNV as a risk factor for psoriasis and analysis of interaction with other genetic risk factors. J Invest Dermatol. 2010;130:979-84
- Taylor W, Gladman D, Helliwell P, et al. Classification criteria for psoriatic arthritis: Development of new criteria from a large international study. Arthritis Rheum. 2006;54:2665-73
- Menter A, Strober BE, Kaplan DH, et al. Joint AAD-NPF guidelines of care for the management and treatment of psoriasis with biologics. J Am Acad Dermatol. 2019;80:1029-72
- 31. Knuckles MLF, Levi E, Soung J. Defining and treating moderate plaque psoriasis: A dermatologist survey. J Dermatolog Treat. 2018;29:658-63
- Sun LD, Cheng H, Wang ZX, et al. Association analyses identify six new psoriasis susceptibility loci in the Chinese population. Nat Genet. 2010;42:1005-9
- Sonnenberg GF, Fouser LA, Artis D. Border patrol: Regulation of immunity, inflammation and tissue homeostasis at barrier surfaces by IL-22. Nat Immunol. 2011;12:383-90
- Ma HL, Liang S, Li J, et al. IL-22 is required for Th17 cell-mediated pathology in a mouse model of psoriasis-like skin inflammation. J Clin Invest. 2008;118:597-607
- 35. Mitra A, Raychaudhuri SK, Raychaudhuri SP. Functional role of IL-22 in psoriatic arthritis. Arthritis Res Ther. 2012;14:R65
- Zheng Y, Danilenko DM, Valdez P, et al. Interleukin-22, a T(H)17 cytokine, mediates IL-23-induced dermal inflammation and acanthosis. Nature. 2007;445:648-51
- Wang B, Han D, Li F, et al. Elevated IL-22 in psoriasis plays an antiapoptotic role in keratinocytes through mediating Bcl-xL/Bax. Apoptosis. 2020;25:663-73
- Yamamoto M, Imai Y, Sakaguchi Y, et al. Serum cytokines correlated with the disease severity of generalized pustular psoriasis. Dis Markers. 2013;34:153-61
- Yu B, Guan M, Peng YH, et al. Copy number variations of interleukin-17F, interleukin-21, and interleukin-22 are associated with systemic lupus erythematosus. Arthritis Rheum. 2011;63:3487-92
- Tokuyama M, Mabuchi T. New treatment addressing the pathogenesis of psoriasis. Int J Mol Sci. 2020;21:7488
- Sbidian E, Chaimani A, Afach S, et al. Systemic pharmacological treatments for chronic plaque psoriasis: a network meta-analysis. Cochrane Database Syst Rev. 2020;1:CD011535
- Bao L, Li J, Perez White BE, et al. Inhibition of dipeptidyl-peptidase 4 induces upregulation of the late cornified envelope cluster in keratinocytes. Arch Dermatol Res. 2021 [Online ahead of print]
- 43. Riveira-Munoz E, He SM, Escaramis G, et al. Meta-analysis confirms the LCE3C_LCE3B deletion as a risk factor for psoriasis in several ethnic groups and finds interaction with HLA-Cw6. J Invest Dermatol. 2011;131:1105-9