



L-DOPA and Its Receptor GPR143: Implications for Pathogenesis and Therapy in Parkinson's Disease

Yoshio Goshima*, Daiki Masukawa, Yuka Kasahara, Tatsuo Hashimoto and Aderemi Caleb Aladeokin

Department of Molecular Pharmacology & Neurobiology, Yokohama City University Graduate School of Medicine, Yokohama, Japan

L-3,4-Dihydroxyphenylalanine (L-DOPA) is the most effective therapeutic agent for Parkinson's disease (PD). L-DOPA is traditionally believed to be an inert amino acid that exerts actions and effectiveness in PD through its conversion to dopamine. In contrast to this generally accepted idea, L-DOPA is proposed to be a neurotransmitter. Recently, GPR143 (OA1), the gene product of *ocular albinism 1* was identified as a receptor candidate for L-DOPA. GPR143 is widely expressed in the central and peripheral nervous system. GPR143 immunoreactivity was colocalized with phosphorylated α -synuclein in Lewy bodies in PD brains. GPR143 may contribute to the therapeutic effectiveness of L-DOPA and might be related to pathogenesis of PD.

OPEN ACCESS

Edited by:

Philippe De Deurwaerdere, Université de Bordeaux, France

Reviewed by:

Erwan Bezard, UMR5293 Institut des Maladies Neurodégénératives (IMN), France Clotilde Mannoury La Cour, Institut de Recherche International Servier, France

*Correspondence:

Yoshio Goshima goshima@med.yokohama-cu.ac.jp

Specialty section:

This article was submitted to Neuropharmacology, a section of the journal Frontiers in Pharmacology

Received: 13 April 2019 Accepted: 30 August 2019 Published: 03 October 2019

Citation:

Goshima Y, Masukawa D, Kasahara Y, Hashimoto T and Aladeokin AC (2019) L-DOPA and Its Receptor GPR143: Implications for Pathogenesis and Therapy in Parkinson's Disease. Front. Pharmacol. 10:1119. doi: 10.3389/fphar.2019.01119 Keywords: L-DOPA, neurotransmitter, G protein-coupled receptor, Parkinson's disease, dopamine, Lewy bodies

INTRODUCTION

Parkinson's disease (PD) results primarily from the degeneration of dopaminergic neurons in the substantia nigra. L-3,4-Dihydroxyphenylalanine (L-DOPA), a precursor of dopamine (DA), replenishes disease-related lower levels of DA by conversion of L-DOPA to DA in the brain by aromatic L-amino acid decarboxylase (AADC) and alleviates motor symptoms of PD. L-DOPA is mainly synthesized from tyrosine by tyrosine hydroxylase (TH), the rate-limiting enzyme for catecholamine synthesis. Synthetic DA agonists have been included in the treatment of PD for several decades, but L-DOPA is still the most effective therapeutic agent and widely considered the gold standard in PD treatment. However, L-DOPA therapy is associated with adverse effects, such as dyskinesias and impulse control disorders, the underlying mechanisms of which are largely unknown (Ahlskog, 2011; Gottwald and Aminoff, 2011; Rascol et al., 2011; Xie et al., 2015; Ivanova and Loonen, 2016; Voon et al., 2017).

In 1986, we first proposed that L-DOPA itself act a neurotransmitter. L-DOPA is released upon neuronal excitation, and L-DOPA exerts biological activity even with inhibition of AADC to prevent DA formation (Goshima et al., 2014). Most of these responses are antagonized by L-DOPA esters, compounds structurally related to L-DOPA. Recently, GPR143, a G protein–coupled receptor (GPCR), the gene product of *ocular albinism 1* (OA1) (Schiaffino et al., 1999), was reported to have binding activities for L-DOPA (Lopez et al., 2008). GPR143 was distributed in the central and peripheral nervous system and was further confirmed to play a role as an L-DOPA receptor *in vivo* (Hiroshima et al., 2014; Masukawa et al., 2014; Fukuda et al., 2015; Masukawa et al., 2017). GPR143 was also found to be localized in Lewy bodies, the histological hallmark of PD (Goshima et al., 2018). In this review, we revisit L-DOPA therapy and PD pathogenesis in light of L-DOPA action as a neurotransmitter.

1

EVIDENCE FOR L-DOPA AS A NEUROTRANSMITTER

L-DOPA has been believed to reside exclusively in the cytoplasm of catecholaminergic neurons only as a precursor of the neurotransmitter DA. Contrary to this generally accepted idea, we found that nerve stimulation elicited the release of L-DOPA in vitro and in vivo experiments (Goshima et al., 1988; Nakamura et al., 1992). The release was dependent on the extracellular Ca2+ and sensitive to tetrodotoxin, a voltage-dependent Na⁺-channel blocker. The L-DOPA release was suppressed by inhibitors of P-type voltage-sensitive Ca2+ channels and of synaptobrevin (Zhu et al., 2004). These findings together suggest that L-DOPA is released via exocytosis (Misu and Goshima, 1993). By using specific antibodies, TH- and L-DOPA-positive but AADCand DA-negative neurons were shown in some brain areas including the nucleus tractus solitarii (NTS), hypothalamic arcuate nucleus, and magnocellular neurosecretory system (Mons et al., 1988; Misu et al., 1996). Ultrastructurally, L-DOPA-positive signals were localized in the terminals with vesicle-like structures in the lateral habenular nucleus of the house-shrew brain (Karasawa et al., 1992). It is possible that L-DOPA release may arise from L-DOPA-containing vesicles, although vesicular L-DOPA transporter(s), if it exists, have not yet been identified. In addition, exogenous application of L-DOPA produced pharmacological actions in the presence of AADC inhibitor. L-DOPA facilitated the impulse-evoked noradrenaline (NA) release from superfused rat hypothalamic slices (Goshima et al., 1986; Goshima et al., 1991b). Structureactivity relationship study of DOPA-related compounds revealed that D-DOPA, the D-isomer of L-DOPA, did not mimic the effect of L-DOPA. In addition, L-DOPA methyl ester (L-DOPA ME) antagonized the action of L-DOPA in a competitive fashion. Thus, the L-DOPA recognition site(s) has high receptor-like specificity, being stereoselective and requiring specific structural features including the catechol moiety in addition to amino and carboxy groups. These findings support the existence of specific molecular recognition site(s) or receptor(s) for L-DOPA.

Previous reports suggested a role of L-DOPA as a neuromodulator in the regulation of motor function. Certain L-DOPA actions were observed only in PD model but not in normal control animals (Ueda et al., 1995a; Ueda et al., 1995b). Using in vivo microdialysis, L-DOPA decreased acetylcholine (ACh) from the striatum of rats lesioned with 6-hydroxydopamine (6-OHDA), but not from that of sham-operated rats. The L-DOPA-induced decrease was not affected by sulpiride, a D₂/D₃ antagonist. These findings suggest that L-DOPA by itself regulates the release of ACh. In addition, L-DOPAsensitive mechanisms were supersensitized in the PD model (Ueda et al., 1995b). A similar supersensitization to L-DOPA was also observed in quinpirole-induced locomotor activity. In normal rats, a highest dose of quinpirole, a D_2/D_3 agonist, slightly increased the total accounts of locomotor activities. Concomitant treatment with noneffective dose of L-DOPA (30 mg/kg) potentiated hyperlocomotion induced by quinpirole. In 6-OHDA rats, a noneffective lower dose of L-DOPA (10 mg/kg) potentiated quinpirole-induced locomotor activities (Nakamura et al., 1994).

L-DOPA probably plays a role as a neurotransmitter of the primary baroreceptor afferents in the lower brain stem. A prominent effect of L-DOPA is its depressor and bradycardic actions when microinjected into the NTS of anesthetized rats (Kubo et al., 1992; Yue et al., 1994; Misu et al., 1996; Goshima et al., 2014). Phenylephrine-induced hypertension triggered L-DOPA release in the NTS and reflex bradycardia temporally associated with the rise and recovery of blood pressure. The L-DOPA release and bradycardia were abolished by denervation of bilateral carotid sinus and aortic nerves, which contained the baroreceptor afferents. Electrical stimulation of aortic nerve released L-DOPA and induced depressor and bradycardic responses, and these responses were antagonized by bilateral injection of L-DOPA ME into the NTS (Yue et al., 1994). The depressor response induced by L-DOPA might possibly be related to orthostatic hypotension, which is often encountered in PD patients on L-DOPA therapy (Connolly and Lang, 2014; De Pablo-Fernandez et al., 2017).

RECEPTOR CANDIDATES TARGETTED BY L-DOPA: THEIR LOCALIZATION AND PHARMACOLOGY IN NORMAL CONDITIONS

These findings together suggest that there is one or more specific receptors for L-DOPA. L-DOPA did not interact with cognate monoamine receptors (Goshima et al., 1991b; Misu et al., 1996). On the other hand, L-DOPA inhibited specific binding of [³H]-AMPA in the brain membrane preparations, and L-DOPA induced current response in Xenopus laevis oocytes expressing AMPA receptors (Miyamae et al., 1999). However, L-DOPA interacted with AMPA receptors with a low affinity (ED₅₀ of 2.2 mM). In addition, DOPA ME and L-DOPA cyclohexyl ester (CHE), L-DOPA antagonists, did not interact with AMPA receptors (Miyamae et al., 1999). These findings suggest that the L-DOPA CHE-sensitive actions of L-DOPA are mediated through recognition sites other than monoamine receptors and AMPA receptors. A clue to the identification of L-DOPA receptors involved genetic analysis of *oa1*, the gene responsible for ocular albinism type 1 (Schiaffino et al., 1999). The gene product of oal referred to as GPR143 is critically involved in organogenesis of the melanosomes. Based on sequence similarities, the protein could not be assigned to any GPCR subfamily. Although binding activities of both DA and L-DOPA to GPR143 were detected, only L-DOPA induced intracellular Ca²⁺ response in cell lines expressing GPR143 (Lopez et al., 2008; Hiroshima et al., 2014). Dopamine also interacted with GPR143 with Ki of 2.4 µM, a comparable value to that for L-DOPA, but produced no functional response on intracellular Ca²⁺ levels in GPR143-expressing CHO cells. Dopamine may therefore act as an antagonist or inverse agonist against GPR143, which is consistent with the structural features and the homology model for GPR143 (Ghosh et al., 2012). These findings indicate that L-DOPA may be an endogenous ligand for GPR143. The role of GPR143 in mediating the L-DOPA response was then examined in the NTS of anesthetized rats. The shRNA knockdown of GPR143 in the NTS abolished or attenuated the depressor response to L-DOPA but not to glutamate microinjected into the NTS. L-DOPA CHE also suppressed the L-DOPA response and displaced the specific binding of [³H]-L-DOPA in CHO cells expressing GPR143. These findings suggest that GPR143 is involved in mediating the L-DOPA actions in the NTS (Hiroshima et al., 2014).

GPR143 is highly expressed in the retinal pigment epithelial cells (Schiaffino et al., 1999), and most studies have been conducted to delineate roles of GPR143 in melanogenesis (Schiaffino, 2010; De Filippo et al., 2017). Using specific antibodies against GPR143 and GPR143-KO mice, we investigated tissue distribution of GPR143. GPR143 immunoreactive cells were localized in the hippocampus, cerebral cortex, striatum, substantia nigra, hypothalamic median eminence and suprachiasmatic nucleus, NTS, and caudal ventrolateral medulla and olfactory bulb (Masukawa et al., 2014). A similar distribution pattern was confirmed in wild-type (Wt) mouse brains (Fukuda et al., 2015). Of note, GPR143 signals were observed in areas involving both hippocampal and cortical circuits as well as in basal ganglia and hypothalamic and brainstem neural networks. In line with GPR143 expression pattern, several observations reported an effect of L-DOPA in enhancing performances in executive verbal tasks and visuospatial working memory (Kulisevsky et al., 1996; Costa et al., 2003; Poletti and Bonuccelli, 2013), as well as in increasing secretion of growth hormone or cortisol (Biermasz et al., 2005; Muller et al., 2011; Marakaki et al., 2015), both of which being regulated by neurosecretory hypothalamic nuclei.

The downstream signaling pathways triggered by L-DOPA through GPR143 *in vivo* remain unknown (Schiaffino, 2010). Biochemical and morphological analysis suggest that Gai3, Go, and Gq are potential G protein partners for GPR143. Among them, Gai3 was shown to coprecipitate and colocalize with GPR143 (Young et al., 2008). In addition, the retinal pigment epithelial cells of Gai3-deficient mice contained fewer and larger melanosomes, and analysis of the optic pathways revealed a significant reduction of ipsilateral retinofugal projections, both of which were similar to the phenotypes of *Oa1/Gpr143*-deficient mice (Young et al., 2008). Although overall picture is far from understood (Schiaffino, 2010), it is likely that Gai3 plays a role at least in melanosome biogenesis and in the development of the optic tract.

There is additional evidence for the role of GPR143 as an L-DOPA receptor in the peripheral cardiovascular system (Masukawa et al., 2017). Phenotypic analysis of *Gpr143* gene-deficient (GPR143-KO) revealed that the pressor response to phenylephrine, an agonist of a_1 -adrenoceptors (α_1 ARs), was attenuated in GPR143-KO compared to that in *Wt* mice. The pressor response to vasopressin was not altered

in GPR143-KO mice. In isolated GPR143-KO peripheral arteries, phenylephrine-induced contraction was reduced, indicating that GPR143 expressed in vascular smooth muscle cells could regulate α_1 ARs -mediated arterial contraction. GPR143-KO smooth muscle cells, phenylephrine-In induced phosphorylation of myosin light chain 2 (MLC2) and ERK phosphorylation were attenuated compared to Wt smooth muscle cells. MLC2 and ERK phosphorylation are causally related to phenylephrine-induced contraction, an effect involving GPR143. Pretreatment with L-DOPA at low nanomolar concentrations augmented the vasoconstricting effect of phenylephrine in isolated arteries from Wt but not GPR143-KO mice. Consistently, exogenously applied L-DOPA at 1 to 10 nM augmented the contractile and intracellular Ca2+ responses to phenylephrine in smooth muscle cells. The binding affinity of phenylephrine against the specific binding of $[{}^{3}H]$ -prazosin, an $\alpha_{1}ARs$ antagonist, was higher in HEK293 cells coexpressing both a1_BAR-Myc and GPR143-EGFP than in cells coexpressing $\alpha_{1B}AR$ -Myc and free-EGFP. Immunoprecipitation assay also revealed that the interaction between GPR143 and $\alpha_{1B}AR$ was enhanced by L-DOPA. Importantly, the enhancement was observed at a concentration comparable to those of plasma L-DOPA (10-20 nM) (Chritton et al., 1997; Goldstein et al., 2003; Masukawa et al., 2017). GPR143, when functionally coupled with $\alpha 1_{B}AR$, may have significantly more affinity for L-DOPA than when not forming heteromers with $a1_BAR$ (Masukawa et al., 2017). Förster resonance energy transfer signal was detected in live HEK293 cells expressing a1_BAR-Venus and GPR143-CFP, but not in those expressing vasopressin receptor AVPR1_A -CFP and GPR143-Venus (Masukawa et al., 2017). The direct interaction between GPR143 and $\alpha_{1B}AR$ was demonstrated in Wt tissue by in situ proximity ligation assay. Furthermore, the stress-induced pressor response and the rise in blood pressure in active phase in GPR143-KO were blunted when compared to Wt mice. Together, these findings indicate that L-DOPA sensitizes contractile response to sympathetic outflow through coupling of GPR143 with aARs in vascular smooth muscle cells (Masukawa et al., 2017; Figure 1) and further suggest the physiological relevance of GPR143 as an L-DOPA receptor.

Recent findings indicate that the interaction between distinct GPCRs, forming hetero-oligomers, can affect affinity for agonists, the performance, and range through which extracellular signals are transmitted to G protein molecules (Ferre et al., 2014; Ferre, 2015; Farran, 2017). The functional interaction of hetero-oligomer GPCRs was observed for DA D1-D3 heterodimer receptors. D1-D3 receptor heterodimer had greater affinity for DA than did DA₁ monomeric receptors without DA₃ receptors (Fiorentini et al., 2008). Coupling between GPR143 and $\alpha 1_{B}AR$ could also occur in the central nervous system, because $\alpha 1_BAR$ was present in the nucleus accumbens and colocalized with dopamine D_1 receptor (Mitrano et al., 2018). These finding suggest that some of the L-DOPA actions may be mediated through possible functional coupling between these GPCRs: D1, al_BAR and GPR143. L-DOPA itself did not interact with adrenergic α-,β-,D₁, and D₂ receptors (Goshima et al., 1991b). However, L-DOPA actions that were suppressed by antagonists for these receptors have been reported. For example, prazosin attenuated L-DOPA-induced



hyperactivity without affecting its anti-PD actions or dyskinesia in 1-methyl-4-phenyl-1,2,3,6- tetrahydropyridine (MPTP)–lesioned macaques (Visanji et al., 2009). L-DOPA produced propranolol (nonselective β -adrenoceptor antagonist)–facilitatory action on the NA release from brain slices (Goshima et al., 1986). Propranolol attenuated L-DOPA–induced efflux of DA from the 6-OHDA–lesioned striatum and reduced L-DOPA–induced dyskinesia (Bhide et al., 2015).

Although GPR143 is the most plausible L-DOPA receptor candidate, we found that L-DOPA induced ptosis in GPR143-KO as well as *Wt* mice pretreated with a central AADC inhibitor (Ueda et al., 2016). Ptosis sometimes associates with short- or long-term L-DOPA treatment (Giladi et al., 1992). These findings raise the possibility that L-DOPA induces actions through both GPR143-dependent and GPR143-independent mechanisms as well as its conversion to DA.

PATHOPHYSIOLOGICAL SIGNIFICANCE OF L-DOPA/GPR143 INTERACTION IN PD

Parkinson's disease is characterized by neuronal loss of dopaminergic neurons, and enzymatic activities of TH and AADC decreased during the progression of the pathology (Lloyd et al., 1975; Nagatsu and Nagatsu, 2016). As mentioned, L-DOPA can induce orthostatic hypotension in the treatment of PD (Connolly and Lang, 2014; De Pablo-Fernandez et al., 2017). Thus, the reduction of TH levels and its subsequent decrease in the synthesis and release of endogenous L-DOPA may contribute to autonomic dysfunctions observed in PD (De Pablo-Fernandez et al., 2017). The possible reduction in endogenous levels of L-DOPA might be related to supersensitization of responses to L-DOPA or DA agonists in PD or experimental PD models (Ueda et al., 1995a; Shimamura et al., 2006; Bhide et al., 2015). On the other hand, long-term L-DOPA treatment could induce down-regulation or desensitization of the receptor(s) for L-DOPA. Thus, changes in GPR143 properties (expression, sensitization) may also arise in PD patients together with the progression of the pathology, which may account for the development of the side effects or loss of efficacy of L-DOPA.

The decrease in the AADC activity may also affect the pharmacological actions of L-DOPA. For example, in intact rat brain slices, L-DOPA increased the release of DA at 0.1 μ M but produced no effect at 1 μ M. In the presence of an AADC inhibitor, L-DOPA (1 µM) inhibited the release of DA (Goshima et al., 1986). Likewise, L-DOPA at 30 nM increased the DA release from intact slices, but decreased the release from the slices of MPTP-treated mice (Goshima et al., 1991a). These results imply that the action of L-DOPA can become predominant over the action of DA during the progression of PD. Another example is the ability of L-DOPA but not DA to induce the release of glutamate from the striatum (Goshima et al., 1993). The respective inhibitory and facilitatory effects of L-DOPA on the release of DA and glutamate seen under the decreased AADC activities (Goshima et al., 1986; Goshima et al., 1993) may be relevant to some adverse effects such as the decreased therapeutic efficacy, dyskinesia, or "on-off" encountered during long-term therapy (Marsden, 1994). The decreased AADC activities may further tip the balance in favor of the L-DOPA actions, because DA may act as an agonist or inverse agonist against GPR143 (Lopez et al., 2008).

Previous clinical data have suggested that L-DOPA either slows the progression of PD or has a prolonged effect on PD symptoms (Fahn et al., 2004). In contrast, a recent trial concluded that treatment with L-DOPA had no disease-modifying effect, either beneficial or detrimental, on early PD among patients who were evaluated over the course of 80 weeks (Verschuur et al., 2019). L-DOPA, DA, and related compounds have the potential to be cytotoxic, since free radical and quinone metabolites of L-DOPA were shown to be toxic *in vitro* and *in vivo* (Graham et al., 1978; Cohen, 1987; Smith et al., 1994). Our demonstration of increased glutamate release by L-DOPA (Goshima et al., 1993) provides *in vivo* evidence for a mechanism that could accelerate the degeneration process. On the other hand, it was shown that chronic L-DOPA promoted recovery in remaining DA neurons, increased mRNA, and induced release of BDNF in the brain of PD models (Murer et al., 1998; Zhang et al., 2006). Thus, specific assays suggest the possibility that L-DOPA might have both beneficial and detrimental effects depending on various conditions such as therapeutic doses of L-DOPA, its metabolism, and the degree of degeneration of DA neurons. Whether GPR143 is required for any long-term effects of L-DOPA is an important issue to be answered in future studies.

In addition to a possible role of GPR143 in symptomaticmodifying effect of L-DOPA, GPR143 might also be involved in the pathogenesis of PD. To gain an insight into this issue, immunohistochemical analysis was conducted of PD brain tissues by using human anti-GPR143 antibody (Goshima et al., 2018). GPR143-immunoreactive neurons with large perikarya and neurites were observed in the midbrain of control and PD brains. The GPR143-immunoreactive signals showed a dot-like pattern in the perikarya, reflecting its localizations to melanosomes or late endosomes/lysosomes (Schiaffino, 2010). Consistently, unlike other GPCRs, an unconventional dileucine motif and a tryptophan-glutamic acid doublet motif were identified in GPR143 within its third cytosolic loop and the C-terminal tail, respectively, both of which may be responsible for driving its intracellular targeting (Piccirillo et al., 2006). In the PD brain tissue, GPR143 was colocalized with phosphorylated a-synuclein in Lewy bodies (Goshima et al., 2018) (Figure 2). It might be possible that accumulation and localization of GPCRs such as GPR143 are related to the pathogenesis of PD or to efficacy and/ or untoward effects of L-DOPA. Interestingly, Parkin-associated endothelin receptor-like receptor (Pael-R) (GPR37) shared some properties common with those of GPR143. GPR37 showed overlapping expression pattern with GPR143 and poor trafficking to the plasma membrane. GPR37 was localized in Lewy bodies (Yang et al., 2003; Murakami et al., 2004) and, like GPR143, possessed high basal activities. Interestingly, overexpression of GPR37 resulted in death of dopaminergic neurons (Leinartaite and Svenningsson, 2017), while GPR37 was contributed to the signaling of certain neuroprotective factors (Meyer et al., 2013). Future studies should address whether GPR143 is also involved in the pathogenesis and the toward and/or untoward actions of L-DOPA in PD (Figure 3).



FIGURE 2 | GPR143-positive (A, C) and GPR143-negative (D) Lewy bodies in neurons in the substantia nigra compacta of PD brain tissue. GPR143-positive signals were blocked by the synthetic peptide (B). Scale bar, 20 µm. (Goshima et al., 2018).



a role in carrying baroreceptor information as a neurotransmitter of the primary baroreceptor afferents terminating in the NTS. ADN, aortic depressor nerve; CSN, carotid sinus nerve; CVLM, caudal ventrolateral medulla; IML, intermediolateral cell column; LC, locus caeruleus; NTS, nucleus tractus solitarii; PHN, posterior hypothalamic nucleus; RVLM, rostral ventrolateral medulla. (See details in Misu et al., 2002) **(B)** GPR143 might be related to PD pathogenesis. **(C)** A possible impact on L-DOPA therapy. The decrease in the AADC activity may affect L-DOPA/DA receptor signaling balance in PD brains.

CONCLUSION

L-DOPA is likely to play a role as a neurotransmitter as well as a precursor of DA. Some critical questions, however, remain unanswered. Does L-DOPA release occur by a vesicular or nonvesicular mechanism? How does GPR143 mediate L-DOPA actions at a single-cell level? Is GPR143 involved in the pathogenesis of PD? Is GPR143 the only functional receptor for L-DOPA? This is the beginning of a new era for the mechanistic study of L-DOPA as a neurotransmitter, its neuronal network specificity, and its pathogenic role, all of which surely have a major impact on L-DOPA therapy in PD.

ETHICS STATEMENT

All the experiments were performed in accordance with the guidelines of ethical committees of Yokohama City University, Tokyo Metropolitan Neurological Hospital and Tokyo Metropolitan Institute of Medical Sciences. Informed consent was obtained from all subjects.

AUTHOR CONTRIBUTIONS

YG, DM, YK, TH and AC all contributed to the writing and editing of this review.

FUNDING

YG is funded by a Grant-in-Aid for Scientific Research (B) (General) (no. 15H04687), Scientific Research (B) (General) (no. 18H02580), the Japanese SRF Grand for Biomedical Research (no. 1565), Uehara Memorial Foundation (no. 201320161 and 201720339), and the fund for Creation of Innovation Centers for

REFERENCES

- Ahlskog, J. E. (2011). Pathological behaviors provoked by dopamine agonist therapy of Parkinson's disease. *Physiol. Behav.* 104, 168–172. doi: 10.1016/j. physbeh.2011.04.055
- Bhide, N., Lindenbach, D., Barnum, C. J., George, J. A., Surrena, M. A., and Bishop, C. (2015). Effects of the beta-adrenergic receptor antagonist propranolol on dyskinesia and L-DOPA-induced striatal DA efflux in the hemi-parkinsonian rat. J. Neurochem. 134, 222–232. doi: 10.1111/jnc.13125
- Biermasz, N. R., Romijn, J. A., Pereira, A. M., and Roelfsema, F. (2005). Current pharmacotherapy for acromegaly: a review. *Expert Opin. Pharmacother* 6, 2393–2405. doi: 10.1517/14656566.6.14.2393
- Chritton, S. L., Chinnow, S. L., Grabau, C., Dousa, M. K., Lucas, D., Roddy, D., et al. (1997). Adrenomedullary secretion of DOPA, catecholamines, catechol metabolites, and neuropeptides. *J. Neurochem.* 69, 2413–2420. doi: 10.1046/j.1471-4159.1997.69062413.x
- Cohen, G. (1987). Monoamine oxidase, hydrogen peroxide, and Parkinson's disease. Adv. Neurol. 45, 119–125. doi: 10.1007/978-1-4613-0759-4_15
- Connolly, B. S., and Lang, A. E. (2014). Pharmacological treatment of Parkinson disease: a review. *JAMA* 311, 1670–1683. doi: 10.1001/jama.2014.3654
- Costa, A., Peppe, A., Dell'agnello, G., Carlesimo, G. A., Murri, L., Bonuccelli, U., et al. (2003). Dopaminergic modulation of visual-spatial working memory in Parkinson's disease. *Dement. Geriatr. Cogn. Disord.* 15, 55–66. doi: 10.1159/000067968
- De Filippo, E., Manga, P., and Schiedel, A. C. (2017). Identification of novel G protein–coupled receptor 143 ligands as pharmacologic tools for investigating X-linked ocular albinism. *Invest. Ophthalmol. Vis. Sci.* 58, 3118–3126. doi: 10.1167/iovs.16-21128
- De Pablo-Fernandez, E., Tur, C., Revesz, T., Lees, A. J., Holton, J. L., and Warner, T. T. (2017). Association of autonomic dysfunction with disease progression and survival in Parkinson disease. *JAMA Neurol.* 74, 970–976. doi: 10.1001/jamaneurol.2017.1125
- Fahn, S., Oakes, D., Shoulson, I., Kieburtz, K., Rudolph, A., Lang, A., et al. (2004). Levodopa and the progression of Parkinson's disease. N. Engl. J. Med. 351, 2498–2508. doi: 10.1056/NEJMoa033447
- Farran, B. (2017). An update on the physiological and therapeutic relevance of GPCR oligomers. *Pharmacol. Res.* 117, 303–327. doi: 10.1016/j. phrs.2017.01.008
- Ferre, S. (2015). The GPCR heterotetramer: challenging classical pharmacology. Trends Pharmacol. Sci. 36, 145–152. doi: 10.1016/j.tips.2015.01.002
- Ferre, S., Casado, V., Devi, L. A., Filizola, M., Jockers, R., Lohse, M. J., et al. (2014). G protein–coupled receptor oligomerization revisited: functional and pharmacological perspectives. *Pharmacol. Rev.* 66, 413–434. doi: 10.1124/ pr.113.008052
- Fiorentini, C., Busi, C., Gorruso, E., Gotti, C., Spano, P., and Missale, C. (2008). Reciprocal regulation of dopamine D1 and D3 receptor function and

Advanced Interdisciplinary Research Areas Program in the Project for Developing Innovation Systems from MEXT (no. 42890001). DM is funded by a Grant-in-Aid for Young Scientists (B) (no. 18K14923) and by the Uehara Memorial Foundation (201810115). TH is funded by a Grant-in-Aid for Scientific Research (C) (General) (no. 18K06896). YK is funded by a Grant-in-Aid for Young Scientists (no. 19K16375). AC is supported by Japanese government (MEXT) scholarship for PhD (no. 8513).

ACKNOWLEDGMENTS

We sincerely thank Drs M Koga, E Seki, N Arai, T Komori, Y Kamikubo, T Sakurai, K Tamura, H Wakui, U Yokoyama, and Y Ishikawa for collaboration. We also thank Dr SM Strittmatter for his critical reading of our manuscript.

trafficking by heterodimerization. Mol. Pharmacol. 74, 59-69. doi: 10.1124/ mol.107.043885

- Fukuda, N., Naito, S., Masukawa, D., Kaneda, M., Miyamoto, H., Abe, T., et al. (2015). Expression of ocular albinism 1 (OA1), 3, 4- dihydroxy- L-phenylalanine (DOPA) receptor, in both neuronal and non-neuronal organs. *Brain Res.* 1602, 62–74. doi: 10.1016/j.brainres.2015.01.020
- Ghosh, A., Sonavane, U., Andhirka, S. K., Aradhyam, G. K., and Joshi, R. (2012). Structural insights into human GPCR protein OA1: a computational perspective. J. Mol. Model 18, 2117–2133. doi: 10.1007/ s00894-011-1228-8
- Giladi, N., Mcmahon, D., Przedborski, S., Flaster, E., Guillory, S., Kostic, V., et al. (1992). Motor blocks in Parkinson's disease. *Neurology* 42, 333–339. doi: 10.1212/WNL42.2.333
- Goldstein, D. S., Eisenhofer, G., and Kopin, I. J. (2003). Sources and significance of plasma levels of catechols and their metabolites in humans. *J. Pharmacol. Exp. Ther.* 305, 800–811. doi: 10.1124/jpet.103.049270
- Goshima, Y., Kubo, T., and Misu, Y. (1986). Biphasic actions of L-DOPA on the release of endogenous noradrenaline and dopamine from rat hypothalamic slices. *Br. J. Pharmacol.* 89, 229–234. doi: 10.1111/j.1476-5381.1986. tb11139.x
- Goshima, Y., Kubo, T., and Misu, Y. (1988). Transmitter-like release of endogenous 3,4-dihydroxyphenylalanine from rat striatal slices. J. Neurochem. 50, 1725– 1730. doi: 10.1111/j.1471-4159.1988.tb02470.x
- Goshima, Y., Misu, Y., Arai, N., and Misugi, K. (1991a). Nanomolar L-DOPA facilitates release of dopamine via presynaptic beta-adrenoceptors: comparative studies on the actions in striatal slices from control and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated C57 black mice, an animal model for Parkinson's disease. Jpn J. Pharmacol. 55, 93–100. doi: 10.1254/ jjp.55.93
- Goshima, Y., Nakamura, F., Masukawa, D., Chen, S., and Koga, M. (2014). Cardiovascular actions of DOPA mediated by the gene product of ocular albinism 1. J. Pharmacol. Sci. 126, 14–20. doi: 10.1254/jphs.14R03CR
- Goshima, Y., Nakamura, S., and Misu, Y. (1991b). L-Dihydroxyphenylalanine methyl ester is a potent competitive antagonist of the L-dihydroxyphenylalanineinduced facilitation of the evoked release of endogenous norepinephrine from rat hypothalamic slices. J. Pharmacol. Exp. Ther. 258, 466–471.
- Goshima, Y., Ohno, K., Nakamura, S., Miyamae, T., Misu, Y., and Akaike, A. (1993). L-DOPA induces Ca(2+)-dependent and tetrodotoxin-sensitive release of endogenous glutamate from rat striatal slices. *Brain Res.* 617, 167–170. doi: 10.1016/0006-8993(93)90631-V
- Goshima, Y., Watanabe, S., Seki, E., Koga, M., Masukawa, D., Nakamura, F., et al. (2018). Immunoreactivity of a G protein–coupled l-DOPA receptor GPR143, in Lewy bodies. *Neurosci. Res.* 148, 49–53. doi: 10.1016/j.neures.2018.12.004
- Gottwald, M. D., and Aminoff, M. J. (2011). Therapies for dopaminergic-induced dyskinesias in Parkinson disease. Ann. Neurol. 69, 919–927. doi: 10.1002/ ana.22423

- Graham, D. G., Tiffany, S. M., Bell, W. R., Jr., and Gutknecht, W. F. (1978). Autoxidation versus covalent binding of quinones as the mechanism of toxicity of dopamine, 6-hydroxydopamine, and related compounds toward C1300 neuroblastoma cells *in vitro*. *Mol. Pharmacol.* 14, 644–653.
- Hiroshima, Y., Miyamoto, H., Nakamura, F., Masukawa, D., Yamamoto, T., Muraoka, H., et al. (2014). The protein Ocular albinism 1 is the orphan GPCR GPR143 and mediates depressor and bradycardic responses to DOPA in the nucleus tractus solitarii. *Br. J. Pharmacol.* 171, 403–414. doi: 10.1111/bph.12459
- Ivanova, S. A., and Loonen, A. J. (2016). Levodopa-induced dyskinesia is related to indirect pathway medium spiny neuron excitotoxicity: a hypothesis based on an unexpected finding. *Parkinsons Dis.* 2016, 6461907. doi: 10.1155/2016/6461907
- Karasawa, N., Isomura, G., and Nagatsu, I. (1992). Production of specific antibody against L-DOPA and its ultrastructural localization of immunoreactivity in the house-shrew (*Suncus murinus*) lateral habenular nucleus. *Neurosci. Lett.* 143, 267–270. doi: 10.1016/0304-3940(92)90280-K
- Kubo, T., Yue, J. L., Goshima, Y., Nakamura, S., and Misu, Y. (1992). Evidence for L-DOPA systems responsible for cardiovascular control in the nucleus tractus solitarii of the rat. *Neurosci. Lett.* 140, 153–156. doi: 10.1016/0304-3940(92)90090-T
- Kulisevsky, J., Avila, A., Barbanoj, M., Antonijoan, R., Berthier, M. L., and Gironell, A. (1996). Acute effects of levodopa on neuropsychological performance in stable and fluctuating Parkinson's disease patients at different levodopa plasma levels. *Brain* 119 (Pt 6), 2121–2132. doi: 10.1093/ brain/119.6.2121
- Leinartaite, L., and Svenningsson, P. (2017). Folding underlies bidirectional role of GPR37/Pael-R in Parkinson disease. *Trends Pharmacol. Sci.* 38, 749–760. doi: 10.1016/j.tips.2017.05.006
- Lloyd, K. G., Davidson, L., and Hornykiewicz, O. (1975). The neurochemistry of Parkinson's disease: effect of L-DOPA therapy. J. Pharmacol. Exp. Ther. 195, 453–464.
- Lopez, V. M., Decatur, C. L., Stamer, W. D., Lynch, R. M., and Mckay, B. S. (2008). L-DOPA is an endogenous ligand for OA1. *PLoS Biol.* 6, e236. doi: 10.1371/ journal.pbio.0060236
- Marakaki, C., Papadimitriou, D. T., Kleanthous, K., Papadopoulou, A., and Papadimitriou, A. (2015). L-DOPA stimulates cortisol secretion through adrenocorticotropic hormone release in short children. *Horm. Res. Paediatr.* 84, 319–322. doi: 10.1159/000439532
- Marsden, C. D. (1994). Problems with long-term levodopa therapy for Parkinson's disease. Clin. Neuropharmacol. 17 (Suppl), S32–S44.
- Masukawa, D., Koga, M., Sezaki, A., Nakao, Y., Kamikubo, Y., Hashimoto, T., et al. (2017). L-DOPA sensitizes vasomotor tone by modulating the vascular alpha1adrenergic receptor. *JCI Insight* 2, 17. doi: 10.1172/jci.insight.90903
- Masukawa, D., Nakamura, F., Koga, M., Kamiya, M., Chen, S., Yamashita, N., et al. (2014). Localization of ocular albinism-1 gene product GPR143 in the rat central nervous system. *Neurosci. Res.* 88, 49–57. doi: 10.1016/j. neures.2014.07.008
- Maurice, P., Kamal, M., and Jockers, R. (2011). Asymmetry of GPCR oligomers supports their functional relevance. *Trends Pharmacol. Sci.* 32, 514–520. doi: 10.1016/j.tips.2011.05.006
- Meyer, R. C., Giddens, M. M., Schaefer, S. A., and Hall, R. A. (2013). GPR37 and GPR37L1 are receptors for the neuroprotective and glioprotective factors prosaptide and prosaposin. *Proc. Natl. Acad. Sci. U S A* 110, 9529–9534. doi: 10.1073/pnas.1219004110
- Misu, Y., and Goshima, Y. (1993). Is L-DOPA an endogenous neurotransmitter? Trends Pharmacol. Sci. 14, 119–123. doi: 10.1016/0165-6147(93)90082-U
- Misu, Y., Goshima, Y., Ueda, H., and Okamura, H. (1996). Neurobiology of L-DOPAergic systems. *Prog. Neurobiol.* 49, 415–454. doi: 10.1016/0301-0082(96)00025-1
- Misu, Y., Goshima, Y., and Miyamae, T. (2002). Is DOPA a neurotransmitter? Trends Pharmacol. Sci. 23, 262–268. doi: 10.1016/S0165-6147(02)02013-8
- Mitrano, D. A., Jackson, K., Finley, S., and Seeley, A. (2018). alpha1b-Adrenergic receptor localization and relationship to the D1-dopamine receptor in the rat nucleus accumbens. *Neuroscience* 371, 126–137. doi: 10.1016/j. neuroscience.2017.11.053
- Miyamae, T., Goshima, Y., Shimizu, M., Shibata, T., Kawashima, K., Ohshima, E., et al. (1999). Some interactions of L-DOPA and its related

compounds with glutamate receptors. Life Sci. 64, 1045-1054. doi: 10.1016/S0024-3205(99)00031-4

- Mons, N., Danel, N., and Geffard, M. (1988). Visualization of L-dihydroxyphenylalanine in rat brain by using specific antibodies. *Brain Res.* 451, 403–407. doi: 10.1016/0006-8993(88)90793-7
- Muller, T., Hellwig, R., and Muhlack, S. (2011). Levodopa induces synthesis of nerve growth factor and growth hormone in patients with Parkinson disease. *Clin. Neuropharmacol.* 34, 101–103. doi: 10.1097/WNF.0b013e318219dad0
- Murakami, T., Shoji, M., Imai, Y., Inoue, H., Kawarabayashi, T., Matsubara, E., et al. (2004). Pael-R is accumulated in Lewy bodies of Parkinson's disease. Ann. Neurol. 55, 439–442. doi: 10.1002/ana.20064
- Murer, M. G., Dziewczapolski, G., Menalled, L. B., Garcia, M. C., Agid, Y., Gershanik, O., et al. (1998). Chronic levodopa is not toxic for remaining dopamine neurons, but instead promotes their recovery, in rats with moderate nigrostriatal lesions. *Ann. Neurol.* 43, 561–575. doi: 10.1002/ana.410430504
- Nagatsu, T., and Nagatsu, I. (2016). Tyrosine hydroxylase (TH), its cofactor tetrahydrobiopterin (BH4), other catecholamine-related enzymes, and their human genes in relation to the drug and gene therapies of Parkinson's disease (PD): historical overview and future prospects. J. Neural Transm. (Vienna) 123, 1255–1278. doi: 10.1007/s00702-016-1596-4
- Nakamura, S., Goshima, Y., Yue, J. L., and Misu, Y. (1992). Transmitter-like basal and K(+)-evoked release of 3,4-dihydroxyphenylalanine from the striatum in conscious rats studied by microdialysis. *J. Neurochem.* 58, 270–275. doi: 10.1111/j.1471-4159.1992.tb09306.x
- Nakamura, S., Yue, J. L., Goshima, Y., Miyamae, T., Ueda, H., and Misu, Y. (1994). Non-effective dose of exogenously applied L-DOPA itself stereoselectively potentiates postsynaptic D2 receptor-mediated locomotor activities of conscious rats. *Neurosci. Lett.* 170, 22–26. doi: 10.1016/0304-3940(94)90229-1
- Piccirillo, R., Palmisano, I., Innamorati, G., Bagnato, P., Altimare, D., and Schiaffino, M. V. (2006). An unconventional dileucine-based motif and a novel cytosolic motif are required for the lysosomal and melanosomal targeting of OA1. J. Cell Sci. 119, 2003–2014. doi: 10.1242/jcs.02930
- Poletti, M., and Bonuccelli, U. (2013). Acute and chronic cognitive effects of levodopa and dopamine agonists on patients with Parkinson's disease: a review. *Ther. Adv. Psychopharmacol.* 3, 101–113. doi: 10.1177/2045125312470130
- Rascol, O., Lozano, A., Stern, M., and Poewe, W. (2011). Milestones in Parkinson's disease therapeutics. *Mov. Disord.* 26, 1072–1082. doi: 10.1002/mds.23714
- Schiaffino, M. V. (2010). Signaling pathways in melanosome biogenesis and pathology. Int. J. Biochem. Cell Biol. 42, 1094–1104. doi: 10.1016/j. biocel.2010.03.023
- Schiaffino, M. V., D'addio, M., Alloni, A., Baschirotto, C., Valetti, C., Cortese, K., et al. (1999). Ocular albinism: evidence for a defect in an intracellular signal transduction system. *Nat. Genet.* 23, 108–112. doi: 10.1038/12715
- Shimamura, M., Shimizu, M., Yagami, T., Funabashi, T., Kimura, F., Kuroiwa, Y., et al. (2006). L-3,4-Dihydroxyphenylalanine–induced c-Fos expression in the CNS under inhibition of central aromatic L-amino acid decarboxylase. *Neuropharmacology* 50, 909–916. doi: 10.1016/j.neuropharm.2006.01.001
- Smith, T. S., Parker, W. D., Jr., and Bennett, J. P., Jr. (1994). L-DOPA increases nigral production of hydroxyl radicals *in vivo*: potential L-DOPA toxicity? *Neuroreport* 5, 1009–1011. doi: 10.1097/00001756-199404000-00039
- Ueda, H., Sato, K., Okumura, F., and Misu, Y. (1995b). L-DOPA inhibits spontaneous acetylcholine release from the striatum of experimental Parkinson's model rats. *Brain Res.* 698, 213–216. doi: 10.1016/0006-8993(95)00870-V
- Ueda, H., Sato, K., Okumura, F., Inoue, A., Nakata, Y., Ozaki, N., et al. (1995a). Supersensitization of neurochemical responses by L-DOPA and dopamine receptor agonists in the striatum of experimental Parkinson's disease model rats. Biomed. Pharmacother. 49, 169–177. doi: 10.1016/0753-3322(96)82617-5
- Ueda, S., Masukawa, D., Koga, M., and Goshima, Y. (2016). l-3,4-Dihydroxyphenylalanine induces ptosis through a GPR143-independent mechanism in mice. J. Pharmacol. Sci. 132, 109–112. doi: 10.1016/j.jphs.2016.08.005
- Verschuur, C. V. M., Suwijn, S. R., Boel, J. A., Post, B., Bloem, B. R., Van Hilten, J. J., et al. (2019). Randomized delayed-start trial of levodopa in Parkinson's disease. N. Engl. J. Med. 380, 315–324. doi: 10.1056/NEJMoa1809983
- Visanji, N. P., Fox, S. H., Johnston, T. H., Millan, M. J., and Brotchie, J. M. (2009). Alpha1-adrenoceptors mediate dihydroxyphenylalanine-induced activity in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine–lesioned macaques. J. Pharmacol. Exp. Ther. 328, 276–283. doi: 10.1124/jpet.108.144097

- Voon, V., Napier, T. C., Frank, M. J., Sgambato-Faure, V., Grace, A. A., Rodriguez-Oroz, M., et al. (2017). Impulse control disorders and levodopa-induced dyskinesias in Parkinson's disease: an update. *Lancet Neurol.* 16, 238–250. doi: 10.1016/S1474-4422(17)30004-2
- Xie, C. L., Zhang, Y. Y., Wang, X. D., Chen, J., Chen, Y. H., Pa, J. L., et al. (2015). Levodopa alone compared with levodopa-sparing therapy as initial treatment for Parkinson's disease: a meta-analysis. *Neurol. Sci.* 36, 1319–1329. doi: 10.1007/s10072-015-2253-7
- Yang, Y., Nishimura, I., Imai, Y., Takahashi, R., and Lu, B. (2003). Parkin suppresses dopaminergic neuron-selective neurotoxicity induced by Pael-R in *Drosophila*. *Neuron* 37, 911–924. doi: 10.1016/S0896-6273(03)00143-0
- Young, A., Powelson, E. B., Whitney, I. E., Raven, M. A., Nusinowitz, S., Jiang, M., et al. (2008). Involvement of OA1, an intracellular GPCR, and G alpha i3, its binding protein, in melanosomal biogenesis and optic pathway formation. *Invest. Ophthalmol. Vis. Sci.* 49, 3245–3252. doi: 10.1167/iovs.08-1806
- Yue, J. L., Okamura, H., Goshima, Y., Nakamura, S., Geffard, M., and Misu, Y. (1994). Baroreceptor-aortic nerve-mediated release of endogenous L-3,4dihydroxyphenylalanine and its tonic depressor function in the nucleus tractus solitarii of rats. *Neuroscience* 62, 145–161. doi: 10.1016/0306-4522(94)90321-2

- Zhang, X., Andren, P. E., and Svenningsson, P. (2006). Repeated l-DOPA treatment increases c-fos and BDNF mRNAs in the subthalamic nucleus in the 6-OHDA rat model of Parkinson's disease. *Brain Res.* 1095, 207–210. doi: 10.1016/j. brainres.2006.04.019
- Zhu, G., Okada, M., Yoshida, S., Hirose, S., and Kaneko, S. (2004). Determination of exocytosis mechanisms of DOPA in rat striatum using *in vivo* microdialysis. *Neurosci. Lett.* 367, 241–245. doi: 10.1016/j.neulet.2004.06.014

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Goshima, Masukawa, Kasahara, Hashimoto and Aladeokin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.