

Research Article

Nitro-Oxidative Stress after Neuronal Ischemia Induces Protein Nitrotyrosination and Cell Death

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Ischemic stroke is an acute vascular event that obstructs blood supply to the brain, producing irreversible damage that affects neurons but also glial and brain vessel cells. Immediately after the stroke, the ischemic tissue produces nitric oxide (NO) to recover blood perfusion but also produces superoxide anion. These compounds interact, producing peroxynitrite, which irreversibly nitrates protein tyrosines. The present study measured NO production in a human neuroblastoma (SH-SY5Y), a murine glial (BV2), a human endothelial cell line (HUVEC), and in primary cultures of human cerebral myocytes (HC-VSMCs) after experimental ischemia *in vitro*. Neuronal, endothelial, and inducible NO synthase (NOS) expression was also studied up to 24 h after ischemia, showing a different time course depending on the NOS type and the cells studied. Finally, we carried out cell viability experiments on SH-SY5Y cells with H₂O₂, a prooxidant agent, and with a NO donor to mimic ischemic conditions. We found that both compounds were highly toxic when they interacted, producing peroxynitrite. We obtained similar results when all cells were challenged with peroxynitrite. Our data suggest that peroxynitrite induces cell death and is a very harmful agent in brain ischemia.

1. Introduction

Ischemic stroke, the most common type of stroke, is an acute vascular accident caused by blockage in a brain vessel, which yields to a lack of oxygen supply that dramatically affects the brain parenchyma and brain vasculature. The tissue surrounding the ischemic lesion where neurons are still alive, but their viability that is highly compromised, called

the penumbra area, is a major target for ischemic stroke treatments [1, 2].

After ischemia, nitric oxide (NO) is released by endothelial cells to recover blood perfusion [3]. The NO is produced by the NO synthases (NOS), a family of enzymes coded by genes located in different chromosomes [3]. The major regulator of the vascular tone is the endothelium, through the endothelial NOS (eNOS). Diffusion of NO

to vascular smooth myocytes produces myorelaxation in a cyclic, guanosine monophosphate-dependent manner [4]. The neuronal (nNOS) and inducible (iNOS) enzymes are expressed in neurons, glial cells, and vascular myocytes [5–12] and may contribute to NO production in ischemic processes. Moreover, a burst of free radicals such as superoxide anion ($O_2^{\bullet-}$) [13, 14] is produced after the ischemic event. This scenario could be very harmful because superoxide anion has a high affinity for NO, higher than for the superoxide dismutase [15, 16], and both compounds react with each other and produce peroxynitrite ($ONOO^-$) [17]. Peroxynitrite has a gaseous nature that allows it to spread around the surrounding tissue to the ischemic focus. Peroxynitrite provokes protein nitrotyrosination, an irreversible chemical process consisting in the addition of a nitro group (NO_2) to the tyrosine residues to generate 3-nitrotyrosine [18, 19]. This posttranslational modification is pathological because it usually impairs the physiological function of the proteins [20, 21]. Evidence of protein nitrotyrosination in ischemic stroke has been reported [22].

The present study analyzed the role of the different cell types from brain parenchyma and brain vessels in NO production. Protein expression and mRNA levels of the different NOS were assessed. Moreover, we studied the effects of peroxynitrite on the viability of the different brain cells.

2. Materials and Methods

2.1. Cell Cultures. Human neuroblastoma cells (SH-SY5Y) were grown in DMEM (Gibco) supplemented with 15% fetal bovine serum (FBS; Gibco) and antibiotics (100 units/mL penicillin and 10^{-6} μ g/mL streptomycin; Gibco). HUVECs were grown in M-199 medium (Gibco) supplemented with 10% FBS, 3.2 mM glutamine (Sigma), and antibiotics. Murine microglial cells (BV2) were grown in RPMI (Gibco) supplemented with 10% FBS and antibiotics. Primary cultures of HC-VSMCs were produced from cerebral arteries (basilar) obtained from autopsies of 4 individuals (55.3 ± 5.6 years) and utilized up to the ten passages [23]. Procedure was approved by the ethics committee of the Institut Mar d'Investigació Mèdica and the Universitat Pompeu Fabra (IMIM-IMAS-UPF). Briefly, pieces of tunica media were incubated with 0.1% collagenase type IV (Sigma) for 35 min at 37°C and cultured to allow HC-VSMC migration to the flask surface. Cells were grown in DMEM with 4.5 g/L glucose (Sigma), 25 mM HEPES (Gibco), 10% FBS, 2.5 μ g/mL amphotericin B (Gibco), 100 units/mL penicillin, and 10^{-6} μ g/mL streptomycin. Myocytes were characterized by immunostaining with mouse anti-smooth muscle α -actin antibody (Ab; Sigma). Cells were used up to ten passages.

2.2. Mouse Embryo Hippocampal Cell Cultures. Hippocampal cells were isolated from 18-day-old CBl mouse embryos. Procedure was approved by the ethics committee of the IMIM-IMAS-UPF. Hippocampi were aseptically dissected and trypsinized. Cells were seeded in DMEM plus 10% horse serum (Gibco) into 1% poly-L-lysine (Sigma) coated coverslips (5×10^4 cells/cover). After 2 h, medium was removed,

and Neurobasal medium containing 1% B27 supplement (Gibco) plus antibiotics was added. Glial proliferation was avoided by adding 2 μ M cytosine arabinoside (Sigma) at day 3 for 24 h. Cultured hippocampal cells were used for the experiments on day 7, when they were considered to be mature neurons [24].

2.3. Human Brain Study. The human cortex sample was obtained from an autopsy of a patient who had a cortical ischemic stroke (Servei d'Anatomia Patològica, Hospital del Mar, Barcelona). The sample was fixed in formalin and embedded in paraffin. Procedure was approved by the ethics committee of the Institut Municipal d'Investigació Mèdica and the Universitat Pompeu Fabra (IMIM-IMAS-UPF).

2.4. In Vitro Ischemia. Ischemia was induced by an oxygen-glucose deprivation protocol (OGD). Briefly, the culture medium was replaced with a glucose-free balanced saline solution containing 116 mM NaCl, 5.4 mM KCl, 0.8 mM $MgSO_4$, 1 mM NaH_2PO_4 , 26.2 mM $NaHCO_3$, and 1.8 $CaCl_2$, incubated for 1 h in a nitrogen chamber at 37°C. The OGD medium was then replaced by culture medium, and cells were incubated for up to 12 h or 24 h after OGD under standard culture conditions.

2.5. Free Radical Production. SH-SY5Y cells were seeded on 1.5% gelatin-coated 35 mm coverslips (1×10^5 cells/well) for image analysis with a Leica TCS SP confocal microscope. SH-SY5Y cells were seeded in a 96-well plate (1×10^3 cells/well) for fluorescence quantification with a Fluostar Optima Microplate Reader (BMG Labtech). In both cases, cells were subjected to OGD with the OGD medium plus 5 μ M 2,7-dichlorofluorescein diacetate (DCF-DA; Sigma). The OGD medium was replaced by culture medium plus 5 μ M DCF-DA, cells were incubated in standard culture conditions, and experimental analysis was completed within 1 h.

2.6. NO Assay. HUVEC, SH-SY5Y, BV2, and HC-VSMC cells were seeded on T-25 flasks (approximately 5×10^6 cells/flask) and subjected to OGD. Control media, 12 h post-ischemic media and 24 h postischemic media were collected, and NO was measured using a nitrate/nitrite colorimetric assay kit (Cayman).

2.7. Immunofluorescence. Hippocampal cells (4×10^4 cells/well) were seeded on 1.5% gelatin coated, 12 mm coverslips. Cells were challenged for 1 h with OGD and maintained in standard culture conditions up to 24 h. Cells were fixed and incubated for 2 h at room temperature (RT) with 1:500 antinitrotyrosine polyclonal Ab (Chemikon), followed by incubation with 1:500 Alexa Fluor 488 goat anti-rabbit polyclonal Ab (Dako) for 1 h at RT. Nuclei were stained in blue with To-pro 3. The human cortex sample was cut at 3 μ m, deparaffinized at 70°C for 1 h, and washed with decreasing concentrations of ethanol. Antigen retrieval was performed with proteinase K at 40 μ g/mL in a 1:1 glycerol and TE buffer solution. Immunostaining was performed with 1:200 mouse monoclonal anti-N-Tyr Ab (Cayman Chem) for 2 h

at RT, followed by 1:1000 Alexa555-bound anti-mouse as secondary Ab (Dako) overnight at 4°C. Sections were stained with To-pro 3 to identify the nuclei. All the coverslips were mounted with Mowiol. Images were taken with a Leica TCS SP confocal microscope and analyzed with Leica confocal software (Leica).

2.8. Protein Identification by Western Blot. Cell cultures were lysed on ice with a solution containing 1M Tris-HCl, 1% Nonidet P-40, 150 mM NaCl, 5 mM EDTA, 1 mM sodium orthovanadate, 1 mM dithiothreitol, pH 7.4, and a protease inhibitor cocktail (Roche). Protein concentration was determined by Bio-Rad protein assay. Protein samples were analyzed using 3–8% Tris-acetate gels (Invitrogen) for NOS detection. Gels were run at 150 V for 1 h and transferred to nitrocellulose membranes (Millipore) at 100 V for 2 h. Membranes were blocked in Tween 20-Tris buffer solution (100 mM Tris-HCl, 150 mM NaCl, and pH 7.5; 0.1% Tween, 5% milk) and incubated for 2 h at 25°C with 1:1,000 anti-nNOS (Santa Cruz Biotechnology), anti-iNOS (Santa Cruz Biotech.), and anti-eNOS (Santa Cruz Biotech.) Abs. Peroxidase-conjugated donkey anti-rabbit and anti-mouse Abs (Amersham Bioscience) were used as secondary Abs at 1:5,000 for 1 h at 25°C. Bands were visualized with Super Signal (Pierce) and Amersham Bioscience Hyperfilm ECL kit.

2.9. RNA Isolation and Reverse Transcriptase-Polymerase Chain Reaction (RT-PCR) Analysis. Total RNA from cell cultures was isolated using Trizol reagent (Invitrogen), following the manufacturer instructions. Briefly, 1 µg of RNA was applied to RT-PCR using the OneStep RT-PCR Kit (Qiagen, Hamburg, Germany). The *iNOS*-specific primers for mice were 5'-CCATCACTGTGTTCCCC-3' and 5'-AAG-GTGGCAGCATCCCC-3' (Genbank accession number: NM_010927). The *iNOS*-specific primers for humans were 5'-CTGCTTGAGGTGGGCGG-3' and 5'-GTGACTCTGACTCGGGACGCC-3' (NM_000625). The *eNOS*-specific primers were 5'-CAAGTATGCCACCAACCGGG-3' and 5'-ACTGAAGGGGGCTGCGG-3' (NM_000603). The *nNOS*-specific primers were 5'-GAGAAGGAGCAGGGGGG-3' and 5'-CACATTGGCTGGGTCCCC-3' (NM_000620). These primers were used to amplify the different NOS. Hypoxanthine phosphoribosyltransferase- (*hprt*-) specific primers (5'-GGCCAGACTTTGTTGGATTTG-3' and 5'-TGCGCTCATCTTAGGCTTTGT-3'; NM000194) were used as positive control. Negative control was performed in the absence of oligonucleotide primers. Results were analyzed with Image Gauge software (Fuji Photo Film Co.).

2.10. Cell Viability Assay. Cells were seeded in 96-well plates at a density of 8,000 cells/100 µL per well and treated with 3-morpholinopyridone hydrochloride (SIN-1), sodium nitroprussiate (SNP), and/or H₂O₂, as described in the corresponding figures. Cells were incubated for 6 or 24 h at 37°C, and cell viability was measured by the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) reduction method. Absorbances at 540 and 650 nm were determined in

a Microplate Reader (Bio-Rad) and expressed as percentage of control.

2.11. Statistical Analysis. Data were expressed as the mean ± SEM of the values from the number of experiments indicated in the corresponding figures. Data were analyzed using one-way ANOVA with Bonferroni *post hoc* analysis.

3. Results

3.1. Ischemia Induces an Increase in Nitrotyrosination, Reactive Oxygen Species (ROS), and NO Production in Brain Cells. A strong labeling for nitrotyrosination was observed in the cortex from a stroke patient, not only in the brain parenchyma but also in the tunica intima and media of the blood vessel (Figure 1(a)). Neurons are very sensitive to oxidative stress. Therefore, we analyzed the protein nitrotyrosination after ischemia in mature mouse hippocampal neurons (Figure 1(b)). We assayed the effect of ischemia by subjecting the hippocampal neurons to OGD for 1 h, followed by reoxygenation. Neurons were maintained in a growth medium containing glucose for 24 h. We observed that hippocampal neurons subjected to ischemia had high levels of nitrotyrosination. The presence of nitrotyrosinated proteins demonstrated the existence of nitro-oxidative stress resulting from ischemia.

To study free-radical production after ischemia, we exposed human neuroblastoma cells (SH-SY5Y) to OGD. After 1 h, SH-SY5Y cells showed a burst in free radical production, measured by DCF oxidation (Figure 1(c), right) and visualized by confocal microscopy (Figure 1(c), left).

In addition to neurons, other brain cell types are found in the ischemic area; all cell types could contribute to the damage by producing NO. After the ischemic challenge, microglia and endothelial cells showed an increase in NO production (Figure 1(d)) that persisted up to 24 h. Endothelial cells and microglia were the highest NO producers in both baseline conditions and after ischemia; SH-SY5Y and brain vascular myocytes had no effect on NO release due to ischemia (Figure 1(d)).

3.2. Protein and mRNA Levels of NOS Types Are Modified in Brain Cells after Ischemia. NO production is dependent on the expression and activity of the different NOS. For this reason we examined the effect of ischemic insult on the protein and mRNA levels of *nNOS*, *eNOS*, and *iNOS*. In SH-SY5Y cells, we studied *nNOS* (Figure 2(a)). At 24 h after ischemia, we observed a significant reduction in *nNOS* protein ($P < 0.05$), but *nNOS* mRNA levels were not significantly affected (Figure 3(a)).

The main responsibility for the NO burst occurring after brain ischemia has classically been attributed to microglial cells. We studied the behavior of *iNOS* protein and mRNA in microglial cells (BV2) challenged with *in vitro* ischemia (Figure 2(b)). We found that *iNOS* is present in non-stimulated control microglial cells. Moreover, *iNOS* expression increased significantly at 12 h after the ischemic challenge ($P < 0.005$). The analysis of *iNOS* mRNA revealed, once

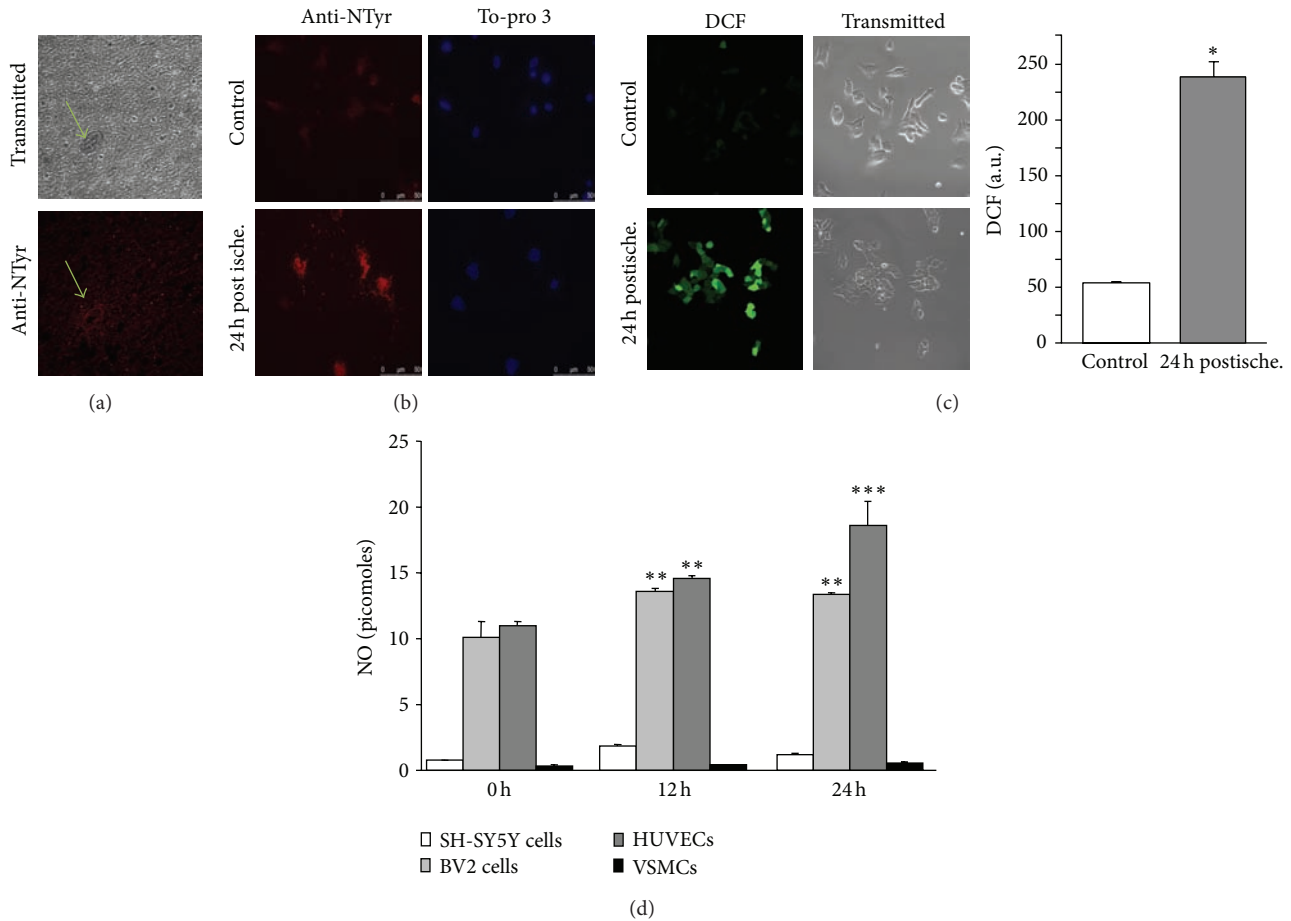


FIGURE 1: *In vitro* ischemia induces nitro-oxidative stress. (a) A representative image of nitrotyrosination (bottom image, stained in red) and bright field (top image) in a section of cortex from a stroke brain analyzed by immunohistochemistry. The arrows indicate blood vessels. (b) Primary hippocampal cells were subjected to *in vitro* ischemia followed by reperfusion with normal medium for 24 h. Nitrotyrosine (red staining) was demonstrated by immunostaining. Nuclei are stained in blue with To-pro 3. (c) Human neuroblastoma cells were subjected to *in vitro* ischemia reperused with normal medium for 24 h. Free radical production was detected by DCF fluorescence and quantified. The mean fluorescence of DCF represents the levels of ROS. Data are mean \pm SEM values of 4 independent experiments. * $P < 0.05$ versus control. (d) NO production (expressed in picomoles) was measured in neurons, microglia, endothelial, and vascular smooth muscle cells challenged with *in vitro* ischemia and later reoxygenated with normal growing medium containing glucose at 0, 12, and 24 h. Data are mean \pm SEM values of 4 independent experiments. ** $P < 0.01$; *** $P < 0.001$ versus controls at 0 h.

again, a drop in the *iNOS* mRNA levels at 12 h ($P < 0.05$) that was normalized 24 h after the ischemic insult (Figure 3(b)).

Regarding the expression of eNOS in endothelial cells (HUVEC), we did not see any significant change in the protein content after *in vitro* ischemia (Figure 2(c)), whereas the mRNA levels are upregulated after the ischemic challenge ($P < 0.05$; Figure 3(c)).

The iNOS and nNOS expression has been reported in systemic rat vascular smooth muscle cells [25–29]. Neither iNOS protein nor *iNOS* mRNA was detected in myocytes (primary cultures of HC-VSMC) in basal conditions or after ischemia (data not shown); nNOS protein and mRNA were clearly identified (Figure 2(d)). nNOS protein is constitutively present in cerebral vascular myocytes. We did not observe any difference in nNOS (Figure 2(d)) and mRNA levels (Figure 3(d)) following exposure to OGD medium.

3.3. ONOO⁻ Induces Cell Death. We used a NO donor (SNP) to demonstrate that NO alone is not toxic to human neuroblastoma cells at high physiological concentrations (Figure 4(a)), tested up to 1.5 mM SNP (Figure 4(a)). However, when SNP was applied together with nontoxic concentrations of H₂O₂ (an oxidative stress source when metabolized by cells), we obtained neurotoxicity due to the formation of ONOO⁻ at 10 ($P < 0.05$) and 20 ($P < 0.05$ and $P < 0.005$) μ M H₂O₂, even with the lowest SNP concentration (0.5 mM), which is closer to the physiological concentrations obtained in NO burst production.

The ONOO⁻ donor (SIN-1) was assayed at increasing concentrations in the different cell types (Figure 4(b)). The threshold to cause cytotoxicity was at 1 mM SIN-1 in endothelial cells and neuroblastoma cells. Myocytes and microglial cells were more susceptible to ONOO⁻, with the cytotoxicity threshold at 100 μ M SIN-1.

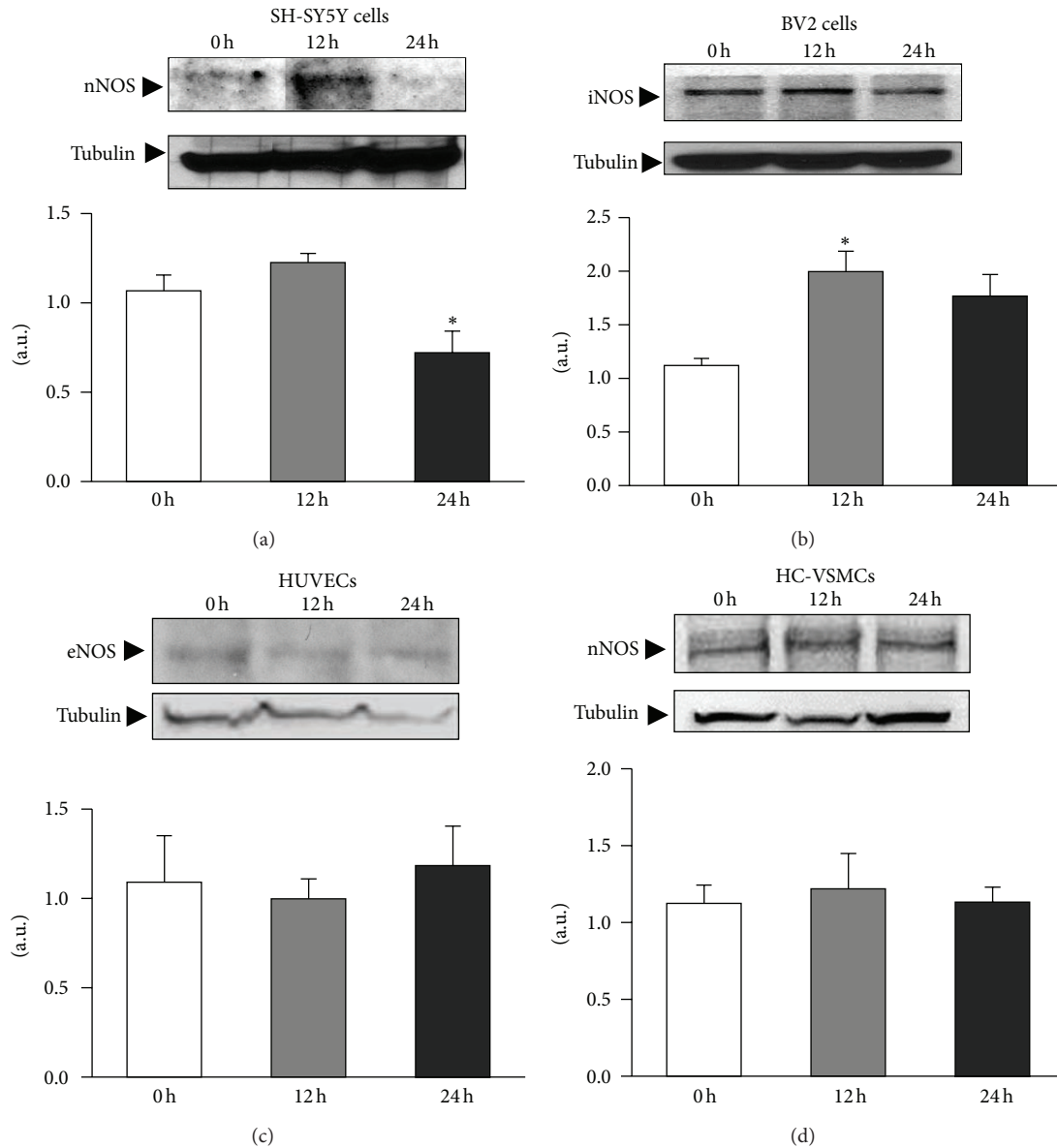


FIGURE 2: NO is produced in different cell types by the different NOS types. Cells were challenged with ischemia and iNOS, eNOS, and nNOS protein levels were studied immediately at time 0, 12, and 24 h after the ischemic challenge. Densitometric analysis of the bands quantified NOS expression relative to tubulin in all cell types. Data are mean \pm SEM values of 6 experiments for microglia and 3 experiments for neurons, endothelial cells, and myocytes. * $P < 0.05$ versus controls at 0 h.

4. Discussion

Stroke generates a cascade of molecular events as a consequence of arrested blood supply [30]. Immediately after the first minutes of ischemic cerebral damage, the activity of eNOS becomes elevated in an attempt to improve blood supply [31]. Our data demonstrate that NO is released after *in vitro* ischemia in brain cells. The NO production is activated up to 24 h after ischemia, mainly in endothelial and microglial cells. At first it might constitute a homeostatic response to increase blood perfusion, but studies performed in iNOS and nNOS knockout (KO) mice suggest that the activation of these two NOS after ischemia is highly harmful to brain cells [32, 33].

The regulation of NO levels depends on the levels and expression of the different NOS. The activity of constitutive NOS (eNOS and nNOS) is regulated by intracellular calcium that increases in response to ischemic challenge. iNOS is regulated at the transcriptional level because it lacks the regulatory arm activated by Ca^{2+} -calmodulin. Its transcription is activated by various stimuli, including LPS [34, 35].

We found significantly increased protein levels of iNOS in BV2 cells at 12 h after *in vitro* ischemia, and baseline levels were not totally recovered at 24 h. The regulation of mRNA expression for iNOS (BV2 cells) showed a decrease in mRNA levels at 12 h after ischemia, which correlates with high protein expression. The high production of NO by microglial

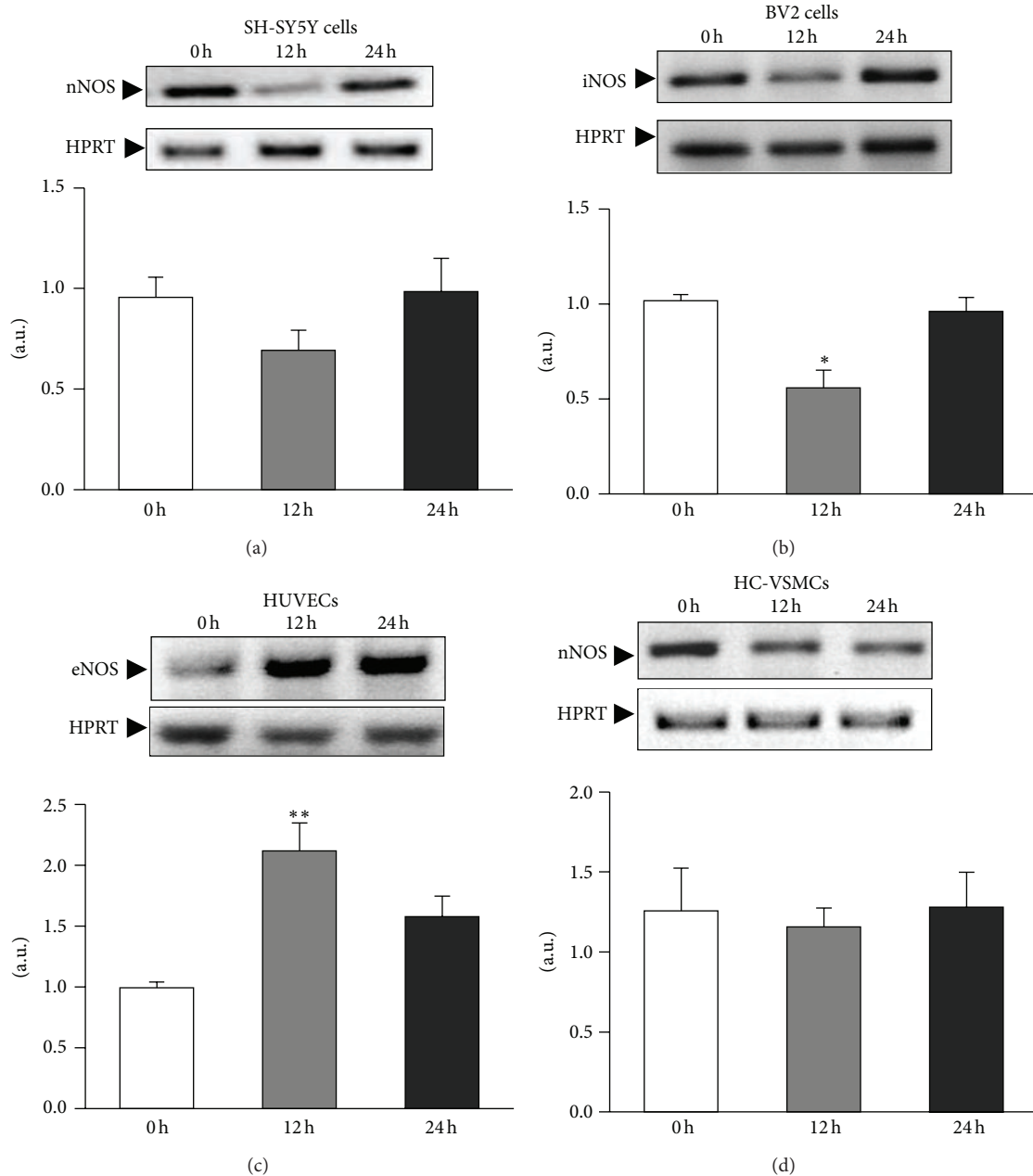


FIGURE 3: Differential expression of mRNA NOS types occurs in different cell types. Cells were challenged with ischemia, and *iNOS*, *eNOS*, and *nNOS* mRNA expression were studied immediately at time 0, 12 and 24 h after the ischemic challenge. The expression of mRNA was assessed by semiquantitative RT-PCR, and bands were quantified by *HPRT* densitometric analysis in all cell types. Data are mean \pm SEM values of 3 experiments in all cell types. * $P < 0.05$; ** $P < 0.01$ versus controls at 0 h.

cells after an ischemic challenge would be related to the nonspecific response of these cells to activation, as occurs with other immune cells [36] before phagocytosis of the cellular debris produced by postischemia necrosis [37]. This NO will not contribute to vasodilation but will mostly react with superoxide anion to form peroxynitrite. The *iNOS* KO mice had less neuronal death after ischemic stroke [32].

Postischemic protein levels (*eNOS* in HUVECs and *nNOS* in SH-SY5Y and HC-VSMCs) had not changed at 24 h. The *eNOS* mRNA was overexpressed at 12 h, which

strongly suggests an inhibitory translational control of *eNOS* expression after ischemia. In one study, neuronal death increased in *eNOS* KO mice after a stroke [38], suggesting a protective role for this enzyme that is likely due to its direct control of blood flow. On the other hand, *nNOS* mRNA levels in SH-SY5Y and HC-VSMCs did not change during postischemic observation.

Oxidative stress plays a key role in ischemic-reperfusion situations [13] because mitochondrial dysfunction leads to a burst in free radical production that cannot be scavenged

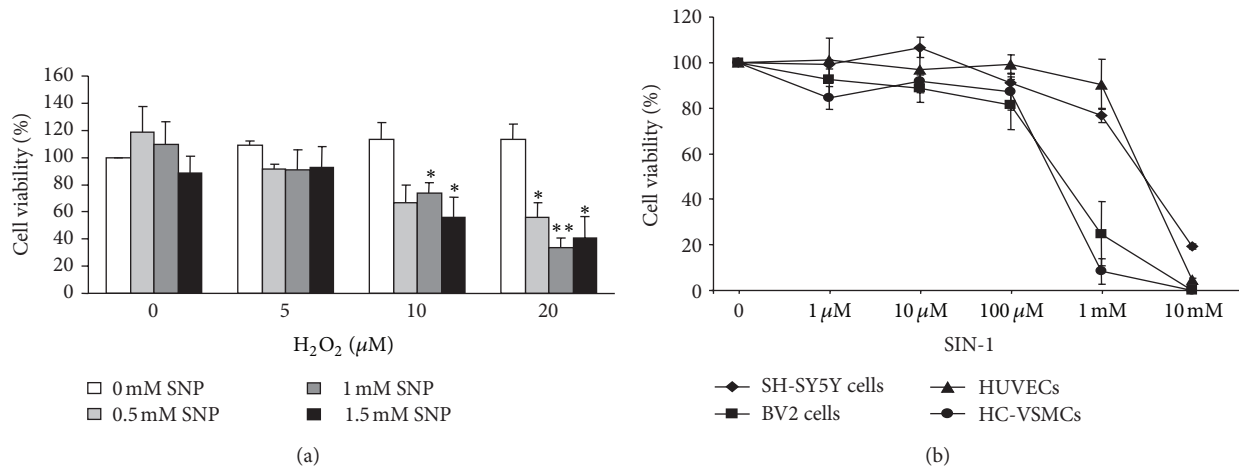


FIGURE 4: ONOO⁻ induces cell death. (a) Human neuroblastoma cells were treated with increasing concentrations of a NO donor (SNP) and H₂O₂ (free radical source). Cells were incubated for 6 h, and cell viability was assayed by MTT reduction. Data are expressed as percentage of control cells. Data are mean ± SEM values of 7 experiments. **P* < 0.05; ***P* < 0.01 versus controls without SNP. (b) Cells were challenged with increasing concentrations of the ONOO⁻ donor SIN-1, and cell viability was assayed by MTT reduction.

by the constitutive antioxidant cellular defense systems [39]. The large amount of free radicals produced during ischemia not only scavenges NO but also transforms it into the toxic ONOO⁻ [17], which yields to the protein nitrotyrosination and cell death observed in the present study. Although ONOO⁻ is a short lived molecule [40], its ability to diffuse through biological membranes can spread its harmful effects into neighboring cells and tissues. This critical process defines survival or death in the penumbra area. Consequently, we analyzed the effects of NO, H₂O₂, and NO plus H₂O₂ on neuronal survival. Neurotoxicity was only induced when cells were challenged with NO plus H₂O₂, which will produce peroxynitrite. Furthermore, we assayed increasing concentrations of a peroxynitrite donor, SIN-1, on all the brain cell types. We found a similar pattern of cell viability for all the cell types at low micromolar concentrations but a major cytotoxicity for glial cells and HC-VSMCs at 1 mM SIN-1. This effect may be related to a lower antioxidant defense. Further work is needed to investigate the possible protective role of antioxidants in the prevention of ischemic damage [41].

5. Conclusion

Our work demonstrates that brain ischemia induces nitrooxidative stress that produces protein nitrotyrosination. The high production of peroxynitrite after ischemia will cause neuronal death and dramatically affect the survival of other brain cells.

Nonstandard Abbreviations Used

Ab: Antibody
 BSA: Bovine serum albumin
 BV2: Mouse microglial cells
 eNOS: Endothelial nitric oxide synthase
 FBS: Fetal bovine serum

HC-VSMC: Human cerebral vascular smooth myocytes
 hPRT: Hypoxanthine phosphoribosyltransferase
 HUVEC: Human umbilical vein endothelial cells
 iNOS: Inducible nitric oxide synthase
 MTT: 3-(4,5-Dimethylthiazol-2-yl)2,5-diphenyltetrazolium bromide
 NO: Nitric oxide
 NOS: Nitric oxide synthase
 O₂⁻: Superoxide anion
 OGD: Oxygen-glucose deprivation
 ONOO⁻: Peroxynitrite anion
 RT: Room temperature
 RT-PCR: Reverse transcriptase-polymerase chain reaction
 SH-SY5Y: Human neuroblastoma cells
 SIN-1: 3-Morpholiniosydnonimine hydrochloride.

Conflict of Interests

The authors indicate no financial conflict of interests.

Acknowledgments

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References

- [1] I. Ferrer and A. M. Planas, "Signaling of cell death and cell survival following focal cerebral ischemia: Life and death struggle in the penumbra," *Journal of Neuropathology and Experimental Neurology*, vol. 62, no. 4, pp. 329–339, 2003.
- [2] K. Jin, X. Wang, L. Xie et al., "Evidence for stroke-induced neurogenesis in the human brain," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 35, pp. 13198–13202, 2006.
- [3] F. X. Guix, I. Uribealago, M. Coma, and F. J. Muñoz, "The physiology and pathophysiology of nitric oxide in the brain," *Progress in Neurobiology*, vol. 76, no. 2, pp. 126–152, 2005.
- [4] P. L. Huang, "Neuronal and endothelial nitric oxide synthase gene knockout mice," *Brazilian Journal of Medical and Biological Research*, vol. 32, no. 11, pp. 1353–1359, 1999.
- [5] I. N. Mungrue, D. S. Bredt, D. J. Stewart, and M. Husain, "From molecules to mammals: what's NOS got to do with it?" *Acta Physiologica Scandinavica*, vol. 179, no. 2, pp. 123–135, 2003.
- [6] P. A. Marsden, H. H. Q. Heng, S. W. Scherer et al., "Structure and chromosomal localization of the human constitutive endothelial nitric oxide synthase gene," *The Journal of Biological Chemistry*, vol. 268, no. 23, pp. 17478–17488, 1993.
- [7] C. J. Lowenstein, C. S. Glatt, D. S. Bredt, and S. H. Snyder, "Cloned and expressed macrophage nitric oxide synthase contrasts with the brain enzyme," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 89, no. 15, pp. 6711–6715, 1992.
- [8] S. L. Elfering, T. M. Sarkela, and C. Giulivi, "Biochemistry of mitochondrial nitric-oxide synthase," *The Journal of Biological Chemistry*, vol. 277, no. 41, pp. 38079–38086, 2002.
- [9] H. J. Cho, Q.-W. Xie, J. Calaycay et al., "Calmodulin is a subunit of nitric oxide synthase from macrophages," *Journal of Experimental Medicine*, vol. 176, no. 2, pp. 599–604, 1992.
- [10] D. S. Bredt and S. H. Snyder, "Nitric oxide: a physiological messenger molecule," *Annual Review of Biochemistry*, vol. 63, pp. 175–195, 1994.
- [11] D. S. Bredt and S. H. Snyder, "Transient nitric oxide synthase neurons in embryonic cerebral cortical plate, sensory ganglia, and olfactory epithelium," *Neuron*, vol. 13, no. 2, pp. 301–313, 1994.
- [12] C. Angeloni and S. Hrelia, "Quercetin reduces inflammatory responses in LPS-stimulated cardiomyoblasts," *Oxidative Medicine and Cellular Longevity*, vol. 2012, Article ID 837104, 8 pages, 2012.
- [13] S. Cuzzocrea, D. P. Riley, A. P. Caputi, and D. Salvemini, "Antioxidant therapy: a new pharmacological approach in shock, inflammation, and ischemia/reperfusion injury," *Pharmacological Reviews*, vol. 53, no. 1, pp. 135–159, 2001.
- [14] M. M. H. El Kossi and M. M. Zakhary, "Oxidative stress in the context of acute cerebrovascular stroke," *Stroke*, vol. 31, no. 8, pp. 1889–1892, 2000.
- [15] R. E. Huie and S. Padmaja, "The reaction of no with superoxide," *Free Radical Research Communications*, vol. 18, no. 4, pp. 195–199, 1993.
- [16] A. Cudd and I. Fridovich, "Electrostatic interactions in the reaction mechanism of bovine erythrocyte superoxide dismutase," *The Journal of Biological Chemistry*, vol. 257, no. 19, pp. 11443–11447, 1982.
- [17] J. S. Beckman, T. W. Beckman, J. Chen, P. A. Marshall, and B. A. Freeman, "Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 87, no. 4, pp. 1620–1624, 1990.
- [18] J. M. Souza, E. Daikhin, M. Yudkoff, C. S. Raman, and H. Ischiropoulos, "Factors determining the selectivity of protein tyrosine nitration," *Archives of Biochemistry and Biophysics*, vol. 371, no. 2, pp. 169–178, 1999.
- [19] M. Brennan, W. Wu, X. Fu et al., "A tale of two controversies. Defining both the role of peroxidases in nitrotyrosine formation in vivo using eosinophil peroxidase and myeloperoxidase-deficient mice, and the nature of peroxidase-generated reactive nitrogen species," *The Journal of Biological Chemistry*, vol. 277, no. 20, pp. 17415–17427, 2002.
- [20] D. K. Newman, S. Hoffman, S. Kotamraju et al., "Nitration of PECAM-1 ITIM tyrosines abrogates phosphorylation and SHP-2 binding," *Biochemical and Biophysical Research Communications*, vol. 296, no. 5, pp. 1171–1179, 2002.
- [21] M. M. Elahi, Y. X. Kong, and B. M. Matata, "Oxidative stress as a mediator of cardiovascular disease," *Oxidative Medicine and Cellular Longevity*, vol. 2, no. 5, pp. 259–269, 2009.
- [22] F. J. Pérez-Asensio, X. De La Rosa, F. Jiménez-Altayó et al., "Antioxidant CR-6 protects against reperfusion injury after a transient episode of focal brain ischemia in rats," *Journal of Cerebral Blood Flow and Metabolism*, vol. 30, no. 3, pp. 638–652, 2010.
- [23] M. Coma, F. X. Guix, G. Ill-Raga et al., "Oxidative stress triggers the amyloidogenic pathway in human vascular smooth muscle cells," *Neurobiology of Aging*, vol. 29, no. 7, pp. 969–980, 2008.
- [24] S. Kaeck and G. Banker, "Culturing hippocampal neurons," *Nature Protocols*, vol. 1, no. 5, pp. 2406–2415, 2006.
- [25] M. L. Arbones, J. Ribera, L. Agullo et al., "Characteristics of nitric oxide synthase type I of rat cerebellar astrocytes," *Glia*, vol. 18, pp. 224–232, 1996.
- [26] R. J. Cork, M. L. Perrone, D. Bridges, J. Wandell, C. A. Scheiner, and R. R. Mize, "A web-accessible digital atlas of the distribution of nitric oxide synthase in the mouse brain," *Progress in Brain Research*, vol. 118, pp. 37–50, 1998.
- [27] L. Kobzik, M. B. Reid, D. S. Bredt, and J. S. Stamler, "Nitric oxide in skeletal muscle," *Nature*, vol. 372, no. 6506, pp. 546–548, 1994.
- [28] N. A. Mohammed, S. Abd El-Aleem, I. Appleton, M. M. Maklout, M. Said, and R. F. T. McMathon, "Expression of nitric oxide synthase isoforms in human liver cirrhosis," *Journal of Pathology*, vol. 200, no. 5, pp. 647–655, 2003.
- [29] K. Y. Xu, D. L. Huso, T. M. Dawson, D. S. Bredt, and L. C. Becker, "Nitric oxide synthase in cardiac sarcoplasmic reticulum," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 96, no. 2, pp. 657–662, 1999.
- [30] M. A. Moro, A. Cárdenas, O. Hurtado, J. C. Leza, and I. Lizasoain, "Role of nitric oxide after brain ischaemia," *Cell Calcium*, vol. 36, no. 3–4, pp. 265–275, 2004.
- [31] C. Depre, J. L. Vanoverschelde, B. Gerber et al., "Correlation of functional recovery with myocardial blood flow, glucose uptake, and morphologic features in patients with chronic left ventricular ischemic dysfunction undergoing coronary artery bypass

- grafting," *Journal of Thoracic and Cardiovascular Surgery*, vol. 113, no. 2, pp. 371–378, 1997.
- [32] C. Iadecola, F. Zhang, R. Casey, M. Nagayama, and M. Elizabeth Ross, "Delayed reduction of ischemic brain injury and neurological deficits in mice lacking the inducible nitric oxide synthase gene," *Journal of Neuroscience*, vol. 17, no. 23, pp. 9157–9164, 1997.
- [33] M. J. L. Eliasson, Z. Huang, R. J. Ferrante et al., "Neuronal nitric oxide synthase activation and peroxynitrite formation in ischemic stroke linked to neural damage," *Journal of Neuroscience*, vol. 19, no. 14, pp. 5910–5918, 1999.
- [34] R. Kamijo, H. Harada, T. Matsuyama et al., "Requirement for transcription factor IRF-1 in NO synthase induction in macrophages," *Science*, vol. 263, no. 5153, pp. 1612–1615, 1994.
- [35] Q. Xie, Y. Kashiwabara, and C. Nathan, "Role of transcription factor NF- κ B/Rel in induction of nitric oxide synthase," *The Journal of Biological Chemistry*, vol. 269, no. 7, pp. 4705–4708, 1994.
- [36] C. Bogdan, "Nitric oxide and the immune response," *Nature Immunology*, vol. 2, no. 10, pp. 907–916, 2001.
- [37] M. Xu and H. Zhang, "Death and survival of neuronal and astrocytic cells in ischemic brain injury: a role of autophagy," *Acta Pharmacologica Sinica*, vol. 32, no. 9, pp. 1089–1099, 2011.
- [38] Z. Huang, P. L. Huang, J. Ma et al., "Enlarged infarcts in endothelial nitric oxide synthase knockout mice are attenuated by nitro-L-arginine," *Journal of Cerebral Blood Flow and Metabolism*, vol. 16, no. 5, pp. 981–987, 1996.
- [39] R. Taffi, L. Nanetti, L. Mazzanti et al., "Plasma levels of nitric oxide and stroke outcome," *Journal of Neurology*, vol. 255, no. 1, pp. 94–98, 2008.
- [40] C. Szabó, H. Ischiropoulos, and R. Radi, "Peroxynitrite: biochemistry, pathophysiology and development of therapeutics," *Nature Reviews Drug Discovery*, vol. 6, no. 8, pp. 662–680, 2007.
- [41] M. K. Hagen, A. Ludke, A. S. Araujo et al., "Antioxidant characterization of soy derived products in vitro and the effect of a soy diet on peripheral markers of oxidative stress in a heart disease model," *Canadian Journal of Physiology and Pharmacology*, vol. 90, pp. 1095–1103, 2012.