



Draft Genome Sequences of *Pseudomonas* spp. Isolated from Berry Surfaces in Commercial Cranberry Bogs in Massachusetts, USA

Marit H. Koszewski,^a Sheyda Motevalli,^a DScott D. Soby^{a,b}

^aBiomedical Sciences Program, College of Graduate Studies, Midwestern University, Glendale, Arizona, USA ^bCollege of Veterinary Medicine, Midwestern University, Glendale, Arizona, USA

Marit H. Koszewski and Sheyda Motevalli contributed equally to this work. Author order was determined alphabetically.

ABSTRACT The surfaces of plants are colonized by a rich diversity of microbes but are largely unexplored. Here, we present the draft genome sequences of five *Pseudomonas* spp. isolated from cultivated cranberry fruit surfaces. Although the isolates represent four different species, their genomes all contain conserved iron sequestration and uptake genes.

The genus *Pseudomonas* (*Gammaproteobacteria*) is widely recognized as being among the most diverse and ubiquitous bacterial taxa, with 242 currently validated species (https://lpsn.dsmz.de/genus/pseudomonas). Members of the genus include human, animal, and plant pathogens (1–4), inhabit diverse habitats (5–8), and play important roles in plant growth, development, and protection from disease (9–11). We recently demonstrated that *Pseudomonas* spp. isolated from cranberry plants produce volatile organic compounds that inhibit the growth of several types of plant-associated fungi and *Phytophthora cinnamomi* (12). Despite their ubiquity and importance, little is known about the *Pseudomonas* spp. that inhabit the surfaces of plant organs or what their functional roles are in those niches. Recently, we explored the bacteria colonizing the surfaces of cranberry plants (*Vaccinium macrocarpon* Ait.) (13–17). The ability to analyze and compare the genomes of these nonpathogenic commensal bacteria is providing new insights into the relationships between plants and their microbiomes and may yield new methods for controlling fungal infections that lead to crop loss.

Bacteria were isolated from berries that were aseptically collected in August 2010 from commercial cranberry bogs. Berries were vortexed in sterile water, and the water was plated on King's medium B (KMB) agar containing $50 \,\mu g \,ml^{-1}$ each of cycloheximide and ampicillin. Single colonies that fluoresced under long-wave UV light were transferred to fresh medium, colony purified 3 times, and stored at -80° C in 34% glycerol. Isolates were placed in the genus Pseudomonas by phenotype and 16S rRNA gene sequences amplified with 27F and 1525R primers using BLAST (18). Taxonomic placement was verified using the Type (Strain) Genome Server (Fig. 1) (19). Isolates were recovered from frozen storage, streaked onto KMB agar, and inoculated into overnight KMB broth cultures for genomic DNA (gDNA) isolation with a DNeasy blood and tissue kit (Qiagen). Genomic DNA libraries (KAPA HyperPlus library preparation kit) were analyzed for fragment size with an Agilent TapeStation and quantified by quantitative PCR (qPCR) (KAPA library quantification kit) with a QuantStudio 5 system (Thermo Fisher Scientific) before sequencing (Illumina MiSeq 2×250 -bp flow cell). Raw reads were assembled using Unicycler+ with SPAdes and Pilon version 1.23 for polishing within the PATRIC Comprehensive Genome Analysis pipeline version 3.6.8 with default settings (http://patricbrc.org) (20) (Table 1). The compiled genome sequences were annotated using RASTtk (21).

Citation Koszewski MH, Motevalli S, Soby SD. 2021. Draft genome sequences of *Pseudomonas* spp. isolated from berry surfaces in commercial cranberry bogs in Massachusetts, USA. Microbiol Resour Announc 10:e00204-21. https://doi.org/10 .1128/MRA.00204-21.

Editor David A. Baltrus, University of Arizona Copyright © 2021 Koszewski et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Scott D. Soby, ssobyx@midwestern.edu.

Received 15 March 2021 Accepted 11 June 2021 Published 8 July 2021

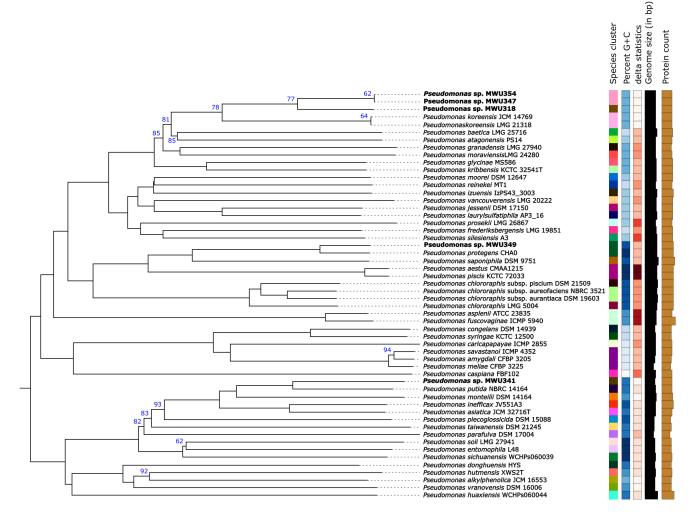


FIG 1 Genome BLAST Distance Phylogeny (GBDP) tree. A phylogenetic tree was constructed with the Type (Strain) Genome Server (19), which produces a GBDP tree by approximating intergenomic relatedness using the MASH algorithm among all type strain genomes in the TYGS database and by extracting and comparing 16S rRNA gene sequences with 12,670 type strains using BLAST as a proxy to identify the 50 closest type strains to calculate precise distances. The tree itself was constructed using FastME version 2.1.4 to infer a balanced minimum evolution tree with branch support (23). The tree represents only the *Pseudomonas* spp. most closely related to the described isolates. Bootstrap support values are shown at the nodes. Nodes without values have 100% bootstrap support. Isolates described in the text are in bold font. Isolates MWU347 and MWU354 are the same species, MWU347/ MWU354, MWU318, and MWU341 represent new species, and MWU349 is *P. protegens*.

Isolate MWU349 is *Pseudomonas protegens sensu lato*, but the other isolates were not assigned to a specific taxon (Fig. 1). MWU354 and MWU347 are members of the same *nova species* but are not clonal isolates. As an indication of the importance of iron sequestration in the berry surface microenvironment (22), each of the isolates has multiple siderophore-related genes, including nonribosomal peptide synthases for the production of pyoverdine-like siderophores. TonB-dependent hemin receptors, iron siderophore sensor proteins, pyoverdine chromophore precursor synthase PvdL, and the iron dicitrate transport protein FecA are conserved across all of the isolates.

Data availability. The *Pseudomonas* sp. strain MWU318, MWU341, MWU347, MWU349, and MWU354 genome sequences have been deposited in GenBank under BioProject number PRJNA691338. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the whole-genome sequence (WGS) and SRA accession numbers in Table 1.

ACKNOWLEDGMENTS

Library construction and Illumina sequencing were performed at the Arizona State University CLAS Genomics Core facility.

isolates
Pseudomonas sp.
of P
enome features
6
TABLE

							0+0				No. of	No. of
	Assigned	Collection	Genome	No. of	N ₅₀ contig	Coverage	content	BioSample	GenBank	SRA accession	coding	siderophore-
lsolate	taxon	site	size (bp)	contigs	size (bp)	(×)	(%)	no.	accession no.	no.	seduences	related genes
MWU318	AWU318 Nov. sp.	West Wareham, MA 6,035,781		33	489,847	174	60.3	SAMN17284639	JAESJL000000000	SRX10176289	5,150	17
MWU341	P. protegens	P. protegens Carver, MA	5,851,990	72	336,634	182	62.4	SAMN17284726	JAERIH000000000	SRX10166717	5,460	21
MWU347	Nov. sp.	East Wareham, MA	6,137,901	45	556,804	87	60.3	SAMN17284874	JAFGZB0000000000	SRX10299995	5,581	17
MWU349	Nov. sp.	East Wareham, MA	6,715,860 18	18	906,199	228	63.3	SAMN17284895	JAFEVP000000000	SRX10166718	6,199	27
MWU354	Nov. sp.	East Wareham, MA 6,137,275 46	6,137,275	46	531,002	92	60.3	SAMN17284896	JAFGZC0000000000	SRX10299996	5,572	17

This research was supported by the College of Graduate Studies and Biomedical Sciences Program, Midwestern University.

We thank Zachary Hummel for the preliminary characterization of the isolates, including analysis of the 16S rRNA genes, and Alisha Harrison for isolation of gDNA and technical support. We acknowledge the generous cooperation of the UMASS Cranberry Station and members of the Massachusetts Cranberry Growers Association for access to plant materials.

REFERENCES

- 1. Pye C. 2018. Pseudomonas otitis externa in dogs. Can Vet J 59:1231–1234.
- Driscoll JA, Brody SL, Kollef MH. 2007. The epidemiology, pathogenesis and treatment of Pseudomonas aeruginosa infections. Drugs 67:351–368. https://doi.org/10.2165/00003495-200767030-00003.
- Hesse C, Schulz F, Bull CT, Shaffer BT, Yan Q, Shapiro N, Hassan KA, Varghese N, Elbourne LDH, Paulsen IT, Kyrpides N, Woyke T, Loper JE. 2018. Genome-based evolutionary history of Pseudomonas spp. Environ Microbiol 20:2142–2159. https://doi.org/10.1111/1462-2920.14130.
- Xin XF, Kvitko B, He SY. 2018. Pseudomonas syringae: what it takes to be a pathogen. Nat Rev Microbiol 16:316–328. https://doi.org/10.1038/nrmicro .2018.17.
- Lopes LD, Davis EW, Pereira E Silva MDC, Weisberg AJ, Bresciani L, Chang JH, Loper JE, Andreote FD. 2018. Tropical soils are a reservoir for fluorescent Pseudomonas spp. biodiversity. Environ Microbiol 20:62–74. https:// doi.org/10.1111/1462-2920.13957.
- Monteil CL, Bardin M, Morris CE. 2014. Features of air masses associated with the deposition of Pseudomonas syringae and Botrytis cinerea by rain and snowfall. ISME J 8:2290–2304. https://doi.org/10.1038/ismej.2014.55.
- Amoozegar MA, Shahinpei A, Sepahy AA, Makhdoumi-Kakhki A, Seyedmahdi SS, Schumann P, Ventosa A. 2014. Pseudomonas salegens sp. nov., a halophilic member of the genus Pseudomonas isolated from a wetland. Int J Syst Evol Microbiol 64:3565–3570. https://doi.org/10.1099/ijs.0.062935-0.
- Vyas P, Rahi P, Gulati A. 2009. Stress tolerance and genetic variability of phosphate-solubilizing fluorescent Pseudomonas from the cold deserts of the trans-Himalayas. Microb Ecol 58:425–434. https://doi.org/10.1007/ s00248-009-9511-2.
- Duke KA, Becker MG, Girard IJ, Millar JL, Dilantha Fernando WG, Belmonte MF, de Kievit TR. 2017. The biocontrol agent Pseudomonas chlororaphis PA23 primes Brassica napus defenses through distinct gene networks. BMC Genomics 18:467. https://doi.org/10.1186/s12864-017-3848-6.
- Mannaa M, Kim KD. 2018. Biocontrol activity of volatile-producing Bacillus megaterium and Pseudomonas protegens against Aspergillus and Penicillium spp. predominant in stored rice grains: study II. Mycobiology 46:52–63. https://doi.org/10.1080/12298093.2018.1454015.
- Ramette A, Frapolli M, Fischer-Le Saux M, Gruffaz C, Meyer J-M, Défago G, Sutra L, Moënne-Loccoz Y. 2011. Pseudomonas protegens sp. nov., widespread plant-protecting bacteria producing the biocontrol compounds 2,4-diacetylphloroglucinol and pyoluteorin. Syst Appl Microbiol 34:180–188. https://doi.org/10.1016/j.syapm.2010.10.005.
- Ebadzadsahrai G, Higgins Keppler EA, Soby SD, Bean HD. 2020. Inhibition of fungal growth and induction of a novel volatilome in response to Chromobacterium vaccinii volatile organic compounds. Front Microbiol 11:1035. https://doi.org/10.3389/fmicb.2020.01035.

- Ebadzadsahrai G, Soby S. 2020. Draft genome sequence of Pseudomonas sp. strain MWU12-2323, isolated from a wild cranberry bog in Truro, Massachusetts. Microbiol Resour Announc 9:e01387-19. https://doi.org/10.1128/ MRA.01387-19.
- 14. Ebadzadsahrai G, Soby S. 2019. 16S rRNA amplicon profiling of cranberry (Vaccinium macrocarpon Ait.) flower and berry surfaces. Microbiol Resour Announc 8:e01479-18. https://doi.org/10.1128/MRA.01479-18.
- Ebadzadsahrai G, Thomson J, Soby S. 2018. Draft genome sequence of Pseudomonas sp. strain MWU13-2860, isolated from a wild cranberry bog in Truro, Massachusetts. Microbiol Resour Announc 7:e01007-18. https:// doi.org/10.1128/MRA.01007-18.
- Ebadzadsahrai G, Thomson J, Soby S. 2018. Draft genome sequence of Pseudomonas sp. strain MWU12-2534b, isolated from a wild cranberry bog in Truro, Massachusetts. Microbiol Resour Announc 7:e01005-18. https://doi.org/10.1128/MRA.01005-18.
- Ebadzadsahrai G, Thomson J, Soby S. 2018. Draft genome sequences of Pseudomonas MWU13-2625 and MWU12-2115, isolated from a wild cranberry bog at the Cape Cod National Seashore. Microbiol Resour Announc 7:e00992-18. https://doi.org/10.1128/MRA.00992-18.
- Auch AF, Henz S, Holland B, Göker M. 2006. Genome BLAST distance phylogenies inferred from whole plastid and whole mitochondrion genome sequences. BMC Bioinformatics 7:350. https://doi.org/10.1186/1471-2105-7-350.
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. Nat Commun 10:2182. https://doi.org/10.1038/s41467-019-10210-3.
- Wattam AR, Davis JJ, Assaf R, Boisvert S, Brettin T, Bun C, Conrad N, Dietrich EM, Disz T, Gabbard JL, Gerdes S, Henry CS, Kenyon RW, Machi D, Mao C, Nordberg EK, Olsen GJ, Murphy-Olson DE, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Vonstein V, Warren A, Xia F, Yoo H, Stevens RL. 2017. Improvements to PATRIC, the all-bacterial Bioinformatics Database and Analysis Resource Center. Nucleic Acids Res 45: D535–D542. https://doi.org/10.1093/nar/gkw1017.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. https://doi.org/10.1038/srep08365.
- Kramer J, Özkaya Ö, Kümmerli R. 2020. Bacterial siderophores in community and host interactions. Nat Rev Microbiol 18:152–163. https://doi.org/ 10.1038/s41579-019-0284-4.
- Lefort V, Desper R, Gascuel O. 2015. FastME 2.0: a comprehensive, accurate, and fast distance-based phylogeny inference program. Mol Biol Evol 32:2798–2800. https://doi.org/10.1093/molbev/msv150.