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Potential pathological and molecular mechanisms of temporomandibular joint osteoarthritis



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KEYWORDS

Temporomandibular joint osteoarthritis; Inflammation; Extracellular matrix; Biological behaviors; Angiogenesis; Molecular pathways Abstract Temporomandibular joint osteoarthritis (TMJ OA) is a progressive degenerative disease of the temporomandibular joint (TMJ). The unclear etiology and mechanisms of TMJ OA bring great difficulties to early diagnosis and effective treatment, causing enormous burdens to patients' life and social economics. In this narrative review, we summarized the main pathological changes of TMJ OA, including inflammatory responses, degeneration of extracellular matrix (ECM), abnormal cell biological behaviors (apoptosis, autophagy, and differentiation) in TMJ tissue, and aberrant angiogenesis. All pathological features are closely linked to each other, forming a vicious cycle in the process of TMJ OA, which results in prolonged disease duration and makes it difficult to cure. Various molecules and signaling pathways are involved in TMJ OA pathogenesis, including nuclear factor kappa-B (NF-kB), mitogen-activated protein kinases (MAPKs), extracellular regulated protein kinases (ERKs) and transforming growth factor (TGF)- β signaling pathways et al. One molecule or pathway can contribute to several pathological changes, and the crosstalk between different molecules and pathways can further lead to a complicated condition TMJ OA. TMJ OA has miscellaneous etiology, complex clinical status, depressed treatment results, and poor prognosis. Therefore, novel in-vivo and in-vitro models, novel medicine, materials, and approaches for therapeutic procedures might be helpful for further investigation of TMJ OA. Furthermore, the role of genetic factors in TMJ OA needs to be elucidated to establish more reasonable and effective clinical strategies for diagnosing and treating TMJ OA.

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Introduction

Temporomandibular joint osteoarthritis (TMJ OA) is a progressive degenerative disease in the temporomandibular joint (TMJ) tissue. Pain, clicking in the temporomandibular region, and abnormal mandibular movements are considered to be the main symptoms of TMJ OA. TMJ OA is the most common subtype of temporomandibular disorders (TMD), affecting about 30% of the population in the Chinese mainland. Furthermore, 14.56% of TMD patients show definite imaging lesions on the articular surface and bone tissue. Aging, gender, and mechanical stress are the most common factors of TMJ OA.¹ Other factors, including trauma, body postures, oral parafunctional habits, and psychological stress, are also associated with the etiology of TMJ OA.² Currently, the main therapeutic principle for TMJ OA is conservative treatment, including joint protection, physical therapy (physical movement exercise and laser therapy), occlusal splints, nonsteroidal antiinflammatory drugs (NSAIDs), and glucosamine sulfate. Unfortunately, due to the ambiguity of etiology and pathogenesis, current conservative therapeutic approaches for TMJ OA seem ineffective for curing this long-lasting disease, Moreover, TMJ OA may finally lead to limitation of jaw movement, and even orofacial deformity in adolescences, which burdens patients' lives and social economy.

Inflammatory responses, degeneration of extracellular matrix (ECM), abnormal cell biological behaviors, and angiogenesis in TMJ tissue are the main pathological changes of TMJ OA.³ Inflammatory responses, including cytokines release, phagocyte chemotaxis, and macrophage polarization, can lead to the breakdown of the homeostasis of TMJ tissue, resulting in the imbalance of the anabolic and catabolic activities leading to ECM degeneration. Inflammatory factors and catabolic product accumulation can further cause hypoxia and oxidative stress to cells in TMJ tissue, leading to abnormal cell biological behaviors and abnormal angiogenesis. Cell and vascular abnormality can cause a more severe inflammatory and degenerative status, forming a vicious circle and a protracted course of TMJ OA (Fig. 1).

Accordingly, we make a comprehensive review of the present studies conducted on pathological and corresponding molecular mechanisms of TMJ OA, aiming to present a promising aspect of utilizing the mechanisms for targeting the suppression of TMJ OA.

Pathological changes in temporomandibular joint osteoarthritis

Inflammatory responses

Inflammation is believed to be one of the most crucial factors in the pathological changes in TMJ OA.⁴ It has been

elucidated that several cytokines, including interleukin (IL) -1 β , IL-6, IL-8, IL-17, and tumor necrosis factor (TNF) - α , are elevated in TMJ synovial fluid of TMJ OA patients.4-6 Among them, IL-1 β plays a pivotal role in exacerbating the pathological process of TMJ OA by promoting the release of matrix-degrading enzymes, inhibiting the differentiation of mesenchymal stem cells and mediating the crosstalk between several pro-inflammatory molecules and pathways.^{5,7–9} The expression of cyclooxygenase (COX) -2 and prostaglandin (PG) E2 is increased in fibroblast-like synoviocytes (FLS) derived from TMJ stimulated with IL-1B, resulting in the activation of ptgs-2/PGE2 pathway and inhibiting the proteoglycan production.^{10,11} Cell migrationinducible chemokines, including monocyte chemoattractant protein (MCP-1, or CCL2), macrophage inflammatory protein-3a (MIP-3a, or CCL20), stromal cell-derived factor 1 (SDF-1, or CXCL12) and intracellular adhesion molecule 1 (ICAM-1), are highly expressed in human TMJ FLS.^{12,13} IL-1 β can interact with high mobility group box protein 1 (HMGB1), amplifying the production of cytokines and chemokines and aggravating inflammation.¹⁴ Furthermore, Toll-like receptor (TLR)-4 can interact with IL-1 β and activate MyD88, leading to NF- κ B pathway activation.^{15,16} Ge et al. demonstrated that NF- κ B and Wnt5a signaling pathways were involved in the inflammatory responses when TMJ condylar chondrocytes (CC) were stimulated with IL-1 β . Moreover, NF- κ B was required to activate Wnt5a when stimulated by IL-1 β .

Epigenetics refers to changes in gene expression that are stable between cell divisions or sometimes generations but do not involve changes in the DNA sequences. The contents of epigenetics include DNA methylation, acetylation, and deacetylation of histone and non-coding RNAs et al. Epigenetics is associated with many different human diseases and is also involved in TMJ OA.¹⁷ Histone deacetylase (HDAC) is a type of protein that promotes the deacetylation of histones, leading to higher condensation of chromatins and inhibition of gene translation.^{17,18} Liao et al. have demonstrated that HDAC10 upregulation can contribute to inflammatory activation of TMJ synovium-derived mesenchymal stem cells (SMSCs) mediated by IL-1 β , intervening in the cartilage repair of TMJ OA.¹⁷ BRD4 is a double bromodomain protein of the BET family binding to the acetylated histone to regulate gene expression. Inhibition of BRD4 can attenuate TREM1-mediated inflammatory responses in TMJ OA.¹⁸ Non-coding RNAs (ncRNAs) are derived from many sequences generating non-protein-coding RNAs in the human genome and have vital functions in translation, RNA splicing, and regulation of gene expression.²⁰ MicroRNAs, circRNAs, and long non-coding RNAs (lncRNAs) are all involved in inflammatory responses of TMJ OA, leading to the release of pro-inflammatory cytokines and matrixdegrading enzymes. LncRNAs, including AK094629 and HOTAIR, can lead to the aggravation of inflammation, while



Fig. 1 Potential pathological changes and the vicious cycle of TMJ OA. The pathological changes of TMJ OA typically include inflammatory response, ECM degeneration, abnormal cell biological behaviors, and aberrant angiogenesis. Inflammatory responses may occur in cartilage, disc, synovium, and all other TMJ tissue, characterized by cytokine release, cell infiltration, M1 macrophage polarization, and excessive oxidative stress. ECM degeneration is mainly caused by overexpression of MMPs and ADAMTS, leading to the degradation of collagens and aggrecan. Various cells in TMJ tissue may display excessive apoptosis, suppressed autophagy, and abnormal differentiation and activation. Neovascularization also occurs and primarily develops into osteophytes. One pathological change can be the causation or the result of other pathological changes, forming a vicious cycle in TMJ OA pathogenesis. Thus, it may prolong the disease process making it hard to cure.

AK094629 can interact with the MAPK pathway.^{20,21} Increased expression of miR-1246 in extracellular vesicles (EVs) derived from M1 macrophages (M1-EVs) has a proinflammatory effect on TMJ OA via activation of the Wnt/ β -catenin pathway.²² Hsa_circ_0000448 is also involved in the progression of TMJ OA by promoting the upregulation of TNF- α through the ceRNA mechanism.²³

Other molecules and pathways also participate in the inflammatory responses of TMJ OA. For example, connexin 43 (Cx43) is localized at the cell surface and is capable of forming hemichannels, which plays a role in mechanotransduction and is widely expressed in chondrocytes and osteocytes. It is reported that fluid flow shear stress (FFSS) on ATDC5 cells leads to increased expression of Cx43, followed by enhanced synthesis and release of PGE2.²⁴ Recently, leptin, a non-glycosylated polypeptide hormone synthesized predominantly by white adipose tissue, has been increasingly proven to be a cytokine-like hormone functioning as an immune response modulator. Xiong et al. showed that leptin could upregulate IL-6 production via JAK/STAT3, p38 MAPK, and PI3K/Akt (protein kinase B) pathways when binding to Ob-Rb in TMJ FLS.²⁵ In addition, considering that the incidence of TMJ OA in females is higher than in males, estrogen is believed

to be involved in TMJ OA. It has been demonstrated that estrogen could interact with cadherin-11, a cell adhesion molecule, to promote M1-like macrophage activation and aggravate the acute inflammation in TMJ tissue.²⁶ Estradiol can also activate the NF- κ B pathway, causing the induction of upregulating pro-inflammatory cytokines such as IL-1 β , IL-6, and COX2.²⁷ Moreover, SDF-1, Dkk-1, and p38 MAPK are also involved in generating and processing TMJ OA.^{28–30}

Several molecules suppress inflammatory responses in TMJ OA through different molecular pathways. In complete Freund's adjuvant-induced (CFA) and disc-perforation-induced TMJ OA model, IL-37 was able to shift M1 macro-phage into the M2 phenotype and inhibited the M1-polarized phenotype through the NLRP3 (NOD-like receptor pyrin domain-containing 3) signaling pathway.^{31,32} The interaction between the tumor necrosis factor receptor OX40 and its ligand OX40L (TNFSF4) can lead to the stimulation of T-cells. After IL-1 β and TNF- α established TMJ CC inflammatory model, miR-145-5p was upregulated, suppressing the progression of TMJ OA by targeting ox40l to regulate T-cell-mediated immune response.³³ In contrast to estrogen, progesterone (P4) displays a protective effect on TMJ OA. P4 replacement attenuates TMJ OA and diminishes the DNA-

binding effect of NF- κ B in a dose-dependent manner in ovariectomized TMJ OA rats. The in-vitro experiment also demonstrated that P4 could partially reverse the transcription of cytokines in TMJ synoviocytes induced by IL-1 β and TNF- α .³⁴ Bmal1, a circadian gene, downregulates IL-6 levels by decreasing extracellular signal-regulated kinase (ERK) phosphorylation in TMJ OA cartilage.³⁵ Other molecules, including 15-deoxy- $\Delta^{12,14}$ -prostaglandin J2 and p21, also play a protective role in TMJ OA by inhibiting the expression and release of pro-inflammatory cytokines IL-1 β , IL-6, IL-12, IL-18, chemokines, including cytokine-induced neutrophil chemoattractant 1 (CINC-1) and matrix-degrading enzymes MMP9, MMP13.^{36,37}

Detailed information about inflammatory responses and corresponding molecular mechanisms in TMJ OA is displayed in Table 1.

Extracellular matrix degradation

ECM, produced by proximal cells and tissues, contains a wide variety of molecules forming a well-organized network, which is the main component of the microenvironment and acts as extrinsic regulators for various physiological and pathological processes.³⁸ The ECM of TMJ cartilage is mainly composed of collagen type I, collagen type II, and proteoglycans. It protects against elastic and shear forces and regulates chondrocytes' metabolism and biological behaviors.^{3,39} ECM degeneration is closely related to inflammatory response, excessive mechanical loading, aging, estrogen level, and osteoclastogenesis in osteoarthritis.^{2,39,40} It starts from the expression of matrix metalloproteinases (MMPs) and a disintegrin and metalloproteinase with thrombospondin motifs (ADAMTS) in the cells.³ MMP and ADAMTS belong to the subfamily of zincdependent proteinases, which play a significant role in repairing and remodeling ECM in both physiological and pathological conditions.^{38,40,41} IL-1 β -induced overexpression of MMP13, ADAMTS4, and ADAMTS5 could lead to cartilage damage in IL-1RA-/- mice.39 In the TMJ OA mouse model, the expression of IL-6 and MMP12 was increased. Overexpression of IL-6 could also lead to the increased expression of ADAMTS4 and ADAMTS5, forming a positive feedback loop and resulting in chronic cartilage degeneration in TMJ.² Another study found that MMP3, one of the predominant proteinases in degrading ECM, was closely linked to heavy masticatory loading, while MMP8 might be protective and related to estrogen levels and aging.⁴⁰ Furthermore, ERK signaling may have a crucial role in controlling the expression of MMPs.⁴

Several intra- or extracellular molecules and epigenetics regulate the ECM degeneration process. In unilateral anterior crossbite (UAC) TMJ OA model, Regulated upon Activation, Normal T Cell Expressed and Presumably Secreted (RANTES) and Stromal cell-derived factor 1 (SDF-1) was significantly upregulated. RANTES and SDF-1 are linked with C–C chemokine receptor type 1 (CCR1) and C-X-C chemokine receptor type 4 (CXCR4), respectively.⁴³ SDF-1 combined with CXCR4 in chondrocytes shifts TGF- β receptor type I (T β RI) to activin receptor-like kinase 1 (ALK1) from activin receptor-like kinase 5 (ALK5).⁴⁴ Both molecules can promote cartilage degeneration and subchondral bone resorption, suggesting a potential link between

Main Factors	Pathways	Effect on Inflammation	References
CRP	-	Aggravation	4
IL-17a	NF-κB, PI3K/ Akt	Aggravation	6
IL-1β	Pin1/NF-κB Wnt5a	Aggravation	7-9,13
COX2	_	Aggravation	10
PGE2	_	Aggravation	11
MIP-3	MAPK, NF-κB	Aggravation	12
HMGB1	_	Aggravation	14
TLR4/MyD88	NF-кВ р38 МАРК	Aggravation	15,16
HDAC(10)	NF-κB	Aggravation	17,18
BRD4	_	Aggravation	19
LncRNA HOTAIR	_	Aggravation	20
LncRNA AK094629	MAPK	Aggravation	21
miR-1246	Wnt/β- catenin	Aggravation	22
hsa_circ_0000448	_	Aggravation	23
Connexin 43	_	Aggravation	24
Leptin	JAK/STAT3, PI3K/Akt, p38 MAPK	Aggravation	25
Estrogen	NF-κB	Aggravation	26,27
SDF-1	-	Aggravation	28
Dkk-1	Wnt/β- catenin NF-κB	Aggravation	29
p38 MAPK	p38 MAPK	Aggravation	30
IL-37	ERK, JNK, NF- κB, p38	Inhibition	31,32
miR-146-5p	_	Inhibition	33
Progesterone (P4)	NF-κB	Inhibition	34
Bmal1	MAPK/ERK	Inhibition	35
15d-PGJ2	_	Inhibition	36
p21	-	Inhibition	37

Abbreviations: CRP: C-reaction protein; IL: interleukin; NF-kB: nuclear factor-kappa B; PI3K: phosphoinositide-3 kinase; Akt: protein kinase B; iNOS: inducible nitric oxide synthase; MMP: matrix metalloproteinase; Pin1: peptidyl-prolyl *cis*-trans isomerase NIMA-interacting 1; HMGB: high mobility group box; TLR: toll-like receptor; SDF-1: stromal cell-derived factor 1; MAPK: mitogen-activated protein kinases; JAK: Janus kinase; STAT3: signal transducer and activator of transcription 3; ERK: Extracellular Signal-Regulated Kinase; JNK: *c*-Jun N-terminal kinase.

inflammatory responses and ECM degeneration. Periostin, a matricellular protein belonging to the fasciclin family, is upregulated by excessive mechanical loading. The increased expression of periostin induces the expression of ADAMTS5 via the NF- κ B pathway, leading to ECM degeneration and condylar resorption.⁴⁵ Osteopontin (OPN), a matricellular protein regulating tissue repair and remodeling, is upregulated in the TMJ OA rat model and activates the NF- κ B signaling pathway to stimulate the expression of MMPs.⁴⁶ HMGB2 participates in TMJ OA and is necessary for TMJ cartilage in sensing pressure loading. Increased level of

HMGB2 is associated with cartilage degeneration induced by excessive mechanical load.⁴⁷ In addition, miR-21-5p also plays a vital role in matrix degradation by targeting growth differentiation factor 5 (Gdf5). Knock-out (KO) of miR-21-5p or Gdf5 stimulation significantly decreases the expression of MMP13 and reverses the degeneration of condylar cartilage.⁴⁸

Estrogen has also been suggested to have a crucial role in ECM degeneration in TMJ OA. The pathogenic effect of estrogen on the degeneration of ECM primarily depends on estrogen receptors (ER) α and β , which are widely expressed in females, especially in TMJ fibrocartilage. When 17 β -estradiol activates its receptor Er α , MMP9 is upregulated via the ERK signaling pathway, and NF- κ B and ELK-1, direct targets of ERK, contribute to estrogen/Er α / ERK-induced MMP9 upregulation.⁴⁹ Another study has revealed that in combination with Er β , estrogen could increase the expression of HIF (hypoxia-inducible factor) -2 α and down-streaming cytokines, MMP13 and COLX, leading to aggravation of ECM degeneration.⁵⁰

The Notch signaling pathway comprises a family of four type-1 transmembrane receptors (Notch 1–4). In the canonical Notch signaling pathway, Notch binds to Notch ligands and activates the release and translocation of the Notch1 intracellular domain (NICD) to the nucleus.⁵¹ The Notch cascade is involved in regulating the behaviors of chondrocytes.³ In the TMJ OA model, IL-1 β stimulates the overexpression of Notch1 and NICD and increases the expression of IL-6 and TNF- α . In contrast, inhibition of Notch1 could suppress cartilage degeneration by balancing MMP and TIMP (tissue inhibitors of matrix metal-loproteinases) -1 via the NF- κ B signaling pathway.⁵¹

Transforming growth factor β (TGF- β) is a multifunctional cytokine and structurally belongs to the TGF- β superfamily, including four subtypes (TGF- β 1, β 2, β 3, β 1 β 2). Studies have shown that TGF- β is one of the most abundant cytokines in bone matrix and is responsible for regulating the remodeling of bone tissues.⁵² The role of the TGF- β signaling pathway is controversial in TMJ OA. It seems that increased expression of TGF-B1 is unnecessary for chondrocytes with completed differentiation. Overexpression of TGF- β 1 involved *p*-Smad2 signaling and HtrA1 production, resulting in MMP13 overproduction and breakdown of cartilage ECM.⁵³ In contrast, another study found that TGFβ1-induced Smad activation could inhibit the expression of MMPs. The crosstalk between TGF-B/Smad3 and S1P/S1P3 signaling pathways could downregulate the expression of MMP9 and MMP13 and induce chondrocyte migration via Rac1, RhoA, and Cdc42.52

Various molecules and pathways inhibit ECM degeneration in TMJ OA. LOXL2, a subtype of the lysyl oxidase (LOX) protein family, is a copper-dependent amine oxidase that remodels ECM and establishes the cross-link between collagens and elastin. Studies have shown that LOXL2 is upregulated in TMJ OA models and may be a protective factor for promoting ECM anabolism and inhibiting catabolism via NF- κ B, ERK, and Integrin/FAK pathways.^{54,55} Lubricin is a large proteoglycan encoded by gene proteoglycan 4 (Prg4) and is pivotal for lubricating the articular surface and maintaining articular integrity. Deletion of Prg4 led to OA-like degeneration of condylar in condyle.⁵⁶ Similarly, type V collagen is essential in establishing and maintaining the ECM. Moreover, the knock-out (KO) of type V collagen could induce the degeneration and malformation of ECM of condylar cartilage.⁵⁷ Deletion of Axin1 contributes to overexpression of MMP13 and ADAMTS5 via Wnt/ β -catenin and fibroblast growth factor (FGF)/ERK1/2 pathways, thus leading to TMJ cartilage degeneration.⁵⁸ Furthermore, Fgfr3 conditional KO (cKO) cartilage expresses higher Runx2, Mmp13, Col10, and Adamts5 via Indian Hedgehog (Ihh) signaling pathway.⁵⁹ In addition, miR-29b attenuates ECM degeneration by rescuing the decrease of the percentage areas of proteoglycan and type II collagen.⁶⁰ (Table 2).

Abnormal cell biological behaviors

Accumulating evidence has shown that the progression of TMJ OA is closely related to tabnormal cell proliferation, differentiation, death, and cell metabolism in cartilage and subchondral bone. These abnormalities in cell biological behaviors may have deteriorative effect on the hemostasis of the TMJ environment, leading to aggravated

Table 2	Molecules and pathways involved in degeneration
of ECM in	TMJ OA.

Main Factors	Pathways	Effect on ECM Degeneration	References
IL-1β, MMP13	_	Aggravation	39
MMP12	_	Aggravation	2
MMP3/8	_	Aggravation	40
MMP1/3/13	ERK	Aggravation	42
RANTES	_	Aggravation	43
SDF-1	_	Aggravation	43,44
Periostin	NF-κB	Aggravation	45
Osteopontin	NF-κB	Aggravation	46
HMGB2	Wnt/β-catenin	Aggravation	47
miR-21-5p		Aggravation	48
Estrogen	ERK, NF-κB	Aggravation	49,50
Notch1	NF-ĸB, Notch	Aggravation	51
Smad3	TGF-β/Smad3 S1P/S1P3	Inhibition	52
TGF-B1	TGF-B1/Smad2	Aggravation	53
LOXL2	NF-κB, ERK, Integrin/FAK	Inhibition	54,55
Lubricin	_	Inhibition	56
COLV	Wnt/β-catenin	Inhibition	57
Axin1	Wnt/β-catenin FGF/ERK1/2	Inhibition	58
FGFR3	lhh	Inhibition	59
miR-29h	_	Inhibition	60

Abbreviations: IL: interleukin; MMP: matrix metalloproteinase; ADAMTS: a disintegrin and metalloproteinase with thrombospondin motifs; NF- κ B: nuclear factor-kappa B; ERK: Extracellular Signal-Regulated Kinase; RANTES: regulated upon activation, normal T cell expressed and presumably secreted; SDF-1: stromal cell-derived factor 1; T β R1: TGF-beta type I receptor; HMGB: high mobility group box; COL: collagen; S1P: sphingosine-1-phosphate; TGF- β : transforming growth factor beta; FAK: focal adhesion kinase; FGF: fibroblast growth factor; FGFR3: fibroblast growth factor receptor type 3. inflammation, degradation of ECM, and remodeling of subchondral bone in TMJ.

Apoptosis

Apoptosis is an active and well-organized cell death under physiological or pathological conditions. Apoptosis-related genes regulate the process of apoptosis, called "programmed cell death" or type I cell death. It is characterized by decreased cell volume, the loss of intercellular connections, and the attachment and detachment of cells from surrounding cells. Apoptosis can be divided into endogenous and exogenous apoptosis, referring to the mitochondrial-related apoptosis pathway and the apoptosis process induced by death receptors respectively, Various factors can induce apoptosis, including trauma, aging, DNA damage, inflammation, and endoplasmic reticulum stress (ERS).^{61,62} It is considered that the increase of apoptosis is one of the essential factors in the progression of TMJ OA. Apoptosis provides space for neovascularization and is the source of abnormal cartilage mineralization, leading to the destruction of cartilage and subchondral bone of TMJ.³

NcRNAs have a crucial role in regulating apoptosis in TMJ OA. In the lipopolysaccharide (LPS)-induced inflammatory model. IncRNA PVT1 was found to exacerbate the expression of TNF- α by sponging the microRNA, miR-211-3p.⁶³ In the synovial fluid of TMJ OA patients, IncRNA HOTAIR is significantly upregulated, and HOTAIR upregulation contributes IL-1 β induced MMP1, MMP3, and MMP9 expression and chondrocyte apoptosis.²⁰ Analogously, circRNA GCN1L1 could promote synoviocyte proliferation, catabolism of cartilage ECM and chondrocyte apoptosis by sponging miR-330-3p and targeting the TNF- α gene.⁶⁴ In addition, miR-15b impedes chondrocyte proliferation by targeting IGF1 and IGF1R and, in turn, promotes chondrocyte apoptosis by targeting BCL2.65 In Complete Freund's Adjuvant (CFA)induced rat TMJ OA model and IL-1_β-induced in-vitro model, miR-101-3p was detected to be downregulated, while miR-101-3p could enhance cell apoptosis induced by IL-1 β targeting the gene ubiquitin-conjugating enzyme 2D1 (UBE2D1) and Frizzled class receptor 4 (FZD4).66

Calcium is crucial in chondrocyte apoptosis, and calcium influx could be elevated under mechanical loading.³ Excessive calcium loading leads to misfolded protein accumulation in the endoplasmic reticulum lumen, termed ER stress (ERS). The mechanistic target of rapamycin kinase (mTOR) is a highly conserved protein kinase with two different functional complexes, mTORC1, and mTORC2. In a study, UAC and FFSS induced calcium overload in chondrocytes in-vivo and in-vitro, which activates the mTORC1/ EIF2AK3/CASP12/DDIT3 signaling pathway and leads to the enhancement of chondrocyte apoptosis.⁶⁷

The role of the canonical Wnt/ β -catenin pathway in TMJ OA is controversial. It has been demonstrated that excessive compressive force could reduce endogenous β -catenin and result in OA-like lesions.³ In contrast, Hui et al. found that in β -catenin conditional activation mice $(\beta$ -cat(ex3)^{Agc1CreER}), condylar cartilage defect was present, and the expression of proliferating cell nuclear antigen (PCNA) and Ki67 was significantly reduced, with cell proliferation decreased, and cell apoptosis increased in TMJ tissue.⁶⁸

The Ihh signaling pathway is also a vital regulator in chondrocyte apoptosis. In a bite-raising TMJ OA model, Ihh, Smoothened (Smo), and Gli zinc finger transcription factor 1 (Gli1) were found to be activated, and the expression of MMP13 and cysteinyl aspartate specific proteinase-3 (caspase-3) were upregulated, leading to hypertrophy and apoptosis of chondrocytes in TMJ OA.⁶⁹

As mentioned above, p21 and Smad3, which are proven to participate in the inhibition of inflammatory response and degeneration of ECM, respectively, also play a role in inhibiting chondrocyte apoptosis.^{37,52} N6-methyladenosine (m⁶A), a type of internal modification in messenger RNA (mRNA), also participates in apoptosis in TMJ OA. Methyltransferase-like 3 (Mettl3) can catalyze Bcl2 mRNA and inhibit chondrocyte apoptosis induced by TNF- α via m⁶A/Ythdf1/Bcl2 signal axis.⁷⁰ These studies indicate that cell apoptosis may be closely related to the inflammatory response and degeneration of ECM in TMJ (Table 3).

Autophagy

Autophagy is another form of cell death known as programmed non-apoptotic cell death, which is essential for cleaning up senescent cells, promoting wound healing and cell renewal. In autophagy, cells degrade their internal damaged organelles and macromolecules through lysosomes, activate autophagy-related genes, and realize the turnover and circulation of intracellular substances. Classical autophagy includes endogenous and exogenous stimulation (DNA damage, cell senescence, and inflammatory environment et al.), autophagy induction, autophagosome formation, and autophagy lysosome formation. Abnormality in chondrocyte autophagy has been proven to be one of the significant factors and characteristics of OA. Multiple pathways, including PI3K/AKT/mTOR, MEK/ERK, and SIRT1/ AMPK pathways, are involved in the regulation of autophagy in the pathogenesis of OA.⁶¹

Table 3	Molecules and pathways involved in apoptosis in
TMJ OA.	

IMJ OA.			
Main Factors	Pathways	Effect on Apoptosis	References
LncRNA PVT1	_	Aggravation	63
LncRNA HOTAIR	-	Aggravation	20
CircRNA GCN1L1	-	Aggravation	64
miR-15b	-	Aggravation	65
miR-101a-3p	-	Aggravation	66
mTORC1	-	Aggravation	67
β-catenin	Wnt/β-catenin	Aggravation	68
lhh	lhh	Aggravation	69
p21	-	Inhibition	37
Smad3	TGF-β/Smad3 S1P/S1P3	Inhibition	52
Mettl3	m6A/Ythdf1/Bcl2	Inhibition	70

Abbreviations: mTORC: mammalian target of rapamycin complex; EIF2AK3: ukaryotic translation initiation factor 2 alpha kinase 3; TGF- β : transforming growth factor beta; S1P: sphingosine-1-phosphate; mettl3: m6A regulators like methyltransferase-like 3; Ythdf1: YT521-B homology (YTH) domain-containing proteins 1; m6A: N6-methyladenosine.

Currently, there are few studies on autophagy in TMJ OA. Nevertheless, autophagy is considered a vital survival mechanism for chondrocytes in TMJ OA.³ Current studies on autophagy in TMJ OA primarily focus on mTOR. According to the study by Liu et al. the expression of mTOR was upregulated in response to IL-1 β , and the ratio of LC3-II/LC3-I was decreased, leading to lowered autophagosome formation.⁷¹ Similarly, another study showed that in the TMJ OA model induced by estrogen deficiency and high bite force, the expression of Beclin-1 and LC3 were significantly downregulated while mTOR was upregulated and autophagic activity was lowered.⁷² Yang et al. revealed that calcium overload induced by excessive mechanical loading promotes ERS, switching autophagy to apoptosis via the p-ERN1/p-PRKAA1/2 (AMPK)/mTORC1 signaling axis.⁶⁷ Fibroblast growth factor receptor 1 (FGFR1) is a tyrosine kinase receptor belonging to the FGF family. Fgfr1 deficiency promotes autophagy activity and attenuates TMJ OA progression.⁷³ In addition, Mettl3 could inhibit autophagy through m⁶A/Ythdf1/Bcl2 signal axis.⁷⁰ (Table 4).

Other cell biological behaviors

Abnormal biological behaviors of cells in TMJ OA also include alteration of osteoclast and osteoblast activity, abnormal differentiation of stem cells and chondrocytes, enhanced fibrogenesis by FLS and increased sensitivity to the stimulations.

One common pathological change TMJ OA brings is abnormal subchondral bone remodeling, which is also one of the first processes and signs. In the early stage, subchondral bone primarily displays bone loss, while cartilage calcification and stiffness of the osteochondral interface occur in the later stage.³ Subchondral bone remodeling is predominantly characterized by the elevated osteoclast activity and decreased osteoblast activity. Several molecules are related to osteoclast activation. Netrin-1, an axon-guiding protein, is upregulated in the synovial fluid of TMJ OA patients and monoiodoacetic acid (MIA)-induced TMJ OA mice and in combination with its receptor, Unc5b, the osteoclast formation is stimulated.74 Wnt5a/receptor tyrosine kinase-like orphan receptor 2 (Ror2) signaling derived from TMJ bone marrow mesenchymal stem cells (BMSCs) is enhanced by UAC, with increased RANKL and

Table 4Molecules and pathways involved in autophagy inTMJ OA.

Main Factors	Pathways	Effect on Autophagy	References
Mettl3	m6A/Ythdf1/Bcl2	Inhibition	70
mTOR	-	Inhibition	71
mTOR, estrogen	-	Inhibition	72
mTORC1	-	Inhibition	67
FGFR1	-	Inhibition	73

Abbreviations: mettl3: m6A regulators like methyltransferaselike 3; Ythdf1: YT521-B homology (YTH) domain-containing proteins 1; m6A: N6-methyladenosine; mTOR: mammalian target of rapamycin; mTORC: mammalian target of rapamycin complex; FGFR1: fibroblast growth factor receptor type 1. CXCL12 expression and osteoclast precursors migration and differentiation via JNK and Ca²⁺/nuclear factor of activated T-cells (NFAT) pathway.⁷⁵ Overactivation of the TGF- $\beta/T\beta$ RI signaling pathway is detected in TMJ OA cartilage and subchondral bone, leading to a disturbed bone marrow microenvironment and abnormal osteoclast activation.⁷⁶ VEGF could initiate TMJ OA by promoting osteoclast differentiation and subchondral bone resorption.⁷⁷ In the incisor malocclusion model, semaphorin 4D (Sema 4D) and Plexin B1, a pair of osteoblast differentiation suppressors, are increased at week 3, accompanied by the increased osteoclast activity, leading to bone remodeling in TMJ.⁷⁸

In contrast, inhibition of abnormal cell biological behaviors can attenuate the severity of TMJ OA and promote cartilage regeneration and bone repair. Two microRNAs, miR-26b and miR-29b play a role in osteoblast activation and osteoclast inhibition, respectively, suppressing abnormal subchondral bone remodeling. 60,79 FGF1 and epidermal growth factor (EGF) activate the MAPK/ERK signaling pathway to suppress the expression of myofibroblast markers, α -smooth muscle actin (α -SMA), and type I collagen, and they can cooperate with inflammatory cytokines, including IL-1 β and TNF- α , to inhibit fibrosis in TMJ OA.⁸⁰ DNA methyltransferase (DNMT) 3b is a member of DNMTs controlling DNA methylation. Dnmt3b gain-offunction in TMJ stem and progenitor cells express more type II collagen and less type X collagen and show enhanced premature differentiation via β-catenin activation, thus alleviating cartilage degradation and progression in TMJ OA.⁸¹

The inhibition of abnormal chondrogenesis and chondrocyte differentiation appears pivotal in promoting correct bone formation and endochondral ossification. Calcium-sensing receptor (CaSR), a member of the class C G protein-coupled receptor (GPCP-C), is widely expressed in maturing and hypertrophic chondrocytes in growth plates and is an essential promoter of chondrocyte terminal differentiation. UAC and FFSS-induced Ca2+ overload and CaSR localization in ER accelerate chondrocyte terminal differentiation and exacerbated matrix degradation in TMJ cartilage.⁸² In contrast, parathyroid hormone-related peptide receptor 1 (PTH1R) activation rescues UAC and FFSSpromoted chondrocyte terminal differentiation and OAlike lesions by inhibiting the lhh signaling pathway.⁸³ Moreover, type VI collagen is one of the primary components in TMJ cartilage. In Col6a2 KO cartilage, reduced bone volume and abnormal chondrocyte differentiation are detected, and type VI collagen possibly regulates endochondral ossification and chondrocyte differentiation via pSmad1/5/8 signaling.⁸⁴ Similarly, in Prg4-null TMJ, ectopic cartilage develops, and ectopic mineralization occurs, leading to irreversible OA-like changes in TMJ, suggesting that Prg4 is crucial for maintaining TMJ integrity and functions.⁸⁵ (Table 5).

Angiogenesis

In addition to inflammatory responses, degeneration of ECM, and abnormal biological behaviors, angiogenesis has been considered to be a crucial pathological change in TMJ OA. In mature, healthy articular cartilage, cells typically

Table 5	Molecules and	pathways involved	in other biological	behaviors in TMJ OA.
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Main Factors	Pathways	Effects	References
Netrin-1	_	Osteoclast activation	74
Wnt5a/Ror2	JNK, Ca ²⁺ /NFAT	Osteoclast activation	75
ΤGF-β/ΤβRΙ	TGF β /Smad2,3	Osteoclast activation	76
VEGF	-	Osteoclast activation	77
Sema 4D, Plexin B1	-	Osteoclast activation Osteoblast inhibition	78
miR-29b	Wnt5a	Osteoclast inhibition	60
miR-26b	Wnt/β-catenin	Osteoblast activation	79
FGF1, EGF + IL-1 β , TNF- α	MAPK/ERK	FLS activity inhibition	80
Dnmt3b	Wnt/β-catenin	Stem/progenitor cells differentiation regulation	81
CaSR	_	Chondrocyte terminal differentiation acceleration	82
PTH1R	lhh	Chondrocyte terminal differentiation inhibition	83
COL VI	-	Chondrocyte differentiation regulation	84
Lubricin (Prg4)	Ihh, BMP	Ectopic chondrogenesis inhibition	85

Abbreviations: Ror2: eceptor tyrosine kinase-like orphan receptor 2; JNK: *c*-Jun N-terminal kinase; NFAT: Nuclear factor of activated T cells; TGF- β : transforming growth factor beta; T β R1: TGF-beta type I receptor; COL: collagen; VEGF: vascular endothelial growth factor; MMP: matrix metalloproteinase; VEGF: vascular endothelial growth factor; FGF: fibroblast growth factor; EGF: epidermal growth factor; Dnmt3b: DNA methylation 3b; CaSR: calcium-sensing receptor; PTH1R: parathyroid hormone type 1 receptor.

produce angiogenic inhibitors, while patients with TMJ OA exhibit higher-density vascularization, which has been detected in histopathological studies.^{86,87} Vascularization in OA has been observed to promote the formation of osteophytes. After osteophyte is formed, vascularization sustains, leading to the formation of osteophyte and destruction of articular cartilage.^{3,87} Neovascularization is accompanied by sympathetic and sensory nerve formation, which may contribute to pain in TMJ OA.⁸⁷ In addition, angiogenesis in OA promotes the transportation of inflammatory factors and degradation and premature mineralization of ECM in cartilage.³

VEGF, a vascular permeability factor and selective endothelial mitogen, has a crucial role in physiological and pathological process of angiogenesis and accumulates in the chondrocytes of mandibular condyles with OA.88,89 Several pathways are involved in the upregulation of VEGF. One of the most studied signaling pathways is the hypoxia-inducible factor (HIF)- 1α /VEGF/Notch pathway. In a study using chronic sleep deprivation (CSD)-induced TMJ OA model, HIF-1a/VEGF/Notch pathway was activated bidirectionally by hypoxia, mediating angiogenesis in TMJ OA.⁹⁰ Dickkopf-related protein 1 (DKK-1), a secreted protein inhibiting Wnt/ β -catenin signaling, is upregulated in concordance with VEGF and could promote nuclear translocation of HIF-1a.⁸⁶ HMGB1 directs the nuclear localization of HIF-1 α and promotes the expression of VEGF via JNK and ERK signaling pathways.⁹¹

ERK signaling pathway is another essential pathway mediating angiogenesis in TMJ OA. The inflammatory cytokine IL-6 could activate estrogen-related receptor γ (ERR γ) via ERK1/2 signaling to promote the synthesis of VEGF, leading to the degradation of ECM and angiogenesis.⁸⁷ The crosstalk between ERK and Notch pathway may be crucial in regulating neovascularization in TMJ OA. As mentioned above, HMGB1 could activate HIF-1 α /VEGF via the ERK pathway.⁹¹ Delta-like ligand 4 (Dll4), an essential component of the Notch pathway functioning mainly in stem cell self-renewal and vascularization, was upregulated in the CSD TMJ OA rat model. It was demonstrated that when VEGF binds to its receptor VEGFR2, ERK1/2 is activated, increasing Dll4 expression.⁸⁹ In addition, IL-1 β could promote VEGF synthesis through the NF- κ B signaling pathway,⁹² while estrogen promotes VEGF synthesis via ER β /HIF-2 α pathway.⁵⁰

In addition, another vascularization regulator, angioprotein (Ang), is also involved in bone remodeling of TMJ. Ang-1, one subtype of Ang, is overexpressed by the activation of IL-1 β via p38 MAPK signaling. The expression of Ang-1 and Ang-2 was increased in a rabbit model treated with biochemical stimulation by experimental forward mandibular positioning (FMP), leading to angiogenesis and remodeling of the mandibular condylar.⁹³

The crosstalk in angiogenesis and corresponding molecular mechanisms of TMJ OA are exhibited in Fig. 2.

Discussion and future directions

The etiology and pathophysiology of TMJ OA are unclear, and there is still no consensus about the precise pathological changes, biomarkers, and pathways in TMJ OA. Nevertheless, emerging studies have suggested that inflammation, ECM degeneration, abnormal biological behaviors, and angiogenesis participate in TMJ OA. Various signaling pathways are involved in the pathogenesis of TMJ OA. Among them, the NF- κ B signaling pathway is essential in mediating inflammatory response and degradation of ECM, which has been demonstrated in many studies. The Wnt/\beta-catenin pathway is a "double-edged sword" under different conditions in TMJ OA. The overexpression of β catenin leads to enhanced chondrocyte apoptosis and OAlike defect, while in a murine model with loss-of-function of $\beta\mbox{-}catenin,$ reduced cell proliferation and impaired cartilage occurred.^{57,68} MAPK and ERK signaling are involved in inflammation and cartilage degeneration and play a critical role in angiogenesis. The Ihh signaling mainly participates in cell behaviors, including apoptosis, ectopic chondrogenesis, chondrocyte terminal differentiation, and crosstalk with PTH1R and BMP signaling pathways. Notch signaling is a pivotal pathway in neovascularization and ECM



Fig. 2 Molecules and pathways Involved in angiogenesis in TMJ OA. VEGF, Ang-1, and Ang-2 are the main pro-angiogenic factors in the pathogenesis of TMJ OA. Among them, VEGF plays a pivotal role in neovascularization. Several signaling pathways, including IL- $1\beta/NF-\kappa B$, IL- $6/ERK1/2/ERR\gamma$, HIF- 1α , Notch1, E2/ER $\beta/HIF-2\alpha$ are involved in this process. Excessive neovascularization leads to pain in the TMJ region, osteophyte formation, and more severe inflammatory responses in TMJ.

degeneration in TMJ OA. Other pathways, including TGF- β / Smad, JNK, and JAK/STAT3 et al. also have a role in TMJ OA development. Notably, estrogen may be one of the primary reasons why the incidence of TMJ OA in females is higher than in males. A higher estrogen level is closely related to inflammatory cytokine release, M1 macrophage polarization, degeneration of ECM, and decreased cell autophagy via NF- κ B, ERK, and mTOR signaling.^{26,49,72} However, a previous study showed that a lack of estrogen reduces subchondral bone volume.⁵⁰ Therefore, considering the complex crosstalk between various pathological changes, molecules, and signaling pathways in TMJ OA, further studies should notice the complicated networks in TMJ OA pathogenesis.

Current in vivo animal models of TMJ OA are established in various methods, including modifying mechanical loading (UAC, bite-raising, prolonged mouth-opening), injection of inflammatory mediators, including MIA, CFA, and IL-1B, surgeries, and gene editing.³ Different animals, such as mice, SD/Wistar rats, New Zealand white rabbits, and pigs. are used to establish animal models of TMJ OA.^{3,81,83} For invitro cell models of TMJ OA, FFSS and injection of IL-1 β or TNF- α are commonly used.³ Different cell types are chosen to establish cell TMJ OA models, such as ATDC5 cells and rat, rabbit, or pig chondrocytes. 35,55,57 Despite various selections in TMJ OA models, it is still difficult to reproduce or intimate the complicated clinical cases of TMJ OA. Moreover, obtaining human fibrocartilage of TMJ seems problematic due to technical and ethical reasons. Further studies must focus on improving the methods of establishing TMJ OA in-vivo and in-vitro models and applying newly developed techniques such as organoid and 3D printing.⁹

Studies have shown that targeting the molecules and pathways mentioned above might be potential novel treatments for TMJ OA (Table 6). Recent studies have demonstrated that extracellular vesicles (EVs) or exosomes derived from stem cells are vital structures in cell therapy⁹⁵ which contain nucleotides and signaling proteins to modulate cell biological behaviors. Wang et al. found that EVs derived from BMSCs could promote cartilage reconstruction via HIPPO and autotaxin/Yes-associated protein (YAP) signaling axis in TMJ OA.⁹⁶ Two studies focusing on miRNAs in EVs and exosomes showed that miR-100-5p and miR-27b-3p displayed anti-inflammatory effects on TMJ OA. MiR-100-5p in exosomes inhibits inflammatory cytokine release and ECM degradation by targeting mTOR,⁹⁵ while miR-27b-3p could promote M2 macrophage differentiation and inhibit M1 polarization by targeting colony-stimulating factor 1 (CSF-1).⁹⁷ Low-intensity pulsed ultrasound (LIPUS), which uses a low frequency (1-3 MHz) and intensity (less than 100 mW/cm²), could attenuate inflammatory responses and degeneration of ECM by targeting HIF-1 α , HIF-2 α , and Zn²⁺ (ZNT)-9.98,99 Other chemical ingredients, exporter including curcumin, glucocorticoids, and etanercept, have their roles in protecting TMJ cartilage and subchondral bone in TMJ OA by attenuating pain, oxidative stress, inflammatory response, and ECM degradation or by enhancing autophagy and chondrogenesis. Unfortunately, current therapy for TMJ OA hardly sets an eye on condyle repair and reconstruction.^{71,100} Various medical treatments have been proven effective in treating TMJ OA in animal and cell models but few clinical trials were constructed. Therefore, it is essential to conduct further studies focusing on novel medicine, materials, and approaches that can promote the

Therapeutic Procedures	Main Factors Affected	Pathways	Effects	References
Exosome	miR-100-5p	_	Attenuating inflammation and	95
	·		ECM degeneration	
BMSC EVs	ATX, YAP	HIPPO	Inducing cartilage reconstruction	96
Exosome	miR-27b-3p	_	Inhibiting M1 polarization	97
LIPUS	ZNT-9	-	Attenuating ECM degeneration	98
LIPUS	HIF-1a, HIF-2a	_	Attenuating inflammation and	99
			ECM degeneration	
Curcumin	Nrf2, ARE	Nrf2/ARE	Attenuating inflammation and	100
			oxidative stress	
Hyaluronic acid (HA)	NLRP3	-	Attenuating inflammation	101
Yohimbine	lkkβ, p65	NF-κB	Attenuating inflammation	102
Methylprednisolone	TNF-α, CRP	_	Attenuating inflammation	103
Glycyrrhizin	HMGB1	NF-κB/Akt	Attenuating inflammation	104
		RAGE/TLR4		
Genistein	p65	NF-κB	Attenuating inflammation	105
Resveratrol	COX2, p65	NF-κB	Attenuating inflammation and apoptosis	106
Etanercept	HIF-2α, EF5	-	Attenuating pain and hypoxia	107
Rapamycin	IL-1β, mTOR, LC3	Wnt/β-catenin	Enhancing autophagy	71
Rebamipide	RANKL	NF-κΒ, ΜΑΡΚ	Inhibiting oxidative stress and osteoclast	108
Strontium ranelate	β-catenin	Wnt/β-catenin	Promoting chondrogenesis	109

Abbreviations: mTOR: mammalian target of rapamycin; BMSC: bone marrow mesenchymal stem cell; EV: extracellular vesicles; ATX: autotaxin; CSF: colony-stimulating factor; ZNT: zinc transporter; MTF: metal responsive transcription factor; HIF: hypoxia-inducible factor; NF- κ B: nuclear factor-kappa B; HMGB: high mobility group box protein; Akt: protein kinase B; RAGE: receptor for advanced glycation end products.

regeneration of TMJ in humans; specifically, more clinical trials are needed on the agenda.

Besides, it has been widely accepted that the cause of TMJ OA is closely related to the combination of physical and psychological factors. However, the susceptibility of TMJ OA is not similar between different individuals. Though it has been suggested that TMD is not a type of hereditary disease, the role of gene mutation and genetic polymorphism in TMJ OA is still unclear.¹¹⁰ Several studies have revealed that genotype plays a vital role in the risk of developing TMD, including TNFA-308 (rs1800629) polymorphism, catechol-O-methyltransferase (COMT) (rs9332377) polymorphism, and an 86-bp variable number tandem repeat (VNTR) in IL-1Ra gene.^{111–113} Studies on genetic factors of TMJ OA can undoubtedly contribute to predicting susceptibility, making a precise diagnosis, and tailoring therapeutic procedures for patients.¹¹⁴

Conclusion

TMJ OA is a complicated disease exhibiting various pathological changes, including inflammatory response, degeneration of ECM, abnormal cell biological behaviors, and aberrant angiogenesis in TMJ tissue. One pathological change is closely linked to others, thus forming a vicious circle in TMJ OA. Many molecules and signaling pathways are involved in the pathogenesis of different sections in TMJ OA and may serve as potential targets to help find effective therapeutic procedures for TMJ OA. Though invivo and in-vitro models of TMJ OA have been successfully established in various ways, it is urgent to develop models closer to human TMJ OA. New medicine, materials, and approaches for treating TMJ OA and the role of genetic factors in TMJ OA still need further investigation.

Declaration of competing interest

The authors have no conflicts of interest relevant to this article.

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References

- 1. Cui C, Zheng L, Fan Y, et al. Parathyroid hormone ameliorates temporomandibular joint osteoarthritic-like changes related to age. *Cell Prolif* 2020;53:e12755.
- 2. Yamashita-Futani Y, Jokaji R, Ooi K, et al. Metalloelastase-12 is involved in the temporomandibular joint inflammatory response as well as cartilage degradation by aggrecanases in STR/Ort mice. *Biomed Rep* 2021;14:51.
- Li B, Guan G, Mei L, et al. Pathological mechanism of chondrocytes and the surrounding environment during osteoarthritis of temporomandibular joint. J Cell Mol Med 2021;25: 4902–11.
- 4. He Y, Zhou M, Jian Z, et al. C-reactive protein knockout attenuates temporomandibular joint inflammation in rats. *J Immunol Res* 2022;2022:8613986.
- Shrivastava M, Battaglino R, Ye L. A comprehensive review on biomarkers associated with painful temporomandibular disorders. *Int J Oral Sci* 2021;13:23.

- 6. Hattori T, Ogura N, Akutsu M, et al. Gene expression profiling of IL-17A-treated synovial fibroblasts from the human temporomandibular joint. *Mediat Inflamm* 2015;2015:436067.
- Liu W, Sun Y, He Y, et al. IL-1β impedes the chondrogenic differentiation of synovial fluid mesenchymal stem cells in the human temporomandibular joint. Int J Mol Med 2017;39: 317-26.
- Yuan F, Xie JL, Liu KY, et al. Xanthan gum protects temporomandibular chondrocytes from IL-1β through Pin1/NF-κB signaling pathway. *Mol Med Rep* 2020;22:1129–36.
- **9.** Ge XP, Gan YH, Zhang CG, et al. Requirement of the NF-κB pathway for induction of Wnt-5A by interleukin-1β in condylar chondrocytes of the temporomandibular joint: functional crosstalk between the Wnt-5A and NF-κB signaling pathways. *Osteoarthritis Cartilage* 2011;19:111–7.
- Kawashima M, Ogura N, Akutsu M, et al. The antiinflammatory effect of cyclooxygenase inhibitors in fibroblast-like synoviocytes from the human temporomandibular joint results from the suppression of PGE2 production. *J Oral Pathol Med* 2013;42:499–506.
- 11. Nazet U, Feulner L, Muschter D, et al. Mechanical stress induce PG-E2 in murine synovial fibroblasts originating from the temporomandibular joint. *Cells* 2021;10:298.
- 12. Akutsu M, Ogura N, Ito K, et al. Effects of interleukin-1β and tumor necrosis factor-α on macrophage inflammatory protein-3α production in synovial fibroblast-like cells from human temporomandibular joints. J Oral Pathol Med 2013;42:491–8.
- **13.** Ibi M, Horie S, Kyakumoto S, et al. Cell-cell interactions between monocytes/macrophages and synoviocyte-like cells promote inflammatory cell infiltration mediated by augmentation of MCP-1 production in temporomandibular joint. *Biosci Rep* 2018;38. BSR20171217.
- 14. Li C, Cai H, Meng Q, et al. IL-1β mediating high mobility group box protein-1 expression in condylar chondrocyte during temporomandibular joint inflammation. J Oral Pathol Med 2016;45:539–45.
- **15.** Liu X, Cai HX, Cao PY, et al. TLR4 contributes to the damage of cartilage and subchondral bone in discectomy-induced TMJOA mice. *J Cell Mol Med* 2020;24:11489–99.
- 16. Lin X, Xie J, Sun S, et al. Toll-like receptor 4 (TLR4) stimulates synovial injury of temporomandibular joint in rats through the activation of p38 mitogen-activated protein kinase (MAPK) signaling pathway. *Med Sci Mon Int Med J Exp Clin Res* 2018; 24:4405–12.
- 17. Liao W, Sun J, Liu W, et al. HDAC10 upregulation contributes to interleukin 1β-mediated inflammatory activation of synovium-derived mesenchymal stem cells in temporomandibular joint. J Cell Physiol 2019;234:12646–62.
- 18. Liao WT, Sun JD, Wang Y, et al. Histone deacetylase inhibitors attenuated interleukin-1β-induced chondrogenesis inhibition in synovium-derived mesenchymal stem cells of the temporomandibular joint. *Bone Joint Res* 2022;11:40–8.
- **19.** Huang Z, Yang R, Zhang L, et al. BRD4 inhibition alleviates mechanical stress-induced TMJ OA-like pathological changes and attenuates TREM1-mediated inflammatory response. *Clin Epigenet* 2021;13:10.
- 20. Zhang C, Wang P, Jiang P, et al. Upregulation of lncRNA HOTAIR contributes to IL-1 β -induced MMP overexpression and chondrocytes apoptosis in temporomandibular joint osteoar-thritis. *Gene* 2016;586:248–53.
- 21. Jia J, Sun J, Liao W, et al. Knockdown of long non-coding RNA AK094629 attenuates the interleukin-1β induced expression of interleukin-6 in synovium-derived mesenchymal stem cells from the temporomandibular joint. *Mol Med Rep* 2020;22: 1195–204.
- 22. Peng S, Yan Y, Li R, et al. Extracellular vesicles from M1polarized macrophages promote inflammation in the

temporomandibular joint via miR-1246 activation of the Wnt/ β -catenin pathway. Ann N Y Acad Sci 2021;1503:48–59.

- 23. Hu Y, Zhu H, Bu L, et al. Expression profile of circular RNA s in TMJ osteoarthritis synovial tissues and potential functions of hsa_circ_0000448 with specific back-spliced junction. *Am J Transl Res* 2019;11:5357–74.
- 24. Zhang J, Zhang HY, Zhang M, et al. Connexin43 hemichannels mediate small molecule exchange between chondrocytes and matrix in biomechanically-stimulated temporomandibular joint cartilage. *Osteoarthritis Cartilage* 2014;22:822–30.
- Xiong H, Li W, Li J, et al. Elevated leptin levels in temporomandibular joint osteoarthritis promote proinflammatory cytokine IL-6 expression in synovial fibroblasts. J Oral Pathol Med 2019;48:251–9.
- **26.** Kou XX, Li CS, He DQ, et al. Estradiol promotes M1-like macrophage activation through cadherin-11 to aggravate temporomandibular joint inflammation in rats. *J Immunol* 2015;194:2810–8.
- Xue XT, Zhang T, Cui SJ, et al. Sexual dimorphism of estrogensensitized synoviocytes contributes to gender difference in temporomandibular joint osteoarthritis. *Oral Dis* 2018;24: 1503–13.
- **28.** Qi D, Sun S, Han L, et al. Stromal cell-derived factor-1 regulates the secretion of interleukin-1β in the temporomandibular joint of rats with synovial inflammation. *J Oral Pathol Med* 2020;49:933–9.
- **29.** de Sousa LM, Dos Santos Alves JM, da Silva Martins C, et al. Immunoexpression of canonical Wnt and NF-κB signaling pathways in the temporomandibular joint of arthritic rats. *Inflamm Res* 2019;68:889–900.
- Jiang L, Lin X, Ji P. Effect of p38 mitogen activated protein kinase inhibitor on temporomandibular joint synovitis induced by occlusal alteration. *J Oral Maxillofac Surg* 2016;74: 1131-9.
- **31.** Luo P, Peng S, Yan Y, et al. IL-37 inhibits M1-like macrophage activation to ameliorate temporomandibular joint inflammation through the NLRP3 pathway. *Rheumatology* 2020;59: 3070–80.
- **32.** Luo P, Feng C, Jiang C, et al. IL-37b alleviates inflammation in the temporomandibular joint cartilage via IL-1R8 pathway. *Cell Prolif* 2019;52:e12692.
- **33.** Yu D, Wei W, Hefeng Y, et al. Upregulated ox40l can be inhibited by miR-146a-5p in condylar chondrocytes induced by IL-1 β and TNF- α : a possible regulatory mechanism in osteo-arthritis. *Int Arch Allergy Immunol* 2021;182:408–16.
- 34. Xue XT, Kou XX, Li CS, et al. Progesterone attenuates temporomandibular joint inflammation through inhibition of NF-κB pathway in ovariectomized rats. *Sci Rep* 2017;7:15334.
- **35.** Chen G, Zhao H, Ma S, et al. Circadian rhythm protein Bmal1 modulates cartilage gene expression in temporomandibular joint osteoarthritis via the MAPK/ERK pathway. *Front Pharmacol* 2020;11:527744.
- 36. Silva Quinteiro M, Henrique Napimoga M, Gomes Macedo C, et al. 15-deoxy-Δ12,14-prostaglandin J2 reduces albumininduced arthritis in temporomandibular joint of rats. Eur J Pharmacol 2014;740:58-65.
- Khurel-Ochir T, Izawa T, Iwasa A, et al. The immunoregulatory role of p21 in the development of the temporomandibular joint-osteoarthritis. *Clin Exp Dent Res* 2021;7:313–22.
- **38.** Simões G, Pereira T, Caseiro A. Matrix metaloproteinases in vascular pathology. *Microvasc Res* 2022;143:104398.
- 39. Tabeian H, Betti BF, Dos Santos Cirqueira C, et al. IL-1β damages fibrocartilage and upregulates MMP-13 expression in fibrochondrocytes in the condyle of the temporomandibular joint. Int J Mol Sci 2019;20:2260.
- 40. Yu J, Mursu E, Typpö M, et al. MMP-3 and MMP-8 in rat mandibular condylar cartilage associated with dietary

loading, estrogen level, and aging. *Arch Oral Biol* 2019;97: 238-44.

- Łukaszewicz-Zając M, Dulewicz M, Mroczko B. A disintegrin and metalloproteinase (ADAM) family: their significance in malignant tumors of the central nervous system (CNS). Int J Mol Sci 2021;22:10378.
- **42.** Ma C, Wu G, Wang Z, et al. Effects of chronic sleep deprivation on the extracellular signal-regulated kinase pathway in the temporomandibular joint of rats. *PLoS One* 2014;9: e107544.
- **43.** Lu L, Zhang X, Zhang M, et al. RANTES and SDF-1 are keys in cell-based therapy of TMJ osteoarthritis. *J Dent Res* 2015;94: 1601–9.
- 44. Qin HJ, Xu T, Wu HT, et al. SDF-1/CXCR4 axis coordinates crosstalk between subchondral bone and articular cartilage in osteoarthritis pathogenesis. *Bone* 2019;125:140–50.
- **45.** Fan B, Liu X, Chen X, et al. Periostin mediates condylar resorption via the NF-κB-ADAMTS5 pathway. *Inflammation* 2020;43:455–65.
- **46.** Ding F, Wang J, Zhu G, et al. Osteopontin stimulates matrix metalloproteinase expression through the nuclear factor- κ B signaling pathway in rat temporomandibular joint and condylar chondrocytes. *Am J Transl Res* 2017;9:316–29.
- **47.** Zhou Y, Lu H, Deng L, et al. HMGB2 is associated with pressure loading in chondrocytes of temporomandibular joint: in vitro and in vivo study. *Cytokine* 2020;126:154875.
- **48.** Zhang A, Ma S, Yuan L, et al. Knockout of miR-21-5p alleviates cartilage matrix degradation by targeting Gdf5 in temporomandibular joint osteoarthritis. *Bone Joint Res* 2020;9: 689–700.
- 49. Ahmad N, Chen S, Wang W, et al. 17β-estradiol induces MMP-9 and MMP-13 in TMJ fibrochondrocytes via estrogen receptor α. J Dent Res 2018;97:1023-30.
- **50.** Ye T, He F, Lu L, et al. The effect of oestrogen on mandibular condylar cartilage via hypoxia-inducible factor- 2α during osteoarthritis development. *Bone* 2020;130:115123.
- 51. Ying W, Yuan F, He P, et al. Inhibition of Notch1 protects against IL-1β-induced inflammation and cartilage destruction in temporomandibular chondrocytes. *Mol Med Rep* 2017;15: 4391–7.
- Mori H, Izawa T, Tanaka E. Smad3 deficiency leads to mandibular condyle degradation via the sphingosine 1phosphate (S1P)/S1P3 signaling axis. Am J Pathol 2015;185: 2742-56.
- Long E, Motwani R, Reece D, et al. The role of TGF-ß1 in osteoarthritis of the temporomandibular joint in two genetic mouse models. *Arch Oral Biol* 2016;67:68–73.
- Alshenibr W, Tashkandi MM, Alsaqer SF, et al. Anabolic role of lysyl oxidase like-2 in cartilage of knee and temporomandibular joints with osteoarthritis. *Arthritis Res Ther* 2017;19: 179.
- 55. Zhang C, Zhu M, Wang H, et al. LOXL2 attenuates osteoarthritis through inactivating Integrin/FAK signaling. *Sci Rep* 2021;11:17020.
- Hill A, Duran J, Purcell P. Lubricin protects the temporomandibular joint surfaces from degeneration. *PLoS One* 2014; 9:e106497.
- 57. Chandrasekaran P, Kwok B, Han B, et al. Type V collagen regulates the structure and biomechanics of TMJ condylar cartilage: a fibrous-hyaline hybrid. *Matrix Biol* 2021;102: 1–19.
- 58. Zhou Y, Shu B, Xie R, et al. Deletion of Axin1 in condylar chondrocytes leads to osteoarthritis-like phenotype in temporomandibular joint via activation of β-catenin and FGF signaling. J Cell Physiol 2019;234:1720-9.
- **59.** Zhou S, Xie Y, Li W, et al. Conditional deletion of fgfr3 in chondrocytes leads to osteoarthritis-like defects in temporomandibular joint of adult mice. *Sci Rep* 2016;6:24039.

- 60. Sun JL, Yan JF, Yu SB, et al. MicroRNA-29b promotes subchondral bone loss in TMJ osteoarthritis. *J Dent Res* 2020;99: 1469–77.
- **61.** Kong H, Wang XQ, Zhang XA. Exercise for Osteoarthritis: a literature review of pathology and mechanism. *Front Aging Neurosci* 2022;14:854026.
- **62.** Yang J, Hu S, Bian Y, et al. Targeting cell death: pyroptosis, ferroptosis, apoptosis and necroptosis in osteoarthritis. *Front Cell Dev Biol* 2022;9:789948.
- **63.** Xu K, Meng Z, Xian XM, et al. LncRNA PVT1 induces chondrocyte apoptosis through upregulation of TNF- α in synoviocytes by sponging miR-211-3p. *Mol Cell Probes* 2020;52: 101560.
- 64. Zhu H, Hu Y, Wang C, et al. CircGCN1L1 promotes synoviocyte proliferation and chondrocyte apoptosis by targeting miR-330-3p and TNF- α in TMJ osteoarthritis. *Cell Death Dis* 2020; 11:284.
- **65.** Cao P, Feng Y, Deng M, et al. MiR-15b is a key regulator of proliferation and apoptosis of chondrocytes from patients with condylar hyperplasia by targeting IGF1, IGF1R and BCL2. *Osteoarthritis Cartilage* 2019;27:336–46.
- 66. Mao D, Wu M, Wei J, et al. MicroRNA-101a-3p could be involved in the pathogenesis of temporomandibular joint osteoarthritis by mediating UBE2D1 and FZD4. J Oral Pathol Med 2021;50:236–43.
- **67.** Yang H, Wen Y, Zhang M, et al. MTORC1 coordinates the autophagy and apoptosis signaling in articular chondrocytes in osteoarthritic temporomandibular joint. *Autophagy* 2020;16: 271–88.
- 68. Hui T, Zhou Y, Wang T, et al. Activation of β-catenin signaling in aggrecan-expressing cells in temporomandibular joint causes osteoarthritis-like defects. Int J Oral Sci 2018;10:13.
- **69.** Long HQ, Tian PF, Guan YX, et al. Expression of Ihh signaling pathway in condylar cartilage after bite-raising in adult rats. *J Mol Histol* 2019;50:459–70.
- **70.** He Y, Wang W, Xu X, et al. Mettl3 inhibits the apoptosis and autophagy of chondrocytes in inflammation through mediating Bcl2 stability via Ythdf1-mediated m6A modification. *Bone* 2022;154:116182.
- 71. Liu W, Luo H, Wang R, et al. Rapamycin-induced autophagy promotes the chondrogenic differentiation of synoviumderived mesenchymal stem cells in the temporomandibular joint in response to IL-1β. *BioMed Res Int* 2020;2020:4035306.
- 72. Zhang J, Zhang S, Qi WJ, et al. Mechanism and potential contributing factors to temporomandibular joint osteoar-thritis. *Oral Dis* 2023;29:1060–9.
- **73.** Wang Z, Huang J, Zhou S, et al. Loss of Fgfr1 in chondrocytes inhibits osteoarthritis by promoting autophagic activity in temporomandibular joint. *J Biol Chem* 2018;293:8761–74.
- 74. Xiao M, Hu Z, Jiang H, et al. The expression of Netrin-1 in the MIA-induced osteoarthritic temporomandibular joint in mice. *Sci Rep* 2021;11:15695.
- **75.** Yang T, Zhang J, Cao Y, et al. Wnt5a/Ror2 mediates temporomandibular joint subchondral bone remodeling. *J Dent Res* 2015;94:803–12.
- **76.** Zheng L, Pi C, Zhang J, et al. Aberrant activation of latent transforming growth factor-β initiates the onset of temporomandibular joint osteoarthritis. *Bone Res* 2018;6:26.
- 77. Shen P, Jiao Z, Zheng JS, et al. Injecting vascular endothelial growth factor into the temporomandibular joint induces osteoarthritis in mice. *Sci Rep* 2015;5:16244.
- Lu L, Huang J, Zhang X, et al. Changes of temporomandibular joint and semaphorin 4D/Plexin-B1 expression in a mouse model of incisor malocclusion. J Oral Facial Pain Headache 2014;28:68–79.
- **79.** Yang J, Xu Y, Xue X, et al. MicroRNA-26b regulates BMSC osteogenic differentiation of TMJ subchondral bone through β-catenin in osteoarthritis. *Bone* 2022;162:116448.

- **80.** Matsumoto S, Yokota S, Chosa N, et al. Receptor tyrosine kinase ligands and inflammatory cytokines cooperatively suppress the fibrogenic activity in temporomandibular-joint-derived fibroblast-like synoviocytes via mitogen-activated protein kinase kinase/extracellular signal-regulated kinase. *Exp Ther Med* 2020;20:1967–74.
- **81.** Zhou Y, Chen M, O'Keefe RJ, et al. Epigenetic and therapeutic implications of dnmt3b in temporomandibular joint osteoar-thritis. *Am J Transl Res* 2019;11:1736–47.
- Zhang M, Yang H, Wan X, et al. Prevention of injury-induced osteoarthritis in rodent temporomandibular joint by targeting chondrocyte CaSR. J Bone Miner Res 2019;34:726–38.
- Yang H, Zhang M, Liu Q, et al. Inhibition of Ihh reverses temporomandibular joint osteoarthritis via a PTH1R signaling dependent mechanism. *Int J Mol Sci* 2019;20:3797.
- **84.** Komori T, Ji Y, Pham H, et al. Type VI collagen regulates endochondral ossification in the temporomandibular joint. *JBMR Plus* 2022;6:e10617.
- **85.** Bechtold TE, Saunders C, Decker RS, et al. Osteophyte formation and matrix mineralization in a TMJ osteoarthritis mouse model are associated with ectopic hedgehog signaling. *Matrix Biol* 2016;52–54:339–54.
- 86. Jiang SJ, Li W, Li YJ, et al. Dickkopf-related protein 1 induces angiogenesis by upregulating vascular endothelial growth factor in the synovial fibroblasts of patients with temporomandibular joint disorders. *Mol Med Rep* 2015;12:4959–66.
- 87. Zhao H, Liu S, Ma C, et al. Estrogen-related receptor γ induces angiogenesis and extracellular matrix degradation of temporomandibular joint osteoarthritis in rats. *Front Pharmacol* 2019;10:1290.
- Ke J, Liu Y, Long X, et al. Up-regulation of vascular endothelial growth factor in synovial fibroblasts from human temporomandibular joint by hypoxia. J Oral Pathol Med 2007; 36:290-6.
- Dong Y, Wu G, Zhu T, et al. VEGF promotes cartilage angiogenesis by phospho-ERK1/2 activation of Dll4 signaling in temporomandibular joint osteoarthritis caused by chronic sleep disturbance in Wistar rats. Oncotarget 2017;8: 17849–61.
- 90. Chen Y, Zhao B, Zhu Y, et al. HIF-1-VEGF-Notch mediates angiogenesis in temporomandibular joint osteoarthritis. *Am J Transl Res* 2019;11:2969–82.
- **91.** Feng Y, Ke J, Cao P, et al. HMGB1-induced angiogenesis in perforated disc cells of human temporomandibular joint. *J Cell Mol Med* 2018;22:1283–91.
- 92. Xu J, Cai H, Meng Q, et al. IL-1β-regulating angiogenic factors expression in perforated temporomandibular disk cells via NFκB pathway. J Oral Pathol Med 2016;45:605–12.
- **93.** Jing Z, Gu Z, Feng J. Forward mandibular positioning enhances the expression of Ang-1 and Ang-2 in rabbit condylar chondrocytes. *Mol Med Rep* 2013;8:1094–8.
- 94. Lu K, Ma F, Yi D, et al. Molecular signaling in temporomandibular joint osteoarthritis. J Orthop Translat 2021;32:21-7.
- **95.** Luo P, Jiang C, Ji P, et al. Exosomes of stem cells from human exfoliated deciduous teeth as an anti-inflammatory agent in temporomandibular joint chondrocytes via miR-100-5p/mTOR. *Stem Cell Res Ther* 2019;10:216.
- **96.** Wang Y, Zhao M, Li W, et al. BMSC-derived small extracellular vesicles induce cartilage reconstruction of temporomandibular joint osteoarthritis via autotaxin-YAP signaling axis. *Front Cell Dev Biol* 2021;9:656153.
- 97. Liu Y, Zhang Z, Wang B, et al. Inflammation-stimulated MSCderived small extracellular vesicle miR-27b-3p regulates macrophages by targeting CSF-1 to promote temporomandibular joint condylar regeneration. *Small* 2022;18:e2107354.

- **98.** He D, Wang J, Li Y, et al. Low-intensity pulsed ultrasound promotes aggrecan expression via ZNT-9 in temporomandibular joint chondrocytes. *Gene* 2021;768:145318.
- **99.** Yang T, Liang C, Chen L, et al. Low-intensity pulsed ultrasound alleviates hypoxia-induced chondrocyte damage in temporomandibular disorders by modulating the hypoxiainducible factor pathway. *Front Pharmacol* 2020;11:689.
- 100. Jiang C, Luo P, Li X, et al. Nrf2/ARE is a key pathway for curcumin-mediated protection of TMJ chondrocytes from oxidative stress and inflammation. *Cell Stress Chaperones* 2020;25:395–406.
- 101. Jia M, Lv Y, Xu Y, et al. A comparative analysis of NLRP3related inflammatory mediators in synovial fluid in temporomandibular joint osteoarthritis and internal derangement. *BMC Muscoskel Disord* 2021;22:229.
- 102. Ou F, Huang Y, Sun J, et al. Yohimbine ameliorates temporomandibular joint chondrocyte inflammation with suppression of NF-kB pathway. *Inflammation* 2021;44:80–90.
- **103.** Fredriksson L, Alstergren P, Kopp S. Tumor necrosis factoralpha in temporomandibular joint synovial fluid predicts treatment effects on pain by intra-articular glucocorticoid treatment. *Mediat Inflamm* 2006;2006:59425.
- **104.** Hu Z, Xiao M, Cai H, et al. Glycyrrhizin regulates rat TMJOA progression by inhibiting the HMGB1-RAGE/TLR4-NF-κB/AKT pathway. *J Cell Mol Med* 2022;26:925–36.
- **105.** Yuan J, Ding W, Wu N, et al. Protective effect of genistein on condylar cartilage through downregulating NF-kB expression in experimentally created osteoarthritis rats. *BioMed Res Int* 2019;2019:2629791.
- **106.** Li W, Hu S, Chen X, et al. The antioxidant resveratrol protects against chondrocyte apoptosis by regulating the COX-2/NF-κB pathway in created temporomandibular osteoarthritis. *Bio-Med Res Int* 2021;2021:9978651.
- 107. Sperry MM, Yu YH, Kartha S, et al. Intra-articular etanercept attenuates pain and hypoxia from TMJ loading in the rat. J Orthop Res 2020;38:1316–26.
- 108. Izawa T, Mori H, Shinohara T, et al. Rebamipide attenuates mandibular condylar degeneration in a murine model of TMJ-OA by mediating a chondroprotective effect and by downregulating RANKL-mediated osteoclastogenesis. *PLoS One* 2016;11:e0154107.
- **109.** Yu H, Liu Y, Yang X, et al. Strontium ranelate promotes chondrogenesis through inhibition of the Wnt/β-catenin pathway. *Stem Cell Res Ther* 2021;12:296.
- 110. Aneiros-Guerrero A, Lendinez AM, Palomares AR, et al. Genetic polymorphisms in folate pathway enzymes, DRD4 and GSTM1 are related to temporomandibular disorder. *BMC Med Genet* 2011;12:75.
- 111. Furquim BD, Flamengui LM, Repeke CE, et al. Influence of TNF-α-308 G/A gene polymorphism on temporomandibular disorder. Am J Orthod Dentofacial Orthop 2016;149:692–8.
- **112.** Tumer MK, Nursal AF, Tekcan A, et al. The IL-1Ra gene variable number tandem repeat variant is associated with susceptibility to temporomandibular disorders in Turkish population. *J Clin Lab Anal* 2018;32:e22255.
- 113. de Souza Tesch R, Ladeira Bonato L, Quinelato V, et al. Evaluation of genetic risk related to catechol-Omethyltransferase (COMT) and β2-adrenergic receptor (ADRB2) activity in different diagnostic subgroups of temporomandibular disorder in Brazilian patients. Int J Oral Maxillofac Surg 2020;49:237–43.
- 114. Sangani D, Suzuki A, VonVille H, et al. Gene mutations associated with temporomandibular joint disorders: a systematic review. *OAli* 2015;2:e1583.