



# Current stem cell treatments for spinal cord injury

R Vawda, J Wilcox, MG Fehlings

## INTRODUCTION

Spinal cord injury (SCI) is a devastating condition associated with significant functional and sensory deficits, emotional, social, and financial burdens, and an increased risk of cardiovascular complications, deep vein thrombosis, osteoporosis, pressure ulcers, autonomic dysreflexia, and neuropathic pain.

The estimated annual global incidence of SCI is 15–40 cases per million. In the USA, approximately 1.275 million individuals are affected, with over 12,000 new cases each year.<sup>1-5</sup> The most common causes of traumatic SCI are road traffic accidents, falls, occupational and sports-related injuries that result in contusion and compression of the spinal cord.<sup>1</sup> Approximately 55% of SCIs occur at the cervical level (C1 to C7-T1) with a mortality of 10% in the first year following injury and an expected lifespan of only 10–15 years post-injury, and thoracic (T1–T11), thoracolumbar (T11–T12 to L1–L2) and lumbosacral (L2–S5) injuries each account for approximately 15% of SCI.<sup>1,4</sup> Depending on the age of the patient, severity, and levels of SCI, the lifetime cost of health care and other injury-related expenses can reach \$25 million.<sup>1-5</sup>

Despite advances in pre-hospital care, medical and surgical management and rehabilitation approaches, many SCI sufferers still experience substantial neurological disability. Intensive efforts are underway to develop effective neuroprotective and regenerative strategies.

Department of University Health Network, Toronto Western Hospital, Toronto, Canada ON M5T 2S8

**Address for correspondence:** Dr. Michael Fehlings, University Health Network, Toronto Western Research Institute, Main Pavilion, 12<sup>th</sup> Floor, 399 Bathurst Street, Toronto, Canada ON M5T 2S8. E-mail: michael.fehlings@uhn.ca

## PATHOPHYSIOLOGY

SCI involves a primary (the physical injury) and a secondary injury (the subsequent cascade of molecular and cellular events which amplify the original injury).<sup>6</sup> The primary injury damages both upper and lower motor neurons and disrupts motor, sensory and autonomic functions. Pathophysiological processes occurring in the secondary injury phase are rapidly instigated in response to the primary injury in an attempt to homeostatically control and minimize the damage. Paradoxically, this response is largely responsible for exacerbating the initial damage and creating an inhibitory milieu that prevents endogenous efforts of repair, regeneration and remyelination. These secondary processes include inflammation, ischemia, lipid peroxidation, production of free radicals, disruption of ion channels, axonal demyelination, glial scarring (astrogliosis), necrosis and programmed cell death. Nevertheless, endogenous repair and regenerative mechanisms during the secondary phase of injury minimize the extent of the lesion (through astrogliosis), reorganize blood supply through angiogenesis, clear cellular debris, and reunite and remodel damaged neural circuits. The spatial and temporal dynamics of these secondary mediators<sup>7</sup> are fundamental to SCI pathophysiology and as such offer exploitable targets for therapeutic intervention.

## CELL THERAPY

A multitude of characteristics of cells tested pre-clinically and clinically make them attractive to potentially address the multifactorial nature of the pathophysiology of secondary SCI – they are anti-inflammatory, immunomodulatory,<sup>8-12</sup> anti-gliotic,<sup>13</sup> pro-oligodendroglial,<sup>14</sup> pro-neuronogenic,<sup>15</sup> and secrete various anti-apoptotic and pro-angiogenic neurotrophic factors. Given the pathophysiological targets of SCI,<sup>7</sup> transplanted cells should: 1) enable regenerating axons to cross barriers; 2) functionally replace lost cells; and/or 3) create an environment supportive of neural repair.<sup>16</sup> However, given the multifactorial nature of SCI and its dynamic pathophysiological consequences, the success of future clinical trials of cell therapy will likely depend on the informed co-administration of multiple strategies, including pharmacological and rehabilitation therapies.<sup>7</sup>

Access this article online	
Quick Response Code:	Website: www.ijonline.com
	DOI: 10.4103/0019-5413.91629

Different sources and types of cells have been and/or are being tested in clinical trials for SCI, including embryonic stem cells (ESCs), neural progenitor cells (NPCs), bone marrow mesenchymal cells (BMSCs) and non-stem cells such as olfactory ensheathing cells and Schwann cells.<sup>17</sup> Other cell types are being developed for the clinic, including other sources of mesenchymal cells (fetal blood,<sup>18</sup> adipose tissue, umbilical cord<sup>19-36</sup>), adult<sup>21,37</sup> and immortalized neural progenitors (PISCES, NCT01151124), skin-derived progenitors,<sup>38-47</sup> induced pluripotent stem cells<sup>48-52</sup> and endogenous spinal cord progenitors<sup>53-58</sup> [Table 1]. The advantages and disadvantages of each cell source and type being considered or already in clinical trials for SCI have been extensively described and compared elsewhere,<sup>17,59-63</sup> and reflect their potential in the clinic [Table 1]. There are currently more than a dozen cell therapy clinical trials for SCI listed on clinicaltrials.gov.<sup>64</sup> Most are Phase I or I/II clinical safety and feasibility studies, indicating that cellular treatments for SCI developed in the laboratory are still in the very early stages of clinical translation.

This editorial will focus specifically on the most widely studied progenitor cells currently in clinical trials for SCI: BMSCs and NPCs.

## BONE MARROW STROMAL CELLS

BMSCs are isolated from the stromal compartment of bone marrow, and fractionated from hematopoietic stem cells by virtue of their adherence to tissue culture plastic and/or their expression of distinct cell surface antigenic markers. They are non-teratogenic, have anti-inflammatory and

immunomodulatory effects<sup>65,66</sup> and secrete neurotrophic factors, making them attractive candidates in CNS cell rescue and as autologous transplanted cellular sources of trophic support for endogenous and co-implanted cells. Despite recurring claims of their neurogenic differentiation potential *in vitro* or *in vivo*, there is no conclusive evidence to support this.<sup>67</sup>

Most studies of BMSCs have found beneficial effects of BMSC administration after thoracic SCI, largely as a result of neurotrophic factor secretion<sup>68,69</sup> and possibly also anti-inflammatory cytokine secretion. Intraspinal as well as intrathecal and systemic (intravenous) routes of delivery have been successful.<sup>70-72</sup> Porcine and non-human primate studies have been carried out to further support their clinical use,<sup>73,74</sup> and as in rodent studies, it has been found that BMSCs promote a certain degree of axonal regrowth and sprouting, at least in transection models,<sup>75</sup> especially when treated with growth factors prior to being implanted.<sup>68</sup>

The inflammatory component of SCI and subsequent demyelination of surviving axons are serious limiting factors in the efficacy of early cell therapy for SCI, as implanted cells are more likely to be eliminated by the host.<sup>76</sup> The immunomodulatory effect of bone marrow-derived cells has been demonstrated in the Canadian Bone Marrow Transplant (BMT) clinical trials for multiple sclerosis led by Freedman.<sup>65,66</sup> It is therefore worth considering interventions to modify the inflammatory milieu in order to enhance donor cell survival and efficacy, as in the newly initiated clinical trial of autologous BMSCs in children suffering from SCI [Table 2] (NCT01328860).

**Table 1: A comparison of the different cell types and sources currently in (\*) or under consideration for clinical trials for SCI**

	WJ/UCM	fMPC	BMSC*	ES*	iPS	fNPC*	aNPC	OEC*	SC*	SKP	Adip MSC
Isolation	Easy	Chal-	Chal-	Challenging	Challenging	Challeng-	Challeng-	Chal-	Chal-	Less	Easy
Practicalities	None	lenging	lenging	Significant	None	ing	ing	lenging	lenging	challenging	Few
Ethical considerations		Signifi-	Consid-			Signifi-	Consider-	Few	Consid-		
Differentiation potential		cant	erable			cant	able		erable		
Bone	✓	✓	✓	Pluripotent	Pluripotent	Neural	Neural	-	-	Peripheral	✓
Fat	✓	✓	✓							myelin	✓
Cartilage	✓	✓	✓								✓
Storage											
Pre-isolation	✓	×	×	×	×	×	×	×	×	×	?
Post-isolation	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
Immunogenicity	Low	Low	Low	Low/?	Low/?	Low	?	Low	Low	Low	Low
Immunosuppressant/anti-inflammatory	✓	×	✓	?	?	✓	?	?	?	?	?
Tumorigenicity	×	×	×	✓	✓	×	×	×	×	?	×
Transfection	✓	✓	✓	✓	✓	✓	✓	✓	✓	?	?
Safety/risk	✓	✓	✓	✓	?	✓	✓	✓	✓	?	✓
Pathotropism	✓	✓	✓	?	?	✓	?	?	?	?	?
Autologous	Potential	No	✓	×	Potential	×	×	✓	✓	✓	✓

WJ – Wharton's jelly; UCM – umbilical cord matrix; BMSC – Bone marrow stromal cells; ES – Embryonic stem; iPS – Induced pluripotent; fNPC and aNPC – Fetal and adult neural precursor cells; OEC – Olfactory ensheathing cells; SC – Schwann cells; SKP – Skin-derived progenitors; AdipMSC – Adipose tissue-derived mesenchymal cells, \*cells in current or past clinical trials for SCI.

**Table 2: Clinical trials of cell therapy for SCI listed on [www.clinicaltrials.gov](http://www.clinicaltrials.gov).**

Status	Study	NCT	Cells	Administration route	Phase	Country	Sponsor/ Investigator	Duration	Numbers enrolled
Recruiting	Transfer of bone marrow derived stem cells for the treatment of SCI	NCT 01162915	Autologous BMSCs, expanded <i>ex vivo</i>	Intrathecal infusion, single dose	I, single center	USA	TCA Cellular Therapy, LLC; Gabriel P. Lasala	July 2010– June 2012	10
Completed	Cell transplant in SCI patients Condition: Chronic SCI Procedure: Physical therapy	NCT 00816803	Autologous bone marrow	?	I/II	Egypt	Cairo University, Cancer Institute of New Jersey, UMDNJ/RWJMS; Hatem E. Sabaawy	May 2005– Dec 2008	80
Recruiting	Transplantation of autologous OECs in complete human SCI Other: Rehabilitation	NCT 01231893	Autologous olfactory mucosa ensheathing cells (OECs) and fibroblasts	Intraspinal	I	Poland	Wroclaw Medical University; Wlodzimierz Jarmundowicz, Pawel Tabakow	May 2008	10
Terminated	Treatment for acute SCI	NCT 00695149	BMSC	Into cerebrospinal fluid	I/II	Japan	Translational Research Informatics Center	July 2005– March 2010	23
Completed	Autologous adipose derived MSCs transplantation in patients with SCI	NCT 01274975	Autologous adipose derived MSCs	Intravenous infusion, $4 \times 10^8$ cells	I	Korea	RNL Bio Company Ltd, SangHan Kim	July 2009– Feb 2010	8
Recruiting	Safety and feasibility of umbilical cord blood cell transplant into injured spinal cord Drug: $\pm$ Methylprednisolone Drug: $\pm$ Lithium	NCT 01046786	Umbilical cord blood mononuclear cell, dose comparison	Intraspinal	I/II	China	China Spinal Cord Injury Network	Jan 2010– June 2012	20
Completed	Safety and efficacy of autologous bone marrow stem cells in treating SCI Condition: Acute, subacute and chronic SCI Procedure: Laminectomy	NCT 01186679	Autologous bone marrow	Intrathecal	I/II	India	International Stemcell Services Limited, Arvind Bhateja	Jan 2008– Aug 2010	12
Enrolling by invitation	Umbilical cord blood mononuclear cell transplant to treat chronic SCI Other: Methylprednisolone, sodium succinate or lithium carbonate plus rehabilitation	NCT 01354483	HLA-matched umbilical cord blood mononuclear cells		I/II	China	Treating Center of Spinal Cord Injury Chengdu Army Kunming General Hospital (Dr Hui Zhu)	Sep 2010– Dec 2012	20
Suspended	Autologous incubated macrophages for patients with complete SCIs condition: Acute SCI	NCT 00073853	Autologous incubated macrophages	Intraspinal	II	USA, Israel	Proneuron Biotechnologies, Marcus Foundation B.I.R.D. (Israel-U.S. Binational Industrial Research and Development); Daniel Lammertse, Nachshon Knoller, Marca Sipski, Edward Benzel	Sep 2003	61

Table 2 contd/-

Table 2 contd/-

Status	Study	NCT	Cells	Administration route	Phase	Country	Sponsor/ Investigator	Duration	Numbers enrolled
Recruiting	Safety study of GRNOPC1 in SCI Condition: Complete T3–T9 level subacute (7–14 days post-injury) SCI	NCT 01217008	GRNOPC1 (ES cell-derived oligodendrocytic progenitors)	Intraspinal, single dose of 2 million cells	I	USA	Geron, Gary K. Steinberg, David Apple, Richard G Fessler, James S Harrop, Shekar Kurpad	Oct 2010– Oct 2012	10
Recruiting	Autologous stem cells for SCI in children Condition: Primary SCI, to minimize secondary SCI	NCT 01328860	Autologous BMSCs	Intravenous infusion	I	USA	Memorial Hermann Healthcare System, James E. Baumgartner	April 2011– Oct 2014	10
Recruiting	Autologous bone marrow stem cell transplantation in patients with spinal cord injury	NCT 01325103	Autologous BMSC	Intraspinal	I	Brazil	Hospital Sao Rafael; Ricardo R. dos Santos	July 2010– Jan 2013	20
Recruiting	Study of human CNS stem cells (HuCNS-SC) in patients with thoracic spinal cord injury Condition: Subacute thoracic (T2–T11) SCI	NCT 01321333	HuCNS-SC cells	Single dose, intramedullary	I/II	Switzerland	StemCells, Inc.; Armin Curt	March 2011–March 2016	12

Ongoing clinical studies and those carried out to date have enrolled small patient numbers and have used autologous marrow-derived cells rather than purified stromal cells.<sup>77-80</sup> A recently published dose-escalation trial examined autologous BMSCs in patients with chronic SCI.<sup>66</sup> Although BMSCs were safe, they were not found to be beneficial in this cohort of patients. Having clearly established the safety and feasibility of the clinical use of BM-derived cells specifically for SCI in these trials, the continued testing of BMSCs in the context of SCI appears justified although the use of this intervention in complete thoracic cases may not be optimal. Based on the mechanism of action of BMSCs, which appear to provide trophic support to the penumbra zone of the acutely and subacutely injured cord, trials in patients with subacute severe, but incomplete spinal cord lesions are a consideration.

The use of BMSCs in SCI does, however, present certain issues. BMSC migration beyond the injection site (for intraspinally delivered cells) is limited, and inter-donor variability in efficacy and immunomodulatory potency might confer variable clinical outcome,<sup>81</sup> making evaluation of efficacy difficult. Studies of BMSCs in cervical contusion-compression models have yet to be carried out. BMSCs have, in all but two studies by the same group, been used in subacute and acute models.<sup>82-85</sup> Based on the limited number of pre-clinical studies in chronic models, it is not yet possible to evaluate their efficacy. It is also not known

whether BMSCs provide functional preservation of axons or *de novo* axonal regrowth across the lesion site in contusion-compression models, which are more appropriate models to distinguish these processes than transection models.<sup>86-89</sup>

## NEURAL PROGENITOR CELLS

NPCs can be generated from ESCs, which are derived from the blastocyst-stage embryo. These cells have indefinite self-renewal capacity and are pluripotent, with the potential to generate all cell types of the body, making them a potentially limitless source of cells for therapy. However, they are not without problems [Table 1], including the moral issues and practical constraints of their embryonic derivation, their karyotypic instability with repeated freeze-thaw cycles,<sup>90,91</sup> and their teratogenic potential in the host.

Pre-clinical studies have shown that animals transplanted with human ESC-derived oligodendrocytic progenitors cells (OPCs) show improvement in functional recovery following SCI.<sup>12,92-96</sup> With this background, extensive pre-clinical studies were conducted by Geron to characterize the safety and efficacy of hESC-OPCs exclusively in rodent models prior to the conduct (not without considerable objection and controversy) of a clinical trial of human ESC-derived OPCs implanted within 2 weeks into patients with thoracic SCI.<sup>97-99</sup> This Geron-sponsored clinical trial is further supported by behavioral and histological data from studies implanting

glial restricted progenitors (GRPs)<sup>100</sup> and OPCs<sup>101,102</sup> isolated from embryonic and post-natal rodents in SCI models, albeit indirectly. Whilst these show predominantly astroglial differentiation of GRP implanted within the blunt contusion-induced thoracic lesion site, there is a shift toward oligodendrocytic specification beyond the injury site correlated with the degree of functional improvement,<sup>103-105</sup> both of which can be enhanced by transduction of factors such as D15A, brain-derived neurotrophic factor (BDNF) and neurotrophin-3 (NT-3).<sup>106</sup> GRP implantation was also shown to be neuroprotective and inhibited neuropathic pain.

Neural progenitor cells can also be derived from several regions of the fetal, post-natal and adult CNS, including the sub-ventricular zone of the brain, the central canal of the spinal cord and the hippocampus. They can be expanded in culture as non-adherent neurospheres and have the potential to generate all three neural cell types under the appropriate conditions. The key advantage of this NPC source is the amenability to *in vitro* manipulation (including immortalization) prior to implantation as well as the lack of tumorigenicity. However, autologous derivation of CNS NPCs would be unfeasible for cell therapy purposes.

On the basis of promising results in clinically relevant primate<sup>107-109</sup> and canine cervical contusion models of SCI and cell number-dependent locomotor recovery in acute, subacute and chronic thoracic rodent models,<sup>110-113</sup> a StemCells Inc-sponsored clinical trial is underway to treat SCI sufferers with non-immortalized fetal human CNS stem cells (HuCNS-SC, NCT01321333). The lack of trials of NPCs in SCI is in spite of the bulk of pre-clinical findings to date in support of the potential of fetal and adult NPCs (particularly the former) in experimental SCI models,<sup>17</sup> and is likely to reflect ethical concerns over their origins and practical issues hindering their isolation and directed differentiation. Another possible explanation for the absence of clinical trials of NPCs for SCI is that the mechanisms through which NPCs provide functional benefit (including immunomodulation and angiogenesis) are only now beginning to be understood. Also, aims of axonal regeneration through the injury site have been replaced pre-clinically by more realistic objectives of remyelination<sup>17</sup> and provision of trophic support for endogenous precursors and axons. This makes NPCs much more promising candidates for cell therapy for SCI and probably heralds their increased use in clinical trials.

## CONCLUSION

Cell therapy can potentially enhance the quality of life of those affected by SCI. The significant advances that have been made on the basis of pre-clinical studies carried out in rodent models of SCI have enabled clinical trials demonstrating the safety of

cell therapy for SCI to proceed and have informed researchers of the knowledge gaps that remain to be addressed. However, rodent contusion/compression models of SCI are generally “incomplete” with partial sparing of motor and sensory functions, and mimic most closely patients with severe, partial lesions with an American Spinal Injury Association (ASIA) impairment scale rating of AIS B or C. Given that most trials of cell therapy have been carried out in AIS A patients (the safest to treat but also the least likely to show cell therapy-induced benefit), there is a need for future clinical trials to include patients modeled in the laboratory. There is a compelling need for preclinical researchers to develop valid models of compressive/contusive cervical SCI given that approximately 50–60% of human SCI involves the cervical region.

## ACKNOWLEDGEMENTS

Thank you to Dr Madeleine O'Higgins, Communications Specialist, for help with the manuscript and submission of this Editorial.

## REFERENCES

1. Sekhon LH, Fehlings MG. Epidemiology, demographics, and pathophysiology of acute spinal cord injury. *Spine (Phila Pa 1976)*, 2001;26(24 Suppl):S2-12.
2. Spinal Cord Injury Facts and Statistics, Rick Hansen Spinal Cord Injury Registry 2006. <http://rickhansenregistry.org/en/news-and-resources/sci-facts.html>.
3. Spinal cord injury facts and figures at a glance, National Spinal Cord Injury Statistical Center: Birmingham, Alabama: 2010. [https://www.nscisc.uab.edu/public\\_content/pdf/Facts%202011%20Feb%20Final.pdf](https://www.nscisc.uab.edu/public_content/pdf/Facts%202011%20Feb%20Final.pdf).
4. One Degree of Separation: Paralysis and Spinal Cord Injury in the United States, New Jersey: Christopher and Dana Reeve Foundation; 2010.
5. Fehlings MG, Tighe A. Spinal cord injury: The promise of translational research. *Neurosurg Focus* 2008;25:E1.
6. Tator CH, Fehlings MG. Review of the secondary injury theory of acute spinal cord trauma with emphasis on vascular mechanisms. *J Neurosurg* 1991;75:15-26.
7. Figley SA, Austin JW, Rowland JW, Fehlings MG. Pathophysiology of Spinal Cord Injury, in *The Cervical Spine* (in review), Philadelphia: Lippincott, Williams and Wilkins; 2011.
8. Wang L, Shi J, van Ginkel FW, Lan L, Niemeyer G, Martin DR, et al. Neural stem/progenitor cells modulate immune responses by suppressing T lymphocytes with nitric oxide and prostaglandin E2. *Exp Neurol* 2009;216:177-83.
9. Pluchino S, Gritti A, Blezer E, Amadio S, Brambilla E, Borsellino G, et al. Human neural stem cells ameliorate autoimmune encephalomyelitis in non-human primates. *Ann Neurol* 2009;66:343-54.
10. Okamura RM, Lebkowski J, Au M, Priest CA, Denham J, Majumdar AS. Immunological properties of human embryonic stem cell-derived oligodendrocyte progenitor cells. *J Neuroimmunol* 2007;192:134-44.
11. Bai L, Lennon DP, Eaton V, Maier K, Caplan AI, Miller SD, et al. Human bone marrow-derived mesenchymal stem cells induce Th2-polarized immune response and promote

- endogenous repair in animal models of multiple sclerosis. *Glia* 2009;57:1192-203.
12. Bottai D, Cigognini D, Madaschi L, Adami R, Nicora E, Menarini M, et al. Embryonic stem cells promote motor recovery and affect inflammatory cell infiltration in spinal cord injured mice. *Exp Neurol* 2010;223:452-63.
  13. Jaderstad J, Jäderstad LM, Li J, Chintawar S, Salto C, Pandolfo M, et al. Communication via gap junctions underlies early functional and beneficial interactions between grafted neural stem cells and the host. *Proc Natl Acad Sci U S A*, 2010;107:5184-9.
  14. Arriola A, Kiel ME, Shi Y, McKinnon RD. Adjunctive MSCs enhance myelin formation by xenogenic oligodendrocyte precursors transplanted in the retina. *Cell Res* 2010;20:728-31.
  15. Zhang J, Wang B, Xiao Z, Zhao Y, Chen B, Han J, et al. Olfactory ensheathing cells promote proliferation and inhibit neuronal differentiation of neural progenitor cells through activation of Notch signaling. *Neuroscience* 2008;153:406-13.
  16. Rowland JW, Hawryluk GW, Kwon B, Fehlings MG. Current status of acute spinal cord injury pathophysiology and emerging therapies: Promise on the horizon. *Neurosurg Focus* 2008;25:E2.
  17. Tetzlaff W, Okon EB, Karimi-Abdolrezaee S, Hill CE, Sparling JS, Plemel JR, et al. A systematic review of cellular transplantation therapies for spinal cord injury. *J Neurotrauma* 2011;28:1611-82.
  18. Campagnoli C, Roberts IA, Kumar S, Bennett PR, Bellantuono I, Fisk NM. Identification of mesenchymal stem/progenitor cells in human first-trimester fetal blood, liver, and bone marrow. *Blood* 2001;98:2396-402.
  19. Ikegami T, Nakamura M, Yamane J, Katoh H, Okada S, Iwanami A, et al. Chondroitinase ABC combined with neural stem/progenitor cell transplantation enhances graft cell migration and outgrowth of growth-associated protein-43-positive fibers after rat spinal cord injury. *Eur J Neurosci* 2005;22:3036-46.
  20. Karimi-Abdolrezaee S, Eftekharpour E, Wang J, Schut D, Fehlings MG. Synergistic effects of transplanted adult neural stem/progenitor cells, chondroitinase, and growth factors promote functional repair and plasticity of the chronically injured spinal cord. *J Neurosci* 2010;30:1657-76.
  21. Bottai D, Madaschi L, Di Giulio AM, Gorio A. Viability-dependent promoting action of adult neural precursors in spinal cord injury. *Mol Med* 2008;14:634-44.
  22. Hofstetter CP, Holmström NA, Lilja JA, Schweinhardt P, Hao J, Spenger C, et al. Allodynia limits the usefulness of intraspinal neural stem cell grafts; directed differentiation improves outcome. *Nat Neurosci* 2005;8:346-53.
  23. Mitchell KE, Weiss ML, Mitchell BM, Martin P, Davis D, Morales L, et al. Matrix cells from Wharton's jelly form neurons and glia. *Stem Cells* 2003;21:50-60.
  24. Wang HS, Hung SC, Peng ST, Huang CC, Wei HM, Guo YJ, et al. Mesenchymal stem cells in the Wharton's jelly of the human umbilical cord. *Stem Cells* 2004;22:1330-7.
  25. Weiss ML, Medicetty S, Bledsoe AR, Rachakatla RS, Choi M, Merchav S, et al. Human umbilical cord matrix stem cells: preliminary characterization and effect of transplantation in a rodent model of Parkinson's disease. *Stem Cells* 2006;24:781-92.
  26. La Rocca G, Anzalone R, Corrao S, Magno F, Loria T, Lo Iacono M, et al. Isolation and characterization of Oct-4+/HLA-G+ mesenchymal stem cells from human umbilical cord matrix: Differentiation potential and detection of new markers. *Histochem Cell Biol* 2009;131:267-82.
  27. Vawda R, Kennea NK, Mehmet H. Stem Cells in Neurodegeneration and Injury, in *Tissue stem cells*, In: Potten CS, Clarke RB, Wilson J and Renehan AG Editors. London: Taylor and Francis Pubs; 2006. p. 271-93.
  28. Vawda R, Woodbury J, Covey M, Levison SW, Mehmet H. Stem cell therapies for perinatal brain injuries. *Semin Fetal Neonatal Med* 2007;12:259-72.
  29. Troyer DL, Weiss ML. Wharton's jelly-derived cells are a primitive stromal cell population. *Stem Cells* 2008;26:591-9.
  30. Weiss ML, Anderson C, Medicetty S, Seshareddy KB, Weiss RJ, VanderWerff I, et al. Immune properties of human umbilical cord Wharton's jelly-derived cells. *Stem Cells* 2008;26:2865-74.
  31. Weiss ML, Mitchell KE, Hix JE, Medicetty S, El-Zarkouny SZ, Grieger D, et al. Transplantation of porcine umbilical cord matrix cells into the rat brain. *Exp Neurol* 2003;182:288-99.
  32. Lund RD, Wang S, Lu B, Girman S, Holmes T, Sauvé Y, et al. Cells isolated from umbilical cord tissue rescue photoreceptors and visual functions in a rodent model of retinal disease. *Stem Cells* 2007;25:602-11.
  33. Ganta C, Chiyo D, Ayuzawa R, Rachakatla R, Pyle M, Andrews G, et al. Rat umbilical cord stem cells completely abolish rat mammary carcinomas with no evidence of metastasis or recurrence 100 days post-tumor cell inoculation. *Cancer Res* 2009;69:1815-20.
  34. Ayuzawa R, Doi C, Rachakatla RS, Pyle MM, Maurya DK, Troyer D, et al. Naive human umbilical cord matrix derived stem cells significantly attenuate growth of human breast cancer cells *in vitro* and *in vivo*. *Cancer Lett* 2009;280:31-7.
  35. Yang CC, Shih YH, Ko MH, Hsu SY, Cheng H, Fu YS. Transplantation of human umbilical mesenchymal stem cells from Wharton's jelly after complete transection of the rat spinal cord. *PLoS ONE* 2008;3:e3336.
  36. Zhang L, Zhang HT, Hong SQ, Ma X, Jiang XD, Xu RX. Cografted Wharton's Jelly Cells-derived neurospheres and BDNF promote functional recovery after rat spinal cord transection. *Neurochem Res* 2009.
  37. Hernandez J, Torres-Espina A, Navarro X. Adult stem cell transplants for spinal cord injury repair: Current state in preclinical research. *Curr Stem Cell Res Ther* 2011;6:273-87.
  38. Biernaskie JA, McKenzie IA, Toma JG, Miller FD. Isolation of skin-derived precursors (SKPs) and differentiation and enrichment of their Schwann cell progeny. *Nat Protoc* 2006;1:2803-12.
  39. Biernaskie J, Miller FD. White matter repair: Skin-derived precursors as a source of myelinating cells. *Can J Neurol Sci* 2010;37 Suppl 2:S34-41.
  40. Biernaskie J, Sparling JS, Liu J, Shannon CP, Plemel JR, Xie Y, et al. Skin-derived precursors generate myelinating Schwann cells that promote remyelination and functional recovery after contusion spinal cord injury. *J Neurosci* 2007;27:9545-59.
  41. Fernandes KJ, McKenzie IA, Mill P, Smith KM, Akhavan M, Barnabé-Heider F, et al. A dermal niche for multipotent adult skin-derived precursor cells. *Nat Cell Biol* 2004;6:1082-93.
  42. Fernandes KJ, Toma JG, Miller FD. Multipotent skin-derived precursors: Adult neural crest-related precursors with therapeutic potential. *Philos Trans R Soc Lond B Biol Sci* 2008;363:185-98.
  43. McKenzie IA, Biernaskie J, Toma JG, Midha R, Miller FD. Skin-derived precursors generate myelinating Schwann cells for the injured and dysmyelinated nervous system. *J Neurosci* 2006;26:6651-60.
  44. Toma JG, Akhavan M, Fernandes KJ, Barnabé-Heider F, Sadikot A, Kaplan DR, et al. Isolation of multipotent adult stem cells from the dermis of mammalian skin. *Nat Cell Biol* 2001;3:78-84.

45. Toma JG, McKenzie IA, Bagli D, Miller FD. Isolation and characterization of multipotent skin-derived precursors from human skin. *Stem Cells* 2005;23:727-37.
46. Fernandes KJ, Kobayashi NR, Gallagher CJ, Barnabé-Heider F, Aumont A, Kaplan DR, et al. Analysis of the neurogenic potential of multipotent skin-derived precursors. *Exp Neurol* 2006;201:32-48.
47. Fernandes KJ, Miller FD. Isolation, expansion, and differentiation of mouse skin-derived precursors. *Methods Mol Biol* 2009;482:159-70.
48. Zhang F, Citra F, Wang DA. Prospects of induced pluripotent stem cell technology in regenerative medicine. *Tissue Eng Part B Rev* 2011;17:115-24.
49. Salewski RP, Eftekharpour E, Fehlings MG. Are induced pluripotent stem cells the future of cell-based regenerative therapies for spinal cord injury? *J Cell Physiol* 2010;222:515-21.
50. Parsons XH, Teng YD, Snyder EY. Important precautions when deriving patient-specific neural elements from pluripotent cells. *Cytotherapy* 2009; p. 1-11.
51. Tsuji O, Miura K, Okada Y, Fujiyoshi K, Mukaino M, Nagoshi N, et al. Therapeutic potential of appropriately evaluated safe-induced pluripotent stem cells for spinal cord injury. *Proc Natl Acad Sci U S A* 2010;107:12704-9.
52. Zhao T, Zhang ZN, Rong Z, Xu Y. Immunogenicity of induced pluripotent stem cells. *Nature* 2011;474:212-5.
53. Hamilton LK, Truong MK, Bednarczyk MR, Aumont A, Fernandes KJ. Cellular organization of the central canal ependymal zone, a niche of latent neural stem cells in the adult mammalian spinal cord. *Neuroscience* 2009;164:1044-56.
54. Meletis K, Barnabé-Heider F, Carlén M, Evergren E, Tomilin N, Shupliakov O, et al. Spinal cord injury reveals multilineage differentiation of ependymal cells. *PLoS Biol* 2008;6:e182.
55. Hawryluk GW, Fehlings MG. The center of the spinal cord may be central to its repair. *Cell Stem Cell* 2008;3:230-2.
56. Yamamoto S, Yamamoto N, Kitamura T, Nakamura K, Nakafuku M. Proliferation of parenchymal neural progenitors in response to injury in the adult rat spinal cord. *Exp Neurol* 2001;172:115-27.
57. Martens DJ, Seaberg RM, van der Kooy D. *In vivo* infusions of exogenous growth factors into the fourth ventricle of the adult mouse brain increase the proliferation of neural progenitors around the fourth ventricle and the central canal of the spinal cord. *Eur J Neurosci* 2002;16:1045-57.
58. Martens DJ, Tropepe V, van Der Kooy D. Separate proliferation kinetics of fibroblast growth factor-responsive and epidermal growth factor-responsive neural stem cells within the embryonic forebrain germinal zone. *J Neurosci* 2000;20:1085-95.
59. Ruff CA, Fehlings MG. Neural stem cells in regenerative medicine: bridging the gap. *Panminerva Med* 2010;52:125-47.
60. Zhang HT, Cheng HY, Cai YQ, Ma X, Liu WP, Yan ZJ, et al. Comparison of adult neurospheres derived from different origins for treatment of rat spinal cord injury. *Neurosci Lett* 2009;458:16-21.
61. Mackay-Sim A, St John JA. Olfactory ensheathing cells from the nose: Clinical application in human spinal cord injuries. *Exp Neurol* 2011;229:174-80.
62. Kwon BK, Hillyer J, Tetzlaff W. Translational research in spinal cord injury: a survey of opinion from the SCI community. *J Neurotrauma* 2010;27:21-33.
63. Kwon BK, Sekhon LH, Fehlings Mg. Emerging repair, regeneration, and translational research advances for spinal cord injury. *Spine (Phila Pa 1976)* 2010;35(21 Suppl):S263-70.
64. Gensel JC, Donnelly DJ, Popovich PG. Spinal cord injury therapies in humans: An overview of current clinical trials and their potential effects on intrinsic CNS macrophages. *Expert Opin Ther Targets* 2011;15:505-18.
65. Atkins H, Freedman M. Immune ablation followed by autologous hematopoietic stem cell transplantation for the treatment of poor prognosis multiple sclerosis. *Methods Mol Biol* 2009;549:231-46.
66. Freedman MS, Bar-Or A, Atkins HL, Karussis D, Frassoni F, Lazarus H, et al. The therapeutic potential of mesenchymal stem cell transplantation as a treatment for multiple sclerosis: Consensus report of the International MSCT Study Group. *Mult Scler* 2010;16:503-10.
67. Lu P, Blesch A, Tuszynski MH. Induction of bone marrow stromal cells to neurons: differentiation, transdifferentiation, or artifact? *J Neurosci Res* 2004;77:174-91.
68. Novikova LN, Brohlin M, Kingham PJ, Novikov LN, Wiberg M. Neuroprotective and growth-promoting effects of bone marrow stromal cells after cervical spinal cord injury in adult rats. *Cytotherapy* 2011;13:873-87.
69. Lu P, Jones LL, Tuszynski MH. Axon regeneration through scars and into sites of chronic spinal cord injury. *Exp Neurol* 2007;203:8-21.
70. Urdzikova L, Jendelová P, Glogarová K, Burian M, Hájek M, Syková E. Transplantation of bone marrow stem cells as well as mobilization by granulocyte-colony stimulating factor promotes recovery after spinal cord injury in rats. *J Neurotrauma* 2006;23:1379-91.
71. Ohta M, Suzuki Y, Noda T, Ejiri Y, Dezawa M, Kataoka K, et al. Bone marrow stromal cells infused into the cerebrospinal fluid promote functional recovery of the injured rat spinal cord with reduced cavity formation. *Exp Neurol* 2004;187:266-78.
72. Fan L, DU F, Cheng BC, Peng H, Liu SQ. Migration and distribution of bone marrow stromal cells in injured spinal cord with different transplantation techniques. *Chin J Traumatol* 2008;11:94-7.
73. Zurita M, Vaquero J. Functional recovery in chronic paraplegia after bone marrow stromal cells transplantation. *Neuroreport* 2004;15:1105-8.
74. Deng YB, Liu XG, Liu ZG, Liu XL, Liu Y, Zhou GQ. Implantation of BM mesenchymal stem cells into injured spinal cord elicits de novo neurogenesis and functional recovery: evidence from a study in rhesus monkeys. *Cytotherapy* 2006;8:210-4.
75. Kim MS, Kang KN, Lee JY, Kim da Y, Lee BN, Ahn HH, et al. Regeneration of completely transected spinal cord using scaffold of poly (D,L-lactide-co-glycolide)/small intestinal submucosa seeded with rat bone marrow stem cells. *Tissue Eng Part A*, 2011;17:2143-52.
76. Okada S, Ishii K, Yamane J, Iwanami A, Ikegami T, Katoh H, et al. *In vivo* imaging of engrafted neural stem cells: Its application in evaluating the optimal timing of transplantation for spinal cord injury. *Faseb J* 2005;19:1839-41.
77. Callera F, do Nascimento RX. Delivery of autologous bone marrow precursor cells into the spinal cord via lumbar puncture technique in patients with spinal cord injury: A preliminary safety study. *Exp Hematol* 2006;34:130-1.
78. Chernykh ER, Stupak VV, Muradov GM, Sizikov MY, Shevela EY, Leplina OY, et al. Application of autologous bone marrow stem cells in the therapy of spinal cord injury patients. *Bull Exp Biol Med* 2007;143:543-7.
79. Yoon SH, Shim YS, Park YH, Chung JK, Nam JH, Kim MO, et al. Complete spinal cord injury treatment using autologous bone marrow cell transplantation and bone marrow stimulation with granulocyte macrophage-colony stimulating factor: Phase I/II

- clinical trial. *Stem Cells* 2007;25:2066-73.
80. Saito F, Nakatani T, Iwase M, Maeda Y, Hirakawa A, Murao Y, et al. Spinal cord injury treatment with intrathecal autologous bone marrow stromal cell transplantation: The first clinical trial case report. *J Trauma* 2008;64:53-9.
  81. Neuhuber B, Timothy Himes B, Shumsky JS, Gallo G, Fischer I. Axon growth and recovery of function supported by human bone marrow stromal cells in the injured spinal cord exhibit donor variations. *Brain Res* 2005;1035:73-85.
  82. Vaquero J, Zurita M, Oya S, Santos M. Cell therapy using bone marrow stromal cells in chronic paraplegic rats: systemic or local administration? *Neurosci Lett* 2006;398:129-34.
  83. Zurita M, Vaquero J. Bone marrow stromal cells can achieve cure of chronic paraplegic rats: functional and morphological outcome one year after transplantation. *Neurosci Lett* 2006;402:51-6.
  84. Zurita M, Vaquero J, Bonilla C, Santos M, De Haro J, Oya S, et al. Functional recovery of chronic paraplegic pigs after autologous transplantation of bone marrow stromal cells. *Transplantation* 2008;86:845-53.
  85. de Haro J, Zurita M, Ayllón L, Vaquero J. Detection of <sup>111</sup>In-oxine-labeled bone marrow stromal cells after intravenous or intralesional administration in chronic paraplegic rats. *Neurosci Lett* 2005;377:7-11.
  86. Kamada T, Koda M, Dezawa M, Anahara R, Toyama Y, Yoshinaga K, et al. Transplantation of human bone marrow stromal cell-derived Schwann cells reduces cystic cavity and promotes functional recovery after contusion injury of adult rat spinal cord. *NeuroPathology* 2011;31:48-58.
  87. Kamada T, Koda M, Dezawa M, Yoshinaga K, Hashimoto M, Koshizuka S, et al. Transplantation of bone marrow stromal cell-derived Schwann cells promotes axonal regeneration and functional recovery after complete transection of adult rat spinal cord. *J Neuropathol Exp Neurol* 2005;64:37-45.
  88. Koda M, Kamada T, Hashimoto M, Murakami M, Shirasawa H, Sakao S, et al. Adenovirus vector-mediated *ex vivo* gene transfer of brain-derived neurotrophic factor to bone marrow stromal cells promotes axonal regeneration after transplantation in completely transected adult rat spinal cord. *Eur Spine J* 2007;16:2206-14.
  89. Koda M, Nishio Y, Kamada T, Someya Y, Okawa A, Mori C, et al. Granulocyte colony-stimulating factor (G-CSF) mobilizes bone marrow-derived cells into injured spinal cord and promotes functional recovery after compression-induced spinal cord injury in mice. *Brain Res* 2007;1149:223-31.
  90. Draper JS, Moore HD, Ruban LN, Gokhale PJ, Andrews PW. Culture and characterization of human embryonic stem cells. *Stem Cells Dev* 2004;13:325-36.
  91. Draper JS, Smith K, Gokhale P, Moore HD, Maltby E, Johnson J, et al. Recurrent gain of chromosomes 17q and 12 in cultured human embryonic stem cells. *Nat Biotechnol* 2004;22:53-4.
  92. Cloutier F, Siegenthaler MM, Nistor G, Keirstead HS. Transplantation of human embryonic stem cell-derived oligodendrocyte progenitors into rat spinal cord injuries does not cause harm. *Regen Med* 2006;1:469-79.
  93. Keirstead HS, Nistor G, Bernal G, Totoiu M, Cloutier F, Sharp K, et al. Human embryonic stem cell-derived oligodendrocyte progenitor cell transplants remyelinate and restore locomotion after spinal cord injury. *J Neurosci* 2005;25:4694-705.
  94. Faulkner J, Keirstead HS. Human embryonic stem cell-derived oligodendrocyte progenitors for the treatment of spinal cord injury. *Transpl Immunol* 2005;15:131-42.
  95. Nistor GI, Totoiu MO, Haque N, Carpenter MK, Keirstead HS. Human embryonic stem cells differentiate into oligodendrocytes in high purity and myelinate after spinal cord transplantation. *Glia* 2005;49:385-96.
  96. Sharp J, Frame J, Siegenthaler M, Nistor G, Keirstead HS. Human embryonic stem cell-derived oligodendrocyte progenitor cell transplants improve recovery after cervical spinal cord injury. *Stem Cells* 2010;28:152-63.
  97. Bretzner F, Gilbert F, Baylis F, Brownstone RM. Target populations for first-in-human embryonic stem cell research in spinal cord injury. *Cell Stem Cell* 2011;8:468-75.
  98. Solbakk JH, Zoloth L. The tragedy of translation: The case of "first use" in human embryonic stem cell research. *Cell Stem Cell* 2011;8:479-81.
  99. Wirth E 3rd, Lebkowski JS, Lebacqz K. Response to Frederic Bretzner et al. "Target populations for first-in-human embryonic stem cell research in spinal cord injury". *Cell Stem Cell* 2011;8:476-8.
  100. Mitsui T, Shumsky JS, Lepore AC, Murray M, Fischer I. Transplantation of neuronal and glial restricted precursors into contused spinal cord improves bladder and motor functions, decreases thermal hypersensitivity, and modifies intraspinal circuitry. *J Neurosci* 2005;25:9624-36.
  101. Bambakidis NC, Miller RH. Transplantation of oligodendrocyte precursors and sonic hedgehog results in improved function and white matter sparing in the spinal cords of adult rats after contusion. *Spine J* 2004;4:16-26.
  102. Lee KH, Yoon DH, Park YG, Lee BH. Effects of glial transplantation on functional recovery following acute spinal cord injury. *J Neurotrauma* 2005;22:575-89.
  103. Enzmann GU, Benton RL, Woock JP, Howard RM, Tsoulfas P, Whittemore SR. Consequences of noggin expression by neural stem, glial, and neuronal precursor cells engrafted into the injured spinal cord. *Exp Neurol* 2005;195:293-304.
  104. Han SS, Liu Y, Tyler-Polsz C, Rao MS, Fischer I. Transplantation of glial-restricted precursor cells into the adult spinal cord: survival, glial-specific differentiation, and preferential migration in white matter. *Glia* 2004;45:1-16.
  105. Hill CE, Proschel C, Noble M, Mayer-Proschel M, Gensel JC, Beattie MS, et al. Acute transplantation of glial-restricted precursor cells into spinal cord contusion injuries: survival, differentiation, and effects on lesion environment and axonal regeneration. *Exp Neurol* 2004;190:289-310.
  106. Cao Q, Xu XM, Devries WH, Enzmann GU, Ping P, Tsoulfas P, et al. Functional recovery in traumatic spinal cord injury after transplantation of multineurotrophin-expressing glial-restricted precursor cells. *J Neurosci* 2005;25:6947-57.
  107. Iwanami A, Kaneko S, Nakamura M, Kanemura Y, Mori H, Kobayashi S, et al. Transplantation of human neural stem cells for spinal cord injury in primates. *J Neurosci Res* 2005;80:182-90.
  108. Iwanami A, Yamane J, Katoh H, Nakamura M, Momoshima S, Ishii H, et al. Establishment of graded spinal cord injury model in a nonhuman primate: The common marmoset. *J Neurosci Res* 2005;80:172-81.
  109. Yamane J, Nakamura M, Iwanami A, Sakaguchi M, Katoh H, Yamada M, et al. Transplantation of galectin-1-expressing human neural stem cells into the injured spinal cord of adult common marmosets. *J Neurosci Res* 2010;88:1394-405.
  110. Cummings BJ, Uchida N, Tamaki SJ, Salazar DL, Hooshmand M, Summers R, et al. Human neural stem cells differentiate and promote locomotor recovery in spinal cord-injured mice. *Proc Natl Acad Sci U S A* 2005;102:14069-74.



111. Salazar DL, Uchida N, Hamers FP, Cummings BJ, Anderson AJ. Human neural stem cells differentiate and promote locomotor recovery in an early chronic spinal cord injury NOD-scid mouse model. *PLoS One* 2010;5:e12272.
112. Hooshmand MJ, Sontag CJ, Uchida N, Tamaki S, Anderson AJ, Cummings BJ. Analysis of host-mediated repair mechanisms after human CNS-stem cell transplantation for spinal cord injury: Correlation of engraftment with recovery. *PLoS One* 2009;4:e5871.
113. Cummings BJ, Uchida N, Tamaki SJ, Anderson AJ. Human neural stem cell differentiation following transplantation into spinal cord injured mice: Association with recovery of locomotor function. *Neurol Res* 2006;28:474-81.

**How to cite this article:** Vawda R, Wilcox J, Fehlings MG. Current stem cell treatments for spinal cord injury. *Indian J Orthop* 2012;46:10-8.

**Source of Support:** Nil, **Conflict of Interest:** None.

## New features on the journal's website

### Optimized content for mobile and hand-held devices

HTML pages have been optimized of mobile and other hand-held devices (such as iPad, Kindle, iPod) for faster browsing speed.

Click on [**Mobile Full text**] from Table of Contents page.

This is simple HTML version for faster download on mobiles (if viewed on desktop, it will be automatically redirected to full HTML version)

### E-Pub for hand-held devices

EPUB is an open e-book standard recommended by The International Digital Publishing Forum which is designed for reflowable content i.e. the text display can be optimized for a particular display device.

Click on [**EPub**] from Table of Contents page.

There are various e-Pub readers such as for Windows: Digital Editions, OS X: Calibre/Bookworm, iPhone/iPod Touch/iPad: Stanza, and Linux: Calibre/Bookworm.

### E-Book for desktop

One can also see the entire issue as printed here in a 'flip book' version on desktops.

Links are available from Current Issue as well as Archives pages.

Click on  View as eBook