

SCIENTIFIC REPORTS



OPEN

Association between aromatase in human brains and personality traits

Kayo Takahashi^{1,2,3}, Takamitsu Hosoya^{1,2,4}, Kayo Onoe^{1,2}, Tadayuki Takashima¹, Masaaki Tanaka³, Akira Ishii³, Yasuhito Nakatomi^{3,5}, Shusaku Tazawa¹, Kazuhiro Takahashi¹, Hisashi Doi^{1,2}, Yasuhiro Wada^{1,2,3} & Yasuyoshi Watanabe^{1,2,3}

Received: 11 January 2018

Accepted: 22 October 2018

Published online: 15 November 2018

Aromatase, an enzyme that converts androgens to estrogens, has been reported to be involved in several brain functions, including synaptic plasticity, neurogenesis, neuroprotection, and regulation of sexual and emotional behaviours in rodents, pathophysiology of Alzheimer's disease and autism spectrum disorders in humans. Aromatase has been reported to be involved in aggressive behaviours in genetically modified mice and in personality traits by genotyping studies on humans. However, no study has investigated the relationship between aromatase in living brains and personality traits including aggression. We performed a positron emission tomography (PET) study in 21 healthy subjects using ¹¹C-cetozole, which has high selectivity and affinity for aromatase. Before performing PET scans, subjects answered the Buss-Perry Aggression Questionnaire and Temperament and Character Inventory to measure their aggression and personality traits, respectively. A strong accumulation of ¹¹C-cetozole was detected in the thalamus, hypothalamus, amygdala, and medulla. Females showed associations between aromatase levels in subcortical regions, such as the amygdala and supraoptic nucleus of the hypothalamus, and personality traits such as aggression, novelty seeking, and self-transcendence. In contrast, males exhibited associations between aromatase levels in the cortices and harm avoidance, persistence, and self-transcendence. The association of aromatase levels in the thalamus with cooperativeness was common to both sexes. The present study suggests that there might exist associations between aromatase in the brain and personality traits. Some of these associations may differ between sexes, while others are likely common to both.

Aromatase is an enzyme that converts androgens to estrogens and is localized not only in the gonads but also in the brain¹. In mammals such as rodents and non-human primates, the regions rich in aromatase are the hypothalamus and amygdala²⁻⁴. In addition, the thalamus also contains high concentrations of aromatase in humans⁵⁻⁷. Aromatase in the brain has been suggested to be related to several brain functions. Aromatase knockout mice display altered aggressive behaviours⁸⁻¹¹, disrupted sexual behaviour^{9,11}, and displayed depressive-like behaviour¹². Human postmortem brain studies demonstrated lower aromatase expression in the hypothalamus of Alzheimer's disease¹³ and depression patients¹⁴, and in the frontal cortex of autism spectrum disorder (ASD) patients^{15,16}, as compared with the same brain regions of control subjects.

The association between aromatase and aggression in animals has been reviewed by Trainor *et al.*¹⁷. Rodent studies suggested that the medial amygdala has been related to aggression¹⁸⁻²⁰. In human studies, fMRI studies showed increased amygdala activation related to aggression^{21,22} and MRI morphometric study demonstrated reduced amygdala volume in healthy volunteers with higher aggression trait²³. The lines of evidence indicate that the amygdala may be a key region in aggression. Besides aggression, there is a study indicating a relation between aromatase gene polymorphism and personality trait, harm avoidance²⁴. So far, however, there was no study on the association between aromatase in living brain and personality traits including aggression. For assessment of personality traits, a questionnaire method is often used. In this study, we employed the Buss-Perry Aggression

¹RIKEN Center for Life Science Technologies, 6-7-3 Minatojima-minamimachi, Chuo-ku, Kobe, Hyogo, 650-0047, Japan. ²RIKEN Center for Biosystems Dynamics Research, 6-7-3 Minatojima-minamimachi, Chuo-ku, Kobe, Hyogo, 650-0047, Japan. ³Department of Physiology, Osaka City University Graduate School of Medicine, 1-4-3 Asahi-cho, Abeno-ku, Osaka, 545-8585, Japan. ⁴Institute of Biomaterials and Bioengineering, Tokyo Medical and Dental University, (TMDU), 2-3-10 Kanda-Surugadai, Chiyoda-ku, Tokyo, 101-0062, Japan. ⁵Department of Metabolism, Endocrinology and Molecular Medicine, Osaka City University Graduate School of Medicine, 1-4-3 Asahi-cho, Abeno-ku, Osaka, 545-8585, Japan. Correspondence and requests for materials should be addressed to Y.W. (email: yywata@riken.jp)

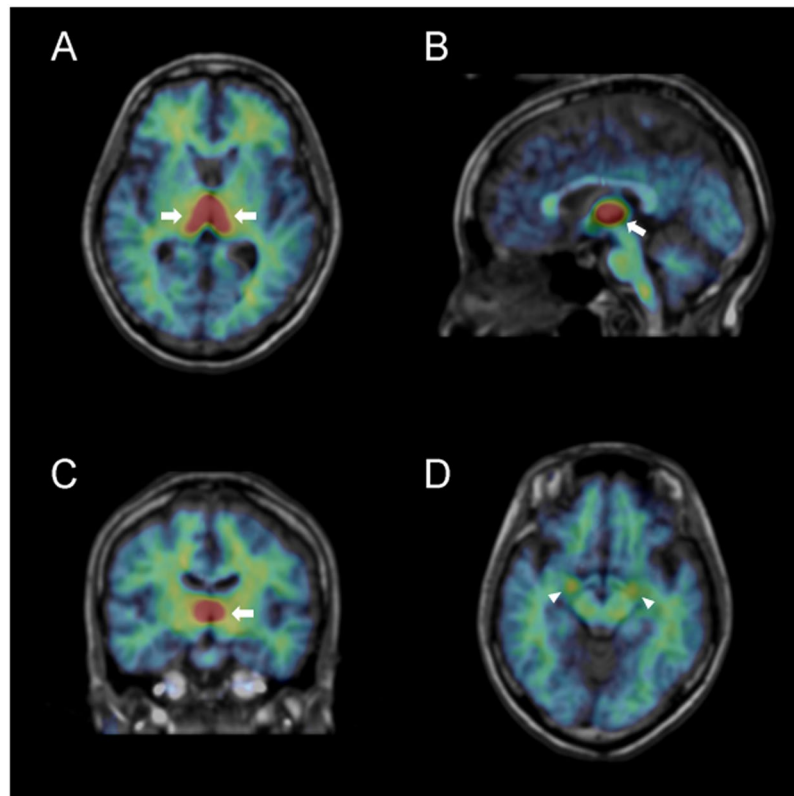


Figure 1. Representative distribution volume ($BP_{ND} + 1$) images of ^{11}C -cetrozole in a female human brain (rainbow colour scale) superimposed on the structural MR image (gray scale) of the same subject. (A) transaxial slice at level of the thalamus; (B) sagittal slice at the midline; (C) coronal slice at the level of the thalamus; (D) transaxial slice at the level of the amygdala. Arrows and arrow heads indicate the thalamus and amygdala, respectively.

Questionnaire (BAQ)^{25,26} for aggression assessment and the Temperament and Character Inventory (TCI)^{27,28}, which can evaluate personality traits, namely, novelty seeking, harm avoidance, reward dependence, persistence, self-directedness, cooperativeness, and self-transcendence.

To investigate molecular dynamics in the living body, positron emission tomography (PET) is a suitable technique, which allows quantitative analysis of compound accumulation in tissues. Previously, a PET study using ^{11}C -vorozole, which was developed as the first PET probe for aromatase imaging, was performed to image aromatase in the brain of healthy men and women^{6,7}. In that study, the authors demonstrated a unique distribution of aromatase in living human brains for the first time. However, a certain amount of nonspecific signals was observed in ^{11}C -vorozole PET images probably caused by unintended reuptake of the radioactive metabolite into the brain.

We have developed a novel PET probe for aromatase imaging to overcome the disadvantages of ^{11}C -vorozole, ^{11}C -cetrozole²⁹. Our previous study showed that ^{11}C -cetrozole had a higher signal-to-noise ratio than ^{11}C -vorozole since almost no radioactive metabolite of ^{11}C -cetrozole was not taken up into the brain, that is, ^{11}C -cetrozole is superior to ^{11}C -vorozole in terms of specificity and metabolic stability²⁹. In the present study, we performed a ^{11}C -cetrozole PET study in healthy subjects to examine the association between aromatase in the brain and personality traits.

Results

Distribution of aromatase in human brain. The binding potential (BP_{ND}) values of ^{11}C -cetrozole, which are an index of aromatase concentration, were calculated in 21 healthy individuals (10 females and 11 males). High BP_{ND} s were found in the thalamus with heterogeneous distribution among the medulla, hypothalamus, and amygdala in both sexes (Figs 1 and 2). Males had higher BP_{ND} values in most of these regions than females, except for in the right hypothalamus; however, a significant sex difference was found only in the left hypothalamus ($P = 0.005$; Fig. 2). There were no regions which BP_{ND} s depended on sex hormone levels in plasma (estradiol for females, and free testosterone for males).

Association between aromatase and personality traits. We assessed associations between BP_{ND} images of ^{11}C -cetrozole and scores of BAQ. Given that earlier studies showed that the amygdala is implicated in aggression^{18–23}, we focused on the amygdala as a volume of interest (VOI). Using Statistical Parametric Mapping 8 software (SPM8, Wellcome Department of Imaging Neuroscience), a voxel-wise analysis corrected by family-wise error rate for aggression scores on the VOI was performed. Apparently, region-specific differences among

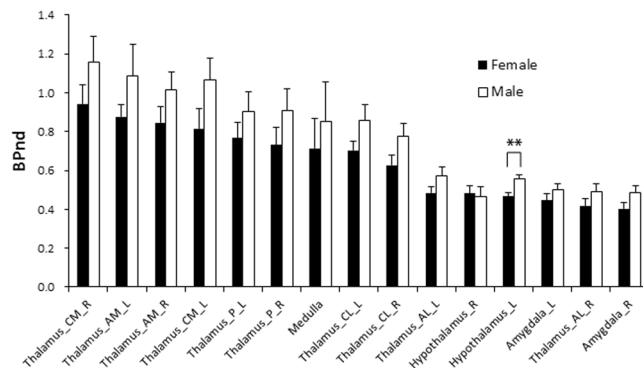


Figure 2. BP_{ND} values for the subregions of thalamus, amygdala, hypothalamus, and medulla. In all regions except for the right hypothalamus, males had higher BP_{ND} values than females. A significant sex difference was observed only in the left hypothalamus (**P = 0.005). AM, anterior medial; AL, anterior lateral; CM, central medial; CL, central lateral; P, posterior; L, left; R, right.

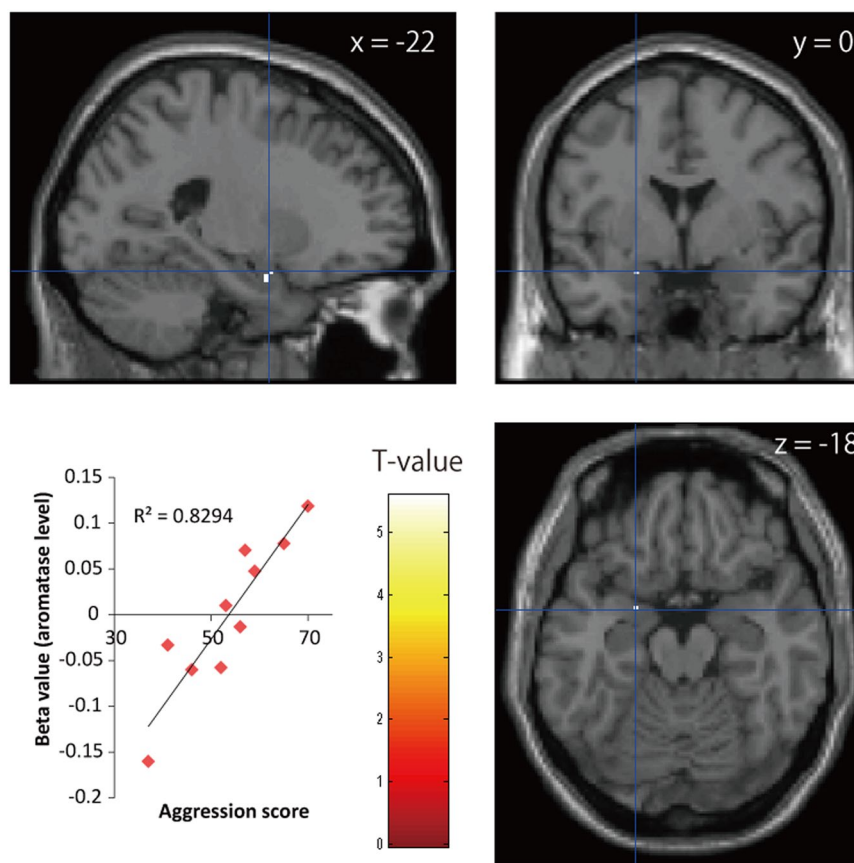


Figure 3. Statistical parametric maps of associations between ¹¹C-cetozole BP_{ND} values in the amygdala of females and aggression scores ($P_{\text{FWE-corr}} < 0.05$, 40 mm³). Peak coordinates ($x = -22$, $y = 0$, $z = -18$) are mapped on the template brain.

individuals are evident and the associations between personality traits and region-specific level of aromatase were also observed (Figs 3 and 4). In females, aggression scores were positively associated with BP_{ND} in the left amygdala ($P_{\text{FWE-corr}} < 0.05$, $R^2 = 0.83$, Fig. 3). In contrast, data from male and combined data from male and female did not exhibit a significant association between aggression scores and BP_{ND} in the amygdala. For an evaluation of individuals' personality traits, subjects answered TCI. Associations between traits and brain regions analysed in each sex group are listed in Table 1. Concerning other traits, sex-specific associations were observed in the amygdala and supraoptic nucleus of hypothalamus and in the inferior parietal gyrus in females. On the other hand, males-specific associations were observed in the anterior cingulate gyrus, supramarginal gyrus, caudate nucleus, pons, and midbrain. When the data of female and male were combined, almost no significant associations were

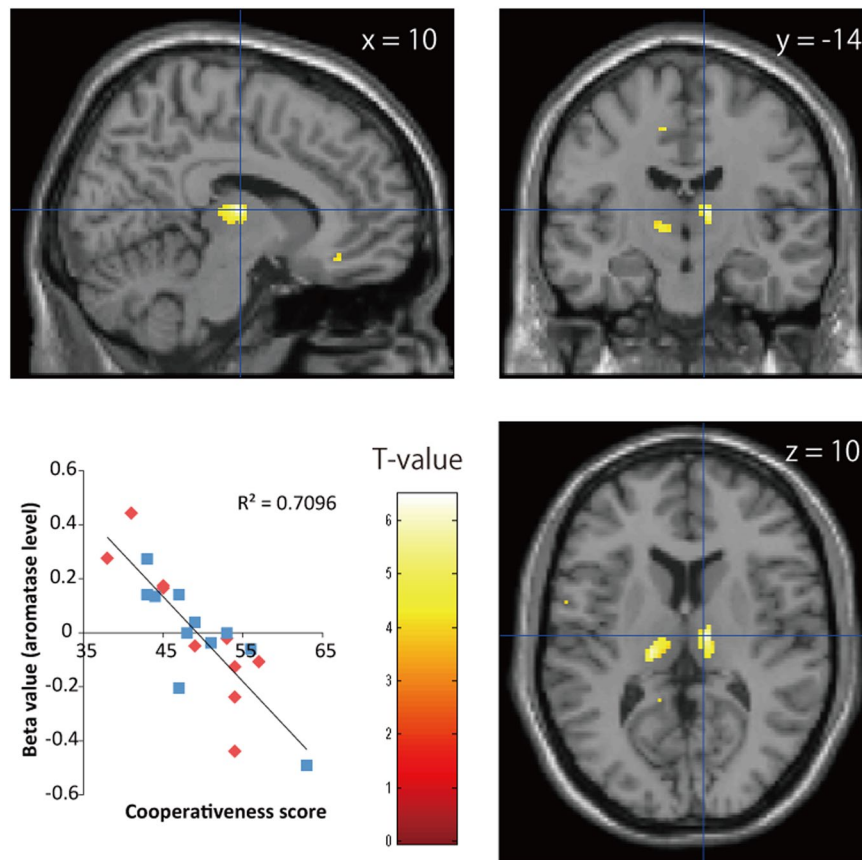


Figure 4. Statistical parametric maps of associations between ^{11}C -cetozole BP_{ND} values in the thalamus and cooperativeness scores in females and males ($P < 0.001$, uncorrected, 640 mm^3). Peak coordinates ($x = 10$, $y = -14$, $z = 10$) are mapped on the template brain. Red diamonds and blue squares represent the data from females and males, respectively.

found, except for cooperativeness. Cooperativeness scores were negatively associated with BP_{ND} in the bilateral thalamus of females and males ($R^2 = 0.71$, Fig. 4). The subregions that associated with cooperativeness were localized to the ventral lateral and ventral posterior parts of the thalamus.

There were 2 regions that exhibited associations between BP_{ND} of ^{11}C -cetozole and female traits, namely, the right supraoptic nucleus of the hypothalamus (SON; MNI: $x = 8$, $y = 2$, $z = -14$) and the right amygdala ($x = 24$, $y = -6$, $z = -20$), although several associations were present in small clusters (Tables 2 and 3, Fig. 5). The traits that associated with BP_{ND} in the SON were novelty seeking (negative, $R^2 = 0.998$), harm avoidance (positive, $R^2 = 0.97$), reward dependence (negative, $R^2 = 0.997$), persistence (negative, $R^2 = 0.98$), cooperativeness (positive, $R^2 = 0.995$), and self-transcendence (positive, $R^2 = 0.997$; Table 2). The traits associated with BP_{ND} in the right amygdala were novelty seeking (negative, $R^2 = 0.999$), persistence (negative, $R^2 = 0.994$), cooperativeness (positive, $R^2 = 0.999$), and self-transcendence (positive, $R^2 = 0.999$; Table 3).

Discussion

In this study, we demonstrated the distribution of aromatase in living human brains using our originally developed PET probe, ^{11}C -cetozole, and suggested that aromatase levels in the brain may relate to personality traits. The first PET scan of brain aromatase using ^{11}C -vorozole was performed by Biegon *et al.*^{6,7}. They demonstrated high levels of aromatase in the thalamus, amygdala, preoptic area, medulla, etc. ^{11}C -Vorozole had high specificity and affinity for aromatase; however, the metabolites of ^{11}C -vorozole were taken up into the brain with radiolabelling²⁹. Thus, the measurements were less quantitative. Aiming for a more quantitative measurement of aromatase, we developed ^{11}C -cetozole as a novel PET probe for aromatase imaging²⁹.

In the present study, we performed PET scans with ^{11}C -cetozole in 21 healthy human subjects. Approximately 50% of parental compound was intact 60 min after injection, indicating that this PET probe was suitable to measure aromatase in a living body (Supplemental Fig. S1). High BP_{ND} s of ^{11}C -cetozole were found in the thalamus with a heterogeneous distribution among the amygdala, hypothalamus, and medulla in both sexes (Figs 1 and 2). The distribution pattern of ^{11}C -cetozole binding was consistent with that of ^{11}C -vorozole⁷ and with immunohistochemistry in the postmortem human brain⁵, suggesting that considerable aromatase enzyme is expressed in the thalamus, hypothalamus, amygdala, and medulla in the human brain. Unlike other mammals that have more aromatase enzyme in male brains, such as rats and monkeys²⁻⁴, there was no distinct sex difference in aromatase levels in our human cohort, except for in the left hypothalamus ($P = 0.005$). However, males showed a tendency towards relatively

Traits	Association	Sex	Region	Side	MNI Coordinates			P value	Z score	Cluster size (mm ³)
					x	y	z			
Novelty Seeking	Negative	F	Hypothalamus(SON)	R	8	2	-14	<0.001	4.04	136
	Negative	F	Amygdala	R	24	-6	-20	<0.001	3.71	80
	Positive	M	Caudate nucleus	R	8	18	-4	<0.001	3.96	104
Harm Avoidance	Negative	M	Pons	R	10	-22	-30	<0.001	3.55	88
	Negative	M	Supramarginal gyrus	R	60	-44	34	<0.001	3.38	88
Reward Dependence	Positive	M	Thalamus	L	-18	-26	6	<0.001	3.79	152
Persistence	Negative	M	Anterior cingulate gyrus	L	-12	42	14	<0.001	3.76	96
	Negative	M	Supramarginal gyrus	R	58	-46	38	<0.001	3.43	96
	Negative	F & M	Lingual gyrus	L	-16	-76	-12	<0.001	3.54	144
Self-Directedness	Positive	F	Inferior parietal gyrus	L	-36	-56	46	<0.001	3.58	96
Cooperativeness	Positive	M	Anterior cingulate gyrus	L	-12	40	20	<0.001	3.94	104
	Negative	F & M	Thalamus	R	10	-14	10	<0.001	4.17	640
	Negative	F & M	Thalamus	L	-16	-24	8	<0.001	3.99	1296
	Negative	M	Superior frontal gyrus	L	-22	34	38	<0.001	3.87	96
	Negative	M	Inferior frontal gyrus, triangular part	L	-48	28	24	<0.001	3.76	168
	Negative	F & M	Inferior frontal gyrus, orbital part	L	-44	24	-6	<0.001	3.76	160
Self-Transcendence	Positive	F	Hypothalamus (SON)	R	8	2	-14	<0.001	4.01	120
	Negative	M	Anterior cingulate gyrus	L	-12	40	20	<0.001	4.02	152
	Negative	M	Midbrain	R	8	-26	-18	<0.001	3.70	80

Table 1. The association between ¹¹C-cetozole BP_{ND} and traits (scores on TCI; P < 0.001, uncorrected, cluster size ≥ 80 mm³).

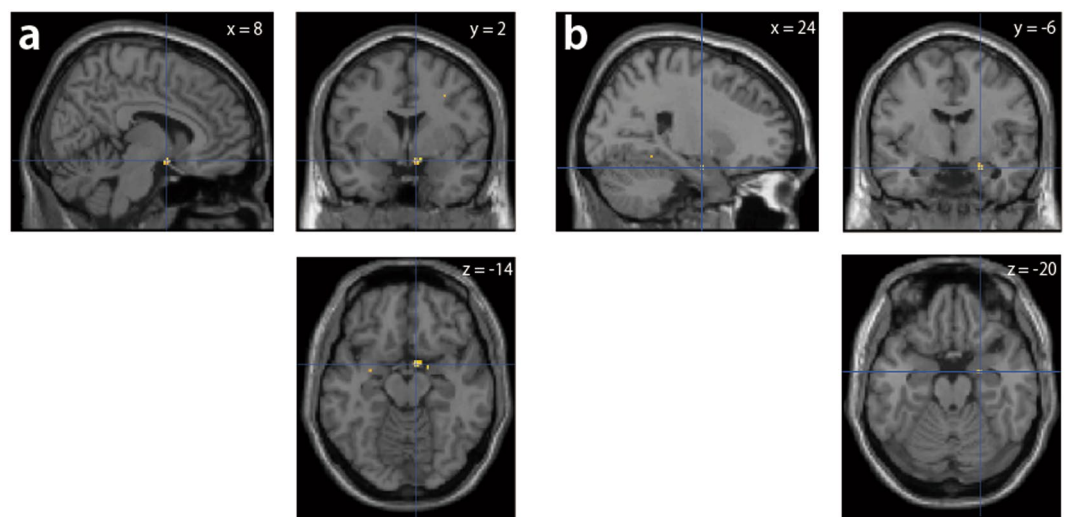


Figure 5. Two regions that exhibited associations between BP_{ND} of ¹¹C-cetozole and female traits are the right supraoptic nucleus of the hypothalamus ((a) SON; MNI: x = 8, y = 2, z = -14) and the right amygdala ((b) x = 24, y = -6, z = -20).

higher aromatase expression in all brain regions than females, except for the right hypothalamus. The reason why males have more aromatase in the brain than females do is considered to compensate lower circulating estrogens since estrogen is an important hormone related to regulation of sexual behaviour and emotions, neural plasticity, neuroprotection, etc. in the brain^{30–32}. In the present study we showed that BP_{ND} of ¹¹C-cetozole varied between individuals, especially in the thalamic subregions. This variability was associated with personality trait variability.

Both females and males showed a negative association between BP_{ND} of ¹¹C-cetozole and cooperative scores in the thalamus region in this study. The subregions that associated with cooperativeness were localized to the ventral lateral and ventral posterior parts of the thalamus. The ventral lateral nucleus of the thalamus contains estrogen receptor β and is known to project to the primary motor cortex³³. Diffusion-weighted imaging studies have segmented the thalamus on a connectivity basis and reported individual variations in segmentation³⁴. Johansen-Berg *et al.*³⁴ reported that the ventral lateral nucleus primarily connected to the prefrontal cortex in some subjects and to the primary motor cortex in others. This individual variation may provide cause of different personality. High aromatase in the thalamus is unique to humans, while monkeys, baboons, and rats have high

Traits	Association	P value	Z score	Cluster size (mm ³)
Novelty Seeking	Negative	<0.001	4.04	136
Harm Avoidance	Positive	<0.001	3.43	16
Reward Dependence	Negative	<0.001	3.06	56
Persistence	Negative	<0.001	3.58	24
Cooperativeness	Positive	<0.001	3.62	32
Self-Transcendence	Positive	<0.001	4.01	120

Table 2. Traits in females that associated with BP_{ND} of ¹¹C-cetozole in the SON ($x = 8, y = 2, z = -14$). No traits in males were associated with BP_{ND} in this region.

amount of aromatase in the amygdala and hypothalamus^{3,4,7,29,35,36}. The characteristically high social abilities of humans such as cooperativeness may be processed in the thalamus through the regulation of estrogens.

In addition to cooperativeness and the thalamus, the traits and associated regions were different between sexes (Table 1). In females, sex-specific associations were observed in the amygdala and supraoptic nucleus of hypothalamus and in the inferior parietal gyrus. On the other hand, males-specific associations were observed in the anterior cingulate gyrus, supramarginal gyrus, caudate nucleus, pons, and midbrain.

The right SON and the right amygdala showed associations between BP_{ND}s of ¹¹C-cetozole and female traits (Fig. 5). The localization of aromatase in the SON was demonstrated previously via immunohistochemistry¹³. Further, the SON is known to be a region in which oxytocin is synthesized³⁷. Oxytocin production is regulated by estradiol³⁸. Regulation of oxytocin by estradiol may affect personality traits given that oxytocin is implicated in stress susceptibility, emotion, memory, and social interaction³⁹. The traits that associated with BP_{ND} in the SON were novelty seeking (negative), harm avoidance (positive), reward dependence (negative), persistence (negative), cooperativeness (positive), and self-transcendence (positive; Table 2). The traits associated with BP_{ND} in the right amygdala were novelty seeking (negative), persistence (negative), cooperativeness (positive), and self-transcendence (positive; Table 3). These 4 traits are consistent with the traits that associated with BP_{ND} in the SON, which suggests functional connectivity between the right amygdala and SON in females.

Since animal and human studies suggested that the amygdala is involved in aggression^{18–23}, the VOI was drawn on the amygdala, and a regression analysis was performed between BP_{ND} of ¹¹C-cetozole in the amygdala and aggression score. There was a positive association between aggression scores and BP_{ND} in the left amygdala in females. Earlier studies found a relationship between aggression and reactivity in the left amygdala, although one of the studies combined female and male data^{21,22}. Another study reported that lower amygdala volume was correlated with more aggression in healthy females²³. Animal studies have demonstrated that aromatase neurons in the medial amygdala regulate male aggression and maternal aggression²⁰. Our results show that aromatase in the female left amygdala is associated with aggression, which indicates that estrogen synthesis in the left amygdala may induce aggression in females.

Furthermore, polymorphisms in the aromatase and estrogen receptor genes have been reported to be associated with harm-avoidance traits^{24,40}. Although we did not perform genetic analyses, gene polymorphisms or DNA methylation may affect the expression of aromatase or hormone receptors. Multi-disciplinary studies that consider gene polymorphisms, epigenetic changes, and imaging are needed in the future.

Our results showed that there may be associations between aromatase in the brain and personality traits, and some of the associations may differ between sexes, while others are likely common to both. Whether sex differences exist in the brain has long been contentious and remains controversial. Classical differences in brain structure, e.g., greater corpus callosum volume in females or enlarged cortical language regions in females, have been dismissed via meta-analysis⁴¹. However, neurochemistry suggests tendencies for females to have higher activity in serotonergic (5-HT transporter, 5-HT_{1A} and 5-HT_{2A} receptors), dopaminergic (dopamine transporter), and GABAergic (neurotransmitter level) systems⁴², which are involved in mood, emotions, and personality traits. An animal study revealed that exogenous estrogen treatments increased the mRNA and protein levels of tryptophan hydroxylase, a rate-limiting enzyme in the production of a serotonin precursor, in the raphe nucleus of spayed rhesus monkeys⁴³. Further studies are needed to clarify the differences and commonalities between sexes.

Previously the results between monoamine oxidase (MAO) level measured with PET and mood/personality disorders were reported^{44,45}. Our present results also show that the aromatase level in different brain sub-regions measured with PET is associated with a variety of personality traits. The dynamics of interaction between sex hormone and monoaminergic neurotransmitter systems are partly regulated by both enzyme levels. The PET studies in combination of both enzymes may be an interesting target in the future study.

There are several limitations of the present study. The associations between TCI scores and BP_{ND} did not survive a family-wise error correction, then were analysed with a significance threshold of $P < 0.001$, uncorrected, $k \geq 10$ voxels. As for the SON and the right amygdala of females, the associations with TCI scores were discussed even though the criteria were not fulfilled ($k < 10$ voxels). When we could increase the number of subjects, the results might become clearer. As regards sex hormone in plasma, we measured testosterone, free testosterone, estradiol, and progesterone in both males and females, however, free testosterone in female ($N = 2$), estradiol in male ($N = 7$) were too little to be quantified. Thus we tested the association between aromatase level and free testosterone in male or estradiol in female.

Although the subjects in this study were all healthy, there were associations between aromatase levels in the brain and personality traits, which suggests that regulation of estrogens might affect personality. Further PET

Traits	Association	P value	Z score	Cluster size (mm ³)
Novelty Seeking	Negative	<0.001	3.71	80
Persistence	Negative	<0.001	3.40	24
Cooperativeness	Positive	<0.001	3.44	32
Self-Transcendence	Positive	<0.001	3.50	40

Table 3. Traits in females that associated with the BP_{ND} of ¹¹C-cetozole in the right amygdala (x = 24, y = -6, z = -20). No traits in males were associated with this region.

studies that include patients with personality disorders using other PET tracers for estrogen and androgen receptors would help clarify the association between sex hormone systems and personality traits.

Methods

Subjects. Subjects were recruited by advertisements at Osaka City University and RIKEN. Twenty-four healthy adults (11 females and 13 males) participated in the present study. Participants were excluded if they had past or current serious medical illnesses and/or organic brain diseases or if they took drugs actin on the central nervous system. All females were not taking oral contraceptives and had regular menstrual cycle. Two females (35 and 45 yrs old) and 2 males (36 and 35 yrs old) received a whole-body PET scan to measure their radiation exposure, and 10 females and 11 males (average age of 34.7 ± 6.4 and 31.7 ± 8.1 yrs, mean \pm SD, respectively) received a brain PET scan. Each subject completed the validated Japanese version of BAQ²⁵ and TCI^{27,28} before PET scanning. All experiments were conducted in compliance with national legislation and the Code of Ethical Principles for Medical Research Involving Human Subjects of the World Medical Association (the *Declaration of Helsinki*) and registered to the UMIN Clinical Trials Registry (No. UMIN000006586). This study was approved by the Ethics Committee of the Kobe Institute of RIKEN and the Osaka City University Graduate School of Medicine. All participants provided written informed consent for participation in the study.

Positron emission tomography. ¹¹C-Cetozole was synthesized at Osaka City University Hospital according to the previously published procedure²⁹. The desired compound was dissolved in a mixture of polysorbate 80 (0.1 ml), propylene glycol (0.9 ml), and saline (9 ml). The identity and concentration of ¹¹C-cetozole were assessed using high-performance liquid chromatography. The specific activity of ¹¹C-cetozole was 81.2 ± 26.0 GBq/ μ mol (mean \pm SD) at administration. The radiochemical purity was greater than 99.5%. The dose of ¹¹C-cetozole was 4.7 ± 1.0 MBq/kg bodyweight. Subjects were positioned in the PET scanner (Biograph-16, Siemens, Knoxville, TN, USA) with their heads lightly tied with bandages to minimize movement during the scans. The left and right median cubital veins were cannulated for blood sampling and radiotracer administration, respectively. Four males received cannulation in the left radial artery instead of the median cubital vein for arterial blood sampling. Before the emission scans, CT scans were performed for head positioning and attenuation correction. At the start of the emission scan, ¹¹C-cetozole was intravenously administered for approximately 30 sec, and the catheter line was flushed with 15–20 ml saline to prevent radiotracer retention. Serial PET scanning of the brain was performed for 60 min in the 3-dimensional dynamic mode in the following frames: 6×10 sec, 6×30 sec, 11×60 sec, and 15×180 sec. Blood samples were taken 5, 10, 20, 30, 45, and 60 min after administration of ¹¹C-cetozole from the venous line and at 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, and 180 sec, and 4, 5, 10, 20, 30, 45, and 60 min after administration from the arterial line. Venous and arterial blood samples taken after 5 min and later were used for radiometabolite analyses.

Magnetic resonance (MR) image. Fine structural whole-brain T1-weighted magnetic resonance anatomical images were acquired using the Philips Achieva 3.0 TX (Royal Philips Electronics, Eindhoven, The Netherlands) with the following parameters: Repetition time = 5.9 msec, echo time = 2.7 msec, flip angle = 12 degrees, slice gap = 0 mm, matrix size = 256, field of view = 220 mm, voxel size = $0.86 \times 0.86 \times 0.90$ mm.

PET image processing. Brain PET images were reconstructed by Fourier rebinning and 2-dimensional filtered backprojection without additional smoothing filters. For quantitative image analyses, PMOD software (PMOD Technologies Ltd., Zurich, Switzerland) was used. VOIs were delineated in the thalamus, amygdala, hypothalamus, and medulla, which are structures known to contain a rich supply of aromatase enzyme^{5–7}, and in the cerebellar lobules on the same individual's MR images and transferred to PET images. Decay-corrected time-activity curves were generated for each brain region, arterial blood plasma, and parent unchanged compound, as measured by thin-layer chromatography (Supplemental Fig. S1). The time-activity curves for plasma and parent unchanged fraction were fitted to a 3-exponential model and a Hill function, respectively. The data with arterial blood sampling were analysed with a Logan plot⁴⁶, and the total distribution volume (Vt) in each brain region was calculated. The data without arterial blood sampling were analysed with a Logan reference tissue model based on average k₂⁴⁷ using the cerebellum as a reference. Nondisplaceable binding potential (BP_{ND}) and distribution volume ratio (DVR), which are linear functions of enzyme availability, were calculated ($DVR = BP_{ND} + 1$). Since no arterial blood sampling is preferable because of possible intense pain caused by arterial puncture, we compared the data with or without arterial blood sampling, that is, the data calculated by Logan plot or by Logan reference tissue model. Vt values of all examined brain regions were divided by the Vt value of the cerebellum and were compared with BP_{ND} values. Since the difference between normalized Vt values and BP_{ND} was $4 \pm 1\%$ (mean \pm SD, N = 4), we decided to employ

the Logan reference tissue model to analyse the remainder of the data. Then, BP_{ND} images were generated by model fitting. After co-registration of PET and MR images, whole-head structural images were normalized to the Montréal Neurological Institute (MNI) T1 image template, with the same parameters applied to the BP_{ND} images. BP_{ND} images were resampled to a voxel size of 2.0 × 2.0 × 2.0 mm using SPM8.

Quantification of aromatase in the rich regions. For measurements of the aromatase level in the brain, VOIs of the amygdala, hypothalamus, thalamus, and medulla were superimposed on BP_{ND} images. Because ¹¹C-cetozole binding was heterogeneously distributed in the thalamus, the thalamus was further divided into 5 subregions, namely, anterior medial, anterior lateral, central medial, central lateral, and posterior parts, consistent with a prior study⁴⁸. BP_{ND} values were extracted from BP_{ND} images. The difference between sexes in each VOI was analysed by a t-test. All P-values were two-tailed, and P values less than 0.05 were considered significant. These analyses were performed with the GraphPad PRISM 5.0 software package (GraphPad Software, Inc., La Jolla, CA).

Statistical analysis of PET images. For aggression, we focused on the amygdala since earlier studies showed that this region is implicated in aggression^{18–23}. A VOI of the amygdala was delineated using the WFU-Pickatlas SPM tool⁴⁹. Using SPM8, voxel-wise analysis was performed on the amygdala of BP_{ND} images by applying the score of BAQ as a covariate. A family-wise error corrected significance threshold was set at P < 0.05 in the amygdala. The associations between scores of TCI and BP_{ND} were analysed as whole-brain analyses with a significance threshold of P < 0.001, uncorrected, k ≥ 10 voxels (=80 mm³). Voxel-wise analyses were performed on the BP_{ND} images using the general linear model in SPM8, with covariates of 7 traits of TCI. Although the criteria were not fulfilled (k < 10 voxels), 6 and 4 traits of TCI showed the associations in the identical coordinates (x = 8, y = 2, z = -14 and x = 24, y = -6, z = -20, respectively), thus these 2 regions were discussed separately.

References

- Simpson, E. R. *et al.* Aromatase—a brief overview. *Annu. Rev. Physiol.* **64**, 93–127 (2002).
- Roselli, C. E., Abdelgadir, S. E. & Resko, J. A. Regulation of aromatase gene expression in the adult rat brain. *Brain Res. Bull.* **44**, 351–357 (1997).
- Abdelgadir, S. E., Roselli, C. E., Choate, J. V. & Resko, J. A. Distribution of aromatase cytochrome P450 messenger ribonucleic acid in adult rhesus monkey brains. *Biol. Reprod.* **57**, 772–777 (1997).
- Takahashi, K. *et al.* Imaging of aromatase distribution in rat and rhesus monkey brains with [¹¹C]vorozole. *Nucl. Med. Biol.* **33**, 599–605 (2006).
- Sasano, H., Takahashi, K., Satoh, F., Nagura, H. & Harada, N. Aromatase in the human central nervous system. *Clin. Endocrinol. (Oxf)*. **48**, 325–329 (1998).
- Biegon, A. *et al.* Aromatase imaging with [N-methyl-¹¹C]vorozole PET in healthy men and women. *J. Nucl. Med.* **56**, 580–585, <https://doi.org/10.2967/jnumed.114.150383> (2015).
- Biegon, A. *et al.* Unique distribution of aromatase in the human brain: *in vivo* studies with PET and [N-methyl-¹¹C]vorozole. *Synapse* **64**, 801–807, <https://doi.org/10.1002/syn.20791> (2010).
- Bakker, J., Honda, S., Harada, N. & Balthazart, J. Restoration of male sexual behavior by adult exogenous estrogens in male aromatase knockout mice. *Horm. Behav.* **46**, 1–10, <https://doi.org/10.1016/j.yhbeh.2004.02.003> (2004).
- Honda, S., Harada, N., Ito, S., Takagi, Y. & Maeda, S. Disruption of sexual behavior in male aromatase-deficient mice lacking exons 1 and 2 of the *cyp19* gene. *Biochem. Biophys. Res. Commun.* **252**, 445–449 (1998).
- Honda, S., Wakatsuki, T. & Harada, N. Behavioral analysis of genetically modified mice indicates essential roles of neurosteroidal estrogen. *Frontiers in endocrinology* **2**, 1–8, <https://doi.org/10.3389/fendo.2011.00040> (2011).
- Sato, T. *et al.* Brain masculinization requires androgen receptor function. *Proc. Natl. Acad. Sci. USA* **101**, 1673–1678, <https://doi.org/10.1073/pnas.0305303101> (2004).
- Dalla, C., Antoniou, K., Papadopoulou-Daifoti, Z., Balthazart, J. & Bakker, J. Oestrogen-deficient female aromatase knockout (ArKO) mice exhibit depressive-like symptomatology. *Eur. J. Neurosci.* **20**, 217–228 (2004).
- Ishunina, T. A. *et al.* Diminished aromatase immunoreactivity in the hypothalamus, but not in the basal forebrain nuclei in Alzheimer's disease. *Neurobiol. Aging* **26**, 173–194 (2005).
- Wu, J. L. *et al.* Aromatase changes in depression: A postmortem and animal experimental study. *Psychoneuroendocrinology* **77**, 56–62, <https://doi.org/10.1016/j.psyneuen.2016.11.026> (2017).
- Sarachana, T., Xu, M., Wu, R. C. & Hu, V. W. Sex hormones in autism: androgens and estrogens differentially and reciprocally regulate RORA, a novel candidate gene for autism. *PLoS one* **6**, e17116, <https://doi.org/10.1371/journal.pone.0017116> (2011).
- Crider, A., Thakkar, R., Ahmed, A. O. & Pillai, A. Dysregulation of estrogen receptor beta (ERbeta), aromatase (CYP19A1), and ER co-activators in the middle frontal gyrus of autism spectrum disorder subjects. *Molecular autism* **5**, 46, <https://doi.org/10.1186/2040-2392-5-46> (2014).
- Trainor, B. C., Kyomen, H. H. & Marler, C. A. Estrogenic encounters: how interactions between aromatase and the environment modulate aggression. *Front. Neuroendocrinol.* **27**, 170–179 (2006).
- Halasz, J., Liposits, Z., Meelis, W., Kruk, M. R. & Haller, J. Hypothalamic attack area-mediated activation of the forebrain in aggression. *Neuroreport* **13**, 1267–1270 (2002).
- Kollack-Walker, S. & Newman, S. W. Mating and agonistic behavior produce different patterns of Fos immunolabeling in the male Syrian hamster brain. *Neuroscience* **66**, 721–736 (1995).
- Unger, E. K. *et al.* Medial amygdalar aromatase neurons regulate aggression in both sexes. *Cell reports* **10**, 453–462, <https://doi.org/10.1016/j.celrep.2014.12.040> (2015).
- Beaver, J. D., Lawrence, A. D., Passamonti, L. & Calder, A. J. Appetitive motivation predicts the neural response to facial signals of aggression. *J. Neurosci.* **28**, 2719–2725, <https://doi.org/10.1523/JNEUROSCI.0033-08.2008> (2008).
- Coccaro, E. F., McCloskey, M. S., Fitzgerald, D. A. & Phan, K. L. Amygdala and orbitofrontal reactivity to social threat in individuals with impulsive aggression. *Biol. Psychiatry* **62**, 168–178, <https://doi.org/10.1016/j.biopsych.2006.08.024> (2007).
- Matthies, S. *et al.* Small amygdala-high aggression? The role of the amygdala in modulating aggression in healthy subjects. *The world journal of biological psychiatry: the official journal of the World Federation of Societies of Biological Psychiatry* **13**, 75–81, <https://doi.org/10.3109/15622975.2010.541282> (2012).
- Matsumoto, Y. *et al.* Effect of the cytochrome P450 19 (aromatase) gene polymorphism on personality traits in healthy subjects. *Behav. Brain Res.* **205**, 234–237, <https://doi.org/10.1016/j.bbr.2009.06.034> (2009).
- Ando, A. *et al.* Development of the Japanese version of the Buss-Perry Aggression Questionnaire (BAQ). *Shinrigaku Kenkyu.* **70**, 384–392 (1999).
- Buss, A. H. & Perry, M. The aggression questionnaire. *J. Pers. Soc. Psychol.* **63**, 452–459 (1992).

27. Kijima, N., Tanaka, E., Suzuki, N., Higuchi, H. & Kitamura, T. Reliability and validity of the Japanese version of the Temperament and Character Inventory. *Psychol. Rep.* **86**, 1050–1058 (2000).
28. Cloninger, C. R., Svrakic, D. M., Przybeck, T. R. & Wetzel, R. D. The Temperament and Character Inventory (TCI): a guide to its development and use. (ed. Cloninger, C. R.) (Center for Psychobiology of Personality, Washington University, St. Louis, Missouri 1994).
29. Takahashi, K. *et al.* 11C-Cetozole: An Improved C-11C-Methylated PET Probe for Aromatase Imaging in the Brain. *J. Nucl. Med.* **55**, 852–857, <https://doi.org/10.2967/jnumed.113.131474> (2014).
30. Azcoitia, I., Arevalo, M. A. & Garcia-Segura, L. M. Neural-derived estradiol regulates brain plasticity. *J. Chem. Neuroanat.* **89**, 53–59, <https://doi.org/10.1016/j.jchemneu.2017.04.004> (2018).
31. Azcoitia, I. *et al.* Brain aromatase is neuroprotective. *J. Neurobiol.* **47**, 318–329 (2001).
32. Blakemore, J. & Naftolin, F. Aromatase: Contributions to Physiology and Disease in Women and Men. *Physiology (Bethesda)* **31**, 258–269, <https://doi.org/10.1152/physiol.00054.2015> (2016).
33. Ostlund, H., Keller, E. & Hurd, Y. L. Estrogen receptor gene expression in relation to neuropsychiatric disorders. *Ann. N. Y. Acad. Sci.* **1007**, 54–63 (2003).
34. Johansen-Berg, H. *et al.* Functional-anatomical validation and individual variation of diffusion tractography-based segmentation of the human thalamus. *Cereb. Cortex* **15**, 31–39, <https://doi.org/10.1093/cercor/bhh105> (2005).
35. Lephart, E. D. A review of brain aromatase cytochrome P450. *Brain research reviews* **22**, 1–26 (1996).
36. Biegon, A. *et al.* Nicotine blocks brain estrogen synthase (aromatase): *in vivo* positron emission tomography studies in female baboons. *Biol. Psychiatry* **67**, 774–777, <https://doi.org/10.1016/j.biopsych.2010.01.004> (2010).
37. Onaka, T. Neural pathways controlling central and peripheral oxytocin release during stress. *J. Neuroendocrinol.* **16**, 308–312, <https://doi.org/10.1111/j.0953-8194.2004.01186.x> (2004).
38. Acevedo-Rodriguez, A., Mani, S. K. & Handa, R. J. Oxytocin and Estrogen Receptor beta in the Brain: An Overview. *Frontiers in endocrinology* **6**, 160, <https://doi.org/10.3389/fendo.2015.00160> (2015).
39. Meyer-Lindenberg, A., Domes, G., Kirsch, P. & Heinrichs, M. Oxytocin and vasopressin in the human brain: social neuropeptides for translational medicine. *Nature reviews Neuroscience* **12**, 524–538, <https://doi.org/10.1038/nrn3044> (2011).
40. Gade-Andavolu, R. *et al.* Association between the estrogen receptor TA polymorphism and Harm avoidance. *Neurosci. Lett.* **467**, 155–158, <https://doi.org/10.1016/j.neulet.2009.10.028> (2009).
41. Eliot, L. The trouble with sex differences. *Neuron* **72**, 895–898, <https://doi.org/10.1016/j.neuron.2011.12.001> (2011).
42. Cosgrove, K. P., Mazure, C. M. & Staley, J. K. Evolving knowledge of sex differences in brain structure, function, and chemistry. *Biol. Psychiatry* **62**, 847–855, <https://doi.org/10.1016/j.biopsych.2007.03.001> (2007).
43. Bethea, C. L., Lu, N. Z., Gundlach, C. & Streicher, J. M. Diverse actions of ovarian steroids in the serotonin neural system. *Front. Neuroendocrinol.* **23**, 41–100 (2002).
44. Kolla, N. J. & Vinette, S. A. Monoamine Oxidase A in Antisocial Personality Disorder and Borderline Personality Disorder. *Curr Behav Neurosci Rep* **4**, 41–48, <https://doi.org/10.1007/s40473-017-0102-0> (2017).
45. Meyer, J. Novel Phenotypes Detectable with PET in Mood Disorders: Elevated Monoamine Oxidase A and Translocator Protein Level. *PET Clin* **12**, 361–371, <https://doi.org/10.1016/j.cpet.2017.02.008> (2017).
46. Logan, J. *et al.* Graphical analysis of reversible radioligand binding from time-activity measurements applied to [N-11C-methyl]-(-)-cocaine PET studies in human subjects. *J. Cereb. Blood Flow Metab.* **10**, 740–747, <https://doi.org/10.1038/jcbfm.1990.127> (1990).
47. Logan, J. *et al.* Distribution volume ratios without blood sampling from graphical analysis of PET data. *J. Cereb. Blood Flow Metab.* **16**, 834–840 (1996).
48. Yasuno, F. *et al.* Low dopamine d(2) receptor binding in subregions of the thalamus in schizophrenia. *Am. J. Psychiatry* **161**, 1016–1022 (2004).
49. Maldjian, J. A., Laurienti, P. J., Kraft, R. A. & Burdette, J. H. An automated method for neuroanatomic and cytoarchitectonic atlas-based interrogation of fMRI data sets. *Neuroimage* **19**, 1233–1239 (2003).

Acknowledgements

We thank Ms. Yumiko Katayama and Ms. Kanako Tajima of the RIKEN Center for Life Science Technologies and Dr. Naohiro Tsuyuguchi, Mr. Takashi Yamanaga, Mr. Hideki Kawahata, and Dr. Hisako Kobata of Osaka City University for their expert technical assistance and Prof. Sanae Fukuda of Kansai University of Welfare Sciences for her support in participant recruitment. This work was supported in part by a consignment expense for molecular imaging research programs entitled “Research Base for Exploring New Drugs” from the Japanese Ministry of Education, Culture, Sports, Science and Technology; and JSPS KAKENHI Grant Numbers 22791155, 25830024. No other potential conflicts of interest relevant to this article exist.

Author Contributions

Kayo T., T.H. and Y. Watanabe designed this research and wrote the paper. K.O., T.T., M.T., A.I., Y.N., S.T. and Y. Wada performed experiments and analysed the data. Kazuhiro T., H.D. provided critical advice and discussion. All authors commented on the manuscript.

Additional Information

Supplementary information accompanies this paper at <https://doi.org/10.1038/s41598-018-35065-4>.

Competing Interests: The authors declare no competing interests.

Publisher’s note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2018