

RESEARCH

Open Access

Tracing dynamic biological processes during phase transition

Tao Zeng¹, Luonan Chen^{1,2*}

From The 5th IEEE International Conference on Computational Systems Biology (ISB 2011)
Zhuhai, China. 02-04 September 2011

Abstract

Background: Phase transition widely exists in the biological world, such as transformation of cell cycle phases, cell differentiation stages, disease development, and so on. Such a nonlinear phenomenon is considered as the conversion of a biological system from one phenotype/state to another. Studies on the molecular mechanisms of biological phase transition have attracted much attention, in particular, on different genotypes (or expression variations) in a specific phase, but with less of focus on cascade changes of genes' functions (or system state) during the phase shift or transition process. However, it is a fundamental but important mission to trace the temporal characteristics of a biological system during a specific phase transition process, which can offer clues for understanding dynamic behaviors of living organisms.

Results: By overcoming the hurdles of traditional time segmentation and temporal biclustering methods, a causal process model (CPM) in the present work is proposed to study the biological phase transition in a systematic manner, i.e. first, we make gene-specific segmentation on time-course expression data by developing a new boundary gene estimation scheme, and then infer functional cascade dynamics by constructing a temporal block network. After the computational validation on synthetic data, CPM was used to analyze the well-known *Yeast* cell cycle data. It was found that the dynamics of the boundary genes are periodic and consistent with the phases of the cell cycle, and the temporal block network indeed demonstrates a meaningful cascade structure of the enriched biological functions. In addition, we further studied protein modules based on the temporal block network, which reflect temporal features in different cycles.

Conclusions: All of these results demonstrate that CPM is effective and efficient comparing to traditional methods, and is able to elucidate essential regulatory mechanism of a biological system even with complicated nonlinear phase transitions.

Introduction

In the biological world, a phase transition can be defined as the transformation of a biological system from one phenotype or state to another, where different phenotypes can be mapped to distinct states. For example, cell cycle is known to have four distinct phases: G1, S, G2 and M phases; cell differentiation contains different stages like cell proliferation, growth arrest and

mature differentiation; and cancer development mainly involves three steps as mutation, promotion and invasion. Obviously, analysing those biological phase transitions will offer valuable clues for understanding life and its dynamics. Therefore, a fundamental but important question is how to trace the temporal characteristics or dynamics of a biological system during a particular phase transition process.

The study on molecular mechanism of biological phase transition has attracted much attention [1-4]. For instance, by modulating the intracellular redox state and measuring cell cycle progression, the redox cycle within the (mammalian) mouse embryonic fibroblast cell cycle

* Correspondence: Inchen@sibs.ac.cn

¹Key Laboratory of Systems Biology, SIBS-Novo Nordisk Translational Research Centre for PreDiabetes, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Shanghai 200031, China
Full list of author information is available at the end of the article

was found to maintain the metabolic processes early in G1 and activate G1-regulatory proteins ahead of entry into S phase [1]. For a well known agricultural pest as migratory locust with a phase transition from the solitary to the gregarious, many down-regulated and some up-regulated genes were found in various organs when arriving to gregarious phase [2], which provides molecular indicators and recovers genetic mechanisms of phase transition in locusts. To determine the dormancy status of raspberry buds whose developmental regulation is helpful to promote the economic values of fruit and horticultural industries, a few significant dormancy-related candidate genes for raspberry buds had been identified by principal component analysis on clones' expressions [5]. Generally speaking, these research works are mainly on the different genotypes or expression variations at the level of individual genes under specific phases. Despite of those progresses, however, there is much less of focus on studying cascade changes or sequential dynamics of genes' or modules' functions at the level of networks during phase transition process.

As well known to us, one gene generally has multiple roles in biological processes but what role at a specific time is still unclear. Thus, identifying a gene functional group or module, which is composed of cooperative genes in biological processes or pathways, can reveal the functional specificity of individual genes or network modules. On the other hand, nowadays, there is rich information on biological processes [6,7], but the information on biological processes generally lacks dynamic features even compared with pathways [8,9]. Hence, in this paper we intend to identify the sequential structure or cascade dynamics of biological processes during phase transitions by developing a general framework for gene-specific segmentation and temporal block network (or network module), in particular on when and what a biological process or function will be cooperatively facilitated by network modules (or gene modules) during a phase transition. Note that, in the previous studies, the term "dynamic biological process" was usually used to refer to the dynamics of some general biological functional work-flow rather than sequential dynamics of biological processes or pathways [10-12]. In contrast, our work focuses on studying conditional and time-dependent behaviours or sequential dynamics of network modules, which are functionally enriched on specific biological processes [13].

The rapid accumulation of temporal gene expression data provides us the opportunity to unveil mechanisms of dynamic processes behind phenotype changes. In particular, a recent work shows that temporal dynamical model has ability to detect the presence and absence of stage/phase specific biological processes in *Yeast* cell cycle and metabolic cycle [13]. But, this model is limited

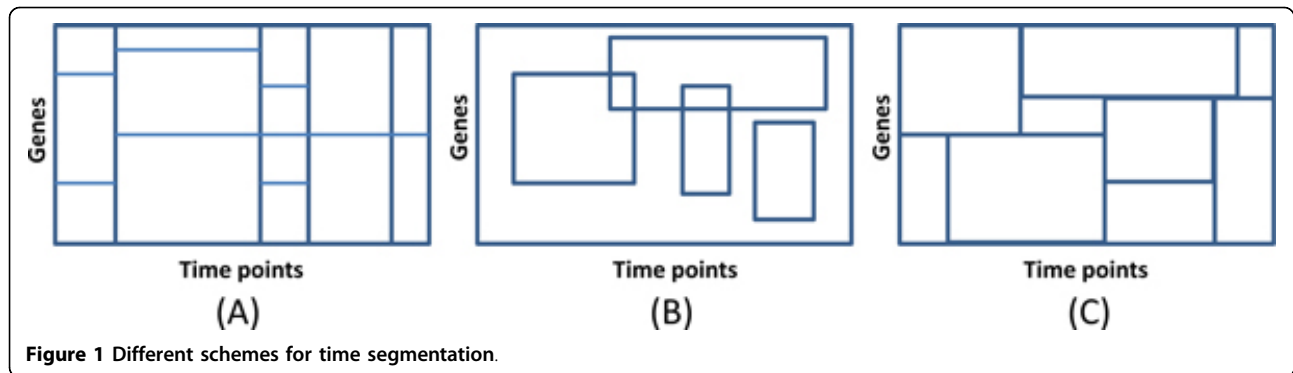
to the analysis on the time segmentation for all genes, by simply using the replicated observations to infer biological processes' temporal coordination. To overcome this problem, a new bicluster-based temporal segmentation method in this paper is developed to build a causal process model (CPM) for identifying the temporal features of biological processes during genotype or system reorganizations. In addition to biological processes and pathways, network modules or protein complexes [14] are used to further illustrate the sequential dynamics of biological systems as the molecular basis of those functional temporal features. Actually, protein modules or protein complexes have been found to play many important roles in biological phase changes, such as, indicator of genetic effect during mammary gland oncogenesis [15], marker of cancer diagnosis and prognosis [16], predictor of genotype-phenotype associations [17,18], and responder of dynamic cues from the environment [19].

In summary, the construction of our causal process model (CPM) includes three steps. First, we identify specific biclusters with linear patterns, and assemble them into temporal blocks representing a group of genes and their time segmentations. Then, each temporal block is refined by conducting functional enrichment analysis. Finally, we infer the sequential or cascade (causal) relations between temporal blocks by a graphical model (e. g., partial correlation) among two groups of genes. Through various experiments, we demonstrate the effect of our method on gene-specific temporal segmentation. In particular, on *Yeast* cell cycle data, we show that the phase division based on CPM is more efficient and effective than the segmentation based on traditional CCC-biclustering method [20]; and in the analysis of phase/cell cycle related biological processes, we found that the group of genes actually displays conditional functional enrichment and protein interaction network rewiring. All those results show that CPM is indeed able to unveil the biological mechanism behind complicated phase transitions.

Method

Causal process model: temporal block based on biclusters' assembler

Unlike traditional time segmentation methods requiring the same division on a time period for all genes [13] (see Figure 1 (A)), the gene-specific time segmentation is considered in the present work. That means, for different genes or gene groups, they can have different corresponding time segmentations based on their expressions, which can be considered as a general framework without the uniform division constraint. This is why the biclustering methodology [21,22] (see Figure 1 (B)), which can group genes and conditions



simultaneously, is adopted. However, as discussed in the study of temporal dynamic model [13], state-of-the-art CCC-biclustering method [20] has the limitation that it usually cannot cover all/most genes and time points. To overcome this problem, an in-house biclustering method (noted as **EBB**: **Error-Bounded Biclustering**) is used to enumerate so-called error-bounded linear patterns, e.g. traditional shifting pattern and scaling pattern [22], which can model a group of genes having similar expression change tendency, and further assemble them into the proposed temporal blocks by estimating the following boundary genes.

The brief framework of EBB includes three main steps: (1) discretizing the raw data matrix to a 0-1 matrix by a referred element in data matrix and a given error bound; (2) building a suffix tree based on 0-1 sequences encoded by rows in the above 0-1 matrix where '0' represents left child node and '1' represents right child node; (3) identifying the deepest right-only node in the suffix tree as a potential bicluster with error-bounded linear pattern. In fact, CCC-biclustering is also an exhaustive method [20], but it adopts a significant trend filtering to handle with the data pre-processing and thereby cannot guarantee to find all potential scaling patterns/linear patterns. This problem leads to loss of most low-signal patterns and some important expression patterns (e.g. linear patterns), which prohibit method itself to explore whole information of data. On the other hand, EBB method seeks linear patterns covering traditional shifting/scaling patterns [22] so that it can identify all interesting expression patterns in theory. Besides, EBB can also keep low-varying signals as many as possible because it uses the error bound but not the tendency bound to discrete the raw data.

As well known to us, biclusters represent similar expression behaviors of a group of genes at the same time points. However, our temporal block gathers those genes with the cooperative expression change during a specific time period, i.e. find those genes which simultaneously obtain or lose similar expression with their partner genes. Qualitatively, a temporal block is a sub-

matrix in the original data to cover the complete biclusters as many as possible but split the known biclusters as few as possible. According to the following concepts and definitions, the genes on so-called temporal boundary are used to divide the whole data matrix into different matrices named as temporal blocks (see Figure 1 (C)).

Definition 1 (Boundary gene and set) Given a data matrix $D = \{d_{m,n}\}_{m \in I, n \in J}$, let a set of gene expression patterns as biclusters $\{P_i = \{(G_i, T_i) | G_i \subseteq I, T_i \subseteq J\}\}_{i=1}^K$. Then, a gene g in I is on the temporal boundary at time point t in J only when its R value is larger than a given threshold θ with default value as one, where R is calculated as formula (1). And all boundary genes at every time point consist of a boundary set $\{BG(t) = \{g | R(g, t) > \theta, g \in I\}\}_{t \in J}$.

$$R(g, t) = \frac{| \{T_i | g \in G_i, t = \min_{\tau \in T_i} \tau\} | \max(1, | \{\tau | \tau \in J, \tau < t\} |)}{| \{T_i | g \in G_i, t \in T_i, t \neq \min_{\tau \in T_i} \tau\} |} \quad (1)$$

Definition 2 (Temporal block) Given a matrix data $D = \{d_{m,n}\}_{m \in I, n \in J}$ and its boundary set BG , the temporal block $B_i = \{(G_i, T_i) | G_i \subseteq I, T_i \subseteq J\}$ should satisfy following conditions:

- (a) $\forall g \in G_i, g \in BG(\min_{\tau \in T_i} \tau)$
- (b) $\forall g \in G_i, g \in I - BG(\min_{\tau \in T_i} \tau - 1)$ or $\min_{\tau \in T_i} \tau = \min_{\tau \in J} \tau$
- (c) $\forall g \in G_i, g \in I - BG(\max_{\tau \in T_i} \tau)$ or $\max_{\tau \in T_i} \tau = \max_{\tau \in J} \tau$
- (d) $\forall g \in G_i, g \in BG(\max_{\tau \in T_i} \tau + 1)$ or $\max_{\tau \in T_i} \tau = \max_{\tau \in J} \tau$
- (e) $\forall G \subseteq G_i, T \subset T_i, (G, T)$ does not satisfy conditions (a)-(d);
- (f) $\forall G \subseteq I - G_i, T = T_i, (G, T)$ does not satisfy conditions (a)-(d).

For convenience, $\min_{\tau \in T_i} \tau$ points the starting point or left-end of temporal block and $\max_{\tau \in T_i} \tau$ points the ending point or right-end of temporal block, which are similar for temporal bicluster. Some additional differences between the proposed temporal block and traditional bicluster will be discussed in the next section.

Causal process model: expansion of temporal block for functional enrichment analysis

Like temporal segmentation, CPM gives a non-overlapping division on the whole data. It means that one gene within one time point at most belongs to one temporal block although this gene can belong to a different temporal block but at a different time, i.e. one temporal block cannot cover any other one in CPM. Taking Figure 2 as an example, six genes $\{g_1, g_2, g_3, g_4, g_5, g_6\}$ might have coherent expression on time points $\{t_3, t_4, t_5, t_6\}$. In order to reflect the different gene reorganization events happening on time points t_2 and t_3 , these genes are divided into two temporal blocks during the co-expression period. This is just the over-division phenomenon in biclustering study which can supply a multi-granularity model for overlapping patterns [23]. When analyzing functional enrichment on temporal blocks, the over-divided genes should be gathered again. This can be easily achieved by the expansion of temporal blocks.

Definition 3 (Expanded temporal block) Given a data matrix $D = \{d_{m,n}\}_{m \in I, n \in J}$ and its temporal block $B_i = \{G_i, T_i | G_i \subseteq I, T_i \subseteq J\}$, the corresponding expanded temporal block $B_i = \{G_i, T_i | G_i \subseteq I, T_i \subseteq J\}$ satisfies: $B_i^* = \{G_i^*, T_i^* | G_i^* \subseteq I, G_i^* \supseteq G_i, T_i^* = T_i\}$. Where, $C_{x,y}$ represents the Pearson coefficient correlation between expression profiles of two genes during the time period T_i^* , and p is a threshold with a default value as 0.8.

Therefore, the temporal blocks are useful to give a global scheme of the data division, and the expanded temporal blocks are suitable to reflect the local property of large data.

Causal process model: temporal block network construction based on partial correlation

In order to extract the cascade dynamics of temporal blocks representing the sequential order of biological processes, there is a need to build a directed network

among different temporal blocks whose qualitative connections are evaluated by the partial correlation [24]. It should be emphasized that, at present, our model concerns the linear relationship (i.e., linear pattern in temporal bicluster) so that the correlation but not mutual information is considered in relationship measurement. And to infer direct but not indirect correlation among genes, we adopted the partial correlation to measure association between two genes by removing the effect of their controlling genes.

Definition 4 (Partial correlation) Given three gene expression profiles or vectors X, Y and Z , the partial correlation between X and Y under condition Z is calculated as:

$$PR(X, Y|Z) = \frac{C_{X,Y} - C_{X,Z}C_{Y,Z}}{\sqrt{1 - C_{X,Z}^2}\sqrt{1 - C_{Y,Z}^2}} \quad (2)$$

where $C_{.,.}$ represents the Pearson coefficient correlation.

Definition 5 (Link strength between temporal blocks) Given two temporal blocks $B_1 = (G_1, T_1)$ and $B_2 = (G_2, T_2)$, if $\min_{\tau \in T_1} \tau \leq \min_{\tau \in T_2} \tau \leq \max_{\tau \in T_1} \tau + 1$, these two blocks have a link with direction from B_1 to B_2 . The link strength between their referred gene expression profiles in the time period $[\min_{\tau \in T_1} \tau, \min(\max_{\tau \in T_1} \tau, \max_{\tau \in T_2} \tau)]$ can be calculated as:

$$LS(B_1, B_2) = \frac{\sum_{X \in G_1} \max_{Y \in G_2} (\min_{Z \in G_2, Z \neq X, Y} |PR(X, Y|Z)|)}{|G_1|} \quad (3)$$

This strength measurement indicates the potential partial relation from genes in a source block B_1 to genes in a target block B_2 . It requires that the gene X in a source can directly interact with gene Y in a target (the correlation between X and Y is maximal as shown in the above definition), or be indirectly related to Y without the conduction from other target genes (the minimal partial correlation between X and Y under the

	t1	t2	t3	t4	t5	t6	t7
g1	0.1	1.1	1.2	0.5	0.4	0.6	1.2
g2	0	1.5	1.3	0.7	0.5	0.2	1.3
g3	0.5	1.4	1.1	0.3	0	0	1.2
g4	0.4	1.4	0.8	0.3	0.3	0	1.5
g5	0.2	0	1.2	0.5	0.3	0.1	1.1
g6	0.3	0	1.1	0.5	0.4	0.3	1.4

Figure 2 Illustration of temporal blocks based on the estimated boundary genes.

control of any Z is maximal as shown in above definition). When the link strength is larger than a threshold with default value as 0.9, the connected temporal blocks are thought to have significant causal relation.

Based on the links (edges) with strengths (weights) among temporal blocks (nodes), the temporal block network (TBN) is constructed for deep analysis on dynamic biological processes. And the execution program (CPM) for temporal blocks can be accessed from <http://www.sysbio.ac.cn/cb/chenlab/software.htm>.

Result and discussion

There are different characteristics between the proposed temporal blocks and traditional biclusters. Due to the module-in-focus property of biclustering, biclusters always have overlap with each other and have less size (i.e., in terms of clusters) than the original data [20]. The redundancy elimination of those overlapped biclusters is still a relevant and open question in the study of biclustering. On the other hand, in the present work, CPM suffers from few effects of potential bicluster redundancy according to the principles of temporal block construction. In order to divide original time course data, the temporal blocks instead of biclusters are used to build the dynamic model constructed by boundary gene estimation so that any temporal block is not a traditional bicluster pattern but a bicluster assembler. In other words, a temporal block does not represent the coherent expression solely as a bicluster but represents the similar expression pattern change events (condition (a) in Definition 2) as the concept of gene reorganization across the neighbouring time windows [13]. With the conditions (b), (c) and (d) in Definition 2, a temporal block can tolerate the so-called disorder period, thereby allowing the boundary genes to present at consecutive time points located at left-end of temporal block. It can also allow the so-called asynchronous ending period, i.e. allow those genes not on temporal boundary when they present at right-end of temporal block or even allow them not belonging to any original bicluster pattern. Besides, temporal blocks also have completeness guaranteed by Conditions (e) and (f) in Definition 2. These advantages of temporal blocks all let them reasonably represent the non-overlapped sub-regions of the original whole data.

For instance, in the matrix (with synthetic R values) of above Figure 2, an element in red representing its gene (row) is on the temporal boundary at its time point (column); an element in blue means that its gene is not on the temporal boundary but at the starting time point of a few biclusters; an element in orange points that its gene is not at the starting time point of any biclusters yet. Therefore, the temporal block $\{(g_5, g_6), (t_3, t_4, t_5, t_6)\}$ is one without either disorder period or asynchronous

ending period, while the temporal block $\{(g_1, g_2, g_3, g_4), (t_2, t_3, t_4, t_5, t_6)\}$ covers a disorder period because genes (g_1, g_2, g_3) are at time points (t_2, t_3) and an asynchronous ending period for genes (g_3, g_4) being at time points (t_5, t_6) .

Furthermore, the time cost of CPM is mainly on the computation of temporal block construction by temporal bicluster mining, which is similar to CCC-biclustering with a polynomial time complexity [20].

Gene-specific temporal segmentation by CPM shown on synthetic data

First of all, we analyzed CPM on a synthetic data in a simple but typical strategy adopted in the previous studies [23]. We produced a random data matrix with 10 rows and 15 columns. Five predefined blocks or patterns with five genes and four consecutive time points were embedded into such a matrix. As the recovering patterns in the above synthetic data are perfect, we used a strict error bound as 0.0001 and minimum bicluster size as $3 \times 3, 3 \times 4, 4 \times 3, 4 \times 4$ respectively to run CPM method (hereafter, the annotation $x \times y$ means that one bicluster contains at least x genes and y time points). Under different parameter settings, the divisions with temporal blocks on the whole synthetic data are shown in Figure 3, where one temporal block is surrounded by a yellow box. We should emphasize two points on these results. One is, for the effect of minimum bicluster size setting, the biclusters with a shorter time period will lead to more sub-blocks due to over-division (3×3 in Figure 3 (A) and 4×3 in Figure 3 (C)) than those with a longer time period (3×4 in Figure 3 (B) and 4×4 in Figure 3 (D)), but all blocks are still reasonable and acceptable. The other is, according to the proposed design principles, each temporal block can cover all time points of a predefined pattern and some asynchronous ending period (e.g. cases shown in Figure 3), in order to tolerate the noise/error and divide the whole data in a unified way. Totally, CPM can simultaneously group genes and find gene-specific time divisions, which cannot usually be obtained by traditional time segmentation methods, and it can further split the whole data matrix into different sub-matrices, which is disregarded in many previous biclustering studies.

Phase description by CPM comparing with CCC-biclustering based method

Then, we analyzed CPM for the *Yeast Cell Cycle of α -factor synchronization experiment of Spellman *et al.** [25]. This dataset comprises two cell cycles, with each cell cycle containing three phases as M/G1, G1&S, and G2&M [13,25]. Every phase crosses three time points in the experiment with a constant time interval as 7 minutes. After using one-way ANOVA [26] to select genes

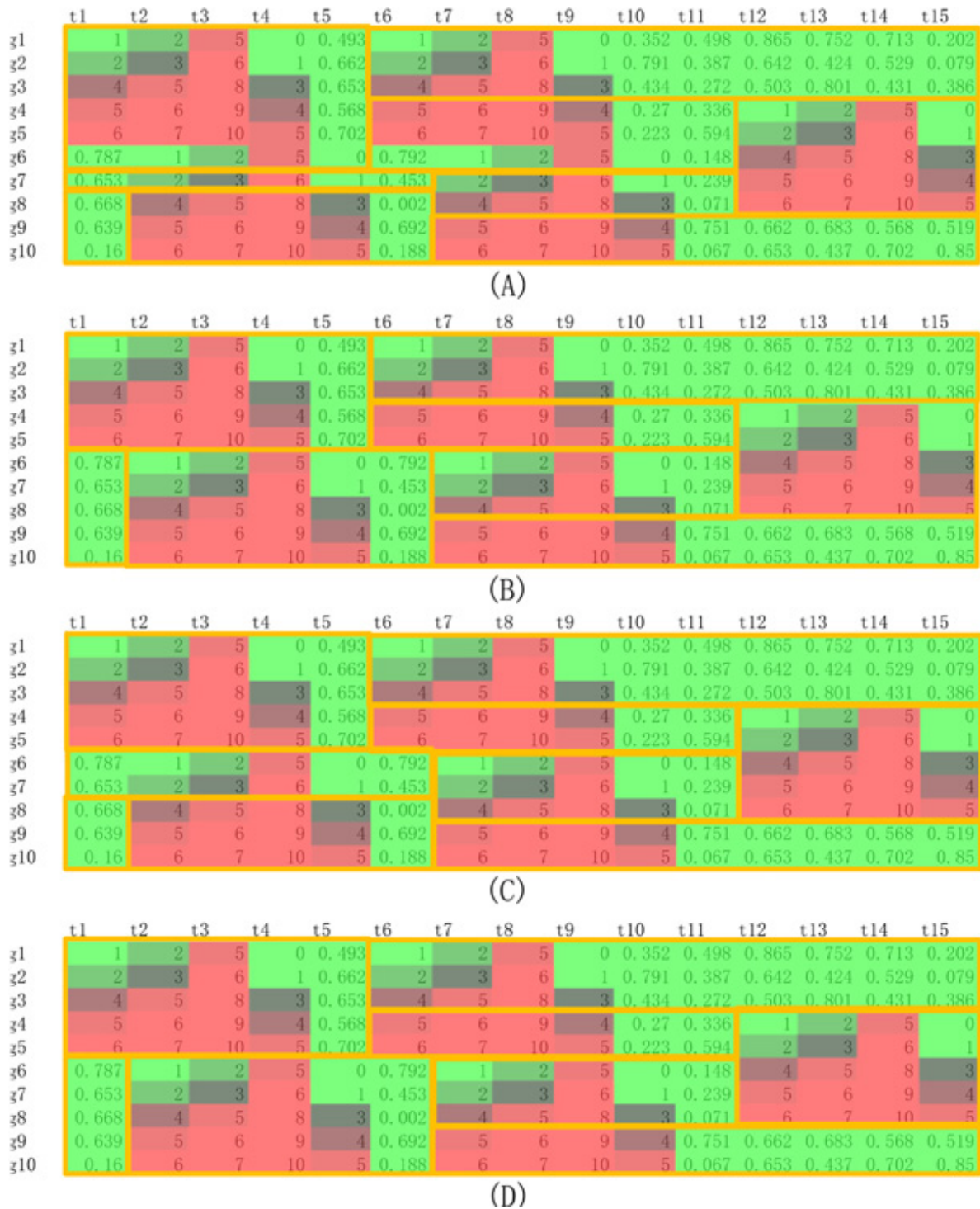


Figure 3 Temporal blocks on synthetic data according to CPM with different parameter settings.

(i.e. setting the number of sample (time point) groups to be six with prior knowledge in six phases of two cell cycles, and the P -value to be based on the F-distribution with significant threshold as 0.05), remaining data

denoted as YCC with 730 genes and 18 time points was used for further analysis. Again, we used different error bounds in $\{0.05, 0.1, 0.15, 0.2, 0.25\}$ and minimum bicluster size as 10^5 (experience values in previous

study) to build CPMs on YCC data for extensive evaluations.

As described before, the boundary genes can be used to trace the role-change events of a group of genes, and their number would increase greatly at a time point around the alternation of phases [13]. Due to the need to cover the possible disorder period, a few boundary genes are not effective on the temporal block construction and others are just the refined boundary genes locating at the left-end (starting time point) of final temporal blocks. According to the statistic of temporal blocks and their depending boundary genes, Figure 4 shows two kinds of distributions of boundary gene numbers under different CPM parameter settings, where the dotted line represents the distribution of the original boundary genes and the solid line represents the distribution of the refined boundary genes. Obviously, the distributions of numbers of the refined boundary genes unveil more convincing phase related characteristics than those of the original boundary genes, thereby confirming the effectiveness of the temporal blocks. Note

that boundary genes mean the refined ones in the following discussions. When the error bound is strictly set to 0.05, the peaks of distributions of boundary genes are always located at the middle time of each phase because genes try to keep their status of steady coordination (note that, the strictest parameter setting as 0.01 results in no bicluster output). When error bound is set to 0.1, the peaks of distributions of boundary genes are always located at the time point of a phase transition because genes usually start to cooperatively facilitate functions at this time and temporal block can cover the potential beginning disorder period. On the other hand, when an error bound is set to 0.15 or even a larger value, distributions of boundary genes cannot keep on their correlations with phases because many noises are introduced to mix up the genes on and not on temporal boundaries. Therefore, CPM can directly use the distributions of boundary genes to trace the critical time points of phase transitions, whose dependent parameter setting will be estimated from both experience of data analyzers and pattern quality evaluation of biclustering.

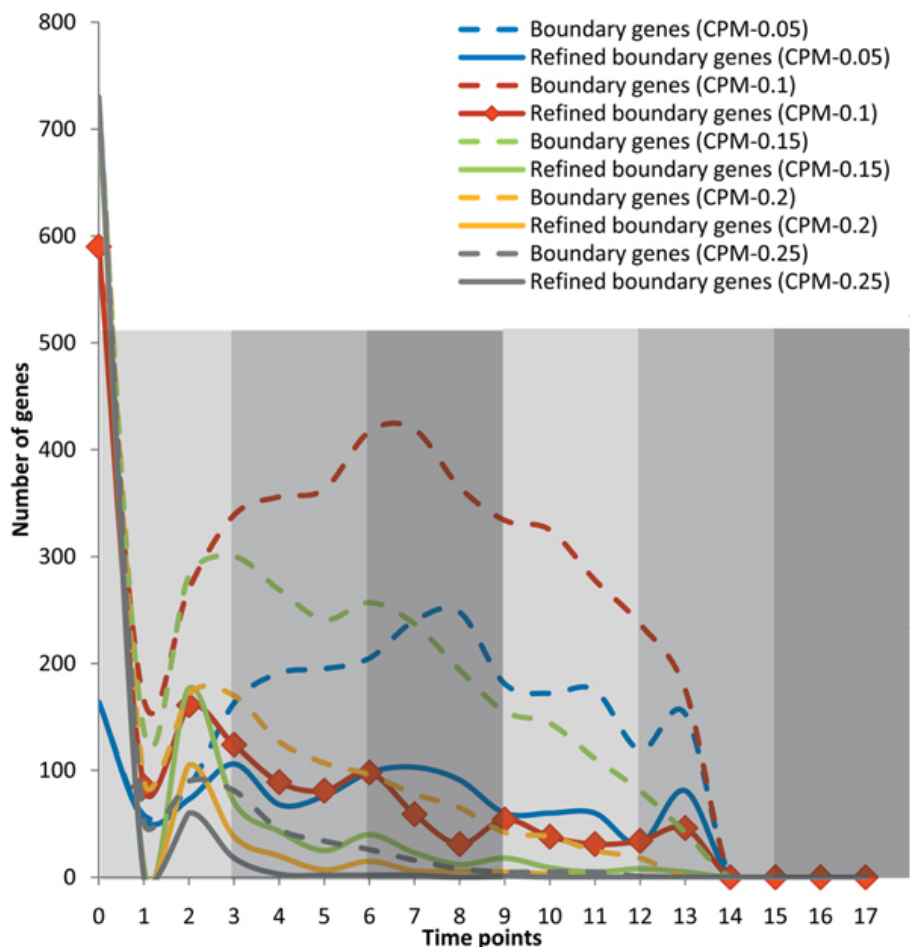


Figure 4 Statistic view of boundary genes by CPM with different parameter settings.

In order to further confirm the efficiency of the proposed (EBB) bicluster-based segmentation method comparing with other bicluster-based methods, we used temporal biclusters produced by CCC-biclustering [20] (under five different parameter settings and with 1.0 as the default value) to assemble temporal blocks again and re-analyzed the relations between developmental stages and distribution of boundary genes. Compared with Figure 4, the results shown in Figure 5 illustrate that CPM is more suitable on phase description than traditional temporal biclustering based segmentation. The further discussion on the differences between bicluster-based segmentation and traditional temporal segmentation is beyond the scope of this paper because they actually belong to two distinct methodology categories like biclustering and clustering.

Temporal trace identification by CPM with functional enrichment analysis

According to the above discussion on parameter setting, we chose the temporal blocks obtained with the most suitable error bound setting as 0.1 to conduct the following functional enrichment analysis [27]. In the temporal block expansion and temporal block network construction, the default thresholds were all used for calculation.

Biological processes during phase transition revealed by CPM and comparison with temporal dynamical model

Due to minimum length requirement of bicluster, the last four time points were not divided in our experiments. That is why we investigated the biological processes enriched in temporal blocks corresponding to the first phase and the latter two phases in each cell

cycle, to compare with temporal dynamical model [13]. Similar to the previous studies, the circular presence and absence of some biological processes in two cell cycles are shown in a chart as Figure 6. The obtained biological processes are close to those identified by temporal dynamic model, such as amino acid biosynthetic process, cell wall chitin biosynthetic process, chromosome condensation and nucleosome assembly [13]. Therefore, CPM indeed can reveal biological processes related to phase transitions, by analyzing the phase segmentation and the temporal block network.

It is worth noting that the potential causal relation between temporal blocks in CPM can further strengthen the cascade relation of phases belonging to intra- or inter-cell cycles. Figure 7 displays the whole temporal block network (where the edges between temporal blocks with same starting points were omitted so as to focus on the major asynchronous temporal relation), in which the nodes represent different temporal blocks denoted as $\{TB_k\}$; the direct edges represent causal relations; the node label shows the id of temporal block k and its time segmentation $[f, t]$ in the form as " $k - [f, t]$ "; and green, blue, yellow and pink nodes mean phase related, cell cycle related, cross-phases related and other kinds of temporal blocks, respectively.

- Obviously, there are direct edges linking temporal block TB_{63} to TB_{77} , and temporal block TB_{21} to TB_{41} . They are actually the phases' relations belonging to intra-cell cycle, which further confirm the phase related biological processes shown in the above Figure 6.

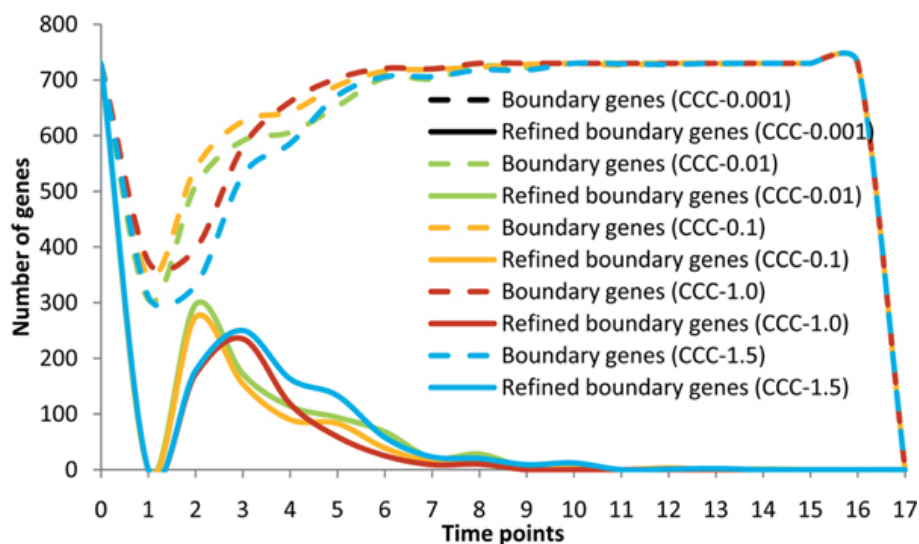


Figure 5 Statistic view of boundary genes by CCC-biclustering based method with different parameter settings.

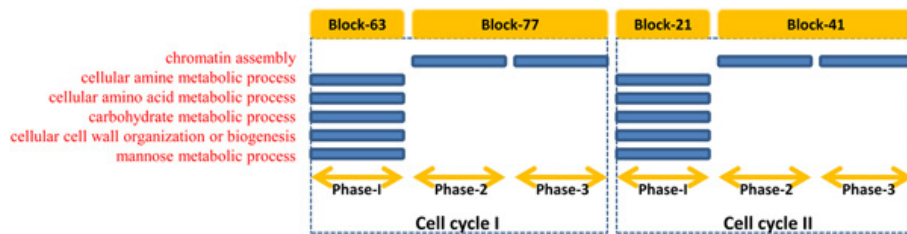


Figure 6 Biological processes with potential circular behaviour enriched in phase related temporal blocks.

- In all temporal blocks, only TB_{82} directly connects TB_{77} and TB_{21} , thereby acting as a bridge of (expression) correlation between the last phases of the first cell cycle and the initial phase of the latter cell cycle. This means that CPM can also identify the phases' relations belonging to inter-cell cycle, and has the ability to infer cascade dynamics of biological functions like biological processes across multiple cell cycles. Note that, the previous temporal dynamic model needs multiple datasets to deduce causal relation between biological processes [13], however, our CPM can infer meaningful functional cascade dynamics during biological transitions even on single dataset. At present, it is actually difficult to deeply discuss the biological processes not starting at a "check point" of some phase or cell cycle due to lack of relevant biological data, however, a few processes like protein-DNA complex assembly or nucleosome assembly enriched in temporal block TB_{82} suggest that some of those functions will hold before entering the next phase or cell cycle.

- As the temporal dynamical model strongly shows the similarity of two cell cycles after α -factor handling [13], CPM can even be used to elucidate the specificities for cell cycle related temporal blocks

TB_{39} and TB_{20} in Figure 7. These two cell cycle related temporal blocks (note that their functional analysis will be discussed in detail in next subsections) have not direct edges between themselves, but they can also be directly connected by temporal block TB_{82} again. This supports the need and importance of novel temporal blocks across neighbouring functional periods which are modelled by the gene-specific temporal segmentation integrated in CPM.

Functional enrichment variance during continuous cell cycles after α -factor treatment

The 1_{st} cell cycle related temporal block TB_{39} covers the former three phases with time points 0-8 and has 12 genes expanded to 432 ones. On the other hand, the 2_{nd} cell cycle related temporal block TB_{20} covers the latter three phases with time points 9-17 and has 42 genes expanded to 400 ones. For those two expanded gene sets, the significant phase (cell cycle)-related biological processes and pathways are listed in Table 1 and 2. Obviously, the 1_{st} cell cycle related genes and 2_{nd} cell cycle related genes have shown several different biological processes annotated in GO [28], and the 1_{st} cell cycle related genes are frequently observed in biological pathways annotated in KEGG and Reactome [29,30]. Therefore such two cell cycles after α -factor treatment

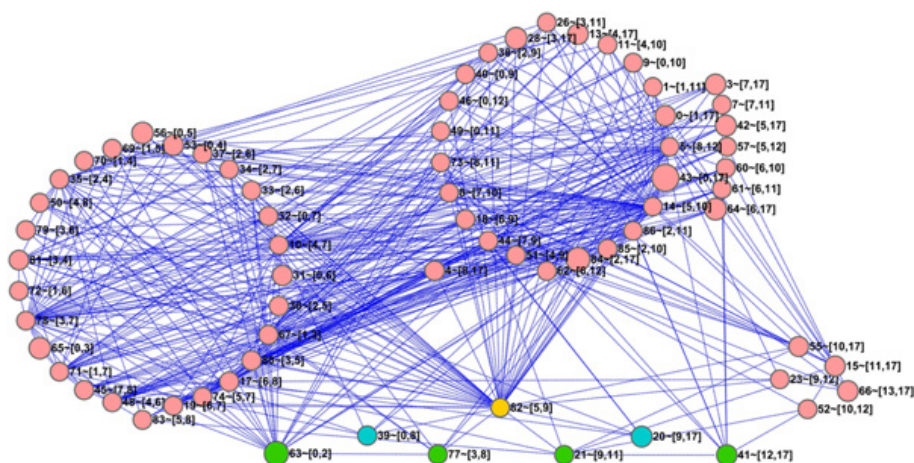


Figure 7 Temporal block network on YCC dataset.

Table 1 Biological processes enriched in two cell cycles according to genes in TB_{39} and TB_{20}

Biological process	cell 1 _{st}	cycle 2 _{nd}
mannose metabolic process	✓	
external encapsulating structure organization	✓	✓
cell wall organization or biogenesis	✓	✓
cell wall organization	✓	✓
cellular cell wall organization or biogenesis	✓	✓
cellular cell wall organization	✓	✓
cytokinetic cell separation	✓	✓
cytokinesis, completion of separation	✓	✓
cytokinesis	✓	✓
transition metal ion transport	✓	
iron ion transport	✓	
chromatin assembly	✓	✓
nucleosome assembly	✓	✓
DNA conformation change		✓
DNA packaging		✓
chromatin assembly or disassembly		✓

can be just thought as two super-phases with distinct dynamical properties, which is helpful to understand the cascade dynamics of complicated biological procedures across multiple phases or cycles.

In addition, in order to re-validate the cell cycle specificity on gene expression of such two temporal blocks, we used the genes in them to conduct hierarchical clustering with appropriate distance measurements [31] respectively on our analyzed dataset and other three independent *Yeast* gene expression datasets which also cover two cell cycles after the α -factor handling. They were downloaded from NCBI GEO with id GDS2318 [32] (one contributed dataset is denoted as YCC-gds2318) and GSE4987 [33] (two contributed datasets as dye-swap technical replicates are denoted as YCC-gse4987-53 and YCC-gse4987-35). On these four datasets YCC (Figure 8 (A)), YCC-gds2318 (Figure 8 (B)), YCC-gse4987-53 (Figure 8 (C)) and YCC-gse4987-35 (Figure 8 (D)) respectively, the genes from TB_{39} can correctly classify almost all time points into two cell cycles disregarding the effect of potential circular expression profiles in cell cycles. According to

Table 2 Biological pathways enriched in two cell cycles according to genes in TB_{39} and TB_{20}

Pathway	cell 1 _{st}	cycle 2 _{nd}
Amino sugar and nucleotide sugar metabolism	✓	✓
Steroid biosynthesis	✓	
Fructose and mannose metabolism	✓	
Regulation of beta-cell development	✓	
Regulation of gene expression in beta cells	✓	

Figure 9, genes from TB_{20} also have good performance on clustering time points from different cell cycles. Considering the existence of missing expressions (filled with zero) of genes in other independent datasets, we only analyzed the molecular network behind such cell cycle specificity on our main YCC dataset in next subsection.

Relation among modules and complexes in protein interaction network rewiring and temporal trace of biological phase transitions

The co-expression network [34] was also used to reflect the potential cell cycle specificity after α -factor treatment through the rewired structures of the protein interaction network (PIN). Given a cell cycle related temporal block $TB(G, T)$, we had a group of genes G and obtained the interactions of these genes' encoding proteins from STRING database [35]; with the information of *Yeast* protein subcellular localization [36] denoted as Yeast-eSLDB, we filtered the interaction by requiring that two proteins involved in one interaction must have a same candidate subcellular localization (this is because one protein may move to several subcellular localizations, and we only consider the location as "Nucleus", which has the most known protein members); based on these co-localization proteins' expression profiles in different cell cycles $\{T_i\}_{i=1,2}$ (for some i , $T = T_i$), we calculated the Pearson coefficient correlation of two proteins with an interaction; combining the proteins and interactions with weights (or correlations), we extracted a PIN conducted co-expression network (PCCN).

Thus, we used the genes in TB_{39} and TB_{20} with their expression profiles during two cell cycles to build four PCCNs. They are denoted as $\{N_i^c\}_{i \in \{1,2\}, c \in \{1,2\}}$, which mean that the genes/proteins in i cell cycle related temporal block have a rewired PCCN in actual c cell cycle. Figure 10(A)-(D) displays N_1^1 , N_1^2 , N_2^1 and N_2^2 respectively. As the above discussion, N_1^1 and N_2^2 should indeed have specific network characteristic corresponding to cell cycles. Generally, the genes represented by nodes in light blue belong to TB_{39} ; the genes represented by nodes in dark blue belong to TB_{20} ; while genes represented by nodes in blue belong to the overlap of such two cell cycle related temporal blocks. Each interaction edge becomes from light & thin to dark & thick when its absolute weight (or correlation) increases. By network visualization of Cytoscape [37], we easily observe the approximate network modules C_2 and C_3 in the Figure 10. The largest protein complex *Nucleosomal protein complex* extracted from the information of *Yeast* protein complexes [38] denoted as Yeast-CYC is also highlighted as another module C_1 . It is interesting that three different

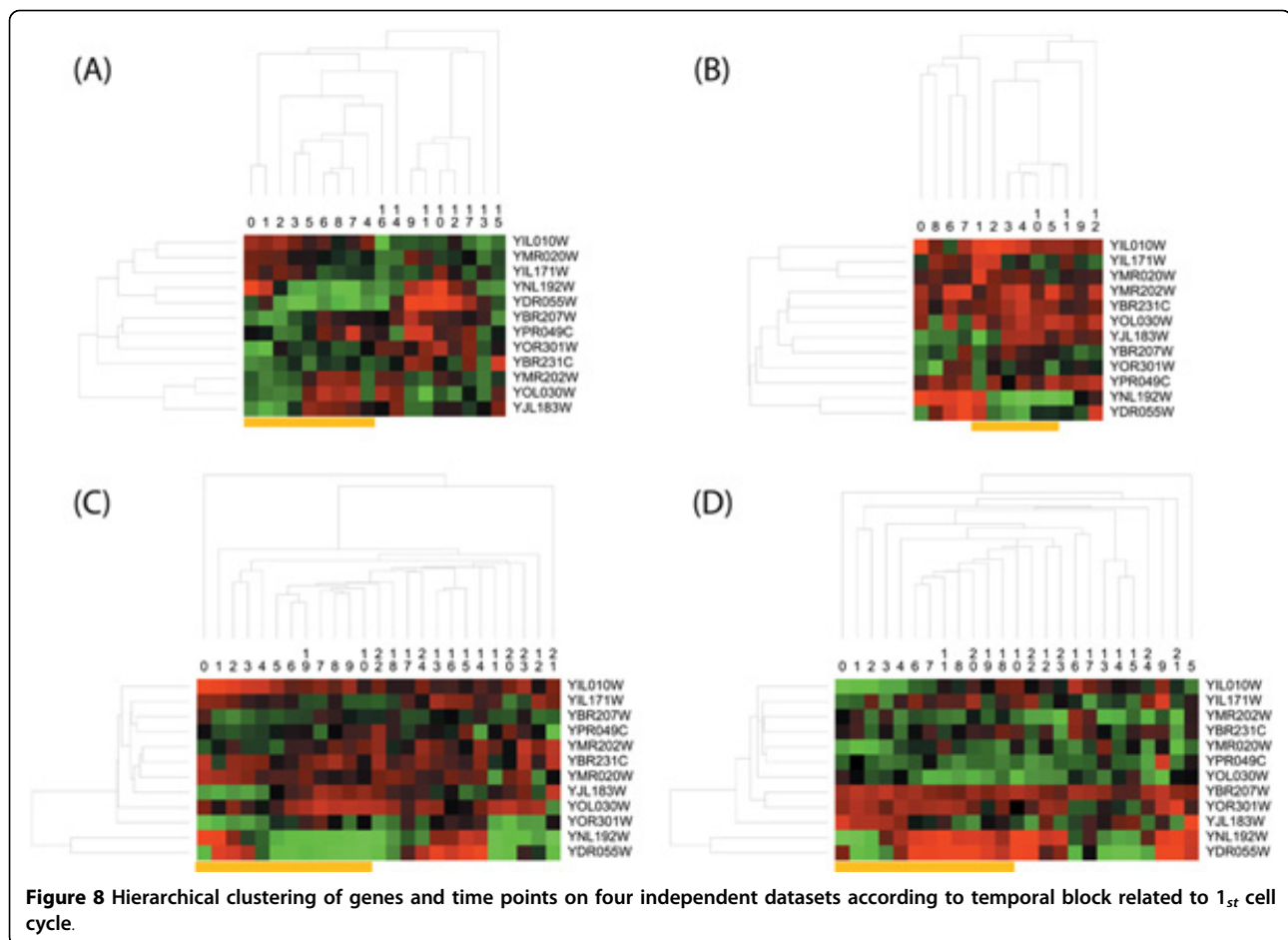


Figure 8 Hierarchical clustering of genes and time points on four independent datasets according to temporal block related to 1st cell cycle.

changes of network rewired profile correspond to the specificities of proteins in cell cycle related temporal blocks.

- For proteins in TB_{39} , they are densely connected to module C_3 in just the first cell cycle but not the second one; while C_3 always has fewer contacts with proteins in TB_{20} so that the presence and absence of relation with module C_3 would be a temporal trace for functional specificity in the first cell cycle.
- For proteins in TB_{39} or TB_{20} , they present strict interactions with module C_2 in the first cell cycle but lose such relation in following cell cycle. This means, in our mathematical model, TB_{39} mainly captures the presence of relation with C_2 while TB_{20} tends to mine the disappearance of relation with the same module.
- Dissimilar from the above two conditions, protein complex C_1 strengthens its relation with proteins in TB_{20} in just the second cell cycle but not the first one. Hence, the varying relation with protein

complex C_1 can be a candidate temporal trace for functional specificity in the second cell cycle.

Therefore, attractively, protein interaction modules and their relations with other proteins above can be thought as the dynamical markers (or temporal traces) of cell cycles in phase transitions. The proposed temporal blocks with the causal process model are indeed effective to efficiently uncover such molecular basis of a biological transition.

Conclusion

To overcome the drawbacks of traditional time segmentation and temporal biclustering methods, the causal process model (CPM) was proposed to study the biological phase transitions in a systematic way. The experimental results validated that CPM can effectively identify gene-specific temporal segmentations by developing a boundary gene estimation scheme, and efficiently infer the potential cascade dynamics of biological processes by constructing a temporal block network. CPM not only has identified the phase-specific dynamic

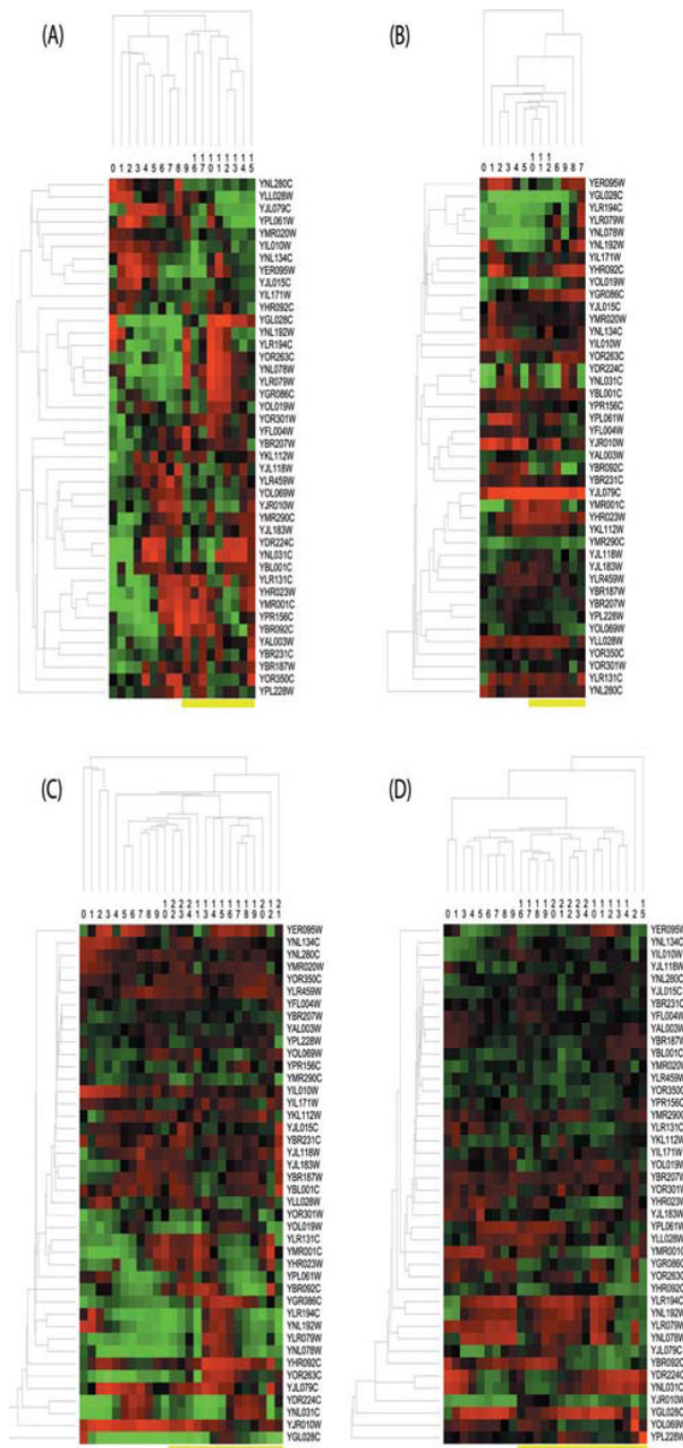


Figure 9 Hierarchical clustering of genes and time points on four independent datasets according to temporal block related to 2nd cell cycle.

biological processes which were found by the traditional dynamic temporal model, but also revealed cell cycle specific rewiring of the protein interaction network which was missed in the previous studies. All in all,

along with the improvement of bicluster enumeration and sparse causal network inference, the proposed CPM can both detect unknown phase transitions in real biological systems, and identify the candidate functional

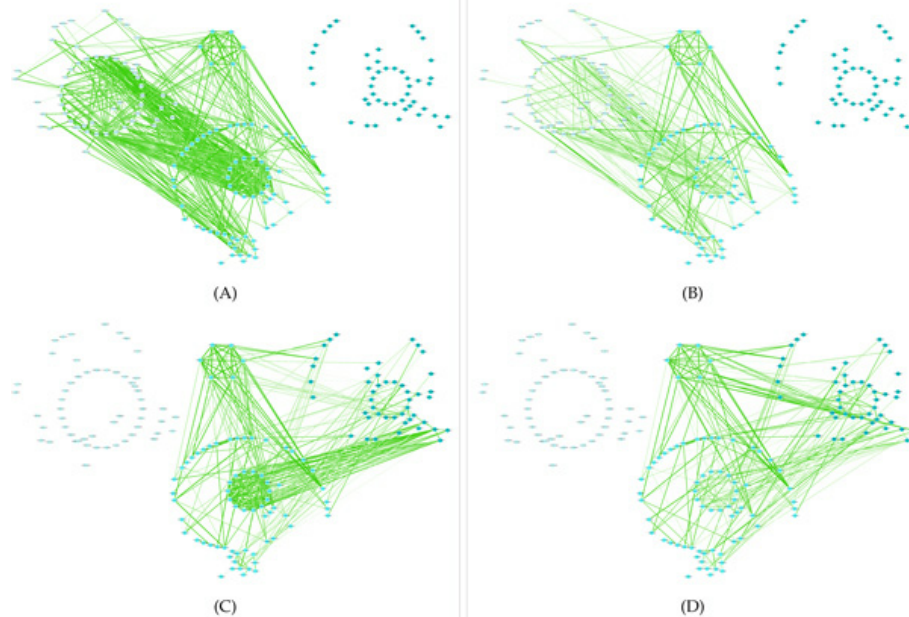


Figure 10 PIN conducted co-expression networks related to genes and cell cycles corresponding to cell cycle related temporal blocks.

cascade dynamics with temporal traces (or dynamical markers) during the transformation of a biological system.

Acknowledgements

Authors were supported by NSFC under Grant No. 91029301, No. 61134013 and No. 61072149. This work was supported by a fund from National Center for Mathematics and Interdisciplinary Sciences of CAS, the Knowledge Innovation Program of CAS with Grant No. KSCX2-EW-R-01, Shanghai Pujiang Program, and China Postdoctoral Science Foundation Funded Project under Grant No. 20110490757. This research was partially supported by the Chief Scientist Program of SIBS of CAS with Grant No. 2009CSP002, and the FIRST program (Aihara Project) from JSPS initiated by CSTP. This article has been published as part of *BMC Systems Biology* Volume 6 Supplement 1, 2012: Selected articles from The 5th IEEE International Conference on Systems Biology (ISB 2011). The full contents of the supplement are available online at <http://www.biomedcentral.com/bmcsystbiol/supplements/6/S1>.

Author details

¹Key Laboratory of Systems Biology, SIBS-Novo Nordisk Translational Research Centre for PreDiabetes, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Shanghai 200031, China. ²Collaborative Research Center for Innovative Mathematical Modelling, Institute of Industrial Science, University of Tokyo, Tokyo 153-8505, Japan.

Authors' contributions

TZ and LC conceived the research. TZ performed the study. LC supervised the project. TZ drafted a version of the manuscript. All authors wrote and approved the manuscript.

Competing interests

The authors declare that they have no competing interests.

Published: 16 July 2012

References

1. Menon SG, Sarsour EH, Spitz DR, Higashikubo R, Sturm M, Zhang H, Goswami PC: **Redox Regulation of the G1 to S Phase Transition in the Mouse Embryo Fibroblast Cell Cycle.** *Cancer Res* 2003, **63**(9):2109-2117.
2. Kang L, Chen X, Zhou Y, Liu B, Zheng W, Li R, Wang J, Yu J: **The analysis of large-scale gene expression correlated to the phase changes of the migratory locust.** *Proc Natl Acad Sci USA* 2004, **101**(51):17611-17615.
3. Chen L, Liu R, Liu Z, Li M, Aihara K: **Detecting early-warning signals for sudden deterioration of complex diseases by dynamical network biomarkers.** *Scientific Reports* 2012, **2**:342.
4. He D, Liu ZP, Chen L: **Coexpression network analysis in chronic hepatitis B and C hepatic lesion reveals distinct patterns of disease progression to hepatocellular carcinoma.** *Journal of Molecular Cell Biology* 2012, **4**:140-152.
5. Mazzitelli L, Hancock RD, Haupt S, Walker PG, Pont SDA, McNicol J, Cardle L, Morris J, Viola R, Brennan R, Hedley PE, Taylor MA: **Co-ordinated gene expression during phases of dormancy release in raspberry (*Rubus idaeus* L.) buds.** *J Exp Bot* 2007, **58**(5):1035-1045.
6. Baudot A, de la Torre V, Valencia A: **Mutated genes, pathways and processes in tumours.** *EMBO Rep* 2010, **11**:805-810.
7. Chen L, Wang R, Zhang XS: **Biomolecular Networks: Methods and Applications in Systems Biology.** *John Wiley and Sons* 2009.
8. Saxena V, Orgill D, Kohane I: **Absolute enrichment: gene set enrichment analysis for homeostatic systems.** *Nucleic Acids Res* 2006, **34**(22):e151.
9. Chen L, Wang R, Li C, Aihara K: **Modelling Biomolecular Networks in Cells: Structures and Dynamics.** Springer-Verlag; 2010.
10. Otsuji T, Okuda-Ashitaka E, Kojima S, Akiyama H, Ito S, Ohmiya Y: **Monitoring for dynamic biological processing by intramolecular bioluminescence resonance energy transfer system using secreted luciferase.** *Anal Biochem* 2004, **329**(2):230-237.
11. Specht A, Bolze F, Omran Z, Nicoud JF, Goeldner M: **Photochemical tools to study dynamic biological processes.** *HFSP J* 2009, **3**(4):255-264.
12. Iola F: **Duarte: Following dynamic biological processes through NMR-based metabonomics: A new tool in nanomedicine?** *J Control Release* 2011, **153**:34-39.
13. Ramakrishnan N, Tadepalli S, Watson LT, Helm RF, Antoniotti M, Mishra B: **Reverse engineering dynamic temporal models of biological processes and their relationships.** *Proc Natl Acad Sci USA* 2010, **107**(28):12511-12516.

14. Choi H, Kim S, Gingras AC, Nesvizhskii AI: **Analysis of protein complexes through model-based biclustering of label-free quantitative AP-MS data.** *Mol Syst Biol* 2010, **6**:385.
15. Conte N, Delaval B, Ginestier C, Ferrand A, Isnardon D, Larroque C, Prigent C, Séraphin B, Jacquemier J, Birnbaum D: **TACC1-chTOG-Aurora A protein complex in breast cancer.** *Oncogene* 2003, **22**:8102-8116.
16. Dutkowski J, Ideker T: **Protein Networks as Logic Functions in Development and Cancer.** *PLoS Comput Biol* 2011, **7**(9):e1002180.
17. Fraser H, Plotkin J: **Using protein complexes to predict phenotypic effects of gene mutation.** *Genome Biol* 2007, **8**(11):R252.
18. Yang P, Li X, Wu M, Kwok CK, Ng SK: **Inferring Gene-Phenotype Associations via Global Protein Complex Network Propagation.** *PLoS ONE* 2011, **6**(7):e21502.
19. Jin R, McCallen S, Liu CC, Xiang Y, Almaas E, Zhou XJ: **Identifying dynamic network modules with temporal and spatial constraints.** *Pac Symp Biocomput* 2009, 203-214.
20. Madeira SC, Teixeira MC, Sa-Correia I, Oliveira AL: **Identification of Regulatory Modules in Time Series Gene Expression Data Using a Linear Time Biclustering Algorithm.** *IEEE/ACM Trans Comput Biol Bioinform* 2010, **7**:153-165.
21. Madeira SC, Oliveira AL: **Biclustering Algorithms for Biological Data Analysis: A Survey.** *IEEE/ACM Trans Comput Biol Bioinform* 2004, **1**:24-45.
22. Aguilar-Ruiz JS: **Shifting and scaling patterns from gene expression data.** *Bioinformatics* 2005, **21**:3840-3845.
23. Gu J, Liu J: **Bayesian biclustering of gene expression data.** *BMC Genomics* 2008, **9**(Suppl 1):S4.
24. Reverter A, Chan EKF: **Combining partial correlation and an information theory approach to the reversed engineering of gene co-expression networks.** *Bioinformatics* 2008, **24**(21):2491-2497.
25. Spellman PT, Sherlock G, Zhang MQ, Iyer VR, Anders K, Eisen MB, Brown PO, Botstein D, Futcher B: **Comprehensive Identification of Cell Cycle-regulated Genes of the Yeast *Saccharomyces cerevisiae* by Microarray Hybridization.** *Mol Biol Cell* 1998, **9**(12):3273-3297.
26. Saeed AI, Sharov V, White J, Li J, Liang W, Bhagabati N, Braisted J, Klapa M, Currier T, Thiagarajan M, Sturn A, Snuffin M, Rezantsev A, Popov D, Ryltsov A, Kostukovich E, Borisovskiy I, Liu Z, Vinsavich A, Trush V, Quackenbush J: **TM4: a free, open-source system for microarray data management and analysis.** *BioTechniques* 2003, **34**(2):374-378.
27. Reimand J, Kull M, Peterson H, Hansen J, Vilo J: **g:Profiler-a web-based toolset for functional profiling of gene lists from large-scale experiments.** *Nucleic Acids Res* 2007, **35**(suppl 2):W193-W200.
28. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, Harris MA, Hill DP, Issel-Tarver L, Kasarskis A, Lewis S, Matese JC, Richardson JE, Ringwald M, Rubin GM, Sherlock G: **Gene ontology: tool for the unification of biology. The Gene Ontology Consortium.** *Nat Genet* 2000, **25**:25-29.
29. Kanehisa M, Goto S: **KEGG: Kyoto Encyclopedia of Genes and Genomes.** *Nucleic Acids Res* 2000, **28**:27-30.
30. Vastrik I, D'Eustachio P, Schmidt E, Joshi-Tope G, Gopinath G, Croft D, de Bono B, Gillespie M, Jassal B, Lewis S, Matthews L, Wu G, Birney E, Stein L: **Reactome: a knowledge base of biologic pathways and processes.** *Genome Biol* 2007, **8**(3):R39.
31. Caraux G, Pinloche S: **PermutMatrix: a graphical environment to arrange gene expression profiles in optimal linear order.** *Bioinformatics* 2005, **21**(7):1280-1281.
32. Pramila T, Miles S, GuhaThakurta D, Jemiolo D, Breeden LL: **Conserved homeodomain proteins interact with MADS box protein Mcm1 to restrict ECB-dependent transcription to the M/G1 phase of the cell cycle.** *Gene Dev* 2002, **16**(23):3034-3045.
33. Pramila T, Wu W, Miles S, Noble WS, Breeden LL: **The Forkhead transcription factor Hcm1 regulates chromosome segregation genes and fills the S-phase gap in the transcriptional circuitry of the cell cycle.** *Gene Dev* 2006, **20**(16):2266-2278.
34. Horvath S, Dong J: **Geometric Interpretation of Gene Coexpression Network Analysis.** *PLoS Comput Biol* 2008, **4**(8):e1000117.
35. Szklarczyk D, Franceschini A, Kuhn M, Simonovic M, Roth A, Minguez P, Doerks T, Stark M, Muller J, Bork P, Jensen LJ, Mering Cv: **The STRING database in 2011: functional interaction networks of proteins, globally integrated and scored.** *Nucleic Acids Res* 2011, **39**(suppl 1):D561-D568.
36. Pierleoni A, Martelli PL, Fariselli P, Casadio R: **eSLDB: eukaryotic subcellular localization database.** *Nucleic Acids Res* 2007, **35**(suppl 1):D208-D212.
37. Smoot ME, Ono K, Ruscheinski J, Wang PL, Ideker T: **Cytoscape 2.8: new features for data integration and network visualization.** *Bioinformatics* 2011, **27**(3):431-432.
38. Pu S, Wong J, Turner B, Cho E, Wodak SJ: **Up-to-date catalogues of yeast protein complexes.** *Nucleic Acids Res* 2009, **37**(3):825-831.

doi:10.1186/1752-0509-6-S1-S12

Cite this article as: Zeng and Chen: Tracing dynamic biological processes during phase transition. *BMC Systems Biology* 2012 **6**(Suppl 1):S12.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

