#### **Meta-Analysis**

## Clinicopathological impacts of high c-Met expression in renal cell carcinoma: a meta-analysis and review

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#### ABSTRACT

c-Met overexpression has been observed in renal cell carcinoma (RCC). However, its clinicopathological impacts remain uncertain. We performed this meta-analysis to evaluate the pathologic and prognostic impacts of high c-Met expression in patients with RCC. A systematic computerized search of the electronic databases PubMed and Embase was performed. From 12 studies, 1,724 patients with RCC were included in the meta-analysis. Compared with RCCs showing low c-Met expression, tumors with high c-Met expression showed significantly higher nuclear grade (odds ratio = 2.45 [95% CI: 1.43–4.19], P = 0.001) and pT stage (odds ratio = 2.18 [95% CI: 1.27–3.72], P = 0.005). In addition, patients with c-Met-high RCC showed significantly worse overall survival than those with c-Met-low tumor (hazard ratio = 1.32 [95% CI: 1.12-1.56], P = 0.0009). In conclusion, this meta-analysis demonstrates that high c-Met expression correlate with significantly worse pathological features and overall survival, indicating c-Met overexpression is a potential adverse prognostic marker for patients with RCC.

#### **INTRODUCTION**

Renal cell carcinoma (RCC) is the most common malignant renal neoplasm, accounting for approximately 85% of kidney cancers [1, 2]. Most patients without metastases can be cured by nephrectomy alone. However, considerable patients have metastatic diseases at the time of diagnosis and nephrectomy is not usually curative for those patients [2, 3]. Until the last decade, immunotherapeutic agents (interferon- $\alpha$  and interleukin-2) had been the main treatment option for patients with metastatic RCC, despite marginal benefits and significant toxicities [4, 5].

With understanding of molecular mechanisms of carcinogenesis, treatment of RCC has dramatically changed. The molecular targeted agents such as sorafenib, sunitinib, axitinib, pazopanib, or temsirolimus are currently recommended with improved outcomes for patients with metastatic RCC [6-10]. However, most tumors eventually develop resistance and their survival benefits are still disappointing. Therefore, efforts to identify novel therapeutic targets and develop more effective targeted drugs are still required. c-Met has recently emerged as a possible therapeutic target in various tumors including RCC [11–13].

c-Met is the tyrosine kinase receptor for hepatocyte growth factor (HGF) and encoded by the proto-oncogene MET located on chromosome 7. The HGF-c-Met signaling pathway regulates multiple cellular functions, including differentiation, proliferation, and angiogenesis [14, 15]. Thus, dysregulation of c-Met and HGF has been implicated in the pathogenesis of cancers. It is related to molecular mechanisms of tumor cell proliferation and survival, invasion, and metastasis [16]. The enhanced expression of c-Met has been observed in various tumors, such as breast cancer [17], lung cancer [18], gastric cancer

[19], colorectal cancer [20], cervix cancer [21], pancreatic cancer [22], and hepatocellular carcinoma [23]. Several meta-analyses in common tumors indicated that high c-Met expression was associated with a poor prognosis [17–23].

The expression of c-Met has also been observed in various cytomorphologic subtypes of RCC [24–39]. High c-Met expression has been associated with poor pathologic features and prognosis in many studies [24–26, 28–31]. However, most studies had a small number of patients, and there has been some conflicts regarding its clinicopathological impacts in RCC [27, 32–34, 38, 39]. Therefore, we performed this meta-analysis to evaluate the pathologic and prognostic roles of c-Met overexpression in patients with RCC.

#### RESULTS

#### **Results of search**

Figure 1 shows flow diagram of search process. A total of 187 relevant studies were initially retrieved, but 171 of them were excluded after screening the titles and abstracts. Of the remaining 16 potentially eligible studies, 4 were further excluded by the inclusion criteria: two had no criteria for c-Met expression status [24, 25] and the others adopted too low cutoff values (< 10%) for

high c-Met expression [26, 27]. Finally, 12 studies were included in the meta-analysis [28–39].

#### Characteristics of the included studies

Table 1 summarizes the main characteristics and clinicopathological outcomes of the 12 included studies. All the studies were performed retrospectively. From the 12 studies, 1,724 patients were included in the metaanalysis. In one study, patients (n = 81) had chromophobe RCC [35]. Two small studies had patients (n = 96) only with papillary RCC (pRCC) [30, 33] and other three had patients (n = 752) only with clear cell RCC (ccRCC) [36, 38, 39]. Almost all patients had received renal surgery as primary treatment for RCC. In two studies [38, 39], patients were treated with sunitinib as a first-line therapy for metastatic RCC. Except for one [32], eleven studies used immunohistochemistry (IHC) to assess c-Met expression status [28–31, 33–39].

#### c-Met expression assignation

There was a marked heterogeneity between the criteria used to dichotomize c-Met status (low c-Met or high c-Met). The criteria were briefly summarized in the Table 1. The rates of high c-Met expression were various, ranging from 16.7% [29, 36] to 80% [30].



Figure 1: Flow diagram of search process.

Author (year) Country	Histology	Methods, antibody, detection kit or immunostainer	No. of pts	Criteria for c-Met <sup>high</sup>	c-Met <sup>high</sup> (%)	OR for NG (95% CI)	OR for pT stage (95% CI)	HR for OS (95% CI)
Pisters <i>et al.,</i> (1997) USA	RCC	IHC with whole slides, Rabbit c-Met Ab	41	$\geq$ 30% of cancer cells	28 (68.3%)	8.1 (0.75–87.23)	0.99 (0.26–3.70)	NA
Inoue <i>et al.,</i> (1998) Japan	RCC	IHC with whole slides, c-Met c-12	120	Positive: higher membrane staining than normal kidney (c-Met <sup>high</sup> : $\geq$ 50% cancer cells with positivity)	20 (16.7%)	2.97 (1.06-8.36)	0.84 (0.25–2.75)	NA
Sweeney et al., (2002) USA	pRCC	IHC with whole slides, NA	50	Intensity: 0 (absent) to 3 ( intense) (c-Met <sup>high</sup> : $\geq$ positive cytoplasmic or membrane staining grade 1 or higher in $\geq$ 10% of cancer cells)	40 (80%)	1.83 (0.45–7.51) P = 0.157	12.76 (0.7 - 233.48) P = 0.004	6.93 (0.92-52.23) P = 0.07
Miyata <i>et al.,</i> (2006) Japan	RCC	IHC with whole slides, Phosphorylated c-Met Ab, DAKO EnVision	114	> 50% of cancer cells with higher staining than normal kidney	73 (64%)	1.70 (0.77–3.75)	2.52 (0.93-6.64)	2.94 (1.12–7.72) <i>P</i> = 0.028
Betsunoh <i>et al.</i> , (2007) Japan	RCC	RT-PCR	66	Tumor/normal ratio $\geq 3$	17 (25.8%)	NA	NA	3.16 (0.10 - 92.3) P = 0.505
Gontero <i>et al.,</i> (2008) Italy	pRCC	IHC with whole slides, Clone DQ 13	46	Cytoplasmic staining $\geq 30\%$ of cancer cells	13 (28.3%)	NA	NA	0.96 (0.38–2.44) <i>P</i> = 0.609
Gibney <i>et al.</i> , (2012) USA	RCC	IHC with TMA, Anti-c-Met Ab (MET4)	317	$\geq$ Cutoff point of median AQUA score (32.5%)	159 (50.2%)	NA	NA	1.36 (1.08-1.74) P = 0.0091
Erlmeier <i>et al.,</i> (2013) Germany	chRCC	IHC with TMA, Anti-MET (AB-103), EnVision Kit	81	Intensity: 0 = no staining; 1 = weak; 2 = moderate; 3 = strong Proportion of staining area: recorded in percent (0–100%) (c-Met <sup>high</sup> : intensity score × percentage score > median)	24 (29.6%)	NA	1.89 (0.39–9.19) P = 0.6	1.90 ( $(0.42-8.57)$ ) P = 0.59
Chen <i>et al.,</i> (2017) China	ccRCC	IHC with TMA Rabbit anti-c-Met Ab, SP-9000 SP link Kit	90	Intensity: $0 = no$ signal; $1 =$ weak; 2 = moderate; $3 =$ strong Positive rate: $0 = no; 1 = 1-25\%;$ 2 = 26-50%; 3 = 51-75%; 4 = 76-100% (c-Met <sup>high</sup> : intensity score × positive rate score $\geq 6$ )	15 (16.7%)	10.42 (1.30–83.37)	7.46 (2.16–25.75)	2.85 (1.21-6.70) P = 0.017
Peltola <i>et al.,</i> (2017) Finland	RCC	IHC with whole slides, Anti-c-Met ready-to-use Mab, BenchMark XT	137	Intensity: 0 = no staining; 1 = weak; 2 = strong; 3 = very strong (c-Met <sup>high</sup> : intensity score 2 or 3)	59 (43.1%)	NA	NA	1.22 ( $0.81-1.82$ ) P = 0.34
Macher- Goeppinger <i>et al.,</i> (2017) Germany	ccRCC	IHC with TMA, Anti-total c-Met (SP44), OptiView DAB IHC Kit	572	Intensity: 0 = negative; 1 = low; 2 = medium; 3 = high Quantity: 0 = no expression; 1 = < 10% of positive cells; 2 = positive in 10–50%; 3 = positive in 51–80%; 4 = positive in $\ge 80\%$ (c-Met <sup>high</sup> : intensity score $\times$ quantity score $\ge 6$ )	184 (32.2%)	NA	NA	$\begin{array}{l} 1.05 \\ (0.69 - 1.61) \\ P = 0.81 \end{array}$
Kammerer- Jacquet et al., (2017) France	ccRCC	IHC with whole slides Anti-total c-Met (SP44), BenchMark XT	90	Intensity: 0 = absent; 1 = weak; 2 = moderate; 3 = strong (c-Met <sup>high</sup> : intensity score 2 or 3)	62 (68.9%)	NA	NA	0.99 (0.56–1.78)

### Table 1: Summary of the 12 included studies

RCC, renal cell carcinoma; ccRCC, clear cell RCC; chRCC, chromophobe RCC; pRCC, papillary RCC; IHC, immunohistochemistry; TMA, tissue microarray; Ab, antibody; Mab, monoclonal antibody; RT-PCR, reverse transcription-polymerase chain reaction; pts, patients; AQUA, automated quantitative analysis; OR, odds ratio; HR, hazard ratio; CI, confidence interval; OS, overall survival; NA, not available

# Impact of high c-Met expression on pathological features

From six studies [28–31, 35, 36], 496 patients were included in the meta-analysis of odds ratios (ORs) with 95% confidence intervals (CIs) for nuclear grade and/or depth of cancer penetration (pT stage).

Compared with RCCs with low c-Met expression, tumors with high c-Met expression showed significantly

higher nuclear grade (II–IV) (OR = 2.45 [95% CI: 1.43–4.19], P = 0.001) (Figure 2A). The fixed-effects model was selected because there was no significant heterogeneity among studies ( $X^2 = 3.95$ , P = 0.41, P = 0%).

In terms of primary tumor stage, there was a positive correlation between c-Met overexpression and higher pT stage (pT3 and pT4) (OR = 2.18 [95% CI: 1.27-3.72], P = 0.005) (Figure 2B). The fixed-effects model was

used because there was little heterogeneity across studies  $(X^2 = 9.11, P = 0.10, I^2 = 45\%)$ .

# Impact of high c-Met expression on overall survival

From ten studies [30–39], 1,563 patients were included in the meta-analysis of HRs with 95% CIs for OS. Patients with c-Met-high RCC showed significantly worse OS than those with c-Met-low tumor (HR = 1.32 [95% CI: 1.12–1.56], P = 0.0009) (Figure 3). The fixed-effects model was selected because there was no significant heterogeneity across the studies ( $X^2 = 11.55$ , P = 0.24,  $I^2 = 22\%$ ).

In the subgroup analyses, however, patients with c-Met-high tumor did not show significantly worse OS than those with c-Met-low tumor both in pRCC

#### A

(HR = 1.36 [95% CI: 0.58–3.16], P = 0.48) (Figure 4A) and ccRCC (HR = 1.29 [95% CI: 0.76–2.19], P = 0.34) (Figure 4B).

#### **Publication bias**

Visual inspection of the funnel plots for nuclear grade, pT stage, and OS showed symmetry, indicating there were no substantial publication biases (Figure 5).

#### **DISCUSSION**

In this meta-analysis, we evaluated the pathologic and prognostic impact of c-Met overexpression in patients with RCC. The results show that high c-Met expression is associated with significantly worse pathological features and prognosis.

			c-Met high	c-Met low		Odds Ratio		Odd	s Ratio		
Study or Subgroup	log[Odds Ratio]	SE	Total	Total	Weight	IV, Fixed, 95% Cl		IV, Fixe	ed, 95% Cl		
Pisters et al. (1997)	2.0919	1.2133	28	13	5.1%	8.10 [0.75, 87.35]				•	
Inoue et al. (1998)	1.0886	0.5268	20	100	27.2%	2.97 [1.06, 8.34]					
Sweeney et al. (2002)	0.6043	0.718	40	10	14.7%	1.83 [0.45, 7.47]			+		
Miyata et al. (2006)	0.5306	0.4039	73	41	46.3%	1.70 [0.77, 3.75]			+		
Chen et al. (2017)	2.3437	1.0615	15	75	6.7%	10.42 [1.30, 83.45]				•	
Total (95% CI)			176	239	100.0%	2.45 [1.43, 4.19]			•		
Heterogeneity: Chi² = 3. Test for overall effect: Z	95, df = 4 (P = 0.41 = 3.26 (P = 0.001)	); I² = 0%					0.01	0.1 Favors (c-Met high	1 I Favors (	10 c-Met low]	100

#### В

			c-Met high	c-Met low		Odds Ratio		Odds F	latio	
Study or Subgroup	log[Odds Ratio]	SE	Total	Total	Weight	IV, Fixed, 95% Cl		IV, Fixed,	95% CI	
Pisters et al. (1997)	-0.0101	0.6774	28	13	16.4%	0.99 [0.26, 3.73]				
Inoue et al. (1998)	-0.1744	0.6117	20	100	20.1%	0.84 [0.25, 2.79]				
Sweeney et al. (2002)	2.5463	1.4821	40	10	3.4%	12.76 [0.70, 233.03]		+	•	
Miyata et al. (2006)	0.9243	0.5014	73	41	29.8%	2.52 [0.94, 6.73]		+		
Erlmeier et al. (2013)	0.6366	0.8061	24	57	11.5%	1.89 [0.39, 9.18]			•	
Chen et al. (2017)	2.0096	0.6322	15	75	18.8%	7.46 [2.16, 25.76]				
Total (95% CI)			200	296	100.0%	2.18 [1.27, 3.72]			•	
Heterogeneity: Chi <sup>2</sup> = 9.11, df = 5 (P = 0.10); l <sup>2</sup> = 45%							0.01	01 1	10	100
Test for overall effect: Z = 2.84 (P = 0.005)							0.01	Favors (c-Met high)	Favors (c-Met low)	

#### Figure 2: Forest plots of odds ratios for nuclear grade (A) and pT stage (B).

			c-Met high	c-Met low		Hazard Ratio		Hazard Ratio
Study or Subgroup	log[Hazard Ratio]	SE	Total	Total	Weight	IV, Fixed, 95% Cl		IV, Fixed, 95% Cl
Sweeney et al. (2002)	1.9359	1.0304	40	10	0.7%	6.93 [0.92, 52.22]		
Miyata et al. (2006)	1.0784	0.4925	73	41	2.9%	2.94 [1.12, 7.72]		
Betsunoh et al. (2007)	1.1506	1.7417	17	49	0.2%	3.16 [0.10, 95.99]		
Gontero et al. (2008)	-0.0408	0.4744	13	33	3.2%	0.96 [0.38, 2.43]		
Gibney et al. (2012)	0.3075	0.1217	159	158	48.0%	1.36 [1.07, 1.73]		<b>₩</b>
Erlmeier et al. (2013)	0.6419	0.7693	24	57	1.2%	1.90 [0.42, 8.58]		
Chen et al. (2017)	1.0473	0.4366	15	75	3.7%	2.85 [1.21, 6.71]		
Peltola et al. (2017)	0.1989	0.2065	59	78	16.7%	1.22 [0.81, 1.83]		- <b>-</b>
Macher-Goeppinger et al. (2017)	0.0488	0.2161	184	388	15.2%	1.05 [0.69, 1.60]		
Kammerer-Jacquet et al. (2017)	-0.0101	0.2936	62	28	8.2%	0.99 [0.56, 1.76]		-+-
Total (95% CI)			646	917	100.0%	1.32 [1.12, 1.56]		•
Heterogeneity: Chi <sup>2</sup> = 11.55, df = 9	(P = 0.24); I <sup>2</sup> = 22%						0.01	0.1 1 10 100
Test for overall effect: Z = 3.32 (P =	0.0009)						5.01	Favors [c-Methigh] Favors [c-Metlow]

#### Figure 3: Forest plot of hazard ratios for overall survival.

MET activation has been proven to play a critical role in the pathogenesis and progression of many tumor types [14-16]. Mechanisms of MET activation include mutations, amplification, and enhanced transcription [40]. Germline *MET* mutations have been identified in hereditary and sporadic pRCC [41]. c-Met was overexpressed in von Hippel-Lindau (VHL) RCC cells due to the upregulation of hypoxia-inducible factors [42]. In addition, VHL mutation and/or loss of heterozygosity have been associated with c-Met expression in ccRCC [25]. Besides pRCC and ccRCC, MET upregulation has also been detected in a rarer subtype, chromophobe RCC [35].

In RCC, many studies have suggested that c-Met expression is associated with significantly inferior clinicopathological features, such as tumor grade [27, 28–31, 36, 38], primary tumor stage [30, 31, 36], lymphatic invasion [27], metastases [30, 35, 38], and worse progression-free survival [37] or OS [31, 34, 36]. However, the pathological or clinical impacts of c-Met expression are not consistent across the studies [33, 38, 39]. In addition, because most studies had a small number of patients and adopted various methods and criteria for c-Met expression status, they could not draw a consensus regarding the clinicopathological roles of c-Met overexpression. With respect to cancer-specific survival, in particular, the prognostic value of c-Met overexpression has been controversial. Gibney et al. evaluated c-Met expression as a prognostic marker in 317 patients with RCC and found that high c-Met expression was an independent predictor of survival (multivariate HR = 1.013 [95% CI: 1.002-1.023, P = 0.015 [34]. Recently, Macher-Goeppinger et al. assessed c-Met expression and MET copy number in 572 patients with ccRCC [38]. Patients with high c-Met expression showed significantly worse OS

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Hazard Ratio log[Hazard Ratio] IV, Fixed, 95% Cl IV. Fixed, 95% Cl Study or Subgroup SE Total Total Weight Sweeney et al. (2002) 1.9359 1.0304 40 10 17.5% 6.93 [0.92, 52.22] Gontero et al. (2008) -0.0408 0.4744 13 82.5% 0.96 [0.38, 2.43] 33 Total (95% CI) 53 43 100.0% 1.36 [0.58, 3.16] Heterogeneity: Chi<sup>2</sup> = 3.04, df = 1 (P = 0.08); l<sup>2</sup> = 67% 0.01 100 01 10 Test for overall effect: Z = 0.71 (P = 0.48) Favors [c-Methigh] Favors [c-Metlow] В Hazard Ratio Hazard Ratio c-Met high c-Met low Study or Subgroup log[Hazard Ratio] SE Total Total Weight IV, Random, 95% Cl IV, Random, 95% CI Chen et al. (2017) 1.0473 0.4366 15 75 23.0% 2.85 [1.21, 6.71] Macher-Goeppinger et al. (2017) 42.5% 1.05 [0.69, 1.60] 0.0488 0.2161 184 388 Kammerer-Jacquet et al. (2017) -0.0101 0.2936 62 28 34.5% 0.99 [0.56, 1.76] 261 100.0% Total (95% CI) 491 1.29 [0.76, 2.19] Heterogeneity: Tau<sup>2</sup> = 0.12; Chi<sup>2</sup> = 4.73, df = 2 (P = 0.09); l<sup>2</sup> = 58% 0.01 0.1 10 100 Test for overall effect: Z = 0.96 (P = 0.34) Favors [c-Methigh] Favors [c-Metlow]

Hazard Ratio

Figure 4: Forest plots of hazard ratios for overall survival in papillary RCC (A) and clear cell RCC (B).

c-Met high c-Met low

than those with c-Met-low tumor (HR = 1.49 [95% CI: 1.11-2.0], P = 0.008) in univariate analysis. In multivariate analysis, however, c-Met overexpression did not remain as an independent prognostic factor (HR = 1.05 [95% CI: 0.69-1.61, P = 0.81].

In the current meta-analysis, we included studies comparing the major pathological features (nuclear grade and pT stage) and cancer-specific survival outcome according to the c-Met expression status. Chen at al. had also performed the similar meta-analysis regarding clinicopatholigical impacts of c-Met expression in RCC [36]. They included studies with no criteria or low threshold for c-Met expression. In this study, however, we excluded two articles with no criteria for c-Met expression [24, 25] and another two with very low cutoff value (IHC staining in < 10% of tumor cells) for high c-Met expression [26, 27]. Compared with RCCs showing low c-Met expression, tumors with high expression showed significantly higher nuclear grade (OR = 2.45, P = 0.001) and pT stage (OR = 2.18, P = 0.005). In addition, patients with c-Met-high RCC showed significantly worse OS than those with c-Met-low tumor (HR = 1.32, P = 0.0009). Our findings indicate that high c-Met expression represent a potential adverse prognostic marker for patients with RCC. In the subgroup analyses, however, the OS failed to show statistically significant difference between patients with c-Met-high tumor and those with c-Met-low tumor in both pRCC (HR = 1.36, P = 0.48) and ccRCC (HR = 1.29, P = 0.34). Because the limited number of studies was included in the subgroup analyses, further studies are needed to evaluate the prognostic role of c-Met expression in the subtypes of RCC.

Several meta-analyses in other cancers have also demonstrated that high c-Met expression is an adverse prognostic marker for survival [17-23]. Thus, inhibition of c-Met/HGF pathway may provide an effective therapeutic strategy for cancers with c-Met overexpression [43]. Based on the scientific rationale to target c-Met, various c-Met inhibitors have been investigated in a variety of cancers, including RCC [12, 13, 44-49]. Cabozantinib is an oral inhibitor of tyrosine kinases including MET, VEGFR, and AXL. The randomized phase 3 METEOR trial compared the efficacy and safety of cabozantinib versus the mTOR inhibitor everolimus in patients with advanced RCC who progressed after previous VEGFR tyrosine-kinase inhibitor treatment [49]. Compared with everolimus, cabozantinib significantly prolonged OS (median 21.4 vs. 16.5 months, HR = 0.66 [95% CI: 0.53-0.83], P = 0.00026) and progression-free survival (median 7.4 vs. 3.9 months, HR = 0.51 [95% CI: 0.41-0.62], P < 0.0001). Based on these results, the U. S. FDA approved cabozantinib for patients with advanced RCC who have received prior anti-angiogenic therapy. In a phase II study with RCC, interestingly, therapeutic response of foretinib (a multi-kinase inhibitor targeting c-Met, VEGF, RON, AXL, and TIE-2 receptors) was closely associated with the germline MET mutations [12]. In addition, the efficacy of c-Met-targeting agents has been associated with high c-Met expression in non-smallcell lung cancer and hepatocellular carcinoma [46, 47]. These results suggest that cancers showing high c-Met expression may be good candidates for c-Met inhibitors.

Recently, MET amplification/upregulation has been proposed as a mechanism of resistance to antiangiogenic therapy [46, 47]. Anti-angiogenic therapy induces hypoxia by decreasing blood supply to tumors, which may upregulate c-Met [42]. c-Met activation in turn promotes tumor invasion and metastasis [16]. Peltola et al. retrospectively analyzed c-Met expression in 137 patients with metastatic RCC treated with sunitinib and found that high c-Met expression was associated with poor survival [38]. c-Met upregulation by anti-VEGF therapy may explain the reason why patients with high c-Met expression achieved less survival benefit from sunitinib. This finding indicates that c-Met expression may serve as a biomarker to predict who benefit less from antiangiogenic therapy. Therefore, those patients with c-Met overexpression might benefit from a dual inhibitor [48].

The major limitation for clinical application of c-Met inhibitors is that there is no consensus of the reliable criteria for high c-Met expression. A variety of methods, such as IHC, Western blot, fluorescence *in situ* hybridization, reverse transcription-polymerase chain reaction (RT-PCR), and molecular invasion probe are currently used to





assess c-Met status, but there are no standardized criteria for overexpression. There are also differences in the IHC criteria for high c-Met expression. The discrepancies in the clinicopathological impacts of c-Met among studies might be due to the different methods and criteria for high c-Met expression. Therefore, the definition of reliable criteria for c-Met status is essential to verify the prognostic role of c-Met expression and develop c-Met inhibitors in solid tumors.

Our study has several inherent limitations. First, the meta-analysis included a small number of studies. Second, all the studies were retrospectively performed. Third, as we mentioned above, the studies used different IHC methods (antibodies, detection kits, tissue samples-whole slide or tissue microarray, staining sites-cytoplasm or membrane) for assessing c-Met expression. In addition IHC criteria to stratify c-Met status were also various among studies. Fourth, because of limited information in most studies, we could not consider the impact of systemic therapies that would have inevitably affected the OS. Finally articles published only in English were included, which might bias the results.

In conclusion, our results show that c-Met overexpression is significantly associated with poor pathological features and prognosis. These findings indicate that high c-Met expression is a potential adverse prognostic marker for patients with RCC. However, larger studies using standardized methods and criteria are still needed to verify the prognostic roles of c-Met expression in various subtypes of RCC.

### **MATERIALS AND METHODS**

#### Search strategy

We performed this meta-analysis according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines [50]. A systematic computerized search of the electronic databases PubMed and Embase (up to May 2017) was done. The search used the following terms: "c-Met" or "Met," "hepatocyte growth factor receptor," and "renal cell carcinoma." The related articles function in the PubMed was also used to identify all related articles. The titles and abstracts of the retrieved studies were initially scanned to exclude irrelevant papers. Then, the potentially relevant articles were reviewed in full text, further excluding those that did not meet the inclusion criteria of this meta-analysis.

#### **Inclusion criteria**

Eligible studies should meet the following inclusion criteria: (i) patients had a pathological diagnosis of RCC; (ii) pathological features (nuclear grade and/or pT stage) or OS were analyzed according to c-Met expression status; (iii) ORs with 95% CIs for pathological features or HR with 95% CI for OS were provided or could be estimated from the data provided; (iv) articles were published in English. Articles with no criteria for c-Met expression status were excluded. We also excluded articles with very low cutoff value (IHC staining in < 10% of tumor cells) for high c-Met expression.

#### **Data extraction**

The required data were collected independently by two investigators (BJK and JHK). If these two authors did not agree, the other investigator (HSK) was consulted to resolve the discrepancies.

The following data were recorded from all eligible studies: the first author's name, publication year, country, histology, nuclear grade, pT stage, methods to test c-Met expression, antibody and detection kit for IHC, number of patients, treatment, cutoff values adopted to dichotomize c-Met expression as 'high' or 'low', and HR with 95% CI for OS and OR with 95% CI for pathological features.

#### Statistical analysis

Statistical values were obtained directly from the original articles. When OR or HR with 95% CI were not provided, the Engauge Digitizer (version 9.1) was used to estimate the needed data from the results and Kaplan-Meier curves. The strength of the association between c-Met overexpression and nuclear grade or pT stage was shown as ORs with their 95% CIs. The effect size of OS was pooled through HR with its 95% CI. The heterogeneity across studies was tested by the Q statistic and the I<sup>2</sup> inconsistency test. The fixed-effects model (Mantel-Haenszel method) was selected for pooling homogeneous outcomes when  $P \ge 0.1$  and  $I^2 \le 50\%$ , whereas the randomeffects model (DerSimonian-Laird method) was applied for pooling heterogeneous outcomes when P < 0.1 and  $l^2 > 50\%$ . The RevMan (version 5.2) was used to combine data and report outcomes. All reported P-values were twosided and P < 0.05 was considered statistically significant. Publication bias was assessed graphically by the funnel plot method [51].

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### **CONFLICTS OF INTEREST**

All authors have declared no competing interest.

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