Computational and Structural Biotechnology Journal 18 (2020) 2568-2572



101010100100100100100100 J O U R N A L

COMPUTATIONAL ANDSTRUCTURAL BIOTECHNOLOGY

journal homepage: www.elsevier.com/locate/csbj

TGIF1-Twist1 axis in pancreatic ductal adenocarcinoma

Mohammed S. Razzaque^{a,*}, Azeddine Atfi^b

^a Department of Pathology, Lake Erie College of Osteopathic Medicine, Erie, PA, USA ^b Department of Pathology and Massey Cancer Center, Virginia Commonwealth University, Richmond, VA, USA

ARTICLE INFO

Article history: Received 18 June 2020 Received in revised form 11 September 2020 Accepted 11 September 2020 Available online 17 September 2020

Keywords: TGIF1 TGF-β Twist1 Tumor suppressor Pancreatic ductal adenocarcinoma

ABSTRACT

TG-interacting factor 1 (TGIF1) exerts inhibitory effects on transforming growth factor-beta (TGF- β) signaling by suppressing Smad signaling pathway at multiple levels. TGIF1 activity is important for normal embryogenesis and organogenesis, yet its dysregulation can culminate in tumorigenesis. For instance, increased expression of TGIF1 correlates with poor prognosis in triple-negative breast cancer patients, and enforced expression of TGIF1 facilitates Wnt-driven mammary tumorigenesis, suggesting that TGIF1 might function as an oncoprotein. Quite surprisingly, TGIF1 has recently been shown to function as a tumor suppressor in pancreatic ductal adenocarcinoma (PDAC), possibly owing to its ability to antagonize the pro-malignant transcription factor Twist1. In this article, we will briefly elaborate on the biological and clinical significance of the unique tumor-suppressive function of TGIF1 and its functional interaction with Twist1 in the context of PDAC pathogenesis and progression.

© 2020 The Author(s). Published by Elsevier B.V. on behalf of Research Network of Computational and Structural Biotechnology. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Contents

1. Introduction 2 2. Dichotomous roles of TGF-β signaling in cancer 2 3. Role of TGIF1 in the progression of PDAC. 2 4. Functional interaction between TGIF1 and Twist1 in PDAC 2 5. Conclusion 2 Declaration of Competing Interest 2 Acknowledgements 2 References 2

1. Introduction

Pancreatic ductal adenocarcinoma (PDAC) is an extremely aggressive tumor and a leading cause of cancer-associated related death [1]. More than 85% of pancreatic cancers are PDAC, and less than 7% of patients with PDAC have a 5-years survival rate [2]. PDAC arise from pancreatic acinar cells or their common progenitor [3]. Early stages of pancreatic tumors are usually symptom-free, and the tumor becomes clinically apparent once cells have invaded the adjacent tissues or metastasize to distant organs.

E-mail address: mrazzaque@lecom.edu (M.S. Razzaque).

Therefore, most of the patients who present with pancreatic cancer symptoms frequently present advanced stages of the disease. The liver, peritoneum, lungs, and bones are the most common sites for pancreatic tumor metastasis [4]. To make the situation worse for the patients, PDAC does not respond well to conventional chemotherapy and radiotherapy [5]. A two-fold increase in the number of PDAC cases and subsequent death is predicted in the USA and worldwide in the next ten years [6,7]. In addition, both obesity and type 2 diabetes are predisposing factors for PDAC [8,9], highlighting the fact that other major life-threatening conditions could contribute to the rising prevalence of PDAC.

The pancreatic intraepithelial neoplasia (PanIN), with activating mutations in the *KRAS* proto-oncogene, usually leads to the formation of more than 90% of PDACs [3,10]. The *KRAS* activating muta-







^{*} Corresponding author at: Department of Pathology, Lake Erie College of Osteopathic Medicine, 1858 West Grandview Boulevard, Erie, PA 16509, USA.

^{2001-0370/© 2020} The Author(s). Published by Elsevier B.V. on behalf of Research Network of Computational and Structural Biotechnology. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

tions are key early genetic markers of the genesis of PDAC, while the subsequent accumulation of inactivating mutations in several tumor suppressor genes (e.g., *p16lNK4A*, *TP53*, and *SMAD4*) are related to tumor progression and late-stage metastasis [3,11]. Genetically, inducing the expression of the most prevalent oncogenic mutation in Kras (Kras^{G12D}) in mouse pancreas leads to the formation of PanINs that occasionally progress to an invasive PDAC phenotype with a very prolonged latency period, suggesting a key role of KRAS in the initiation of human PDAC [12,13]. Interestingly, combining Kras^{G12D} expression with genetic inactivation of the tumor suppressor *Smad4*, which encodes for an essential component of the canonical transforming growth factor-beta (TGF- β) signaling pathway, led to a dramatic acceleration of PDAC, highlighting the importance of this pathway in restricting PDAC progression [11].

2. Dichotomous roles of TGF-β signaling in cancer

Canonical TGF- β signaling is initiated by the interaction of the ligand with two transmembrane receptors called type I (TBRI) and type II ($T\beta RII$) [14,15]. The $T\beta RII$ is constitutively active while the T β RI is inactive in the absence of ligand, [16]. TGF- β binding allows the formation of an oligomeric complex in which TBRII phosphorylates T_BRI, leading to the activation of its kinase activity. The activated TβRI then phosphorylates Smad2 and Smad3, which subsequently form a complex with Smad4, and the complexes translocate into the nucleus to regulate the expression of TGF-B genes through interacting with transcriptional cofactors or transcriptional corepressors [15,17]. As for the roles of TGF-β pathways in tumorigenesis, reduced signaling through the "loss of function" mutations of TBRII has been detected in a significant number of colorectal and gastric carcinomas with microsatellite instability [18]. Impaired TGF- β signaling due to the deletion of *SMAD4* is also detected in 16-25% of colorectal cancer and up to 50% in PDAC

[19]. Quite paradoxically, sustained activation of TGF-β can exacerbate tumor progression rather than exerting tumor-suppressive effects. For instance, increased expression of TGF-β has been shown to be associated with poor prognosis and decreased survival in colorectal cancer patients [20,21]. Other studies have shown that TGFβ-mediated angiogenesis could also enhance tumor growth [22,23]. Thus, the TGF- β pathway appears to have both anti- and pro-tumorigenic functions [24]. In the early stage of tumorigenesis, activation of TGF-β signaling can induce cell cycle arrest and apoptotic cell death, while in the later stages, TGF-β activation can promote tumor progression, invasion and metastasis by facilitating the epithelial-to-mesenchymal transition (EMT) as well as increasing cancer cell motility [25,26]. These opposing functions during the different stages of tumorigenesis are known as the "TGF-B paradox" [27]. Recently, we have shown that sustained activation of TGF-B signaling through the deletion of the *Tgif1* gene promotes the progression of PDAC, shedding new mechanistic insight into how TGF-β signaling can exert its bimodal function in this aggressive malignancy [28].

3. Role of TGIF1 in the progression of PDAC

TGIF1 is a suppressor of TGF- β signaling, acting primarily either by preventing Smad2 phosphorylation [29] or by facilitating the ubiquitin-dependent degradation of Smad2 [30] (Fig. 1). TGIF1 maintains cellular homeostasis by influencing cell differentiation and proliferation [31,32]. Mutations in the human *TGIF1* gene are associated with holoprosencephaly, a congenital developmental anomaly of the forebrain [33]. In direct support of the role of TGIF1 in holopresencephaly, ablating the mouse *Tgif1* gene resulted in a defective brain development phenotype with features reminiscent of holoprosencephaly [34]. Besides its role in development, TGIF1 has been shown to play oncogenic roles in a wide variety of human hematological and solid malignancies. For instance, in myeloge-



Fig. 1. Simplified diagram showing canonical TGF-β signaling. Note TGIF1 exerts an antagonistic effect on TGF-β signaling.

nous leukemia, an association between TGIF1 expression and poor patient outcomes was noted [35]. In similar lines of study, *TGIF1* gene overexpression is correlated with poor survival in urinary tract urothelial carcinoma and triple-negative breast cancer [36,37]. Moreover, constitutive TGIF1 protein stabilization occurs in promyelocytic leukemia initiated by the oncogenic fusion protein PML-RAR α [32], providing strong indications that TGIF1 function to promote tumorigenesis. TGIF1 is deemed to contribute to tumorigenesis through the suppression of the TGF- β cytostatic program, which is known to orchestrate cell cycle arrest and apoptotic cell death in many cell systems [38].

Perhaps unexpectedly, we found that genetic inactivation of Tgif1 in mice accelerated Kras^{G12D}-driven PDAC, suggesting that TGIF1 might also have a dual role in tumorigenesis, as does TGFβ signaling [39]. Mice with global *Tgif1* knockout alone did not show any major pancreatic structural or functional changes. Similarly, targeted ablation of *Tgif1* in pancreatic progenitor cells did not result in any pancreatic defects. Even though targeted pancreatic deletion of Tgif1 caused an elevated TGF-B/Smad signaling, none of the Tgif1^{KO} mice developed pancreatic neoplasms in the 18-month follow up period, indicating that activation of TGF-β signaling does not play a major role in pancreas biology and function [39]. Similar results were reported by a separate group, showing no pancreatic developmental defects or pancreatic tumor formation in mice with targeted pancreatic deletion of *Tgif1* [40]. Consistent with the absence of no obvious histological changes between the wild-type and mutant mice, the glucose tolerance tests were similar, indicating that TGIF1 is dispensable for pancreas development and physiology [40].

As discussed earlier, the expression of Kras^{G12D} in mice leads to the development of PanINs that eventually progress to PDAC after a long latency period, typically within 8 to 12 months of age. Despite having no effect on the pancreas in a wild-type background, we found that ablating *Tgif1* from these mutant mice accelerated the onset of PDAC formation and progression, as gauged by the dramatic decrease in survival of mice, which rarely exceed two months [39]. What was also interesting is the observation that Tgif1 inactivation in Kras^{G12D}-bearing mice also bolstered the metastatic behaviors of PDAC, providing us with a powerful tool to investigate mechanistic paradigms of PDAC metastasis [39], which remain poorly understood. Again, these results were independently validated by a separate group, which demonstrated using similar genetic approaches, that TGIF1 deficiency in the pancreas is enough to accelerate PDAC in cooperation with Kras^{G12D} [40]. Given the sustained activation of TGF- β signaling in these mice, it is likely that combined Kras and TGF-β activation synergistically promotes pancreatic tumorigenesis with increased metastatic potentials, therefore resulting in reduced survival. It is also worth restating that the inactivation of TGF-β signaling through genetic deletion of Smad4 also facilitates Kras^{G12D}-driven PDAC, which fits well with the dichotomous role of TGF- β signaling as both tumor suppressor and tumor promoter, depending on the stage of tumor progression. Given these similarities in phenotypes following TGIF1 or Smad4 inactivation, it would be interesting to investigate whether TGIF1 and Smad4 converge to regulate biological processes, whether pro- or anti-tumorigenic, that are instrumental to PDAC progression. Regardless, the available experimental data indicate that the underlying mechanisms of TGIF1-mediated PDAC are partly driven by Twist1 [39].

4. Functional interaction between TGIF1 and Twist1 in PDAC

Twist1 is a pro-malignant transcription factor that plays important role in tumor invasion and metastasis in a wide variety of human malignancies, including PDAC [41]. Twist was first detected

in Drosophila [42], and subsequently Twist isoforms were identified in humans and mice [43,44]. Twist1 is important for regulating the activity of genes that are essential for embryogenesis and organogenesis [44-46]. Mutation of the human TWIST1 gene resulted in craniosynostosis (premature closure of the sutures between the bones of the skull), as detected in the patients with Saethre-Chotzen syndrome [47]. Increased expression of Twist1 has been shown to be associated with various human malignant tumors, including PDAC [41,48]. Recently, the induction of Twist1 in muscle progenitor cells has been demonstrated to drive muscle cachexia during the progression of PDAC [49,50]. One of the possible mechanisms of PDAC-induced muscle cachexia is through the tumor-derived Activin A, which acts on the skeletal muscle cells to increase the expression of Twist1, in turn inducing the expression of the muscle-specific ubiquitin ligases (MuRF1 and Atrogin1) to drive muscle protein degradation and subsequent muscle cachexia in PDAC [49.50]. These studies highlight that Twist1 plays a dual role in PDAC, acting in the tumor to orchestrate invasion and metastasis, and in muscle to orchestrate PDAC-associated muscle cachexia.

Endogenous interaction between TGIF1 and Twist1 has been detected in pancreatic extracts of wild-type mice, and as expected, no such interaction was found in pancreatic extracts obtained from *Tgif1* null mice [39]. Ablating *Tgif1* resulted in an increased abundance of the Twist1 protein, while forced expression of TGIF1 reduced the levels of endogenous Twist1 protein and mRNA. Of note, the TGIF1-Twist1 interaction seemed to be independent of TGF- β signaling [39]. Interestingly, Twist1 can induce its own expression, and such auto-transcriptional activity could be blocked by TGIF1. Furthermore, strong binding of endogenous TGIF1 to the Twist1 promoter has been detected both in vitro and in vivo. Based on these and other findings, it appears likely that TGIF1 acts as a direct transcriptional repressor for the *Twist1* gene.

Earlier studies have shown that Twist1 promotes tumorigenesis by inducing cell invasion and metastasis, possibly through creating a microenvironment for EMT [51–54]. A molecular hallmark of EMT is manifested by the loss of E-cadherin expression [55]. Twist1 is thought to reduce the expression of E-cadherin, which leads to disassembly of the epithelial cell–cell interaction to help in cell invasion and migration. Twist1-induced suppression of Ecadherin expression could be blunted by inducing the expression of TGIF1, and this phenomenon is associated with reduced PDAC metastasis [39]. Collectively, these observations implicate TGIF1 as a novel tumor suppressor gene in PDAC, likely functioning through suppression of Twist1 to restrain PDAC progression and metastasis.

In addition to promoting tumor invasion and metastasis, Twist1 could also help in the growth of various human tumors by modulating the functions of several tumor suppressors and oncogenic signaling pathways [48,56]. For instance, Twist1 has been found to directly suppress the expression of the tumor suppressor p16Ink4A, thereby allowing tumor cells to escape cell senescence, which is essential for tumorigenesis in Kras^{G12D} mutant mice [48]. Consistent with this notion, genetically inactivating Twist1 in Kras^{G12D} mice completely blocked the PDAC phenotype [39]. Twist1 inactivation in Kras^{G12D} mice with targeted pancreatic deletion of Tgif1 resulted in increased expression of Cadherin-1 and p16Ink4A, concurring with concomitantly decreased expression of the mesenchymal marker Vimentin [39]. The underlying mechanism of how and why TGIF1 inactivation accelerates Kras^{G12D}induced progression of PDAC appeared to be linked directly to Twist1 hyperactivation. In the absence of TGIF1, sustained Twist1 occurs and ultimately promotes tumor growth, invasion, and metastasis to increase the overall mortality in PDAC. In patients with PDAC patients, a 3-fold higher level of Twist1 has been detected in tumor tissues as compared to the healthy normal tissues [57], further supporting the oncogenic role of Twist1 in human PDAC.

Most of the tumor cells utilize aerobic glycolysis to induce uncontrolled proliferation and perhaps evade cell death. Unlike normal cells, tumor cells preferentially metabolize glucose (glycolysis) instead of (mitochondrial) oxidative phosphorylation to generate energy, even when oxygen is sufficiently available, a phenomenon called aerobic glycolysis (Warburg effect) [58]. Warburg effect can create a tumor microenvironment favorable to cancer cell proliferation. The Warburg effect changes reactive oxygen species (ROS) production, and dysregulated ROS can influence cell signaling cascade by impacting phosphatase and tensin homolog (PTEN) and tyrosine phosphatases activities to create a mitogenic milieu for the tumor cells [59]. Of note, Twist1 is a regulator of aerobic glycolysis in PDAC. Using human pancreatic cancer cell lines, Twist1 has shown to transcriptionally regulate the expression of key glycolytic genes, such as GLUT1, HK2, ENO1, and PKM2 to promote the Warburg effect [60]. Whether TGIF1 functions to limit this pro-tumorigenic activity of Twist1 remains to be established. Besides promoting Twist1 activity, inactivation of TGIF1 could create a microenvironment for the generation of tumor-associated macrophages (TAMs), which in turn contribute to the growth of PDAC [40]. In fact, TGIF1 inactivation results in the increased production of certain cytokines and chemokines in the murine model of PDAC. Most notably, tumor cells produce colony-stimulating factor-1 (CSF-1), vascular endothelial growth factor (VEGF), C-C motif chemokine ligand 2 (CCL2), IL-4, IL-10, IL-13, and TGF-B among others. These factors can act as chemoattractant to recruit monocytes and eventually differentiate those cells into the M2like macrophage phenotype [61]. In PDAC patients, a higher number of M2-like TAMs has been reported to be associated with metastasis and poor prognosis. In studies using CD68 as a panmacrophage marker and CD204 as a marker for M2 macrophage, increased numbers of M2-phenotype macrophages were detected in patients with invasive PDAC, as compared to the patients with chronic pancreatitis [62]. Increased numbers of M2 macrophages correlated with the enlarged tumor size, metastatic behavior, and shortened survival of PDAC patients [62]. From a translational perspective, suppressing macrophage recruitment resulted in reduced hepatic metastasis in the experimental model of the pancreatic tumor [63], while enhancing macrophage phenotype towards an M2 phenotype resulted in an increased metastatic spreading [64]. Again, loss of TGIF1 function in PDAC could induce TAM polarization towards the M2-like macrophage phenotype to promote pancreatic tumorigenesis [40].



Fig. 2. Simplified diagram showing various cellular events that are mediated by the disruption of the TGIF1-Twist1 axis in the genesis of PDAC.

5. Conclusion

Recent studies from independent groups have convincingly shown that enhancing TGF-B signaling by inactivating *Tgif1* in the pancreatic epithelium resulted in a highly aggressive and metastatic PDAC phenotype in mice, partly mediated through facilitating Kras-driven tumorigenesis. Although there was no death in pancreatic deleted Tgif1 mice during the entire observation period exceeding 18 months, all the *Kras^{G12D};Tgif1^{KO}* double mutant mice died within 19 weeks [39]. These results clearly suggest that enhanced TGF-β signaling, due to selective pancreatic inactivation of Tgif1, could not avert the genesis of PDAC, when Kras activity is normal. The available evidence implicates TGIF1 as a tumor suppressor in PDAC owing to its ability to inhibit the progression of Kras-initiated pancreatic tumor formation, possibly by inhibiting the formation of EMT and consequently minimizing metastasis (Fig. 2). The tumor suppressor effects of TGIF1 are partly exerted through antagonizing the pro-tumorigenic activity of Twist1, which is known to play key roles in malignant transformation and tumor progression and metastasis. Manipulating the TGIF1-Twist1 interaction and subsequent signaling might be a valid therapeutic target to reduce disease burden for the patients with PDAC. In this context, recent studies showed that inhibition of Twist1 activity by the small molecule IO1 was able to restrain tumor growth in vivo [65], underscoring Twist1 as an attractive candidate target for anti-cancer therapy. Of particular relevance, JQ1 analogs are currently under clinical trials for a variety of malignancies, hinting at the possibility that targeting the TGIF1-Twist1 axis could hold promise for designing breakthrough therapeutic strategies with immediate clinical applicability in fatal PDAC.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

Thanks to Dr. Nuraly Akimbekov of Al-Farabi Kazakh National University (Kazakhstan) for help in drawing the illustrations, and to Ms. Margo Wolfe for proofreading the manuscript and providing useful suggestions. The original works that formed the basis of this article were supported by the NIH grants (R01CA210911 and R01CA240484) to A.A.

References

- Jemal A, Siegel R, Xu J, Ward E. Cancer statistics. CA Cancer J Clin 2010;60 (2010):277–300.
- [2] Kern SE, Shi C, Hruban RH. The complexity of pancreatic ductal cancers and multidimensional strategies for therapeutic targeting. J Pathol 2011;223:295–306.
- [3] Hezel AF, Kimmelman AC, Stanger BZ, Bardeesy N, Depinho RA. Genetics and biology of pancreatic ductal adenocarcinoma. Genes Dev 2006;20:1218–49.
- [4] Vincent A, Herman J, Schulick R, Hruban RH, Goggins M. Pancreatic cancer. Lancet 2011;378:607–20.
- [5] Winter JM, Brennan MF, Tang LH, D'Angelica MI, Dematteo RP, Fong Y, et al. Survival after resection of pancreatic adenocarcinoma: results from a single institution over three decades. Ann Surg Oncol 2012;19:169–75.
- [6] Quante AS, Ming C, Rottmann M, Engel J, Boeck S, Heinemann V, et al. Projections of cancer incidence and cancer-related deaths in Germany by 2020 and 2030. Cancer Med 2016;5:2649–56.
- [7] Rahib L, Smith BD, Aizenberg R, Rosenzweig AB, Fleshman JM, Matrisian LM. Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. Cancer Res 2014;74:2913–21.
- [8] Font-Burgada J, Sun B, Karin M. Obesity and cancer: the oil that feeds the flame. Cell Metab 2016;23:48–62.
- [9] Rahn S, Zimmermann V, Viol F, Knaack H, Stemmer K, Peters L, et al. Diabetes as risk factor for pancreatic cancer: Hyperglycemia promotes epithelial-

mesenchymal-transition and stem cell properties in pancreatic ductal epithelial cells. Cancer Lett 2018;415:129–50.

- [10] Almoguera C, Shibata D, Forrester K, Martin J, Arnheim N, Perucho M. Most human carcinomas of the exocrine pancreas contain mutant c-K-ras genes. Cell 1988;53:549–54.
- [11] Iacobuzio-Donahue CA, Velculescu VE, Wolfgang CL, Hruban RH. Genetic basis of pancreas cancer development and progression: insights from whole-exome and whole-genome sequencing. Clin Cancer Res 2012;18:4257–65.
- [12] Hingorani SR, Petricoin EF, Maitra A, Rajapakse V, King C, Jacobetz MA, et al. Preinvasive and invasive ductal pancreatic cancer and its early detection in the mouse. Cancer Cell 2003;4:437–50.
- [13] Tuveson DA, Shaw AT, Willis NA, Silver DP, Jackson EL, Chang S, et al. Endogenous oncogenic K-ras(G12D) stimulates proliferation and widespread neoplastic and developmental defects. Cancer Cell 2004;5:375–87.
- [14] Heldin CH, Miyazono K, ten Dijke P. TGF-beta signalling from cell membrane to nucleus through SMAD proteins. Nature 1997;390:465–71.
- [15] Wu G, Chen YG, Ozdamar B, Gyuricza CA, Chong PA, Wrana JL, et al. Structural basis of Smad2 recognition by the Smad anchor for receptor activation. Science 2000;287:92–7.
- [16] Wrana JL, Attisano L, Wieser R, Ventura F, Massague J. Mechanism of activation of the TGF-beta receptor. Nature 1994;370:341–7.
- [17] Shi Y, Massague J. Mechanisms of TGF-beta signaling from cell membrane to the nucleus. Cell 2003;113:685–700.
- [18] Ogino S, Kawasaki T, Ogawa A, Kirkner GJ, Loda M, Fuchs CS. TGFBR2 mutation is correlated with CpG island methylator phenotype in microsatellite instability-high colorectal cancer. Hum Pathol 2007;38:614–20.
- [19] Takayama T, Miyanishi K, Hayashi T, Sato Y, Niitsu Y. Colorectal cancer: genetics of development and metastasis. J Gastroenterol 2006;41:185–92.
- [20] Calon A, Espinet E, Palomo-Ponce S, Tauriello DV, Iglesias M, Cespedes MV, et al. Dependency of colorectal cancer on a TGF-beta-driven program in stromal cells for metastasis initiation. Cancer Cell 2012;22:571–84.
- [21] Friedman E, Gold LI, Klimstra D, Zeng ZS, Winawer S, Cohen A. High levels of transforming growth factor beta 1 correlate with disease progression in human colon cancer. Cancer Epidemiol Biomarkers Prev 1995;4:549–54.
- [22] Batlle R, Andrés E, Gonzalez L, Llonch E, Igea A, Gutierrez-Prat N, et al. Regulation of tumor angiogenesis and mesenchymal–endothelial transition by p38α through TGF-β and JNK signaling. Nat Commun 2019;10:3071.
- [23] Goumans MJ, Liu Z, ten Dijke P. TGF-beta signaling in vascular biology and dysfunction. Cell Res 2009;19:116–27.
- [24] Principe DR, Doll JA, Bauer J, Jung B, Munshi HG, Bartholin L, et al. TGF-beta: duality of function between tumor prevention and carcinogenesis. J Natl Cancer Inst 106 2014:djt369.
- [25] Drabsch Y, ten Dijke P. TGF-beta signalling and its role in cancer progression and metastasis. Cancer Metastasis Rev 2012;31:553–68.
- [26] Tian M, Neil JR, Schiemann WP. Transforming growth factor-beta and the hallmarks of cancer. Cell Signal 2011;23:951–62.
- [27] Wendt MK, Tian M, Schiemann WP. Deconstructing the mechanisms and consequences of TGF-beta-induced EMT during cancer progression. Cell Tissue Res 2012;347:85–101.
- [28] Parajuli P, Nguyen TL, Prunier C, Razzaque MS, Xu K, Atfi A. Pancreatic cancer triggers diabetes through TGF-beta-mediated selective depletion of islet betacells. Life Sci Alliance 2020;3.
- [29] Seo SR, Ferrand N, Faresse N, Prunier C, Abecassis L, Pessah M, et al. Nuclear retention of the tumor suppressor cPML by the homeodomain protein TGIF restricts TGF-beta signaling. Mol Cell 2006;23:547–59.
- [30] Seo SR, Lallemand F, Ferrand N, Pessah M, L'Hoste S, Camonis J, et al. The novel E3 ubiquitin ligase Tiul1 associates with TGIF to target Smad2 for degradation. EMBO J 2004;23:3780–92.
- [31] Bartholin L, Melhuish TA, Powers SE, Goddard-Leon S, Treilleux I, Sutherland AE, et al. Maternal Tgif is required for vascularization of the embryonic placenta. Dev Biol 2008;319:285–97.
- [32] Prunier C, Zhang MZ, Kumar S, Levy L, Ferrigno O, Tzivion G, et al. Disruption of the PHRF1 tumor suppressor network by PML-RARalpha drives acute promyelocytic Leukemia pathogenesis. Cell Rep 2015.
 [33] El-Jaick KB, Powers SE, Bartholin L, Myers KR, Hahn J, Orioli IM, et al.
- [33] El-Jaick KB, Powers SE, Bartholin L, Myers KR, Hahn J, Orioli IM, et al. Functional analysis of mutations in TGIF associated with holoprosencephaly. Mol Genet Metab 2007;90:97–111.
- [34] Kuang C, Xiao Y, Yang L, Chen Q, Wang Z, Conway SJ, et al. Intragenic deletion of Tgif causes defects in brain development. Hum Mol Genet 2006;15:3508–19.
- [35] Hamid R, Patterson J, Brandt SJ. Genomic structure, alternative splicing and expression of TG-interacting factor, in human myeloid leukemia blasts and cell lines. BBA 2008;1779:347–55.
- [36] Zhang MZ, Ferrigno O, Wang Z, Ohnishi M, Prunier C, Levy L, et al. TGIF governs a feed-forward network that empowers Wnt signaling to drive mammary tumorigenesis. Cancer Cell 2015;27:547–60.
- [37] Yeh BW, Wu WJ, Li WM, Li CC, Huang CN, Kang WY, et al. Overexpression of TG-interacting factor is associated with worse prognosis in upper urinary tract urothelial carcinoma. Am J Pathol 2012;181:1044–55.

Computational and Structural Biotechnology Journal 18 (2020) 2568-2572

- [38] Massague J. TGFbeta in Cancer. Cell 2008;134:215-30.
- [39] Parajuli P, Singh P, Wang Z, Li L, Eragamreddi S, Ozkan S, et al. TGIF1 functions as a tumor suppressor in pancreatic ductal adenocarcinoma. EMBO J 2019;38: e101067.
- [40] Weng CC, Hsieh MJ, Wu CC, Lin YC, Shan YS, Hung WC, et al. Loss of the transcriptional repressor TGIF1 results in enhanced Kras-driven development of pancreatic cancer. Mol Cancer 2019;18:96.
- [41] Qin Q, Xu Y, He T, Qin C, Xu J. Normal and disease-related biological functions of Twist1 and underlying molecular mechanisms. Cell Res 2012;22:90–106.
- [42] Thisse B, el Messal M, Perrin-Schmitt F. The twist gene: isolation of a Drosophila zygotic gene necessary for the establishment of dorsoventral pattern. Nucleic Acids Res 1987;15:3439–53.
- [43] Wolf C, Thisse C, Stoetzel C, Thisse B, Gerlinger P, Perrin-Schmitt F. The Mtwist gene of Mus is expressed in subsets of mesodermal cells and is closely related to the Xenopus X-twi and the Drosophila twist genes. Dev Biol 1991;143:363–73.
- [44] Wang SM, Coljee VW, Pignolo RJ, Rotenberg MO, Cristofalo VJ, Sierra F. Cloning of the human twist gene: its expression is retained in adult mesodermallyderived tissues. Gene 1997;187:83–92.
- [45] Pan D, Fujimoto M, Lopes A, Wang YX. Twist-1 is a PPARdelta-inducible, negative-feedback regulator of PGC-1alpha in brown fat metabolism. Cell 2009;137:73–86.
- [46] el Ghouzzi V, Le Merrer M, Perrin-Schmitt F, Lajeunie E, Benit P, Renier D, et al. Mutations of the TWIST gene in the Saethre-Chotzen syndrome. Nat Genet 1997;15:42–6.
- [47] Howard TD, Paznekas WA, Green ED, Chiang LC, Ma N, Ortiz de Luna RI, et al. Mutations in TWIST, a basic helix-loop-helix transcription factor, in Saethre-Chotzen syndrome. Nat Genet 1997;15:36–41.
- [48] Lee KE, Bar-Sagi D. Oncogenic KRas suppresses inflammation-associated senescence of pancreatic ductal cells. Cancer Cell 2010;18:448–58.
- [49] Parajuli P, Kumar S, Loumaye A, Singh P, Eragamreddy S, Nguyen TL, et al. Twist1 Activation in Muscle Progenitor Cells Causes Muscle Loss Akin to Cancer Cachexia. Dev Cell 2018;45:712–725 e716.
- [50] Razzaque MS, Atfi A. Regulatory role of the transcription factor Twist1 in cancer-associated muscle cachexia. Front Physiol 2020;11:662. <u>https://doi. org/10.3389/fphys.2020.00662</u>.
- [51] D'Angelo RC, Liu XW, Najy AJ, Jung YS, Won J, Chai KX, et al. TIMP-1 via TWIST1 induces EMT phenotypes in human breast epithelial cells. Mol Cancer Res 2014;12:1324–33.
- [52] Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, et al. The epithelialmesenchymal transition generates cells with properties of stem cells. Cell 2008;133:704–15.
- [53] Martin A, Cano A. Tumorigenesis: Twist1 links EMT to self-renewal. Nat Cell Biol 2010;12:924–5.
- [54] Valastyan S, Weinberg RA. Tumor metastasis: molecular insights and evolving paradigms. Cell 2011;147:275–92.
- [55] Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. Cell 2009;139:871–90.
- [56] Piccinin S, Tonin E, Sessa S, Demontis S, Rossi S, Pecciarini L, et al. A "twist box" code of p53 inactivation: twist box: p53 interaction promotes p53 degradation. Cancer Cell 2012;22:404–15.
- [57] Wang J, Nikhil K, Viccaro K, Chang L, Jacobsen M, Sandusky G, et al. The Aurora-A-Twist1 axis promotes highly aggressive phenotypes in pancreatic carcinoma. J Cell Sci 2017;130:1078–93.
- [58] Martinez-Outschoorn UE, Peiris-Pages M, Pestell RG, Sotgia F, Lisanti MP. Cancer metabolism: a therapeutic perspective. Nat Rev Clin Oncol 2017;14:11-31.
- [59] Locasale JW, Cantley LC. Metabolic flux and the regulation of mammalian cell growth. Cell Metab 2011;14:443–51.
- [60] Wang XX, Yin GQ, Zhang ZH, Rong ZH, Wang ZY, Du DD, et al. TWIST1 transcriptionally regulates glycolytic genes to promote the Warburg metabolism in pancreatic cancer. Exp Cell Res 2020;386:111713.
- [61] Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. Cell 2010;141:39–51.
 [62] Yoshikawa K, Mitsunaga S, Kinoshita T, Konishi M, Takahashi S, Gotohda N,
- [62] Yoshikawa K, Mitsunaga S, Kinoshita T, Konishi M, Takahashi S, Gotohda N, et al. Impact of tumor-associated macrophages on invasive ductal carcinoma of the pancreas head. Cancer Sci 2012;103:2012–20.
- [63] Mitchem JB, Brennan DJ, Knolhoff BL, Belt BA, Zhu Y, Sanford DE, et al. Targeting tumor-infiltrating macrophages decreases tumor-initiating cells, relieves immunosuppression, and improves chemotherapeutic responses. Cancer Res 2013;73:1128–41.
- [64] Tugues S, Honjo S, Konig C, Noguer O, Hedlund M, Botling J, et al. Genetic deficiency in plasma protein HRG enhances tumor growth and metastasis by exacerbating immune escape and vessel abnormalization. Cancer Res 2012;72:1953–63.
- [65] Shi J, Wang Y, Zeng L, Wu Y, Deng J, Zhang Q, et al. Disrupting the interaction of BRD4 with diacetylated Twist suppresses tumorigenesis in basal-like breast cancer. Cancer Cell 2014;25:210–25.