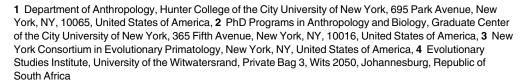




Papio Cranium from the Hominin-Bearing Site of Malapa: Implications for the Evolution of Modern Baboon Cranial Morphology and South African Plio-Pleistocene Biochronology

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Abstract

A new partial cranium (UW 88-886) of the Plio-Pleistocene baboon *Papio angusticeps* from Malapa is identified, described and discussed. UW 88-886 represents the only non-hominin primate yet recovered from Malapa and is important both in the context of baboon evolution as well as South African hominin site biochronology. The new specimen may represent the first appearance of modern baboon anatomy and coincides almost perfectly with molecular divergence date estimates for the origin of the modern *P. hamadryas* radiation. The fact that the Malapa specimen is dated between ~2.026–2.36 million years ago (Ma) also has implications for the biochronology of other South African Plio-Pleistocene sites where *P. angusticeps* is found.

Introduction

The recent discovery of fossiliferous deposits at Malapa, South Africa, has had a major impact on the study of human evolution [1–3]. Over the past few years, three *Australopithecus sediba* partial skeletons have been described [1, 4–13], and the preparation of additional specimens (including other partial skeletons) is currently underway. Along with the hominin material, numerous faunal specimens from Malapa have also been recovered [2, 14–15], and most are still awaiting formal analysis and description. The description and analysis of the fauna are crucial to understanding aspects of paleoecology, biochronology, and mammalian evolution during the African Plio-Pleistocene. In this paper, we describe the only known non-hominin primate specimen from Malapa, UW 88–886, a specimen of the fossil baboon *Papio angusticeps*. Furthermore, we evaluate *Papio angusticeps* in the broader context of modern baboon craniodental evolution and South African Plio-Pleistocene biochronology.



study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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Geological context

Malapa cave is hosted in the stromatolite-rich, chert-free dolomite of the Lyttelton Formation of the late Archaean (2.64–2.5 Ga) Malmani Subgroup [2], and presently lies near the base of a cave system that was originally >30-m deep prior to erosion by valley incision. Minimal limestone mining in the early 20th century exposed in situ cave deposits [2]. Two shallow pits have been recognized at Malapa, of which the main (Pit 1) contains the well- preserved and welldated (\sim 1.977 Ma) Au. sediba fossil remains [1-2]. The cave deposits comprise six distinct stratigraphic lithofacies assigned, A to F, from base to top [3]. Facies A and B represent the oldest deposits that were partially eroded before they were covered by flowstone (Flowstone 1, Fig 1). Facies D overlies Flowstone 1, followed by Facies E and topped by Facies F, while Facies C occurs as a remnant between the dolomite on the western sidewall of Pit 1 and Facies B and E (and possibly D) [3]. Flowstone 1 is dated to 2.026 ± 0.021 Ma, suggesting that Facies A and B are older than 2 million years [2]. The *P. angusticeps* partial cranium was found in a block of calcified clastic sediments within two metres of Pit 1 and was almost certainly moved to this position by early mining activities. The cranium is embedded within fine-grained clastic dark brown sediment with clastic sandstone. It is unlikely that the specimen comes from Facies A, D and E for a couple of reasons. First, Facies A and D are comprised of course grain sediments, both abundant in fossil remains, and second, Facies E sediments are best characterized as coarse to fine upwards and also contain abundant fossil remains. It is also unlikely that the specimen derives from Facies C and F, since Facies C lacks clastic intercalations while Facies F is devoid of sandstone intercalations. We also considered the possibility that the specimen came from a nearby cave. Indeed, there are two caves in close proximity to Malapa, both less than 150 metres south of the Malapa pits. Currently there are no indications of fossil remains in situ or in the mining dumps near these caves.

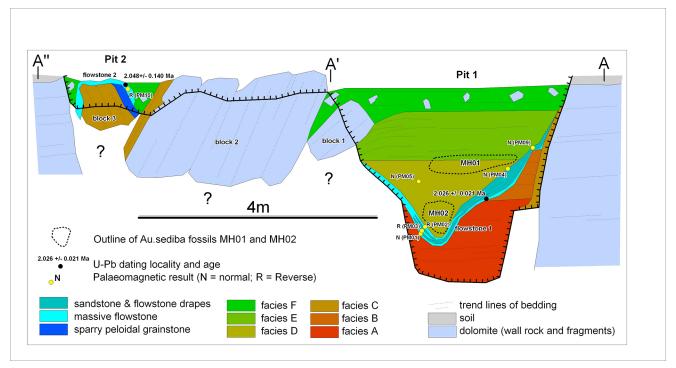


Fig 1. Cross-section sketch map through Malapa showing facies A-F and the U-Pb samples. Modified from [3].

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Therefore, it is most parsimonious to presume that the block containing *P. angusticeps* was removed from Pit 1, from Facies B, and the resulting age of this specimen is thus $> 2.026 \pm 0.021$ Ma based on the U-Pb dates of the overlying Flowstone 1 (Fig 1). Geological studies indicate that the different Facies were deposited in quick succession of each other with limited transport distance [2], and the discovery of *Equus* sp., which has its earliest recorded appearance in Africa at ~ 2.36 Ma [16–18], in Facies D, provides a probable maximum age for the site more generally. Therefore, the probable maximum age for this specimen is ~ 2.36 million years ago (Ma) based on the appearance of *Equus* sp. in Facies D [2].

Material and Methods

UW 88-886 is housed in the Evolutionary Studies Institute at the University of Witwatersrand in Johannesburg, Republic of South Africa. Where available, observations and measurements were taken on original specimens using digital calipers and recorded to the nearest tenth of a millimeter. Additional comparative observations and measurements were taken from highquality casts, the online PRIMO database (access courtesy of Eric Delson), as well as from data kindly provided by Mark Collard to CCG (see Supporting Information, Table A in S1 Dataset). To test for statistical significance between various Papio populations, one-way ANOVAs with post-hoc comparisons were computed. Comparative specimens were examined by the authors at the University of Witwatersrand Anatomy Department, Johannesburg (UW-AD), the Bernard Price Institute, Johannesburg (BPI), the Evolutionary Studies Institute, Johannesburg (ESI), the Ditsong National Museum of Natural History, Pretoria (formerly the Transvaal Museum, TM), the Iziko South African Museum, Cape Town (SAM), the University of California Museum of Paleontology, Berkeley (UCMP), and the American Museum of Natural History (AMNH), New York. Bernhard Zipfel, Bonita de Klerk, Steph Potze, Graham Avery and Kerwin von Willingh, Pat Holroyd and Leslea Hlusko, and Eric Delson and Eileen Westwig kindly provided access to primate specimens at UW-AD, BPI/ESI, TM, SAM, UCMP, and AMNH, respectively. For raw measurements used in this study, please see the Supporting Information (Table A in S1 Dataset). All necessary permits and permissions were obtained for the described study, which complied with all relevant regulations. We thank the Nash family and the Malapa Nature Reserve for permission to access the site. We thank the South African Heritage Resource Agency for permission to excavate, Permit No. 80/08/09/001/51 to LRB and JMK.

Systematic paleontology

Order Primates Linnaeus, 1758
Suborder Anthropoidea Mivart, 1864
Infraorder Catarrhini E. Geoffroy, 1812
Superfamily Cercopithecoidea Gray, 1821
Family Cercopithecidae Gray, 1821
Subfamily Cercopithecinae Gray, 1821
Tribe Papionini Burnett, 1828
Genus Papio Erxleben, 1777
Papio angusticeps (Broom, 1940)

Preservation. The specimen is sheared nearly in half along an approximate sagittal plane thus preserving most of the right part of the cranium. There is little postdepositional compaction with minor broken areas in the maxillary and infratemporal fossae. The preserved portion of the cranium is approximately half-filled with fine sediment capped by a thin layer of flowstone; stage 1 weathering is apparent [19].





Fig 2. Photographs of UW 88–886, male *P. angusticeps* in lateral (top left), oblique (top right), dorsal (middle), and inferior (bottom) views. Note the strong maxillary ridges, deep maxillary fossae, strong temporal lines, and tall malar region, distinctive of *P. angusticeps* males. Scale bar in each panel = 1 cm.

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Description. U.W. 88–886 is a partial cranium preserving most of the right facial skeleton and the right half of the neurocranium (Figs $\underline{2}$ and $\underline{3}$). The overall size, strong maxillary ridges, and strong temporalis muscle markings suggest the specimen is most likely a male. In addition,



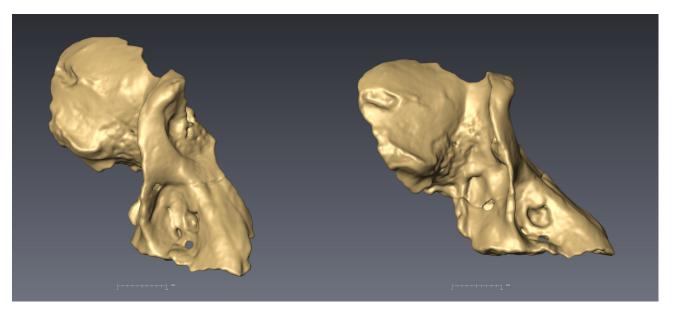


Fig 3. CT Scans of UW 88–866 in oblique (left) and lateral (right) views. Again, note the strong maxillary ridges, deep maxillary fossae, strong temporal lines, and tall malar region, distinctive of *P. angusticeps* males.

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the overall size of the specimen, apparent anteorbital drop, long and narrow muzzle, strong maxillary ridges, deep maxillary fossae, and tall malar region make the specimen referable to *Papio* among Plio-Pleistocene cercopithecoid taxa, generally, and *P. angusticeps* among Plio-Pleistocene *Papio* taxa, more specifically (<u>Table 1</u>). Unfortunately, the dentition is not preserved, with the exception of remnants of the M² and M³ roots. The facial skeleton includes

Table 1. Summary of features found in UW 88-886 compared with modern and Plio-Pleistocene Papio taxa.

		<u> </u>		<u> </u>	
	UW-88-886	P. angusticeps	P. izodi	P. robinsoni	P. hamadryas ssp.
Maxillary fossae development	Well-developed, invade infraorbital plate	Well-developed, invade infraorbital plate	Absent to weakly developed in males, weakly to well-developed and extending up to the infraorbital plate in females	Slightly to well- developed, extend up to infraorbital plate	Well-developed, extend up to and sometimes invade infraorbital plate
Maxillary ridge development in males	Present	Present	Absent to weakly present	Present	Present
Mandibular corpus fossa development	X	Present and well- developed in males, variably present in females	Absent	Present, slightly developed	Present and well-developed in males, variably present to present and well-developed in females
Relative malar height	Tall	Tall	Short	Tall	Tall
Anteorbital Drop	Present?	Present	Variably present	Present	Present
Muzzle shape	Long and narrow	Long and narrow	Short and broad	Long and narrow	Long and narrow
Relative tooth size	X	Small	Large	Small	Small
Relative orbit size	Intermediate	Small	Large	Small	Small
Estimated body mass	~21 kg?	~21 kg (Males), ~15 kg (Females)	~20 kg (Males) ~15 kg (Females)	~29 kg (Males) ~18 kg (Females)	~16–32 kg (Males) ~10–16 kg (Females)

Notes: See text and figures for additional details. Body mass estimates from [39].

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Table 2. Comparison of selected morphological features in "small-bodied" Papio species.

Taxon	Feature	n	Mean	Range	Significance
Papio izodi	Orbit Height	10	27.4	24–33	> P. angusticeps, p < 0.01 > P. h. cynocephalus, p < 0.01
P. angusticeps	Orbit Height	5	21.8	20–24	< P. izodi, p < 0.01 P. h. cynocephalus, n.s.
P. h. cynocephalus	Orbit Height	21	23.8	18–27	< P. izodi, p < 0.01 P. angusticeps, n.s.
UW 88-886	Orbit Height	1	(27.2)	-	-
Papio izodi	Orbit Area	8	780	624–957	> P. angusticeps, p < 0.01 P. h. cynocephalus, n.s.
P. angusticeps	Orbit Area	5	576	542-629	< P. izodi, p < 0.01 P. h. cynocephalus, n.s.
P. h. cynocephalus	Orbit Area	6	711	577–870	P. izodi, n.s. P. angusticeps, n.s.
UW 88-886	Orbit Area	1	(707)	-	-
Papio izodi	Relative Malar Height	7	0.94	0.79-1.04	< P. angusticeps, p = 0.02 < P. h. cynocephalus, p < 0.01
P. angusticeps	Relative Malar Height	5	1.29	1.08-1.51	> P. izodi, p = 0.02 P. h. cynocephalus, n.s.
P. h. cynocephalus	Relative Malar Height	21	1.22	0.80-1.65	> P. izodi, p < 0.01 P. angusticeps, n.s.
UW 88-886	Relative Malar Height	1	(1.26)	-	-

Notes: Results from one-way ANOVA with Tukey's Honestly Significant Difference post-hoc comparisons for those variables with equal variances and Games-Howell post-hoc comparisons for those variables with unequal variances. Because orbit height, orbit area, and malar height all scale allometrically, the most meaningful comparisons are among taxa of similar body size. The estimated mass for *P. angusticeps* averages ~21 kg for males and ~15 for females [39]. *P. izodi* is estimated at ~20 kg for males and ~15 for females [39]. The most similar extant taxon in terms of body mass is *P. h.* cynocephalus, ~23 kg for males and 12.5 kg for females, which is why *P. h. cynocephalus* is used in the above comparisons. All specimens were pooled regardless of sex in order to increase sample size. For sex-specific values, see Table 3. n.s. = non-significant. Note that *P. angusticeps* and *P. h.* cynocephalus are both significantly different from *P. izodi*, but not from each other. Results for all comparisons are the same if UW 88–886 is included in the *P. angusticeps* sample. Orbit height defined as the maximum distance between the inferior and superior orbit borders. Orbit width defined as the maximum distance between the lateral and medial orbit borders. Orbit area is defined as orbit width x orbit height. Malar height defined as the distance between orbitale inferior/zygoorbitale and zygomaxillare inferior. Relative malar height defined as malar height/orbit height. *P. angusticeps* specimens include CO 100, CO 135A/B, CO 101, GV 4040, and HGD 1249. *P. izodi* specimens include TP 12, SAM 11728, T10, T13, UCMP 125854, UCMP 125855, UCMP 125856, STS 262, T89-11-1, and SWP UN-2. Values for each taxon represent averages. Numbers in parentheses represent estimates. For boxplots with ranges, see also Fig 4 and Table A in S1 Dataset.

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portions of the right orbit, right zygomatic, right nasal, and right half of the muzzle including most of the maxilla and a small remnant of the premaxilla (Figs 2 and 3). In addition to the right side of the face, fragments of right frontal, parietal, temporal and zygomatic arch are also preserved. The occipital, basicranium, most of the dorsal surface of the neurocranium, and left side of the cranium have not been recovered.

The upper part of the face is distinguished by tall, oval-shaped orbits. Because only half of the orbital rim is preserved (the inferior, lateral, and superolateral aspects), it is impossible to measure the exact size of the orbit, but the orbital height and width can be estimated (Tables 2 and 3; Figs 2-4). Thus, UW 88–886 appears to have large orbits compared to other specimens of *P. angusticeps* (a feature more typical of *Papio izodi* among Plio-Pleistocene *Papio* taxa), but additional specimens preserving the entire orbit are needed to confirm this morphology as typical of the Malapa population. Given the other features present in the specimen (see below) and the low sample size of known *P. angusticeps* orbits, it seems most reasonable to accept that UW 88–886 simply adds to the known variation of *P. angusticeps* at this time. In fact, including UW 88–886 in the existing *P. angusticeps* sample of orbit height measurements results in a range of variation (20–27 mm) similar to that seen in a small sample of extant *P. h. cynocephalus* (18–27 mm, variances statistically equivalent, Levene statistic p = 0.344; see Tables 2 and 3; Fig 4). From the preserved anatomy of the upper face and orbital region, particularly the nasals

Table 3. Available measurements (in mm) of UW 88-886 Compared with modern and Plio-Pleistocene Papio taxa.

	M	٩	<i>9</i>	۹.	P. izodi	g.	P.h.	P.h.	P.h.	P. h.	P. h.	P. h.	P. h.	P. h.
	-88 886	<i>angusticeps</i> males		<i>izodi</i> males	females	<i>robinsoni</i> females	<i>cynocephalus</i> males	<i>cynocephalus</i> females	<i>kindae</i> males	<i>kindae</i> females	<i>anubis</i> males	<i>anubis</i> females	ursinus males	ursinus females
Orbit height	(27.2)	21.3 (n = 2)		28.7 (n = 4)		25.8 (n = 3)	24.5 (<i>n</i> = 14)	22.8 (n = 9)	24.7 (n = 8)	23.0 (n = 7)	25.6 (<i>n</i> = 11)	25.3 $(n = 12)$	25.6 (n = 6)	24.6 (n = 3)
Orbit	(26)	27.9 (n = 2)	25.5 (n = 3)	29.5 $(n = 2)$		29.6 (<i>n</i> = 3)	30.7 (n = 3)	27.6 (n = 3)	29.1 (n = 8)	26.7 (n = 7)	31.0 (n = 7)	29.0 $(n = 4)$	33.1 (n = 6)	28.6 ($n = 3$)
Orbit area	(707.2)	593.9 (n = 2)	563.5 (n = 3)	892.5 $(n = 2)$		763.0 $(n = 3)$	791.2 (n = 3)	630.0 (n = 3)	718.0 ($n = 8$)	611.8 $(n = 7)$	782.3 $(n = 7)$	678.6 $(n = 4)$	847.9 (n = 6)	702.9 $(n = 3)$
Malar height	34.2	31.3 (n = 3)	25.6 (<i>n</i> = 4)	26.1 $(n = 3)$	25.0 ($n = 4$)	33 (n = 1)	31.8 (<i>n</i> = 14)	24.6 (n = 9)	27.6 (n = 7)	23.6 $(n = 7)$	34.9 ($n = 10$)	29.5 ($n = 12$)	43.6 (<i>n</i> = 5)	32.7 $(n = 3)$
Relative malar height	(1.3)	1.4 (n = 2)	1.2 (n = 3)	0.9 (n = 3)		1.2 (<i>n</i> = 1)	1.3 $(n = 14)$	1.1 (n = 9)	1.1 (n = 7)	1.0 $(n = 7)$	1.4 (n = 10)	1.2 $(n = 12)$	$\frac{1.7}{(n=5)}$	1.3 $(n = 3)$

Notes: Orbit height defined as the maximum distance between the inferior and superior orbit borders. Orbit width defined as the maximum distance between the lateral and medial zygomaxillare inferior. Relative malar height defined as malar height/orbit height. P. angusticeps specimens include KA 194, KA 195, CO 100, CO 135A/B, CO 101, GV 4040, and specimens include UCMP 56797, UCMP 56786, and M3147. No measurements were available for P. robinsoni males. Numbers in parentheses represent estimates. Values for HGD 1249. P. izodi specimens include TP 7, TP 12, SAM 11728, T10, T13, UCMP 125854, UCMP 125855, UCMP 125856, STS 262, T89-11-1, and SWP UN-2. P. robinsoni orbit borders. Orbit area is defined as orbit width x orbit height and is measured in mm². Malar height defined as the distance between orbitale inferior/zygoorbitale and each taxon represent averages. See also Fig 4 and Table A in S1 Dataset.

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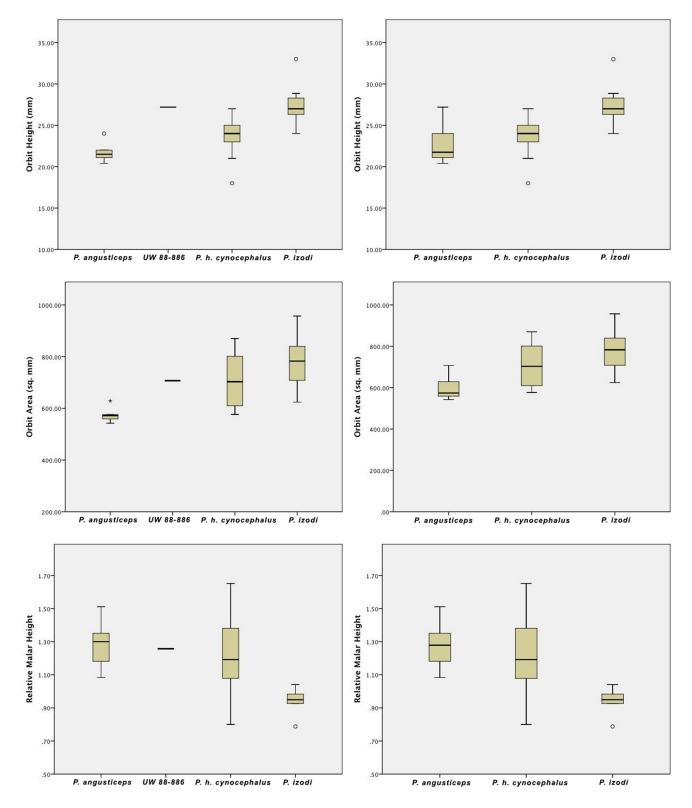


Fig 4. Available craniometric comparisons in *P. angusticeps,* **UW 88–886,** *P. h. cynocephalus,* **and** *P. izodi.* Top Row: Boxplots of orbit height considering UW 88–886 separately (left) and within *P. angusticeps* (right). Note that *P. izodi* has significantly taller orbits than both *P. angusticeps* and *P. h. cynocephalus.* UW 88–886 has tall orbits compared to other *P. angusticeps* specimens, but within a reasonable range of expected variation for a species. Middle Row: Boxplots of orbit area (mm²) considering UW 88–886 separately (left) and within *P. angusticeps* (right). *P. izodi* has significantly larger orbits



than *P. angusticeps*, but a non-significant difference compared to *P. h. cynocephalus*. The difference between *P. angusticeps* and *P. h. cynocephalus* is also non-significant. UW 88–886 appears to have large orbits compared to other *P. angusticeps* specimens, but again within a reasonable range of expected variation for a species. Bottow Row: Boxplots of relative malar height considering UW 88–886 separately (left) and within *P. angusticeps* (right). *P. izodi* has a significantly shorter malar height compared to *P. angusticeps* and *P. h. cynocephalus*, but the difference between *P. angusticeps* and *P. h. cynocephalus* is non-significant. UW 88–886 is closest to the average of other *P. angusticeps* specimens. See also Table 2 and Fig 5.

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and maxilla just medial to the infraorbital border, it appears that a pronounced anteorbital drop would have been present.

The zygomatic bone bulges outward slightly lateral to a series of infraorbital/maxillary foramina, then pinches inferiorly, forming sharp cheekbones bordering the pronounced infraorbital/maxillary fossa. The right lateral border of the orbit is intact from the frontal-zygomatic suture to the beginning of the temporal process of the zygomatic. The zygomatic arch is not preserved (Figs 2 and 3). Malar height, as characterized by the minimum distance from inferior orbital margin to the inferior border of zygomatic process of the maxilla, is relatively tall in UW 88–886, a feature it shares with other *P. angusticeps* specimens and similarly-sized modern *P. hamadrayas* baboons (e.g., *P. h. cynocephalus*) to the exclusion of *P. izodi* (Tables 1–3; Figs 4 and 5). On the lateral orbit border, there are two zygomatico-facial foramina, the larger in the central body of the zygoma and the smaller is placed superiorly near the frontal-zygomatic suture. The zygomatic root begins above the maxillary alveoli at the level of the M²–M³ contact. The lateral margin of the zygomatic appears to flare slightly outward (laterally).



Fig 5. Comparison of morphology in UW 88–886 (left), *P. angusticeps* males (CO 100, center), and *P. izodi* males (TP 89-11-1, right). Top: Lateral view, specimens scaled to approximately the same cranial height. Note the tall malar region (black bar), prominent maxillary ridges and deep maxillary fossae (white arrows) in UW 88–886 and *P. angusticeps* compared with *P. izodi*. Bottom: Dorsal view, specimens scaled to approximately the same cranial width. Note the longer, narrower muzzle in *P. angusticeps* compared to *P. izodi*, and again the prominent maxillary ridges and deep maxillary fossae in UW 88–886 and *P. angusticeps* compared with *P. izodi*.

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The preserved portion of the muzzle dorsum is flat to slightly rounded moving medially, and there is strong development of a prominent maxillary ridge bordering the lateral aspect of the muzzle dorsum. The superior rim of the nasal aperture is broken in this specimen; however, what is preserved of the superior portion of the rim appears to have been rounded. The premaxillae do not extend up along the nasals as far as in some other specimens of *P. angusticeps* (e.g., female CO 101). There is a slight groove medial to the canine juga extending to a large infraorbital foramen. Lateral and inferior to this foramen, there are several smaller maxillary foramina. The lateral portion of the muzzle is dominated by a deep and pronounced maxillary fossa extending into the zygomatic and maxillary bone inferior to the orbital margin (i.e., the infraorbital plate).

In dorsal view, there is evidence for a strong supraorbital region based on the preserved morphology of the right superolateral orbit border. The superolateral portion of the orbital rim is relatively flat and the supraorbital torus has a sharp border demarcated posteriorly by the temporal line with a distinct lip above the post-orbitally constricted frontal bone. There is marked post-orbital constriction with evidence of a probable post-orbital sulcus, not as much constriction as in *Theropithecus*, but a similar degree to that observed in typical male *P. angusticeps* and *Papio hamadryas* specimens (Figs 2–5). Also similar to *P. angusticeps* and modern *Papio hamadryas* males, in particular, UW 88–886 clearly displays pinched temporal lines as far medially as they are preserved, although it is unclear whether they would have met posteriorly to form a slight sagittal crest. The temporal lines emanate from the supero-lateral corner above the orbits medial to the post-orbital constriction of the frontals.

Discussion

Based on preserved morphology, UW 88–886, a presumed male, is most similar to the South African taxon *Papio angusticeps* among known Plio-Pleistocene baboon populations. The combination of an anteorbital drop, long and narrow muzzle, well-developed maxillary ridges, well-developed and deep maxillary fossae, and a relatively tall malar region make the specimen statistically distinct from other Plio-Pleistocene *Papio* species and most similar to *P. angusticeps* as well as modern *P. hamadryas* subspecies (Tables <u>1–3;</u> Figs <u>4</u> and <u>5</u>). The identification of *P. angusticeps* at Malapa has broader implications for baboon evolution and hominin biochronology as well.

Modern baboons (extant *P. hamadryas* ssp.) represent one of the most successful living primate radiations, with populations distributed all across sub-Saharan Africa and into the Arabian Peninsula. Despite their evolutionary success, modern baboon origins in the fossil record are not well understood or agreed upon. In East Africa prior to the Middle Pleistocene, there are craniodental fragments from various localities that possibly represent the genus *Papio*, but none of these fossils are definitively diagnostic (e.g., [20–25]). Fossils more clearly attributable to the modern *Papio hamadryas* population appear in the Middle Pleistocene at Olduvai Gorge, Asbole, and Bodo [26–29].

During the South African Plio-Pleistocene, at least two species of "small-bodied" papionins exist that have been typically placed into the genus *Papio*: *P. izodi* and *P. angusticeps*. *P. izodi* is currently recognized at Taung as well as Members 2 and 4 at Sterkfontein [30–33], suggesting an age range anywhere from ~3.7–2.0 Ma depending on the estimate (e.g., see [30, 34] for Taung; see [34–35] for most recent estimates of Members 2 and 4 at Sterkfontein). *P. angusticeps* on the other hand, is securely documented at sites generally suggested to be less than ~2.0 Ma, such as Kromdraai A and Cooper's A. One exception might be the *P. angusticeps* population at Haasgat, which was recently argued to be slightly older, between ~2.58–1.95 Ma, with a more likely range for most fossils between ~2.3–2.0 Ma [36–37]. The age of *P. angusticeps* at Gladysvale is difficult to assess at this time as much of the faunal material was initially collected from ex situ breccia blocks of unknown provenience [34].



While some authors lump both *P. izodi* and *P. angusticeps* into a single taxon (*P. izodi* sensu lato; e.g., [22, 38]), other experts recognize them as different taxa (e.g., [30–31, 39]). Some authors even suggest that *P. angusticeps* is most likely a subspecies of the modern baboon radiation (i.e., *P. hamadryas angusticeps*) and note many similarities with modern *P. h. kindae* and *P. h. cynocephalus* populations, in particular [30, 33, 40]. Similar to East Africa, specimens definitively attributable to *P. hamadryas* appear in southern Africa by the Middle Pleistocene, best represented by the distinct subspecies *P. h. botswanae* in Botswana [41] and the South African Pleistocene/Holocene *P. "spelaeus*" specimen of unknown provenience that appears to represent a large *P. h. ursinus* male [22, 41–42].

P. angusticeps shares features with modern P. hamadryas such as a relatively tall malar region, relatively small orbits, a definitive anteorbital drop, deep maxillary/suborbital fossae, pronounced maxillary ridges in males, a definitive anteorbital drop, a relatively narrow muzzle, a relatively long muzzle, and relatively small teeth (Table 1; Fig 5). When Freedman [42] transferred the species to Papio from Parapapio (originally Parapapio angusticeps Broom, 1940), he also noted that P. angusticeps cranial morphology closely resembles that of P. h. ursinus, except that the maxillary ridges are slightly weaker and the maxillary fossae are slightly deeper in P. angusticeps. P. izodi, on the other hand, is much more primitive in that it displays a relatively short malar region, relatively large orbits, a variable anteorbital drop, shallower maxillary/suborbital fossae (particularly in males), little to no development of the maxillary ridges in males, a relatively broad muzzle, a relatively short muzzle, and relatively large teeth (Tables 1 and 3; Fig 5; see also [30–31, 33]). In fact, one recent phylogenetic analysis places P. izodi outside of the crown African papionins and questions its status as a member of the modern genus Papio at all [33]. In any case, it is clear that the morphology of P. izodi is distinct from that of P. angusticeps as well as the modern baboon populations, P. hamadryas ssp. (Tables 1–3; Figs 4 and 5).

Excluding P. izodi, there are a few additional South African fossil Papio specimens that are relevant to the origins of the modern baboon. Papio robinsoni is a large form of Papio (larger on average than P. izodi, P. angusticeps, and most modern subspecies) securely documented at sites estimated to be anywhere between ~2.6–1.5 Ma: Swartkrans Member 1, Skurweburg, Bolt's Farm Pit 23, and Drimolen; it is also less securely identified at Sterkfontein Member 4 [30, 42-46]. While P. robinsoni has also been argued to be a member of the modern Papio hamadryas radiation by some (e.g., [30, 33, 38, 44]), there are possible reasons to suspect it is distinct. P. robinsoni clearly displays synapomorphies with modern P. hamadryas such as a distinct anteorbital drop, large overall size, definitive maxillary ridges, a relatively long and narrow muzzle, relatively small orbits, relatively small teeth, and definitive maxillary fossae in males (Tables 1 and 3). However, there are also consistent differences between P. robinsoni and modern P. hamadryas ssp. P. robinsoni displays weaker development of the maxillary and mandibular corpus fossae than modern baboons (Table 1), maxillae that occasionally meet in the midline (covering the nasals near nasion), and nasals that lie below the level of the maxillary ridges on the muzzle dorsum [40, 42]. Our interpretation is that these features are primitive relative to modern baboons, particularly the slight development of the facial fossae. P. angusticeps, on the other hand, displays no features that are obviously distinct from the modern P. hamadryas populations (see Tables 1 and 3), suggesting that it is possibly the first well-documented modern P. hamadryas subspecies in the fossil record (i.e., P. h. angusticeps). While P. angusticeps displays slightly deeper fossae and less-developed maxillary ridges on average when compared to P. h. ursinus [42], similar morphology to P. angusticeps can be found among the other P. hamadryas susbspecies. A more complete and formal taxonomic reassessment of P. angusticeps and P. robinsoni is beyond the scope of this paper, but future studies may further support the sinking of *P. angusticeps* into *P. hamadrayas* (as originally suggested by Delson [30]), depending on the species concept being used. In particular, given the quantitative



morphometric differences documented among modern *P. hamadryas* populations [41, 47], future 3D morphometric studies including both *P. angusticeps* and *P. robinsoni* may help illuminate the affinities of these taxa relative to modern *P. hamadryas* populations and relative to the recently described *P. h. botswanae*.

Interestingly, recent molecular dates place the origin of the modern P. hamadryas radiation between ~1.8 and 2.2 Ma in South Africa [48-51], which closely brackets the estimated age of the Malapa specimen at ~2.026-2.36 Ma. Thus, the P. angusticeps specimen described here may not only represent the earliest appearance of *P. angusticeps*, but potentially the earliest appearance of the modern baboon population in the fossil record, in almost perfect agreement with molecular estimates. While it is true that the molecular clock must always be calibrated through the fossil record and, therefore, there is always a bit of circular reasoning involved, it is still noteworthy and perhaps reassuring whenever new fossil discoveries seem to confirm molecular estimates, thereby lending support to hypothesized divergence dates. Because the estimated age of the Malapa fauna seems well-constrained around the dated ~2.026 Ma flowstone (see Geological Context above), the presence of *P. angusticeps* also assists in the biochronological assessment of other sites where *P. angusticeps* is documented, including Gladysvale, Haasgat, Kromdraai A, and Cooper's A [30, 33, 42, 44, 52-53]. In particular, the Malapa specimen supports recent revised estimates for the age of the Haasgat fauna (and P. angusticeps) between ~2.3-2.0 Ma, and suggests that Haasgat and Malapa together document a more secure first appearance date (FAD) for P. angusticeps between ~2.3-2.0 Ma. Cercopithecoid taxa such as P. angusticeps have long been used as important biochronological markers in age assessments of the South African Plio-Pleistocene hominin sites (e.g., [30-31, 44, 32, 54]), thus a more certain FAD for P. angusticeps may consequently lead to a slight adjustment in the estimated chronology of other hominin sites pending future faunal correlation analyses.

Supporting Information

S1 Dataset. Raw measurements used in this study (Table A). (XLS)

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Author Contributions

Conceived and designed the experiments: CCG CMS JMK LRB. Performed the experiments: CCG CMS. Analyzed the data: CCG CMS. Contributed reagents/materials/analysis tools: CCG CMS JMK LRB. Wrote the paper: CCG CMS JMK LRB.



References

- Berger LR, de Ruiter DJ, Churchill SE, Schmid P, Carlson KJ, Dirks PHGM, et al. (2010) Australoptihecus sediba: a new species of Homo-like Australopith from South Africa. Science 328: 195–204. doi: 10.1126/science.1184944 PMID: 20378811
- Dirks PHGM, Kibii JM, Kuhn BF, Steininger C, Churchill SE, Kramers JD, et al. (2010) Geological setting and age of Australopithecus sediba from Southern Africa. Science 328: 205–208. doi: 10.1126/science.1184950 PMID: 20378812
- Pickering R, Dirks PHGM, Jinnah Z, de Ruiter DJ, Churchill SE, Herries AIR, et al. (2011) Australopithecus sediba at 1.977 Ma and implications for the origins of the genus Homo. Science 333: 1421–1423. doi: 10.1126/science.1203697 PMID: 21903808
- Carlson KJ, Stout D, Jashashvili T, de Ruiter DJ, Tafforeau P, Carlson K, et al. (2011) The endocast of MH1, Australopithecus sediba. Science 333: 1402–1407. doi: 10.1126/science.1203922 PMID: 21903804
- Kibii JM, Churchill SE, Schmid P, Carlson KJ, Reed ND, de Ruiter DJ, et al. (2011) A partial pelvis of Australopithecus sediba. Science 333: 1407–1411. doi: 10.1126/science.1202521 PMID: 21903805
- Kivell TL, Kibii JM, Churchill SE, Schmid P, Berger LR (2011) Australopithecus sediba hand demonstrates mosaic evolution of locomotor and manipulative abilities. Science 333: 1411–1417. doi: 10.1126/science.1202625 PMID: 21903806
- Zipfel B, DeSilva JM, Kidd RS, Carlson KJ, Churchill SE, Berger LR (2011) The foot and ankle of Australopithecus sediba. Science 333: 1417–1420. doi: 10.1126/science.1202703 PMID: 21903807
- Churchill SE, Holliday TW, Carlson KJ, Jashashvili T, Macias ME, Mathews S, et al. (2013) The upper limb of Australopithecus sediba. Science 340: doi: 10.1126/science.1233477
- de Ruiter DJ, DeWitt TJ, Carlson KB, Brophy JK, Schroeder L, Ackermann RR, et al. (2013) Mandibular remains support taxonomic validity of *Australopithecus sediba*. Science 340: doi: 10.1126/science. 1232997
- DeSilva JM, Holt KG, Churchill SE, Carlson KJ, Walker CS, Zipfel B, et al. (2013) The lower limb and mechanics of walking in *Australopithecus sediba*. Science 340: doi: 10.1126/science.1232999
- Irish JD, Guatelli-Steinberg D, Legge SS, de Ruiter DJ, Berger LR (2013) Dental morphology and the phylogenetic "place" of Australopithecus sediba. Science 340: doi: 10.1126/science.1233062
- Schmid P, Churchill SE, Nalla S, Weissen E, Carlson KJ, de Ruiter DJ, et al. (2013) Mosaic morphology in the thorax of Australopithecus sediba. Science 340: doi: 10.1126/science.1234598
- Williams SA, Ostrofsky KR, Frater N, Churchill SE, Schmid P, Berger LR (2013) The vertebral column of Australopithecus sediba. Science 340: doi: 10.1126/science.1232996
- 14. Kuhn BF, Werdelin L, Hartstone-Rose A, Lacruz RS, Berger LR (2011) Carnivoran remains from the Malapa hominin site, South Africa. PLoS One: doi: 10.1371/journal.pone.0026940
- 15. Hartstone-Rose A, Kuhn BF, Nalla S, Werdelin L, Berger LR (2013) A new species of fox from the Australopithecus sediba type locality, Malapa, South Africa. T Roy Soc S Afr 68: 1–9.
- Brown FH, McDougall I, Davies I, Maier R (1985) In Delson E, Editor. Ancestors: The Hard Evidence. New York: Liss. pp. 82–90.
- Berger LR, Lacruz R, De Ruiter DJ (2002) Brief Communication: Revised age estimates of Australopithecus-bearing deposits at Sterkfontein, South Africa. Am J Phys Anthropol 119: 192–197. PMID: 12237940
- 18. Brugal J P, Roche H, Kibunjia M (2003) Faunes et paléoenvironnements des principaux sites archéologiques plio-pléistocènes de la formation de Nachukui (Ouest-Turkana, Kenya). C R Palevol 2: 675–684.
- **19.** Behrensmeyer AK (1978) Taphonomic and ecologic information from bone weathering. Paleobiology 4: 150–162.
- Leakey MG, Leakey REF (1976) Further Cercopithecinae (Mammalia, Primates) from the Plio/Pleistocene of East Africa. Fossil Vertebrates of Africa 4: 121–146.
- Leakey MG, Delson E (1987) Fossil Cercopithecidae from the Laetoli Beds. In: Leakey MD, Harris JM, editors. The Pliocene Site of Laetoli, northern Tanzania. Oxford: Oxford University Press. pp. 91–107.
- Jablonski NG (2002) Fossil Old World monkeys: the late Neogene radiation. In: Hartwig WC, editor. The Primate Fossil Record. Cambridge: Cambridge University Press. pp. 255–299.
- 23. Jablonski NG, Leakey MG (2008) The importance of the Cercopithecoidea from the Koobi Fora Formation in the context of primate and mammalian evolution. In: Jablonski NG, Leakey MG, editors. Koobi Fora Research Project. The fossil monkeys, Vol 6. San Francisco: California Academy of Sciences. pp. 397–416.



- Jablonski NG, Frost SR (2010) Cercopithecoidea. In: Werdelin L, Sanders WJ, editors. Cenozoic mammals of Africa. Berkeley: University of California Press. pp. 393

 –428.
- Harrison T (2011) Cercopithecoids: (Cercopithecidae, Primates). In: Harrison T, editor. Paleontology and geology of Laetoli: human evolution in context. Fossil hominins and the associated fauna, vertebrate paleobiology and paleoanthropology, Vol 2. New York: Springer. pp. 83–139.
- Remane A (1925) Der fossile Pavian (Papio sp.) von Oldoway nebst Bemerkungen u" ber die Gattung Simopithecus C.W. Andrews. Wissenschaftliche Ergebnisse der Oldoway Expedition 2: 83–90.
- Alemseged Z, Geraads D (2000) A new Middle Pleistocene fauna from the Busidima-Telalak region of the Afar, Ethiopia. C R Acad Sci IIA 331:549–556.
- Frost SR (2007) Fossil Cercopithecidae from the Middle Pleistocene Dawaitoli Formation, Middle Awash Valley, Afar Region, Ethiopia. Am J Phys Anthropol 134: 460–471. PMID: 17668443
- Frost SR, Alemseged Z, (2007) Middle Pleistocene fossil Cercopithecidae from Asbole, Afar Region, Ethiopia. J Hum Evol 53: 227–259. PMID: 17658583
- **30.** Delson E (1988) Chronology of South African australopith site units. In: Grine FE, editor. Evolutionary History of the "Robust" Australopithecines. New York: Aldine de Gruyter. pp. 317–324.
- McKee JK (1993) Taxonomic and evolutionary affinities of Papio izodi fossils from Taung and Sterkfontein. Palaeontol Afr 30: 43–49.
- **32.** Heaton JL (2006) Taxonomy of the Sterkfontein fossil Cercopithecinae: the Papionini of Members 2 and 4 (Gauteng, South Africa). Ph. D. Dissertation, Indiana University, Bloomington.
- Gilbert CC (2013) Cladistic analysis of extant and fossil African papionins using craniodental data. J Hum Evol 64: 399–433. doi: 10.1016/j.jhevol.2013.01.013 PMID: 23490264
- 34. Herries AIR, Pickering R, Adams JW, Curnoe D, Warr G, Latham AG, Shaw J (2013) A multi-disciplinary perspective on the age of Australopithecus in Southern Africa. In: Reed KE, Fleagle JG, Leakey RE, editors. The Paleobiology of Australopithecus. Dordrecht: Springer. pp. 21–40.
- 35. Granger DE, Gibbon RJ, Kuman K, Clarke RJ, Bruxelles L, Caffee MW (2015) New cosmogenic burial ages for Sterkfontein Member 2 Australopithecus and Member 5 Olduwan. Nature: doi: 10.1038/ nature14268
- **36.** Adams JW, Kegley ADT, Krigbaum J (2013) New faunal stable carbon isotope data from the Haasgat HGD assemblage, South Africa, including the first reported values for *Papio angusticeps* and *Cercopithecoides haasgati*. J Hum Evol 64: 693–698. doi: 10.1016/j.jhevol.2013.02.009 PMID: 23541178
- 37. Herries AIR, Kappen P, Kegley ADT, Patterson D, Howard DL, de Jonge MD, Potze S, Adams JW (2014) Palaeomagnetic and synchrotron analysis of >1.95 Ma fossil-bearing palaeokarst at Haasgat, South Africa. S Afr J Sci 110: 1–12. Available: http://dx.doi.org/10.1590/sajs.2014/20130102).
- 38. Szalay FS, Delson E (1979) Evolutionary History of the Primates. New York: Academic Press. 580 p.
- Delson E, Terranova CJ, Jungers WL, Sargis EJ, Jablonski NG, Dechow PC (2000) Body mass in Cercopithecidae (Primates, Mammalia): estimation and scaling in extinct and extant taxa. Anthropol Pap Am Mus 83: 1–159.
- Gilbert CC, Frost SR, Delson E (2013) Origins and evolution of the modern baboon, Papio hamadryas: evidence from the South African Plio-Pleistocene fossil record. Am J Phys Anthropol 56S: 129.
- **41.** Williams BA, Ross CF, Frost SR, Waddle DM, Gabadirwe M, Brook GA (2012) Fossil Papio cranium from! Ncumtsa (Koanaka) Hills, western Ngamiland, Botswana. Am J Phys Anthropol 149: 1–17.
- 42. Freedman L (1957) The fossil Cercopithecoidea of South Africa. Ann Transvaal Mus 23: 121-262.
- Eisenhart WL (1974) The fossil cercopithecoids of Makapansgat and Sterkfontein. Unpublished A.B. Thesis, Harvard College, Cambridge, Massachusetts.
- **44.** Delson E (1984) Cercopithecid biochronology of the African Plio-Pleistocene: Correlation among eastern and southern hominid-bearing localities. Cour Forsch Inst Senckenberg 69: 199–218.
- Keyser AW, Menter CG, Moggi-Cecchi J, Pickering TR, Berger LR (2000) Drimolen: a new hominidbearing site in Gauteng, South Africa. S Afr J Sci 96: 193–197.
- **46.** Herries AIR, Curnoe D, Adams JW (2009) A multi-disciplinary seriation of early *Homo* and *Paranthropus* bearing palaeocaves in southern Africa. Quatern Int 202: 14–28.
- 47. Frost SR, Marcus LF, Bookstein FL, Reddy DP, Delson E (2003) Cranial allometry, phylogeography, and systematics of large-bodied papionins (Primates: Cercopithecinae) inferred from geometric morphometric analysis of landmark data. Anat Rec Part A 275A: 1048–1072.
- Newman TK, Jolly CJ, Rogers J (2004) Mitochondrial phylogeny and systematics of baboons (*Papio*).
 Am J Phys Anthropol 124: 17–27. PMID: <u>15085544</u>
- Wildman DE, Bergman TJ, al-Aghbari A, Sterner KN, Newman TK, Phillips-Conroy JE, et al. (2004)
 Mitochondrial evidence for the origin of hamadryas baboons. Mol Phylo Evol 32: 287–296.



- Zinner D, Groenveld LF, Keller C, Roos C (2009) Mitochondrial phylogeography of baboons (*Papio* spp.)- indication for introgressive hybridization? BMC Evol Biol 9: 83–97. doi: <u>10.1186/1471-2148-9-83</u>
 PMID: <u>19389236</u>
- Zinner D, Wertheimer J, Liedigk R, Groeneveld LF, Roos C (2013) Baboon phylogeny as inferred from complete mitochondrial genomes. Am J Phys Anthropol 150: 133–140. doi: 10.1002/ajpa.22185 PMID: 23180628
- Berger LR, Keyser AW, Tobias PV (1993) Brief Communication: Gladysvale: first early hominid site discovered in South Africa since 1948. Am J Phys Anthropol 92: 107–111. PMID: 8238287
- McKee JK, Keyser AW (1995) Craniodental remains of Papio angusticeps from the Haasgat cave site, South Africa. Int J Primatol 16: 823–841.
- 54. McKee JK, Thackeray JF, Berger LR (1995) Faunal assemblage seriation of Southern African Pliocene and Pleistocene fossil deposits. Am J Phys Anthropol 96: 235–250. PMID: 7785723