

REVIEW

Intricate Relationship Between Adaptive and Innate Immune System in Allergic Contact Dermatitis

Muhammad Azeem^{a,1}, Hidaya Kader^{b,1}, Andreas Kerstan^c, Helal F. Hetta^{d,e}, Edgar Serfling^a, Matthias Goebeler^c, and Khalid Muhammad^{b,*}

^aDepartment of Molecular Pathology, Institute of Pathology, University of Würzburg, Würzburg, Germany; ^bDepartment of Biology, College of Science, United Arab Emirates University, Al Ain, United Arab Emirates; ^cDepartment of Dermatology, Venereology and Allergology, University Hospital Würzburg, Würzburg, Germany; ^dDepartment of Medical Microbiology and Immunology, Faculty of Medicine, Assiut University, Assiut, Egypt; ^eDepartment of Internal Medicine, University of Cincinnati College of Medicine, Cincinnati, OH, USA

Allergic contact dermatitis (ACD) is a complex immunological allergic disease characterized by the interplay between the innate and adaptive immune system. Initially, the role of the innate immune system was believed to be confined to the initial sensitization phase, while adaptive immune reactions were linked with the advanced elicitation phase. However, recent data predicted a comparatively mixed and interdependent role of both immune systems throughout the disease progression. Therefore, the actual mechanisms of disease progression are more complex and interlinked. The aim of this review is to combine such findings that enhanced our understanding of the pathomechanisms of ACD. Here, we focused on the main cell types from both immune domains, which are involved in ACD, such as CD4⁺ and CD8⁺ T cells, B cells, neutrophils, and innate lymphoid cells (ILCs). Such insights can be useful for devising future therapeutic interventions for ACD.

INTRODUCTION

Allergic contact dermatitis (ACD) is an inflammatory skin disease, affecting around 15% of the population worldwide [1]. In Europe, 40% of occupational health problems are related to the skin and 90% of them account

for contact dermatitis [2]. Recent data showed that almost 27% of European adults are sensitized to at least one contact allergen and they are on the verge of developing ACD [3]. Clinically, pruritus, vesicles, erythema, and dry scaly patches on the skin characterize the disease. The most commonly affected parts of the body are hands, arms, and

*To whom all correspondence should be addressed: Dr. Khalid Muhammad, Department of Biology, College of Science, United Arab Emirates University, Al Ain, United Arab Emirates; Tel: +971 3 713 6517, Fax: +971 3 713 4927; Email: k.muhammad@uaeu.ac.ae; ORCID iD: 0000-0001-6488-1722.

Abbreviations: ACD, allergic contact dermatitis; ILCs, innate lymphoid cells; DCs, dendritic cells; APC, antigen-presenting cells; DTH, delayed type hypersensitivity; dLN, skin-draining lymph nodes; Bregs, regulatory B cells; Treg, regulatory T cells.

Keywords: Allergic contact dermatitis, Lymphocytes, Innate immune cells, Adaptive immune cells.

Author Contributions: KM conceptualized the manuscript. AM, HK, and KM wrote the manuscript with the help of AK, HFH, ES, and MG. All authors were involved in critically reading and revising the manuscript. ¹These authors contributed equally to this work.

face [4-6]. The symptoms of the disease affect the quality of life and put a strong socio-economic impact of approximately 1 billion dollar annual loss in the US alone [7]. Therefore, there is a strong need of (pre)clinical research to better understand the immunological mechanisms of ACD and to find out effective treatment strategies. In this article, we focus on the recent advancements made by the interconnecting role of innate and adaptive immune cells in ACD. We encompass our current understandings about the pathomechanisms of ACD that would be helpful for planning future therapeutics.

PATHOPHYSIOLOGY OF THE DISEASE

ACD is a common skin disease evoked by delayed type hypersensitivity (DTH) responses to low molecular weight chemicals and metal allergens. The disease pathology is comprised of two distinct phases; the initial sensitization and subsequently, the elicitation phase. The sensitization phase starts when the susceptible individuals encounter skin contact with the allergen for the first time. Allergens are small molecular weight molecules such as urushiol, poison ivy, fragrance mixes, wool alcohol, rubber mixes, methylisothiazolinone, or metal ions such as nickel, chromium, cobalt, and gold [8-11]. These low molecular weight allergens are incomplete antigens (so-called haptens). After passing through the skin barrier they bind to cellular proteins to become immunogenic by triggering T cell responses [12-14]. Two distinct subsets of dendritic cells (DCs) are involved in allergen uptake and subsequent presentation to T cells. Tissue-resident langerin⁺ epidermal DCs, known as Langerhans cells (LCs) reside in the epidermis while langerin⁻ dermal dendritic cells (DDCs) patrol in the dermis [15,16]. After taking up hapten-protein complexes they migrate to skin-draining lymph nodes (dLN) where they act as antigen-presenting cells (APCs) (Figure 1). In dLN, the APCs prime naïve CD4⁺ and CD8⁺ T cells by presenting the antigen in the context of MHC (I or II) cell surface molecules. This exposure triggers the differentiation and proliferation of CD8⁺ and CD4⁺ T cells into IFN γ producing cytotoxic T cells (Tc) and helper T (Th) cells, respectively [17,18].

Those T cells then reside in the dLN as effector memory cells until the next exposure of the same allergen. The elicitation phase of ACD is the clinically obvious phase in which inflammatory signs become visible in the patient. In case of allergen re-exposure in a sensitized individual, effector memory cells proliferate and subsequently migrate from dLN to the site of allergen contact. This activation and migration of T cells is mediated by the allergen-induced production of chemokines and cytokines from native DDCs, LCs, and keratinocytes [19-22]. The recruitment of activated T cells results in epidermal tissue destruction, such as vesicle and blister

formation, erythema, itch, and other inflammatory signs [23]. Recently, such acute responses have been attributed to skin tissue-resident memory T (Trm) cells. Trm cells induce acute allergic responses as fast as 24 hours after allergen exposure [24]. The effector T cells of ACD mainly produce inflammatory cytokines including interferon (IFN)- γ , interleukin (IL)-1 α , IL-6, IL-17, IL-26, Tumor necrosis factor (TNF)- α , and IL-23 at the site of inflammation and promote further recruitment of cytotoxic T cells and innate immune cells to enhance the allergic responses [25-28]. At the same time, the regulatory arm of adaptive immune system, namely Foxp3⁺ IL-10⁺ regulatory T cells (Tregs) and IL10⁺ regulatory B cells (Bregs) downregulate the inflammatory responses of cytotoxic T cells and innate immune cells [29,30]. In this review we have discussed the role of all these important cells that mediate the disease initiation, progression, maintenance, and suppression.

ROLE OF ADAPTIVE IMMUNITY IN ACD

Adaptive immune responses in ACD start with the sensitization phase when hapten bearing APCs migrate from skin to local lymph nodes and present the antigens to naïve T cells [31]. This antigen presentation through MHCI or MHCII molecules activates naïve CD8⁺ or CD4⁺ T cells, respectively [32]. It leads to priming and clonal expansion of antigen-specific T cells and later on to their migration to the skin upon the second antigen exposure resulting in DTH reaction. DTH is a T cell-mediated inflammatory response evolving 24 to 48 hours after allergen exposure [33]. The “classical cells” of DTH are CD4⁺ T cells, but in case of ACD the major T cell effector functions are attributed to CD8⁺ T cells. This was demonstrated by monoclonal antibody (mAb)-dependent depletion of CD4⁺ or CD8⁺ T cells *in vivo* in the contact hypersensitivity (CHS) mouse model of ACD. The depletion of CD8⁺ cells showed a marked reduction in the CHS reactions while depletion of CD4⁺ cells resulted in a strong enhancement of CHS [34]. In another CHS study, mice with an inactivated MHC I gene within the CD8 compartment (leading to a drastic decrease in CD8⁺ T cells) showed a diminished allergic response to 1-Fluoro-2,4-dinitrobenzene (DNFB), one of the classical obligatory contact-sensitizing haptens used to evoke CHS. On the other hand, MHC II knock-out mice lacking CD4⁺ T cells showed a fulminant allergic response [35].

Crosstalk between CD4⁺ and CD8⁺ T cells in ACD

There are numerous studies which indicate an important role of different CD8⁺ cytotoxic T cells (IFN γ secreting Tc1 while IL4 and IL5 secreting Tc2 cells) in ACD. In addition, keratinocytes also play a significant

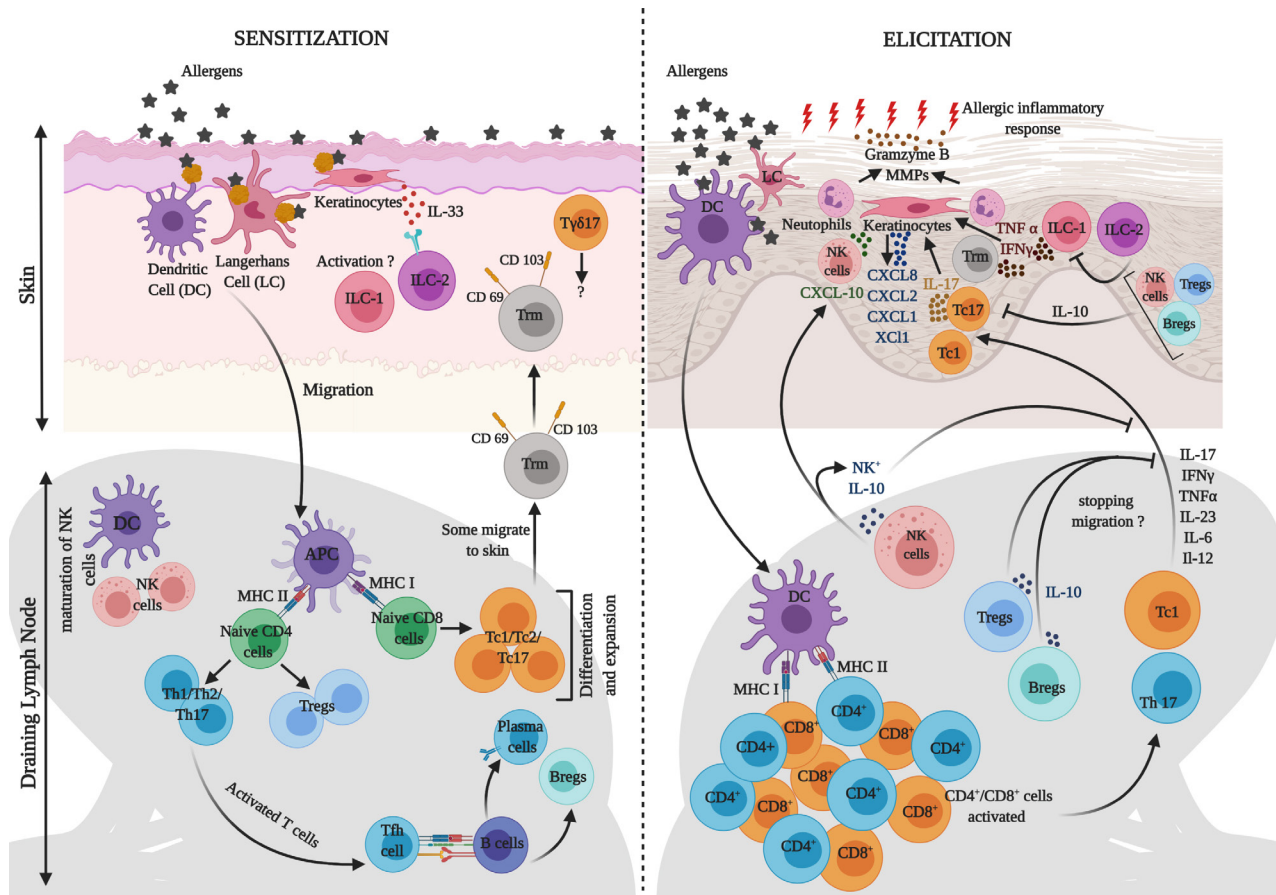


Figure 1. Pathophysiology of allergic contact dermatitis (ACD) showing the immunological response during the sensitization and elicitation phases.

role by secreting chemokines and cytokines in response to allergen application. In the early stages of the elicitation phase, these chemokines attract Tc1 and Tc2 cells towards the skin where they induce tissue damage and acute inflammatory signs [36]. In the later stage, IFN γ and TNF α prime keratinocytes for expression of ICAM-I and MHC II molecules, leading to the recruitment of CD4⁺ Th1 cells. The recruitment of CD8⁺ Tc1 and Tc2 cells is largely independent of IFN γ [37]. Therefore, one may speculate that CD8⁺ T cells induce early cytotoxicity and edematous inflammation, while CD4⁺ T cells act later by inducing more modulatory effects.

The inflammatory effects of CD8⁺ and CD4⁺ are dependent on a library of different cytokines. It includes IL-1 α , IL-6, IL-12, IL-17, IL-22, IL-23, TNF α , TGF β , and IFN γ , secreted by lymphocytes, APCs, and epithelial cells (e.g. keratinocytes) [38,39]. IL-1 α is part of the IL-1 family of cytokines which includes 11 pro-inflammatory and anti-inflammatory cytokines [40]. IL-1 α and IL-1 β are pro-inflammatory cytokines and both bind to the IL-1 receptor (IL-1R). Their activity is modulated by an intrinsic IL-1R antagonist (IL-1Ra). Other members of the family include three isoforms of IL-36 which bind

to its respective receptor (IL-36R) and are responsible for inflammatory responses. Alike IL-1, IL36 mode of action can be blocked by IL-36Ra. IL-18 and IL-33 are also part of the IL-1 family and produce inflammatory responses by binding to IL-18R α and interleukin-1 receptor-like 1(ST2) receptor respectively [41]. The disturbed balance between pro- and anti-inflammatory IL-1 family cytokines is linked to the development of ACD. Mattii *et al.* have reported an increase in IL-1 β , IL-36, and IL-33 in skin biopsies from ACD patients [42].

In the context of ACD, IL-17 is a signature cytokine and therefore is a major player for inducing inflammatory symptoms [43,44]. Both CD4⁺ and CD8⁺ T cells known as Th17 and Tc17 respectively, can produce IL-17 [45] and a crosstalk between these cells play an important role in ACD. Murine Th17 polarization starts after naïve CD4⁺ T cells have experienced antigen signals in the presence of IL-23, IL-1 β , and IL-12. In addition, IL-6 inhibits the Treg pathway and allows TGF β to act as a driver for Th17 polarization. After differentiation, Th17 cells secrete IL-21, IL-22, IL-17A, and TGF β [46]. Similar polarizing conditions are required for the development of Tc17 cells with the dual capacity to secrete IL-17 and

IFN γ [47]. The role of IL-17 in ACD was first reported by detecting IL-17 mRNA in skin samples of nickel allergic patients [48]. Later on, *in vitro* studies showed that CD4⁺ T cells are capable of producing IL-17 and IFN γ after stimulation with PMA and ionomycin [49]. In humans the role of Th17/Tc17 is linked to the elicitation phase of ACD. Zhao *et al.* showed a significant increase in Th17/Tc17 infiltration in human ACD skin biopsies along with marked expression of ROR γ t and other Th17/Tc17 specific cytokines (IL-17A, IL-17F, IL-21, and IL-22) [50]. Furthermore, *in vivo* evidence for a role of Th17 cells in CHS was presented by using IL-17^{-/-} mice. These mice displayed a decreased ear inflammation after allergen exposure and ear swelling, surrogate marker of skin inflammation, was diminished by adoptive transfer of CD4⁺ T cells from wild type mice [51]. On the other hand, the role of Tc17 cells in CHS/ACD has been underestimated as compared to Th17 cells. The effector function of Tc17 cells in CHS were first reported by *in vivo* depletion of CD4⁺ and CD8⁺ T cells before DNFB sensitization. Mice depleted of CD8⁺ T cells showed reduced ear swelling, while CD4⁺ depletion enhanced the inflammation [52]. For further confirmation, DNFB-primed CD8⁺ T cells were adoptively transferred in Rag-1^{-/-} mice. Upon elicitation, these mice exhibited a prominent CHS reaction that was inhibited by IL-17 neutralization. These findings suggest a context-dependent role of both cell types in CHS, depending on the mouse model and the nature of the allergen used. It is possible that CD8⁺ T cells become dominant during CHS responses by inducing Fas-mediated apoptosis of CD4⁺ T cells after allergen application. However, in the absence of CD8⁺ T cells, CD4⁺ T cells are fully capable of inducing hypersensitive response [53].

Besides IL-17, Th17/Tc-generated IL-26 is another cytokine involved in ACD. IL-26 is an important member of the IL-10 cytokine family and is predominantly produced by Th17, Th1, and NK cells [54,55]. Previously, it was known for its anti-microbial activity by recruitment of various innate immune cells and Th17 cells. More recently, however, IL-26 generated by Th17 cells was reported to be important for the development of ACD in humans [56] implying that IL-26 is not only important for antimicrobial activity but also inflammatory responses.

ROLE OF IL-17 IN ACD

There are six different types of IL-17 cytokines (IL17 A-F), produced as homo and heterodimers from CD4⁺ T, CD8⁺ T, $\gamma\delta$ T, natural killer (NK) cells, neutrophils, mast cells, and epithelial cells [44]. The most potent proinflammatory type is IL-17A, which exerts its functions by inducing various pro-inflammatory molecules and recruiting different types of cells [57]. It induces the production of granulocyte-colony stimulating factor

(G-CSF) and granulocyte-macrophage CSF (GM-CSF), thereby attracting neutrophils and monocytes to the site of inflammation [58,59]. In addition to IL-6 and TNF α cytokines, IL17 also stimulates the production of various chemokines such as CXCL2, CXCL5, CXCL10, CCL20, and CCL2, which are responsible for the recruitment of monocytes and macrophages [60,61]. Notably, CCL20 binds to its receptor CCR6, which is preferentially expressed on Th17/Tc17 cells thereby supporting a positive skin inflammation feedback loop [62]. Moreover, IL-17 induces the production of matrix metalloproteinases (MMP1-3) in myeloid skin infiltrating cells resulting in tissue damage by extracellular matrix proteins [63]. In conclusion, IL-17 exerts its inflammatory role by bridging adaptive immune responses to the innate immune system and inducing tissue damage. Contrary to the tissue-damaging effect of IL-17, a recent finding also suggested its homeostatic role. In this study, mice deficient for neonatal V γ 2TCR⁺ T $\gamma\delta$ 17 cells were used as an atopic dermatitis (AD) mouse model. The study showed that IL-17 controls AD development by maintaining balance in the population of $\alpha\beta$ T cells, skin commensal bacteria, and rate of basal keratinocyte differentiation [64]. This indicated that neonatal $\gamma\delta$ T cells behave like innate regulatory T cells in the skin. Such a homeostatic role of IL-17 needs further investigation in the context of ACD/CHS which would further enhance our understanding of the functionality of IL-17 during different inflammatory disorders.

Role of IL-10 in ACD

ACD is a self-limiting disease, once the contact allergen is removed, inflammatory symptoms begin to resolve. On the cellular level, the resolution is dependent on regulatory T (Tregs) and B (Bregs) cells [65,66]. Tregs cause suppression of activated T cells by expressing the anti-inflammatory cytokines IL-10 and TGF β . They also induce cellular anergy through surface-bound molecules such as PD-1 and CTLA-4 [67-69]. In case of chronic ACD, activated CD8⁺ T cells turn into tissue-resident memory T cells (CD8⁺ Trm) which are capable of eliciting a fulminant immune response upon re-exposure to allergen [24,70]. It has been reported that the activity of these Trm cells is controlled by inhibitory checkpoint receptors such as CTLA-4, PD-1, and TIM-3 [71]. Removal of these checkpoint inhibitors facilitates the induction of recurrent allergic response even with low amount of allergen. Apart from anergic T cell markers, IL-10 acts as a major suppressive agent of Tregs in ACD. Other than B and T cells, IL-10 may be produced by numerous cell types including DCs, neutrophils, macrophages and NK cells [72]. IL-10 exerts its suppressive function by activation of STAT3. Activated STAT3 modifies the pro-inflammatory transcriptome directly or indirectly by inducing transcriptional or post transcriptional modifications

respectively [73]. On other hand, it also starts the transcription of suppressor of cytokine signaling 3 (SOCS3) and IL-1Ra. SOCS3 downregulates the TNF α production by inhibiting the activation of mitogen activated protein kinase (MAPK) and NF- κ B-signaling pathways which lead to further reduce the expression of pro-inflammatory cytokines [74-76]. Therefore, IL-10 producing Tregs are in the focus of therapeutic approaches for ACD treatment. Despite the production of IL-10 from CD8⁺ T cells, CD8 Tregs appeared to be dispensable for the resolution of CHS. Mice deficient for CD8⁺IL-10⁺ T cells showed CHS responses comparable to wild-type controls. Even in the absence of CD4⁺ T cells, IL-10 production from CD8⁺ T cells could not resolve inflammation [77]. Therefore, it is implicated that in terms of ACD/CHS, IL10 producing CD8⁺ T cells may play an effector role while regulatory CD4⁺ T cells can exert a regulatory role in controlling allergic responses.

B CELLS IN ACD

B cells play crucial roles in protection against infectious diseases by contributing to the immune responses through antigen presentation, cytokine, and antibody production. In addition to antibody-secreting B cells, Bregs have been reported [78-80]. Through the action of IL10 Bregs exert immunosuppressive capacities to modulate or control various inflammatory diseases [79,81,82]. Several studies have revealed the significance of B cells in CHS/ACD [83,84]. B cell deficiency worsened the CHS responses, hence providing a protective role of B cells in allergic inflammatory diseases [66]. The ablation of the B cell-specific peroxisome proliferator-activated receptor γ (PPAR- γ), a member of the nuclear hormone receptor superfamily, impaired the regulatory function of B cells in a CHS mouse model. This resulted in enhanced CHS responses after 48 hours of challenge. It was observed that the expression of IL-10 was significantly decreased by CD19⁺ B cells and CD5⁺ CD1d^{hi} cells in PPAR- γ -deficient mice, contributing to the reduced regulatory function of B cells. However, the development of B cells was unaffected by the loss of PPAR- γ [85]. It is suggested that the immunosuppressive effects of UVB irradiation in a CHS mouse model were attributed to the inhibition of T cell proliferation by Bregs. The Bregs were induced by UVB via Toll-Like receptor (TLR) 4 whereby the deficiency of TLR-4 not only impaired the inhibitory function of B cells but also reduced the therapeutic effects of UVB on CHS responses [86]. Further data on Bregs have shown that the presence of two distinct Bregs subsets, namely splenic CD1d^{hi} CD5⁺ B cells and peritoneal B1 cells, inhibit the CHS response. Of interest, splenic CD1d^{hi} CD5⁺ B cells suppress the acute phase of CHS whereas the less significant peritoneal B1a cells are believed to help in

suppressing the late remission phase. However, the role of B1 cells in CHS remains yet controversial [66].

ROLE OF THE INNATE IMMUNE SYSTEM IN ACD

Traditionally, the role of the innate immune system is believed to be restricted to the sensitization phase of ACD, which is characterized by antigen presentation by DCs and early responses by macrophages, natural killer cells, and neutrophils. However, in recent studies the role of the innate immune system has found to be expanded till later stage of elicitation. Studies on metal allergy indicate that innate immune system play pivotal role in CHS responses [11,14,87]. Schmidt *et al.* have shown that Nickel ions interact with three histidine residues (H431/H456/H458) within the interaction domain of human TLR-4 and trigger the formation of tetrameric complexes consisting of two TLR4 and two myeloid differentiation protein 2 (MD-2) co-receptor molecules. These complexes initiate a downstream signal cascade that lead to the activation of pro-inflammatory nuclear factor κ B (NF- κ B) transcription factors [88]. Here we want to focus on different innate immune cells as well as newly identified innate lymphoid cells (ILCs).

Neutrophils in ACD

Neutrophils are among the first recruited cells to the site of inflammation and their recruitment is dependent on C-X-C chemokines such as CXCL8, CXCL2, CXCL1, and CXCL5 [89]. Pathological function of neutrophils relies on the metalloproteinase and granzyme B dependent degradation of extra cellular matrix and attraction of macrophages to the site of inflammation [90]. More than two decades ago, Laan *et al.* showed that IL-17 plays an important role in neutrophil recruitment via C-X-C chemokine release [91,92]. Perturbation of neutrophil infiltration to the site of allergen exposure leads to reduced allergic responses. Leukotriene B4 is a potent chemo attractant for neutrophils in allergic diseases [93]. Epicutaneous application of 2,4,6 Trinitrochlorobenzene (TNCB) onto Leukotriene B4 (Ltb4)r1^{-/-} deficient mice showed a decreased neutrophil infiltration to ear skin and reduced inflammation as compared to wild type mice [94]. Moreover, depletion of neutrophils showed that they are important for both stages of ACD. Their role in the sensitization and elicitation phases was determined by using Mcl1^{ΔMyelo} neutrophil-deficient mice and by anti-Ly6G dependent neutrophil depletion. Adoptive transfer of lymph node cells from wild type mice sensitized by TNCB showed that CHS responses cannot be recapitulated in Mcl1^{ΔMyelo} mice [95]. This indicates that neutrophils are actively involved in both phases of ACD. Moreover, it is proposed that neutrophils facilitate the

depletion of T cells and ILCs, suggesting a much more complex association between neutrophils and adaptive immune responses [96,97].

Innate Lymphoid Cells in ACD

Innate lymphoid cells (ILCs) belong to the lymphoid lineage and share a common origin with T, B, and NKT cells. During development, upregulation of transcription factor T cell factor 1 (Tcf-1) in common lymphoid progenitor (CLP) cells gives rise to early innate lymphoid progenitor (EILP) cells. From this stage, two developmental branches appear, one leads to EOMES⁺ NK cells, and the other gives rise to three different types of ILCs (ILC 1-3) [98].

Natural killer cells: NK cells have recently been identified as an important cell type for ACD. They produce IFN γ and regarded as innate counterparts of CD8⁺ T cells. Human CD56^{high} CD16⁻ CD62L⁻ NK cells were characterized for the first time by Carbone *et al.* for their role in ACD. In mice, NK cells have two further subdivisions, CD49a⁺ DX5⁺ conventional NK (cNK) cells and CD49a⁺ DX5⁻ liver resident NK cells [99]. Mice NK cells also express CD62L and CCR7 for localizing to secondary lymphoid organs where they foster DC maturation and T cell priming [100]. EOMES⁺ cNK cells are among the first responders in CHS after allergen exposure and evoke an acute inflammatory response within 24 hours of allergen application [101]. Their recruitment to skin is also facilitated by the CXCL10 chemokine as they express CXCR3, CCR5, and CCR6 chemokine receptors [102]. Besides cNK cells, another unique type of NK cells, with restricted repertoire of TCR $\alpha\beta$ chains known as NKT cells, is involved in CHS responses [103]. NKT cells detect glycolipid antigens by CD1d (MHC analogue) bound on LCs, DCs, and keratinocytes [104,105], which is followed by production of IFN γ , TNF α , and the induction of apoptosis in affected cells [106]. Cytotoxicity of NKT cells can be controlled by inhibiting the CD1d mediated function of antigen presentation, which provides a potential window for therapeutic intervention of ACD [107].

NK cells alone can induce CHS responses independent of T and B cells. Ly49 C-I⁺ NK cells can mediate an antigen-specific long lasting CHS reaction. Likewise, DNFB application can induce fulminant inflammatory responses in Rag1^{-/-} mice and localization of NK cells to inflamed skin. Moreover, transfer of NK cells from DNFB-sensitized Rag2^{-/-} mice to naïve mice reproduced the CHS response, which indicates a capacity of NK cells for retaining hapten-specific memory [108]. Although NK cells constitute a minor population of skin infiltrating cells in CHS, they contribute massively to the development and regulation of CHS responses. Of interest, NK cells also produce IL-10, thereby suppressing CD8⁺ T cell

mediated ACD, independent of Tregs [109,110]. These findings suggest a broad and comprehensive role of NK cells in ACD and propose important intervention opportunities for therapeutic purposes.

Innate lymphoid cells (ILC1-3): Due to the expression of the transcription factor Tbet and of IFN- γ production, ILC1 is considered to be a close counterpart of Th1 cells. NK cells also express Tbet but can be distinguished from ILC1 cells, as the later do not express EOMES [111,112]. Besides NK cells, ILC1 is also reported as an important mediator of acute inflammatory responses in CHS. They produce Th1 dominant CHS responses parallel to CD8⁺ T cells. The application of contact allergens like TNCB and oxazolone, induce type I contact hypersensitivity responses which are dominated by CD8⁺ T, NK, and ILC1 cells [113,114]. Recently, Rafei-Shamsabadei *et al.* characterized the ear-infiltrating ILCs by studying CHS to TNCB in ILC reporter mice. Acute inflammatory responses appeared as early as 24 hours after allergen application and were characterized by elevated levels of NK and ILC1 cells [97]. ILC2 appeared in the later stage (48-72 h) with a characteristic type II cytokine production (IL-13 and IL-5). This indicated a function of ILC-2 in the resolution of the inflammatory response. The role of ILC2 was further investigated by using anti-CD90.2 mAb for depleting ILCs in Rag1^{-/-} mice or by using Rora^{sg/flox1} 17r^{Cre/+} mice which lack ILC2 cells. Application of allergen in these mice showed significantly enhanced and prolonged inflammatory responses [97]. In conclusion, this data supports the effector function of ILC1 cells in type I CHS response, while ILC2 cells appear to exert regulatory functions to keep the immune response in a homeostatic balance.

ILC3 cells, on the other hand, share similarities with Th17 and Tc17 cells because of ROR γ t expression and production of type III cytokine such as IL-17 and IL-22 [115,116]. Such kind of IL-17 and IL-22 producing ILC3 cells were found to be involved in psoriasis [117,118]. During the developmental process, ILC3 bifurcates into two subtypes, namely CCR6⁺ ROR γ t⁺ cells and CCR6⁻ ROR γ t⁺ Tbet⁺ cells. The CCR6⁺ fraction of ILC3 cells comprise lymphoid tissue inducer (LTi) cells, which are important for the development of lymphoid organs at the embryonic stage [119-121]. In mice, the CCR6⁻ fraction gradually loses ROR γ t expression and up regulates Tbet and converts into INF γ -producing cells resembling ILC1 cells [122,123]. Based on their production of type III cytokines and involvement in allergic skin diseases, the role of ILC3 cells in ACD needs to be investigated in more details.

CONCLUSION

Understanding the pathomechanism of ACD is

essential for the development of adequate therapies. Although T cells are major players in the clinical phase of the disease, their activation and maintenance of cytotoxic effects is dependent on innate immune cells. Identification of roles of NK cells and ILCs in the active phase of the disease may propose novel options for therapeutic interventions. Moreover, different cell types are involved in a context-dependent manner in ACD, which are believed to be linked with the type of allergen. It is established that the activity of Tregs can be manipulated for suppression of ACD, but identification of the homeostatic role of $\gamma\delta 17$ cells and NK cells gave new insights into immunosuppressive functions in skin inflammation. Finally, it is obvious that ILCs are involved in ACD development, but there are many missing points to establish their networking with $CD4^+$ and $CD8^+$ T cells. Such gaps need to be filled to more precisely understand the interplay of the innate and adaptive immune system in ACD.

Acknowledgments: The authors acknowledge the funding support of their research by grants of the *Interdisziplinäres Zentrum für Klinische Forschung (IZKF)* Würzburg (KM and AK) and UAE University-start up grant# G00003347 & UAEU-UPAR-Grant#G00003458 (KM). BioRender was used to create the figure.

REFERENCES

- Kostner L, Anzengruber F, Guillod C, Recher M, Schmid-Grendelmeier P, Navarini AA. Allergic contact dermatitis. *Immunol Allergy Clin North Am*. 2017 Feb;37(1):141–52.
- Alfonso JH, Bauer A, Bensefa-Colas L, Boman A, Bubas M, Constandt L, et al. Minimum standards on prevention, diagnosis and treatment of occupational and work-related skin diseases in Europe - position paper of the COST Action StanDerm (TD 1206). *J Eur Acad Dermatol Venereol*. 2017 Jun;31 Suppl 4:31–43.
- Diepgen TL, Ofenloch RF, Bruze M, Bertuccio P, Cazzaniga S, Coenraads PJ, et al. Prevalence of contact allergy in the general population in different European regions. *Br J Dermatol*. 2016 Feb;174(2):319–29.
- Nassau S, Fonacier L. Allergic Contact Dermatitis. *Med Clin North Am*. 2020 Jan;104(1):61–76.
- Owen JL, Vakharia PP, Silverberg JI. The role and diagnosis of allergic contact dermatitis in patients with atopic dermatitis. *Am J Clin Dermatol*. 2018 Jun;19(3):293–302.
- Kohl L, Blondeel A, Song M. Allergic contact dermatitis from cosmetics. Retrospective analysis of 819 patch-tested patients. *Dermatology*. 2002;204(4):334–7.
- Qin R, Lampel HP. Review of occupational contact dermatitis—top allergens, best avoidance measures. *Curr Treat Options Allergy*. 2015;2(4):349–64.
- Mowad CM, Anderson B, Scheinman P, Pootongkam S, Nedorost S, Brod B. Allergic contact dermatitis: patient management and education. *J Am Acad Dermatol*. 2016 Jun;74(6):1043–54.
- Oosterhaven JA, Uter W, Aberer W, Armario-Hita JC, Ballmer-Weber BK, Bauer A, et al.; ESSCA Working Group. European Surveillance System on Contact Allergies (ESSCA): contact allergies in relation to body sites in patients with allergic contact dermatitis. *Contact Dermat*. 2019 May;80(5):263–72.
- Raghavan B, Martin SF, Esser PR, Goebeler M, Schmidt M. Metal allergens nickel and cobalt facilitate TLR4 homodimerization independently of MD2. *EMBO Rep*. 2012 Dec;13(12):1109–15.
- Adam C, Wohlfarth J, Haubmann M, Sennefelder H, Rodin A, Maler M, et al. Allergy-Inducing Chromium Compounds Trigger Potent Innate Immune Stimulation Via ROS-Dependent Inflammasome Activation. *J Invest Dermatol*. 2017 Feb;137(2):367–76.
- Kaplan DH, Igyártó BZ, Gaspari AA. Early immune events in the induction of allergic contact dermatitis. *Nat Rev Immunol*. 2012 Jan;12(2):114–24.
- Davis MD, Wang MZ, Yiannias JA, Keeling JH, Connolly SM, Richardson DM, et al. Patch testing with a large series of metal allergens: findings from more than 1,000 patients in one decade at Mayo Clinic. *Dermatitis*. 2011 Sep-Oct;22(5):256–71.
- Schmidt M, Goebeler M. Immunology of metal allergies. *J Dtsch Dermatol Ges*. 2015 Jul;13(7):653–60.
- Toebak MJ, Gibbs S, Bruynzeel DP, Scheper RJ, Rustemeyer T. Dendritic cells: biology of the skin. *Contact Dermat*. 2009 Jan;60(1):2–20.
- Romani N, Clausen BE, Stoitzner P. Langerhans cells and more: langerin-expressing dendritic cell subsets in the skin. *Immunol Rev*. 2010 Mar;234(1):120–41.
- Honda T, Egawa G, Grabbe S, Kabashima K. Update of immune events in the murine contact hypersensitivity model: toward the understanding of allergic contact dermatitis. *J Invest Dermatol*. 2013 Feb;133(2):303–15.
- Vocanson M, Hennino A, Chavagnac C, Saint-Mezard P, Dubois B, Kaiserlian D, et al. Contribution of $CD4^+$ and $CD8^+$ T-cells in contact hypersensitivity and allergic contact dermatitis. *Expert Rev Clin Immunol*. 2005 May;1(1):75–86.
- Enk AH, Katz SI. Early molecular events in the induction phase of contact sensitivity. *Proc Natl Acad Sci USA*. 1992 Feb;89(4):1398–402.
- Homey B, Wang W, Soto H, Buchanan ME, Wiesenborn A, Catron D, et al. Cutting edge: the orphan chemokine receptor G protein-coupled receptor-2 (GPR-2, CCR10) binds the skin-associated chemokine CCL27 (CTACK/ALP/ILC). *J Immunol*. 2000 Apr;164(7):3465–70.
- Martin SF. Contact dermatitis: from pathomechanisms to immunotoxicology. *Exp Dermatol*. 2012 May;21(5):382–9.
- Martin SF, Esser PR, Weber FC, Jakob T, Freudenberg MA, Schmidt M, et al. Mechanisms of chemical-induced innate immunity in allergic contact dermatitis. *Allergy*. 2011 Sep;66(9):1152–63.
- Akiba H, Kehren J, Ducluzeau MT, Krasteva M, Horand F, Kaiserlian D, et al. Skin inflammation during contact hypersensitivity is mediated by early recruitment of $CD8^+$ T cytotoxic 1 cells inducing keratinocyte apoptosis. *J Immunol*. 2002 Mar;168(6):3079–87.
- Gaide O, Emerson RO, Jiang X, Gulati N, Nizza S, Desmaires C, et al. Common clonal origin of central and resident

- memory T cells following skin immunization. *Nat Med*. 2015 Jun;21(6):647–53.
25. Shabgah AG, Fattahi E, Shahneh FZ. Interleukin-17 in human inflammatory diseases. *Postepy Dermatol Alergol*. 2014 Aug;31(4):256–61. doi: 10.5114/pdia.2014.40954.
 26. Heuffler C, Topar G, Koch F, Trockenbacher B, Kämpgen E, Romani N, et al. Cytokine gene expression in murine epidermal cell suspensions: interleukin 1 beta and macrophage inflammatory protein 1 alpha are selectively expressed in Langerhans cells but are differentially regulated in culture. *J Exp Med*. 1992 Oct;176(4):1221–6.
 27. Silvestre MC, Reis VM. Evaluation of the profile of inflammatory cytokines, through immunohistochemistry, in the skin of patients with allergic contact dermatitis to nickel in the acute and chronic phases. *An Bras Dermatol*. 2018 Nov/Dec;93(6):829–35.
 28. Itoh T, Hatano R, Komiya E, Otsuka H, Narita Y, Aune TM, et al. Biological Effects of IL-26 on T Cell-Mediated Skin Inflammation, Including Psoriasis. *J Invest Dermatol*. 2019 Apr;139(4):878–89.
 29. Zhang H, Kong H, Zeng X, Guo L, Sun X, He S. Subsets of regulatory T cells and their roles in allergy. *J Transl Med*. 2014 May;12(1):125.
 30. Kim HS, Lee MB, Lee D, Min KY, Koo J, Kim HW, et al. The regulatory B cell-mediated peripheral tolerance maintained by mast cell IL-5 suppresses oxazolone-induced contact hypersensitivity. *Sci Adv*. 2019 Jul;5(7):eaav8152. <https://doi.org/10.1126/sciadv.aav8152>.
 31. Weston WL, Bruckner A. Allergic contact dermatitis. *Pediatr Clin North Am*. 2000 Aug;47(4):897–907.
 32. Esser PR, Martin SF. Pathomechanisms of contact sensitization. *Curr Allergy Asthma Rep*. 2017 Nov;17(12):83.
 33. Luo Y, Dorf ME. Delayed-type hypersensitivity. *Curr Protoc Immunol*. 2003;55(1):4.5.1–4.5.
 34. Gocinski BL, Tigelaar RE. Roles of CD4+ and CD8+ T cells in murine contact sensitivity revealed by in vivo monoclonal antibody depletion. *J Immunol*. 1990 Jun;144(11):4121–8.
 35. Bour H, Peyron E, Gaucherand M, Garrigue JL, Desvignes C, Kaiserlian D, et al. Major histocompatibility complex class I-restricted CD8+ T cells and class II-restricted CD4+ T cells, respectively, mediate and regulate contact sensitivity to dinitrofluorobenzene. *Eur J Immunol*. 1995 Nov;25(11):3006–10.
 36. Traidl C, Sebastiani S, Albanesi C, Merk HF, Puddu P, Girolomoni G, et al. Disparate cytotoxic activity of nickel-specific CD8+ and CD4+ T cell subsets against keratinocytes. *J Immunol*. 2000 Sep;165(6):3058–64.
 37. Cavani A, Albanesi C, Traidl C, Sebastiani S, Girolomoni G. Effector and regulatory T cells in allergic contact dermatitis. *Trends Immunol*. 2001 Mar;22(3):118–20.
 38. Raphael I, Nalawade S, Eagar TN, Forsthuber TG. T cell subsets and their signature cytokines in autoimmune and inflammatory diseases. *Cytokine*. 2015 Jul;74(1):5–17.
 39. Cox MA, Harrington LE, Zajac AJ. Cytokines and the inception of CD8 T cell responses. *Trends Immunol*. 2011 Apr;32(4):180–6.
 40. Dinarello CA. Interleukin-1 in the pathogenesis and treatment of inflammatory diseases. *Blood*. 2011 Apr;117(14):3720–32.
 41. Sims JE, Smith DE. The IL-1 family: regulators of immunity. *Nat Rev Immunol*. 2010 Feb;10(2):89–102.
 42. Mattii M, Ayala F, Balato N, Filotico R, Lembo S, Schiattarella M, et al. The balance between pro- and anti-inflammatory cytokines is crucial in human allergic contact dermatitis pathogenesis: the role of IL-1 family members. *Exp Dermatol*. 2013 Dec;22(12):813–9.
 43. Peiser M. Role of Th17 cells in skin inflammation of allergic contact dermatitis. *Clin Dev Immunol*. 2013;2013:261037. doi: 10.1155/2013/261037.
 44. Pappu R, Ramirez-Carrozzi V, Sambandam A. The interleukin-17 cytokine family: critical players in host defence and inflammatory diseases. *Immunology*. 2011 Sep;134(1):8–16.
 45. Mills KH. Induction, function and regulation of IL-17-producing T cells. *Eur J Immunol*. 2008 Oct;38(10):2636–49.
 46. Miossec P, Korn T, Kuchroo VK. Interleukin-17 and type 17 helper T cells. *N Engl J Med*. 2009 Aug;361(9):888–98.
 47. Srenathan U, Steel K, Taams LS. IL-17+ CD8+ T cells: Differentiation, phenotype and role in inflammatory disease. *Immunol Lett*. 2016 Oct;178:20–6.
 48. Albanesi C, Cavani A, Girolomoni G. IL-17 is produced by nickel-specific T lymphocytes and regulates ICAM-1 expression and chemokine production in human keratinocytes: synergistic or antagonistic effects with IFN- γ and TNF- α . *J Immunol*. 1999 Jan;162(1):494–502.
 49. Ghoreschi K, Laurence A, Yang XP, Tato CM, McGeachy MJ, Konkel JE, et al. Generation of pathogenic T(H)17 cells in the absence of TGF- β signalling. *Nature*. 2010 Oct;467(7318):967–71.
 50. Zhao Y, Balato A, Fischelevich R, Chapoval A, Mann DL, Gaspari AA. Th17/Tc17 infiltration and associated cytokine gene expression in elicitation phase of allergic contact dermatitis. *Br J Dermatol*. 2009 Dec;161(6):1301–6.
 51. Nakae S, Komiya Y, Nambu A, Sudo K, Iwase M, Homma I, et al. Antigen-specific T cell sensitization is impaired in IL-17-deficient mice, causing suppression of allergic cellular and humoral responses. *Immunity*. 2002 Sep;17(3):375–87.
 52. He D, Wu L, Kim HK, Li H, Elmetts CA, Xu H. CD8+ IL-17-producing T cells are important in effector functions for the elicitation of contact hypersensitivity responses. *J Immunol*. 2006 Nov;177(10):6852–8.
 53. Martin SF, Dudda JC, Delattre V, Bachtanian E, Leicht C, Burger B, et al. Fas-mediated inhibition of CD4+ T cell priming results in dominance of type 1 CD8+ T cells in the immune response to the contact sensitizer trinitrophenyl. *J Immunol*. 2004 Sep;173(5):3178–85.
 54. Stephen-Victor E, Fickenscher H, Bayry J. IL-26: an emerging proinflammatory member of the IL-10 cytokine family with multifaceted actions in antiviral, antimicrobial, and autoimmune responses. *PLoS Pathog*. 2016 Jun;12(6):e1005624.
 55. Wilson NJ, Boniface K, Chan JR, McKenzie BS, Blumenschein WM, Mattson JD, et al. Development, cytokine profile and function of human interleukin 17-producing helper T cells. *Nat Immunol*. 2007 Sep;8(9):950–7.
 56. Caiazzo G, Di Caprio R, Lembo S, Raimondo A, Scala E, Patruno C, et al. IL-26 in allergic contact dermatitis: resource in a state of readiness. *Exp Dermatol*. 2018

- Jun;27(6):681–4.
57. Xu S, Cao X. Interleukin-17 and its expanding biological functions. *Cell Mol Immunol.* 2010 May;7(3):164–74.
 58. Laan M, Prause O, Miyamoto M, Sjöstrand M, Hytönen AM, Kaneko T, et al. A role of GM-CSF in the accumulation of neutrophils in the airways caused by IL-17 and TNF- α . *Eur Respir J.* 2003 Mar;21(3):387–93.
 59. Hirai Y, Iyoda M, Shibata T, Kuno Y, Kawaguchi M, Hizawa N, et al. IL-17A stimulates granulocyte colony-stimulating factor production via ERK1/2 but not p38 or JNK in human renal proximal tubular epithelial cells. *Am J Physiol Renal Physiol.* 2012 Jan;302(2):F244–50.
 60. Ruddy MJ, Shen F, Smith JB, Sharma A, Gaffen SL. Interleukin-17 regulates expression of the CXC chemokine LIX/CXCL5 in osteoblasts: implications for inflammation and neutrophil recruitment. *J Leukoc Biol.* 2004 Jul;76(1):135–44.
 61. Hartupee J, Liu C, Novotny M, Li X, Hamilton T. IL-17 enhances chemokine gene expression through mRNA stabilization. *J Immunol.* 2007 Sep;179(6):4135–41.
 62. Harper EG, Guo C, Rizzo H, Lillis JV, Kurtz SE, Skorcheva I, et al. Th17 cytokines stimulate CCL20 expression in keratinocytes in vitro and in vivo: implications for psoriasis pathogenesis. *J Invest Dermatol.* 2009 Sep;129(9):2175–83.
 63. Agarwal S, Misra R, Aggarwal A. Interleukin 17 levels are increased in juvenile idiopathic arthritis synovial fluid and induce synovial fibroblasts to produce proinflammatory cytokines and matrix metalloproteinases. *J Rheumatol.* 2008 Mar;35(3):515–9.
 64. Spidale NA, Malhotra N, Frascoli M, Sylvia K, Miu B, Freeman C, et al. Neonatal-derived IL-17 producing dermal $\gamma\delta$ T cells are required to prevent spontaneous atopic dermatitis. *eLife.* 2020 Feb;9:e51188.
 65. Noval Rivas M, Chatila TA. Regulatory T cells in allergic diseases. *J Allergy Clin Immunol.* 2016 Sep;138(3):639–52.
 66. Noh G, Lee JH. Regulatory B cells and allergic diseases. *Allergy Asthma Immunol Res.* 2011 Jul;3(3):168–77.
 67. Cavani A, Nasorri F, Prezzi C, Sebastiani S, Albanesi C, Girolomoni G. Human CD4⁺ T lymphocytes with remarkable regulatory functions on dendritic cells and nickel-specific Th1 immune responses. *J Invest Dermatol.* 2000 Feb;114(2):295–302.
 68. O'Garra A, Vieira P. Regulatory T cells and mechanisms of immune system control. *Nat Med.* 2004 Aug;10(8):801–5.
 69. Taylor A, Verhagen J, Akdis CA, Akdis M. T regulatory cells in allergy and health: a question of allergen specificity and balance. *Int Arch Allergy Immunol.* 2004 Sep;135(1):73–82.
 70. Mackay LK, Rahimpour A, Ma JZ, Collins N, Stock AT, Hafon ML, et al. The developmental pathway for CD103⁺CD8⁺ tissue-resident memory T cells of skin. *Nat Immunol.* 2013 Dec;14(12):1294–301.
 71. Gamradt P, Laoubi L, Nosbaum A, Mutez V, Lenief V, Grande S, Redoulès D, Schmitt AM, Nicolas JF, Vocanson M. Inhibitory checkpoint receptors control CD8⁺ resident memory T cells to prevent skin allergy. *J Allergy Clin Immunol.* 2019 Jun;143(6):2147–2157.e9. doi: 10.1016/j.jaci.2018.11.048.
 72. Boyman O, Werfel T, Akdis CA. The suppressive role of IL-10 in contact and atopic dermatitis. *J Allergy Clin Immunol.* 2012 Jan;129(1):160–1.
 73. Murray PJ. The primary mechanism of the IL-10-regulated antiinflammatory response is to selectively inhibit transcription. *Proc Natl Acad Sci USA.* 2005 Jun;102(24):8686–91.
 74. Berti FC, Pereira AP, Cebinelli GC, Trugilo KP, Brajão de Oliveira K. The role of interleukin 10 in human papilloma virus infection and progression to cervical carcinoma. *Cytokine Growth Factor Rev.* 2017 Apr;34:1–13.
 75. Williams L, Foxwell B. STAT3 is the dominant mediator of the anti-inflammatory effects of IL-10 in human macrophages: 3.2. *Immunol Suppl.* 2004;113:9–10.
 76. Berlato C, Cassatella MA, Kinjyo I, Gatto L, Yoshimura A, Bazzoni F. Involvement of suppressor of cytokine signaling-3 as a mediator of the inhibitory effects of IL-10 on lipopolysaccharide-induced macrophage activation. *J Immunol.* 2002 Jun;168(12):6404–11.
 77. Dolch A, Kunz S, Dorn B, Roers A, Martin SF, Jakob T. Contact allergens induce CD8⁺ T cell-derived interleukin 10 that appears dispensable for regulation of contact hypersensitivity. *Exp Dermatol.* 2017 May;26(5):449–51.
 78. Katz SI, Parker D, Turk JL. B-cell suppression of delayed hypersensitivity reactions. *Nature.* 1974 Oct;251(5475):550–1.
 79. Fillatreau S, Sweeney CH, McGeachy MJ, Gray D, Anderson SM. B cells regulate autoimmunity by provision of IL-10. *Nat Immunol.* 2002 Oct;3(10):944–50.
 80. Grän F, Kerstan A, Serfling E, Goebeler M, Muhammad K. Current Developments in the Immunology of Psoriasis. *Yale J Biol Med.* 2020 Mar;93(1):97–110.
 81. Alrefai H, Muhammad K, Rudolf R, Pham DA, Klein-Hessling S, Patra AK, et al. NFATc1 supports imiquimod-induced skin inflammation by suppressing IL-10 synthesis in B cells. *Nat Commun.* 2016 May;7(1):11724.
 82. Yanaba K, Bouaziz JD, Haas KM, Poe JC, Fujimoto M, Tedder TF. A regulatory B cell subset with a unique CD-1dhiCD5⁺ phenotype controls T cell-dependent inflammatory responses. *Immunity.* 2008 May;28(5):639–50.
 83. Wang K, Tao L, Su J, Zhang Y, Zou B, Wang Y, et al. TLR4 supports the expansion of FasL⁺CD5⁺CD1dhi regulatory B cells, which decreases in contact hypersensitivity. *Mol Immunol.* 2017 Jul;87:188–99.
 84. Kim HS, Lee MB, Lee D, Min KY, Koo J, Kim HW, et al. The regulatory B cell-mediated peripheral tolerance maintained by mast cell IL-5 suppresses oxazolone-induced contact hypersensitivity. *Sci Adv.* 2019 Jul;5(7):eaav8152. <https://doi.org/10.1126/sciadv.aav8152>.
 85. Su J, Wang K, Zhou X, Wang Y, Xu J, Tao L, et al. B-cell-specific-peroxisome proliferator-activated receptor γ deficiency augments contact hypersensitivity with impaired regulatory B cells. *Immunology.* 2019 Mar;156(3):282–96.
 86. Liu X, Huang H, Gao H, Wu X, Zhang W, Yu B, et al. Regulatory B cells induced by ultraviolet B through toll-like receptor 4 signalling contribute to the suppression of contact hypersensitivity responses in mice. *Contact Dermatol.* 2018 Feb;78(2):117–30.
 87. Buters J, Biedermann T. Chromium(VI) Contact Dermatitis: Getting Closer to Understanding the Underlying Mech-

- anisms of Toxicity and Sensitization! *J Invest Dermatol.* 2017 Feb;137(2):274–7.
88. Schmidt M, Raghavan B, Müller V, Vogl T, Fejer G, Tchaptchet S, et al. Crucial role for human Toll-like receptor 4 in the development of contact allergy to nickel. *Nat Immunol.* 2010 Sep;11(9):814–9.
 89. Kobayashi Y. The role of chemokines in neutrophil biology. *Front Biosci.* 2008 Jan;13(1):2400–7.
 90. Wang M, Qin X, Mudgett JS, Ferguson TA, Senior RM, Welgus HG. Matrix metalloproteinase deficiencies affect contact hypersensitivity: stromelysin-1 deficiency prevents the response and gelatinase B deficiency prolongs the response. *Proc Natl Acad Sci USA.* 1999 Jun;96(12):6885–9.
 91. Laan M, Cui ZH, Hoshino H, Lötval J, Sjöstrand M, Gruenert DC, et al. Neutrophil recruitment by human IL-17 via C-X-C chemokine release in the airways. *J Immunol.* 1999 Feb;162(4):2347–52.
 92. Liu R, Lauridsen HM, Amezcua RA, Pierce RW, Jane-Wit D, Fang C, et al. IL-17 promotes neutrophil-mediated immunity by activating microvascular pericytes and not endothelium. *J Immunol.* 2016 Sep;197(6):2400–8.
 93. Ohnishi H, Miyahara N, Gelfand EW. The role of leukotriene B(4) in allergic diseases. *Allergol Int.* 2008 Dec;57(4):291–8.
 94. Oyoshi MK, He R, Li Y, Mondal S, Yoon J, Afshar R, et al. Leukotriene B4-driven neutrophil recruitment to the skin is essential for allergic skin inflammation. *Immunity.* 2012 Oct;37(4):747–58.
 95. Weber FC, Németh T, Csepregi JZ, Dudeck A, Roers A, Ozsvári B, et al. Neutrophils are required for both the sensitization and elicitation phase of contact hypersensitivity. *J Exp Med.* 2015 Jan;212(1):15–22.
 96. Clark RA, Watanabe R, Teague JE, Schlapbach C, Tawa MC, Adams N, Dorosario AA, Chaney KS, Cutler CS, Leboeuf NR, Carter JB, Fisher DC, Kupper TS. Skin effector memory T cells do not recirculate and provide immune protection in alemtuzumab-treated CTCL patients. *Sci Transl Med.* 2012 Jan 18;4(117):117ra7. doi: 10.1126/scitranslmed.3003008.
 97. Rafei-Shamsabadi DA, van de Poel S, Dorn B, Kunz S, Martin SF, Klose CS, et al. Lack of type 2 innate lymphoid cells promotes a type I-driven enhanced immune response in contact hypersensitivity. *J Invest Dermatol.* 2018 Sep;138(9):1962–72.
 98. Rafei-Shamsabadi DA, Klose CS, Halim TY, Tanriver Y, Jakob T. Context Dependent Role of Type 2 Innate Lymphoid Cells in Allergic Skin Inflammation. *Front Immunol.* 2019 Nov;10:2591.
 99. Peng H, Jiang X, Chen Y, Sojka DK, Wei H, Gao X, et al. Liver-resident NK cells confer adaptive immunity in skin-contact inflammation. *J Clin Invest.* 2013 Apr;123(4):1444–56.
 100. Campbell JJ, Qin S, Unutmaz D, Soler D, Murphy KE, Hodge MR, et al. Unique subpopulations of CD56+ NK and NK-T peripheral blood lymphocytes identified by chemokine receptor expression repertoire. *J Immunol.* 2001 Jun;166(11):6477–82.
 101. Ghosh K, Capell BC. The senescence-associated secretory phenotype: critical effector in skin cancer and aging. *J Invest Dermatol.* 2016 Nov;136(11):2133–9.
 102. Carbone T, Nasorri F, Pennino D, Donnarumma M, Garcovich S, Eyerich K, et al. CD56 highCD16 - NK cell involvement in cutaneous lichen planus. *Eur J Dermatol.* 2010 Nov-Dec;20(6):724–30.
 103. Godfrey DI, MacDonald HR, Kronenberg M, Smyth MJ, Van Kaer L. NKT cells: what's in a name? *Nat Rev Immunol.* 2004 Mar;4(3):231–7.
 104. Gober MD, Fischelevich R, Zhao Y, Unutmaz D, Gaspari AA. Human natural killer T cells infiltrate into the skin at elicitation sites of allergic contact dermatitis. *J Invest Dermatol.* 2008 Jun;128(6):1460–9.
 105. Sieling PA. CD1-Restricted T cells: T cells with a unique immunological niche. *Clin Immunol.* 2000 Jul;96(1):3–10.
 106. Deniz G, van de Veen W, Akdis M. Natural killer cells in patients with allergic diseases. *J Allergy Clin Immunol.* 2013 Sep;132(3):527–35.
 107. Balato A, Zhao Y, Harberts E, Groleau P, Liu J, Fischelevich R, et al. CD1d-dependent, iNKT-cell cytotoxicity against keratinocytes in allergic contact dermatitis. *Exp Dermatol.* 2012 Dec;21(12):915–20.
 108. O'Leary JG, Goodarzi M, Drayton DL, von Andrian UH. T cell- and B cell-independent adaptive immunity mediated by natural killer cells. *Nat Immunol.* 2006 May;7(5):507–16.
 109. Pallmer K, Oxenius A. Recognition and regulation of T cells by NK cells. *Front Immunol.* 2016 Jun;7:251.
 110. Rodriguez-Barboza JI, del Rio-Gonzalez ML, Ferreras MC, Buhler L, Fernandez-Renedo C, Perez-Simon JA. NK Cells Regulate CD8 T Cell-Mediated Allogeneic Rejection in Immunocompetent Recipients across an MHC Class I Mismatched Barrier. *Transplantation.* 2018;102:S694–5.
 111. Gordon SM, Chaix J, Rupp LJ, Wu J, Madera S, Sun JC, et al. The transcription factors T-bet and Eomes control key checkpoints of natural killer cell maturation. *Immunity.* 2012 Jan;36(1):55–67.
 112. Daussy C, Faure F, Mayol K, Viel S, Gasteiger G, Charrier E, et al. T-bet and Eomes instruct the development of two distinct natural killer cell lineages in the liver and in the bone marrow. *J Exp Med.* 2014 Mar;211(3):563–77.
 113. Martin SF, Jakob T. From innate to adaptive immune responses in contact hypersensitivity. *Curr Opin Allergy Clin Immunol.* 2008 Aug;8(4):289–93.
 114. Lass C, Merfort I, Martin SF. In vitro and in vivo analysis of pro- and anti-inflammatory effects of weak and strong contact allergens. *Exp Dermatol.* 2010 Nov;19(11):1007–13.
 115. Eberl G, Colonna M, Di Santo JP, McKenzie AN. Innate lymphoid cells. Innate lymphoid cells: a new paradigm in immunology. *Science.* 2015 May;348(6237):aaa6566. <https://doi.org/10.1126/science.aaa6566>.
 116. Spits H, Artis D, Colonna M, Dieffenbach A, Di Santo JP, Eberl G, et al. Innate lymphoid cells—a proposal for uniform nomenclature. *Nat Rev Immunol.* 2013 Feb;13(2):145–9.
 117. Boniface K, Guignouard E, Pedretti N, Garcia M, Delwail A, Bernard FX, et al. A role for T cell-derived interleukin 22 in psoriatic skin inflammation. *Clin Exp Immunol.* 2007 Dec;150(3):407–15.
 118. Caproni M, Antiga E, Melani L, Volpi W, Del Bianco E, Fabbri P. Serum levels of IL-17 and IL-22 are reduced by

- etanercept, but not by acitretin, in patients with psoriasis: a randomized-controlled trial. *J Clin Immunol*. 2009 Mar;29(2):210–4.
119. Eberl G, Marmon S, Sunshine MJ, Rennert PD, Choi Y, Littman DR. An essential function for the nuclear receptor RORgamma(t) in the generation of fetal lymphoid tissue inducer cells. *Nat Immunol*. 2004 Jan;5(1):64–73.
120. Sun Z, Unutmaz D, Zou YR, Sunshine MJ, Pierani A, Brenner-Morton S, et al. Requirement for RORgamma in thymocyte survival and lymphoid organ development. *Science*. 2000 Jun;288(5475):2369–73.
121. Bar-Ephraïm YE, Mebius RE. Innate lymphoid cells in secondary lymphoid organs. *Immunol Rev*. 2016 May;271(1):185–99.
122. Klose CS, Kiss EA, Schwierzeck V, Ebert K, Hoyler T, d'Hargues Y, et al. A T-bet gradient controls the fate and function of CCR6-RORγt+ innate lymphoid cells. *Nature*. 2013 Feb;494(7436):261–5.
123. Rankin LC, Groom JR, Chopin M, Herold MJ, Walker JA, Mielke LA, et al. The transcription factor T-bet is essential for the development of NKp46+ innate lymphocytes via the Notch pathway. *Nat Immunol*. 2013 Apr;14(4):389–95.