

## UPLC-MS/MS Profile Combined With RNA-Seq Reveals the Amino Acid Metabolism in *Zanthoxylum bungeanum* Leaves Under Drought Stress

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Hu H, Fei X, He B, Chen X, Ma L, Han P, Luo Y, Liu Y and Wei A (2022) UPLC-MS/MS Profile Combined With RNA-Seq Reveals the Amino Acid Metabolism in Zanthoxylum bungeanum Leaves Under Drought Stress. Front. Nutr. 9:921742. doi: 10.3389/fnut.2022.921742 Zanthoxylum bungeanum leaves have a unique taste and incomparable nutritional value and hence are popular as a food item and traditional medicine in China. However, the studies on the metabolites in Z. bungeanum leaves are quite limited, especially for amino acids. Therefore, this study explored the amino acid component in Z. bungeanum leaves and also the accumulation mechanism under drought stress in two Z. bungeanum cultivars using the widely targeted metabolome combined with transcriptome analysis. A total of 56 amino acids and their derivatives were identified in Z. bungeanum leaves, including eight essential amino acids. The total amino acid content with most individual amino acids increased under progressive drought stress. More differentially accumulated amino acids (DAAs) and differentially expressed genes (DEGs) were found in FJ (Z. bungeanum cv. 'Fengjiao') than in HJ (Z. bungeanum cv. 'Hanjiao'). The orthogonal projections to latent structures discriminant analysis identified nine and seven indicator DAAs in FJ and HJ leaves, respectively. The weighted gene co-expression network analysis (WGCNA) showed that the green module was significantly correlated with most indicator DAAs and revealed the important role of FBA3, DELTA-OAT, PROC, and 15 transcription factor genes in regulating the amino acid synthesis. Furthermore, the correlation analysis and redundancy analysis (RDA) identified four candidate synthesis genes (ASNS, AK, ASPS, and PK) in amino acid biosynthesis pathway. This study provided useful information for the development of Z. bungeanum leaves in food and nutrition industry and also laid the foundations for future molecular breeding.

Keywords: Zanthoxylum bungeanum leaves, amino acid, metabolome, transcriptome, redundancy analysis

### INTRODUCTION

Zanthoxylum bungeanum Maxim., a member of Rutaceae family, is an economically important plant. Z. bungeanum originated in East Asia and is now widely cultivated in China, Japan, South Korea, and Southeast Asian countries (1). As the main product, the peel of Z. bungeanum is an indispensable condiment in Chinese cuisine (1) and an important traditional Chinese herb with various therapeutic effects (2). The Z. bungeanum leaves are another important product of Z. bungeanum. In recent years, Z. bungeanum leaves have been shown to have a great potential as a food item because of their unique tingling taste, favorable aromatic flavor, and abundant nutrients. For example, the sprouts or young leaves of Z. bungeanum are usually eaten with other vegetables in a salad (3). They are also processed as canned pickles by the food industry for sale. The extract or volatile oil of Z. bungeanum leaves is used as a condiment and edible additive in foods (4). At the same time, the Z. bungeanum leaves can be made into green tea through the roasting process. Moreover, the extract of *Z. bungeanum* leaves can also be used as a food preservative (5). Therefore, Z. bungeanum leaves have broad prospects in the food industry.

Amino acids are the basic units in protein synthesis (6) and are also important raw materials for other secondary metabolites, such as terpenoids, flavonoids, and alkaloids (7, 8). Currently, 22 amino acids for synthesizing proteins have been discovered, including 20 major amino acids and two amino acids encoded by termination codons. The amino acids that the human body needs but cannot synthesize by itself or whose synthesis rate cannot meet the needs of human metabolism are called essential amino acids (9). The United Nations Food and Agriculture Organization stipulates that the percentages of eight kinds of essential amino acids are as follows: leucine 17.2%, isoleucine 12.9%, valine 14.1%, lysine 12.5%, threonine 10%, methionine 10.7%, phenylalanine 19.5%, and tryptophan 3.1%. In addition, histidine is an essential amino acid during infancy because the body cannot synthesize this amino acid during this period (10). Appropriate intake of amino acids is of great significance for regulating human function and maintaining human health (6). Glutamate is an amino acid that can promote red blood cell production, improve brain cell nutrition, and activate the mind (11). Phenylalanine is the raw material for the production of epinephrine, thyroxine, and melanin. Phenylalanine and tyrosine together synthesize important neurotransmitters and hormones, which are involved in the body's glucose metabolism and fat metabolism (12). Tryptophan can improve sleep quality by promoting the secretion of serotonin and melatonin, and regulate the body's biological clock (13). Threonine mainly functions in the gut for synthesizing mucosal proteins and modulating the immune response of the gut (14). Lysine can promote protein absorption and utilization, improve nervous system function, and enhance immune system function (15).

Amino acids are important nutrients in most natural crops and are important evaluation indicators of crop quality. A large number of studies focused on the content, ratio, and synthesis mechanism of amino acid components in various plant products, such as tea leaves (16), *Ginkgo* leaves (17), walnut

(18), and *Lycium barbarum* (19). However, the amino acid component and content in *Z. bungeanum* leaves were rarely mentioned. For plants themselves, the amino acid composition has important implications for regulating plant physiological processes, including plant growth, flowering, and fruiting (20), as well as responses to abiotic stresses (21). Besides, the content of amino acids is easily affected by the environment (21). With the deterioration of the global environment, researches on the quality and yield of economical plant products in harsh environments, including drought, low temperature, and salinization, have attracted increasing attentions (22). *Z. bungeanum* is mainly grown in arid and semi-arid regions. Hence, it is of great significance to study the effect of drought stress on the nutrient composition of leaves.

Recent years witnessed the successful application of multiple omics in investigating the mechanism of metabolite synthesis. For example, Zhang et al. (23) identified the sabinene synthase gene in citrus leaves and flowers using metabolic and transcript data. Liu et al. (24) explored the volatile compounds and revealed the inner metabolism for flavor in apple fruit through the integration of transcriptome and metabolite profiling. Li et al. (25) investigated metabolite accumulation and the molecular mechanisms of flavonoid biosynthesis in jujube leaves using metabolic and transcriptomic profiles. Moreover, Xia et al. (26) integrated metabolic profiling and transcriptome to unveil the pigment accumulation metabolism in Lonicera japonica flowers. Therefore, in this study, the integrative analysis of transcriptome and metabolome was used to investigate the amino acid content in the leaves of Z. bungeanum in response to drought with three specific objectives: (1) to uncover the amino acid components in Z. bungeanum leaves, (2) to explore the effects of drought on amino acid content in leaves of two Z. bungeanum cultivars with different tolerance, and (3) to reveal the molecular mechanism of amino acid metabolism in Z. bungeanum leaves. The results of this study might provide an insight for the development of Z. bungeanum leaves in the food industry and also a pivotal clue for the molecular breeding of Z. bungeanum.

#### MATERIALS AND METHODS

#### **Plant Material and Sample**

The mature seeds of two Z. bungeanum cultivars (FJ, Z. bungeanum cv. 'Fengjiao'; HJ, Z. bungeanum cv. 'Hanjiao') were obtained from the Pickly Ash Experimental Station of Northwest A&F University in Fengxian, Shannxi Province, China ( $33^{\circ}59'6.55''$ N,  $106^{\circ}39'29.38''$ E). After cleaning and airdrying, the Z. bungeanum seeds were sown in a soil mixture of perlite, vermiculite, and chernozem, and then placed in a research greenhouse of Northwest A&F University in Yangling, Shannxi Province, China. After germination, the healthy seedlings were transplanted into cultivar pots, with one seedling per pot. Then, these seedlings were cultivated for 3 months in the greenhouse with temperature at  $25 \pm 2^{\circ}$ C and soil moisture at  $85\% \pm 1\%$ . After that, 36 Z. bungeanum healthy seedlings of the same size were selected in each cultivar and the water supply was stopped for 15 days. The top three fully expanded mature leaves were sampled with liquid nitrogen on days 0 (D1), 6 (D2), 9 (D3), and 15 (D4) and stored in a  $-80^{\circ}$ C freezer. Three biological replications in each sample were performed, with three seedlings in each replication.

#### **Total Amino Acid Content**

The total amino acid content was determined using assay kits (BC1570, Beijing Solarbio Science & Technology Co., Ltd., Beijing, China) following the manufacturer's protocols. In this study, 0.1 g leaf samples were ground into powder with liquid nitrogen and used for the total amino acid determination. The absorbance for all samples was measured at 570 nm using a microplate reader (Infinite M200pro, Tecan, Switzerland).

# Amino Acid Profile in *Zanthoxylum bungeanum* Leaves

The 24 *Z. bungeanum* leaf samples were vacuum freeze-dried with a freeze drier (Scientz-100F, Zhejiang, China) and then ground into powders using a grinding mill (MM 400, Retsch, Germany). One-hundred milligrams of the powders were fully mixed with 1.2 mL of 70% aqueous methanol ( $\nu/\nu$ ) with a Vortex-6 (Kylin-Bell, Jiangsu, China). After standing for 12 h at 4°C, the homogenate was centrifuged at 12,000 rpm at 4°C for 10 min (5424R, Eppendorf Co., Shanghai, China). The supernatant was collected and filtered through a 0.22-µm organic nylon needle filter (SCAA-104, ANPEL, Shanghai, China) into a brown sample bottle.

The quantitative and qualitative analyses of amino acids were performed by Wuhan MetWare Biotechnology Co., Ltd. (Wuhan, China) (27). Ultra-performance liquid chromatography (UPLC) analysis was carried out on the Shimadzu Nexera X2 instrument (Shimadzu, Japan) with an Agilent SB-C18 column (1.8  $\mu$ m, 2.1  $\times$  100 mm). The solvent A (ultrapure water with 0.1% formic acid) and solvent B (acetonitrile with 0.1% formic acid) were used as mobile phases for amino acid metabolites. The procedure was set as follows: 0 min, 5%; 0-9 min, increased to 95%; 9-10 min, 95%; 10-11.10 min, reduced to 5%; and 11.10-14 min, 5%. The tandem mass spectrometry (MS/MS) analysis was performed using an Applied Biosystems 4,500 QTRAP instrument (ABI, MA, United States) equipped with an ESI turbo ion-spray interface. The ESI source was operated as follows: ion source (turbo spray), source temperature (550°C), and ion-spray voltage (5500 V/-4500 V).

#### **RNA Extraction and Sequencing**

Biomarker Technologies Co., Ltd. (Beijing, China) performed the RNA sequencing of 24 *Z. bungeanum* leaf samples. Total RNA from the *Z. bungeanum* leaves was extracted following the instruction on a Tiangen RNA Pure kit for plants (Tiangen, Beijing, China). The integrity and concentration of extracted RNA were determined using an Agilent 2100 Bioanalyzer (Agilent Technologies, Inc., CA, United States). The RNA with an OD260/280 value between 1.8 and 2.2 and an OD260/230 value of more than 2.0 was of high quality. The high-quality RNA was used to construct cDNA libraries employing an NEBNext Ultra RNA Library Prep Kit for Illumina (New England Biolabs, MA, United States). The Illumina HiSeq 2500 platform (Illumina, Inc., CA, United States) was used to sequence the resulting libraries. HISAT2 (28) was used to efficiently align RNA sequencing reads to reference Z. bungeanum genome. StringTie was used to assemble the aligned reads and calculate the fragments per kilobase of transcript per million fragments mapped (FPKM) (29). Differential expression analysis between sample groups was performed using DESeq2 (30), and genes with fold change (FC)  $\geq$  2 and false discovery rate (FDR) < 0.01 were defined as differentially expressed genes (DEGs). The functions of DEGs were annotated using GO (Gene Ontology), KO [Kyoto encyclopedia of genes and genomes (KEGG) Ortholog database), KOG/COG (Clusters of Orthologous Groups of proteins], Nr (NCBI non-redundant protein sequences), Nt (NCBI non-redundant nucleotide sequences), Pfam (Protein family), and Swiss-Prot (a manually annotated and reviewed protein sequence database). The transcriptome raw data has been uploaded to the NCBI SRA database with the accession number PRINA784034.1

#### **Co-expression Network Construction**

Weighted gene co-expression network analysis (WGCNA) was performed using the R package (version: 1.70-3) based on the DEGs with a similarity threshold 0.5 (31). The contents of important differentially accumulated amino acids (DAAs) under drought stress were input as the trait file. The modules with a high correlation to amino acids were identified. The co-expression network was constructed using the top 150 core genes in the green module and visualized using Cytoscape 3.9.1 (Java 11.0.6).

#### Quantitative Real-Time Polymerase Chain Reaction

The specific quantitative primers for quantitative real-time polymerase chain reaction (qRT-PCR) were designed using Primer Premier 6.0 (PREMIER Biosoft, CA, United States) and are provided in Supplementary Table 1. A Tiangen RNA Pure kit for plants (Tiangen, Beijing, China) was used to extract the total RNA from the 24 Z. bungeanum leaf samples. We measured the concentration and integrity of total RNA using a NanoDrop 2000 spectrophotometer (Thermo Scientific, DE, United States). The total RNA with high quality was used to synthesize first-strand cDNA using a PrimeScript RT reagent kit with gDNA Eraser (Takara Biotechnology Inc., Dalian, China). qRT-PCR was performed in a 10 µL volume using SYBR Green Premix Pro Taq HS qPCR Kit (Accurate Biotechnology Co., Ltd., Hunan, China). The operational procedure in a CFX96 Real-Time System (Bio-Rad Laboratories, Inc., CA, United States) was set as follows: 1 cycle at 98°C for 30 s; 38 cycles at 95°C for 5 s, 56°C for 30 s, and 72°C for 30 s; and 4°C for the remaining. The relative transcription abundance was calculated by the  $2^{-\Delta} \Delta CT$  method (32). ZbUBA and ZbUBQ were selected as internal reference genes to balance the gene expression (33).

<sup>&</sup>lt;sup>1</sup>https://dataview.ncbi.nlm.nih.gov/object/PRJNA784034?reviewer= v79bcipc97tajk7ppuam33d88t

#### **Statistical Analyses**

The experimental data were processed using SPSS 23.0 (SPSS Inc., IL, United States), and significant differences among samples were analyzed using Duncan's test in a one-way analysis of variance (P < 0.05). Correlation analysis and hierarchical cluster heatmap analysis (HCA) were performed using the online software "Wu Kong" platform.<sup>2</sup> Principal component analysis (PCA), orthogonal projections to latent structures discriminant analysis (OPLS-DA), and redundancy analysis (RDA) were performed using the OmicShare tools online platform.<sup>3</sup> The column chart was illustrated using Origin2021 (Originlab, MA, United States). The heatmaps of gene expression and amino acid abundance were drawn using TBtools (v. 1.098696, JAVA, China). For each sample, three replicates were included.

#### RESULTS

#### Accumulation of Amino Acid Metabolites in *Zanthoxylum bungeanum* Leaves Under Drought Stress

The qualitative and quantitative detection of amino acids were performed in 24 Z. bungeanum leaf samples to explore the amino acid metabolites in Z. bungeanum leaves. A total of 56 kinds of amino acids and derivatives, including eight essential amino acids, were identified (Supplementary Table 2). On the basis of the abundance of 56 amino acids, PCA was carried out on the 24 samples (including 3 replicates) (Figure 1A). The first two principle components explained 97.4% of the variation (PC1, 73.7%; PC2, 23.7%). FJ and HJ were separated along with PC1, and samples in different stages were separated by PC2. The results suggested that the abundance of amino acids was diverse in two Z. bungeanum cultivars, and drought stress had a large influence on amino acids in Z. bungeanum leaves. The intragroup correlation analysis showed a high correlation between biological replicates but a low correlation between cultivars, suggesting that the metabolite data were accurate and repeatable (Figure 1B). The total amino acid content increased with drought stress in both Z. bungeanum cultivars and was significantly higher in FJ than in HJ after D3 stage (Figure 1C). The HCA was performed to explore the variation in individual amino acid metabolite (Figure 1D). As a result, 56 amino acids were divided into three clusters: I, II, and III. Cluster I comprised 23 amino acid metabolites, including L-methionine, L-threonine, L-arginine, L-tryptophan, and L-cysteine. The contents of these amino acids were higher in FJ than in HJ. Likewise, 23 amino acids were contained in cluster III, including L-cystine, L-proline, L-histidine, L-valine, L-lysine, and L-leucine. Interestingly, the amino acids contents in both cultivars presented an increasing trend in cluster III (Supplementary Figure 1). In Cluster II, only 10 amino acids were present with higher content in HJ than in FJ. With regard to the total variation in 56 amino acids, the violin diagram on the basis of the content of 56 amino acids was illustrated. The total trend showed that the amino acids accumulated in both cultivars, especially after D3 stage, and the amino acids contents were higher in FJ than in HJ, which was in line with the total amino acid content.

## The Differentially Accumulated Amino Acids in Different Groups

The amino acids with FC > 2 or FC < 0.05 were defined as DAAs. Venn diagrams combined with column chart was used to depict differential metabolites in different comparison groups (Figure 2A). A total of 4, 16, and 21 DAAs were found in F1vsF2, F1vsF3, and F1vsF4, respectively. Interestingly, three DAAs (L-norleucine, L-histidine, and glutathione reduced form) were overlapped in three comparison groups. We speculated that these three amino acids might be particularly sensitive to drought, and thus their accumulation was disturbed in each stage of drought stress. A total of 4, 4, and 15 DAAs were found in H1vsH2, H1vsH3, and H1vsH4, respectively; no DAAs coexisted in the three comparisons. The fewer DAAs in HJ than in FJ implied that the amino acids in HJ were more stable under drought stress. A total of 14, 17, 13, and 17 DAAs were observed in F1vsH1, F2vsH2, F3vsH3, and F4vsH4, respectively. The result indicated differences in amino acid contents in Z. bungeanum leaves between the two cultivars.

We screened indicator DAAs with a variable influence on projection (VIP) > 1 using OPLS-DA, and these DAAs had a large contribution rate in the comparison group (Figure 2B). In F4vsF1, nine indicator DAAs were identified, namely, L-phenylalanine, pipecolic acid, L-tyrosine, L-valine, L-tryptophan, cycloleucine, L-arginine, L-isoleucine, and L-leucine, indicating that drought mainly affected the change in the amino acid content in FJ. In H4vsH1, seven important DAAs were L-phenylalanine, L-tyrosine, L-valine, L-tryptophan, pipecolic acid, L-leucine, and L-isoleucine, indicating that the seven amino acids were largely responsible for the variation in amino acids under drought stress in HJ. Notably, these seven amino acids were also included in important DAAs for FJ. In addition, six indicator DAAs (pipecolic acid, L-tyramine, cycloleucine, L-tryptophan, reduced glutathione, and L-phenylalanine) were found in HJvsFJ, indicating that these amino acids were the main reason for the difference in amino acids in Z. bungeanum leaves between two cultivars.

#### Transcription Analysis in *Zanthoxylum bungeanum* Leaves Under Drought Stress

The gene expression levels of two *Z. bungeanum* cultivars in four drought stages were analyzed using RNA-seq. A total of 85,942 genes were assembled and annotated in this transcriptome, and the Q30 base percentage in each sample was more than 92.96%. In FJ, 43,536, 44,647, 44,548, and 40,155 unigenes (FPKM > 1) were present in F1, F2, F3, and F4, respectively (**Figure 3A**), and 35,842 common genes were across the four periods. In HJ, 42,218, 43,294, 42,682, and 41,901 unigenes (FPKM > 1) were found in H1, H2, H3, and H4, respectively, and 36,456 genes existed commonly in the four periods (**Figure 3D**).

<sup>&</sup>lt;sup>2</sup>https://www.omicsolution.com/wkomics/main/

<sup>&</sup>lt;sup>3</sup>https://www.omicshare.com/tools



analysis of 24 ZB leaf samples. (C) lotal amino acid content of ZB leaves under progressive drought stress. Hed columns indicating the total amino acid content in ZB cultivar FJ; green columns indicating the total amino acid content in ZB cultivar HJ. D1, D2, D3, and D4 indicating the ZB leaves sampled 0, 6, 9, and 15 days after drought treatment. (D) Hierarchical clustering heatmap analysis (HCA) indicating the abundance of 56 individual amino acids in ZB leaves. Red and blue representing the high and low amino acid abundance, respectively. The above violin diagram and curve chart indicating the total variation in the 56 individual amino acids. F1, F2, F3, and F4 indicating the FJ samples in D1, D2, D3, and D4, respectively; H1, H2, H3, and H4 indicating the HJ samples in D1, D2, D3, and D4, respectively.

According to differential expression analysis between sample groups, the genes with FDR < 0.05 and a FC  $\geq$  2.0 were considered as DEGs. In FJ, 14,819 DEGs were identified under drought stress, with 6,089 genes upregulated and 8,730 genes downregulated (**Figure 3B**). In HJ, 13,779 DEGs were identified across the four stages, with 5,052 DEGs upregulated and 8,574 DEGs downregulated (**Figure 3E**). KEGG analysis and GO analysis were used to explore the possible functions of these DEGs. The KEGG bubble diagram showed the top 20 KEGG pathways enriched by these DEGs, eight of which were related to amino acid

biosynthesis and mechanism in both cultivars (**Figures 3C,F**). Additionally, starch and sucrose metabolism and mitogenactivated protein kinase (MAPK) signaling pathways were also highly enriched. In FJ, the top 20 GO terms were divided into three categories, named as biological process (BP), molecular function (MF), and cellular component (CC), including translation (GO:0006412) and amide biosynthesis process (GO:0043604) in BP, structural constituent of ribosome (GO:0003735) and structural molecule activity (GO:0005198) in MF, and cytosolic ribosome (GO:0022626) in CC (**Figure 3G**). In HJ, only two GO classes were contained in the top 20 GO terms,



FIGURE 2 | Differentially analysis of amino acid in ZB leaves. (A) Venn diagrams of differentially accumulated amino acids (DAAs). The below columns presented the numbers of each group. (B) Orthogonal projections to latent structures discriminant analysis (OPLS-DA) in F4vsF1, H4vsH1, and HJvsFJ. The table on the top right position in each diagram showed the indicator DAAs and the corresponding variable influence on projection (VIP) values.

including maturation of lsu-rRNA (GO:0000470) and ribosomal large subunit biogenesis (GO:0042273) in BP, and small-subunit processome (GO:0032040) and nucleolar part (GO:0044452) in CC (**Figure 3H**). In general, the DEGs under drought stress were related to the amino acid biosynthesis and metabolism, as well as plant signal transcription.

#### Weighted Gene Co-expression Network Analysis of Differentially Expressed Genes

Weighted gene co-expression network analysis was performed based on the DEGs combined with nine indicator DAAs in OPLS-DA under drought stress to investigate the molecular mechanism of amino acid biosynthesis in *Z. bungeanum* leaves under drought stress (**Figure 4** and **Supplementary Figure 1A**). As a result, eight modules were produced according to the co-expression relationship (**Figure 4B**). More than 1,000 genes were observed in magenta, brown, and green modules; however, less than 100 genes existed in dark green, blue, and royal blue modules. Blue, green, and orange modules were positively related to most of these amino acids. Relatively higher correlation coefficients existed in green module, which significantly correlated to blue (R = 0.79, P = 0.002) and orange (R = 0.71, P = 0.04) modules (Supplementary Figure 1B). The overall gene expression pattern varied among the eight modules, which was visualized in a heatmap combined with a column chart (Figure 4C). Notably, the expression levels of most genes in the green module increased under drought stress and were higher in FJ than in HJ, which was in accordance to the varied trend in the contents of most amino acids. Taken together, the green module was selected as the most important module for further experimentation. The GO analysis showed that a large number of DEGs were enriched in metabolic process, response to stimulus and signaling in BP, cell and membrane in CC, and catalytic activity and transporter activity in MF (Supplementary Figure 2A). The KEGG analysis revealed



that most DEGs in the green module were significantly enriched in plant hormone and signal transduction, MAPK signaling pathway, and biosynthesis and metabolism of multiple amino acids, such as cysteine and methionine metabolism, arginine and proline metabolism, and phenylalanine tryptophan tyrosine biosynthesis (**Supplementary Figure 2B**).

The gene co-expression network was constructed using the top 150 core genes in the green module (Figure 5). Among



the 150 genes, three members were involved in amino acid biosynthesis pathway, named fructose-bisphosphate aldolase 3 (FBA3), ornithine aminotransfera (DELTA-OAT), and pyrroline-5-carboxylate reductase (PROC). Transcriptional factors (TFs) play important roles in regulating the expression level of target genes. A total of 15 TF genes were found in the coexpression network, including two AP2/ERFs (EVM0014104 and EVM0095435), eight HD-ZIPs (EVM0001528, EVM0020518, EVM0028024, EVM0043364, EVM0049040, EVM0058931, EVM0061236, and EVM0071427), two HSFs (EVM0020970 and EVM0054003), two MYBs (EVM0002246 and EVM0097168), and one WRKY (EVM0075014). Further, 10 hub genes were screened out according to the connectivity, including DELTA-OAT and HD-ZIP; their expression level increased with drought stress, which is shown by the column chart. Therefore, the 3 structure genes and 15 TF genes might play important roles in amino

acid accumulation in *Z. bungeanum* leaves. Additionally, the expression patterns of these 18 genes were analyzed using qRT-PCR (**Supplementary Figure 3**). The relative expression levels of these 18 genes were consistent with FPKM values in RNA-seq, which validated the reliability of our transcriptome data.

#### Gene Expression Level and Amino Acid Content in the Amino Acid Biosynthesis Pathway

A pathway diagram with gene expression and amino acids abundance was constructed in *Z. bungeanum* leaves according to the amino acid biosynthesis pathway reported in model plants (**Figure 6**). A total of 15 amino acids and 19 DEGs were mapped to the pathway. The contents of threonine, cysteine, and methionine decreased in both cultivars under drought



stress. Consistently, the expression levels of D-3-phosphoglycerate dehydrogenase (PGDH) and serine hydroxymethyltransferase (SHM) also decreased. The histidine content was high under drought; however, a decreased expression level was presented in imidazoleglycerol-phosphate dehydratase (HIS) and ribose-5-phosphate isomerase (RPI). The contents of isoleucine, valine, and leucine increased with the increased expression levels of pyruvate kinase 1 (PK) and branched-chain-aminoacid aminotransferase (BCAT). Differently, the tryptophan accumulated with the increased expression of *indole-3-glycerol* phosphate synthase (IGPS), and the contents of tyrosine and phenylalanine increased with the decreased expression of arogenate dehydratase/prephenate dehydratase 6 (ADT). Tricarboxylic acid (TCA) cycle is an important respiration pathway whose intermediate products also participate in amino acid synthesis. For example, oxaloacetate was the product of aspartate and lysine; a-ketoglutarate was converted into

glutamate by aspartate aminotransferase 3 (ASP3), and glutamate could be further converted into proline and arginine. In *Z. bungeanum* leaves, the lysine content increased with the higher *aspartokinase* (AK) gene expression level. The content of proline and glutamate increased with the elevated expression of ASP3 and PROC. Generally speaking, most amino acids varied in accordance with the genes encoding synthesis enzymes despite several exceptions.

### Correlation Analysis and Redundancy Analysis Between Amino Acid Content and Expression of Synthesis-Related Genes

The intergroup correlation between amino acid content and related genes in the biosynthesis pathway was analyzed in FJ and HJ to further explore the important genes involved in amino



dehydratase/prephenate dehydratase 6; ASP3, aspartate aminotransferas 4-decarboxylase; ARGG, argininosuccinate synthase.

acid synthesis (**Figures 7A,B**). In FJ, the gene expression of *ASP3, ASNS, PROC, BCAT,* and *IGPS* positively correlated to the content of most amino acids, including arginine, histidine, isoleucine, leucine, lysine, phenylalanine, proline, tryptophan, tyrosine, and valine as well as the total amino acid content. Methionine content highly correlated with the expression of *RPI, ADT6, SHM, ASB1, AROC,* and *MDH (R > 0.8, P < 0.05),* while threonine content positively correlated with the expression of *RPI, ASB1, and MDH.* In HJ, the gene expression levels of *AK, PK, MS, ASP3, ASNS, IDH, PROC, BCAT,* and *IGPS* were positively associated with the contents of most amino acids, including histidine, leucine, lysine, tryptophan, and valine. The arginine

content positively correlated with the expression of *RPI*, *HIS*, and *ADT6*. The methionine content positively correlated with the expression of *PGDH* and *SHM*. Besides, the total amino acid content highly correlated with the expression of *PK*, *ASP3*, *IDH*, *PROC*, and *BCAT*. These genes with positive correlation might be important in the biosynthesis of amino acids. Notably, TCA might play important roles in amino acid biosynthesis in HJ rather than FJ because of the positive correlation of the expression of *MS* and *IDH* with the contents of most amino acids in the biosynthesis pathway.

RDA was widely used to reveal the important factors in metabolite synthesis. RDA was performed using the contents



of 15 amino acids and the expression levels of 19 synthesisrelated genes in the aforementioned pathway. The expression levels of the genes related to amino acid synthesis explained 96.79 and 95.91% of the contribution rate in FJ and HJ, respectively, indicating that the RDA results were reliable and repeatable. In both FJ and HJ, a remarkable separation was found among samples in D1, D3, and D4, which was not significant between D1 and D2. The results indicated that drought stress had a significant influence on amino acids after mid drought stage. In FJ, the expression levels of ASP3 and ASNS mostly correlated with the contents of tryptophan, histidine, phenylalanine, tyrosine, valine, and proline. The expression levels of ARGE and PK were significantly associated with the contents of isoleucine, leucine, and lysine. Besides, AK might play an important role in cysteine synthesis due to the smallest angle. However, the expression level of the other six genes did not significantly

correlate with the contents of the aforementioned amino acids. In HJ, the expression of *ASNS*, *AK*, *ASPS*, and *PK* positively correlated with the 10 amino acids. In general, *ASNS*, *AK*, *ASPS*, and *PK* played important roles in amino acid biosynthesis in *Z. bungeanum* leaves.

#### DISCUSSION

The young leaves and buds of *Z. bungeanum* are a popular food, with unique taste and abundant nutrition. The mature *Z. bungeanum* leaves can be served as a food condiment and a Chinese medicinal herb. So far, the understanding of the metabolic components in *Z. bungeanum* leaves is quite limited. This study revealed the amino acid profile in *Z. bungeanum* leaves using a wide-targeted metabolome and investigated the

amino acid synthesis mechanism in *Z. bungeanum* leaves under drought stress by the conjoint analysis of the transcriptome and metabolome. The results increased our understanding of amino acid components in *Z. bungeanum* leaves, amino acid accumulation pattern under drought stress, and the corresponding mechanism, as well as the key genes regulating amino acid synthesis, thereby providing reference for plantbased food industry and laying the foundation for further molecular breeding.

Amino acids are the basic units of protein synthesis, which is of extraordinary significance to living organisms. Totally, 20 basic amino acids are involved in protein synthesis and eight of these are classified as essential amino acids according to the demand of human body, including leucine, isoleucine, lysine, methionine, tryptophan, valine, threonine, and phenylalanine. These essential amino acids need to be obtained from the daily diet. Therefore, understanding the amino acid composition in food is of great significance for a reasonable diet. The content and composition of amino acids in plant-based foods have been extensively studied, such as Torreya grandis (34), green tea (16), Ginkgo leaves (17), and date palm leaves (35). In this study, 56 types of amino acids and their derivatives, including eight essential amino acids and histidine, were identified in Z. bungeanum leaves using UPLC-MS/MS (Figure 1 and Supplementary Table 2). The result suggested that the Z. bungeanum leaves were rich in amino acids. The ingestion of Z. bungeanum leaves could help supplement the amino acid content in the human body.

Unlike animals, plants are stationary and cannot move to avoid unfavorable environments. In the long-term evolution process, higher plants have evolved different mechanisms to reduce the damage caused by unfavorable environment, and many of these mechanisms are related to amino acid metabolism (21). A general accumulation of free amino acids is usually observed in different plants exposed to drought stress; however, the reason for accumulation may be different for individual amino acids (36, 37). Consistently, the total free amino acid content increased especially in the mid and late drought stage in Z. bungeanum leaves (Figure 1C). The accumulation of proline and y-aminobutyric acid (GABA) not only regulated the osmotic pressure but also served as reactive oxygen species scavengers to reduce oxidative toxicity (21, 38). Proline synthesis was significantly stimulated under drought stress (39). Additionally, glutathione (GSH), which functioned as an antioxidant, played numerous roles in plant cells, including stress tolerance, regulation of cellular redox balance, modulation of the expression of stress-related genes, and plant growth and senescence (40). The glutathione-ascorbate cycle is considered one of the most pivotal metabolic pathways for the detoxification of  $H_2O_2$  (41). The overexpression of GSH biosynthesis genes enhanced the drought tolerance in transgenic plants (42). Besides, the exogenous application of GSH was successful in increasing the plant tolerance to drought stress (43). In Z. bungeanum leaves, the contents of both proline and reduced glutathione were upregulated under drought conditions (Figure 1D), suggesting that the upregulated amino acid content contributed to the drought

tolerance in Z. bungeanum. In addition, amino acids are precursors of some signaling molecules that help regulate other substances to regulate drought responses. For example, arginine is the precursor substance of polyamines, which is important in drought response and widely investigated in many crops (44). Lysine can be converted into immune signaling molecule N-hydroxy pipecoline (45). In FJ leaves, the contents of these two amino acids were upregulated under drought stress. Furthermore, our transcriptome analysis found that the DEGs related to amino acid synthesis were largely enriched in plant hormone and signal transduction (**Supplementary Figure 2**).

Differential analysis of amino acid metabolites showed more DAMs in FJ than in HJ at three drought stages, which was in agreement with DEGs (Figures 2, 3). This result indicated that HJ, the tolerant Z. bungeanum cultivar, had a more stable homeostatic system under drought conditions, thus reducing the disturbance from adverse environments. OPLS-DA was used to screen out the DAMs with a significant contribution to the differential group, and these were considered as indicator DAMs (46). The OPLS-DA identified nine indicator DAAs (L-phenylalanine, pipecolic acid, L-tyrosine, L-valine, L-tryptophan, cycloleucine, L-arginine, L-isoleucine, and L-leucine) in FJ and seven indicator DAAs (L-phenylalanine, L-tyrosine, L-valine, L-tryptophan, pipecolic acid, L-leucine, and L-isoleucine) in HJ (Figure 2B). We considered these DAAs to be the drought biomarkers in Z. bungeanum leaves. In addition, the amount of DAAs in the two cultivars was not significantly influenced by the drought stage, implying that the differences in amino acids existed among different Z. bungeanum cultivar leaves. An in-depth amino acids analysis in the leaves of different Z. bungeanum cultivars might be of great significance for Z. bungeanum reasonable utilization in the food and nutrition industry in the future.

Integrating metabolome and transcriptome is an effective tool to study metabolic mechanisms (47). Based on similar trends in the contents of indicator DAAs (input as the trait file) and the transcriptional metabolism levels of DEGs, a WGCNA was performed to explore the mechanism by which drought regulated amino acid accumulation. The green module significantly positively correlated with most indicator amino acids (Figure 4B). The functional annotation showed that the DEGs in this module were related to amino acid synthesis and metabolism (Supplementary Figure 2), indicating that WGCNA was meaningful for the amino acid changes in this study. In the green module, 150 core genes were selected to construct the co-expression network (Figure 5). Among these, three genes were annotated as amino acidrelated genes, namely, FBA3, DELTA-OAT, and PROC. The results of both RNA-seq and qRT-PCR showed the increased trend of FBA3, DELTA-OAT, and PROC, which was consistent with the contents of phenylalanine, tyrosine, valine, tryptophan, leucine, and isoleucine as well as total amino acid content, indicating that these three genes were important for amino acid accumulation. TFs were proteins that could modulate the rate of gene transcription by binding to the 5'-upstream region of target genes (48). They played an important role in metabolite synthesis by regulating gene transcription, protein synthesis, and subsequent altered cellular function (34, 49). For example, proline accumulation was regulated by TFs associated with phosphate starvation (50). In this study, 15 TF genes were identified in the co-expression network, including two *AP2/ERFs*, eight *HD-ZIPs*, two *HSFs*, two *MYBs*, and one *WRKY*, suggesting the possible regulatory role of these TFs in amino acid biosynthesis.

The amino acid biosynthesis pathway is complex and a cross-talk of C metabolism and N metabolism (17). Most of the basic amino acids are biosynthesized using the intermediates of the Embden-Meyerhof pathway and the TCA cycle as the carbon chain backbone, apart from aromatic amino acids and histidine (51). Aromatic amino acids are derived from erythrose-4-phosphate, an intermediate of pentose phosphate pathway, and histidine is biosynthesized using ATP and phosphoribosyl pyrophosphate (51). Based on the data in the transcriptome and metabolome, we constructed an amino acid biosynthesis pathway in Z. bungeanum leaves under drought conditions (Figure 6). Among the 15 amino acids, most amino acid contents were consistent with their synthetic gene expression, with some exceptions, such as ADT and phelylalanine, HIS and histinidine, and MS and methionine. We speculated that the degradation and transformation downstream of these metabolites might be enhanced.

A large scale of secondary metabolites with multiple healthcare functions was derived from the shikimic acid pathway of aromatic amino acids (phenylalanine, tyrosine, and tryptophan) (52). Furthermore, phenylalanine is an important intermediate in the flavonoid synthesis pathway. In this study, drought induced the accumulation of these three aromatic amino acids in Z. bungeanum leaves, which provided more sufficient raw materials for synthesizing secondary metabolites. This was in line with the results of our previous studies, which showed an increase in the flavonoid content in Z. bungeanum leaves under drought conditions (53). Long-chain amino acids (isoleucine, leucine, and valine), which are important supplements for athletes (54), accumulated in the Z. bungeanum leaves under drought stress. Therefore, we speculated that Z. bungeanum leaves had the potential to develop sports drinks; however, more in-depth validation needs to be performed. On the whole, drought leads to an upregulation of most amino acids, including seven essential amino acids. The correlation analysis and RDA were carried out on the amino acid content and gene expression level in the amino acid biosynthesis pathway to explore the pivotal synthesis-related genes in amino acid biosynthesis (Figure 7). As a result, four amino acid synthesisrelated genes having a significant positive correlation with most of the amino acids were sheltered, named ASP3, ASNS, PK, and AK. In a recent study, the overexpression of four unigenes positively correlated with the contents of at least 10 amino acids contributing to amino acid biosynthesis in Torreya grandis (34). Thus, these four genes can be used as candidate genes for improving amino acid content in Z. bungeanum leaves in future studies.

#### CONCLUSION

In this study, 56 amino acids and their derivatives were identified in the Z. bungeanum leaves using UPLC-MS/MS, including eight essential amino acids. Drought stress caused an increase in the total amino acid content and the contents of most amino acids, including seven essential amino acids, which indicated that the drought environment increased the amino acid nutrition level in Z. bungeanum leaves. WGCNA identified three amino acid synthesis-related genes, FBA3, DELTA-OAT, and PROC, and 15 TF genes, which might participate in the biosynthesis of amino acids in Z. bungeanum leaves under drought conditions. In addition, the correlation analysis and RDA analysis screened out four candidate genes (ASNS, AK, ASPS, and PK) in amino acid biosynthesis in the leaves of two Z. bungeanum cultivars. This study provided useful information for the development of food nutrition and also important clues for the cultivation of high-amino acid *Z. bungeanum* leaves.

#### DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/ **Supplementary Material**.

## AUTHOR CONTRIBUTIONS

HH: methodology, software, data curation, and writing—original draft. XF: supervision and writing—review and editing. BH: validation and investigation. LM and XC: data curation. PH and YLu: investigation. YLi: resources and supervision. AW: funding acquisition and resources. All authors contributed to the article and approved the submitted version.

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#### SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnut.2022. 921742/full#supplementary-material

#### REFERENCES

- Zhang M, Wang J, Zhu L, Li T, Jiang W, Zhou J, et al. Zanthoxylum bungeanum Maxim. (Rutaceae): a systematic review of its traditional uses, botany, phytochemistry, pharmacology, pharmacokinetics, and toxicology. Int J Mol Sci. (2017) 18:2172. doi: 10.3390/ijms18102172
- Okagu IU, Ndefo JC, Aham EC, Udenigwe CC. *Zanthoxylum* species: a review of traditional uses, phytochemistry and pharmacology in relation to cancer, infectious diseases and sickle cell anemia. *Front Pharmacol.* (2021) 12:713090. doi: 10.3389/fphar.2021.713090
- Deng Z, Sun B, Kang K, Dong Y. Analysis of the main nutritional labeling in the tender bud of *Zanthoxylum bungeanum*. J Northwest For Coll. (2005) 20:179–80.
- Zhang Y, Luo Z, Wang D, He F, Li D. Phytochemical profiles and antioxidant and antimicrobial activities of the leaves of *Zanthoxylum bungeanum*. *Sci World J.* (2014) 2014:181072. doi: 10.1155/2014/181072
- Li J, Hui T, Wang F, Li S, Cui B, Cui Y, et al. Chinese red pepper (*Zanthoxylum bungeanum* Maxim.) leaf extract as natural antioxidants in salted silver carp (*Hypophthalmichthys molitrix*) in dorsal and ventral muscles during processing. *Food Control.* (2015) 56:9–17.
- Church DD, Hirsch KR, Park S, Kim IY, Gwin JA, Pasiakos SM, et al. Essential amino acids and protein synthesis: insights into maximizing the muscle and whole-body response to feeding. *Nutrients*. (2020) 12:3717. doi: 10.3390/ nu12123717
- Itam M, Mega R, Tadano S, Abdelrahman M, Matsunaga S, Yamasaki Y, et al. Metabolic and physiological responses to progressive drought stress in bread wheat. Sci Rep. (2020) 10:17189. doi: 10.1038/s41598-020-74303-6
- 8. Wang G, Xu M, Wang W, Galili G. Fortifying horticultural crops with essential amino acids: a review. *Int J Mol Sci.* (2017) 18:1306. doi: 10.3390/ijms18061306
- 9. Lopez MJ, Mohiuddin SS. *Biochemistry, Essential Amino Acids*. Treasure Island, FL: StatPearls Publishing LLC (2022).
- Yu Y-M, Fukagawa NK. Chapter 2–Protein and amino acids. 11th ed. In: Marriott BP, Birt DF, Stallings VA, Yates AA editors. *Present Knowledge in Nutrition*. Cambridge, MA: Academic Press (2020). p. 15–35.
- Stanimirovic DB, Ball R, Small DL, Muruganandam A. Developmental regulation of glutamate transporters and glutamine synthetase activity in astrocyte cultures differentiated *in vitro*. *Int J Dev Neurosci*. (1999) 17:173–84. doi: 10.1016/s0736-5748(99)00028-3
- Tessari P, Vettore M, Millioni R, Puricelli L, Orlando R. Effect of liver cirrhosis on phenylalanine and tyrosine metabolism. *Curr Opin Clin Nutr Metab Care*. (2009) 13:81–6. doi: 10.1097/MCO.0b013e32833383af
- Minet-Ringuet J, Le Ruyet P, Tomé D, Even PA. Tryptophan-rich protein diet efficiently restores sleep after food deprivation in the rat. *Behav Brain Res.* (2004) 152:335–40. doi: 10.1016/j.bbr.2003.10.018
- 14. Tang Q, Tan P, Ma N, Ma X. Physiological functions of threonine in animals: beyond nutrition metabolism. *Nutrients*. (2021) 13:2592.
- Fini M, Torricelli P, Giavaresi G, Carpi A, Nicolini A, Giardino R. Effect of L-lysine and L-arginine on primary osteoblast cultures from normal and osteopenic rats. *Biomed Pharmacother*. (2001) 55:213–20. doi: 10.1016/S0753-3322(01)00054-3
- Salman S, Y1lmaz C, Gökmen V, Özdemir F. Effects of fermentation time and shooting period on amino acid derivatives and free amino acid profiles of tea. *LWT*. (2021) 137:110481. doi: 10.1016/j.lwt.2020.110481
- Guo Y, Wang M, Gao C, Fu F-F, Wang T, El-Kassaby YA, et al. Amino acid metabolism reprogramming in response to changing growth environment in ginkgo biloba leaves. *LWT*. (2021) 144:111276. doi: 10.1016/j.lwt.2021.111276
- Liu B, Liang J, Zhao D, Wang K, Jia M, Wang J. Morphological and compositional analysis of two walnut (*Juglans regia* L.) cultivars growing in China. *Plant Foods Hum Nutr.* (2020) 75:116–23. doi: 10.1007/s11130-019-00794-y
- Wang Y, Liang X, Li Y, Fan Y, Li Y, Cao Y, et al. Changes in metabolome and nutritional quality of *Lycium barbarum* fruits from three typical growing areas of China as revealed by widely targeted metabolomics. *Metabolites*. (2020) 10:46. doi: 10.3390/metabo10020046
- Mondal MF, Asaduzzaman M, Tanaka H, Asao T. Effects of amino acids on the growth and flowering of *Eustoma grandiflorum* under autotoxicity in closed hydroponic culture. *Sci. Hortic.* (2015) 192:453–9. doi: 10.1016/j.scienta.2015. 05.024

- Batista-Silva W, Heinemann B, Rugen N, Nunes-Nesi A, Araújo WL, Braun HP, et al. The role of amino acid metabolism during abiotic stress release. *Plant Cell Environ.* (2019) 42:1630–44. doi: 10.1111/pce.13518
- 22. Zhu JK. Abiotic stress signaling and responses in plants. *Cell.* (2016) 167:313–24. doi: 10.1016/j.cell.2016.08.029
- Zhang H, Chen M, Wen H, Wang Z, Chen J, Fang L, et al. Transcriptomic and metabolomic analyses provide insight into the volatile compounds of citrus leaves and flowers. *BMC Plant Biol.* (2020) 20:7. doi: 10.1186/s12870-019-2 222-z
- 24. Liu X, Hao N, Feng R, Meng Z, Li Y, Zhao Z. Transcriptome and metabolite profiling analyses provide insight into volatile compounds of the apple cultivar 'ruixue' and its parents during fruit development. *BMC Plant Biol.* (2021) 21:231. doi: 10.1186/s12870-021-03032-3
- Li S, Deng B, Tian S, Guo M, Liu H, Zhao X. Metabolic and transcriptomic analyses reveal different metabolite biosynthesis profiles between leaf buds and mature leaves in *Ziziphus jujuba* Mill. *Food Chem.* (2021) 347:129005. doi: 10.1016/j.foodchem.2021.129005
- 26. Xia Y, Chen W, Xiang W, Wang D, Xue B, Liu X, et al. Integrated metabolic profiling and transcriptome analysis of pigment accumulation in *Lonicera japonica* flower petals during colour-transition. *BMC Plant Biol.* (2021) 21:98. doi: 10.1186/s12870-021-02877-y
- Liu Y, Lv J, Liu Z, Wang J, Yang B, Chen W, et al. Integrative analysis of metabolome and transcriptome reveals the mechanism of color formation in pepper fruit (*Capsicum annuum* L.). *Food Chem.* (2020) 306:125629. doi: 10.1016/j.foodchem.2019.125629
- 28. Kim D, Langmead B, Salzberg SL. Hisat: a fast spliced aligner with low memory requirements. *Nat Methods*. (2015) 12:357–60. doi: 10.1038/nmeth.3317
- Pertea M, Pertea GM, Antonescu CM, Chang T-C, Mendell JT, Salzberg SL. Stringtie enables improved reconstruction of a transcriptome from Rna-Seq reads. *Nat Biotechnol.* (2015) 33:290–5. doi: 10.1038/nbt.3122
- Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for Rna-Seq data with Deseq2. *Genome Biol.* (2014) 15:550. doi: 10.1186/s13059-014-0550-8
- Langfelder P, Horvath S. Wgcna: an R package for weighted correlation network analysis. *BMC Bioinformatics*. (2008) 9:559. doi: 10.1186/1471-2105-9-559
- Schmittgen TD, Livak KJ. Analyzing real-time pcr data by the comparative Ct method. Nat Protoc. (2008) 3:1101–8. doi: 10.1038/nprot.2008.73
- Fei X, Shi Q, Yang T, Fei Z, Wei A. Expression stabilities of ten candidate reference genes for Rt-Qpcr in *Zanthoxylum bungeanum* maxim. *Molecules (Basel, Switzerland).* (2018) 23:802. doi: 10.3390/molecules2304 0802
- Lou H, Yang Y, Zheng S, Ma Z, Chen W, Yu C, et al. Identification of key genes contributing to amino acid biosynthesis in *Torreya grandis* using transcriptome and metabolome analysis. *Food Chem.* (2022) 379:132078. doi: 10.1016/j.foodchem.2022.132078
- Du B, Kruse J, Winkler JB, Alfarray S, Schnitzler JP, Ache P, et al. Climate and development modulate the metabolome and antioxidative system of date palm leaves. J Exp Bot. (2019) 70:5959–69. doi: 10.1093/jxb/erz361
- 36. Martinelli T, Whittaker A, Bochicchio A, Vazzana C, Suzuki A, Masclaux-Daubresse C. Amino acid pattern and glutamate metabolism during dehydration stress in the 'resurrection' plant *Sporobolus stapfianus*: a comparison between desiccation-sensitive and desiccation-tolerant leaves. J Exp Bot. (2007) 58:3037–46. doi: 10.1093/jxb/erm161
- Showler AT. Effects of water deficit stress, shade, weed competition, and kaolin particle film on selected foliar free amino acid accumulations in cotton, *Gossypium hirsutum* (L.). *J Chem Ecol.* (2002) 28:631–51. doi: 10.1023/A: 1014556515489
- Shelp BJ, Aghdam MS, Flaherty EJ. Γ-Aminobutyrate (Gaba) regulated plant defense: mechanisms and opportunities. *Plants (Basel, Switzerland)*. (2021) 10:1939. doi: 10.3390/plants10091939
- Verslues PE, Sharma S. Proline metabolism and its implications for plantenvironment interaction. *Arabidopsis Book*. (2010) 8:e0140. doi: 10.1199/tab. 0140
- Gill SS, Anjum NA, Hasanuzzaman M, Gill R, Trivedi DK, Ahmad I, et al. Glutathione and glutathione reductase: a boon in disguise for plant abiotic stress defense operations. *Plant Physiol Biochem.* (2013) 70:204–12. doi: 10. 1016/j.plaphy.2013.05.032

- Noctor G, Foyer CH. Ascorbate and glutathione: keeping active oxygen under control. Annu Rev Plant Physiol Plant Mol Biol. (1998) 49:249–79. doi: 10. 1146/annurev.arplant.49.1.249
- 42. Gaber A, Burritt DJ, Hossain MA. Transgenic plants over-expressing glutathione biosynthetic genes and abiotic stress tolerance. In: Hossain MA, Mostofa MG, Diaz-Vivancos P, Burritt DJ, Fujita M, Tran L-SP editors. *Glutathione in Plant Growth, Development, and Stress Tolerance.* Cham: Springer International Publishing (2017). p. 397–412.
- Nahar K, Hasanuzzaman M, Alam MM, Fujita M. Glutathione-induced drought stress tolerance in mung bean: coordinated roles of the antioxidant defence and methylglyoxal detoxification systems. *AoB Plants*. (2015) 7:lv069. doi: 10.1093/aobpla/plv069
- Winter G, Todd CD, Trovato M, Forlani G, Funck D. Physiological implications of arginine metabolism in plants. *Front Plant Sci.* (2015) 6:534. doi: 10.3389/fpls.2015.00534
- Hartmann M, Zeier T, Bernsdorff F, Reichel-Deland V, Kim D, Hohmann M, et al. Flavin monooxygenase-generated N-hydroxypipecolic acid is a critical element of plant systemic immunity. *Cell.* (2018) 173:456–69.e16. doi: 10.1016/ j.cell.2018.02.049
- 46. Thévenot EA, Roux A, Xu Y, Ezan E, Junot C. Analysis of the human adult urinary metabolome variations with age, body mass index, and gender by implementing a comprehensive workflow for univariate and opls statistical analyses. J Proteome Res. (2015) 14:3322–35. doi: 10.1021/acs.jproteome. 5b00354
- Gong C, Diao W, Zhu H, Umer MJ, Zhao S, He N, et al. Metabolome and transcriptome integration reveals insights into flavor formation of 'crimson' watermelon flesh during fruit development. *Front Plant Sci.* (2021) 12:629361. doi: 10.3389/fpls.2021.629361
- Riechmann JL, Heard J, Martin G, Reuber L, Jiang C, Keddie J, et al. Arabidopsis transcription factors: genome-wide comparative analysis among eukaryotes. *Science (New York, NY)*. (2000) 290:2105–10. doi: 10.1126/science. 290.5499.2105
- 49. Ma W, Kang X, Liu P, She K, Zhang Y, Lin X, et al. The Nac-like transcription factor Csnac7 positively regulates the caffeine biosynthesis-related gene Yhnmt1 in Camellia sinensis. *Hortic Res.* (2022) 9:uhab046. doi: 10.1093/hr/ uhab046

- Aleksza D, Horváth G, Sándor G, Szabados L. Proline accumulation is regulated by transcription factors associated with phosphate starvation. *Plant Physiol.* (2017) 175:00791.2017. doi: 10.1104/pp.17.0 0791
- D'Este M, Alvarado-Morales M, Angelidaki I. Amino acids production focusing on fermentation technologies-a review. *Biotechnol Adv.* (2018) 36:14–25. doi: 10.1016/j.biotechadv.2017. 09.001
- Tzin V, Galili G. New insights into the shikimate and aromatic amino acids biosynthesis pathways in plants. *Mol Plant*. (2010) 3:956–72. doi: 10.1093/mp/ ssq048
- Hu H, Fei X, He B, Luo Y, Qi Y, Wei A. Integrated analysis of metabolome and transcriptome data for uncovering flavonoid components of *Zanthoxylum bungeanum* Maxim. Leaves under drought stress. *Front Nutr.* (2021) 8:801244. doi: 10.3389/fnut.2021.801244
- Wolfe RR. Branched-chain amino acids and muscle protein synthesis in humans: myth or reality? J Int Soc Sports Nutr. (2017) 14:30. doi: 10.1186/ s12970-017-0184-9

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