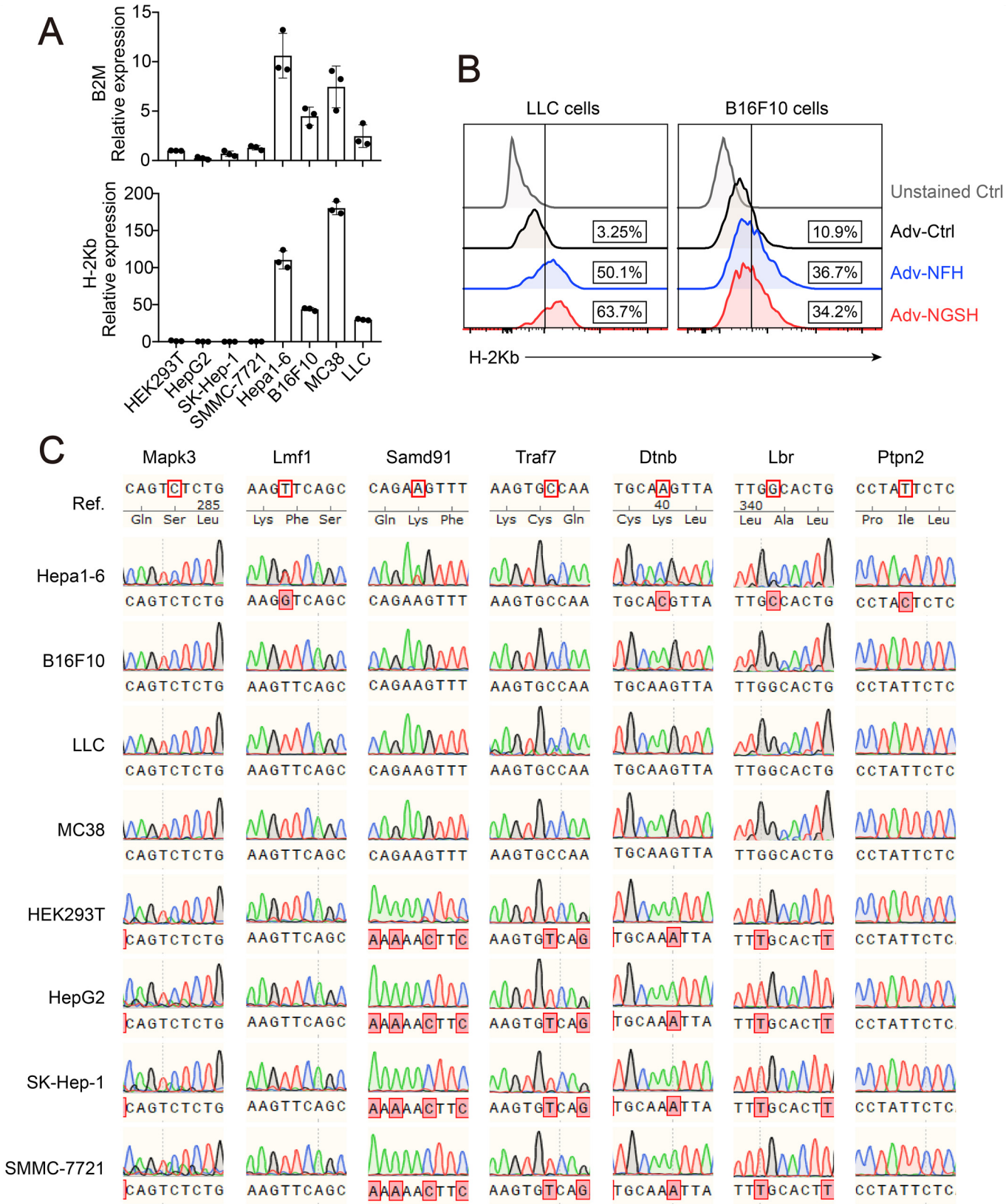


Expanded View Figures

Figure EV1. Expression of B2M, H-2Kb and the neoantigens in murine and human cell lines.

(A) With indicated primers, the relative expression of B2M and H-2Kb was quantitatively analyzed in HEK293T, HepG2, SK-Hep-1, SMMC-7721, Hepa1-6, B16F10, MC38, and LLC cells ($n = 3$). (B) LLC and B16F10 cells were treated with PBS, Adv-Ctrl (1 MOI), Adv-NFH (1 MOI), or Adv-NGSH (1 MOI) for 24 h. The cells were collected and stained by a H-2Kb antibody (Biolegend, 116505, clone AF6-88.5, 5 $\mu\text{g}/\text{ml}$) for 30 min before subjecting to flow cytometry analysis. (C) Sanger sequencing results demonstrated the existence of the neoantigens in indicated cell lines. Data are shown as mean \pm SD for 3 biological replicates.



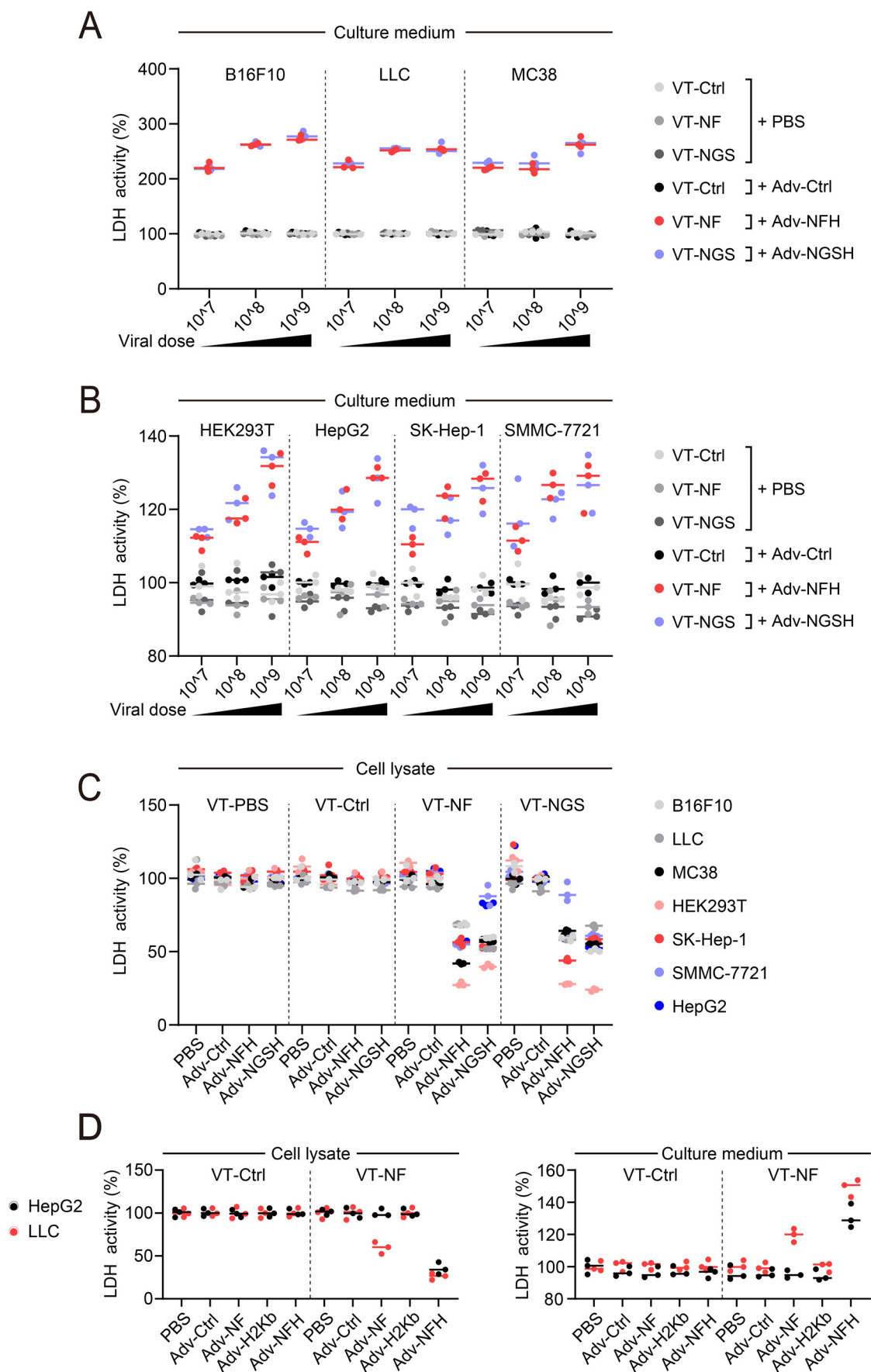
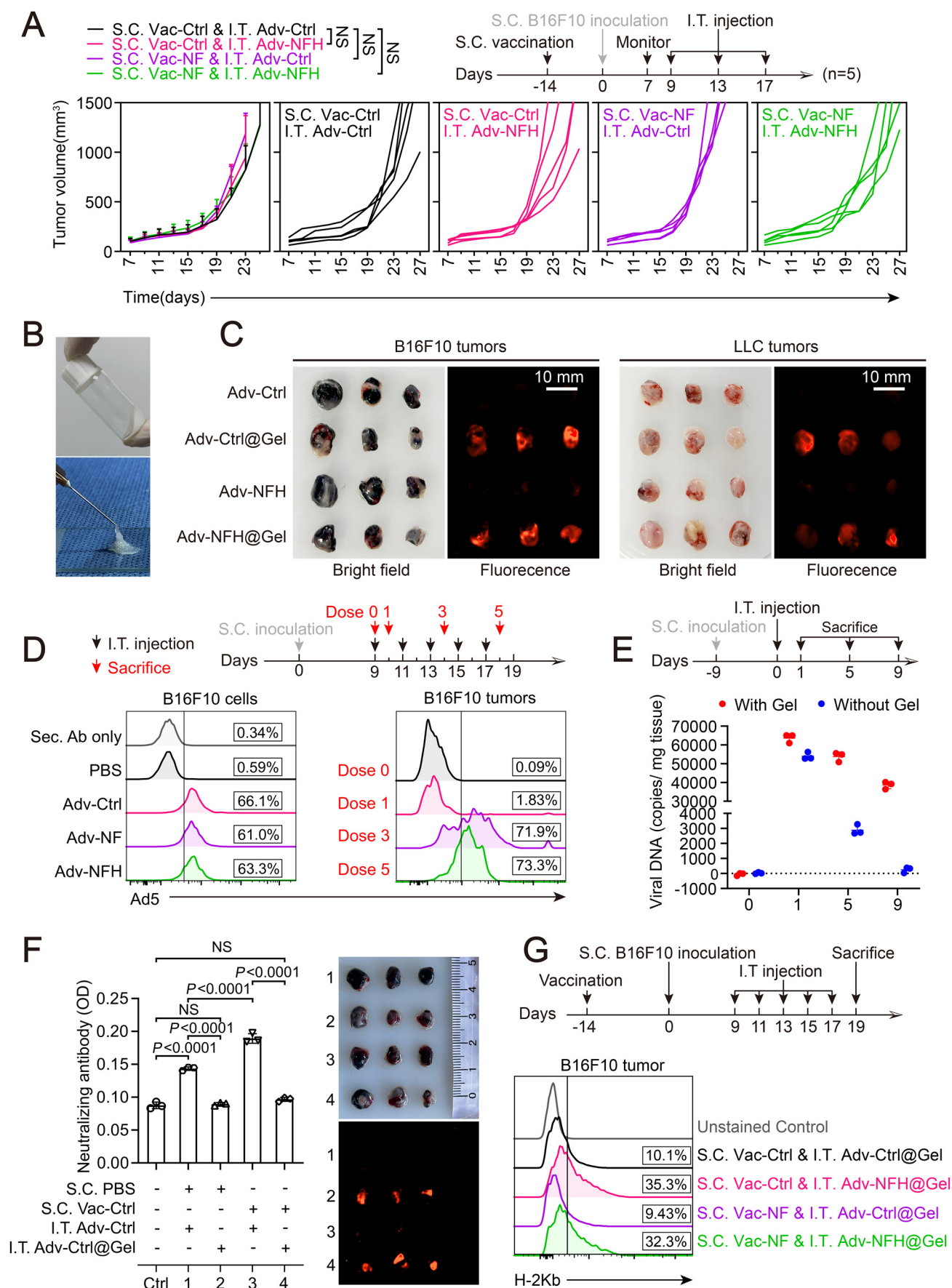


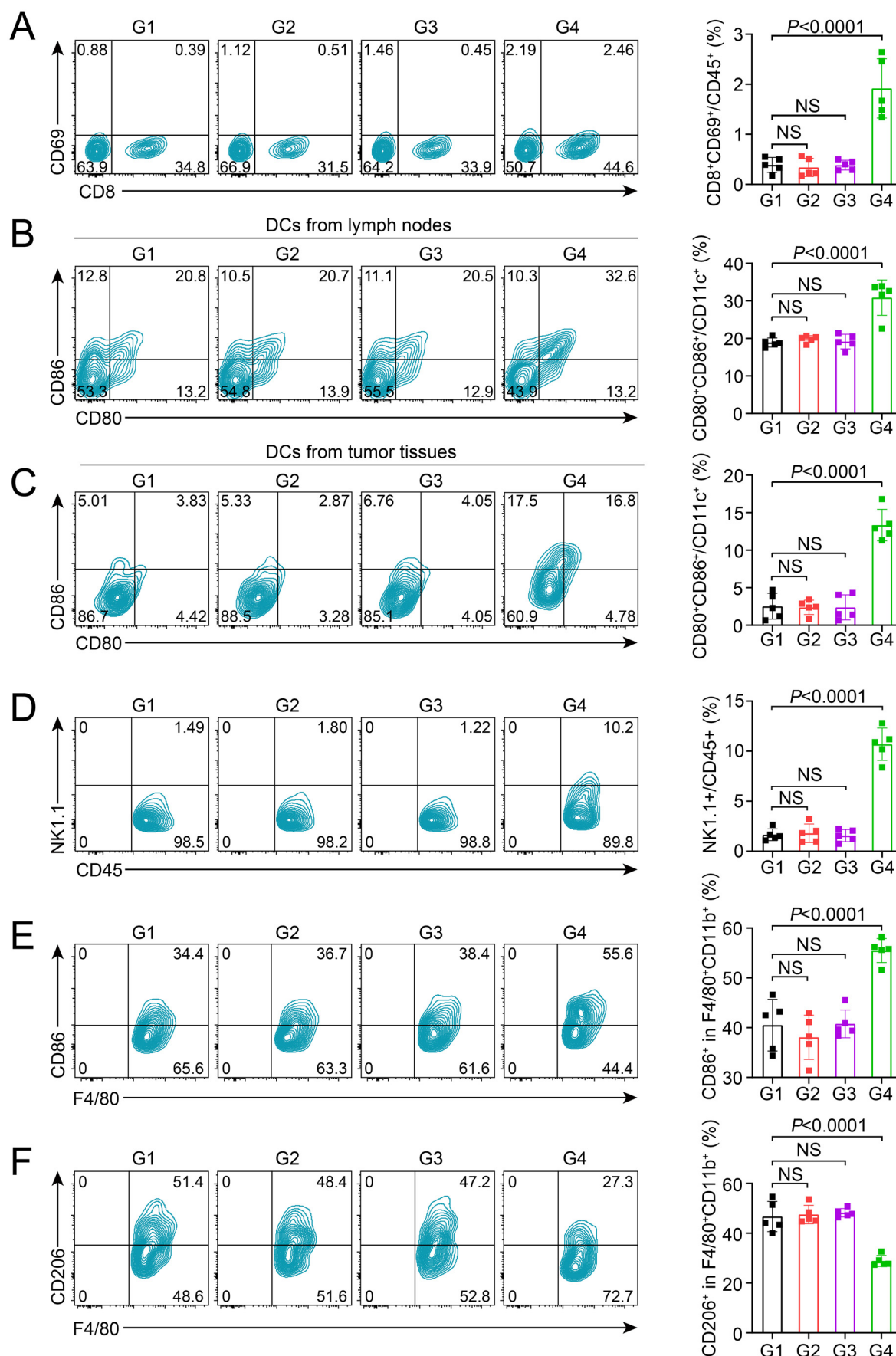
Figure EV2. Delivery of neoantigen peptide-MHC complexes redirected the cytotoxicity of neoantigen vaccine-primed T cells.

(A) LDH assay demonstrated the dose-dependent cytotoxicity of adenovirus-primed T cells in murine tumor cells that infected with Adv-NFH or Adv-NGSH. Samples were the culture medium of the murine cells assessed in Fig. 2E ($n = 3$). (B) Dose-dependent cytotoxicity of adenovirus-primed T cells in human tumor cells infected by Adv-NFH or Adv-NGSH. Samples were the culture medium of the human cells used in Fig. 2E ($n = 3$). (C) LDH assay in remnant tumor cells showed the specificity of the redirected cytotoxicity ($n = 3$). The assay was performed by following the experimental protocol in Fig. 2E. Mice were vaccinated with 2×10^9 VP of adenoviruses for 14 d before harvesting the SMNCs to prepare related T cells (VT-PBS, VT-Ctrl, VT-NF, and VT-NGS). Tumor cells, including B16F10, LLC, MC38, HEK293T, SK-Hep-1, SMMC-7721, and HepG2, were pre-treated with 1 MOI of indicated viruses and co-cultured with the prepared T cells at an E:T ratio of 1:3 for 48 h. (D) LDH assay demonstrated the functionality of MHC-I overexpression. After 48 h infection with 1 MOI of indicated viruses, HepG2 and LLC cells were co-cultured with VT-Ctrl or VT-NF at a ratio of 1:3 for 48 h before assessing the cell viability ($n = 3$). Data are shown as mean \pm SD for 3 biological replicates.



◀ **Figure EV3. Improving the efficacy of virotherapy with hydrogel encapsulation.**

(A) Overall and individual tumor growth curves demonstrated an integrative therapy of subcutaneous B16F10 tumor model with S.C. vaccination (Vac-Ctrl or Vac-NF, 2×10^9 VP) and I.T. virotherapy (Adv-Ctrl or Adv-NFH, 10^9 VP) ($n = 5$). (B) Representative images demonstrated the successful preparation of injectable silk hydrogel. (C) Hydrogel encapsulation potentiated the intratumoral infection of adenoviruses ($n = 3$). (D) Quantification of intratumoral infection rate of hydrogel encapsulated Adv-Ctrl at early and late time point with an anti-adenovirus antibody (Merck, AB1056, 1:500) and Alexa FluorTM 488 secondary antibody (Thermo Fisher, A-11055, 2 μ g/ml). Before the analysis, the functionality of the primary antibody was validated by staining B16F10 cells that have been infected with adenoviruses (Adv-Ctrl, Adv-NF, or Adv-NFH) in vitro at a MOI of 1 for 24 h (left panel). (E) qPCR assay quantified the viral DNA levels in the tumor tissues. Adv-Ctrl was prepared either in hydrogel (With Gel) or PBS (Without Gel) and injected into the tumors. Tumor tissues were subsequently collected at various time points for analysis. (F) Adenovirus neutralizing antibody assay and fluorescent imaging showed the protection of adenoviral vectors from host immunity by hydrogel encapsulation ($n = 3$). P value: Ctrl vs. 1, $P = 3.380 \times 10^{-7}$; Ctrl vs. 2, $P = 0.982$; Ctrl vs. 4, $P = 0.199$; 1 vs. 2, $P = 4.752 \times 10^{-7}$; 1 vs. 3, $P = 2.034 \times 10^{-6}$; 3 vs. 4, $P = 1.457 \times 10^{-9}$. (G) MHC status of B16F10-established tumors following the intratumoral injection of hydrogel encapsulated adenoviruses. Statistical analysis was performed using one-way ANOVA (F) and two-way ANOVA (A). Data are shown as mean \pm SD for 3 biological replicates.



◀ **Figure EV4. Integrative therapy remodeled the tumor microenvironment.**

Results of flow cytometry analysis showed the composition of CD69⁺CD8⁺ T cells in tumors (A), CD80⁺CD86⁺ DCs in lymph nodes (B), CD80⁺CD86⁺ DCs (C), NK1.1⁺ NK cells (D), CD86⁺ macrophages (E) and CD206⁺ macrophages (F) in tumors ($n = 5$). PE-Cyanine7-conjugated anti-NK1.1 (eBioscience, 25-5941-82, 2.5 $\mu\text{g}/\text{ml}$), FITC-conjugated anti-F4/80 (BioLegend, 123107, 2.5 $\mu\text{g}/\text{ml}$), APC-conjugated anti-CD11b (eBioscience, 17-0112-82, 1.25 $\mu\text{g}/\text{ml}$), and PE-conjugated anti-CD206 (eBioscience, 12-2061-82, 1.25 $\mu\text{g}/\text{ml}$) antibodies were used for analysis. *P* value (A): G2, $P = 0.995$; G3, $P = 0.999997$; G4, $P = 6.875 \times 10^{-6}$. *P* value (B): G2, $P = 0.946$; G3, $P = 0.999$; G4, $P = 1.359 \times 10^{-5}$. *P* value (C): G2, $P = 0.999$; G3, $P = 0.999$; G4, $P = 1.063 \times 10^{-7}$. *P* value (D): G2, $P = 0.996$; G3, $P = 0.999$; G4, $P = 1.202 \times 10^{-9}$. *P* value (E): G2, $P = 0.755$; G3, $P = 0.9996$; G4, $P = 8.186 \times 10^{-5}$. *P* value (F): G2, $P = 0.987$; G3, $P = 0.924$; G4, $P = 8.309 \times 10^{-6}$. Statistical analysis was performed using one-way ANOVA (A–F). Data are shown as mean \pm SD for 3 biological replicates.

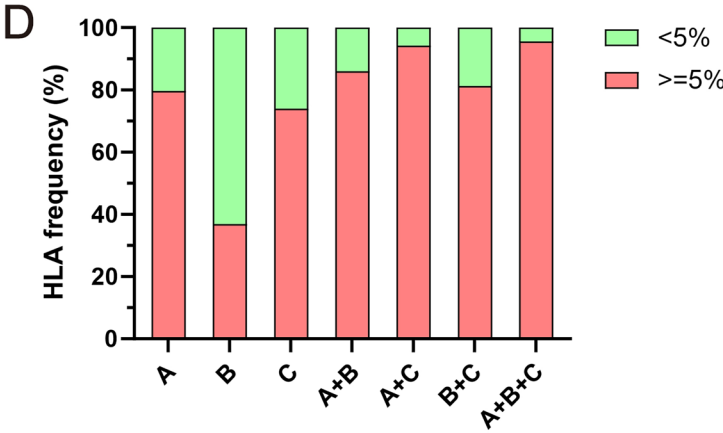
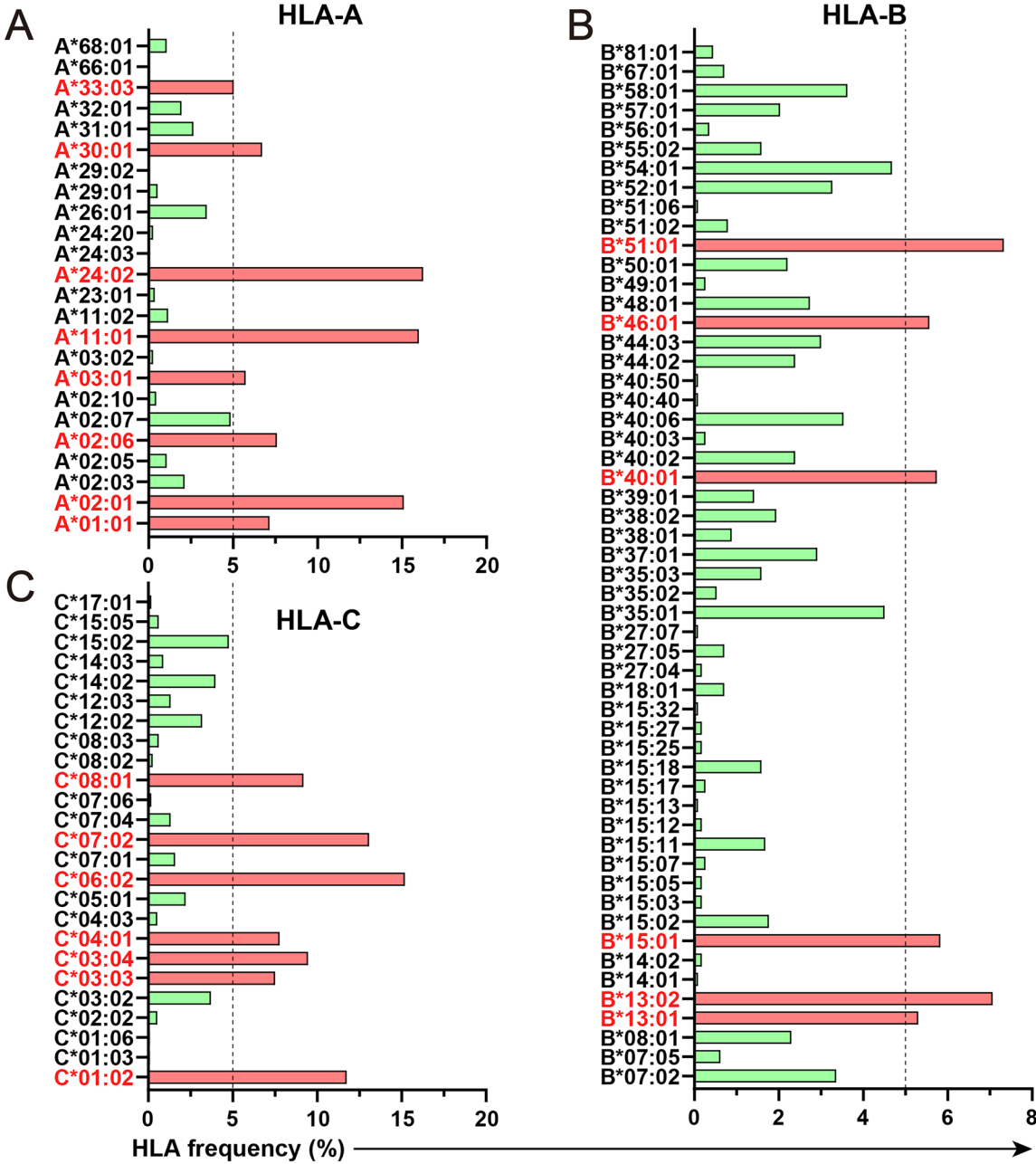


Figure EV5. Coverage of HLA molecules indicates the applicability of the integrative therapy in human.

Allele frequencies of HLA-A (A), HLA-B (B), and HLA-C (C) in the 741 individuals from Han population residing in Shanxi Province of China. The data was downloaded from PGG.MHC website: <https://pog.fudan.edu.cn/pggmhc/#/population>, and all the alleles were plotted. The 5% cut-off value was indicated by the dotted line. Alleles with frequencies <5% and ≥5% were, respectively, indicated by green and red columns. The coverage of alleles with frequencies ≥5% was calculated and demonstrated as cumulative histogram (D), in which HLA-A, HLA-B, and HLA-C were, respectively, abbreviated as A, B, and C. The cumulative results (≥5%) were as follows: A (79.59%), B (36.84%), C (73.94%), A + B (85.94%), A + C (94.17%), B + C (81.27%), A + B + C (95.58%).