

Validation of a monoclonal antibody-based immunofluorescent assay to detect *Burkholderia pseudomallei* in blood cultures

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Background: Identification of *Burkholderia pseudomallei*, the cause of melioidosis, using routine methods takes several days. Use of a monoclonal antibody-based immunofluorescent assay (IFA) on positive blood cultures may speed diagnosis.

Methods: We tested the diagnostic accuracy of the IFA on 545 blood cultures positive for Gram-negative organisms at Udon Thani Hospital, Thailand, between June 2015 and August 2016.

Results: Sensitivity of the IFA was 100% and specificity was 99.6%. The median decrease in time to pathogen identification between the IFA result and routine methods was 28 h (IQR 25–51), p<0.0001.

Conclusions: The IFA accurately expedites the diagnosis of melioidosis.

Keywords: Bacteremia, Burkholderia pseudomallei, Diagnosis, Immunofluorescent assay, Melioidosis, Thailand

Introduction

Melioidosis is caused by infection with the Gram-negative bacillus *Burkholderia pseudomallei*. Northeast Thailand is a hyperendemic zone for melioidosis due to the widespread presence of *B. pseudomallei* in the soil and water in this region.^{1,2} The clinical presentation of melioidosis is difficult to distinguish from other aetiologies of blood stream infection, pneumonia and sepsis. Due to intrinsic antibiotic resistance of *B. pseudomallei*, treatment of melioidosis requires the third generation cephalosporin ceftazidime or a carbapenem for at least 2 weeks followed by up to 20 weeks of trimethoprim-sulfamethoxazole. Mortality rates from melioidosis in Northeast Thailand average 40%.³ Rapid diagnosis is therefore essential. However, definitive diagnosis of melioidosis requires culture and subsequent identification of the organism which may take several days.

Previous studies have evaluated diagnostic strategies to accelerate the identification of *B. pseudomallei* and to ensure initiation of appropriate antibiotic treatment. One such strategy is the use of a monoclonal antibody-based immunofluorescent assay (IFA) to test blood cultures that flag positive. The IFA monoclonal antibody (mAb) is 4B11, which is specific for an exopolysaccharide of *B. pseudomallei.*^{4,5} This assay has been successfully used at Sappasithiprasong Hospital, Ubon Ratchathani, Thailand.⁵ We report here on the successful implementation of the IFA assay at another referral hospital in a hyperendemic melioidosis zone, Udon Thani Hospital, Udon Thani, Thailand.

Materials and methods

We implemented the IFA on blood culture specimens collected by hospital staff from June 2015 to August 2016. This was done as part of a large prospective multi-center study of melioidosis. Routine practice is to take 10 ml of blood for adults or 5 ml of blood for children and inoculate this into a 30 ml bottle (Aerobic BacT/Alert FA Plus; Biomérieux, Marcy-l' Étoile, France). The bottle is incubated in an automated BacT/ALERT 3D instrument at 37°C. Once the instrument flags a bottle as positive, it is removed for Gram stain and sub-culture. Bacterial

© The Author 2017. Published by Oxford University Press on behalf of Royal Society of Tropical Medicine and Hygiene. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/ by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited. isolates are subsequently identified using standard laboratory methodology derived from the Work Manual for Laboratory Identification of Bacteria and Fungi for Regional and General Hospitals, Department of Medical Science, Ministry of Public Health, Thailand. The IFA was performed once daily on 1 ml of fluid taken from blood culture bottles identified as positive by the BacT/ALERT 3D instrument and determined to have Gram-negative bacteria by Gram stain. In a Class II biosafety cabinet, $10 \mu l$ of working mAb-IFA reagent containing $5 \mu g/m l$ of mAb 4B11 and 20µg/ml of Alexa Fluor 488 conjugatedgoat anti-mouse IgG (Molecular Probes, Carlsbad, CA, USA) in phosphate-buffered saline (PBS).^{4,5} were mixed with an equal volume of blood culture fluid on a glass slide, a coverslip applied, and the slide left at room temperature for 10 min. Bacteria were observed using a fluorescent microscope (Olympus, Tokyo, Japan) at 1000x magnification.

Results

A total of 553 sequential blood cultures were positive with Gramnegative bacilli during the study period. IFA was performed on 545 specimens. The IFA results for these specimens were compared to standard techniques used to identify *B. pseudomallei* (Table 1). The sensitivity of IFA was 100%, specificity 99.6%, positive predictive value 97.4%, and negative predictive value 100%. The two discordant specimens that were positive by IFA but negative for *B. pseudomallei* by routine methods were identified as *Escherichia coli*. One of these *E. coli* isolates cultured on agar was available for subsequent testing with IFA and the result was negative.

For 533 specimens with reliable timestamp data available, the median time from collection of the blood culture specimen to identification of *B. pseudomallei* using IFA was 41 h (IQR 24–47). The median time from collection of the blood culture specimen to identification of *B. pseudomallei* using standard techniques was 72 h (IQR 65–94). The median decrease in time between the IFA result and standard technique result was 28 h (IQR 25–51), p<0.0001.

Discussion

Our results confirm the excellent test characteristics of the monoclonal antibody IFA for *B. pseudomallei* identification in positive

Table 1. Diagnostic accuracy of an immunofluorescent assay for

 Burkholderia pseudomallei

IFA	Routine identification methods		Total
_	Positive	Negative	
Positive	74	2	76
Negative	0	469	469
Total	74	471	545

IFA: immunofluorescent assay.

blood cultures in a melioidosis-endemic area. These characteristics are comparable to the original report of this test elsewhere in Thailand,⁵ confirming validity of the assay. Notably, in the previous study the IFA was performed on all positive blood cultures, whereas in this study we restricted testing to positive blood cultures with Gram-negative organisms. The reason for false positive results for *E. coli* isolates does not appear to be due to cross-reactivity of the monoclonal antibody to *E. coli* but may be due to other unknown factors including operator error.

We also provide precise quantitation of the decreased time to bacterial identification with the IFA. We show that the diagnosis of melioidosis can be made at least one day earlier than with standard diagnostic methods, a statistically and clinically significant decrease. In light of the high mortality rate of hospitalized patients with melioidosis in Thailand, this result is highly informative for clinicians and facilitates administration of appropriate therapy for *B. pseudomallei*.

A limitation to this study is that one trained technician as part of a research protocol performed the IFA. The diagnostic accuracy of the test and timeliness of the results may be lower when implemented in a busy clinical microbiology laboratory. However, the IFA can typically be performed in less than 30 min. Chemical inactivation of the bacteria prior to IFA testing is presently being investigated to increase safety for laboratory staff.

Conclusions

We show excellent diagnostic accuracy of an IFA for detection of *B. pseudomallei* in blood cultures at a Thai referral hospital, and demonstrate a reduced time to diagnosis. These data provide important confirmation of validity of the IFA and support the widespread use of the test for prompt diagnosis of melioidosis in areas where the infection is endemic.

Authors' contributions: AD performed IFA. SP prepared IFA reagent. AS and YJ performed routine bacterial identification at Udon Thani Hospital. AD, PE and RP collected data. TEW performed statistical analysis. NC and TEW supervised the study and wrote the manuscript. All authors read and approved the final manuscript. NC and TEW are guarantors of the paper.

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Competing interests: None declared.

Ethical approval: The study was approved by the Ethics Committee of Faculty of Tropical Medicine, Mahidol University (approval number MUTM 2015-002-01) and Ethics Committee of Udon Thani Hospital (approval number 0032.102/318).

References

- 1 Chantratita N, Wuthiekanun V, Limmathurotsakul D et al. Genetic diversity and microevolution of *Burkholderia pseudomallei* in the environment. PLoS Negl Trop Dis 2008;2:e182. doi:10.1371/journal. pntd.0000182.
- 2 Wuthiekanun V, Limmathurotsakul D, Chantratita N et al. Burkholderia pseudomallei is genetically diverse in agricultural land in Northeast Thailand. PLoS Negl Trop Dis 2009;3:e496. doi:10.1371/ journal.pntd.0000496.
- 3 Wiersinga WJ, Currie BJ, Peacock SJ. Melioidosis. N Engl J Med 2012; 367:1035-44. doi:10.1056/NEJMra1204699.
- 4 Tandhavanant S, Wongsuvan G, Wuthiekanun V et al. Monoclonal antibody-based immunofluorescence microscopy for the rapid identification of *Burkholderia pseudomallei* in clinical specimens. Am J Trop Med Hyg 2013;89:165–8. doi:10.4269/ajtmh.13-0066.
- 5 Chantratita N, Tandhavanant S, Wongsuvan G et al. Rapid detection of Burkholderia pseudomallei in blood cultures using a monoclonal antibody-based immunofluorescent assay. Am J Trop Med Hyg 2013; 89:971-2. doi:10.4269/ajtmh.13-0212.