



Article

Prevalence of Alpha(α)-Thalassemia in Southeast Asia (2010–2020): A Meta-Analysis Involving 83,674 Subjects

Lucky Poh Wah Goh , Eric Tzyy Jiann Chong and Ping-Chin Lee *

Biotechnology Programme, Faculty of Science and Natural Resources, Universiti Malaysia Sabah, Kota Kinabalu 88400, Sabah, Malaysia; luckygoh@hotmail.com (L.P.W.G.); eric_ctj@live.com (E.T.J.C.)

* Correspondence: leepc@ums.edu.my

Received: 7 August 2020; Accepted: 24 September 2020; Published: 9 October 2020



Abstract: Alpha(α)-thalassemia is a blood disorder caused by many types of inheritable α -globin gene mutations which causes no-to-severe clinical symptoms, such as Hb Bart's hydrops fetalis that leads to early foetal death. Therefore, the aim of this meta-analysis was to provide an update from year 2010 to 2020 on the prevalence of α -thalassemia in Southeast Asia. A systematic literature search was performed using PubMed and SCOPUS databases for related studies published from 2010 to 2020, based on specified inclusion and exclusion criteria. Heterogeneity of included studies was examined with the I² index and Q-test. Funnel plots and Egger's tests were performed in order to determine publication bias in this meta-analysis. Twenty-nine studies with 83,674 subjects were included and pooled prevalence rates in this meta-analysis were calculated using random effect models based on high observed heterogeneity (I² > 99.5, *p*-value < 0.1). Overall, the prevalence of α -thalassemia is 22.6%. The highest α -thalassemia prevalence was observed in Vietnam (51.5%) followed by Cambodia (39.5%), Laos (26.8%), Thailand (20.1%), and Malaysia (17.3%). No publication bias was detected. Conclusions: This meta-analysis suggested that a high prevalence of α -thalassemia occurred in selected Southeast Asia countries. This meta-analysis data are useful for designing thalassemia screening programs and improve the disease management.

Keywords: prevalence; α -thalassemia; Southeast Asia; meta-analysis; haematological disorder

1. Introduction

Thalassemia is the most common hereditary red blood cell disorder which causes anemias due to defective genes that code for globin proteins synthesis [1]. The inheritance of the thalassemia genotype could result in the individual being either a carrier or a patient. There are two major types of thalassemia: (1) alpha (α) and (2) beta (β), in which the former is the most common form of thalassemia worldwide especially in Southeast Asia populations [2,3]. Both α - and β -thalassemia arise from genetic defects in α and/or β -globin genes, which regulate the number of globin chains in red blood cells. Genetic defects in either α and/or β -globin genes can cause imbalance in numbers of α and β chains in red blood cells. This leads to the manifestation of clinical conditions known as α - or β -thalassemia. The most common genetic defect in α -thalassemia is a deletion in the α -globin gene involving one or both globin genes such as $-\alpha^{3.7}$, $-\alpha^{4.2}$, $-\text{SEA}$, $-\text{THAI}$, $-\alpha^{\text{CD59}}$, $-\alpha^{20.5}$, $-\alpha^{\text{IVS I-1}}$ and others (Figure 1) [4–6]. A highly severe form of deletion α -thalassemia, known as Haemoglobin (Hb) Bart's hydrops fetalis, is a homozygous α^0 -thalassemia deletion with a complete loss of functional α -globin that leads to foetal death or death shortly after birth. Currently, there is no effective treatment for this disease [7,8].

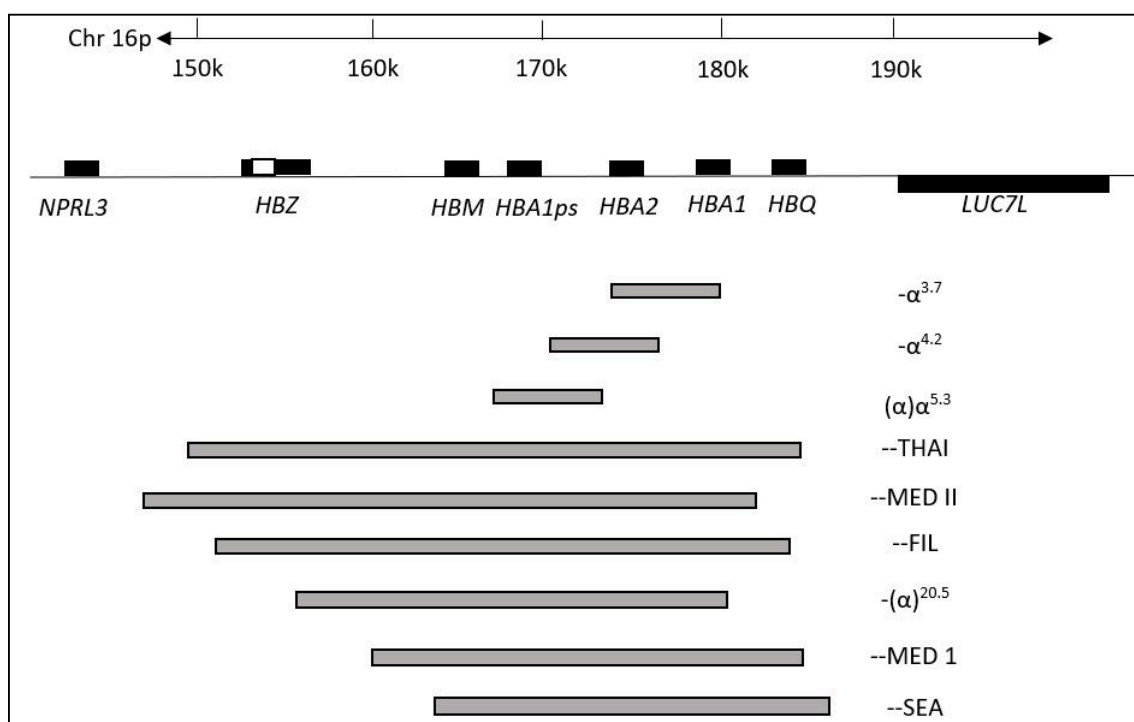


Figure 1. Some common changes in α -thalassemia in Southeast Asia. The genes are shown in boxes with a scale in kilobases (kb). The most common deletions of α -thalassemia mutations are indicated by grey bars indicating the length of deletion. (Adapted from Farashi & Hartevelde, 2018 [6]).

There are also cases of non-deletional mutations in the α -globin gene such as Hb Constant Spring (CS), which is caused by a nucleotide substitution in the termination codon TAA \rightarrow CAA and also Hb Pakse ($-\alpha^{4PS}$), which is caused by the termination codon (UAA \rightarrow CAA). This results in the elongation of the α -globin chain protein and causes severe anemia with serious complications that include liver impairment, cardiac disease and endocrine disorder [9,10]. Some cases of Hb CS require red blood cell transfusions when Hb drop to dangerously low levels [11]. Most importantly, an individual can carry a single or multiple type form of the mutation (deletional/non-deletional), which give rise to different clinical manifestations and complicates diagnosis as well as treatments for α -thalassemia [12]. Both deletional and non-deletional α -thalassemia prevalence rates are highly important in determining the overall severity of clinical symptoms in a region or a specific population.

Despite the detrimental effects of thalassemia, evidence shows that thalassemia confers a protective effect against hyperparasitemia due to malaria infection [13]. *Plasmodium vivax* parasitemia were two to three times lower in thalassemia patients as compared to malaria cases in people without thalassemia. However, the protective effect of thalassemia against parasitemia was not observed in a study conducted in Papua New Guinea in children aged 3–21 months [14]. Therefore, there is a natural selection force which leads to the prevalence of thalassemia cases in malaria-endemic areas [15].

Most of the α -thalassemia cases reveal some abnormalities in their red cell index. According to the British Committee for Standards in Haematology, a value of <27 pg in the average amount of Hb found in red blood cells (also referred as Mean Corpuscular Hemoglobin) is the primary screening threshold to quantify Hb subtypes [16]. However, subjects with a single gene deletion or carriers of the mutation in the non-severe form of α -thalassemia may present a normal Hb level. Different heterozygosity or homozygosity of gene deletions or mutations in the α -thalassemia gene gives different phenotypes, which complicates treatment [12]. Therefore, diagnosing of non-severe forms of α -thalassemia is a highly challenging task.

Several countries in the Southeast Asia region have reported the prevalence of α -thalassemia in different ethnic groups independently, revealing that the prevalence of α -thalassemia differs from

country to country with different ethnicities [4,5,17,18]. However, no studies have systematically meta-analyzed the prevalence and epidemiology of α -thalassemia—where the results of these similar studies are quantitatively combined—for this region. Therefore, the aim of this meta-analysis was to provide an update (from 2010 until 2020) using data concerning α -thalassemia prevalence in Southeast Asia, focusing on Cambodia, Laos, Malaysia, Thailand, and Vietnam. The study outcome on the perspective of α -thalassemia prevalence in Southeast Asia could aid in designing healthcare policies for α -thalassemia screening in large populations and provide better genetic counselling programs.

2. Materials and Methods

2.1. Study Guidelines and Literature Search

PRISMA guidelines were followed for conducting and reporting the results of this meta-analysis (Table S1) [19]. PubMed (Table S2) and SCOPUS (Table S3) databases were searched up to March 2020 with the lower limit set to January 2010 using related terms, including alpha, thalassemia, southeast, and Asian.

2.2. Selection of Studies and Data Extraction

All search results were screened by two investigators, and all potential studies were independently reviewed to be included in this meta-analysis. The main inclusion criteria were: (1) studies published in English in which the prevalence of α -thalassemia (including all deletional and non-deletional mutations) in Southeast Asian countries were reported; and (2) the study was a peer-reviewed publication. Studies that did not report cross-sectional, observational, cohort, or prevalence of α -thalassemia were excluded. We identified additional eligible studies based on references that were cited in the relevant articles. When publications overlapped, only the study with the largest or the most recent data was included in this meta-analysis. Data, including first author's name, publication year, country, sample size, and prevalence of α -thalassemia of the included studies, were extracted and documented by the reviewing investigators. A total number of 278 articles were included in this meta-analysis (Tables S4 and S5). The study selection and review process are illustrated in Figure 2.

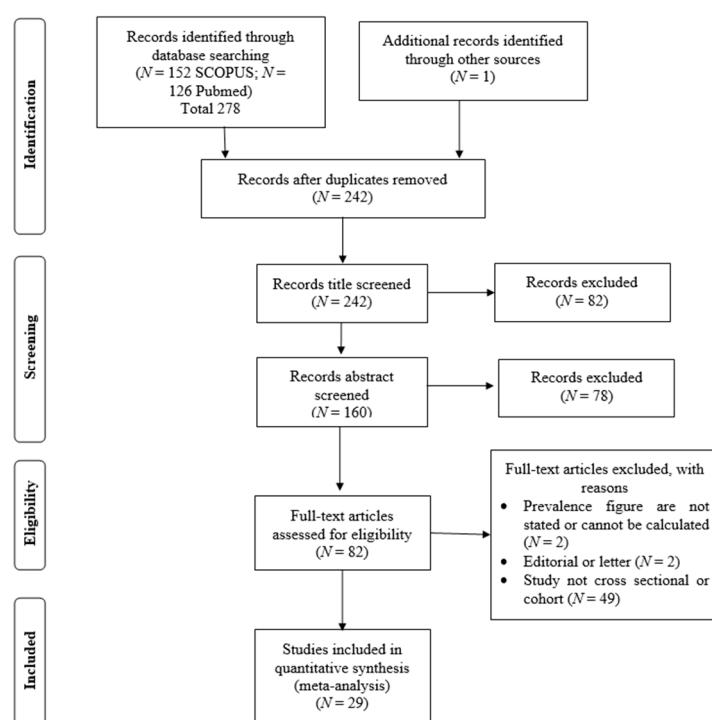


Figure 2. Flow diagram of the systemic literature search in this study.

2.3. Statistical Analyses

The prevalence of α -thalassemia was calculated for each study with the number of reported α -thalassemia cases as the numerator and the total sample size as the denominator. Homogeneity across studies was investigated with the I^2 index (represented as percentage) and Q-test (represented as a p -value) that indicated heterogeneity between studies. It was reported that an I^2 value $> 75\%$ and Q-test with a p -value < 0.1 was regarded as high heterogeneity [20,21]. A random effects model was used to combine individual effect sizes to create pooled α -thalassemia prevalence if a significantly high heterogeneity was observed. If other results were obtained, a fixed effects model was utilized. A forest plot was generated to illustrate the prevalence of each study with a 95% confidence interval (95% CI) that contributed to the analysis along with the combined prevalence rate. A subsequent meta-analysis was also performed based on each respective country. Funnel plots and Egger's tests of asymmetry were performed to identify any bias within the results [22,23]. All analyses were performed with Comprehensive Meta-Analysis version 2 software (Biostat, Inc., New Jersey, USA) [24].

3. Results

3.1. Study Characteristics

Twenty-nine studies with a total number of 83,674 subjects were included in this meta-analysis after a detailed assessment of records obtained from the database and additional searching. These studies were published between January 2010 and October 2019. Among all included studies, two were carried out in Cambodia, three in Laos, five in Malaysia, 20 in Thailand, and two in Vietnam (Table 1). The main characteristics of the studies included in the meta-analysis were recorded and are shown in Table 1.

Table 1. Characteristics of studies included in the meta-analysis.

Author [Reference]	α -Thalassemia Genotyping Method	Genotypes Found in the Study	Country	Specific Ethnic ¹	Events ²	Total ³
Munkongdee et al., 2016 [25]	Polymerase chain reaction (PCR)	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, α^{CS} , α^{Ps}	Cambodia	N/A	646	1631
Jomoui et al., 2017 [26]	PCR	$_{-SEA}$	Cambodia	N/A	7	21
Wongprachum et al., 2012 [27]	PCR	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, $_{-THAI}$, α^{CS} , α^{Ps} , $\alpha^{Q-Thailand}$	Laos	N/A	130	411
Jomoui et al., 2017 [26]	PCR	$_{-SEA}$	Laos	N/A	28	52
Tritipsombut et al., 2012 [28]	PCR	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, α^{CS} , α^{Ps}	Laos	N/A	30	349
Azma et al., 2012 [29]	PCR		Malaysia	N/A	14	400
Azma et al., 2014 [4]	PCR	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, α^{CS} , α^{CD59} , α^{IVS1-1}	Malaysia	Malay, Chinese, Indian, Other	736	1623
Jameela et al., 2011 [30]	PCR	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, $_{-FIL}$, α^{125}	Malaysia	Malay, Chinese, Indian, Sikh, Iban	10	310
Mohd Yatim et al., 2014 [31]	PCR	$-\alpha^{3.7}$, $_{-SEA}$, α^{CS} , α^{CD59}	Malaysia	Malay	28	68
Tan et al., 2010 [3]	PCR	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, $_{-THAI}$, $_{-FIL}$, α^{CS} , α^{125}	Malaysia	Kadazandusun	42	125
Charoenkwan et al., 2010 [32]	PCR	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, $_{-Q-Thailand}$, α^{CS}	Thailand	N/A	142	566
Lithanatudom et al., 2016 [17]	PCR	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, $_{-THAI}$, α^{CS} , α^{Ps}	Thailand	Yong, Yuan, Lue, Khuen, Blang, Mon, Paluang, Lawa	33	141
Nillakupt et al., 2012 [33]	PCR	$-\alpha^{3.7}$, $_{-SEA}$, α^{CS} , α^{Ps}	Thailand	N/A	47	266
Pongjantharasatien et al., 2016 [34]	PCR	$_{-SEA}$, $_{-THAI}$, $_{-FIL}$, $_{-\alpha^{thal-1}}$	Thailand	N/A	4555	31,632
Pichanun et al., 2010 [35]	PCR	$-\alpha^{3.7}$, α^{CS} , α^{Ps}	Thailand	N/A	36	587
Pharephan et al., 2016 [36]	PCR	$-\alpha^{3.7}$, $-\alpha^{4.2}$, $_{-SEA}$, α^{CS}	Thailand	N/A	229	638

Table 1. Cont.

Author [Reference]	α -Thalassemia Genotyping Method	Genotypes Found in the Study	Country	Specific Ethnic ¹	Events ²	Total ³
Panyasai et al., 2016 [37]	PCR	$-\alpha^{3,7}, -\alpha^{QT}, -SEA,$	Thailand	N/A	51	23,914
Panomai et al., 2010 [38]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, -THAI, \alpha^{CS}, \alpha^{Ps}$	Thailand	N/A	40	190
Prayalaw et al., 2014 [39]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, \alpha^{CS}, -\alpha^{Q-Thailand}$	Thailand	N/A	75	300
Seeratanachot et al., 2015 [40]	Realtime-PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA$	Thailand	N/A	62	250
Wisedpanichkij et al., 2015 [41]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, \alpha^{CS}$	Thailand	N/A	409	578
Uaprasert et al., 2013 [42]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, \alpha^{CS}$	Thailand	N/A	67	241
Srivorakun et al., 2011 [43]	PCR	$-\alpha^{3,7}, -SEA, \alpha^{CS}$	Thailand	N/A	44	226
Tritipsombut et al., 2012 [28]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, \alpha^{CS}, \alpha^{Ps}$	Thailand	N/A	85	1460
Chaibunruang et al., 2013 [44]	PCR	$-SEA, -THAI$	Thailand	N/A	1874	12,525
Kulaphisit et al., 2017 [5]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, -THAI, \alpha^{CS}, \alpha^{Ps}$	Thailand	Yong, Lue, Yuan, Shan, Khuen, Htin, Paluang, Blang, Lawa, Mon, Skaw Karen, Pwo Karen, Padong Karen	124	668
Thanyaornwanya et al., 2019 [45]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, \alpha^{CS}, \alpha^{Ps}$	Thailand	N/A	676	1192
Jomoui et al., 2017 [26]	PCR	$-SEA$	Thailand	N/A	66	96
Mankhenthong et al., 2019 [46]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, -THAI, \alpha^{CS}$	Thailand	N/A	118	1290
Pata et al., 2019 [47]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, -THAI, \alpha^{CS}$	Thailand	N/A	82	195
O'Riordan et al., 2010 [18]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, -THAI, -FIL, \alpha^{CS}$	Vietnam	Kinh, Dao, Tay, Nung, S'Tieng, M'Nong, Rac lay, E De	996	1431
Hoa Nguyen et al., 2014 [48]	PCR	$-\alpha^{3,7}, -\alpha^{4,2}, -SEA, -THAI, -SEA, \alpha^{CS}, \alpha^{Ps}$	Vietnam	Có-Tu	98	298
Total					11,580	83,674

¹ N/A: Not available due to ethnicities were not reported by the study. ² Events: Number of subjects carrying alpha-thalassemia. ³ Total: Total number of subjects.

3.2. Meta-Analysis Outcomes

The meta-analysis was conducted using a random effects model found significant heterogeneity showed an $I^2 > 99.5\%$ and p -value < 0.001 in overall and all subgroups except Cambodia (Table 2). The forest plot showed that the overall prevalence rate of α -thalassemia occurrence in this meta-analysis was 0.226 (95% CI = 0.168–0.296; $I^2 = 99.5\%$; p -value < 0.1) (Figure 3). In the subgroup analysis based on country, Vietnam had the highest prevalence rate (51.5%) of α -thalassemia followed by Cambodia (39.5%) Laos (26.8%), Thailand (20.1%), and Malaysia (17.3%) (Figure 4).

Table 2. Prevalence rate and heterogeneity of α -thalassemia in overall and subgroups of the study.

Heterogeneity		Prevalence Rate (95% CI)	Sample Size (N)	No. of Studies (N)	Subgroups
I^2 (%)	p -Value				
99.53	<0.001	0.226 (0.168–0.296)	83,674	32	Overall
0	0.560	0.395 (0.372–0.419)	1652	2	Cambodia
97.26	<0.001	0.268 (0.096–0.559)	812	3	Laos
98.20	<0.001	0.173 (0.060–0.407)	2526	5	Malaysia
99.47	<0.001	0.201 (0.143–0.273)	76,955	20	Thailand
99.22	<0.001	0.515 (0.190–0.828)	1729	2	Vietnam

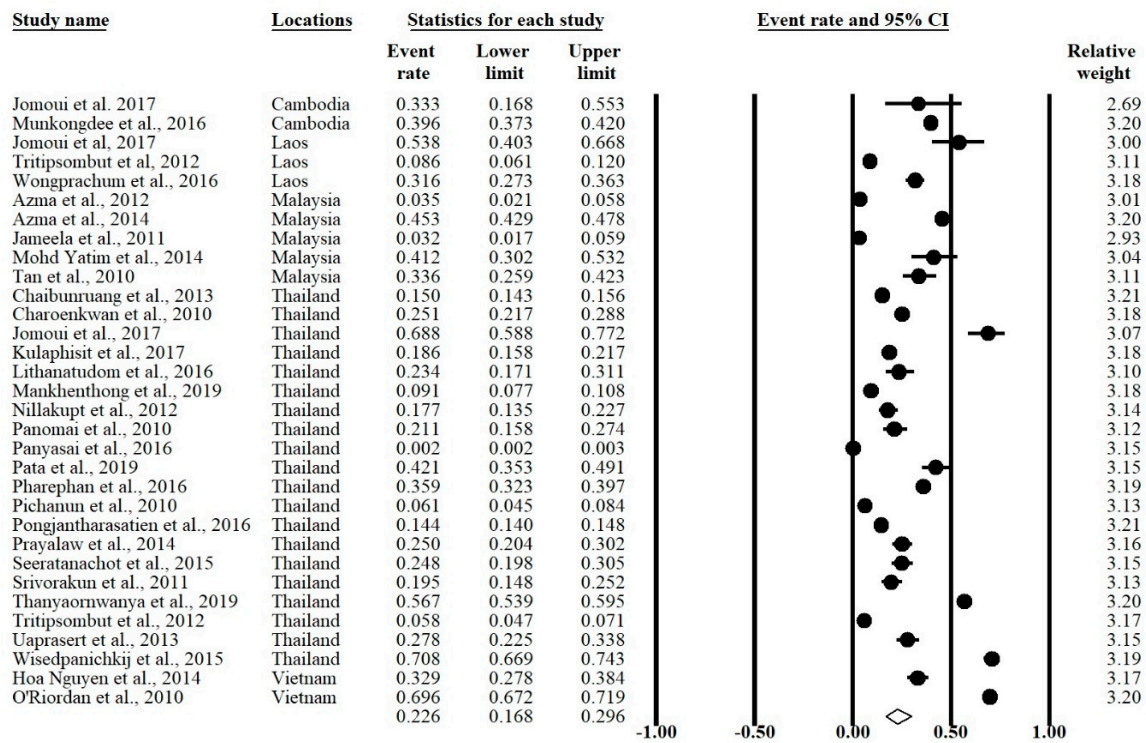


Figure 3. Forest plot of α -thalassemia overall prevalence using random effects model.

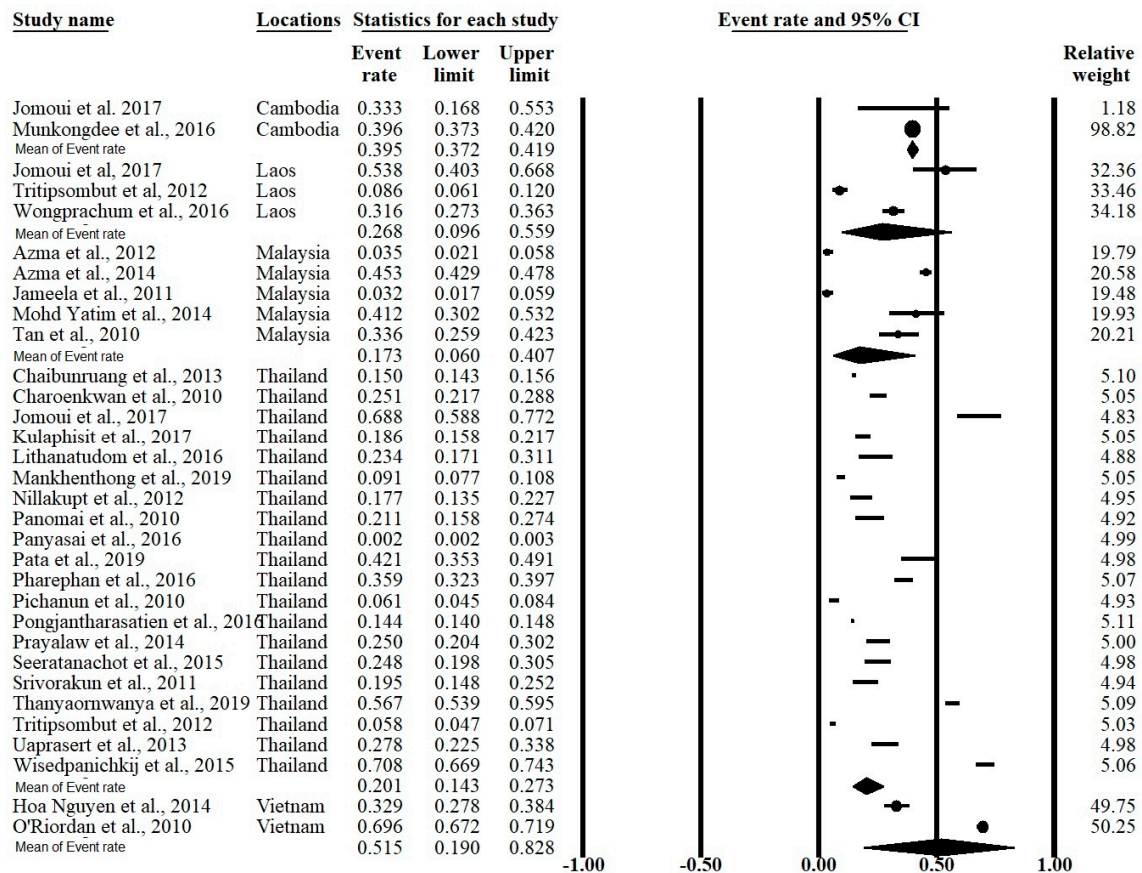


Figure 4. Forest plot of the α -thalassemia prevalence grouped according to country.

3.3. Publication Bias

Funnel plots and Egger's tests were performed to estimate the publication bias of the included literature. The shape of the funnel plot revealed obvious evidence of symmetry (Figure 5). The value for Egger's test was t -value = 1.24 with a p -value = 0.112.

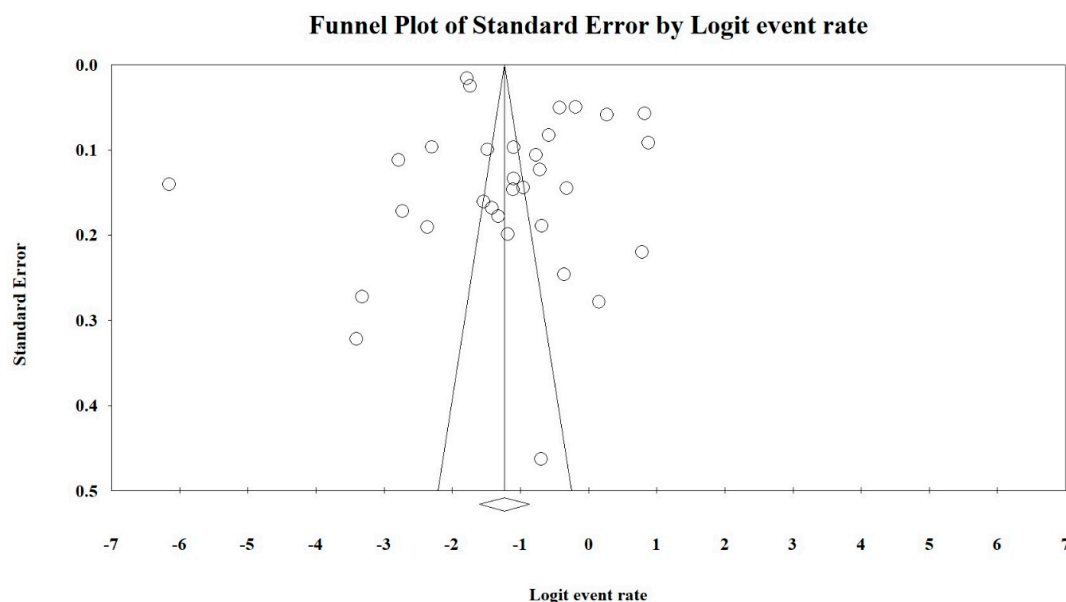


Figure 5. Funnel plot of the overall prevalence of α -thalassemia in this study.

4. Discussion

This study is the first to report the prevalence of α -thalassemia in the Southeast Asia region over the past 10 years (2010–2020). We did not obtain any α -thalassemia-related studies that fulfilled our inclusion and exclusion criteria from other Southeast Asian countries, including Brunei, Indonesia, Myanmar, Philippines, Singapore, and Timor-Leste. Using a random effects model, the overall prevalence rate of α -thalassemia in the included countries was 22.6%, which indicated a significant reduction of ~50% of the prevalence in the Southeast Asia region since 2008. The World Health Organization had reported the α -thalassemia prevalence as 44.6% in 2008 [49]. India and Brazil, were reported at about 12% and 19.2%, respectively [50,51]. The prevalence rates were high in countries such as UAE, Oman and Saudi Arabia at 50% [52].

The high prevalence of α -thalassemia in the past was due to the lack of knowledge regarding the seriousness of this disease among the populations in these countries, especially those living in rural areas with limited access to education and those who could not afford to obtain the proper education similar to that found in urban areas. α -thalassemia is an inherited disease and the mutations may pass from parent to child, affecting the haemoglobin production. Hence, the educational and screening campaigns regarding this disease conducted by the representative bodies (either government or non-government organization) have successfully reduced the prevalence of α -thalassemia in the Southeast Asia region.

Random effects models were used, which are based on the assumption that the true effect could vary between studies [51]. The existence of publication bias in this meta-analysis was determined using a funnel plot and Egger's tests. The shape of the funnel plot in this meta-analysis showed an obvious symmetry, indicating the risk of publication bias is significantly low. This hypothesis was also supported by statistical evidence from Egger's test (t -value = 1.24; p -value = 0.112) in which the publication bias did not significantly exist in this meta-analysis. Therefore, we concluded that there was no publication bias detected in this meta-analysis.

When stratified according to country, Vietnam has the leading α -thalassemia prevalence rate of 51.5%. The high prevalence rate is probably due to one of the observational studies conducted in Vietnam focusing on the country's minority ethnic groups and thus, this likely skewed the actual prevalence rate [51]. The prevalence of α -thalassemia in Cambodia was the second highest (39.5%) when compared to other countries included in this meta-analysis. However, since there was only two studies that reported the prevalence of α -thalassemia Cambodia and Vietnam, more data are required to estimate the actual prevalence rate of this disease in both of these countries. The prevalence of α -thalassemia in Laos, Malaysia, and Thailand was quite similar, ranging from 17.3% to 26.8%. Alpha thalassemia is an inheritable disease where the presence of multiple deletional and non-deletional mutations can cause severe clinical complications. The presence of α -thalassemia major causes hydrops fetalis and prenatal deaths [4,5]. Therefore, the low prevalence of α -thalassemia in these countries and this region indicates that genetic screening for α -thalassemia mutations in the parents could be done in a population focused approach. Allele frequency and genetic diversity amongst the different populations provide information that can be used effectively in designing thalassemia prevention programs [53].

Thalassemia patients suffers from anemia caused by the imbalance of globin chains and impairment of haemoglobin solubility of erythrocytes. The reduced globin chains were shown to impair the cytoadherence of *Plasmodium* [54,55]. These abnormalities of erythrocytes have been shown to confer a protective effect against malaria infection [12,56]. Hence, there is a natural selection pressure which causes thalassemia becoming prevalent in countries with high incidence of malaria. Our meta-analysis was not exhaustive, however it shows that Vietnam had the highest prevalence rate (51.5%) and the lowest malaria cases (5794 cases) among the countries included in our study which supported the protective factor of thalassemia [57].

There are several limitations that should be addressed in this meta-analysis. Firstly, only data from certain Southeast Asian countries were available to be included in this meta-analysis; therefore, the calculated α -thalassemia-related prevalence rate in this study was specific to selected Southeast Asian regions. Besides, only studies from 2010 to 2020 were included in this meta-analysis, and it is possible for studies published before the year 2010 that meet the inclusion criteria but were not included in this meta-analysis, as this study focused on the prevalence rates from recent past 10 years only. We also did not include ethnic stratification because the majority of the studies included in this analysis did not report the ethnic group in the subject population. Alpha thalassemia genotype stratification was not performed due to inconsistencies in the reporting of genotypes in the studies included in this analysis.

5. Conclusions

This is the first meta-analysis that investigated α -thalassemia prevalence in Southeast Asian countries, and the findings suggest high prevalence of α -thalassemia events in certain countries which warrants attention as α -thalassemia major could cause severe health complications and impose a substantial burden to the health authority and families. The data in this meta-analysis may be beneficial to the representative bodies in designing educational and screening campaigns regarding this disease in order to further reduce α -thalassemia rates in these countries.

Supplementary Materials: The following are available online at <http://www.mdpi.com/1660-4601/17/20/7354/s1>, Table S1: PRISMA checklist of items to include when reporting a systematic review or meta-analysis, Table S2: Initial literature search from Pubmed, Table S3: Initial literature search from SCOPUS, Table S4: The SCOPUS literature ($N = 152$) included in the analysis, Table S5: The Pubmed literature ($N = 126$) included in the analysis.

Author Contributions: Conceptualization, L.P.W.G. and P.-C.L.; methodology, L.P.W.G. and E.T.J.C.; validation, E.T.J.C.; investigation, L.P.W.G. and E.T.J.C.; writing—original draft, L.P.W.G.; writing—reviewing and editing, L.P.W.G. and P.-C.L.; resources, P.-C.L.; supervision, P.-C.L. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Higgs, D.R.; Engel, J.D.; Stamatoyannopoulos, G. Thalassemia. *Lancet* **2012**, *379*, 73–383. [[CrossRef](#)]
2. Rosnah, B.; Rosline, H.; Zaidah, A.W.; Noor Haslina, M.N.; Marini, R.; Shafini, M.Y.; Nurul Ain, F.A. Detection of common deletional alpha-thalassemia spectrum by molecular technique in Kelantan, Northeastern Malaysia. *ISRN Hematol.* **2010**, *2012*, 462969. [[CrossRef](#)] [[PubMed](#)]
3. Tan, J.I.M.A.; Lee, P.C.; Wee, Y.C.; Tan, K.L.; Mahali, N.F.; George, E.; Chua, K.H. High prevalence of alpha and beta-thalassemia in the Kadazadusuns in East Malaysia: Challenges in providing effective health care for an indigenous group. *J. Biomed. Biotechnol.* **2010**, *2010*, 706872. [[CrossRef](#)] [[PubMed](#)]
4. Azma, R.Z.; Ainoon, O.; Hafiza, A.; Azlin, I.; Noor Farisah, A.R.; Nor Hidayati, S.; Noor Hamidah, H. Molecular characteristic of alpha thalassemia among patients diagnosed in UKM medical centre. *Malays. J. Pathol.* **2014**, *36*, 27–32.
5. Kulaphisit, M.; Kampuansai, J.; Leecharoenkiat, K.; Wathikthinnakon, M.; Kangwanpong, D.; Munkongdee, T.; Svasti, S.; Fucharoen, S.; Smith, D.R.; Lithanatudom, P. A comprehensive ethnic-based analysis of alpha thalassemia allele frequency in northern Thailand. *Sci. Rep.* **2017**, *7*, 4690. [[CrossRef](#)]
6. Farashi, S.; Harteveld, C.L. Molecular basis of α -thalassemia. *Blood Cells Mol. Dis.* **2018**, *70*, 43–53. [[CrossRef](#)]
7. Chui, D.H.K. Alpha-Thalassemia: Hb H disease and Hb Bart's hydrops fetalis. *Ann. N. Y. Acad. Sci.* **2005**, *1054*, 25–32. [[CrossRef](#)]
8. Weatherall, D.J.; Clegg, J.B. *The Thalassemia Syndrome*, 4th ed.; Blackwell Scientific Publication: Oxford, UK, 2011.
9. Casale, M.; Meloni, A.; Filosa, A.; Cuccia, L.; Caruso, V.; Palazzi, G.; Rita Gamberini, M.; Pitrolo, L.; Caterina Putti, M.; Giuseppe D'Ascola, D.; et al. Multiparametric Cardiac Magnetic Resonance Survey in Children with Thalassemia Major. *Circ. Cardiovasc. Imaging* **2015**, *8*, e003230. [[CrossRef](#)]
10. Kurtoglu, A.U.; Kurtoglu, E.; Temizkan, A.K. Effect of iron overload on endocrinopathies in patients with beta-thalassaemia major and intermedia. *Endokrynol. Pol.* **2012**, *63*, 260–263.
11. Suthat, F.; Pranee, W. Haemoglobinopathies in Southeast Asian. *Indian J. Med. Res.* **2011**, *134*, 498–506.
12. Galanello, R.; Cao, A. Alpha-thalassemia. *Gen. Med.* **2011**, *13*, 83–88. [[CrossRef](#)]
13. Kuesap, J.; Chaijaroenkul, W.; Rungsahirunrat, K.; Pongjantharasatien, K.; Na-Bangchang, K. Coexistence of Malaria and Thalassemia in malaria endemic areas of Thailand. *Korean J. Parasitol.* **2015**, *53*, 265–270. [[CrossRef](#)] [[PubMed](#)]
14. Rosanas-Uegell, A.; Senn, N.; Raru, P.; Aponte, J.J.; Reeder, J.C.; Siba, P.M.; Michon, P.; Mueller, I. Lack of associations of α (+)-thalassemia with the risk of Plasmodium falciparum and Plasmodium vivax infection and disease in a cohort of children aged 3–21 months from Papua New Guinea. *Int. J. Parasitol.* **2012**, *42*, e1000097. [[CrossRef](#)]
15. Vento, S.; Cainelli, F.; Cesario, F. Infections and thalassemia. *Lancet Infect. Dis.* **2006**, *6*, 226–233. [[CrossRef](#)] [[PubMed](#)]
16. Ryan, K.; Bain, B.J.; Worthington, D.; James, J.; Plews, D.; Mason, A.; Roper, D.; Rees, D.C.; de la Salle, B.; Streetly, A.; et al. Significant haemoglobinopathies: Guidelines for screening and diagnosis. *Br. J. Haematol.* **2010**, *149*, 35–49. [[CrossRef](#)]
17. Lithanatudom, P.; Khampan, P.; Smith, D.R.; Svasti, S.; Fucharoen, S.; Kangwanpong, D.; Kampuansai, J. The prevalence of alpha-thalassaemia amongst Tai and Mon-Khmer ethnic groups residing in northern Thailand: A population-based study. *Hematology* **2016**, *21*, 480–485. [[CrossRef](#)]
18. O'Riordan, S.; Hien, T.T.; Miles, K.; Allen, A.; Quyen, N.N.; Hung, N.Q.; Anh, D.Q.; Tuyen, L.N.; Khia, D.B.; Thai, C.Q.; et al. Large scale screening for haemoglobin disorders in southern Vietnam: Implications for avoidance and management. *Br. J. Haematol.* **2010**, *150*, 359–364. [[CrossRef](#)]
19. Moher, D.; Loberati, A.; Tetzlaff, J.; Altman, D.G. The PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. *PLoS Med.* **2009**, *6*, e1000097. [[CrossRef](#)]
20. Dersimonian, R.; Laird, N. Meta-analysis in clinical trials. *Control. Clin. Trials.* **1986**, *7*, 177–188. [[CrossRef](#)]
21. Higgins, J.P.; Thompson, S.G.; Deeks, J.J.; Altman, D.G. Measuring inconsistency in meta analyses. *BMJ* **2003**, *327*, 557–560. [[CrossRef](#)]

22. Egger, M.; Smith, G.D.; Schneider, M.; Minder, C. Bias in meta-analysis detected by a simple, graphical test. *BMJ* **1997**, *315*, 629–634. [[CrossRef](#)] [[PubMed](#)]
23. Light, R.J.; Pillemer, D.B. *Summing up: The Science of Reviewing Research*; Harvard University Press: Cambridge, MA, USA, 1984.
24. Borenstein, M.; Hedges, L.; Higgins, J.; Rothstein, H.R. Comprehensive meta-analysis version 2. *Englewood* **2005**, *104*, 188–191.
25. Munkongdee, T.; Tanakulmas, J.; Butthep, P.; Winichagoon, P.; Main, B.; Yiannakis, M.; George, J.; Devenish, R.; Fucharoen, S.; Svasti, S. Molecular epidemiology of hemoglobinopathies in Cambodia. *Hemoglobin* **2016**, *40*, 163–167. [[CrossRef](#)] [[PubMed](#)]
26. Jomoui, W.; Fucharoen, G.; Sanchaisuriya, K.; Charoenwijitkul, P.; Maneesarn, J.; Xu, X.; Fucharoen, S. Genetic origin of α^0 -thalassemia (SEA deletion) in Southeast Asian populations and application to accurate prenatal diagnosis of Hb Bart's hydrops fetalis syndrome. *J. Hum. Gen.* **2017**, *62*, 747–754. [[CrossRef](#)] [[PubMed](#)]
27. Wongprachum, K.; Sanchaisuriya, K.; Dethvongphanh, M.; Norcharoen, B.; Vidamaly, V.; Sanchaisuriya, P.; Fucharoen, S.; Fucharoen, G.; Schelp, F.P. Molecular heterogeneity of thalassemia among pregnant Laotian women. *Acta Hematol.* **2016**, *135*, 65–69. [[CrossRef](#)] [[PubMed](#)]
28. Tritipsombut, J.; Sanchaisuriya, K.; Phollarp, P.; Bouakhasith, D.; Sanchaisuriya, P.; Fucharoen, G.; Fucharoen, S.; Schelp, F.P. Micromapping of thalassemia and hemoglobinopathies in different regions of northeast Thailand and Vientiane, Laos people's democratic republic. *Hemoglobin* **2012**, *36*, 47–56. [[CrossRef](#)] [[PubMed](#)]
29. Azma, R.Z.; Ainoon, O.; Azlin, I.; Hamenuddin, H.; Hadi, N.A.; Tatt, W.K.; Syazana, I.N.; Asmaliza, A.M.; Das, S.; Hamidah, N.H. Prevalence of iron deficiency anaemia and thalassemia trait among undergraduate medical students. *Clin. Ter.* **2012**, *163*, 287–291.
30. Jameela, S.; Sharifah Sabirah, S.O.; Babam, J.; Phan, C.L.; Visalachy, P.; Chang, K.M.; Salwana, M.A.; Zuridah, A.; Subramanian, Y.; Rahimah, A. Thalassemia screening among students in a secondary school in Ampang, Malaysia. *Med. J. Malays.* **2011**, *66*, 522–524.
31. Mohd Yatim, N.F.; Abd Rahim, M.; Menon, K.; Al-Hassan, F.M.; Ahmad, R.; Manocha, A.B.; Saleem, M.; Yahaya, B.H. Molecular characterization of α and β -thalassaemia among malay patients. *Int. J. Mol. Sci.* **2014**, *15*, 8835–8845. [[CrossRef](#)]
32. Charoenkwan, P.; Taweephol, R.; Sirichotiyakul, S.; Tantiprabha, W.; Sae-Tung, R.; Suanta, S.; Sakdasirithaporn, P.; Sanguansermisri, T. Cord blood screening for α -thalassemia and hemoglobin variants by isoelectric focusing in northern Thai neonates: Correlation with genotypes and hematologic parameters. *Blood Cells Mol. Dis.* **2010**, *45*, 53–57. [[CrossRef](#)]
33. Nillakupt, K.; Nathalang, O.; Arnutti, P.; Jindadamrongwech, S.; Boonsiri, T.; Panichkul, S.; Areekul, W. Prevalence and hematological parameters of thalassemia in the Kradarn subdistrict Chachoengsao province, Thailand. *J. Med. Assoc. Thai.* **2011**, *95*, S124–S132.
34. Pongjantharasatien, K.; Banyatsuppasin, W.; Pounsawat, S.; Jindadamrongwech, S. Occurrence of the $-\text{SEA}$, $-\text{THAI}$, and $-\text{FIL}$ α -thalassemia-1 carriers from a 7-year study at Ramathibodi hospital, Bangkok, Thailand. *Hemoglobin* **2016**, *40*, 283–284. [[CrossRef](#)] [[PubMed](#)]
35. Pichanun, D.; Munkongdee, T.; Klamchuen, S.; Butthep, P.; Winichagoon, P.; Fucharoen, S.; Svasti, S. Molecular screening of the Hbs constant spring (codon 142, TAA>CAA, $\alpha 2$) and paksé (codon 142, TAA>TAT, $\alpha 2$) mutations in Thailand. *Hemoglobin* **2010**, *34*, 582–586. [[CrossRef](#)] [[PubMed](#)]
36. Pharephan, S.; Sirivatanapa, P.; Makonkawkeyoon, S.; Tuntiwechapikul, W.; Makonkawkeyoon, L. Prevalence of α -thalassemia genotypes in pregnant women in northern Thailand. *Indian J. Med. Res.* **2016**, *143*, 315–322. [[CrossRef](#)]
37. Panyasai, S.; Fucharoen, G.; Fucharoen, S. Hemoglobin variants in Northern Thailand: Prevalence, heterogeneity and molecular characteristics. *Genet. Test Mol. Biomark.* **2016**, *20*, 37–43. [[CrossRef](#)]
38. Panomai, N.; Sanchaisuriya, K.; Yamsri, S.; Sanchaisuriya, P.; Fucharoen, S.; Schelp, F.P. Thalassemia and iron deficiency in a group of northeast Thai school children: Relationship to the occurrence of anaemia. *Eur. J. Pediatr.* **2010**, *169*, 1317–1322. [[CrossRef](#)]
39. Prayalaw, P.; Fuchafoen, G.; Fucharoen, S. Routine screening for α -thalassemia using an immunochromatographic strip assay for haemoglobin Bart's. *J. Med. Screen.* **2014**, *21*, 120–125. [[CrossRef](#)]
40. Seeratanachot, T.; Shimbhu, D.; Charoenkwan, P.; Sanguansermisri, T. Detection of deletion α^+ -thalassemia mutation [$-\alpha$ (3.7), $-\alpha$ (4.2)] by quantitative PCR assay. *Southeast Asian J. Trop. Med. Public Health* **2015**, *46*, 110–115.

41. Wisedpanichkij, R.; Jindadamrongwech, S.; Butthep, P. Identification of Hb constant spring (HBA2: c.427T>C) by an automated high performance liquid chromatography method. *Hemoglobin* **2015**, *39*, 190–195. [[CrossRef](#)]
42. Uaprasert, N.; Settapiboon, R.; Amomsiriwat, S.; Sarnthammakul, P.; Thanapat, T.; Rojnuckarin, P.; Sutcharithchan, P. Diagnostic utility of isoelectric focusing and high performance liquid chromatography in neonatal cord blood screening for thalassemia and non-sickling hemoglobinopathies. *Clin. Chim. Acta.* **2013**, *427*, 23–26. [[CrossRef](#)]
43. Srivorakun, H.; Fucharoen, G.; Changtrakul, Y.; Komwilaisak, P.; Fucharoen, S. Thalassemia and hemoglobinopathies in South East Asia newborns: Diagnostic assessment using capillary electrophoresis system. *Clin. Biochem.* **2011**, *44*, 406–411. [[CrossRef](#)] [[PubMed](#)]
44. Chaibunruang, A.; Prommetta, S.; Yamsri, S.; Fucharoen, G.; Sae-Ung, N.; Sanchaisuriya, K.; Fucharoen, S. Molecular and hematological studies in a large cohort of α^0 -thalassemia in northeast Thailand: Data from a single referral centre. *Blood Cells Mol. Dis.* **2013**, *51*, 89–93. [[CrossRef](#)] [[PubMed](#)]
45. Thanyaornwanya, C.; Singha, K.; Fucharoen, G.; Sanchaisuriya, K.; Thepphitak, P.; Wintachai, P.; Karnpean, R.; Fucharoen, S. Molecular characteristics of α^+ -thalassemia (3.7 kb deletion) in Southeast Asia: Molecular subtypes, haplotypic heterogeneity, multiple founder effects and laboratory diagnosis. *Clin. Biochem.* **2019**, *71*, 31–37. [[CrossRef](#)]
46. Mankhenthong, K.; Phusua, A.; Suantan, S.; Srisittipoj, P.; Charoenkwan, P.; Sanguansermisri, T. Molecular characteristics of thalassemia and haemoglobin variants in prenatal diagnosis program in northern Thailand. *Int. J. Hematol.* **2019**, *110*, 474–481. [[CrossRef](#)]
47. Pata, S.; Laopajon, W.; Pongpaiboon, M.; Thongkum, W.; Polpong, N.; Munkongdee, T.; Paiboonsukwong, K.; Fucharoen, S.; Tayapiwatana, C.; Kasinrer, W. Impact of the detection of ζ -globin chains and haemoglobin Bart's using immunochromatographic strip test for α^0 -thalassemia ($-\text{SEA}$) differential diagnosis. *PLoS ONE* **2019**, *14*, e0223996. [[CrossRef](#)]
48. Hoa Nguyen, V.; Sanchaisuriya, K.; Wongprachum, K.; Nguyen, M.D.; Phan, T.T.; Vo, V.T.; Sanchaisuriya, P.; Fucharoen, S.; Schelp, F.P. Hemoglobin constant spring is markedly high in women of an ethnic minority group in Vietnam: A community-based survey and hematologic features. *Blood Cell Mol. Dis.* **2014**, *52*, 161–165. [[CrossRef](#)]
49. Modell, B.; Darlison, M. Global epidemiology in haemoglobin disorders and derived service indicators. *Bull. World Health Organ.* **2008**, *86*, 480–487. [[CrossRef](#)]
50. Nadkarni, A.; Phanasgaonkar, S.; Colah, R.; Mohanty, D.; Ghosh, K. Prevalence and Molecular Characterization of α -Thalassemia Syndromes among Indians. *Genet. Test.* **2008**, *12*, 177–180. [[CrossRef](#)]
51. Souza, A.E.S.; Cardoso, G.L.; Takashi, S.Y.L.; Guerreiro, J.F. α -Thalassemia (3.7 kb deletion) in a population from the Brazilian Amazon region: Santarém, Pará State. *Genet. Mol. Res.* **2009**, *8*, 477–481. [[CrossRef](#)]
52. AL-Awamy, B.H. Thalassemia syndromes in Saudi Arabia. Meta-analysis of local studies. *Saudi Med. J.* **2000**, *21*, 8–17.
53. Kee, B.P.; Lian, L.H.; Lee, P.C.; Lai, T.X.; Chua, K.H. Genetic data for 15 STR loci in a Kadazan-Dusun population from East Malaysia. *Genet. Mol. Res.* **2011**, *10*, 739–743. [[CrossRef](#)] [[PubMed](#)]
54. Forget, B.G.; Bunn, H.F. Classification of the disorders of haemoglobin. *Cold Spring Harb. Perspect. Med.* **2013**, *3*, a011684. [[CrossRef](#)]
55. Krause, M.A.; Diakite, S.A.S.; Lopera-Mesa, T.M.; Amaratunga, C.; Arie, T.; Traore, K.; Doumbia, S.; Konate, D.; Keefer, J.R.; Diakite, M.; et al. α -thalassemia impairs the cytoadherence of Plasmodium falciparum-infected erythrocytes. *PLoS ONE* **2012**, e37214. [[CrossRef](#)] [[PubMed](#)]
56. Gundula, M.-O.; Gros, P. Erythrocyte variants and the nature of their malaria protective effect. *Cell. Microbiol.* **2005**, *7*, 753–763. [[CrossRef](#)]
57. World Health Organization. *World Malaria Report 2019*; World Health Organization: Geneva, Switzerland, 2018.

