

A Flexible Electrochemiluminescence Sensor Equipped With Vertically Ordered Mesoporous Silica Nanochannel Film for Sensitive Detection of Clindamycin

Xinjie Wei^{1†}, Xuan Luo^{2†}, Shuai Xu², Fengna Xi^{2*} and Tingting Zhao^{1*}

¹Guangxi Medical University Cancer Hospital, Guangxi Medical University, Nanning, China, ²Department of Chemistry, Key Laboratory of Surface and Interface Science of Polymer Materials of Zhejiang Province, Zhejiang Sci-Tech University, Hangzhou, China

OPEN ACCESS

Edited by:

Yan Zhang, University of Jinan, China

Reviewed by:

Chuanxiang Chen, Jiangsu University of Science and Technology, China Qiang Ma, Jilin University, China

*Correspondence:

Fengna Xi fengnaxi@zstu.edu.cn Tingting Zhao gxzhaott@163.com [†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Analytical Chemistry, a section of the journal Frontiers in Chemistry

Received: 09 February 2022 Accepted: 11 March 2022 Published: 06 April 2022

Citation:

Wei X, Luo X, Xu S, Xi F and Zhao T (2022) A Flexible Electrochemiluminescence Sensor Equipped With Vertically Ordered Mesoporous Silica Nanochannel Film for Sensitive Detection of Clindamycin. Front. Chem. 10:872582. doi: 10.3389/fchem.2022.872582 Fast, convenient, and highly sensitive detection of antibiotic is essential to avoid its overuse and the possible harm. Owing to enrichment effect and antifouling ability of ultrasmall nanochannels, the vertically ordered mesoporous silica nanochannel film (VMSF) has great potential in the development of the facile electrochemiluminescence (ECL) sensor for direct and sensitive analysis of antibiotics in complex samples. In this study, we demonstrated a flexible ECL sensor based on a cost-effective electrode covered with a VMSF for sensitive detection of clindamycin. Polyethylene terephthalate coated with indium tin oxide (PET-ITO) is applied as a flexible electrode to grow VMSF using the electrochemically assisted self-assembly (EASA) method. The negatively charged VMSF nanochannels exhibit significant enrichment toward the commonly used cationic ECL luminophores, tris(2,2bipyridyl) dichlororuthenium (II) (Ru (bpy)₃²⁺). Using the enhanced ECL of Ru (bpy)₃²⁺ by clindamycin, the developed VMSF/PET-ITO sensor can sensitively detect clindamycin. The responses were linear in the concentration range of 10 nM-25 μ M and in the concentration range of 25–70 µM. Owing to the nanoscale thickness of the VMSF and the high coupling stability with the electrode substrate, the developed flexible VMSF/PET-ITO sensor exhibits high signal stability during the continuous bending process. Considering high antifouling characteristic of the VMSF, direct analysis of clindamycin in a real biological sample, human serum, is realized.

Keywords: electrochemiluminescence sensor, flexible electrode, vertically ordered mesoporous silica nanochannel film, sensitive detection, clindamycin

INTRODUCTION

Antibiotics, drugs that can inhibit and kill bacteria, have been widely used in medical and healthcare, livestock and poultry breeding, agricultural production, and other industries. However, the excessive and irrational use of antibiotics is now widespread. Antibiotic residues can cause serious harm to human health including carcinogenic, teratogenic, and mutagenic effects (Wang et al., 2008; Wong et al., 2016; Hadi and Honarmand, 2017; Paul et al., 2017). In addition, prolonged exposure to such drugs can lead to severe antibiotic-resistant infections. For instance, clindamycin is a lincosamide

1

antibiotic, which is widely used for the treatment of bacterial infections. The side effects of clindamycin include dry skin, nausea, vomiting, and constipation (Liu et al., 2010; Habib et al., 2011; Girondi et al., 2018; Mohammad Beigi et al., 2020; Liang et al., 2021). Researchers are also highly concerned about the bacterial resistance of clindamycin due to its overuse. Therefore, rapid, low-cost, and highly sensitive detection of antibiotics is crucial to standardize the use of antibiotics and avoid harm.

Until now, the developed technologies for the detection of antibiotics include capillary electrophoresis (CE), highperformance liquid chromatography (HLPC), ultrahighperformance liquid chromatography-electrospray tandem mass spectrometry (UPLC-ESI-MS/MS), and electrochemical sensors (Liu et al., 2010; Habib et al., 2011; Hadi and Honarmand, 2017; Girondi et al., 2018; Mohammad Beigi et al., 2020; Liang et al., 2021). However, these methods usually suffer time-consuming detection, expensive instruments, complicated operation, or low sensitivity. Electrochemiluminescence (ECL), where chemiluminescence (CL) is triggered and controlled by electrochemical methods, elegantly combines the electrochemical and optical methods (Chen et al., 2011; Li et al., 2017; Zhai et al., 2017; Ma et al., 2019). Compared with photoluminescence with photo-excitation, ECL has advantages of near-zero background, high detection sensitivity, simplified optical instrument, and good temporal/spatial resolution (Wang et al., 2020; Zhang et al., 2020; Wang et al., 2021a; Huang et al., 2021; Ma et al., 2021; Yang et al., 2021; Zhu et al., 2021). Thus, ECL has emerged as a powerful tool in clinical diagnosis, drug analysis, immunoassay, environmental analysis, etc. In addition to constructing an ECL sensor on conventional electrochemical electrodes, development of a flexible, cost-efficient, and disposable ECL sensor with high sensitivity and its application in sensitive detection of antibiotic is of great significance.

The modification of the supporting electrode can significantly affect the performance of ECL sensors. As functional nanomaterials in molecular sieves, nanofluids, sensing and energy conversion, solid-state nanofilms (SSN) have attracted great attention owing to advantages of tunable nanopores, high specific surface area, and intelligent control of molecular transport (Zhou et al., 2019a; Zhou et al., 2019b; Zhou et al., 2020a; Ding et al., 2020; Li et al., 2020). Among the developed SSN, vertically ordered mesoporous silica nanochannel film (VMSF) exhibits high chemical and thermal stability, ordered and vertically aligned nanochannels (nanopores), uniform pore size distribution, high pore density, and ease of modification (Nasir et al., 2018; Jiokeng et al., 2019; Ullah et al., 2021; Walcarius, 2021). The ultrasmall (usually 2-3 nm) and high-density (up to $4 \sim 8 \times 10^{12}$ /cm²) nanochannel arrays make it possible to significantly improve the sensor performance as electrode modification materials (Mi et al., 2020; Li et al., 2021a; Asadpour et al., 2021; Li et al., 2021b; Xiao et al., 2021). On the one hand, a large number of silanol groups (Si-OH) with low pKa (~2) endow the VMSF with a negatively charged surface, which can accelerate the transfer of positively charged

molecules to the electrode surface, leading to a significant enrichment effect. On the other hand, ultrasmall nanochannels of the VMSF can exclude large-sized substances, which can effectively suppress the interference of coexisting substances in complex samples, providing high antifouling ability and good signal stability (Yan et al., 2020a; Yan et al., 2020b; Lin et al., 2020; Liu et al., 2020; Ma et al., 2020; Yan et al., 2021a; Wang et al., 2021b; Yan et al., 2021b; Qiu et al., 2021; Yanyan et al., 2021). Therefore, the VMSF has great potential in the development of facile ECL sensors for direct and sensitive analysis of antibiotics in complex samples.

work. demonstrate flexible In this we а electrochemiluminescence sensor based on covering the electrode with vertically ordered mesoporous silica nanochannel films, which enable direct and sensitive detection of clindamycin in human serum. As shown in Figure 1, polyethylene terephthalate coated with indium tin oxide (PET-ITO), which is flexible, cheap, and suitable for the fabrication of portable, integrated, or disposable sensors, is used as the supporting electrode to grow VMSFs using the electrochemically assisted self-assembly (EASA) method. The significant enrichment toward the commonly used cationic ECL luminophores tris(2,2-bipyridyl) dichlororuthenium (II) $[Ru (bpy)_3^{2+}]$ is proved resulting from the negatively charged VMSF nanochannels. As clindamycin can act as coreactant to promote the ECL of Ru $(bpy)_3^{2+}$, the developed VMSF/PET-ITO sensor can detect clindamycin with high sensitivity. In addition, high signal stability during the continuous blending process is revealed, showing the unique properties of flexible electrodes. Owing to the high antifouling characteristic of the VMSF, direct analysis of clindamycin in real biological sample, human serum, is also realized.

MATERIALS AND METHODS

Chemicals and Materials

Cetyltrimethylammonium bromide (CTAB), tetraethoxysilane (TEOS), clindamycin hydrochloride, tris(2, 2'-bipyridyl) ruthenium (II), acetone, Na₂HPO₄, NaH₂PO₄, potassium hydrogen phthalate (KHP), NH₄OH (25%), and H₂O₂ (30%) were purchased from Aladdin Chemistry Co., Ltd. (China). Polyethylene terephthalate coated with indium tin oxide (PET-ITO) with surface resistivity of $5.5-6.1 \Omega$ /sq was purchased from Zhuhai Kaivo Optoelectronic Technology (China). Human blood serum was provided by the Center for Disease Control and Prevention (Hangzhou, China). Ultrapure water (18.2 M Ω cm) prepared using Mill-Q system (Millipore, United States) was used to prepare the aqueous solutions in this work.

Measurements and Instrumentations

Transmission electron microscopic (TEM) images were taken on a JEM-2100 transmission electron microscope (JEOL Ltd., Japan) using an operating voltage of 200 kV. Before measurement, the VMSF was scrapped from PET-ITO and dispersed in ethanol. Copper grid was applied to support the VMSF in TEM characterization. Scanning electron microscopic



(SEM) image was taken under a field-emission scanning electron microscopy (S-4800, Hitachi, Japan). CHI 832C electrochemical workstation (CH Instrument, China) and MPI-E workstation (Remex Analysis, China) were used for ECL measurement. The detection cell was fixed in a dark detection chamber and had a volume of ~7 ml. Electrochemical measurements including cyclic voltammetry (CV) and electrochemical impedance spectroscopy (EIS) were carried out on an AutoLab electrochemical station (PGSTAT302N, Metrohm, Switzerland). Three electrode system was used in both electrochemical and ECL investigations. Briefly, bare or modified PET-ITO electrodes act as a working electrode. Ag/AgCl electrode (saturated with KCl solution) is applied as the reference electrode and Pt sheet electrode is the counter electrode.

Growth of Vertically Ordered Mesoporous Silica Nanochannel Film on Polyethylene Terephthalate Coated With Indium Tin Oxide

To improve the hydrophilicity of PET-ITO, the electrode was immersed in a mixed solution containing ultrapure water/ hydrogen peroxide/ammonium hydroxide (5:1:1, v/v) under dark conditions for 1.5 h. Then, the electrode was washed thoroughly with ultrapure water. The VMSF was equipped on PET-ITO using the electrochemically assisted self-assembly method. Briefly, 20 ml NaNO₃ (0.1 mol/L, pH = 3), 20 ml ethanol, 2.833 g TEOS, and 1.585 g CTAB were mixed, and the obtained mixture was stirred for 2.5 h to obtain the precursor solution. For VMSF growth, a constant current (-0.35 mA/cm²) was applied on PET-ITO for 10 s. Then the electrode was quickly rinsed with ultrapure water. After drying under N₂ stream, the electrode with surfactant micelles (SM) in the nanochannels (SM@VMSF/PET-ITO) was aged at 120°C for 10 h. Then, SM was removed by treating the SM@VMSF/PET-ITO in acetone and stirring for 30 min. Finally, the VMSF/PET-ITO with open nanochannels was obtained.

Electrochemiluminescence Determination of Clindamycin

Phosphate-buffered saline (PBS, 0.01 mol/L, pH 7.0) containing Ru (bpy)₃²⁺ (10 μ mol/L) was used as the supporting solution to detect clindamycin. When different concentrations of clindamycin were added, the ECL signal on the VMSF/PET-ITO electrode generated during CV scan (potential ranged from 0 to 1.4 V at a scan rate of 100 mV/s) was recorded. The standard addition method was used to evaluate the reliability of the developed ECL sensor in analysis of a real sample. Briefly, the human serum was diluted by a factor of 50 with PBS. Then, ECL detection was carried out after adding a certain amount of clindamycin.

RESULTS AND DISCUSSION

Facile Coupling Vertically Ordered Mesoporous Silica Nanochannel Film on Flexible Polyethylene Terephthalate Coated With Indium Tin Oxide Electrode and Charge Selectivity of Nanochannels

The flexible electrode is a research hotspot in recent years (Zhou et al., 2020b; İnce and Sezgintürk, 2021; Avelino et al., 2021). Compared with traditional hard electrodes, the flexible electrode has great potential in flexible or wearable sensors owing to the unique abilities of folding, bending, and multi-layer winding. Therefore, a simple and efficient modification of the flexible electrode to improve the detection sensitivity and antifouling ability in the complex sample is of great significance. **Figure 1** is



the schematic illustration for coupling VMSF on the flexible PET-ITO electrode using the electrochemically assisted self-assembly (EASA) method. Among the developed strategies to prepare the VMSF, the EASA method is fast and efficient which can achieve rapid growth of the VMSF within 5-30 s at room temperature. Briefly, the VMSF with a pore diameter of 2-3 nm is formed by using cetyltrimethylammonium bromide (CTAB) micelles (SM) as the template. The mechanism lies in the controlled condensation kinetics of siloxane around the SM template under the *in situ* pH gradient resulting from the reduction of protons and water molecules when a negative voltage is applied to the electrode. Researchers have systematically investigated the effects of experimental conditions on the film formation and properties of VMSF (Herzog et al., 2013). It has been proven that the electrode surface cannot be completely covered by the VMSF, if a short electrodeposition time and a small current density are employed. On the contrary, a long electrodeposition time and a large current density lead to a thick film, which in turn weakens the permeability of the film. If the concentration of silane precursor is too small, the electrodes will not be fully covered by the film. However, a higher precursor concentration will result in a film that is too thick and prone to cracking during the aging stage. After VMSF growth, the resulting electrode contains surfactant SM (SM@VMSF/PET-ITO), which blocks the nanochannels. The efficient removal of SM in the VMSF is the key to obtain open nanochannels. The most common method to remove SM is to soak the SM@VMSF/PET-ITO electrode in HClethanol solution. However, PET will get damaged in HCl medium because of its hydrolysis reaction in strong acid. Thus, acetone is applied to remove SM in this work. Figure 2A shows the CV curves of Ru (NH₃)³⁺ obtained on different electrodes including bare PET-ITO, SM@VMSF/PET-ITO and VMSF/PET-ITO, respectively. To investigate the effect of soaking time in acetone on the removal of SM, VMSF/PET-ITO electrodes prepared using different soaking time under stirring are studied. As shown, no Faraday current is observed on the SM@VMSF/PET-ITO electrode. This is because the micelles

block the nanochannels so that the redox probes cannot reach the electrode surface. The result also proves that the VMSF is a complete film without cracks. In contrast, the VMSF/PET-ITO electrode without SM has significant current responses. In addition, the signal of Ru $(NH_3)_6^{3+}$ on the VMSF/PET-ITO electrode remains unchanged after the VMSF/PET-ITO electrode is soaked in acetone for 30 min, indicating efficient removal of SM. Thus, 30 min of soaking in acetone is chosen for further investigation.

Compared with bare PET-ITO, the VMSF/PET-ITO electrode shows enhanced signal for Ru (NH₃)³⁺, proving remarkable enrichment effect toward positively charges species. The deprotonation of silanols (pKa ~2) on the surface of VMSF nanochannels leads to negatively charged surface, which attracts the positively charged Ru (NH₃)³⁺ through electrostatic interactions. The successful growth of the VMSF and the charge selectivity of nanochannels are also confirmed by electrochemical impedance spectroscopy (EIS) obtained in the anionic $[Fe(CN)_6]^{3-/4}$ probe (Figure 2B). As shown, bare PET-ITO, SM@VMSF/PET-ITO, and VMSF/PET-ITO exhibit significantly different charge transfer resistance (Rct), which is related to the diameter of the curve semicircle in the high-frequency region. When SM exist in nanochannels, the SM@VMSF/PET-ITO electrode exhibits the largest Rct owing to the blocking of nanochannels. When SM is removed, the Rct for VMSF/PET-ITO electrode remarkably decreases. However, the Rct for the VMSF/PET-ITO electrode is still larger than that of bare PET-ITO, indicating the electrostatic repulsion of negatively charged nanochannels to negatively charged probes.

Morphology of Vertically Ordered Mesoporous Silica Nanochannel Film Nanochannels

The structure and morphology of the VMSF are characterized by TEM and SEM. **Figure 3A** is the top-view TEM images of





the VMSF at different magnification. As shown, the VMSF has a uniformly distributed nanopore structure without cracks or defects in the observed range. The highly ordered nanopores have a diameter of 2–3 nm. The cross-sectional TEM image of the VMSF in **Figure 3A** reveals nanochannels parallel to each other. The cross-sectional SEM image of the VMSF/PET-ITO electrode shows a three-layered structure, which corresponds to PET substrate, ITO layer, and VMSF layer from bottom to top. In addition, VMSF has a flat structure with a thickness of ~97 nm.

Enhanced Electrochemiluminescence by Clindamycin and Vertically Ordered Mesoporous Silica Nanochannel Film Nanochannels

Electrochemiluminescence has the unique advantage of simple instruments, low background, and easy operation. Although many ECL luminophores such as acridan ester, ruthenium chelate, luminol, and nanoemitters have been developed, tris(2,2'-bipyridyl) ruthenium (II) [Ru (bpy)₃²⁺] is still widely used due to high stability, solubility, and reversible electrochemical behavior (Zhou

et al., 2019a; Zhou et al., 2020a; Ding et al., 2020; Li et al., 2020). As shown in Figure 4A, when clindamycin is $(bpy)_3^{2+}$, added to Ru the ECL signal significantly enhances, indicating that clindamycin can promote the ECL emission. Owing to the quaternary amine structure in the clindamycin molecule, the possible ECL mechanism might be similar to that of Ru (bpy)₃²⁺/tri-npropylamine (TPrA) system. The ECL reaction pathway can be elucidated in the following equations (Eqs 1-5). Briefly, Ru $(bpy)_3^{2+}$ can be oxidized to Ru $(bpy)_3^{3+}$ when an oxidation potential of 1.4 V is applied to the electrode. At the same time, clindamycin is oxidized to radical cation (clindamycin^{.+}) followed with deprotonation to reduced product 1(clindamycin[•]), which form the Ru $(bpy)_{3}^{3+}$ undergoes redox reaction with to excited state [Ru $(bpy)_3^{2+*}$]. Then, generate the ECL is observed when the excited state returns to the ground state.

$$\operatorname{Ru}(\operatorname{bpy})_{3}^{2+} - e^{-} \to \operatorname{Ru}(\operatorname{bpy})_{3}^{3+}$$
(1)

Clindamycin – $e^- \rightarrow$ Clindamycin⁺⁺ (2)

 $Clindamycin^{\bullet+} \rightarrow Clindamycin^{\bullet} + H^{+}$ (3)



$$\operatorname{Ru}(\operatorname{bpy})_{3}^{3+} + \operatorname{Clindamycin}^{\bullet} \to \operatorname{Ru}(\operatorname{bpy})_{3}^{2+*}$$

$$\operatorname{Ru}(\operatorname{bpy})_{3}^{2+*} \to \operatorname{Ru}(\operatorname{bpy})_{3}^{2+} + \operatorname{hv}$$
(5)

It is worth noting that the ECL signal on the VMSF/PET-ITO electrode is about 7 times of that on the bare PET-ITO electrode (**Figure 4B**), indicating that the nanochannels of the VMSF have significant signal amplification effect. This was attributed to the enrichment of positively charged ECL probes by VMSF nanochannels.

Optimized Conditions for Electrochemiluminescence Detection of Clindamycin

To achieve the best detection performance, the concentration and pH of the supporting electrolyte are optimized. As shown in Supplementary Figure S1A (in supporting information, SI), the ECL intensity gradually decreases when the ionic strength of the supporting buffer increases. This is because the thickness of the electric of VMSF nanochannels is inversely double laver proportional to the ionic strength. The increase in thickness of the electric double layer of the nanochannels enhances the electrostatic adsorption of Ru $(bpy)_3^{2+}$, resulting in more ECL probes on the electrode surface. As can be seen from Supplementary Figure S1B (SI), the ECL intensity on the electrode gradually increases as the pH of the buffer increases from 4 to 8. This might be ascribed to the enhanced deprotonation of the radical cation of clindamycin (clindamycin⁺) to form strong reducing agent (clindamycin, Eq. 3), which facilitates the redox reaction with Ru $(bpy)_3^{3+}$ and the formation of the excited state (Ru $(bpy)_3^{2+*}$) (Leland and Powell, 1990). The phenomenon further confirms the coreactant mechanism for the enhanced ECL signal. The decrease of ECL signal at pH 9 might result from the instability of the VMSF at high alkaline medium. Considering the stability of the VMSF and the neutral

environment of common physiological samples, pH 7 is chosen for further experiments.

Sensitive Electrochemiluminescence Detection of Clindamycin

Under the optimized conditions, ECL detection of clindamycin is investigated. Figure 5A illustrates the ECL intensity on the VMSF/PET-ITO electrode in the presence of different concentrations of clindamycin. The ECL intensity (I_{ECL}) is linearly correlated to the concentration of clindamycin (C) in the range from 10 nmol/L to 25 μ mol/L (I_{ECL} = 227.2C + 1,106, $R^2 = 0.9941$) and from 25 µmol/L to 70 µmol/L (I_{ECL} = 82.45C + 4,635, $R^2 = 0.9995$) (Figure 5B). The limit of detection (LOD) is based on a signal-to-noise ratio of 3 (S/N = 3) is 4 nmol/L. Owing to the enrichment effect of VMSF nanochannels toward Ru $(bpy)_3^{2+}$, the LOD from the VMSF/PET-ITO electrode is lower than that obtained using square wave voltammetry (SWV) detection on a glassy carbon electrode modified with graphene oxide and gold nanoparticles within a film of cross-linked chitosan with epichlorohydrin (AuNP-GO-CTS-ECH/GCE electrode) or CV detection on edge-plane pyrolytic graphite electrode (EPPG) (Wong et al., 2016; Hadi and Honarmand, 2017). The LOD is also lower than that obtained using capillary electrophoresis (CE) or CE-ECL (Wang et al., 2008; Paul et al., 2017). Thus, the constructed ECL sensor has the advantages of simple preparation and high detection performance.

High Stability of Vertically Ordered Mesoporous Silica Nanochannel Film/ Polyethylene Terephthalate Coated With Indium Tin Oxide Electrode Under Continuous Bending

The signal stability under large deformation is a key factor in evaluating the performance of flexible electrodes. We compared the signals on the VMSF/PET-ITO electrode





before and after continuous bending. The micelle-containing electrode, SM@VMSF/PET-ITO electrode, is first investigated with continuous bending for 30 times. As shown in **Figure 6A**, CV curves of Ru $(NH_3)_6^{3+}$ on the original or bended electrode are quite similar. There was no significant Faraday signal before and after folding of the electrode. Thus, no crack of the VMSF layer appears during the continuous bending process, proving high stability of the flexible electrode. When the micelles are removed from the

electrode that was folded 30 times, the opening of the nanochannels leads to significant electrochemical signals of the redox probe. In addition, the stability of the ECL signal on the electrode during the continuous bending is also investigated. As shown in **Figure 6B**, the electrode before and after bending for 10 or 30 times exhibits unchanged ECL intensity, indicating high signal stability. This phenomenon is attributed to the nanoscale

thickness of the VMSF and the high coupling stability with the electrode substrate.

Antifouling of Vertically Ordered Mesoporous Silica Nanochannel Film/ Polyethylene Terephthalate Coated With Indium Tin Oxide Electrode and Real Sample Analysis

Biological, food, and environmental samples usually have complex matrix containing proteins, starch, surfactants, etc., which often contaminates the electrode surface and reduces signal accuracy. Thus, improving the antifouling ability of the electrode is of great significance for direct electroanalysis of complex samples. To evaluate the antifouling ability of the developed VMSF/PET-ITO sensor, bovine serum albumin (BSA), starch, lignin and sodium dodecyl sulfonate (SDS) are applied as the possible interfering substances. The ECL of Ru $(bpy)_3^{2+}$ with clindamycin in the absence or presence of one of these substances on both VMSF/PET-ITO and PET-ITO electrodes are given in Figure 7. As shown, the ECL signal significantly reduced on PET-ITO when one of the aforementioned substances is added, indicating remarkable fouling of the electrode. On the contrary, no significant change of ECL signal is observed on the VMSF/PET-ITO electrode, indicating high antifouling performance resulting from the remarkable molecular sieving ability of the VMSF.

Considering the antifouling ability of the VMSF, the developed VMSF/PET-ITO sensor is applied to detect clindamycin in human serum samples (diluted by a factor by 50) using the standard addition method (**Supplementary Table S1** in SI). The recovery rate of other three artificial added clindamycin concentration is between 97.6 and 104.0% with low relative standard deviation (RSD, $\leq 4.7\%$), indicating good reliability in the direct measurement of clindamycin in real complex samples.

CONCLUSION

In summary, we demonstrate a cost-effective and disposable electrochemiluminescence sensor based on the flexible electrode covered with a vertically ordered mesoporous silica nanochannel film for sensitive detection of clindamycin. The polyethylene terephthalate coated with indium tin oxide (PET-

REFERENCES

- Asadpour, F., Mazloum-Ardakani, M., Hoseynidokht, F., and Moshtaghioun, S. M. (2021). *In Situ* monitoring of Gating Approach on Mesoporous Silica Nanoparticles Thin-Film Generated by the EASA Method for Electrochemical Detection of Insulin. *Biosens. Bioelectron.* 180, 113124. doi:10.1016/j.bios.2021.113124
- Avelino, K. Y. P. S., Oliveira, L. S., Lucena-Silva, N., Andrade, C. A. S., and Oliveira, M. D. L. (2021). Flexible Sensor Based on Conducting Polymer and Gold Nanoparticles for Electrochemical Screening of HPV Families in Cervical Specimens. *Talanta* 226, 122118. doi:10.1016/j.talanta.2021.122118

ITO) is applied as the flexible electrode to easily grow the VMSF using the electrochemically assisted self-assembly (EASA) method. The negatively charged VMSF nanochannels exhibit significant enrichment toward the commonly used cationic ECL luminophores, tris(2,2-bipyridyl) dichlororuthenium (II) $[Ru (bpy)_3^{2+}]$. Using the enhanced ECL of Ru $(bpy)_3^{2+}$ by clindamycin, the developed VMSF/PET-ITO sensor can detect clindamycin with high sensitivity. Owing to the nanoscale thickness of the VMSF and the high coupling stability with the electrode substrate, the developed VMSF/PET-ITO sensor exhibits high signal stability during the continuous bending process. Considering high antifouling characteristic of the VMSF, direct analysis of clindamycin in human serum is realized. Because the PET-ITO electrode is easy to be miniaturized or patterned, the proposed strategy of modifying flexible electrode based on the nanochannel is expected to open up a new research field in flexible, wearable, or miniaturized sensors.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**; further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

XW, XL, and SX curated the data. FX supervised and wrote and edited the review. TZ wrote and edited the review.

FUNDING

The authors gratefully acknowledge the financial support from the National Natural Science Foundation of China (No. 81860512), and the Zhejiang Provincial Natural Science Foundation of China (LY20B050007).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fchem.2022.872582/full#supplementary-material

- Chen, X.-m., Su, B.-y., Song, X.-h., Chen, Q.-a., Chen, X., and Wang, X.-r. (2011). Recent Advances in Electrochemiluminescent Enzyme Biosensors. *Trac Trends Anal. Chem.* 30, 665–676. doi:10.1016/j.trac.2010.12.004
- Ding, J., Li, X., Zhou, L., Yang, R., Yan, F., and Su, B. (2020). Electrodeposition of Nickel Nanostructures Using Silica Nanochannels as Confinement for Low-Fouling Enzyme-free Glucose Detection. J. Mater. Chem. B 8, 3616–3622. doi:10.1039/c9tb02472g
- Girondi, N. G., Barreto., F., Pigatto, M. C., and Dalla Costa, T. (2018). Sensitive Analytical Method to Quantify Clindamycin in Plasma and Microdialysate Samples: Application in a Preclinical Pharmacokinetic Study. *J. Pharm. Biomed. Anal.* 153, 57–62. doi:10.1016/j.jpba.2018.02.005

- Wei et al.
- Habib, I. H. I., Rizk, M. S., and El-Aryan, T. R. (2011). Determination of Clindamycin in Dosage Forms and Biological Samples by Adsorption Stripping Voltammetry with Carbon Paste Electrode. *Pharm. Chem. J.* 44, 705–710. doi:10.1007/s11094-011-0548-4
- Hadi, M., and Honarmand, E. (2017). Application of Anodized Edge-Plane Pyrolytic Graphite Electrode for Analysis of Clindamycin in Pharmaceutical Formulations and Human Urine Samples. *Russ. J. Electrochem.* 53, 380–390. doi:10.1134/s1023193517040061
- Herzog, G., Sibottier, E., Etienne, M., and Walcarius, A. (2013). Electrochemically Assisted Self-Assembly of Ordered and Functionalized Mesoporous Silica Films: Impact of the Electrode Geometry and Size on Film Formation and Properties. *Faraday Discuss.* 164, 259–273. doi:10.1039/c3fd00021d
- Huang, Z., Li, Z., Chen, Y., Xu, L., Xie, Q., Deng, H., et al. (2021). Regulating Valence States of Gold Nanocluster as a New Strategy for the Ultrasensitive Electrochemiluminescence Detection of Kanamycin. *Anal. Chem.* 93, 4635–4640. doi:10.1021/acs.analchem.1c00063
- Ince, B., and Sezgintürk, M. K. (2021). A High Sensitive and Cost-Effective Disposable Biosensor for Adiponectin Determination in Real Human Serum Samples. Sens. Actuators, B 328, 129051. doi:10.1016/j.snb.2020.129051
- Jiokeng, S. L. Z., Tonle, I. K., and Walcarius, A. (2019). Amino-attapulgite/ mesoporous Silica Composite Films Generated by Electro-Assisted Self-Assembly for the Voltammetric Determination of Diclofenac. Sensors Actuators B: Chem. 287, 296–305. doi:10.1016/j.snb.2019.02.038
- Leland, J. K., and Powell, M. J. (1990). Electrogenerated Chemiluminescence: An Oxidative-Reduction Type ECL Reaction Sequence Using Tripropyl Amine. J. Electrochem. Soc. 137, 3127–3131. doi:10.1149/1.2086171
- Li, G., Belwal, T., Luo, Z., Li, Y., Li, L., Xu, Y., et al. (2021). Direct Detection of Pb2+ and Cd2+ in Juice and Beverage Samples Using PDMS Modified Nanochannels Electrochemical Sensors. *Food Chem.* 356, 129632. doi:10.1016/j.foodchem. 2021.129632
- Li, L., Chen, Y., and Zhu, J.-J. (2017). Recent Advances in Electrochemiluminescence Analysis. Anal. Chem. 89, 358–371. doi:10.1021/ acs.analchem.6b04675
- Li, X., Zhou, L., Ding, J., Sun, L., and Su, B. (2020). Platinized Silica Nanoporous Membrane Electrodes for Low-Fouling Hydrogen Peroxide Detection. *ChemElectroChem* 7, 2081–2086. doi:10.1002/celc.202000321
- Li, Z., He, Y., Klausen, L. H., Yan, N., Liu, J., Chen, F., et al. (2021). Growing Vertical Aligned Mesoporous Silica Thin Film on Nanoporous Substrate for Enhanced Degradation, Drug Delivery and Bioactivity. *Bioactive Mater.* 6, 1452–1463. doi:10.1016/j.bioactmat.2020.10.026
- Liang, X., Gou, Z., Wang, X., Wang, Y., Yue, J., Li, N., et al. (2021). Simultaneous Quantification of Cefuroxime and Clindamycin in Human Lumbar Anulus Fibrosus, Nucleus Pulposus and Serum via UPLC-MS/MS. J. Chromatogr. B 1165, 122522. doi:10.1016/j.jchromb.2021.122522
- Lin, J., Li, K., Wang, M., Chen, X., Liu, J., and Tang, H. (2020). Reagentless and Sensitive Determination of Carcinoembryonic Antigen Based on a Stable Prussian Blue Modified Electrode. RSC Adv. 10, 38316–38322. doi:10.1039/ d0ra06751b
- Liu, X., Li, H., Zhou, H., Liu, J., Li, L., Liu, J., et al. (2020). Direct Electrochemical Detection of 4-aminophenol in Pharmaceuticals Using ITO Electrodes Modified with Vertically-Ordered Mesoporous Silica-Nanochannel Films. J. Electroanalytical Chem. 878, 114568. doi:10.1016/j. jelechem.2020.114568
- Liu, Y.-M., Shi, Y.-M., Liu, Z.-L., and Peng, L.-F. (2010). Sensitive Determination of Tilmicosin, Erythromycin Ethylsuccinate and Clindamycin by CE with Electrochemiluminescence Detection Using Azithromycin as Internal Standard and its Applications. J. Sep. Sci. 33, 1305–1311. doi:10.1002/jssc. 200900843
- Ma, C., Cao, Y., Gou, X., and Zhu, J.-J. (2019). Recent Progress in Electrochemiluminescence Sensing and Imaging. *Anal. Chem.* 92, 431–454. doi:10.1021/acs.analchem.9b04947
- Ma, C., Wu, S., Zhou, Y., Wei, H. F., Zhang, J., Chen, Z., et al. (2021). Bio-Coreactant-Enhanced Electrochemiluminescence Microscopy of Intracellular Structure and Transport. Angew. Chem. Int. Ed. 60, 4907–4914. doi:10.1002/ anie.202012171
- Ma, X., Liao, W., Zhou, H., Tong, Y., Yan, F., Tang, H., et al. (2020). Highly Sensitive Detection of Rutin in Pharmaceuticals and Human Serum Using ITO Electrodes Modified with Vertically-Ordered Mesoporous Silica-Graphene

Nanocomposite Films. J. Mater. Chem. B 8, 10630-10636. doi:10.1039/ d0tb01996h

- Mi, X., Li, H., Tan, R., and Tu, Y. (2020). Dual-modular Aptasensor for Detection of Cardiac Troponin I Based on Mesoporous Silica Films by Electrochemiluminescence/electrochemical Impedance Spectroscopy. Anal. Chem. 92, 14640–14647. doi:10.1021/acs.analchem.0c03130
- Mohammad Beigi, S., Mesgari, F., and Hosseini, M. (2020). Detection of Clindamycin in Pharmaceutical Products Using an Electrochemiluminescence Electrode Based on a Composite of Ru(bpy)2³⁺, Eu₂O₃ Nanoparticle and Chitosan. *Anal. Bioanal. Electrochem.* 12, 733–746.
- Nasir, T., Herzog, G., Hébrant, M., Despas, C., Liu, L., and Walcarius, A. (2018). Mesoporous Silica Thin Films for Improved Electrochemical Detection of Paraquat. ACS Sens. 3, 484–493. doi:10.1021/acssensors.7b00920
- Paul, P., Duchateau, T., Sänger-van de Griend, C., Adams, E., and Van Schepdael, A. (2017). Capillary Electrophoresis with Capacitively Coupled Contactless Conductivity Detection Method Development and Validation for the Determination of Azithromycin, Clarithromycin, and Clindamycin. J. Sep. Sci. 40, 3535–3544. doi:10.1002/jssc.201700560
- Qiu, G., Han, Y., Zhu, X., Gong, J., Luo, T., Zhao, C., et al. (2021). Sensitive Detection of Sulfide Ion Based on Fluorescent Ionic Liquid–Graphene Quantum Dots Nanocomposite. *Front. Chem.* 9, 316. doi:10.3389/fchem. 2021.658045
- Ullah, W., Herzog, G., Vilà, N., and Walcarius, A. (2021). Electrografting and Electropolymerization of Nanoarrays of PANI Filaments through Silica Mesochannels. *Electrochemistry Commun.* 122, 106896. doi:10.1016/j.elecom. 2020.106896
- Walcarius, A. (2021). Electroinduced Surfactant Self-Assembly Driven to Vertical Growth of Oriented Mesoporous Films. Acc. Chem. Res. 54, 3563–3575. doi:10. 1021/acs.accounts.1c00233
- Wang, F., Liu, Y., Fu, C., Li, N., Du, M., Zhang, L., et al. (2021). Paper-based Bipolar Electrode Electrochemiluminescence Platform for Detection of Multiple miRNAs. Anal. Chem. 93, 1702–1708. doi:10.1021/acs.analchem.0c04307
- Wang, J., Peng, Z., Yang, J., Wang, X., and Yang, N. (2008). Detection of Clindamycin by Capillary Electrophoresis with an End-Column Electrochemiluminescence of tris(2,2'-bypyridine)Ruthenium(II). *Talanta* 75, 817–823. doi:10.1016/j.talanta.2007.12.019
- Wang, M., Lin, J., Gong, J., Ma, M., Tang, H., Liu, J., et al. (2021). Rapid and Sensitive Determination of Doxorubicin in Human Whole Blood by Vertically-Ordered Mesoporous Silica Film Modified Electrochemically Pretreated Glassy Carbon Electrodes. RSC Adv. 11, 9021–9028. doi:10.1039/d0ra10000e
- Wang, Y., Zhao, G., Chi, H., Yang, S., Niu, Q., Wu, D., et al. (2020). Selfluminescent Lanthanide Metal-Organic Frameworks as Signal Probes in Electrochemiluminescence Immunoassay. J. Am. Chem. Soc. 143, 504–512. doi:10.1021/jacs.0c12449
- Wong, A., Razzino, C. A., Silva, T. A., and Fatibello-Filho, O. (2016). Square-wave Voltammetric Determination of Clindamycin Using a Glassy Carbon Electrode Modified with Graphene Oxide and Gold Nanoparticles within a Crosslinked Chitosan Film. Sensors Actuators B: Chem. 231, 183–193. doi:10.1016/j.snb. 2016.03.014
- Xiao, Y., Chen, S., Zhang, S., Wang, G., Yi, H., Xin, G.-Z., et al. (2021). Mesoporous Silica-Mediated Controllable Electrochemiluminescence Quenching for Immunosensor with Simplicity, Sensitivity and Tunable Detection Range. *Talanta* 231, 122399. doi:10.1016/j.talanta.2021.122399
- Yan, F., Ma, X., Jin, Q., Tong, Y., Tang, H., Lin, X., et al. (2020). Phenylboronic Acid-Functionalized Vertically Ordered Mesoporous Silica Films for Selective Electrochemical Determination of Fluoride Ion in Tap Water. *Mikrochim Acta* 187, 470–478. doi:10.1007/s00604-020-04422-4
- Yan, F., Chen, J., Jin, Q., Zhou, H., Sailjoi, A., Liu, J., et al. (2020). Fast One-step Fabrication of a Vertically-Ordered Mesoporous Silica-Nanochannel Film on Graphene for Direct and Sensitive Detection of Doxorubicin in Human Whole Blood. J. Mater. Chem. C 8, 7113–7119. doi:10.1039/d0tc00744g
- Yan, F., Luo, T., Jin, Q., Zhou, H., Sailjoi, A., Dong, G., et al. (2021). Tailoring Molecular Permeability of Vertically-Ordered Mesoporous Silica-Nanochannel Films on Graphene for Selectively Enhanced Determination of Dihydroxybenzene Isomers in Environmental Water Samples. J. Hazard. Mater. 410, 124636. doi:10.1016/j.jhazmat.2020.124636
- Yan, F., Wang, M., Jin, Q., Zhou, H., Xie, L., Tang, H., et al. (2021). Verticallyordered Mesoporous Silica Films on Graphene for Anti-fouling

Electrochemical Detection of Tert-Butylhydroquinone in Cosmetics and Edible Oils. *J. Electroanalytical Chem.* 881, 114969. doi:10.1016/j.jelechem.2020. 114969

- Yang, X.-y., Wang, Y.-z., Wang, L.-l., Zhu, J. W., Zhao, J., Zong, H.-l., et al. (2021). Bipolar Electrode Ratiometric Electrochemiluminescence Biosensing Analysis Based on boron Nitride Quantum Dots and Biological Release System. *Biosens. Bioselectron.* 191, 113393. doi:10.1016/j.bios.2021.113393
- Yanyan, Z., Lin, J., Xie, L., Tang, H., Wang, K., and Liu, J. (2021). Onestep Preparation of Nitrogen-Doped Graphene Quantum Dots with Anodic Electrochemiluminescence for Sensitive Detection of Hydrogen Peroxide and Glucose. *Front. Chem.* 9, 394. doi:10.3389/fchem. 2021.688358
- Zhai, Q., Li, J., and Wang, E. (2017). Recent Advances Based on Nanomaterials as Electrochemiluminescence Probes for the Fabrication of Sensors. *ChemElectroChem* 4, 1639–1650. doi:10.1002/celc.201600898
- Zhang, Y., Xu, J., Zhou, S., Zhu, L., Lv, X., Zhang, J., et al. (2020). DNAzyme-Triggered Visual and Ratiometric Electrochemiluminescence Dual-Readout Assay for Pb(II) Based on an Assembled Paper Device. Anal. Chem. 92, 3874–3881. doi:10.1021/acs.analchem.9b05343
- Zhou, L., Cheng, C., Li, X., Ding, J., Liu, Q., and Su, B. (2020). Nanochannel Templated Iridium Oxide Nanostructures for Wide-Range pH Sensing from Solutions to Human Skin Surface. *Anal. Chem.* 92, 3844–3851. doi:10.1021/acs. analchem.9b05289
- Zhou, L., Hou, H., Wei, H., Yao, L., Sun, L., Yu, P., et al. (2019). *In Vivo* monitoring of Oxygen in Rat Brain by Carbon Fiber Microelectrode Modified with Antifouling Nanoporous Membrane. *Anal. Chem.* 91, 3645–3651. doi:10. 1021/acs.analchem.8b05658

- Zhou, P., Yao, L., Chen, K., and Su, B. (2020). Silica Nanochannel Membranes for Electrochemical Analysis and Molecular Sieving: a Comprehensive Review. *Crit. Rev. Anal. Chem.* 50, 424–444. doi:10.1080/10408347.2019.1642735
- Zhou, P., Yao, L., and Su, B. (2019). Fabrication, Characterization, and Analytical Application of Silica Nanopore Array-Modified Platinum Electrode. ACS Appl. Mater. Inter. 12, 4143–4149. doi:10.1021/acsami.9b20165
- Zhu, L., Lv, X., Li, Z., Shi, H., Zhang, Y., Zhang, L., et al. (2021). All-sealed Paper-Based Electrochemiluminescence Platform for On-Site Determination of lead Ions. *Biosens. Bioelectron.* 192, 113524. doi:10.1016/j.bios.2021.113524

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Wei, Luo, Xu, Xi and Zhao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.