

Research Article

Intraperitoneal Exposure to Nano/Microparticles of Fullerene (C₆₀) Increases Acetylcholinesterase Activity and Lipid Peroxidation in Adult Zebrafish (*Danio rerio*) Brain

Gonzalo Ogliari Dal Forno,¹ Luiza Wilges Kist,^{1,2} Mariana Barbieri de Azevedo,¹ Rachel Seemann Fritsch,¹ Talita Carneiro Brandão Pereira,¹ Roberta Socoowski Britto,^{3,4} Sílvia Stanisçuaski Guterres,⁵ Irene Cledes Külkamp-Guerreiro,⁵ Carla Denise Bonan,^{2,6} José María Monserrat,^{3,4,7} and Maurício Reis Bogo^{1,2}

¹ Laboratório de Biologia Genômica e Molecular, Faculdade de Biociências, Pontifícia Universidade Católica do Rio Grande do Sul, Avenida Ipiranga 6681 Sala 134, 90619-900 Porto Alegre, RS, Brazil

² Instituto Nacional de Ciência e Tecnologia Translacional em Medicina (INCT-TM), 90035-003 Porto Alegre, RS, Brazil

³ Universidade Federal do Rio Grande-FURG, Instituto de Ciências Biológicas (ICB), Avenida Itália Km 8 s/n, 96208-900 Rio Grande, RS, Brazil

⁴ Programa de Pós-Graduação em Ciências Fisiológicas-Fisiologia Animal Comparada, FURG, Avenida Itália Km 8 s/n, 96208-900 Rio Grande, RS, Brazil

⁵ Faculdade de Farmácia, Universidade Federal do Rio Grande do Sul, Avenida Ipiranga 2752, 90610-000 Porto Alegre, RS, Brazil

⁶ Laboratório de Neuroquímica e Psicofarmacologia, Faculdade de Biociências, Pontifícia Universidade Católica do Rio Grande do Sul, Avenida Ipiranga 6681, 90619-900 Porto Alegre, RS, Brazil

⁷ Instituto Nacional de Ciência e Tecnologia de Nanomateriais de Carbono, 31270-901 Belo Horizonte, MG, Brazil

Correspondence should be addressed to Maurício Reis Bogo; mbogo@pucrs.br

Received 5 April 2013; Accepted 29 May 2013

Academic Editor: Qaisar Mahmood

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Even though technologies involving nano/microparticles have great potential, it is crucial to determine possible toxicity of these technological products before extensive use. Fullerenes C₆₀ are nanomaterials with unique physicochemical and biological properties that are important for the development of many technological applications. The aim of this study was to evaluate the consequences of nonphotoexcited fullerene C₆₀ exposure in brain acetylcholinesterase expression and activity, antioxidant responses, and oxidative damage using adult zebrafish as an animal model. None of the doses tested (7.5, 15, and 30 mg/kg) altered AChE activity, antioxidant responses, and oxidative damage when zebrafish were exposed to nonphotoexcited C₆₀ nano/microparticles during 6 and 12 hours. However, adult zebrafish exposed to the 30 mg/kg dose for 24 hours have shown enhanced AChE activity and augmented lipid peroxidation (TBARS assays) in brain. In addition, the up-regulation of brain AChE activity was neither related to the transcriptional control (RT-qPCR analysis) nor to the direct action of nonphotoexcited C₆₀ nano/microparticles on the protein (*in vitro* results) but probably involved a posttranscriptional or posttranslational modulation of this enzymatic activity. Taken together these findings provided further evidence of toxic effects on brain after C₆₀ exposure.

1. Introduction

Nanoscience investigates nanoscale phenomena and serves as the foundation for nanotechnology, which develops practical

applications for nanomaterials (particles typically with sizes in the 1–100 nm range but not exclusively) [1]. Due to their composition, small size, and shape, nanomaterials exhibit novel properties for diverse applications that have already

yielded in a variety of commercially available products [2]. As a consequence, it is expected that both humans and environmental systems will be increasingly exposed to nanomaterials in the next years. Nanotoxicology [3] and nanoecotoxicology [4] are emerging disciplines that arose to address the potential human and environmental health effects of nanomaterials' exposure.

Fullerenes are molecular materials that call attention after the first preparation of C_{60} which is an allotrope of carbon consisting of 60 carbon atoms joined to form a cage-like structure [5]. C_{60} exhibits unique physical and chemical properties for many technological applications, such as electronics, superconductors, cosmetics [6], and, more recently, drug and gene delivery [7]. Although the toxic effects of C_{60} are still mostly unknown, especially those related to neurotoxicity, some general aspects have already been addressed. For instance, C_{60} is reported to be cytotoxic to various mammalian cells [8] and it has been shown to induce lipid peroxidation in human cells [9], in brain of juvenile largemouth bass (*Micropterus salmoides*) [10], and in common carp (*Cyprinus carpio*) [11]. Moreover, C_{60} was classified as "very toxic" to aquatic organisms by the grid for the potential ecotoxicological hazard evaluation which is based mainly on toxicity to fish, *Daphnia*, and algae and in data about degradability of the substance [4].

In cholinergic neurotransmission, choline acetyltransferase (ChAT) is responsible for the synthesis of acetylcholine (ACh) in the presynaptic neuron. After its release into the synaptic cleft, ACh promotes the activation of metabotropic muscarinic and ionotropic nicotinic cholinergic receptors. The reaction responsible for the maintenance of levels of ACh is catalyzed by two cholinesterases (ChE): acetylcholinesterase (AChE) (E.C. 3.1.1.7) and butyrylcholinesterase (BuChE) (E.C. 3.1.1.8) [12]. The zebrafish (*Danio rerio*) has long been considered a powerful animal model for studying several biological events. More recently, zebrafish become also a valuable model to environmental and toxicological studies [13]. It has been demonstrated that BuChE is not encoded in the zebrafish genome, but AChE is encoded by a single gene that has been cloned, sequenced, and functionally detected in zebrafish brain [14].

AChE activity has been widely used as a bioindicator of environmental exposure to neurotoxic pollutants. For example, the inhibition of AChE as a biomarker for assessment of the exposure of organisms to organophosphate and carbamate insecticides is well known [15]. The inhibition of brain AChE activity of aquatic species by toxic substances such as methanol [16], heavy metals mercury, and lead [17], neurotoxins [18], and organochlorine pesticide endosulfan [19] also has been well established. On the other hand, AChE activation has also been demonstrated as a consequence of exposure to neurotoxic compounds such as aluminum [20] and ethanol [21].

Therefore, considering that (i) both humans and environmental systems will be increasingly exposed to C_{60} in a near future, (ii) the neurotoxic effects of C_{60} are far from being completely understood, (iii) measurement of AChE activity in organisms is used as a biomarker of environmental

contamination due to neurotoxic substances, (iv) determination of oxidative stress parameters is recognized as a tool to evaluate toxicity mediated by small particles exposure, and (v) zebrafish is a well-established organism for toxicological analysis, the aim of the present study was to evaluate the nonphotoexcited C_{60} effects in brain AChE activity and its gene expression pattern. Furthermore, we have analyzed the effects of the nonphotoexcited C_{60} in parameters related to antioxidant defenses and lipid peroxidation in the brain of zebrafish.

2. Materials and Methods

2.1. Chemicals. Fullerene (C_{60} , 99.5% purity) was purchased from Aldrich (Milwaukee, WI, USA); DMSO was purchased from Fisher Scientific (Pittsburgh, PA, USA) and Trizma Base; ethylenediaminetetraacetic acid (EDTA), ethylene glycol bis (beta aminoethylether)-N,N,N',N'-tetraacetic acid (EGTA), sodium citrate, Coomassie Blue G, bovine serum albumin, acetylthiocholine, 5,5'-dithiobis-2-nitrobenzoic acid (DTNB) HEPES, BHT (99%), 2,2'-azobis (2-methylpropionamide) dihydrochloride (ABAP), and 1,1,3,3-tetramethoxypropane were purchased from Sigma Chemical Co. (St. Louis, MO, USA). KCl and SDS (90%) were purchased from Labsynth (Brazil). Tetramethoxypropane (TMP) and 2',7'-dichlorodihydrofluorescein diacetate were purchased from Acros Organics (Morris Plains, NJ, USA) and Molecular Probes Inc. (Eugene, OR, USA) respectively. $MgCl_2$ and Acetic acid 99.7% were purchased from Isofar and Vetec (Brazil), respectively. TRIzol reagent, Platinum Taq DNA Polymerase, and SYBR Green I were purchased from Invitrogen (Carlsbad, CA, USA). ImProm-II Reverse Transcription System was purchased from Promega (Madison, WI, USA). All other reagents used were of analytical grade.

2.2. Animals. Adult wild-type zebrafish (*Danio rerio*, Cyprinidae) of both sexes (6–9 months old) were obtained from a specialized supplier (Redfish Agroloja, RS, Brazil). Animals were kept at a density of up to five animals per liter in 50 L housing tanks with tap water that was previously treated with Tetra's AquaSafe (to neutralize chlorine, chloramines, and heavy metals present in the water that could be harmful to fish) and continuously aerated (7.20 mg O_2 /L) at $26 \pm 2^\circ C$, under a 14/10 h light/dark controlled photoperiod. Animals were acclimated for at least two weeks before the experiments and were fed three times a day with TetraMin Tropical Flake fish food. The fish were maintained healthy and free of any signs of disease and were used according to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health. All procedures in the present study were approved by the Animal Ethics Committee of the Pontifical Catholic University of Rio Grande do Sul (PUCRS), protocol number 10/00185-CEUA.

2.3. C_{60} Suspension. Suspensions of C_{60} in DMSO were prepared as previously described [23, 24] with modifications.

Briefly, 7.6 mg of C_{60} was added to 0.5 mL of DMSO and sonicated for 3 h. The concentrated suspension of C_{60} obtained was diluted in water to result in suspensions that when intraperitoneally injected (mean injection volume was 10 μ L) get the doses of 7.5, 15, and 30 mg/kg fish (12.5% DMSO). For this reason, the C_{60} suspensions were named as 7.5, 15, and 30. The doses and times of exposure were selected based on previous studies where multiwalled carbon nanotubes were intravenously injected in mice [25, 26], and functionalized single-walled carbon nanotubes were intraperitoneally injected in rats [27]. C_{60} suspensions were further sonicated for one hour prior to use. To avoid C_{60} 's photoexcitation, the C_{60} suspensions were prepared and stored in a dark condition.

2.4. Characterization of C_{60} Suspensions. The C_{60} suspensions (7.5, 15, and 30) were characterized in terms of particle size distribution [28]. The mean diameter over the volume and number distribution ($d_{4,3}$) was determined by laser diffractometry (Mastersizer 2000, Malvern Instruments, UK). The value of SPAN was utilized to determine particle size distribution according to (1), where $d_{0.9}$, $d_{0.1}$, and $d_{0.5}$ are the particle diameters determined at 90%, 10%, and 50% cumulative undersized volumes, respectively,

$$\text{SPAN} = \frac{d_{0.9} - d_{0.1}}{d_{0.5}}. \quad (1)$$

2.5. Animal Procedures. Intraperitoneal (i. p.) injection was adopted as administration route in the *in vivo* protocols (i) to avoid the photoexcitation of the C_{60} material and (ii) to ensure that exposure concentrations are in line with target values. Intraperitoneal injections were conducted using a 3/10-mL U-100 BD ultra-fine short insulin syringe 8 mm (5/16") \times 31G short needle (Becton Dickinson and Company, NJ, USA) according to the protocol established by Phelps and colleagues [29]. Briefly, the volume injected into the animal (mean injection volume was 10 μ L) was adjusted to the fish bodyweight (mean mass of the animals was 0.75 g \pm 0.06 g/mean \pm SEM) to achieve 7.5, 15, and 30 mg/kg based on a previous study [30]. The animals of the control group received the same volume of saline solution, and the animals of the vehicle control received the same volume of 12.5% DMSO. Anesthesia of the animals prior to the injection was obtained by its immersion in a solution of benzocaine (1 mM in MeOH 1%) until the animal showed a lack of motor coordination and reduced respiratory rate. The anesthetized animal was gently placed in a water-soaked gauze-wrapped hemostat with the abdomen facing up and the head of the fish positioned at the hinge of the hemostat (the pectoral fins were used as a landmark on the abdomen). The needle was inserted parallel to the spine in the midline of the abdomen posterior to the pectoral fins. The injection procedure was conducted in such a way as to guarantee that the animal did not spend more than 10 s out of the water. After the injection, the animals were placed in a separate tank with highly aerated nonchlorinated tap water (25 \pm 2°C) to facilitate recovery from the anesthesia. Saline solution was used as control. All the animals that recovered within 2-3 min following the injection continued

in the experiment, while animals that did not recover during this period were discarded. Six, twelve, or twenty-four hours after the injection, the animals were euthanized.

2.6. In Vitro Assays of AChE Activity. *In vitro* assays were performed as previously described [31, 32] in order to evaluate if 7.5, 15, and 30 C_{60} suspensions might have a direct effect on the enzyme. Briefly, 33 μ L of C_{60} suspensions were added to the reaction medium before the preincubation with the enzyme-containing lysate from zebrafish brain homogenate and maintained during the enzyme assays. Control treatments with equal volume of vehicle (DMSO 12.5%) were performed to exclude the DMSO effect on the enzyme activities.

2.7. Determination of AChE Activity. Zebrafish were euthanized, and their whole brains were removed by dissection. The brains (two whole brains for each sample) were homogenized on ice in 60 volumes (v/w) of Tris-citrate buffer (50 mM Tris, 2 mM EDTA, 2 mM EGTA, pH 7.4, adjusted with citric acid), in a glass-Teflon homogenizer. The rate of acetylthiocholine hydrolysis (ACSch, 0.88 mM) was assessed in a final volume of 300 μ L with 11 mM phosphate buffer, pH 7.5, and 0.22 mM DTNB using a method previously described [33]. Before the addition of substrate, samples containing protein (5 μ g) and the reaction medium described above were preincubated for 10 min at 25°C. The hydrolysis of substrate was monitored by the formation of thiolate dianion of DTNB at 412 nm for 2-3 min (intervals of 30 s) in a microplate reader. Controls without the homogenate preparation were performed in order to determine the nonenzymatic hydrolysis of the substrate. The linearity of absorbance against time and protein concentration was previously determined. The AChE activity was expressed as micromoles of thiocholine (SCh) released per hour per milligram of protein. All enzyme assays were evaluated in triplicate, and at least four independent experiments were performed.

2.8. Gene Expression Analysis by Quantitative Real-Time RT-PCR (RT-qPCR). Gene expression analysis was carried out only when kinetic alteration occurred. For this reason, immediately after 24 hours of intraperitoneal injection (C_{60} suspension 30), the animals were euthanized by decapitation. For each sample, a pool of three zebrafish whole brains was used. Total RNA was isolated with Trizol reagent (Invitrogen, Carlsbad, CA, USA) in accordance with the manufacturer's instructions. The total RNA was quantified by spectrophotometry and the cDNA was synthesized with ImProm-II Reverse Transcription System (Promega) from 1 μ g total RNA, following the manufacturer's instructions. Quantitative PCR was performed using SYBR Green I (Invitrogen) to detect double-strand cDNA synthesis. Reactions were done in a volume of 25 μ L using 12.5 μ L of diluted cDNA (1:100 for *Ef1 α* and *Rpl13 α* ; and 1:20 for *ache*), containing a final concentration of 0.2x SYBR Green I (Invitrogen), 100 μ M dNTP, 1x PCR Buffer, 3 mM MgCl₂, 0.25 U Platinum Taq DNA Polymerase (Invitrogen), and 200 nM of each reverse and forward primers (Table 1). The PCR cycling conditions

TABLE 1: PCR primers design.

Proteins	Primer sequences (5'-3')	Accession number (mRNA)
<i>EF1α</i> *	F: CTGGAGGCCAGCTCAAACAT R: ATCAAGAAGAGTAGTACCGCTAGCATTAC	NSDART0000023156
<i>Rpl13α</i> *	F: TCTGGAGGACTGTAAGAGGTATGC R: AGACGCACAATCTTGAGAGCAG	NM_212784
<i>ache</i> **	F: GCTAATGAGCAAAAAGCATGTGGGCTTG R: TATCTGTGATGTTAAGCAGACGAGGCAGG	NM_131846

*According to Tang et al. [22].

**Designed by the authors.

were an initial polymerase activation step for 5 min at 95°C, 40 cycles of 15 s at 95°C for denaturation, 35 s at 60°C for annealing, and 15 s at 72°C for elongation. At the end of cycling protocol, a melting curve analysis was included and fluorescence measured from 60 to 99°C. Relative expression levels were determined with 7500 Fast Real-Time System Sequence Detection Software v.2.0.5 (Applied Biosystems). The efficiency per sample was calculated using LinRegPCR 11.0 Software (<http://LinRegPCR.nl>) and the stability of the references genes, *EF1 α* and *Rpl13 α* (*M-value*), and the optimal number of reference genes according to the pairwise variation (*V*) were analyzed by GeNorm 3.5 Software (<http://medgen.ugent.be/genorm/>). Relative RNA expression levels were determined using the $2^{-\Delta\Delta CT}$ method.

2.9. Antioxidant Capacity against Peroxyl Radicals. Total antioxidant capacity against peroxyl radicals was performed according Amado and colleagues [34], employing the thermal decomposition of 2,2'-azobis 2 methylpropionamide dihydrochloride (ABAP; 4 mM) as peroxyl radical generator. Reactive oxygen species (ROS) concentration was detected with the fluorescent probe 2',7'-dichlorofluorescein diacetate (H₂DCF-DA) in a final concentration of 40 μ M (480 and 525 nm forexcitation and emission, resp.). The relative difference between ROS area with and without ABAP was considered a measure of antioxidant capacity, with high area difference meaning low antioxidant capacity, since high fluorescence levels were obtaining after adding ABAP, meaning low competence to neutralize peroxyl radicals.

2.10. Measurement of Lipid Peroxidation. Lipid peroxidation was measured through the determination of thiobarbituric acid reactive substances (TBARS), following the methodology of Oakes and van der Kraak [35]. Brain homogenates (10 μ L) were added to a reaction mixture made with 150 μ L of 20% acetic acid, 150 μ L of thiobarbituric acid (2.4%), 50 μ L of Milli-Q water and 20 μ L of sodium dodecyl sulfate (SDS, 8.1%). Samples were heated at 95°C during 30 min, and after cooling by 10 min, 100 μ L of Milli-Q water and 500 μ L of *n*-butanol were added. After centrifugation (3,000 \times g during 10 min at 15°C), the organic phase (150 μ L) was placed in a microplate reader, and the fluorescence registered after excitation at 520 nm and emission of 580 nm. The concentration of TBARS (nmol/mg of wet tissue) was calculated employing tetramethoxypropane (TMP) as standard.

2.11. Protein Determination. Protein was measured by the Coomassie blue method [36] using bovine serum albumin as standard.

2.12. Statistical Analysis. AChE activity and antioxidant analyses were expressed as means \pm S.E.M. and analyzed by one-way analysis of variance (ANOVA). Post hoc comparisons were made using Tukey's test and orthogonal comparisons. Before ANOVA, its assumptions (normality and variances homogeneity) were checked. Molecular data were expressed as means \pm S.E.M. and analyzed by Student's *t*-test. In every case, the significance level was fixed in 5% ($\alpha = 0.05$).

3. Results

3.1. C₆₀ Suspensions of Nano/Microparticles. Although DMSO is known to show low toxicity by itself [24, 37], appropriate experimental controls must be employed to eliminate its influence. In this study, the DMSO was diluted to result in 12.5% DMSO as a final concentration. Any signal of toxicity, that is, mortality or even transient alterations in behavior, was observed in the vehicle control group (12.5% DMSO). In addition, control group (saline) and vehicle control group were never statistically different in the conditions tested.

The nano/microparticles mean diameters over the volume showed wide distributions (60 nm–316 μ m; 69 nm–1,905 μ m; 182 nm–208 μ m, for suspensions 7.5, 15, and 30, resp.), with most abundant sizes in the micrometric range (Figures 1(a), 1(c), and 1(e)). The SPAN values increased with the dilution of suspensions (5.843, 3.003, and 1.607 to suspensions 7.5, 15, and 30, resp.), showing a narrow size distribution with lower dilutions. On the other hand, it is important to emphasize that when considering the mean diameters over the number distribution, we observed that the greater part of particles were under nanometric sizes (Figures 1(b), 1(d), and 1(f)).

3.2. Acetylcholinesterase Enzymatic Activity and Gene Expression. The effect of different C₆₀ concentrations and times of exposure on brain AChE activity was demonstrated by performing (*in vivo*) experiments using adult zebrafish. None of the concentrations tested (7.5, 15, and 30 mg/kg) altered AChE activity when zebrafish were exposed to C₆₀ during 6 hours (Figure 2(a)) and 12 hours (Figure 2(b)).

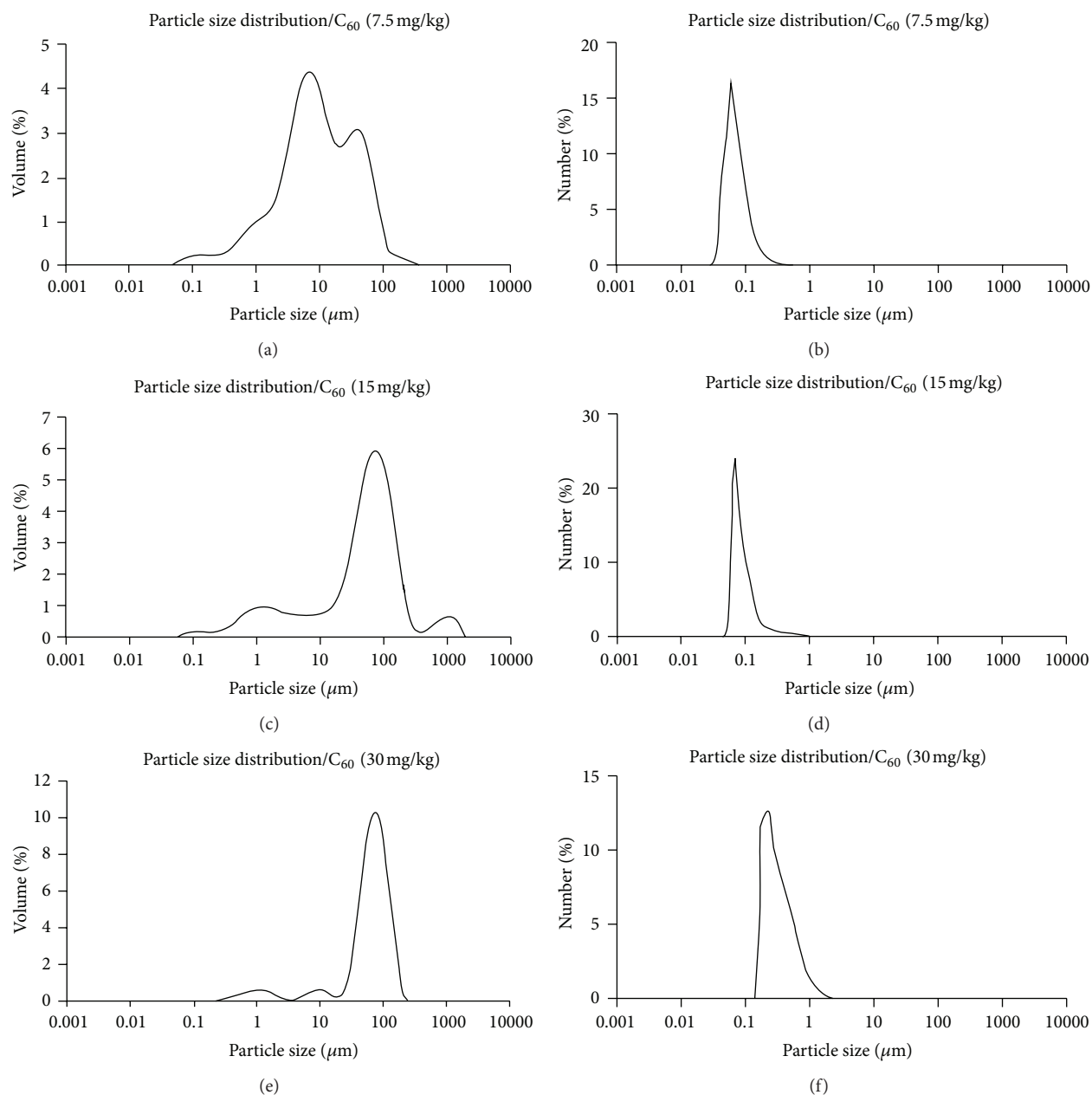


FIGURE 1: Size distribution in suspensions of nano/microparticles. Mean diameters distribution of fullerene C_{60} suspensions (7.5, 15, and 30) was determined over the volume ((a), (c), and (e)) and over the number ((b), (d), and (f)) of the nano/microparticles.

However, the analysis for 24 hours demonstrated that animals treated with the concentration of 30 mg/kg presented a significant increase in AChE activity ($28.54 \pm 3.72 \mu\text{mol SCh}\cdot\text{h}^{-1}\cdot\text{mg protein}^{-1}$; $P = 0.0001$) when compared to saline ($12.19 \pm 0.55 \mu\text{mol SCh}\cdot\text{h}^{-1}\cdot\text{mg protein}^{-1}$; $P = 0.0001$) and to the vehicle control group ($15.46 \pm 0.57 \mu\text{mol SCh}\cdot\text{h}^{-1}\cdot\text{mg protein}^{-1}$; $P = 0.0001$) (Figure 2(c)). The upregulation of brain AChE activity after exposure to C_{60} (30 mg/kg for 24 hours) could be a consequence of transcriptional control. In order to determine if transcriptional regulation of AChE gene has occurred, a RT-qPCR analysis was performed. The results have shown that AChE

transcript levels were not enhanced when compared to the vehicle control group ($P = 0.6695$; Figure 3) suggesting that the activation of brain AChE is not directly related with the transcriptional control.

3.3. In Vitro Effects of C_{60} Suspensions on Acetylcholinesterase Activity. To verify whether C_{60} nano/microparticles might have a direct effect on the enzyme, we tested the *in vitro* effect of C_{60} suspensions on AChE activity in zebrafish brain. The results showed that C_{60} suspensions did not bring about any alteration in AChE activity ($P = 0.7701$; Figure 4).

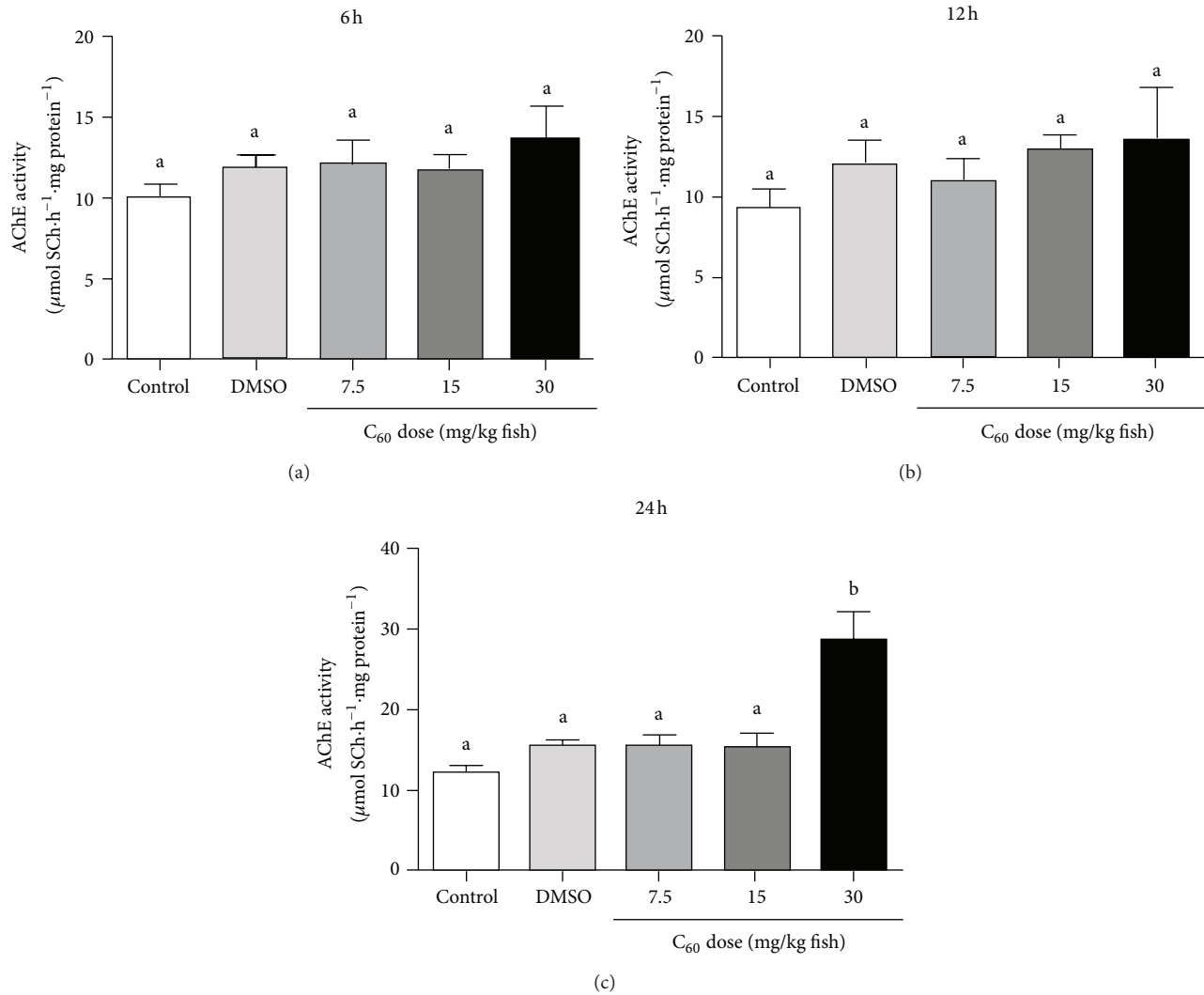


FIGURE 2: *In vivo* AChE activity in zebrafish brain after 06 (a), 12 (b), and 24 hours (c) of fullerene C₆₀ exposure at distinct concentrations (7.5–30 mg/kg fish). Bars represent the mean \pm SEM of at least three different experiments, each one performed in triplicate. The specific enzyme activity is reported as micromoles of thiocholine released per hour per milligram of protein. Bars represent the mean \pm SEM of at least three independent experiments, each one performed in triplicate. Different letters indicate significant differences ($P < 0.05$) between groups.

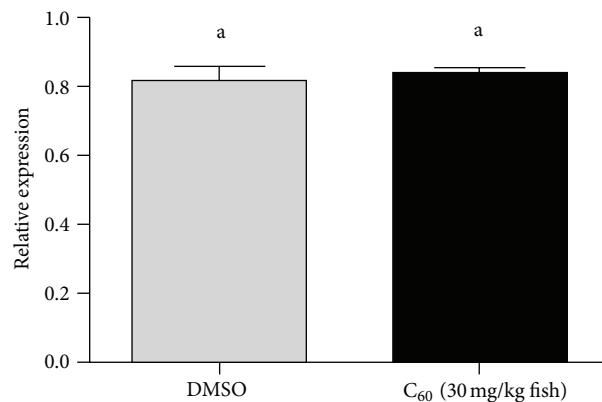


FIGURE 3: RT-qPCR analysis. Relative *ache* expression profile after fullerene C₆₀ exposure (30 mg/kg fish for 24 hours) on zebrafish brain. Bars represent the mean \pm SEM.

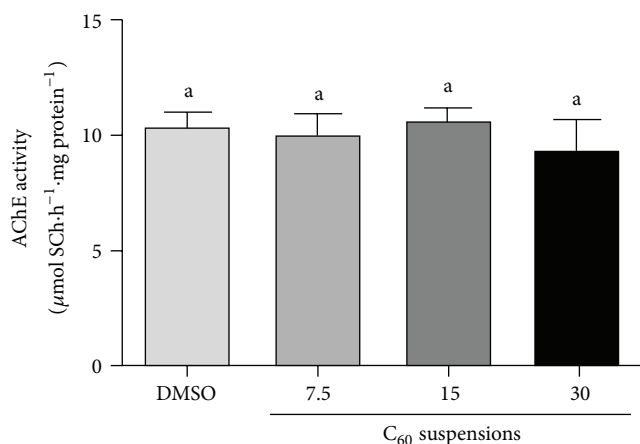


FIGURE 4: *In vitro* AChE activity. *In vitro* effect of different C₆₀ suspensions (7.5, 15, and 30) on ACh hydrolysis in zebrafish brain. Bars represent the mean ± SEM of at least three different experiments, each one performed in triplicate.

3.4. Antioxidant Analysis. The total antioxidant competence against peroxy radicals showed an augmented response (lower relative area) in brains of zebrafish exposed to C₆₀ (15 mg/kg) for 6 hours when compared to zebrafish exposed to C₆₀ (7.5 and 30 mg/kg) for 6 hours ($P = 0.0209$; Figure 5(a)). No other differences were observed under the experimental conditions (Figures 5(b) and 5(c)). Oxidative damage, measured by lipid peroxidation (TBARS assays), showed a pro-oxidant condition elicited by C₆₀ at the highest dose (30 mg/kg) after 24 hours ($P = 0.0194$; Figures 6(a), 6(b), and 6(c)).

4. Discussion

Although technologies evolving nano/microparticles have considerable potential in diverse applications, it is crucial to determine possible toxicity of these technological products before extensive use. Little is known about the toxic effects of fullerenes in brain. At present, only few studies presenting contradictory findings have evaluated possible neurotoxic effects of fullerenes exposure. For instance, it was already suggested that C₆₀ did not cross the blood-brain barrier [38], whereas the results obtained by Mokrushin [39] suggested that fullerenes possess marked neurotropic properties and are neurotoxic substances irreversibly blocking the electrical activity of the nervous tissue.

Neurotoxicity of C₆₀ in fish species has been previously reported [11, 40, 41]. Generation of reactive oxygen species (ROS) by C₆₀ is influenced in part by the presence and type of illumination due to the photoexcitation of C₆₀ by UV and visible light [42] or even to by-products of the organic solvents employed to prepare C₆₀ suspensions [40]. For this reason, the C₆₀ suspensions were prepared under the protection of light, and i.p. injection was adopted as administration route in the *in vivo* protocols, an experimental condition that avoids the influence of light in the analyzed variables. Also, *in vitro* assays were run in darkness.

The characterization of the size and stability of C₆₀ nanoparticles in suspension is very important to evaluate their toxicity once particle size can change during the preparation of the suspension, dilution, and exposure [11]. In this study, the nano/microparticles mean diameters over the volume in the C₆₀ suspensions showed wide distributions with most abundant sizes in the micrometric range. The C₆₀ has a tendency to form aggregates very easily [7], and this may be a possible cause of this wide distribution. In contrast, the nano/microparticles mean diameters over the number of distribution in the C₆₀ suspensions demonstrated that the greater part of particles was under nanometric sizes. Totsuka and colleagues [43] also observed wide distributions by dynamic light scattering in formulations manufactured with C₆₀. Take into account that particle sizes limit their ability to translocate to different tissues [3, 44] the assessment of the distribution of C₆₀ material on zebrafish brain would be useful to reinforce our findings. Nevertheless, studies have demonstrated that particles even larger than the nano/microparticles of this study were able to reach brain. For instance, the study of Sarlo and colleagues [45] showed that 1000 nm latex fluorospheres were recovered from rat brains one and 24 hours after intravenous (i.v.) injection. In addition, the study by Zhu and colleagues [41] showed that exposure to fullerene aggregates suspended in water (with average diameters of approximately 349 and/or 1,394 nm) decreased glutathione in brain of juvenile carps (*Carassius auratus*).

In the present study, we have evaluated the effect of different C₆₀ doses (7.5, 15, and 30 mg/kg) and different times of exposure (6, 12, and 24 hours) on AChE activity and *ache* expression in zebrafish brain. In the concentrations tested, only the animals exposed to 30 mg/kg for 24 hours have shown enhanced AChE activity. The RT-qPCR results suggested that the activation of brain AChE is not directly related with the transcriptional control. The *in vitro* results indicated that none of the C₆₀ suspensions had a direct effect on the enzyme. It is important to highlight that *in vitro* experiments do not evaluate the influence of other indirect mechanisms such as cell signaling pathways. Altogether, our results indicate that the effect of intraperitoneal exposure to nano/microparticles of fullerene (C₆₀) on brain AChE activity was neither related to AChE gene activation nor to the direct action of this molecular material on this enzyme but probably involved a posttranscriptional or posttranslational modulation of this enzymatic activity.

Moreover, we have shown the effects of C₆₀ exposure over the antioxidant competence and lipid peroxidation in zebrafish brain. The results demonstrated that the exposition to 30 mg/kg during 24 hours did not alter the antioxidant competence and yielded in higher levels of lipid peroxidation. The lack in the antioxidant response to a pro-oxidant situation could explain the augmented lipid peroxidation found in zebrafish brain. In addition, Totsuka and colleagues [43] reported increased micronuclei frequencies, induced DNA damage, and increased mutant frequencies after C₆₀ nano/microparticles suspension exposure.

AChE is indispensable for terminating acetylcholine-mediated neurotransmission at cholinergic synapses [46].

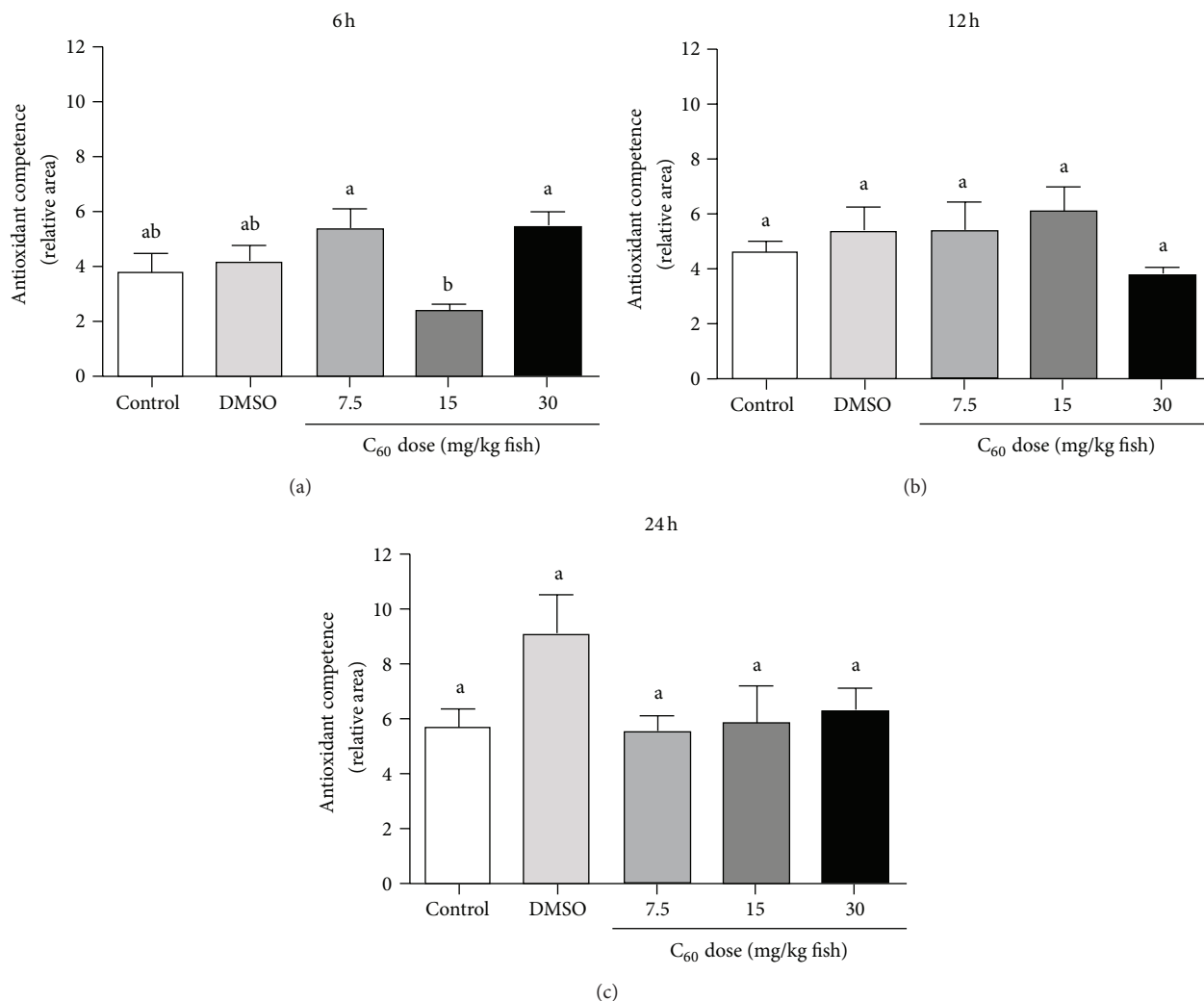


FIGURE 5: Antioxidant capacity. Total antioxidant capacity against peroxy radical in zebrafish brain after 06 (a), 12 (b), and 24 hours (c) of fullerene C₆₀ exposure at distinct concentrations (7.5–30 mg/kg fish). Bars represent the mean±SEM of at least three independent experiments. Different letters indicate significant differences ($P < 0.05$) between groups.

In this context, AChE is inhibited by organophosphorus and carbamate insecticides and by neurotoxins, which are structural analogues of acetylcholine [47]. In addition, there are lines of evidence to suggest that AChE contributes to diverse physiological processes through its involvement in the regulation of cell proliferation, differentiation, and survival. As a consequence, more recently AChE has been redefined as an important regulator of apoptosis, because it can be induced by a variety of apoptotic stimuli [48, 49]. It is well known that apoptosis underlies the neurotoxic effects of various compounds. Moreover, zebrafish brain AChE activation has also been demonstrated as a consequence of exposure to known neurotoxic compounds, including aluminum [20] and ethanol [21] and to the cyanobacterial toxin microcystin-LR [50].

The antioxidant or pro-oxidant effects induced by C₆₀ exposure are still a debatable issue [40, 51]. C₆₀ is photoexcited under UV or visible light [42], a condition, for example,

that elicited lipid peroxidation in brains of common carp (*C. carpio*) [11]. On the other hand, the absence of light did not completely inhibit oxidative stress generation in embryonic zebrafish after C₆₀ exposure [52]. Moreover, cell membrane lipid peroxidation was suggested as the main mechanism of toxicity caused by fullerenes' exposure [10, 11]. In addition, the study by Sayes and colleagues [9] reported cytotoxicity (cell apoptosis) in three human cell cultures including astrocytes caused by cell membrane lipid peroxidation due to exposure to nano-C₆₀ (a water-soluble fullerene species).

The results presented in this paper provide further experimental evidence that C₆₀ exposure can be neurotoxic. Adult zebrafish exposed to nonphotoexcited C₆₀ nano/microparticles (30 mg/kg for 24 hours) have shown enhanced AChE activity and augmented lipid peroxidation in brain. The study by Melo and colleagues [53] proposed a mechanism to explain enhanced AChE activity mediated by oxidative stress generation in cultured retinal cells exposed

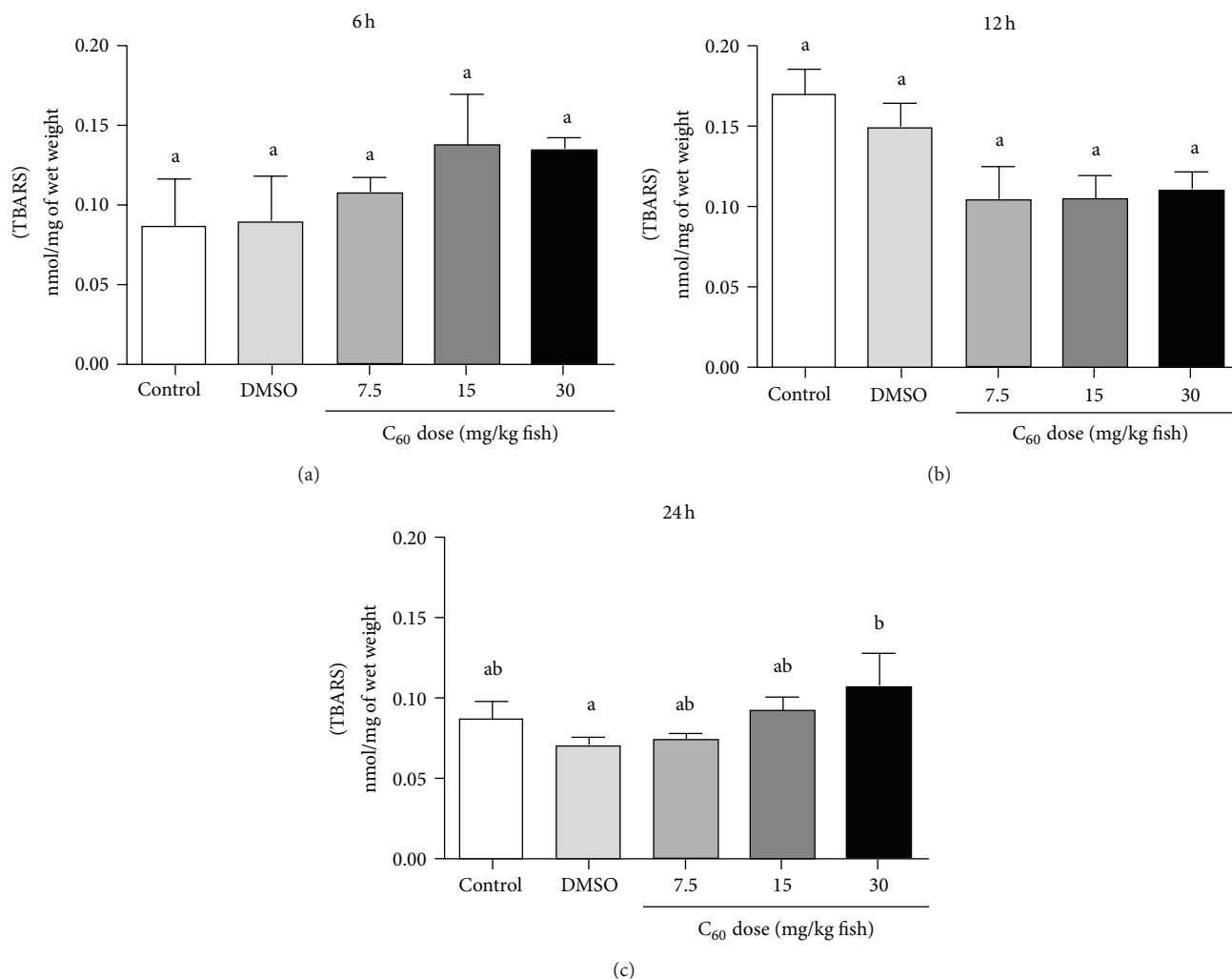


FIGURE 6: Lipid oxidative damage. Concentration of thiobarbituric acid reactive substances (TBARS; nmol/mg of wet weight) in zebrafish brain after 06 (a), 12 (b) and 24 hours (c) of fullerene C₆₀ exposure at distinct concentrations (7.5–30 mg/kg fish). Bars represent the mean ± SEM of at least three independent experiments. Different letters indicate significant differences ($P < 0.05$) between groups.

to amyloidogenic peptide $A\beta_{25-35}$. The incubation with $A\beta_{25-35}$ led to an increment of ROS formation and increased significantly lipid peroxidation levels which decreased cell membrane order and ultimately led to the exposure of more active sites of the AChE. As a consequence, AChE activity was increased. It is possible to speculate that similar events had occurred in zebrafish brain. However, further studies must be performed in order to evaluate this hypothesis.

Abbreviations

ChAT: Choline acetyltransferase
 ACh: Acetyl choline
 AChE: Acetylcholinesterase
 BuChE: Butyrylcholinesterase
 UV: Ultraviolet light
 ACAP: Antioxidant capacity
 TBARS: Thiobarbituric acid reactive substances

EDTA: Ethylenedioxy-diethylene-dinitrilo-tetraacetic acid
 EGTA: Ethylene glycol bis(beta amino ethylether)-N,N,N',N'-tetraacetic acid
 DTNB: 5,5'-Dithiobis-2-nitrobenzoic acid
 HEPES: 4-(2-Hydroxyethyl)-1-piperazineethanesulfonic acid
 BHT: Butylated hydroxytoluene
 ABAP: 2,2'-Azobis(2-methylpropionamide) dihydrochloride
 KCl: Potassium chloride
 SDS: Sodium dodecyl sulfate
 TMP: Tetramethoxypropane
 DMSO: Dimethyl sulfoxide
 MeOH: Methanol
 ACSCh: Acetylthiocholinex hydrolysis
 SCh: Thiocholine
 ROS: Reactive oxygen species.

Conflict of Interests

The authors do not have any conflict of interests.

Acknowledgments

This work was supported by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) (“Nanotoxicologia ocupacional e ambiental: subsídios científicos para estabelecer marcos regulatórios e avaliação de riscos”-Proc. 552131/2011-3) and DECIT/SCTIEMS through CNPq and Fundação de Amparo à Pesquisa do Estado do Rio Grande do Sul (FAPERGS) (Proc. FAPERGS 10/0036-5-PRONEX). G. O. Dal Forno, M. B. de Azevedo and R. S. Britto were recipients of fellowships from Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES). L. W. Kist and R. S. Fritsch were recipient of fellowships from CNPq. S. S. Guterres, C. D. Bonan, J. M. Monserrat, and M. R. Bogo are productivity research fellows from CNPq. The logistic and material support from the Instituto Nacional de Ciência e Tecnologia de Nanomateriais de Carbono (CNPq) was essential for the execution of the present study.

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