

RESEARCH

Open Access

Establishment of a self-propagating population of the African malaria vector *Anopheles arabiensis* under semi-field conditions

Kija RN Ng'habi^{1,2*}, Dickson Mwasheshi¹, Bart GJ Knols^{3,4}, Heather M Ferguson⁵

Abstract

Background: The successful control of insect disease vectors relies on a thorough understanding of their ecology and behaviour. However, knowledge of the ecology of many human disease vectors lags behind that of agricultural pests. This is partially due to the paucity of experimental tools for investigating their ecology under natural conditions without risk of exposure to disease. Assessment of vector life-history and demographic traits under natural conditions has also been hindered by the inherent difficulty of sampling these seasonally and temporally varying populations with the limited range of currently available tools. Consequently much of our knowledge of vector biology comes from studies of laboratory colonies, which may not accurately represent the genetic and behavioural diversity of natural populations. Contained semi-field systems (SFS) have been proposed as more appropriate tools for the study of vector ecology. SFS are relatively large, netting-enclosed, mesocosms in which vectors can fly freely, feed on natural plant and vertebrate host sources, and access realistic resting and oviposition sites.

Methods: A self-replicating population of the malaria vector *Anopheles arabiensis* was established within a large field cage (21 × 9.1 × 7.1 m) at the Ifakara Health Institute, Tanzania that mimics the natural habitat features of the rural village environments where these vectors naturally occur. Offspring from wild females were used to establish this population whose life-history, behaviour and demography under semi-field conditions was monitored over 24 generations.

Results: This study reports the first successful establishment and maintenance of an African malaria vector population under SFS conditions for multiple generations (> 24). The host-seeking behaviour, time from blood feeding to oviposition, larval development, adult resting and swarming behaviour exhibited by *An. arabiensis* under SFS conditions were similar to those seen in nature.

Conclusions: This study presents proof-of-principle that populations of important African malaria vectors can be established within environmentally realistic, contained semi-field settings. Such SFS will be valuable tools for the experimental study of vector ecology and assessment of their short-term ecological and longer-term evolutionary responses to existing and new vector control interventions.

Background

In Africa, current frontline strategies for reducing malaria transmission rely on the use of residual insecticides through application on insecticide-treated nets (ITNs) and indoor residual spraying (IRS). The contribution of these strategies to reduce child mortality and

morbidity has been considerable [1-4]. However, these approaches are facing challenges and limitations as the mosquito vectors they target are increasingly becoming resistant to insecticides [5,6] and many exhibit behavioural plasticity (e.g. biting and resting outside of houses, or early in the evening) that limits their contact with insecticides [7], indicating that these strategies alone may not be sufficient and that new control strategies are needed to supplement them [8].

* Correspondence: kija@ihi.or.tz

¹Biomedical and Environmental Thematic Group, Ifakara Health Institute, Box 53, Ifakara, Tanzania

Full list of author information is available at the end of the article

One of the challenges undermining contemporary vector control strategies is the limited understanding of the ecological complexities that allow vector populations to persist and evade control approaches. Taking the example of African malaria vectors, there is insufficient understanding of mosquito life-history processes that occur outside of the domestic environment (e.g. houses) where they usually bite, including oviposition, larval development, sugar feeding, mating and dispersal [8-10]. Most vector control studies are understandably focused on developing and evaluating specific interventions. While such studies provide the ultimate evidence for evaluating whether to adopt a particular strategy, failure to concurrently measure the ecological parameters of the target vector population during the trial means that little evidence is available to interpret why an intervention failed, and what aspects of its implementation could be modified to achieve greater success. Paying explicit attention to mosquito ecology is vital not only for interpretation of why some otherwise well proven interventions are less effective than expected in different ecological settings, but also for identifying other vulnerabilities in the mosquito life cycle that could be targeted by novel methods.

Gaining insight into the ecological processes of malaria vectors can be both logistically difficult and expensive in natural field settings. This is due to the lack of sampling tools for reliably measuring the abundance and behavioural diversity of different species, genotypes, sexes and life-history stages of mosquito vectors inside and outside of domestic environments, and because of the substantial heterogeneity in their density over time and space [11,12]. As a result of these inherent challenges, many researchers adopt a laboratory experimental approach to quantify key aspects of mosquito life-history and demography. However, it is recognized that laboratory conditions may be insufficient to adequately represent vector fitness and behaviour in nature. Furthermore, the artificial feeding and rearing regimes used in laboratory colonies have been associated with the appearance of behaviours [13,14] and phenotypic traits [15] that are atypical of corresponding field populations. Consequently, in order to progress understanding of vector ecology beyond the limitations of current field and laboratory approaches, there is an urgent need for more environmentally realistic experimental systems where mosquito vector behaviour, ecology and population dynamics can be studied in a natural context over multiple generations.

Contained semi-field systems (SFS) have been proposed as more realistic and reliable experimental tools for the characterization and manipulation of vector ecology [16,17]. An SFS is defined as an experimental

mesocosm, situated within the natural environment of the target vector population and exposed to similar climatic conditions, within which all natural dietary and habitat resources for their life-cycle completion are present [16,17]. The movement of insect vectors into or out of the SFS is typically prevented by netting which blocks their dispersal, but not natural airflow or climatic influences. A key benefit of SFS is that they permit the maintenance of relatively large vector populations in a situation where mating and other behavioural activities can occur more naturally than in the laboratory, and where inbreeding may be less likely to occur. It is thus expected that the demographics, genetic composition, behaviour and life-history of vectors maintained under SFS conditions will be much more representative of wild populations than laboratory colonies. Another advantage of such systems is that in contrast to field studies, the exposure of workers to pathogens such as malaria can be eliminated (e.g by restricting access to potential sources of infection). In the absence of such risk, researchers can conduct a wider range of experimental manipulations, including exposure to mosquito biting, that would be ethically unacceptable in the field. Finally by facilitating detailed study of a defined vector population over time, SFS provide a unique opportunity to investigate their evolutionary as well as ecological dynamics in response to experimental manipulations that mimic the effect of predicted environmental change or interventions; a feat very difficult to achieve under natural field conditions.

Although the value of SFS as an experimental tool for insect disease vectors is increasingly recognized [17-21], few such systems have been successfully established [16,17], and none so far have reported the successful maintenance of a vector population for multiple generations. Here we report the first successful long-term establishment (> 24 generations) of an African malaria vector population under SFS conditions within an area of endemic transmission in southern Tanzania [17]. The study focused on the establishment of *An. arabiensis*, a widespread vector of malaria in Africa [22,23]. Historically, *Anopheles gambiae* s.s. has been recognized as the most important vector of malaria in Africa. However, the abundance and distribution of this vector species is shrinking in many parts of the continent following the widespread use of ITNs, with its sister species *An. arabiensis* playing an increasingly important role in maintaining transmission [23-25]. This is because *An. gambiae* s.s. is more endophilic while *An. arabiensis* is more exophilic and less susceptible to indoor control measures. Given the growing importance of *An. arabiensis*, there is increased interest in obtaining knowledge of its ecology to stimulate new approaches for its control.

Methods

Experimental set-up

A large netting-enclosed semi field system (SFS) was constructed for the study of malaria vector ecology at the Ifakara Health Institute (IHI) in southern Tanzania [17]. This 1800 m³ facility is situated on the main campus of the IHI which is within the Kilombero valley (7°44"- 9°29"S/35°33" -36°56" E), an area of high malaria endemicity where intense levels of transmission are maintained year round [26]. Although all of the three main major African vector species are found in this area (*Anopheles arabiensis*, *An. funestus* and *An. gambiae s. s.*), transmission is largely dominated by *An. arabiensis* which composes >85% of the vector population in most areas [25].

One large experimental chamber (21 × 9.1 × 7.1 m) of the SFS was designated for the establishment of a long-term *An. arabiensis* population. This chamber is enclosed from the surrounding environment by PVC-coated polyester netting (346 holes per inch², Polytex UK) with the interior set up to mimic the natural habitat features of the rural village environments where *An. arabiensis* are typically found. The floor of the chamber was covered with 30 cm of soil obtained from the nearby area, and the vegetation emerging from seeds therein was allowed to grow naturally (Figure 1a). In addition, a variety of other food crops which are normally cultivated around rural homesteads were also planted in the system (e.g. banana plants [*Musa paradisiacal*], papaya [*Carica papaya*], and sweet potatoes

[*Ipomoea batatas*]). Furthermore, castorbean plants (*Ricinus communis*), on which anopheline mosquitoes have been observed to rest and feed upon in East Africa [27,28] were also planted. Although the outer walls of the experimental chamber were separated from the surrounding environment by netting, the decision was taken to cover the roof with polyethylene plastic (plastic film, Filclair Serren Industry N.V.) to provide flexibility to experimentally manipulate rainfall in future research, and also to protect the chamber from the rare but extremely heavy rains that occasionally occur in Kilombero. Consequently, vegetation within the system was watered by sprinklers three times each week. A traditional mud walled house, cow shed and a chicken coop (Figure 1a, c, d) were constructed following local design to provide adult resting sites [17]. Clay pots (n = 23) partially filled with water (to provide humidity) were also distributed throughout the chamber (Figure 1b) to provide additional refuge sites to adult mosquitoes [29]. Clay pots were made locally following the design typically used for water storage and cooking in Kilombero.

In the wild, *An. arabiensis* typically lay their eggs in small, shallow sunlit water pools ranging in size from puddles up to large swamps [30,31], and can include man-made as well as natural water holding bodies that are free from canopy cover [32,33]. To mimic natural sites, artificial larval habitats (Figure 2a) of variable sizes (large, medium and small) were made by half filling plastic basins with a base of soil, and then adding water to



Figure 1 The inside of the SFS experimental chamber where the *An. arabiensis* population was established, showing:

- (A) mud-walled house, natural vegetation and planted food crops,
- (B) clay pots used as outdoor resting sites, (C) chicken coop and
- (D) cow shed.

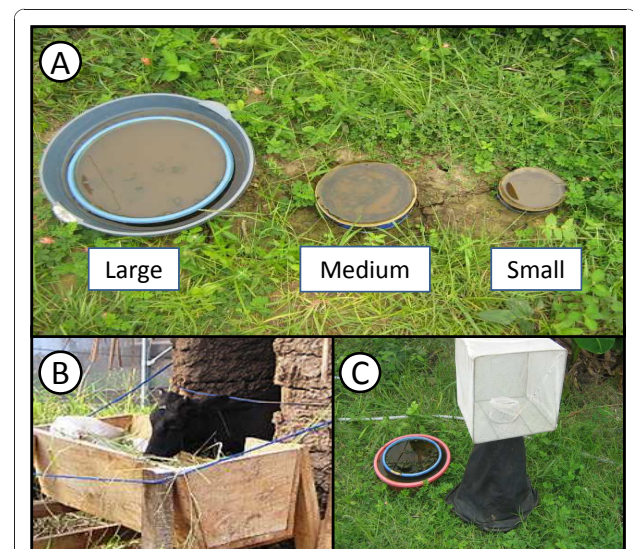


Figure 2 Mosquito habitat features within the SFS: (A) Larval habitats of large (diameter 43.5 cm, maximum depth 5 cm), medium (diameter 19.5, maximum depth 4 cm) and small size (diameter 13.5, maximum depth 3 cm) within the SFS, (B) a calf host used to provide blood meals to adult females, and (C) a trap for collecting adults emerging from a larval habitat.

surface level (Figure 2a). This design gave females the opportunity to land and lay their eggs on wet soil or shallow water as they do in nature [34]. The soil layer acted as a source for microbial and/or algal growth which provides food for larval growth and development [35]. Twenty large artificial larval habitats (diameter 43.5 cm, maximum depth 5 cm) were made by burying plastic containers to ground level. In addition, five medium-sized (diameter 19.5, maximum depth 4 cm) and five small (diameter 13.5, maximum depth 3 cm) artificial larval habitats (Figure 2a) were also distributed throughout the compartment. Water depth was maintained in these habitats through daily replenishment with tap water.

Establishment of *An. arabiensis*

The population of *An. arabiensis* established in the SFS was founded from a wild population in the nearby village of Sagamaganga (~20 km from the IHI, -8.0667 S; 36.8000 E). This village is situated along the flood plains of the Kilombero River where anopheline larval habitats are abundant. Pilot work had shown that *An. arabiensis* constitutes approximately 90% of the *An. gambiae s.l.* complex in this area (Mayagaya & Ferguson, pers. comm). Live blood-fed females that were morphologically identified as *An. gambiae s.l.* were collected from houses and animal sheds using mouth aspirators in May 2008. Collections were made continuously until enough females were obtained to produce the target number of larvae for release into the SFS (provisionally set at 3000 for release over one week in May 2008). On the day of their capture, blood-fed females were transported to the IHI semi-field insectary where they were transferred into individual cups for oviposition. After oviposition, wild-collected females were killed and subjected to PCR analysis to confirm their species [36]. Larval offspring of all females (N = 560) identified as *An. arabiensis* were pooled and continuously added to the artificial larval habitats in the SFS during the week of release (Figure 1c).

Prior to the release of larvae into the SFS, a variety of other invertebrates that naturally occur in and around houses in rural Tanzania (e.g. praying mantids [Mantidae], grasshoppers [Acridoidea], ants [Formicidae]) and important *Anopheles* predators such as jumping spiders (Salticidae) [37] were observed in the SFS. These predators probably entered the system in soil and building material during the chamber set-up, and were allowed to establish within it. After introduction, larvae were not provided with any source of food other than the microorganisms growing naturally within larval habitats.

Adult maintenance and blood feeding

Adult mosquitoes emerging from larval habitats were allowed to fly freely in the SFS and feed on plant and vertebrate host sources available within it. A major issue

when establishing long-term mosquito populations in SFS is to ensure containment and prevent the accidental introduction of malaria parasites that could infect mosquitoes and pose an infection risk to researchers. To achieve this, four steps were routinely undertaken. First, mosquito containment was ensured by the installment of a triple door entry-system which prevented direct entrance or accidental mosquito escape (each door is opened and closed independently). Second, the integrity of the outer netted walls and roof was checked thrice weekly during routine inspection. Third, blood meals were provided only from calves (which *An. arabiensis* feed on as well as humans under natural conditions; Figure 2b). As cattle are dead-end hosts for human malaria parasites, mosquitoes that have fed only on them are incapable of becoming infected or transmitting parasites. Host blood was provided to mosquitoes by introducing a calf into the SFS every evening from 7.00 PM - 7.00 AM for five consecutive nights each week. Finally, all research staff working in the area were screened for malaria parasites on a weekly basis using a rapid diagnostic kit before being allowed to enter the SFS. Any staff that tested positive for malaria were immediately given a full course of anti-malarial treatment (artemisinin combination therapy), and restricted from entering the SFS for at least 2 weeks after their infection was cleared.

Regular entomological monitoring

Daily temperatures inside microhabitats within the SFS was monitored by placing data loggers (Tiny tag™) in aquatic larval habitats and potential adult resting sites (the house, cow shed, chicken coop and clay pots). Water temperature was monitored only in large and small larval habitats by submerging data loggers within them (Tiny tag™). After the introduction of *An. arabiensis*, larval habitats were inspected daily for the presence of larvae and pupae. Larval development was monitored for eighteen days from the day that first instars were released into ten larval habitats. Emerging adults were monitored by setting up emergence traps (Figure 2c) over all large larval habitats. Emerging adults captured in these traps were counted and then released into the SFS. *Anopheles arabiensis* population growth over the first five generations within the SFS was assessed from the number of adults emerging (measured by capture from emergence traps). The length of time between consecutive mosquito generations was also estimated. This was done by adding together the number of days from the time of blood-feeding to the first observation of first instars within larval habitats, the subsequent number of days required for these first instars to emerge as pupae, and the estimated number of days between pupal emergence and the resultant females taking their first blood meal.

An experiment was set up to assess the effect of larval habitat size on larval survival. Here, subsets of large, medium and small larval habitats were covered with netting to prevent oviposition from free flying females. A fixed number of 100 first instar larvae (from gravid females collected in the SFS) were added to these habitats to monitor their development. This experiment was run from the 5th-14th generation to achieve a total of twenty replicates from each larval habitat size class. Habitats to which larvae were added were checked on a daily basis, with the total number of pupae that successfully developed being counted. These pupae were subsequently transferred to another larval habitat in the SFS for emergence.

The resting behaviour of both males and females was assessed by counting the number of adults observed inside the mud house, cow shed and chicken coop (all considered 'indoor' habitats), and the clay pots (outdoors) during the day with the aid of a flash light and counter. The assessment was conducted for three consecutive days from generation 2 to 7, and was repeated again at generation 24. The age structure of males and females in the SFS was estimated from a random sub-sample of 35 males and 176 females that were collected at generation 20. Dissections were performed on this subset to assess their approximate age from their reproductive morphology, with female age being estimated from parity status as indicated from their ovaries [38] and male age from the number of spermatocysts and morphology of their accessory glands [39]. Swarming behaviour was also assessed by daily inspection for aggregations of males in flight at dusk (between 7-8 pm).

Data analysis

Analysis of variance (ANOVA) was used to assess temperature differences between larval habitats and resting sites inside the SFS using the SPSS statistical package (13.0 for windows). Generalized linear models were used to test whether larval survival (as assessed by the proportion of pupae emerging from the 100 instars initially present) varied between larval habitats of different size classes (R statistical software). Here 'habitat size' was treated as a main effect, and the generation on which observations were made as a random effect. The daily larval survival rate in large larval habitats was also estimated using the formula $S = p^{1/t}$, where p is the proportion of first instar larvae that survive to pupation and t is the mean time to pupation in days [40]. Similarly, generalized linear models were used to test whether the proportion of *An. arabiensis* adults resting in indoor versus outside habitats varied between sexes.

Results

Microclimatic conditions

Average water temperatures within the large and small larval habitats were within 1.2°C of one another (Table 1).

Due to the limited availability of data loggers, it was not possible to simultaneously measure the temperature inside all of the four potential adult resting sites and outside air conditions in the SFS, however measurements made within these microhabitats at different times of year suggested that all fell within the natural temperature range of *An. arabiensis* (Table 1). Concurrent measurements made within the mud house, cow shed and chicken coop over a one week period in July 2010 indicated that air temperature varied significantly between these resting sites ($F_{2, 2263} = 45.2, P < 0.001$, Table 1). Temperature within the chicken coop was approximately 1°C warmer than the mud house ($P < 0.001$) and cow shed ($P < 0.001$) respectively, with no significant difference in temperature between the house and cow shed ($P = 0.71$).

Larval development and population growth

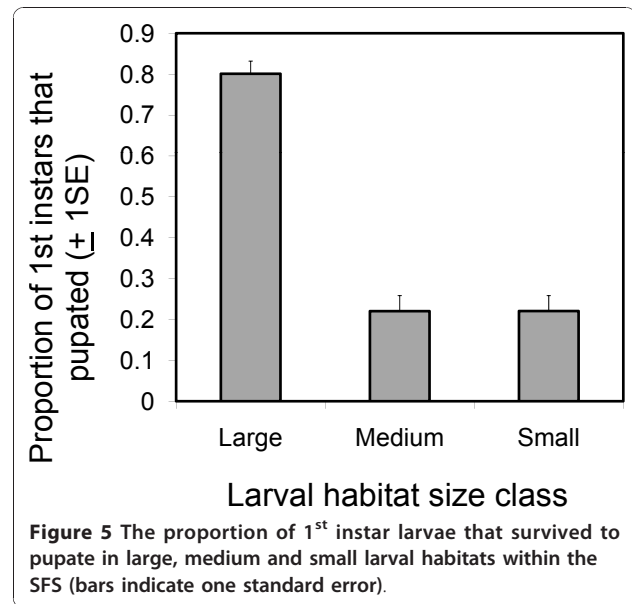
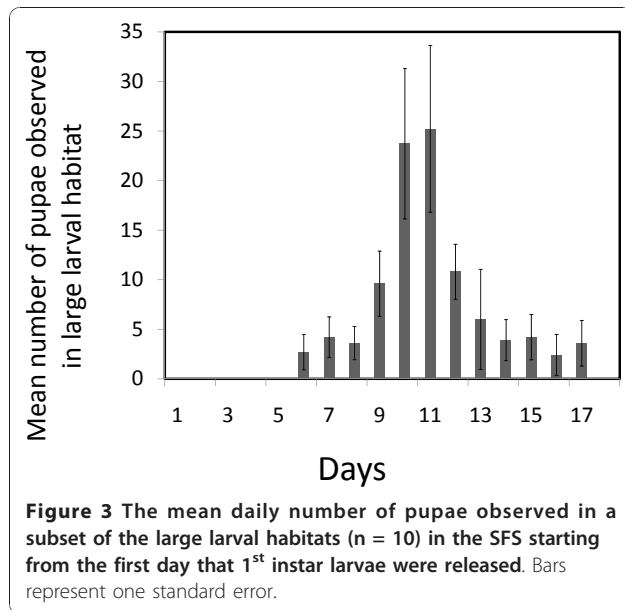
The time required for larvae to develop and pupate after the first release into large habitats ranged from 6 to 17 days, with a median time of 11.5 days (Figure 3). Although the exact number of generations that passed since the founder generation could not be precisely established due to overlapping generations, we conservatively estimated the average time between successive generations as 22 days. This was computed by summing the estimated median development time from 1st instar larvae to pupae (11.5 days), the assumed time from pupation to adult emergence (2 days [41]), the estimated time from adult emergence to blood feeding (estimated at an average of 3 days under field conditions [42]), and the estimated number of days it took from the time that females blood fed in the SFS to the first appearance of 1st instar in larval habitat (5 days).

The number of pupae emerging within the SFS was observed to increase during the first five generations, indicating that the SFS population was growing (Figure 4). The survival of larvae to pupation varied significantly between different larval habitat size classes ($\chi^2 = 62.44$,

Table 1 Mean temperatures (°C) of mosquito larval habitats and adult resting sites inside the SFS

Description	Mean temperature (°C)	Period of measurement
<i>Larval habitat</i>		
Large size	24.9 (0.05)	July 1 st - Oct 15 2008
Small size	25.3 (0.03)	Oct 15-21 2008
<i>Resting site</i>		
Inside mud house	24.39 (0.17)	16-28 July 2010
Inside cow shed	24.23 (0.09)	16-28 July 2010
Inside chicken coop	25.43 (0.09)	16-28 July 2010
Outdoor clay pot	25.47 (0.06)	15-30 Oct 2008
Air temperature	31.30 (0.28)	Feb 29-Mar 9 2008

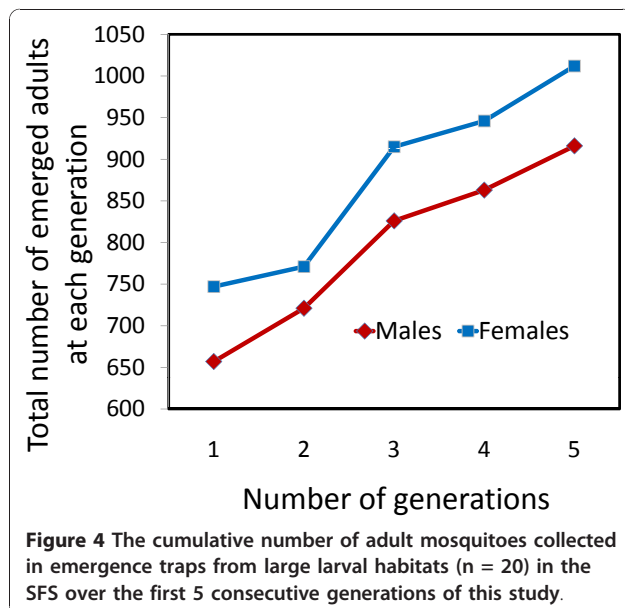
Values in brackets represent one standard error.



$p < 0.001$ Figure 5). Specifically, the pupation rate in the large-sized habitats (69%) was almost 4 times greater than in the small and medium habitats (Figure 5). The daily larval survival rate in large larval habitats was estimated to be 0.962 per day. Due to the labour intensive nature of performing daily larval abundance and adult resting behaviour surveys, the population size in the SFS was monitored for only 5 out of the 24 generations monitored here. However, a general approximation of the average abundance of each *An. arabiensis* generation could in principle be obtained by summing up the

number of emerging adults daily in all larval habitats, collected in emergence traps over each 22 day period (assumed generation length).

Larval predation by ants was observed in the SFS. On several occasions ants were observed on the sides of aquatic habitats, carrying parts of mosquito larvae or adults that had probably been attacked during eclosion. Initially, an attempt was made to reduce ant-related predation by placing larval habitat bowls within a second water-filled outer bowl to provide a protective moat (Figure 1c). At first larvae were only observed within the inner bowl of larval habitats. However from the 3rd generation onwards, females began ovipositing within moat barriers and also inside the shallow water pools within clay pots (Figure 1b), thus ant-predation could not be completely prevented.



Adult feeding and survival

Of the 35 dissected males which were collected on the same day from the SFS (generation 20), only four males (11.4%) were classified as being in the ≤ 4 days old age category [39]. The remaining 31 males (88.8%) had a number of spermatocysts ranging from 0-2 which are estimated to correspond to an age of > 4 days post-emergence. Of the 176 females collected, 42 and 26 were observed to be either blood fed or gravid respectively and were not dissected. The ovaries of the remaining 108 unfed females were dissected to determine their reproductive history. Within this sample, 38 (35.2%) were virgins, 40 (37.04%) nulliparous and 30 (27.8%) had previously laid eggs. It was not possible to precisely age-grade the parous class into gonotrophic cycles.

Resting behaviour and reproduction

Both sexes (male [n = 841] and female [n = 1872]) of *An. arabiensis* adults were observed in all resting sites (Figure 6). While both sexes were more frequently found resting in the outdoor clay pots than in indoor environments, males were substantially more exophilic than females ($\chi_3^2 = 223.05$, $P < 0.001$, Figure 6). Restricting analysis to indoor resting mosquitoes, females were more likely to be found resting in the cow shed than in the house or chicken coop ($\chi_2^2 = 433.93$, $p < 0.001$). Unlike females, males were less selective in their use of indoor resting sites ($\chi_2^2 = 0.39$, $p = 0.83$, Figure 6). During generations 4 and 5, observations were made 3 times each week for the presence of adult mating swarms (at dusk). Of the 24 nights on which observations were made, swarms were detected on 14 occasions (58.3% of the time). When observed, swarms consisted of 50-100 males that formed at dusk (at approximately 7.30 PM, on one half of the SFS where the horizon was visible), and lasted for 15-20 minutes. Also males and females in copula were observed in these swarms.

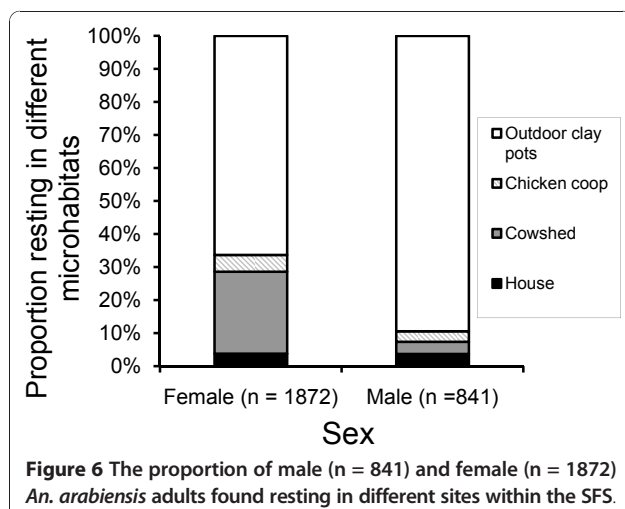
Discussion

This study reports the first successful establishment of a self-replicating population of an African malaria vector, *An. arabiensis*, in a contained semi-field system. It is suggested that a major contributing factor to the successful establishment of this population was the close concurrence between the environmental conditions of the SFS and those experienced by mosquitoes in nature. The mean daily temperatures recorded in all SFS larval habitats were within the range reported for natural *An. gambiae s.l.* larval habitats (20°C-36°C) [43,44] and never exceeded their upper tolerance limit of 40°C [34,44,45]. The time required for *An. arabiensis* to

develop from larvae to pupae within these habitats (6 - 17 days) was similar to the reported range (8-18 days) for *An. gambiae s.l.* in the wild [46]; with the median larval development under both our SFS (11.5 days) and field conditions (e.g. 11.9 days [41]) being close. During their development, the only source of food that would have been available to larvae was microbes and algae that developed naturally within their aquatic habitats. These resources are the primary food source of larvae in natural populations [35], and likely played a similar role in the SFS.

The survival of larvae to pupation within the SFS was highly dependent on larval habitat size, with the pupation rate in 'large' habitats being four times higher than in medium and small habitats. The pupation rate within large habitats corresponds to a daily larval survival of 96.2%, which is in line with what has been reported in other semi-field settings (95.7%) [40], but slightly higher than what has been reported in some field populations (e.g. 85% [47]). The moderately higher pupation rate observed here may be a result of reduced predation and competition within our SFS relative to field conditions. It is hypothesized that the enhanced pupal productivity of the 'large' larval habitats in the SFS is a function of the greater amount of food resources (algae and microbes) they can support relative to smaller habitats. In the absence of predators and pathogens, food availability within larval habitats is a key predictor of the number of adults that emerge from them [43]. The total quantity of microbial growth in larval habitats is related to both surface area and volume [30] and thus are greater in the larger than small habitats where larvae may have experienced more intense resource competition.

Similar to larval development, the behaviour of adult mosquitoes within the SFS was also consistent with what has been reported in nature. The mean temperatures within available adult resting sites varied by no more than 1.3 °C, and all were within the tolerance range of *An. arabiensis* [48,49]. However, there was considerable variation in resting preference. Both males and females were more likely to rest in clay pots (outdoors) than in indoor sites; confirming the previously demonstrated exophilic tendency of this species in the wild [50,51]. Although *An. arabiensis* is known to be substantially more exophilic than *An. gambiae s.s.*, it has not previously been possible to estimate the relative proportion of outdoor resting within natural populations. If our SFS results are typical of natural populations, it suggests that up to 60% of resting adults may be missed by surveys and control measures targeted indoors. Restricting consideration to indoor resting sites, females were more selective than males. Specifically, females were more likely to rest in the cow shed than in other sites, whereas indoor resting males occurred with similar frequency in all three resting sites. The closer



association of females with the cow shed suggests they prefer to rest close to the host blood source, and may be most efficiently targeted there.

Several other aspects of the adult behaviour and life history of *An. arabiensis* within the SFS conformed to what is known of their natural ecology. For example, female *An. arabiensis* in the SFS readily blood-fed on cows and were able to maintain their population exclusively on this host type as has been reported in other field populations in East Africa [52]. In nature, plants are thought to be a major source of sugar for both male and female mosquitoes [27,28]. Mosquitoes can readily imbibe and digest plant juices and nectar to enhance their survival [53,54]. As males rarely survive for more than 48 h without a sugar source [54], evidence that a large proportion of adult males (at generation 20) within the SFS (80%) were four days or older indicates that they were feeding on plant nectar sources within it. These plant sugar sources may also have been used as nutritional supplements by females. Further studies such as gut content analysis could confirm the extent of *An. arabiensis* reliance on plant nectar, and which types are preferred.

Analysis of the age structure of male and female *An. arabiensis* at generation 20 indicated that a significant proportion of both male and females survived beyond the minimum period required to reproduce. In this study, approximately 80% of males were estimated as being > 4 days, the period beyond the peak of *An. gambiae s.l.* mating activity [55,56]. Similar analysis of adult females indicated that 28% of females had survived through their first gonotrophic cycle (estimated as < 4 days). The observed parous rate of 0.28 corresponds to an estimated daily survival rate of 0.53, which falls within the range of reported for adult *An. gambiae s.l.* daily survival during the dry (0.49) and wet seasons (0.84) in East African populations [25,57,58]. Although the age-grading methods used in this study provide a general indication of adult mosquito age, they could not precisely estimate how long individuals survived beyond four days. Further age-grading studies using chronological age estimation methods to more precisely estimate the life span are recommended [59].

Another natural adult behaviour observed within this SFS population was swarming. Swarming has been suggested as the primary reproductive strategy of *An. gambiae s.l.* mosquitoes [60,61], and has been documented in several wild populations in East [62] and West Africa [61,63]. However in some parts of East Africa, male anopheline swarms have been difficult to observe, possibly due to the fact that they occur at dusk when visibility is poor [61,62,64], or because these populations deploy alternative strategies such as mating indoors [65]. Due to its inconsistent occurrence (observed on 58.3% of

occasions), we could not establish whether swarming was the primary mating strategy of mosquitoes in the SFS. Further investigation is needed to identify the mating strategies of *An. arabiensis* both in the SFS and the wild population from which they were established.

The SFS approach adopted provides useful opportunities for characterizing mosquito demography, life-history and behaviour traits that are difficult to measure in nature, and poorly represented in the laboratory [11,66]. This advantage will be particularly strong for vector species that are difficult to colonize under laboratory conditions (e.g. *An. funestus* and *Mansonia annulata*) and/or sample in the wild; as the relatively more natural conditions within the SFS may prove more amenable for their establishment. Although it is argued that biological inferences generated in SFS provide a more accurate representation of field populations than laboratory colonies, this approach also has limitations. For example in the SFS, there were no interactions between *An. arabiensis* and other mosquito species in larval habitats as occurs in nature [67]. Furthermore the high availability of aquatic sites within the SFS probably minimized cannibalism and other types of intraspecific competition [68]. Also unlike field settings, pressure from insecticides or other vector control interventions was absent in the SFS, and the diversity of natural predators and competitors was probably under represented. Furthermore although humans are a common host for *An. arabiensis* in many African settings, this host type was not available in the SFS during their typical host-seeking period (e.g. 10 pm -5 am [69]). Consequently, *An. arabiensis* within this system were not exposed to human malaria parasites, which may have eliminated another source of selection pressure that acts on natural mosquito populations. However, as only 1-2% of *An. gambiae s.l.* become infected even in highly endemic settings, it is perhaps unlikely that parasites are a significant source of selection [70]. Finally, there have been some accounts that the host preference of vectors (from an insectary population) when assayed under semi-field conditions give a biased representation of natural feeding preferences [14]. Whether SFS populations that have been established directly from a wild population and consistently maintained on natural host types may also develop atypical preferences is not yet unknown, but careful and repeated monitoring is required to ensure that these and other behavioural traits of SFS populations reliably approximate natural vector ecology. Supplementing stable population within SFS routinely with fresh materials from the field may be another option to circumvent this, but requires further study. Thus it is cautioned that SFS studies should be seen as a bridging ground between lab and field, and not replacement of field studies. Furthermore, where possible observations

from these SFS studies should be verified under natural conditions.

In addition to elucidating fundamental aspects of vector ecology, SFS studies have a variety of other potential uses including the preliminary evaluation and optimization range of vector control interventions and trapping methods. SFS can be used to develop and optimize new field sampling tools and trapping methods, and identify the most promising candidates to take forward for full field testing within a relatively short period of time (e.g. optimal doses for repellents/attractants [71]). Additionally, the long-term establishment of vector populations allows the testing of some important evolutionary questions that are difficult to monitor in open field populations, such as prediction of the nature of behavioural and physiological resistance strategies against specific interventions. Of particular relevance is the use of SFS studies to determine the feasibility of disease control strategies based on the release of sterile and/or genetically-modified refractory mosquitoes [72]. Despite the optimism that such novel control strategies have garnered [72], so far investigation of their feasibility has been largely based on laboratory studies [72]. It remains unknown whether mosquitoes carrying GM sterility or refractory traits would be fit enough to compete for female mates with their wild counterparts [72]. As unrestricted field trials of GM mosquitoes are unlikely to be authorized before all potential biosecurity risks have been evaluated [72], testing the viability of these mosquitoes under contained semi-field settings will be a mandatory first step before proceeding

Conclusions

The present study reports the successful establishment of a self-propagating malaria vector population in an enclosed semi natural environment. Such populations provide a valuable new research tool for the experimental study of malaria vector ecology, evolution and control under environmental conditions that are largely representative of natural conditions. Climatic conditions within the SFS were broadly similar to those within natural *Anopheles* transmission settings and mosquitoes within the system exhibited similar demographic, life-history and behavioural traits to those in the field. This development will help facilitate the development and optimization of many vector control strategies, including the long-awaited transfer of new genetic control technologies from the laboratory to field application.

Acknowledgements

We would like to thank the Entomology team at the IHI, specifically, Ally Daraja, Paulina Kasanga and Hasani Mtambala for helping with mosquito collections in the field. We also thank Prof. Marcel Dicke for reviewing this manuscript prior to submission. We acknowledge financial support from the

IAEA and the Caroline MacGillavry Fund (KNAW, The Netherlands) through a fellowship awarded to KRN. This research was supported by a VIDJ grant (#864.03.004) from the Dutch Scientific Organisation (NWO) to BGJK. We also acknowledge support from BBSRC through a fellowship awarded to HMF (BB/D020042/1).

Author details

¹Biomedical and Environmental Thematic Group, Ifakara Health Institute, Box 53, Ifakara, Tanzania. ²Wageningen University and Research Centre, Wageningen, The Netherlands. ³Division of Infectious Diseases, Tropical Medicine & AIDS, Academic Medical Center, F4-217, Meibergdreef 9, 1105 AZ Amsterdam, The Netherlands. ⁴K&S Consulting, Kalkestraat 20 6669 CP Dodewaard, The Netherlands. ⁵Institute of Biodiversity, Animal Health and Comparative Medicine, University of Glasgow, G12 8QQ, Glasgow, UK.

Authors' contributions

KRN, BGJK and HMF designed this study. KRN and DM carried out the laboratory and SFS work. KRN analyzed and interpreted the data. KRN, HMF, BGJK drafted the manuscript. All authors read and approved the manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 2 August 2010 Accepted: 8 December 2010

Published: 8 December 2010

References

1. Fegan GW, Noor AM, Akhwale WS, Cousens S, Snow RW: **Effect of expanded insecticide-treated bednet coverage on child survival in rural Kenya: a longitudinal study.** *The Lancet* 2007, **370**:1035-1039.
2. Lindblade KA, Eisele TP, Gimnig JE, Alaii JA, Odhiambo F, ter Kuile FO, Hawley WA, Wannemuehler KA, Phillips-Howard PA, Rosen DH: **Sustainability of reductions in malaria transmission and infant mortality in western Kenya with use of insecticide-treated bednets 4 to 6 years of follow-up.** *JAMA* 2004, **291**:2571-2580.
3. Müller O, Traoré C, Kouyaté B, Yé Y, Frey C, Coulibaly B, Becher H: **Effects of insecticide-treated bednets during early infancy in an African area of intense malaria transmission: a randomized controlled trial.** *Bull WHO* 2006, **84**:120-126.
4. Abdulla S, Gemperli A, Mukasa O, Armstrong Schellenberg JR, Lengeler C., Vounatsou P, Smith T: **Spatial effects of the social marketing of insecticide-treated nets on malaria morbidity.** *Trop Med Inter Health* 2005, **10**:11-18.
5. Müller P, Warr E, Stevenson BJ, Pignatelli PM, Morgan JC, Steven AC, Yawson EY, Mitchell SN, Ranson H, Hemingway J, et al: **Field-caught permethrin-resistant *Anopheles gambiae* overexpress CYP6P3, a P450 that metabolises pyrethroids.** *PLoS Genet* 2008, **4**:e1000286.
6. Ranson H, Abdallah H, Badolo A, Guelbeogo WM, Kera-Hinzoumbé C, Yangalbé-Kalnoné E, Sagnon N, Simard F, Coetzee M: **Insecticide resistance in *Anopheles gambiae*: data from the first year of a multi-country study highlight the extent of the problem.** *Malar J* 2009, **8**:299.
7. Geissbühler Y, Chaki P, Emidi B, Govella NJ, Shirima R, Mayagaya V, Mtasiwa D, Mshinda H, Fillingier U, Lindsay SW, et al: **Interdependence of domestic malaria prevention measures and mosquito-human interactions in urban Dar es Salaam, Tanzania.** *Malar J* 2007, **6**:126.
8. Ferguson HM, Dornhaus A, Beeche A, Borgemeister C, Gottlieb M, Mulla MS, Gimnig JE, Fish D, Killeen GF: **Ecology: a prerequisite for malaria elimination and reduction.** *PLoS Med* 2010, **7**:e1000303.
9. Lyimo I, Ferguson HM: **Ecological and evolutionary determinants of host species choice in mosquito vectors.** *Trends Parasitol* 2009, **25**:189-196.
10. Müller G, Schlein Y: **Sugar-questing mosquitoes in arid areas gather on scarce blossoms that can be used for control.** *Inter J Parasitol* 2006, **36**:1077-1080.
11. Magbity EB, Lines JD: **Spatial and temporal distribution of *Anopheles gambiae* s.l. (Diptera:Culicidae) in two Tanzanian villages: implications for designing mosquito sampling routines.** *Bull Entomol Res* 2002, **92**:483-488.
12. Lindsay SW, Parson L, Thomas CJ: **Mapping the ranges and relative abundance of the two principle African malaria vectors, *Anopheles gambiae sensu stricto* and *An. arabiensis*, using climate data.** *Proc R Soc London B* 1998, **265**:847-854.

13. Pates HV, Takken W, Stuke K, Curtis CF: **Differential behaviour of *Anopheles gambiae sensu stricto* (Diptera: Culicidae) to human and cow odours in the laboratory.** *Bull Entomol Res* 2001, **91**:289-296.
14. Lefèvre T, Gouagna LC, Dabire KR, Elguero E, Fontenille D, Costantini C, Thomas F: **Evolutionary lability of odour-mediated host preference by the malaria vector *Anopheles gambiae*.** *Trop Med Inter Health* 2009, **14**:228-236.
15. Huho BJ, Ng'habi KR, Killeen GF, Nkwengulila G, Knols BGJ, Ferguson HM: **Nature beats nurture: A case study of the physiological fitness of free-living and laboratory-reared males *Anopheles gambiae s.l.*** *J Exp Biol* 2007, **210**:2939-2947.
16. Knols BGJ, Njiru BNN, Mathenge EM, Mukabana WR, Beier JC, Killeen GF: **Malariasphere: A greenhouse-enclosed simulation of a natural *Anopheles gambiae* (Diptera: Culicidae) ecosystem in Western Kenya.** *Malar J* 2002, **1**:19.
17. Ferguson HM, Ng'habi KR, Walder T, Kadungula D, Moore SJ, Lyimo I, Russell TL, Urassa H, Mshinda H, Killeen GF, et al: **Establishment of a large semi-field system for experimental study of African malaria vector ecology and control in Tanzania.** *Malar J* 2008, **7**:158.
18. Stone CM, Taylor RM, Foster WA: **An effective indoor mesocosm for studying populations of *Anopheles gambiae* in temperate climates.** *J Am Mosq Contr Assoc* 2009, **25**:514-516.
19. Knols BGJ, Bossin HC, Mukabana WR, Robinson AS: **Transgenic mosquitoes and the fight against malaria: managing technology push in a turbulent GMO world.** *Am J Trop Med Hyg* 2007, **77**:232-242.
20. Knols BGJ, Louis CE: **Bridging laboratory and field research for genetic control of disease vectors.** Dordrecht, The Netherlands.: Springer; 2006, 225.
21. Knols BGJ, Njiru BNN, Mukabana RW, Mathenge EM, Killeen GF: **Contained semi-field environments for ecological studies on transgenic African malaria vectors.** In *Ecology of transgenic mosquitoes*. Edited by: Scott TW, Takken W. Dordrecht, The Netherlands.: Springer; 2003:99-106.
22. Coetzee M: **Distribution of the African malaria vectors of the *Anopheles gambiae* complex.** *Am J Trop Med Hyg* 2004, **70**:103-104.
23. Bayoh MN, Mathias DK, Odiero MR, Mutuku FM, Kamau L, Gimnig JE, Vulule JM, Hawley WA, Hamel Mj, Walker ED: ***Anopheles gambiae*: historical population decline associated with regional distribution of insecticide-treated bed nets in western Nyanza Province, Kenya.** *Malar J* 2010, **9**:62.
24. Lindblade KA, Gimnig JE, Kamau L, Hawley WA, Odhiambo F, Olang G, ter Kuile FO, Vulule JM, Slutskera L: **Impact of sustained use of insecticide-treated bednets on malaria vector species distribution and Culicine mosquitoes.** *J Med Entomol* 2006, **43**:428-432.
25. Russell TL, Lwetojiera DW, Maliti D, Chipwaza B, Kihonda J, Charlwood JD, Smith TA, Lengeler C, Mwanjangala MA, Nathan R, et al: **Impact of promoting longer-lasting insecticide treatment of bed nets upon malaria transmission in a rural Tanzanian setting with pre-existing high coverage of untreated nets.** *Malar J* 2010, **9**:187.
26. Killeen GF, Tami A, Kihonda J, Okumu FO, Kotas ME, Grundmann H, Kasigudi N, Ngonyani H, Mayagaya V, Nathan R, et al: **Cost-sharing strategies combining targeted public subsidies with private-sector delivery achieve high bednet coverage and reduced malaria transmission in Kilombero Valley, southern Tanzania.** *BMC Infect Dis* 2007, **7**:121.
27. Impoinvil D, Kongere JO, Foster WA, Njiru BN, Killeen GF, Githure JI, Beier JC, Hassanali A, Knols BGJ: **Feeding and survival of the malaria vector *Anopheles gambiae* on plants growing in Kenya.** *Med Vet Entomol* 2004, **18**:108-115.
28. Manda H, Gouagna LC, Foster WA, Jackson RR, Beier JC, Githure JI, Hassanali A: **Effect of discriminative plant-sugar feeding on the survival and fecundity of *Anopheles gambiae*.** *Malar J* 2007, **6**:113.
29. Odiero M, Bayoh MN, Gimnig J, Vulule J, Irungu L, Walker E: **Sampling outdoor, resting *Anopheles gambiae* and other mosquitoes (Diptera: Culicidae) in western Kenya with clay pots.** *Journal of Medical Entomology* 2007, **44**:14-22.
30. Gimnig JE, Ombok M, Kamau L, Hawley WA: **Characteristics of larval anopheline (Diptera: Culicidae) habitats in Western Kenya.** *J Med Entomol* 2001, **38**:282-288.
31. Minakawa N, Sonye G, Mogi M, Yan G: **Habitat characteristics of *Anopheles gambiae s.s.* larvae in a Kenyan highland.** *Med Vet Entomol* 2004, **18**:301-305.
32. Minakawa N, Sonye G, Yan G: **Relationships between occurrence of *Anopheles gambiae s.l.* (Diptera: Culicidae) and size and stability of larval habitats.** *J Med Entomol* 2005, **42**:295-300.
33. Munga S, Minakawa N, Zhou G, Barrack OO, Githeko AK, Yan G: **Oviposition site preference and egg hatchability of *Anopheles gambiae*: effects of land cover types.** *J Med Biogr* 2005, **42**:993-997.
34. Huang J, Walker ED, Vulule J, Miller JR: **The influence of darkness and visual contrast on oviposition by *Anopheles gambiae* in moist and dry substrates.** *Phy Entomol* 2007, **32**:34-40.
35. Kaufman MG, Wanja E, Maknoja S, Bayoh MN, Vulule JM, Walker ED: **Importance of algal biomass to growth and development of *Anopheles gambiae* larvae.** *J Med Entomol* 2006, **43**:669-676.
36. Scott JA, Brogdon WG, Collins FH: **Identification of single specimens of the *Anopheles gambiae* complex by the polymerase chain reaction.** *Am J Trop Med Hyg* 1993, **49**:520-529.
37. Jackson RR, Nelson XJ, Sune GO: **A spider that feeds indirectly on vertebrate blood by choosing female mosquitoes as prey.** *PNAS USA* 2005, **102**:15155-15160.
38. Charlwood JD, Birley MH, Dagaró H, Paru R, Holmes PR: **Assessing survival rates of *Anopheles farauti* (Diptera: Culicidae) from Papua New Guinea.** *J Anim Ecol* 1985, **54**:1003-1016.
39. Huho BJ, Ng'habi KR, Killeen GF, Nkwengulila G, Knols BGJ, Ferguson HM: **A reliable morphological method to assess the age of male *Anopheles gambiae*.** *Malar J* 2006, **5**:62.
40. Paaijmans KP, Huijben S, Githeko AK, Takken W: **Competitive interactions between larvae of the malaria mosquitoes *Anopheles arabiensis* and *Anopheles gambiae* under semi-field conditions in western Kenya.** *Acta Trop* 2009, **109**:124-130.
41. Mwangangi JM, Muturi EJ, Shililu J, Muriu SM, Jacob B, Kabiru EW, Mbogo CM, Githure J, Novak R: **Survival of immature *Anopheles arabiensis* (Diptera: Culicidae) in aquatic habitats in Mwea rice irrigation scheme, central Kenya.** *Malar J* 2006, **5**:114.
42. Charlwood JD, Pinto J, Sousa CA, Ferreira C, Petrarca V, do Rosario VE: **'A mate or a meal'-pre-gravid behaviour of female *Anopheles gambiae* from the islands of Sao Tome and Principe, West Africa.** *Malar J* 2003, **2**:9.
43. Ye-Ebiyo Y, Pollack RJ, Kiszewski A, Spielman A: **Enhancement of development of larval *Anopheles arabiensis* by proximity to flowering maize (*Zea mays*) in turbid water and when crowded.** *Am J Trop Med Hyg* 2003, **68**:748-752.
44. Huang J, Walker ED, Vulule J, Miller JR: **Daily temperature profiles in and around Western Kenyan larval habitats of *Anopheles gambiae* as related to egg mortality.** *Malar J* 2006, **5**:87.
45. Haddow AJ: **Measurements of temperature and light in artificial pools with reference to the larval habitat of *Anopheles (Myzomyia) gambiae*, Giles, and *An. (M.) funestus*, Giles.** *Bull Entomol res* 1943, **34**:89-93.
46. Pfaehler O, Oulo DO, Gouagna LC, Githure J, Guerin PM: **Influence of soil quality in the larval habitat on development of *Anopheles gambiae* Giles.** *J Vect Ecol* 2006, **31**:400-405.
47. Grieco JP, Rejmánková E, Achee NL, Klein CN, Andre R, Roberts D: **Habitat suitability for three species of *Anopheles* mosquitoes: Larval growth and survival in reciprocal placement experiments.** *J Vect Ecol* 2007, **32**:176-187.
48. Gray EM, Bradley TJ: **Physiology of desiccation of resistance in *Anopheles gambiae* and *Anopheles arabiensis*.** *Am J Trop Med Hyg* 2005, **73**:553-559.
49. Kirby MJ, Lindsay SW: **Responses of adult mosquitoes of two sibling species, *Anopheles arabiensis* and *Anopheles gambiae s.s.* (Diptera: Culicidae) to high temperatures.** *Bull Entomol Res* 2004, **94**:441-448.
50. Gillies MT, DeMeillon B: **The Anophelinae of Africa South of the Sahara (Ethiopian zoogeographical region).** Johannesburg: South African Institute for Medical Research; 1968.
51. Coluzzi M, Sabatini A, Della Torre A, Di Deco MA, Petrarca V: **A polytene chromosome analysis of the *Anopheles gambiae* species complex.** *Science* 2002, **298**:1415-1418.
52. White GB, Magayuka SA, Boreham PFL: **Comparative studies on sibling species of the *Anopheles gambiae* Giles complex (Diptera: Culicidae): bionomics and vectorial activity of species A and species B at Segera, Tanzania.** *Bull Entomol Res* 1972, **62**:295-317.
53. Foster WA: **Mosquito sugar feeding and reproductive energetics.** *Ann Rev Entomol* 1995, **40**:443-474.

54. Gary RE, Foster WA: *Anopheles gambiae* feeding and survival on honeydew and extra-floral nectar of peridomestic plants. *Med Vet Entomol* 2004, **18**:102-107.
55. Charlwood JD, Jones MDR: Mating behaviour in the mosquito, *Anopheles gambiae* s.l. Close range and contact behaviour. *Phy Entomol* 1979, **4**:111-120.
56. Chambers GM, Klowden MJ: Age of *Anopheles gambiae* Giles at time of mating influences female oviposition. *Jvect Ecol* 2001, **26**:196-201.
57. Ijumba JN, Mwangi RW, Beier JC: Malaria transmission potential of *Anopheles* mosquitoes in the Mwea-Tebere irrigation scheme, Kenya. *Med Vet Entomol* 1990, **4**:425-432.
58. Gillies MT, Wilkes TJ: A study of the age-composition of populations of *Anopheles gambiae* Giles and *An. funestus* Giles in North-Eastern Tanzania. *Bull Entomol Res* 1965, **56**:237-262.
59. Mayagaya VS, Michel K, Benedict MQ, Killeen GF, Wirtz RA, Ferguson HM, Dowell FE: Non-destructive determination of age and species of *Anopheles gambiae* s.l. using near-infrared spectroscopy. *Am J Trop Med Hyg* 2009, **81**:622-630.
60. Charlwood JD, Jones MDR: Mating behaviour in the mosquito, *Anopheles gambiae* s.l. II. Swarming behaviour. *Phy Entomol* 1980, **5**:315-320.
61. Diabate A, Baldet T, Brengues C, Kengne P, Dabire KR, Simard F, Chandre F, Hougard JM, Hemingway J, Ouedraogo JB, et al: Natural swarming behaviour of the molecular M form of *Anopheles gambiae*. *Trans R Soc Trop Med Hyg* 2003, **97**:713-716.
62. Marchand RP: Field observation on swarming and mating in *Anopheles gambiae* mosquitoes in Tanzania. *Nether J zool* 1984, **34**:367-387.
63. Charlwood JD, Pinto J, Sousa CA, Madsen H, Ferreira C, do Rosario VE: The swarming and mating behaviour of *Anopheles gambiae* s.s. (Diptera: Culicidae) from Sao Tome Island. *J Vect Ecol* 2002, **27**:178-183.
64. Charlwood JD, Thompson R, Madsen H: Observations on the swarming and mating behaviour of *Anopheles funestus* from southern Mozambique. *Malar J* 2003, **2**:2.
65. Dao A, Adamou A, Yaro AS, Maïga HM, Kassogue Y, Traoré SF, Lehmann T: Assessment of alternative mating strategies in *Anopheles gambiae*: Does mating occur indoors? *J Med Entomol* 2008, **45**:643-652.
66. Ribeiro JMC, Seulu F, Abose T, Kidane G, Teklehaimanot A: Temporal and spatial distribution of anopheline mosquitoes in an Ethiopian village: implications for malaria control strategies. *Bull WHO* 1996, **74**:299-305.
67. Muturi EJ, Kim C, Jacob B, Murphy S, Novak RJ: Interspecies predation between *Anopheles gambiae* s.s. and *Culex quinquefasciatus* larvae. *J Med Entomol* 2010, **47**:287-290.
68. Koenraad CJM, Takken W: Cannibalism and predation among larvae of the *Anopheles gambiae* complex. *Med Vet Entomol* 2003, **17**:61-66.
69. Gillies MT: Age-groups and the biting cycle in *Anopheles gambiae*. A preliminary investigation. *Bull Entomol Res* 1957, **48**:553-559.
70. Ferguson HM, Read AF: Why is the effect of malaria parasites on mosquito survival still unresolved? *Trends Parasitol* 2002, **18**:256-261.
71. Okumu FO, Titus E, Mbeyela E, Killeen GF, Moore SJ: Limitation of using synthetic human odours to test mosquito repellents. *Malar J* 2009, **8**:150.
72. Marshall JM, Taylor CE: Malaria control with transgenic mosquitoes. *PLoS Med* 2009, **6**:e1000020.

doi:10.1186/1475-2875-9-356

Cite this article as: Ng'habi et al.: Establishment of a self-propagating population of the African malaria vector *Anopheles arabiensis* under semi-field conditions. *Malaria Journal* 2010 **9**:356.

Submit your next manuscript to BioMed Central
and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

