

Evaluation of antibacterial, angiogenic, and osteogenic activities of green synthesized gap-bridging copper-doped nanocomposite coatings

Dan Huang¹
Kena Ma^{1,2}
Xinjie Cai^{1,2}
Xu Yang³
Yinghui Hu¹
Pin Huang¹
Fushi Wang¹
Tao Jiang^{1,2}
Yining Wang^{1,2}

¹The State Key Laboratory Breeding Base of Basic Science of Stomatology (Hubei- MOST) & Key Laboratory of Oral Biomedicine Ministry of Education, School & Hospital of Stomatology, Wuhan University, ²Department of Prosthodontics, Hospital of Stomatology, Wuhan University, Wuhan, China; ³Department of Oral Biology, School of Dental Medicine, University of Pittsburgh, Pittsburgh, PA, USA

Abstract: Titanium (Ti) and its alloys have been widely used in clinics for years. However, their bio-inert surface challenges application in patients with compromised surgical conditions. Numerous studies were conducted to modify the surface topography and chemical composition of Ti substrates, for the purpose of obtaining antibacterial, angiogenic, and osteogenic activities. In this study, using green electrophoretic deposition method, we fabricated gap-bridging chitosan-gelatin (CSG) nanocomposite coatings incorporated with different amounts of copper (Cu; 0.01, 0.1, 1, and 10 mM for Cu I, II, III, and IV groups, respectively) on the Ti substrates. Physicochemical characterization of these coatings confirmed that Cu ions were successfully deposited into the coatings in a metallic status. After rehydration, the coatings swelled by 850% in weight. Mechanical tests verified the excellent tensile bond strength between Ti substrates and deposited coatings. All Cu-containing CSG coatings showed antibacterial property against both Gram-negative *Escherichia coli* and Gram-positive *Staphylococcus aureus*. The antibacterial property was positively correlated with the Cu concentration. In vitro cytocompatibility evaluation demonstrated that activities of bone marrow stromal cells were not impaired on Cu-doped coatings except for the Cu IV group. Moreover, enhanced angiogenic and osteogenic activities were observed on Cu II and Cu III groups. Overall, our results suggested that Cu-doped CSG nanocomposite coating is a promising candidate to functionalize Ti materials with antibacterial, angiogenic, and osteogenic properties.

Keywords: copper, gap-bridging, antibacterial, angiogenic, osteogenic

Introduction

Since the pioneering work of Ödman et al,¹ titanium (Ti) and its alloys have been increasingly used in clinics for biomedical application because of their excellent-physicochemical properties and superior biocompatibility.^{2,3} However, clinicians are still confronting challenges like biomaterial-associated infections (BAIs) and poor bone osseointegration,^{4,5} owing to the lack of antibacterial, osteogenic, and angiogenic activities of bare Ti materials.

A well-known approach to functionalize the Ti surface is to incorporate antibiotics and various growth factors, such as gentamicin, bone morphogenetic protein-2, and vascular endothelial growth factor (VEGF).⁶⁻⁸ However, antibiotics are unstable and can be easily decomposed,⁹ growth factors are generally expensive, and overdose might cause adverse effects. Due to these drawbacks, their clinical application is limited and alternative methods are required. Recently, more and more research projects aiming to improve the performance of Ti-based materials have focused on surface loading of inorganic metal ions, which showed stable and durable biological properties.¹⁰⁻¹²

Correspondence: Tao Jiang; Yining Wang
Department of Prosthodontics, Hospital of Stomatology, Wuhan University, Luoyu Road 237, Hongshan District, Wuhan, 430079, China
Tel +86 27 8768 6318
Fax +86 27 8787 3260
Email jiangtao2006@whu.edu.cn; wang_yn@whu.edu.cn

Among them, copper (Cu) has drawn our attention for its balanced property between antibacterial activity and cytotoxicity.¹³ Similar to many other noble metals, Cu is known to inhibit bacterial viability by denaturing the respiratory enzyme or destroying cell membrane.¹⁴ Moreover, it has demonstrated outstanding ability to stimulate angiogenic and osteogenic differentiation of stem cells.^{15–19} More precisely, Cu was thought to promote angiogenesis by inducing the expression of hypoxia inducible factor (HIF-1 α) and VEGF, therefore artificially mimicking hypoxia, which played a critical role in blood vessel formation. In the work of Yu et al¹⁶ and Burghardt et al,²⁰ Cu-bearing surfaces manifested considerable antibacterial, osteogenic, and angiogenic properties. Inspired by these advantages of Cu, we attempted to introduce this inorganic element in the surface functionalization of biomedical Ti-based materials.

There is a variety of strategies to functionalize the surface of Ti substrates, including plasma immersion ion implantation,¹⁶ hydrothermal method,²¹ magnetron co-sputtering process,²² and electrophoretic deposition (EPD).^{23,24} In our previous studies, EPD technique was routinely used to prepare organic chitosan-gelatin (CSG) nanosphere coating on Ti substrates.^{25–27} It showed favorable mechanical property, swelling ability, biodegradability, and biocompatibility. Moreover, the mild coating condition (room temperature, water solution) and convenient protocol make it a promising candidate for further loading of functional agents like metal ions.^{27,28}

In this study, Cu-doped CSG nanocomposite coatings were successfully fabricated by an environmentally friendly EPD process (green chemistry) without the use of harsh, toxic, and costly chemicals.^{29–31} As far as we know, this is the first time Cu-doped organic-inorganic coatings have been synthesized to functionalize the Ti surface. The antibacterial ability of the coatings against Gram-negative *Escherichia coli* (*E. coli*) and Gram-positive *Staphylococcus aureus* (*S. aureus*) was examined. Furthermore, rat bone marrow mesenchymal stromal cells were used to investigate the cytocompatibility, angiogenic, and osteogenic potential of the coatings. This study may highlight the potential use of the Cu-doped CSG coatings in clinical application.

Materials and methods

Materials

Commercially pure Ti disks (15 mm in diameter, 1 mm thick, grade 2) were provided by Baoji Titanium Industry Co, Ltd (China) and pretreated according to our previous study.²⁵ Briefly, the disks were sandblasted with large grits and acid etched with HCl/H₂SO₄ for 30 minutes at 60°C, and

then ultrasonically cleaned in acetone, ethanol, and Milli-Q water sequentially. Chitosan (MW 1,000,000 Da) with a deacetylation degree >95% was obtained from Golden-Shell Biochemical Co, Ltd (Zhejiang, China). Gelatin (type A, G1890) derived from acid-cured tissue was supplied by Sigma Aldrich (St Louis, MO, USA). All the other chemical reagents were local products of analytical grade.

Preparation of electrophoretic solutions and EPD process

The CSG solution was prepared as in our previous study.²⁵ In brief, 0.6 g chitosan was dissolved in 80 mL 0.04 M hydrochloric acid with magnetic stirring at room temperature for 2 hours. The pH value was adjusted to 4.0 using 0.5 M NaOH; then, 1.4 g gelatin was dissolved into the chitosan solution at 60°C for 1.5 hours, and the total volume of the solution was brought to 100 mL with Milli-Q water. Cu chloride was added into the CSG solution to obtain the final electrophoretic solution. In total, five groups were made, namely CSG, Cu I, Cu II, Cu III, and Cu IV, with Cu concentrations of 0, 0.01, 0.1, 1, and 10 mM (Table 1).

During the process of EPD, the Ti substrate was used as the cathode with a parallel platinum plate as the anode. The distance between the two electrodes was 50 mm. EPD was carried out using a direct current power supply (model 6614C; Agilent Technologies, Santa Clara, CA, USA) by keeping the current at 15 mA for 2 minutes. After the deposition, the coated Ti was disconnected from the power supply, removed from the electrophoretic solutions, rinsed gently with Milli-Q water, and then air-dried overnight for future study.

Characterization of the deposited coatings

Surface topography and physicochemical characterization

Immediately after EPD, the freshly deposited coatings were diluted with Milli-Q water and then examined by transmission electron microscopy (TEM; JEOL JEM 2100; JEOL Ltd, Tokyo, Japan) to observe the contents. Optical photographs of the fresh coatings were taken using a camera

Table 1 Concentration of chitosan, gelatin, and copper chloride in the electrophoretic deposition solution

Concentration	CSG	Cu I	Cu II	Cu III	Cu IV
Chitosan (g/100 mL)	0.6	0.6	0.6	0.6	0.6
Gelatin (g/100 mL)	1.4	1.4	1.4	1.4	1.4
CuCl ₂ (mM)	0	0.01	0.1	1	10

Abbreviation: CSG, chitosan-gelatin.

(Nikon 90; Nikon Corporation, Tokyo, Japan). In addition, the coatings were visualized by fluorescence microscopy (Nikon TE-2000). After being air-dried, optical images of the coatings were obtained again and the coatings were observed using field emission scanning electron microscopy (FESEM; Carl Zeiss Meditec AG, Jena, Germany). For qualitative elemental analysis of the coatings, energy-dispersive X-ray spectrometry (EDS; Quanta-200; FEI, Eindhoven, the Netherlands) was used. Surface chemistry was investigated with attenuated total reflection Fourier transform infrared spectroscopy (FTIR; Thermo Nicolet 5700; Thermo Fisher Scientific, Waltham, MA, USA). In addition, the chemical state of Cu in the coatings was studied by X-ray photoelectron spectroscopy (XPS; Kratos XSAM 800; Kratos Analytical, Manchester, UK).

Cu entrapment and release assay

To investigate the total amount of Cu entrapped in the coatings, fresh coatings were collected and diluted to 10 mL after being treated with hydrogen nitrate (HNO_3); then, the samples were analyzed for concentration of Cu using an atomic absorption spectrophotometer (AAS; ContrAA 700; Analytik Jena AG, Germany). Each group had three disks for the test.

For the Cu release assay, each sample was soaked in 5 mL phosphate-buffered saline (PBS) at 37°C without agitation. At 6 hours, 12 hours, 24 hours, 3 days, 7 days, 14 days, 21 days, and 28 days, all the immersion solution was extracted, and replaced by fresh PBS. The extracted solution was treated with HNO_3 and analyzed by AAS for the concentration of Cu.

In order to observe the morphology of the Cu compounds released from the coatings, the dried coatings were immersed in 5 mL Milli-Q water for 24 hours, and the released suspension was diluted with Milli-Q water followed by TEM analysis.

Swelling ratio

Swelling characteristic of the coatings was determined by immersing the coated Ti substrates into PBS solution for 2, 10, 20, 30, 40, 50, 60, 90, 120 minutes. The swelling ratio was determined as: $\text{SR} = (W_2 - W_1) / (W_1 - W_0)$, where SR is the swelling ratio of the coating, W_0 is the weight of the pure Ti disk, W_1 is the dry weight of the coated disk, and W_2 is the wet weight of the coated disk after immersion in PBS at a certain time point.

Degradation rate

Degradation of the coatings was performed in sterilized PBS (pH 7.4) containing $1.5 \mu\text{g mL}^{-1}$ lysozyme at 37°C . After

incubation at predetermined time intervals (6 hours, 12 hours, 24 hours, 3 days, 7 days, 14 days, 21 days, and 28 days), the samples were removed from the solution, rinsed thoroughly with Milli-Q water, and dried in vacuum for 3 days at room temperature. After measuring the weight, the same disks were put back in degradation solution. The dry weight of each pure Ti disk and coated disk was measured at the start of the assay (W_1 , W_2) and at the specific time point (W_3). The degradation rate of the coatings in vitro was expressed as the percentage mass loss (D), which was calculated as: $D = (W_2 - W_3) / (W_2 - W_1)$.

Mechanical property

In order to investigate the interface between the coatings and Ti substrates, the shear and tensile bond strength were examined according to our previous study.³² Briefly, the coating surface was made to adhere to another Ti plate ($20 \times 10 \times 1 \text{ mm}^3$) or stainless steel studs (10 mm in diameter, 15 mm thick) using instant gel adhesive (Loctite 454; Henkel Loctite, Düsseldorf, Germany), and cured for 24 hours at ambient temperature. The samples were vertically fixed at the crosshead of the electrical mechanical Instron Model 4465 load frame (Instron Corporation, Norwood, MA, USA) and the long axis of the test samples coincided with the direction of applied forces. The mechanical tests were conducted at a constant displacement of 0.50 mm per minute until failure was reached, which was evidenced by a drop in load.

In vitro antibacterial study

Observation of the bacteria by FESEM

E. coli (ATCC 25922) and *S. aureus* (ATCC 25923) were used to assess the antibacterial effect of various samples. Before assay, *E. coli* and *S. aureus* were cultured in Luria-Bertani and tryptic soy broth medium at 37°C on a shaker bed at 200 rpm for 6–12 hours, and the concentration was adjusted to 10^5 – 10^6 colony-forming units mL^{-1} . For the FESEM examination, the specimens were cultured with 2 mL bacterial suspension for 6 hours, and the samples were removed from the medium, fixed overnight at 4°C , dehydrated through a series of ethanol (30, 50, 75, 90, 95, and 100 v/v%), then gold sputtered in vacuum before observation.

Live/Dead staining

In the Live/Dead staining assay, the bacterial solution, which was cultured with the samples for 6 hours, was stained with SYTO 9 and propidium iodide dyes according to the protocol of the Live/Dead BacLight Bacterial Viability Kits (L7012; Thermo Fisher Scientific) and immediately observed under fluorescence microscopy (Nikon TE-2000). In addition, the

number of live and dead bacterial cells per image was analyzed using ImageJ software (National Institutes of Health, Rockville, MD, USA).

Quantitative measurements of antibacterial property

To quantify the antibacterial ability of the samples, the specimens were incubated with bacterial suspension at 37°C for 6 hours; then, the culture medium was diluted and the number of viable bacteria was counted by spread plate method. At the same time, bacteria that adhered on the sample surface were collected by mild ultrasonication in 1 mL culture medium, and counted by the same method. The antibacterial rates on surface and medium were calculated according to the following formulas: 1) antibacterial rate for adherent bacteria on the sample: $R_a = (A-B)/A \times 100\%$ and 2) antibacterial rate for planktonic bacteria in the culture medium: $R_p = (C-D)/C \times 100\%$.³³ A represents the average number of bacteria adhered on the CSG coatings, B is the average number of bacteria adhered on the Cu-doped coatings, C is the average number of viable bacteria in the culture medium of the CSG group, and D is the average number of viable bacteria in the culture medium of Cu-containing groups.

Leakage of protein from bacteria

Similar to the antibacterial test, samples were cocultured with bacterial suspension at 37°C for 6 hours. The supernatant (1 mL) was collected by centrifugation and frozen at -20°C immediately. The concentration of proteins was determined by bicinchoninic acid (BCA) protein assay kit (23235; Thermo Fisher Scientific).

In vitro cellular study

Culture of rat bone marrow stromal cells (BMSCs)

BMSCs were harvested from 6-week-old male Wistar rats according to established procedures. All experimental protocols of animals in this study were approved by the Animal Care and Experiment Committee of Wuhan University, and followed the Guide for the Care and Use of Laboratory Animals of Wuhan University. In brief, after cutting off both ends of rat femurs at the epiphysis, the bone marrow was quickly rinsed out with alpha minimum essential medium (α -MEM; SH30265; Hyclone, Logan, UT, USA) supplemented with 10% fetal bovine serum (SH30068; Hyclone) and 1% penicillin-streptomycin (SV30010; Hyclone). Flushed cells were incubated at 37°C in a humidified 5% CO₂ atmosphere. After 24 hours, the non-adherent cells were removed by changing the culture medium. Adhered cells were cultured with the medium refreshed every 3 days. When cells reached 80%–90% confluence, they were detached by 0.25%

trypsin/EDTA (SH30042; Hyclone) and seeded at a density of $2 \times 10^4/\text{cm}^2$. Cells of passage 2–4 were used in the study.

Cell adhesion and morphology

To observe the cell morphology, cells were seeded directly on the coated Ti samples and examined by FESEM. After 1 day of culture, the cells were gently washed with PBS three times, fixed with 2.5% glutaraldehyde overnight at 4°C, dehydrated through a series of graded ethanol, and gold sputtered in vacuum before FESEM observation.

For the immunofluorescence assay, the BMSCs were fixed with 4% paraformaldehyde for 15 minutes, washed with PBS, and permeabilized with 0.3% Triton X-100 for 15 minutes. The cells were then incubated in 1% bovine serum albumin for 30 minutes to block the nonspecific binding sites. Afterwards, the filamentous actin of the cells was stained by rhodamine phalloidin (R-415 kit; Molecular Probes, Thermo Fisher Scientific) for 20 minutes and the nuclei were stained with 2-(4-amidinophenyl)-6-indolecarbamide dihydrochloride (Thermo Fisher Scientific) for 15 minutes, successively. The samples were then rinsed again with PBS and stored in glycerol. Immunofluorescence images were obtained using fluorescence microscopy (Nikon TE-2000).

Cell proliferation

Cell proliferation was analyzed by cell counting kit-8 (CCK-8; Dojindo Laboratories, Kumamoto, Japan). On day 1, 4, and 7, the cell culture plates were washed with PBS, and supplied with 400 μL α -MEM containing 10% CCK-8. Then, the culture plates were incubated at 37°C for 1 hour and the absorbance value of the supernatant at 450 nm was read using an ELX808 ultra microplate reader (BioTek Instruments Inc, Winooski, VT, USA).

Cytotoxicity of coatings versus pure Cu ions

The cytotoxicity of the deposited coatings was compared with the pure Cu ions. First, medium extracts were obtained by placing the coated disks in 24-well tissue plates with cell culture medium for 24 hours. Then, culture medium with the same concentration of Cu was prepared by adding Cu chloride solution into the medium. Finally, BMSCs were incubated with both kinds of medium for 24 hours; the medium was then displaced by CCK-8 solution, and the optical density value at 450 nm was read.

Real-time quantitative polymerase chain reaction (RT-qPCR) for angiogenesis-related gene expression

At time points of 3, 7, 14 days, RT-qPCR was executed to assess angiogenesis-related gene expressions of various

samples in BMSCs. Total RNA was extracted using 1 mL Trizol reagent (Takara, Shiga, Japan), the complementary DNA (cDNA) was reverse-transcribed from 1 mg total RNA using a PrimeScript 1st Strand cDNA Synthesis Kit (Takara). Afterwards, the expressions of key angiogenic factors including HIF-1 α and VEGF were measured on Bio-Rad MyiQ single color RT-PCR system using an SYBR green PCR kit (Takara). The relative expressions of target genes were normalized to that of the housekeeping gene glyceraldehyde-3-phosphate dehydrogenase.

Alkaline phosphatase (ALP) activity and RT-qPCR for osteogenesis-related gene expression

For ALP activity assay, BMSCs were cultured with medium supplemented with 10 nM dexamethasone, 10 mM β -glycerolphosphate, and 50 μ g/mL ascorbic acid (Sigma Aldrich). After being cultured for 7 and 14 days, cells were lysed with 0.1% Triton X-100. The total cellular ALP activity of the samples was measured by reacting with p-nitrophenyl phosphate (P5994; Sigma Aldrich) in alkaline buffer solution (A9226; Sigma Aldrich) at 37°C for 1 hour. The absorbance value at 405 nm was measured with an enzyme linked immunosorbent assay microplate reader using 4-nitrophenol as standard (0–25 nM; N7660; Sigma Aldrich). The intracellular

total protein content was determined by the BCA kit (23235; Thermo Fisher Scientific). Finally, the ALP activity was normalized to the total protein content.

To further determine the role of Cu-CSG coatings in osteogenic activity in vitro, the expressions of osteogenesis-related genes including ALP, osteocalcin (OCN), runt-related transcription factor 2 (Runx2), and collagen I (Col-1) in BMSCs were evaluated by RT-qPCR according to the method described before.

Statistical analysis

One-way analysis of variance and post hoc Tukey testing were carried out using SPSS 16.0 (SPSS Inc, Chicago, IL, USA) for statistical analysis. *, **, and *** stand for $P < 0.05$, $P < 0.01$, and $P < 0.001$, respectively. $P < 0.05$ was considered to be statistically significant.

Results

Physicochemical characterization of the deposited coatings

Surface topography and microstructural characterization

Figure 1A shows that with increasing amount of Cu in the EPD suspension, the color of the solution changed gradually

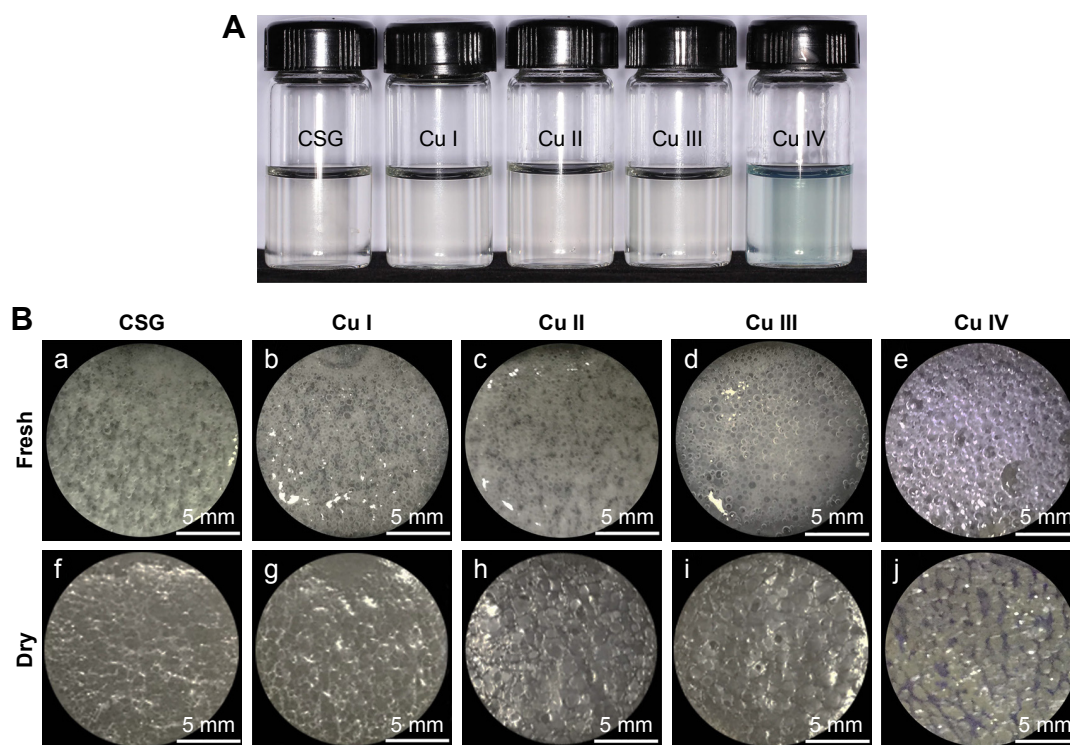


Figure 1 (A) Optical images of the electrophoretic solution and (B) the coatings.

Note: (B) shows optical images of the (a–e) fresh and (f–j) dry coatings.

Abbreviation: CSG, chitosan-gelatin.

from transparent to light blue. Upon visual inspection, porous hydrogel coatings were uniformly deposited on the substrates after EPD (Figure 1B, upper row), and the sample of the Cu IV group presented a purplish color. After being air-dried, the coatings showed a thin film-like structure on the Ti disks (Figure 1B, lower row).

As shown in Figure 2A, monodispersed nanoscale particles were observed in the fresh coatings of all five groups. These particles were formed by a cluster of smaller nanoparticles. Fluorescence images of the wet coatings displayed

macroporous morphology, with pore sizes ranging from 20 to 300 μm (Figure 2Ba–Be). Furthermore, similar to the optical images, the purple fluorescence became obvious with increasing amounts of Cu in the coatings.

Under low magnification of FESEM (Figure 2Bf–Bo), microscale particles were homogeneously distributed in the Cu-containing groups, while the CSG coating displayed a smooth surface morphology. Nanoscale structures were found in high magnification FESEM images (Figure 2Bp–Bt). In the Cu I–III groups, the nanoscale

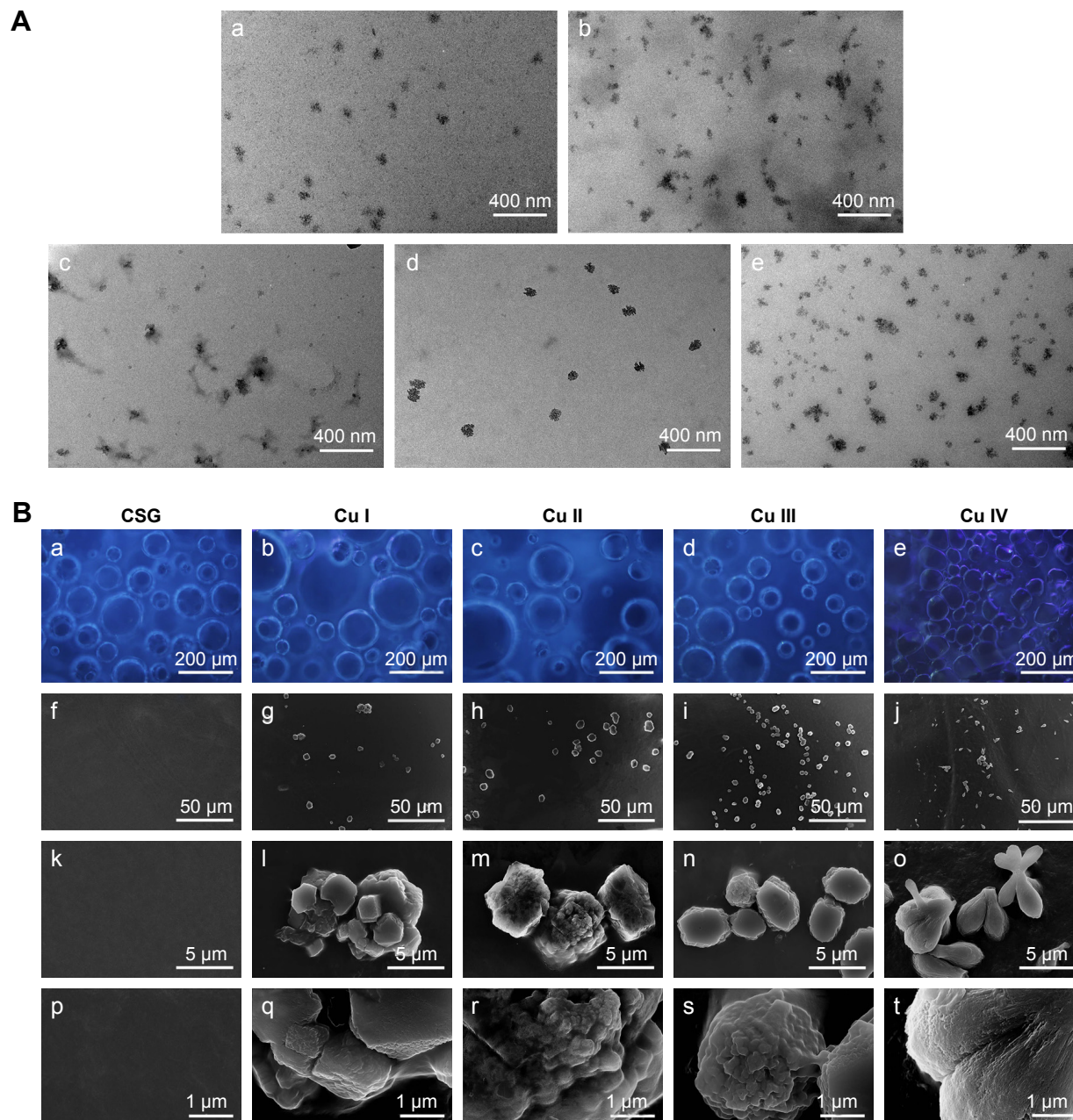


Figure 2 Micro-topography of the coatings. (A) TEM images of the freshly made coatings: (a) CSG; (b) Cu I; (c) Cu II; (d) Cu III; (e) Cu IV. (B) (a–e) Fluorescence images, (f–o) low magnification FESEM, (p–t) high magnification FESEM.

Abbreviations: CSG, chitosan-gelatin; TEM, transmission electron microscopy; FESEM, field emission scanning electron microscopy.

structures exhibited a nanosphere-like structure, while the Cu IV group showed a condensed bundle-like morphology with nano-size diameter.

EDS analysis confirmed the presence of Cu in Cu II–IV coatings while the spectra of Cu I group showed no peak of Cu, probably due to the extreme low concentration (Figure 3A). The relative amount of Cu in the coatings increased from Cu II to IV. CSG coating did not show any Cu peak.

The FTIR spectra (Figure 3B) of all coatings exhibited strong absorption peaks at about 1,151, 1,068, and 1,030 cm^{-1} , which were characteristic of C–N stretching, C–O stretching, and O–H bending in chitosan.³⁴ In addition, the characteristic peaks of gelatin at 1,635, 1,556 cm^{-1} due to amide I, –CONH– stretching and amide II, –NH₂ stretching were also observed.³⁵ In the four Cu-containing groups, slight

changes occurred in the position and intensity of the peaks compared with the CSG group.

The XPS full spectra obtained from the surfaces of samples are shown in Figure 3C. On the basis of these results, the Cu element was detected in the Cu III and Cu IV group with contents of 0.25% and 2.74%, respectively. As presented in Figure 3D, the Cu 2p doublet of the Cu IV group located at 952.25 eV and 932.74 eV corresponded with the binding energies of Cu 2p_{1/2} and Cu 2p_{3/2}, respectively, in metallic Cu.¹⁶

Cu entrapment and release assay

Figure 4A shows that the amount of Cu entrapped in each sample was $0.177 \pm 0.024 \mu\text{g}$ for the Cu I group, $1.308 \pm 0.346 \mu\text{g}$ for the Cu II group, $11.92 \pm 2.154 \mu\text{g}$ for the Cu III group, and $68.29 \pm 2.306 \mu\text{g}$ for the Cu IV group.

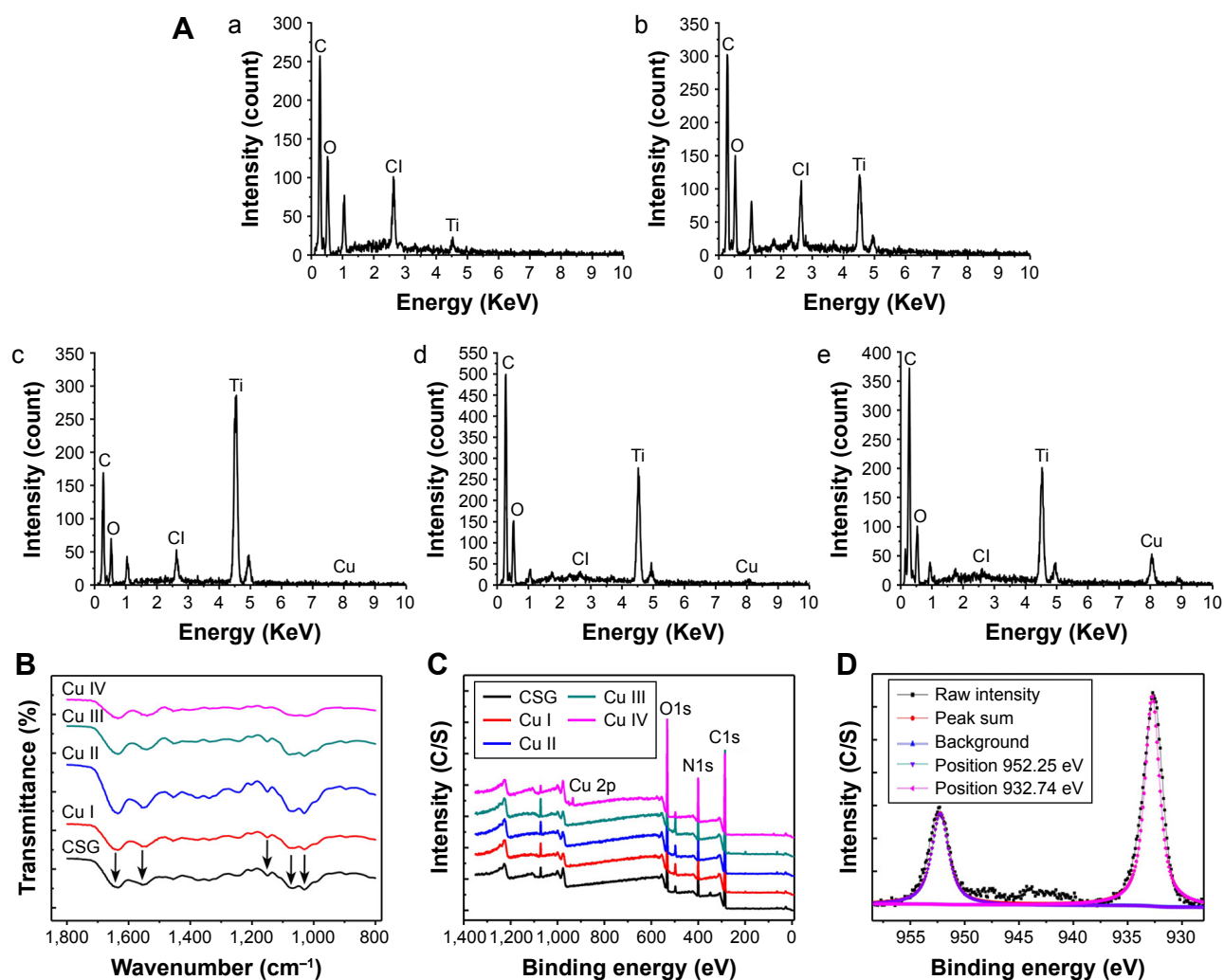


Figure 3 (A) EDS of the coatings: (a) CSG; (b) Cu I; (c) Cu II; (d) Cu III; (e) Cu IV. (B) ATR-FTIR spectra of CSG and Cu-containing coatings. (C) Surface XPS full spectra of coatings with/without Cu doping. (D) High-resolution XPS spectra of Cu 2p of the Cu IV coatings.

Abbreviations: CSG, chitosan-gelatin; EDS, energy-dispersive X-ray spectrometry; ATR-FTIR, attenuated total reflection Fourier transform infrared spectroscopy; XPS, X-ray photoelectron spectroscopy.

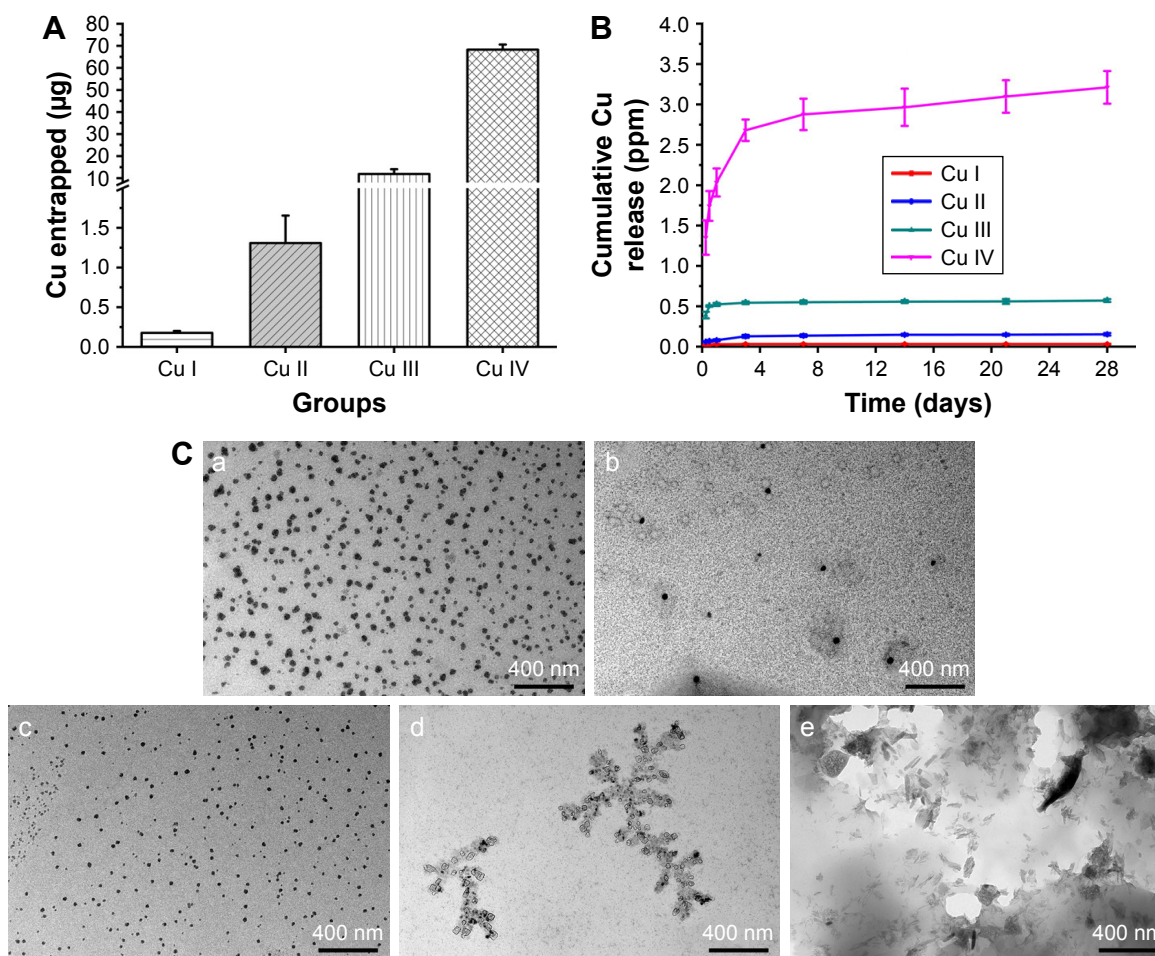


Figure 4 (A) Cu entrapment of the copper-containing coatings. (B) Cumulative release profiles of copper released from copper-doped coatings in PBS. (C) TEM images of the released nanoparticles: (a) CSG; (b) Cu I; (c) Cu II; (d) Cu III; (e) Cu IV.

Abbreviations: CSG, chitosan-gelatin; PBS, phosphate-buffered saline; TEM, transmission electron microscopy.

The percentages of entrapped Cu to the entire coating were about 0.00268%, 0.02%, 0.25%, and 2.2% by weight, respectively, for Cu I–IV groups (data not shown).

In the Cu release assay (Figure 4B), the Cu I, Cu II, and Cu III coatings showed an initial burst release on the first day of observation, and very little amount of Cu was released in the following days. However, in the Cu IV group, sustained release could be observed throughout the whole experiment. Significant differences in the amount of released Cu could be found between Cu I group and Cu II or Cu III group. The percentages of released Cu during the 28 days were about 90%, 58.4%, 23.9%, and 23%, respectively, for Cu I–IV groups.

TEM images (Figure 4C) display the morphology of the nanocomposites released from the coatings. For the CSG, Cu I, and Cu II groups, nanosphere structure could be observed. In the Cu III group, square and oval shaped nano-scale particles appeared, which were arranged in dendritic

structure. For the Cu IV group, the released particles presented a rod-like morphology with diameters of 2–20 nm and lengths of 20–200 nm.

Swelling ratio

As presented in Figure 5A, the swelling ratio of all groups shot up to about 1,300% in 30 minutes. After that, the ratio decreased gradually and remained stable at around 850%. This is also supported by the thickness measurement, which showed that the dried coatings and the PBS-soaked coatings were about 35 and 390 µm, respectively.

Degradation rate

The degradation rate of the coatings is shown in Figure 5B. The weight of all the samples decreased sharply in the first 7 days, followed by relatively little change in the following days. It is noteworthy that the weight loss rate decreased as the amount of Cu in the coating increased. At the end of the

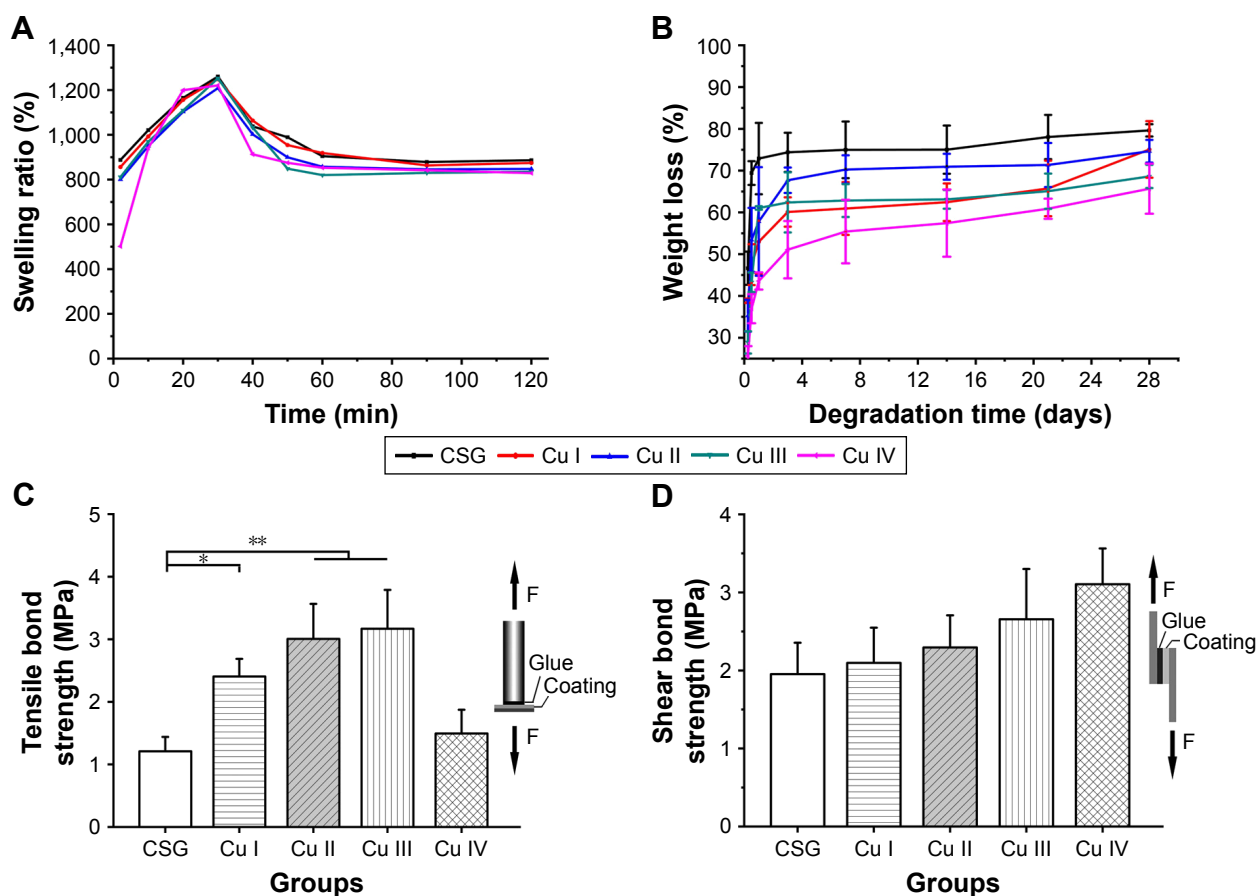


Figure 5 (A) Swelling ratio and (B) degradation rate of all the coatings. (C) Tensile bond strengths and (D) shear bond strengths of different coatings on titanium substrates. $n=3$; * $P<0.05$, ** $P<0.01$.

Abbreviation: CSG, chitosan-gelatin.

study, the weight loss of the CSG and Cu I–IV groups was 79.6%, 75.0%, 74.6%, 68.6%, and 65.7%, respectively.

Mechanical property

During the mechanical tests, all the coatings were detached from the substrate, and failures occurred at the coating–Ti interface rather than the coating–gel interface. As shown in Figure 5C, the tensile bond strength of the coatings increased when Cu was introduced in Cu I–III groups. However, in the Cu IV group, the tensile bond strength had no difference compared with the CSG group. As for the shear bond strength (Figure 5D), there was no significant difference among all the groups.

In vitro antibacterial study

Observation of the bacteria by FESEM

Bacteria adhered on the surface of the coatings were observed by FESEM. It was shown that the amounts of *E. coli* and *S. aureus* on the surfaces decreased with increasing Cu concentration in the coatings (Figures 6A–E and 7A–E).

Furthermore, *E. coli* of CSG, Cu I, Cu II, and Cu III groups showed typical intact and rod-like morphology, while in the Cu IV group, the bacteria were shriveled to spherical shape, even with the membrane perforated (Figure 6E, white arrow). As for *S. aureus*, the bacteria that spread on the CSG, Cu I, and Cu II groups exhibited typical spherical morphology, while shriveled and deformed membranes (Figure 7D and E, white arrow) were detected in Cu III and Cu IV groups.

Live/Dead staining

Figures 6F–T and 7F–T show the fluorescent images of bacteria of various groups after Live/Dead staining. For both *E. coli* and *S. aureus*, as Cu concentration increased from CSG to Cu IV, the green fluorescent intensity (live bacterial colonies) decreased, while the red fluorescent intensity (dead bacterial colonies) increased. A quantitative analysis of Live/Dead staining is displayed in Figure 8A and B. The counts of live bacteria decreased markedly (from 1,444 to 67 for *E. coli*, from 645 to 60 for *S. aureus*) and the counts of dead bacteria drastically increased (from 60 to 1,143 for

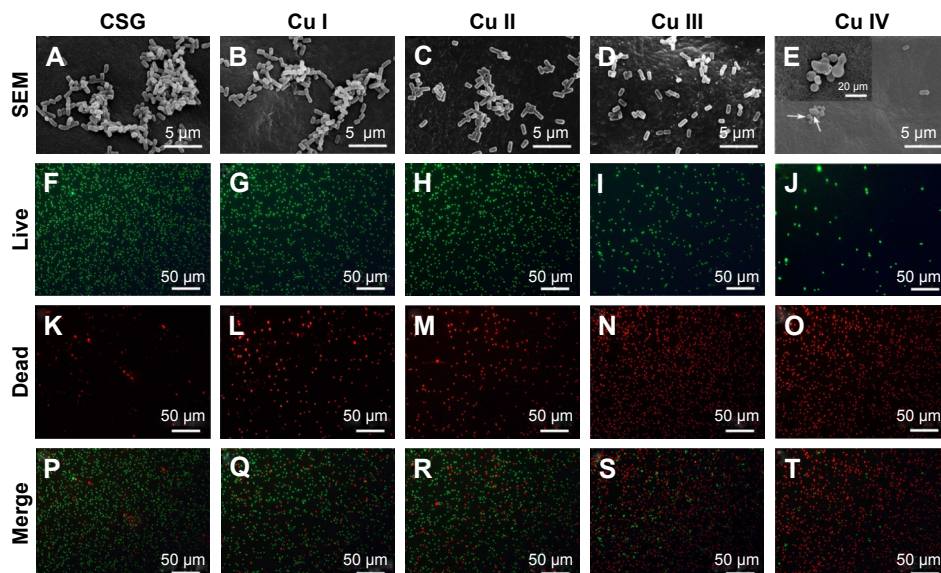


Figure 6 (A–E) FESEM morphologies of *E. coli* adhered on the deposited coatings. The arrows in (E) indicate that the bacteria of group Cu IV were shriveled to spherical shape, even with the membrane perforated. (F–T) Live/Dead stain fluorescent images of *E. coli* after dyeing with SYTO 9 and PI. **Abbreviations:** CSG, chitosan-gelatin; FESEM, field emission scanning electron microscopy; PI, propidium iodide.

E. coli, from 59 to 787 for *S. aureus*) as the concentration of Cu increased.

Quantitative measurements of antibacterial property

To quantify the antibacterial ability of Cu-containing samples against both species, the bacteria that adhered on the surfaces and were suspended in the culture medium were recultivated on agar plates according to the bacteria counting method (Figure 8C). As shown in Figure 8D, for *E. coli*, the Ra values of Cu I, Cu II, Cu III, and Cu IV groups were 12.9%, 31.2%, 83.9%, and 97.8%, respectively, and Rp

values were 21.1%, 37.3%, 85.2%, and 98%, respectively. As for *S. aureus*, Ra was 65.9%, 68.6%, 84.7%, and 91.7%, respectively, and Rp values were 69.4%, 69.2%, 83.2%, and 94.8%, respectively.

Leakage of protein from bacteria

Results suggested that only the Cu IV group could enhance membrane leakage of proteins in *E. coli* compared with the CSG group. As for *S. aureus*, the leakage of proteins in the Cu III and Cu IV coatings was significantly higher than that in the CSG group (Figure 9).

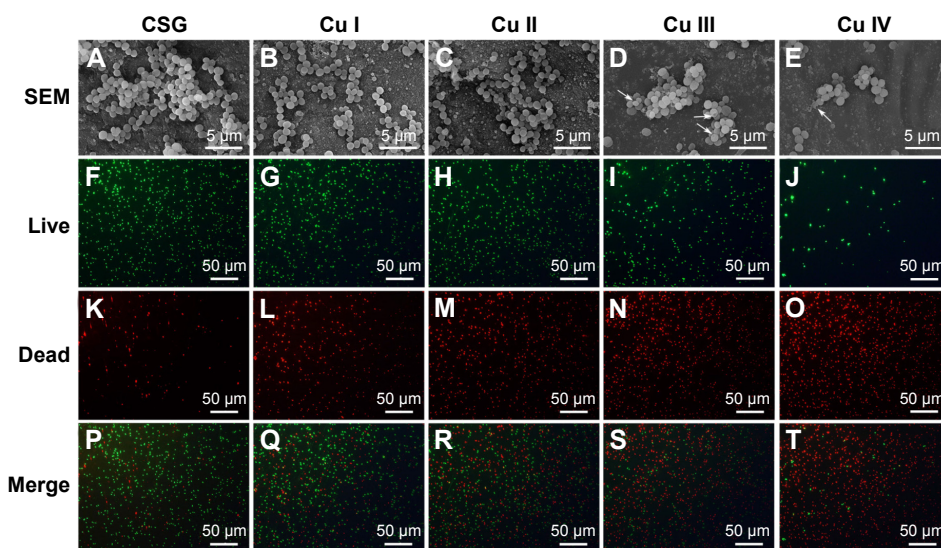


Figure 7 (A–E) FESEM morphologies of *S. aureus* adhered on the deposited coatings. The arrows in D and E indicated that shriveled and deformed membranes were presented by bacteria on Cu III and Cu IV groups. (F–T) Live/Dead stain fluorescent images of *S. aureus* after dyeing with SYTO 9 and PI. **Abbreviations:** CSG, chitosan-gelatin; FESEM, field emission scanning electron microscopy; PI, propidium iodide.

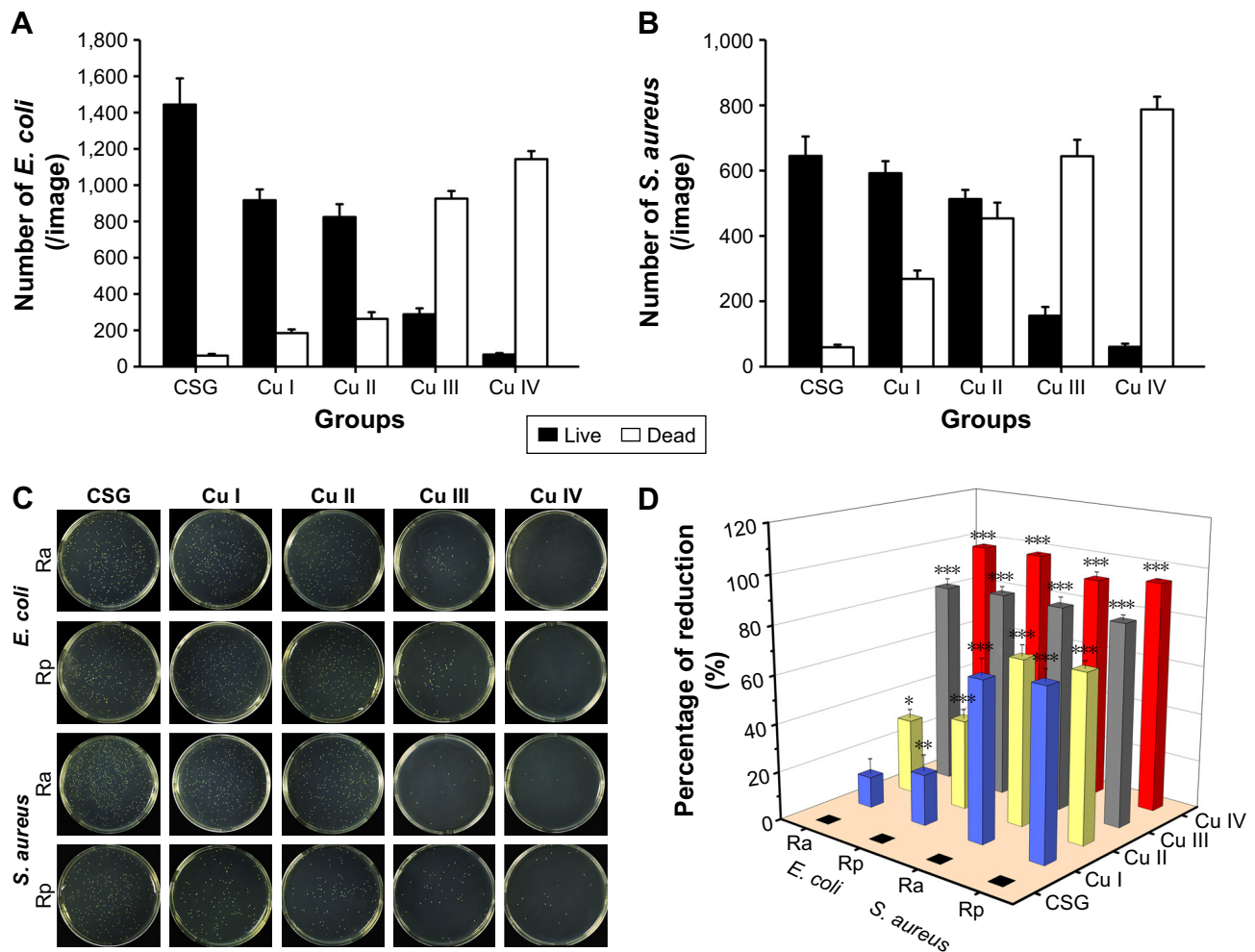


Figure 8 Number of Live/Dead *E. coli* per image (A) and *S. aureus* per image (B) after dyeing with SYTO 9 and PI. (C) Typical photographs of recultivated bacterial colonies on agar and (D) corresponding antibacterial ratios of Ra and Rp. n=3; *P<0.05, **P<0.01, and ***P<0.001 versus the CSG group. **Abbreviations:** CSG, chitosan-gelatin; PI, propidium iodide; Ra, antibacterial rate for adherent bacteria; Rp, antibacterial rate for planktonic bacteria.

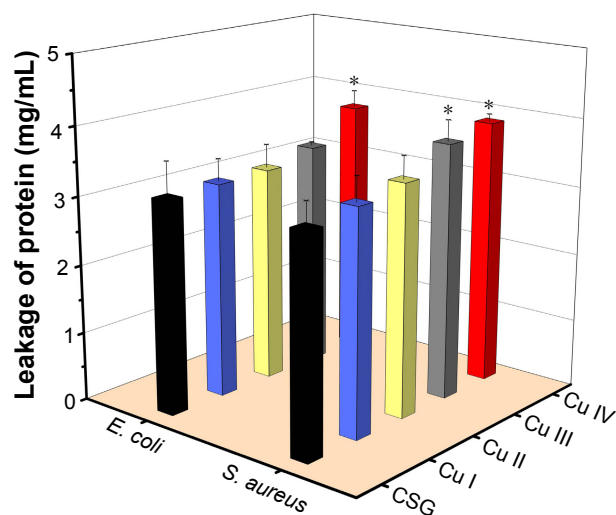


Figure 9 Effects of Cu-containing samples on the membrane leakage of proteins from *E. coli* and *S. aureus* after 6 hours of incubation. n=3; *P<0.05 versus the CSG group. **Abbreviation:** CSG, chitosan-gelatin.

In vitro cellular study

Cell adhesion and morphology

In order to observe cell adhesion and morphology, BMSCs cultured on different samples for 24 hours were examined by FESEM and fluorescence microscopy (Figure 10A). FESEM results indicated that Cu-incorporated coatings could support BMSCs attachment and spreading. In addition, cytoskeleton staining of all groups exhibited a polygonal shape with extended filopodia and lamellipodia (marked with white arrow). No apparent differences were found except that the cell density of the Cu IV group was less than all other groups.

Cell proliferation

The CCK-8 assay results in Figure 10B show the viability and proliferation of BMSCs in various samples. In general, cell population in all samples increased from day 1 to 7 with

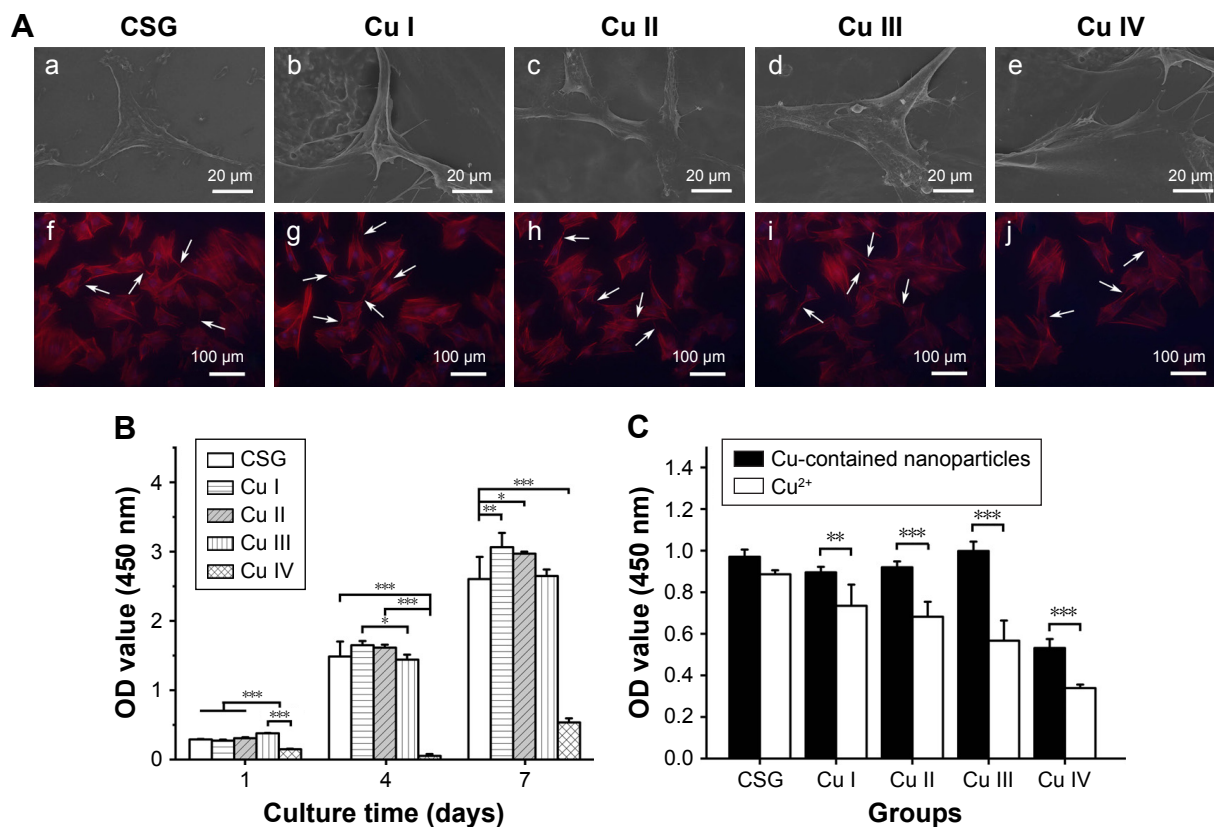


Figure 10 (A) (a–e) FESEM images and (f–j) fluorescence images of BMSCs after 1 day of culture. The arrows indicate the extended filopodia and lamellipodia of the BMSCs. (B) Cell viability of BMSCs cultured at specific time intervals. (C) Cell viability of BMSCs cultured with medium extracts and medium with the same concentration of copper ions. $n=6$; $*P<0.05$, $**P<0.01$, and $***P<0.001$.

Abbreviations: CSG, chitosan-gelatin; FESEM, field emission scanning electron microscopy; BMSCs, bone marrow stromal cells; OD, optical density.

the exception that cell number of the Cu IV group dropped on day 4. Among all Cu deposited coatings, Cu I, II, and III groups manifested similar growth trends as CSG group. On day 7, Cu I and II even showed higher activity than the CSG group. However, cell activity of the Cu IV group was significantly lower than that of the CSG group at all time points and was only 25% of CSG on day 7. This suggests that high Cu concentration can inhibit BMSCs activity.

Cytotoxicity of the coatings compared with pure Cu ions

CCK-8 tests were conducted to compare the cytotoxicity of the Cu organic compounds and pure Cu ions. As shown in Figure 10C, cell activity of the Cu compounds group was significantly higher than Cu ion group with the same concentration in all Cu-doped groups. These results indicated that cytotoxicity of the Cu ions was reduced when assembling with the organic compounds via EPD.

RT-qPCR for angiogenesis-related gene expression

Key angiogenic factors (HIF-1 α and VEGF) in BMSCs were evaluated using RT-qPCR assay after culturing with various samples for 3, 7, and 14 days (Figure 11). The results are

expressed as relative levels over the CSG group. HIF-1 α and VEGF gene expressions followed similar patterns (Cu III > Cu II > Cu I \approx CSG > Cu IV). In particular, Cu II and Cu III groups demonstrated significantly upregulated genes expressions of angiogenic factors compared with the CSG group, while the Cu IV group showed remarkably lower expressions of these genes.

ALP activity and RT-qPCR for osteogenesis-related gene expression

The quantitative analysis shown in Figure 12A revealed that ALP activity of BMSCs in Cu I, Cu II, and Cu III groups increased as Cu loading ascended, and it was significantly higher in the Cu III group than CSG group.

RT-qPCR analysis (Figure 12B–E) show outstanding performance of Cu III coatings. BMSCs expressions of osteogenesis-related genes, including ALP (early marker for osteogenic differentiation), OCN (late marker for osteogenic differentiation), Runx2 (critical transcription factor for bone formation), and Col-1 (main content of bone extracellular matrix) were dramatically upregulated in Cu III samples compared with other groups, especially on day 14. All other groups were in a similar range of expression except that

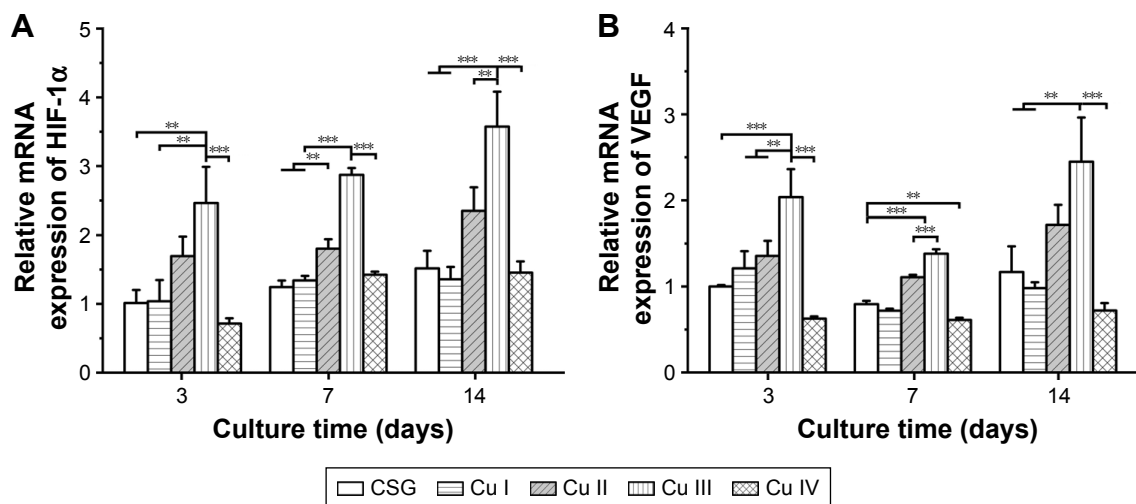


Figure 11 Gene expression levels of angiogenesis-related markers: (A) HIF-1 α and (B) VEGF in BMSCs cultured with various samples at specific time intervals. n=3; ** $p < 0.01$, *** $p < 0.001$.

Abbreviations: CSG, chitosan-gelatin; HIF-1 α , hypoxia inducible factor-1 α ; VEGF, vascular endothelial growth factor; BMSCs, bone marrow stromal cells.

Col-1 expression was apparently lower in the Cu IV group than in the CSG group.

Discussion

In the present study, we have successfully fabricated Cu-doped CSG nanocomposite coatings on Ti substrates by an ecofriendly EPD method (the mechanism is illustrated in

Scheme 1). As described in our previous study, chitosan is a linear polysaccharide with a pK_a value near 6.3,³⁶ and it can be dissolved in HCl solution (pH =4) due to protonation of amino groups. Once gelatin is dissolved in chitosan solution, colloidal polyelectrolyte complexes are formed by the electrostatic interaction between $-\text{NH}_3^+$ groups on chitosan and $-\text{COO}^-$ groups on gelatin. When Cu chloride solution

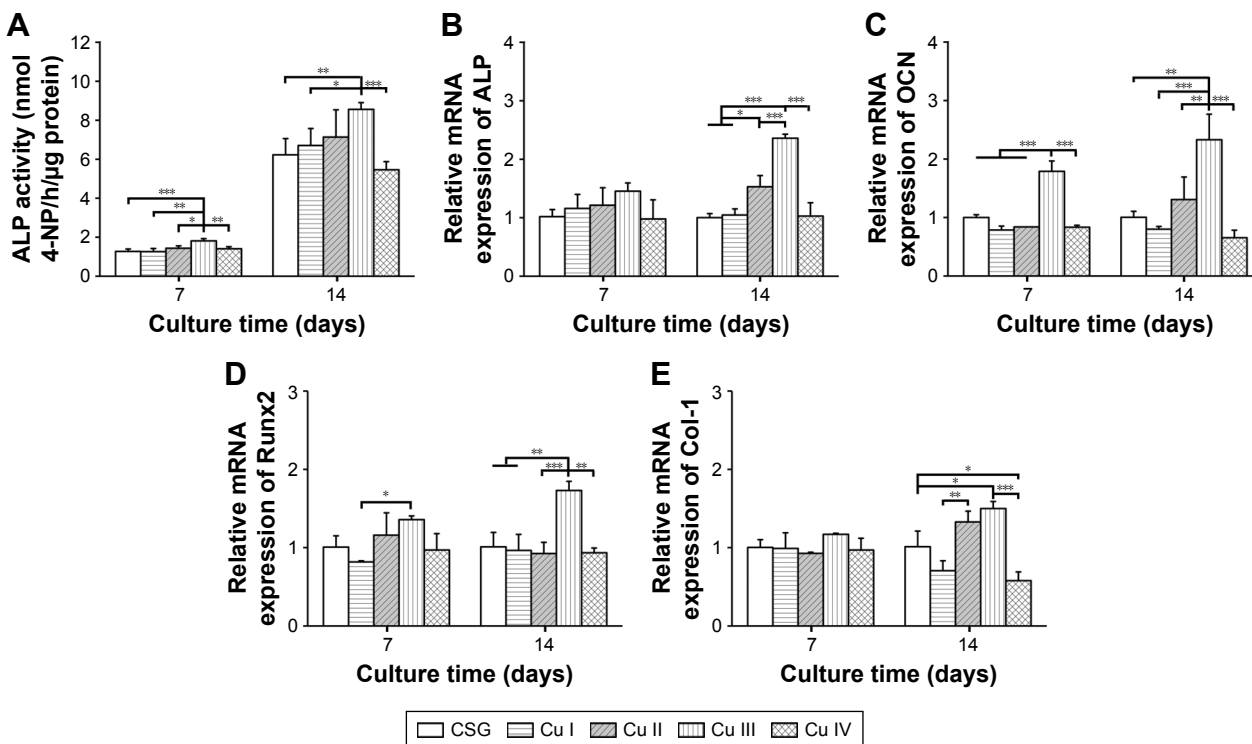
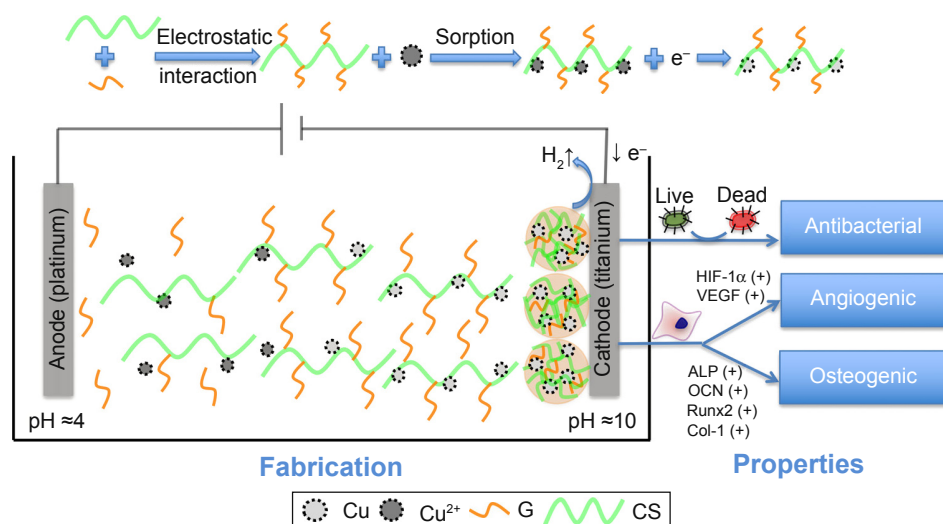


Figure 12 (A) ALP activity and gene expression levels of osteogenesis-related markers: (B) ALP, (C) OCN, (D) Runx2, (E) Col-1 in BMSCs cultured with various samples at specific time intervals. n=3; * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$.

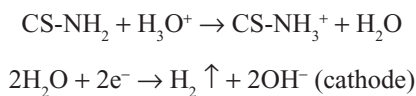
Abbreviations: CSG, chitosan-gelatin; ALP, alkaline phosphatase; OCN, osteocalcin; Runx2, runt-related transcription factor 2; Col-1, collagen I; BMSCs, bone marrow stromal cells.



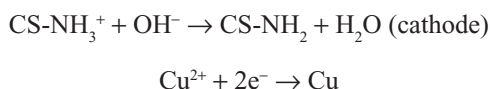
Scheme 1 Fabrication and properties of Cu-CSG nanocomposite coatings via EPD.

Abbreviations: G, gelatin; CS, chitosan; CSG, chitosan-gelatin; EPD, electrophoretic deposition; HIF-1 α , hypoxia inducible factor-1 α ; VEGF, vascular endothelial growth factor; ALP, alkaline phosphatase; OCN, osteocalcin; Runx2, runt-related transcription factor 2; Col-1, collagen I.

was added into the CSG suspension, a stable colloid system of Cu-CSG complexes was formed through chelation or electrostatic attraction between Cu and chitosan.³⁷ During the EPD process, the pH value around the cathode rose up to 10 due to the electrolysis of water,³⁸ which produced bubbles of hydrogen gas.



Consequently, chitosan confronted a high pH than its pK_a near the cathode, where chitosan's amine groups were deprotonated and became insoluble. In this way, the Cu-CSG organic-inorganic complexes were deposited on the cathode plate together with trapped hydrogen bubbles (Figures 1B, upper row and 2B1–B5), which made the coating porous after being air-dried (Figure 1B, lower row). In this process, Cu ions were reduced to metallic Cu, which were embedded in the polymer coatings. As shown in Figure 1B, the coatings became more purple, indicating the reduction of Cu^{2+} ions to metallic Cu.³⁹ This speculation was also identified by the XPS result (Figure 3D), which showed the presence of metallic Cu in the coating of the Cu IV group, while the other groups did not show the peak of Cu probably because of the low content. In addition, the organic chitosan and gelatin in the coatings can work as stabilizers to protect metallic Cu against oxidation.^{40,41}



FESEM (Figure 2Bf–Bo) demonstrated that microparticles appeared in the polymer matrix after the addition of Cu, and its density increased as the Cu concentration became higher. Particularly, these microparticles were composed of smaller nanoscale structures (Figure 2Bp–Bt). Combining these with the TEM results of fresh coatings (Figure 2A), we speculated that during the drying process, nanoparticles underwent aggregation and developed into the microparticles.

All Cu-containing nanocomposite coatings presented a biphasic release of Cu with an initial burst release followed by sustained release (Figure 4B). The burst release profile could assure high drug concentration in the local area, whereas continuous release in the following days guaranteed the enduring effects of the functionalized coatings. These results indicated that the organic CSG coating could serve as a drug carrier enabling the enduring release of Cu. In addition, Cu compounds released from the coatings displayed nanoscale structures (Figure 4C).

The study on the swelling ratio of the coatings indicated that incorporation of Cu with organic CSG coatings did not influence the swelling property significantly (Figure 5A). All dried coatings could swell >10 times after immersion in PBS solution, and recovered to three-dimensional macroporous structures. The enlargement in volume enabled the coating to fill in small gaps (gap-bridging) surrounding the substrate, which was crucial for its stabilization,⁴ while most inorganic coatings did not have such a property. Tensile bond strength in the substrate coating interface is another important factor affecting stabilization. In our previous study, the CSG coating

alone could withstand interfacial stress during surgical insertion process.²⁶ In the current study, the tensile bone strength of the CSG coating was even enhanced by the incorporation of Cu (Figure 5C).

Nowadays, BAI is becoming one of the major causes for implantation failure.⁴² On one hand, surgeons should follow strict disinfection procedures to avoid contamination. On the other, demands were made for Ti substrates with antibacterial capability. Our results demonstrated that the Cu-containing nanocomposite coatings exhibited antibacterial activity against both *E. coli* and *S. aureus*, the most common Gram-negative and Gram-positive microbial pathogens in BAI, respectively (Figures 6–8). With the increasing Cu concentration in CSG coating, the antibacterial ability became more prominent. The results of the present study are in agreement with other reports that proposed the antibacterial ability of Cu. Furthermore, Cu gained importance as a novel antimicrobial agent due to its strong antibacterial activity against not only traditional microorganisms but also several multidrug-resistant organisms.^{43,44} Compared with other metal ions (including silver, zinc, cadmium, lead), Cu has displayed higher antimicrobial efficacy against resistant strains under certain conditions.^{45,46} In addition, researchers found that Cu ions could destroy the permeability of bacterial membranes, promote the generation of bacterial-killing reactive oxygen species (ROS), and disturb the process of gene replication.¹⁴ In our research, similar results showed that Cu-bearing coatings undermined the permeability of bacterial membranes, and led to the leakage of bacterial proteins (Figure 9). However, further research is needed to illustrate the exact mechanism of antibacterial ability of Cu-CSG coatings.

It has been widely accepted that surface chemistry of biomaterials plays a decisive role in biological reactions of surrounding tissue and cells.^{47,48} In this study, we investigated the biocompatibility of Cu-CSG coatings using BMSCs, which is an important cell source in bone tissue engineering. Cell proliferation tests revealed that all Cu-doped coatings were favorable for the cell proliferation except Cu IV group (Figure 10B). One explanation would be that excessive Cu was released from Cu IV coatings and led to the formation of abundant hydroxyl radicals which were toxic to cells.⁴⁹ Interestingly, we noticed that compared with free Cu ions, the Cu-CSG group with the same Cu concentration in cell culture medium displayed significantly higher cell activity (Figure 10C), indicating that the cytotoxicity of the inorganic Cu ions was actually reduced when co-deposited with the organic CSG polymer via EPD. One possible reason for this

phenomenon was that the combination of Cu with biocompatible chitosan could reduce its cytotoxicity by preventing the generation of ROS.⁵⁰

Osteogenesis and angiogenesis are intricately linked, and osteogenesis would not be possible without angiogenesis.^{51–53} Many studies revealed that Cu could promote angiogenesis, possibly through mimicking the hypoxic microenvironment, where HIF-1 α and VEGF were activated to promote neovascularization and tissue regeneration.^{49,54} Our results demonstrated that Cu-doped coatings induced angiogenesis-related gene expression of BMSCs in a dose-dependent manner (Figure 11). Expressions of HIF-1 α and VEGF in Cu II and Cu III groups were significantly elevated compared with the CSG group, whereas the high concentration of Cu in Cu IV group inhibited the expression. The results of ALP activity and RT-qPCR of osteogenesis-related genes showed that osteogenic differentiation of BMSCs on the Cu-containing coatings increased steadily as Cu entrapment ascended, but detrimental influence was displayed in the Cu IV group (Figure 12). Moreover, we also found that the expression of collagen II (Col-2; typical for chondrogenesis) did not increase significantly when the coatings were incorporated with Cu, and the expression level of Col-1 was extremely higher compared with that of Col-2 (Figure S1A and B). These results further confirmed our hypothesis that Cu-doped materials in our study had osteogenic activities but no chondrogenic abilities. Furthermore, Cu-doped coatings did not affect the expression of collagen X (Figure S1C), indicating that there was no hypertrophy during the osteogenic differentiation of BMSCs.⁵⁵ Therefore, our results indicated that Cu element in an appropriate concentration range could improve angiogenic and osteogenic activities of CSG coatings.

To determine the proper Cu concentration with balanced antibacterial activity and low cytotoxicity, we involved four concentrations ranging from 0.01 to 10 mM. The antibacterial activity increased as Cu concentration increased and peaked at 10 mM. However, at this concentration, the coating showed drastic cytotoxicity. Taking together the effects on angiogenic and osteogenic genes expression, we believe that a Cu concentration around 1 mM will guarantee considerable antibacterial activity and benefit angiogenic and osteogenic differentiation while cell activity is not greatly disturbed.

Conclusion

For the first time, we successfully synthesized a Cu-doped organic-inorganic CSG coating on Ti substrate using green EPD technique. The obtained Cu-CSG nanocomposite

coating, which is a gap-bridging coating with low cytotoxicity, was demonstrated to possess antibacterial, angiogenic, and osteogenic properties. Overall, the Cu-CSG coating showed great potential for improving the biological performance of Ti-based materials, although in vivo evaluation is necessary for further research.

Acknowledgments

This study was supported by the National Natural Science Foundation of China (no 81371170, 81470771, and 81571011) and the Fundamental Research Funds for the Central Universities (no 2042017kf0075).

Author contributions

All authors contributed toward data analysis, drafting and revising the paper and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

References

- Ödman J, Lekholm U, Jemt T, Brånemark PI, Thilander B. Osseointegrated titanium implants – a new approach in orthodontic treatment. *Eur J Orthod*. 1988;10(2):98–105.
- Geetha M, Singh AK, Asokamani R, Gogia AK. Ti based biomaterials, the ultimate choice for orthopaedic implants – a review. *Prog Mater Sci*. 2009;54(3):397–425.
- Liu X, Chu PK, Ding C. Surface modification of titanium, titanium alloys, and related materials for biomedical applications. *Mater Sci Eng R Rep*. 2003;47(3–4):49–121.
- Raphel J, Holodniy M, Goodman SB, Heilshorn SC. Multifunctional coatings to simultaneously promote osseointegration and prevent infection of orthopaedic implants. *Biomaterials*. 2016;84:301–314.
- Gristina AG. Biomaterial-centered infection: microbial adhesion versus tissue integration. *Science*. 1987;237(4822):1588–1595.
- Stigter M, Bezemer J, Groot KD, Layrolle P. Incorporation of different antibiotics into carbonated hydroxyapatite coatings on titanium implants, release and antibiotic efficacy. *J Control Release*. 2004;99(1):127–137.
- Wu C, Chang J. Multifunctional mesoporous bioactive glasses for effective delivery of therapeutic ions and drug/growth factors. *J Control Release*. 2014;193:282–295.
- Schliephake H, Rublack J, Förster A, Schwenzer B, Reichert J, Scharnweber D. Functionalization of titanium implants using a modular system for binding and release of VEGF enhances bone-implant contact in a rodent model. *J Clin Periodontol*. 2015;42(3):302–310.
- Camposcia D, Montanaro L, Speziale P, Arciola CR. Antibiotic-loaded biomaterials and the risks for the spread of antibiotic resistance following their prophylactic and therapeutic clinical use. *Biomaterials*. 2010;31(25):6363–6377.
- Mouriño V, Cattalini JP, Boccaccini AR. Metallic ions as therapeutic agents in tissue engineering scaffolds: an overview of their biological applications and strategies for new developments. *J R Soc Interface*. 2011;9(68):401–419.
- Ren N, Li J, Qiu J, et al. Nanostructured titanate with different metal ions on the surface of metallic titanium: a facile approach for regulation of rBMSCs fate on titanium implants. *Small*. 2014;10(15):3169–3180.
- Kim HS, Kim YJ, Jang JH, Park JW. Surface engineering of nanostructured titanium implants with bioactive ions. *J Dent Res*. 2016;95(5):558–565.
- Heidenau F, Mittelmeier W, Detsch R, et al. A novel antibacterial titania coating: metal ion toxicity and in vitro surface colonization. *J Mater Sci Mater Med*. 2005;16(10):883–888.
- Li M, Ma Z, Zhu Y, et al. Toward a molecular understanding of the antibacterial mechanism of copper-bearing titanium alloys against *Staphylococcus aureus*. *Adv Healthcare Mater*. 2016;5(5):557–566.
- Wang H, Zhao S, Zhou J, et al. Evaluation of borate bioactive glass scaffolds as a controlled delivery system for copper ions in stimulating osteogenesis and angiogenesis in bone healing. *J Mater Chem B*. 2014;2(48):8547–8557.
- Yu L, Jin G, Ouyang L, Wang D, Qiao Y, Liu X. Antibacterial activity, osteogenic and angiogenic behaviors of copper-bearing titanium synthesized by PIII&D. *J Mater Chem B*. 2016;4(7):1296–1309.
- Wu Q, Li J, Zhang W, et al. Antibacterial property, angiogenic and osteogenic activity of Cu-incorporated TiO₂ coating. *J Mater Chem B*. 2014;2(39):6738–6748.
- Wu C, Zhou Y, Xu M, et al. Copper-containing mesoporous bioactive glass scaffolds with multifunctional properties of angiogenesis capacity, osteostimulation and antibacterial activity. *Biomaterials*. 2013;34(2):422–433.
- Zheng K, Dai X, Lu M, Hüser N, Taccardi N, Boccaccini AR. Synthesis of copper-containing bioactive glass nanoparticles using a modified Stöber method for biomedical applications. *Collids Surf B Biointerfaces*. 2016;150(2017):159–167.
- Burghardt I, Lüthen F, Prinz C, et al. A dual function of copper in designing regenerative implants. *Biomaterials*. 2015;44(44):36–44.
- Zhao L, Wang H, Huo K, et al. The osteogenic activity of strontium loaded titania nanotube arrays on titanium substrates. *Biomaterials*. 2013;34(1):19–29.
- Andersen OZ, Offermanns V, Sillassen M, et al. Accelerated bone ingrowth by local delivery of strontium from surface functionalized titanium implants. *Biomaterials*. 2013;34(24):5883–5890.
- Besra L, Liu M. A review on fundamentals and applications of electrophoretic deposition (EPD). *Prog Mater Sci*. 2007;52(1):1–61.
- Song J, Chen Q, Zhang Y, et al. Electrophoretic deposition of chitosan coatings modified with gelatin nanospheres to tune the release of antibiotics. *ACS Appl Mater Interfaces*. 2016;8(22):13785–13792.
- Jiang T, Zhang Z, Zhou Y, et al. Surface functionalization of titanium with chitosan/gelatin via electrophoretic deposition: characterization and cell behavior. *Biomacromolecules*. 2010;11(5):1254–1260.
- Ma K, Cai X, Zhou Y, Zhang Z, Jiang T, Wang Y. Osteogenic property of a biodegradable three-dimensional macroporous hydrogel coating on titanium implants fabricated via EPD. *Biomed Mater*. 2014;9(1):015008.
- Cai X, Ma K, Zhou Y, Jiang T, Wang Y. Surface functionalization of titanium with tetracycline loaded chitosan–gelatin nanosphere coatings via EPD: fabrication, characterization and mechanism. *RSC Adv*. 2016;6(9):7674–7682.
- Ma K, Dan H, Jing C, et al. Surface functionalization with strontium-containing nanocomposite coatings via EPD. *Collids Surf B Biointerfaces*. 2016;146:97–106.
- Tokarek K, Hueso JL, Kutrowski P, Stochel G, Kyzio A. Green synthesis of chitosan-stabilized copper nanoparticles. *Eur J Inorg Chem*. 2013;2013(28):4940–4947.
- Thekkae Padil VV, Černik M. Green synthesis of copper oxide nanoparticles using gum karaya as a biotemplate and their antibacterial application. *Int J Nanomedicine*. 2013;8(8):889–898.
- Iravani S. Green synthesis of metal nanoparticles using plants. *Green Chem*. 2011;13(10):2638–2650.
- Zhang Z, Jiang T, Ma K, Cai X, Zhou Y, Wang Y. Low temperature electrophoretic deposition of porous chitosan/silk fibroin composite coating for titanium biofunctionalization. *J Mater Chem*. 2011;21(21):7705–7713.

33. Zhao L, Wang H, Huo K, et al. Antibacterial nano-structured titania coating incorporated with silver nanoparticles. *Biomaterials*. 2011; 32(24):5706–5716.
34. Manjubala I, Scheler S, BöSSERT J, Jandt KD. Mineralisation of chitosan scaffolds with nano-apatite formation by double diffusion technique. *Acta Biomater*. 2006;2(1):75–84.
35. Hong S, Hu X, Fei Y, Bei J, Wang S. Combining oxygen plasma treatment with anchorage of cationized gelatin for enhancing cell affinity of poly(lactide-co-glycolide). *Biomaterials*. 2007;28(29): 4219–4230.
36. Sorlier P, Denuzière A, Viton C, Domard A. Relation between the degree of acetylation and the electrostatic properties of chitin and chitosan. *Biomacromolecules*. 2001;2(3):765–772.
37. Guibal E. Interactions of metal ions with chitosan-based sorbents: a review. *Sep Purif Technol*. 2004;38(1):43–74.
38. Cheng Y, Luo X, Betz J, et al. In situ quantitative visualization and characterization of chitosan electrodeposition with paired sidewall electrodes. *Soft Matter*. 2010;6(14):3177–3183.
39. Patel A, Shukla P, Rufford T, et al. Catalytic reduction of NO by CO over copper-oxide supported mesoporous silica. *Appl Catal A Gen*. 2011;409(23):55–65.
40. Usman MS, El Zowalaty ME, Shameli K, Zainuddin N, Salama M, Ibrahim NA. Synthesis, characterization, and antimicrobial properties of copper nanoparticles. *Int J Nanomedicine*. 2013;8(1): 4467–4479.
41. Chatterjee AK, Sarkar RK, Chattopadhyay AP, Aich P, Chakraborty R, Basu T. A simple robust method for synthesis of metallic copper nanoparticles of high antibacterial potency against *E. coli*. *Nanotechnology*. 2012;23(8):85103–85113.
42. Busscher HJ, van der Mei HC, Subbiahdoss G, et al. Biomaterial-associated infection: locating the finish line in the race for the surface. *Sci Transl Med*. 2012;4(153):2569–2575.
43. Paladini F, Pollini M, Sannino A, Ambrosio L. Metal-based antibacterial substrates for biomedical applications. *Biomacromolecules*. 2015; 16(7):1873–1885.
44. Grass G, Rensing C, Solioz M. Metallic copper as an antimicrobial surface. *Appl Environ Microbiol*. 2011;77(5):1541–1547.
45. Michels HT, Noyce JO, Keevil CW. Effects of temperature and humidity on the efficacy of methicillin-resistant *Staphylococcus aureus* challenged antimicrobial materials containing silver and copper. *Lett Appl Microbiol*. 2009;49(2):191–195.
46. Chudobova D, Dostalova S, Ruttkay-Nedecky B, et al. The effect of metal ions on *Staphylococcus aureus* revealed by biochemical and mass spectrometric analyses. *Microbiol Res*. 2015;170(4687):147–156.
47. Fernandezyague MA, Abbah SA, Mcnamara L, Zeugolis DI, Pandit A, Biggs MJ. Biomimetic approaches in bone tissue engineering: integrating biological and physicochemical strategies. *Adv Drug Deliv Rev*. 2015;84:1–29.
48. Yang X, Wang D, Liang Y, et al. A new implant with solid core and porous surface: the biocompatibility with bone. *J Biomed Mater Res A*. 2014;102(7):2395–2407.
49. Martin F, Linden T, Katschinski DM, et al. Copper-dependent activation of hypoxia-inducible factor (HIF)-1: implications for ceruloplasmin regulation. *Blood*. 2005;105(12):4613–4619.
50. Worthington KL, Adamcakovadodd A, Wongrakpanich A, et al. Chitosan coating of copper nanoparticles reduces in vitro toxicity and increases inflammation in the lung. *Nanotechnology*. 2013;24(39):395101.
51. Carano RAD, Filvaroff EH. Angiogenesis and bone repair. *Drug Discov Today*. 2003;8(21):980–989.
52. Bose S, Fielding G, Tarafder S, Bandyopadhyay A. Understanding of dopant-induced osteogenesis and angiogenesis in calcium phosphate ceramics. *Trends Biotechnol*. 2013;31(10):594–605.
53. Riddle RC, Khatri R, Schipani E, Clemens TL. Role of hypoxia-inducible factor-1 α in angiogenic–osteogenic coupling. *J Mol Med (Berl)*. 2009;87(6):583–590.
54. Weidemann A, Johnson RS. Biology of HIF-1 α . *Cell Death Differ*. 2008;15(4):621–627.
55. Pelttari K, Winter A, Steck E, et al. Premature induction of hypertrophy during in vitro chondrogenesis of human mesenchymal stem cells correlates with calcification and vascular invasion after ectopic transplantation in SCID mice. *Arthritis Rheum*. 2006;54(10):3254–3266.

Supplementary material

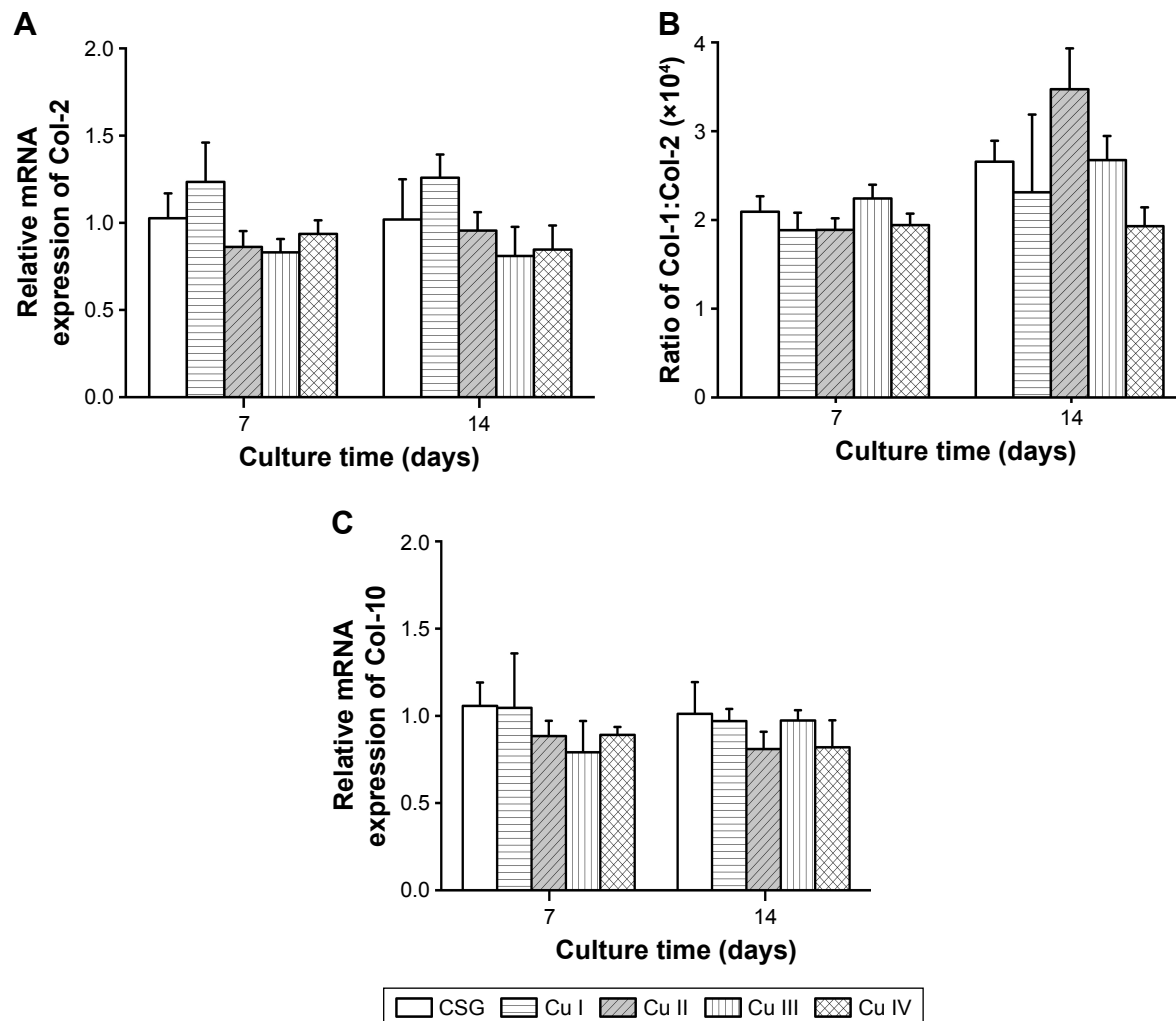


Figure S1 (A) Gene expression level of collagen II, (B) ratio of collagen I:collagen II, and (C) collagen X in BMSCs cultured with various samples at specific time intervals.

Notes: n=3; no significant differences were found between the CSG group and copper-containing groups.

Abbreviations: CSG, chitosan-gelatin; Col-I, collagen I; Col-2, collagen II; Col-10, collagen X; BMSCs, bone marrow stromal cells.

International Journal of Nanomedicine

Publish your work in this journal

The International Journal of Nanomedicine is an international, peer-reviewed journal focusing on the application of nanotechnology in diagnostics, therapeutics, and drug delivery systems throughout the biomedical field. This journal is indexed on PubMed Central, MedLine, CAS, SciSearch®, Current Contents®/Clinical Medicine,

Submit your manuscript here: <http://www.dovepress.com/international-journal-of-nanomedicine-journal>

Dovepress

Journal Citation Reports/Science Edition, EMBase, Scopus and the Elsevier Bibliographic databases. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.